Electronic Supplementary Information

Uranyl Nitrate as a Recyclable Homogeneous Photocatalyst for

Selective Cross-Coupling of N-Substituted Amines and Indoles

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I Mechanistic Study

1. Radical Quenching Experiment

For the cross-coupling reaction of secondary anilines and indoles:

The reaction was conducted under standard conditions with 5 equiv 2,2,6,6-tetramethyl-1piperinedinyloxy (TEMPO).



Scheme S1. Radical quenching experiment of 3a.

For the cross-coupling reaction of tetrahydroisoquinolines and indoles:

The reaction was conducted under standard conditions with 2.0 equiv 2,2,6,6-tetramethyl-1piperinedinyloxy (TEMPO).



Scheme S2. Radical quenching experiment of 5a.

2. Ultraviolet-Visible Absorption Experiments

Ultraviolet-visible absorption experiments were performed using a Hitachi U-2910 spectrophotometer. In each experiment, the varying samples were combined in MeCN in screw-top 1.0 cm quartz cuvettes. The concentration of each component was 4×10^{-3} M. Uranyl cation was approved to serve as photosensor at 420 nm.



Figure S1. UV-Vis experiments of UO₂(NO₃)₂·6H₂O, trifluoroacetic acid, ethyl (4methoxyphenyl)glycinate (**1a**) and 1*H*-indole (**2a**).

Ultraviolet-visible absorption experiments were performed using a Hitachi U-2910 spectrophotometer. In each experiment, the varying samples were combined in the mixture solvent $H_2O/MeCN$ (v/v 1:1) in screw-top 1.0 cm quartz cuvettes. The concentration of each component was 4×10^{-3} M. Uranyl cation was approved to serve as photosensor at 416 nm.



Figure S2. UV-Vis experiments of $UO_2(NO_3)_2 \cdot 6H_2O$, *N*-phenyl-tetrahydroisoquinoline (**4a**) and 1,2-dimethyl-1*H*-indole (**2d**).

3. Stern-Volmer Fluorescence Quenching Experiments with UO₂(NO₃)₂·6H₂O

Fluorescence quenching studies were performed using a Hitachi F-2700 Fluorescence Spectrophotometers. In each experiment, the photoredox catalyst and varying concentrations of quencher were combined in MeCN in screw-top 1.0 cm quartz cuvettes. For the emission quenching of $UO_2(NO_3)_2$ ·6H₂O, the photoredox catalyst concentration was 4×10^{-3} M and the solution was irradiated at 420 nm.



Figure S3. Stern-Volmer plots of fluorescence quenching experiments.

Fluorescence quenching studies were performed using a Hitachi F-2700 Fluorescence Spectrophotometers. In each experiment, the photoredox catalyst and varying concentrations of quencher were combined in the mixture solvent H₂O/MeCN (v/v 1:1) in screw-top 1.0 cm quartz cuvettes. For the emission quenching of UO₂(NO₃)₂·6H₂O, the photocatalyst concentration was 4×10^{-3} M and the solution was irradiated at 416 nm.



Figure S4. Stern-Volmer plots of fluorescence quenching experiments.

4. Control Experiments

Homo-Coupling Reaction of 1*H*-indole (2a)

To a stirring solution of UO₂(NO₃)₂·6H₂O (1.0 mg, 0.002 mmol) in 1.0 mL MeCN were added trifluoroacetic acid (11.4 mg, 0.1 mmol) and 1*H*-indole **2a** (12.3 mg, 0.105mmol). The solution was irradiated by blue LEDs (456 nm 40W) with cooling fan to keep the reaction temperature near 25 °C under air for 3 h. The mixture was concentrated in vacuo. Dichloromethane (2.0 mL) and water (1.0 mL) were then added. The mixture was extracted with dichloromethane (2.0 mL). The combined organic phases were purified by column chromatography on silica gel to give the desired product dimer product **2a'** (2.4 mg, 73% (brsm)). **2a'**: ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.92 (s, 2 H), 7.48–7.46 (m, 2 H), 7.33–7.31 (m, 2 H), 7.15–7.11 (m, 2 H), 7.02–7.01 (m, 2 H), 6.99–6.94 (m, 4 H), 6.64-6.61 (m, 1 H), 6.57–6.55 (m, 1 H), 4.93–4.71 (m, 1 H), 3.43 (d, *J* = 7.4 Hz, 2 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 136.72, 130.49, 127.07, 126.99, 122.00, 121.95, 119.80, 119.78, 119.29, 115.85, 111.17, 37.23, 34.63; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₄H₂₀N₃⁺ 350.1652, found 350.1657.



Scheme S3. Homo-coupling reaction of 1*H*-indole.

Homo-Coupling Reaction of 1,2-Dimethyl-1*H*-indole (2d)

To a stirring solution of UO₂(NO₃)₂·6H₂O (2.0 mg, 0.004 mmol) in 1.0 mL H₂O was added 1,2dimethyl-1*H*-indole **2d** (7.3 mg, 0.05 mmol). The solution was irradiated by blue LEDs (456 nm 40W) with cooling fan to keep the reaction temperature near 25 °C for 18 h. The mixture was extracted with EtOAc (3 × 2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography on silica gel to give the desired product dimer product **2d'** (2.2 mg, 43% (brsm)). **2d'**: ¹H NMR (500 MHz, Dimethyl sulfoxide-*d*₆): δ = 7.60–7.57 (m, 1 H), 7.52–7.48 (m, 1 H), 7.42–7.36 (m, 1 H), 7.18 (d, *J* = 8.1 Hz, 1 H), 7.06–7.02 (m, 1 H), 6.99 (d, *J* = 8.4 Hz, 1 H), 6.87–6.83 (m, 1 H), 6.76–6.73 (m, 1 H), 3.65 (s, 3 H), 2.81 (s, 3 H), 2.29 (s, 3 H), 1.75 (s, 3 H); ¹³C NMR (126 MHz, Dimethyl sulfoxide-*d*₆): δ = 202.6, 158.7, 137.9, 136.2, 135.9, 126.5, 124.4, 120.3, 119.1, 118.8, 117.5, 116.6, 109.5, 108.8, 106.0, 70.4, 29.3, 27.6, 21.6, 11.4; HRMS (*m/z*): [M + H]⁺ calcd for C₂₀H₂₀N₂O⁺ 305.1648, found 305.1656.



Scheme S4. Homo-coupling reaction of 1,2-dimethyl-1*H*-indole.

Homo-Coupling Reaction of *N*-Phenyl-tetrahydroisoquinoline (4a)

To a stirring solution of UO₂(NO₃)₂·6H₂O (2.0 mg, 0.004 mmol) in 1.0 mL H₂O was added *N*-phenyltetrahydroisoquinoline **4a** (20.9 mg, 0.1 mmol). The solution was irradiated by blue LEDs (456 nm) with cooling fan to keep the reaction temperature near 25 °C for 18 h. The mixture was extracted with EtOAc (3 × 2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography on silica gel to give the desired product dimer product **4a'** (8.6 mg, 66% (brsm)). **4a'**: ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.30–7.28 (m, 1 H), 7.22–7.15 (m, 5 H), 7.10–7.02 (m, 4 H), 6.98–6.96 (m, 3 H), 6.88–6.84 (m, 3 H), 6.78–6.71 (m, 2 H), 5.33 (d, *J* = 13.5 Hz, 2 H), 3.58–3.19 (m, 4 H), 2.91–2.50 (m, 4 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 149.76, 149.65, 136.68, 136.16, 135.80, 129.57, 129.42, 129.40, 128.30, 127.82, 127.77, 127.28, 127.13, 125.90, 125.52, 117.40, 117.24, 113.98, 113.02, 64.55, 63.28, 44.37, 43.13, 27.45, 27.30; HRMS (*m*/*z*): [M + H]⁺ calcd for C₃₀H₂₉N₂⁺ 417.2325, found 417.2326.



Scheme S5. Homo-coupling reaction of N-phenyltetrahydroisoquinoline.

Reaction of N-Phenyl-tetrahydroisoquinoline (4a) with excess O₂

To a stirring solution of UO₂(NO₃)₂·6H₂O (2.0 mg, 0.004 mmol) in 1.0 mL acetone was added *N*-phenyl-tetrahydroisoquinoline **4a** (20.9 mg, 0.1 mmol). The solution was irradiated by blue LEDs (456 nm) with cooling fan to keep the reaction temperature near 25 °C for 18 h in an atmosphere of O₂. The mixture was extracted with EtOAc (3 × 2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography on silica gel to give the mixed products **4ab** and **4ab'** (9.1 mg, 41%). **4ab**: ¹H NMR (500 MHz, Chloroform-*d*): δ = 8.17 (d, J = 7.8 Hz,1H), 7.46-7.48 (m, 1H), 7.37-7.46 (m, 5H), 7.24-7.27 (m, 2H), 3.99-4.01(m, 2H), 3.14-3.16 (m, 2H) ; ¹³C NMR (126 MHz, Chloroform-*d*): δ = 164.3, 143.3, 138.4, 132.2, 129.9, 129.0, 128.9, 127.3, 127.1, 126.4, 125.5, 49.5, 28.8; HRMS (*m*/z): [M + H]⁺ calcd for C₁₅H₁₄NO⁺ 224.1070, found 224.1076. **4ab'**: ¹H NMR (500 MHz, Chloroform-*d*): δ = 8.49-8.47 (m, 1H), 7.70-7.67 (m, 1H), 7.57-7.54 (m, 1H), 7.54-7.48 (m, 3H), 7.46-7.39 (m, 3H), 7.19 (d, J = 7.4 Hz, 1H), 6.57 (d, J = 7.4 Hz, 1H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 162.1, 141.4, 137.1, 132.6, 132.2, 129.3, 128.3, 128.1, 127.2, 126.9, 126.6, 126.0, 106.2; HRMS (*m*/z): [M + Na]⁺ calcd for C₁₅H₁₄NO⁺ 244.0733, found 244.0736.



Scheme S6. Reaction of N-Phenyl-tetrahydroisoquinoline with excess O2

II Experimental Procedures and Spectroscopic Data of Compounds

1. General Information

Acetonitrile was purchased in anhydrous form and used without further purification. Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Solvents for chromatography were used as supplied by Adamas-beta[®]. Reactions were monitored by thin layer chromatography (TLC) carried out on Millipore Sigma glass TLC plates (silica gel 60 coated with F₂₅₄, 250 μ m) using UV light for visualization and aqueous ammonium cerium nitrate/ammonium molybdate or basic aqueous potassium permanganate as developing agent. SiliaFlash[®] P60 silica gel (particle size: 40–63 μ m, pore size: 60 Å) was used for flash column chromatography. NMR spectra were recorded on a Bruker Avance 500 MHz NMR spectrometer. The spectra were calibrated by using residual undeuterated solvents (for ¹H NMR) and deuterated solvents (for ¹³C NMR) as internal references: undeuterated chloroform ($\delta_{\rm H} = 7.26$ ppm) and CDCl₃ ($\delta_{\rm C} = 77.16$ ppm); undeuterated DMSO ($\delta_{\rm H} = 2.50$ ppm) and DMSO-d₆ ($\delta_C = 39.52$ ppm); undeuterated methanol ($\delta_H = 3.31$ ppm) and methanol-d₄ ($\delta_C =$ 49.00 ppm). The following abbreviations are used to designate multiplicities: s = singlet, d = doublet, t = doublettriplet, q = quartet, m = multiplet, quint = quintet, br = broad. Melting points (m.p.) are uncorrected and were recorded on an SGW X-4 apparatus. High-resolution mass spectra (HRMS) were recorded on an Agilent MSD-Trap-XCT or Q-Tof micro mass spectrometer. Ultraviolet-visible absorption experiments were performed using a Hitachi U-2910 spectrophotometer. Fluorescence quenching studies were performed using a Hitachi F-2700 Fluorescence Spectrophotometers.

2. Typical Procedure for the Preparation of 4-Methoxyanilines

The substrates **1a-1z** were synthesized according to literature procedures.¹ A typical procedure is described as following for the synthesis of **1a**: To a solution of the aromatic amine (1.5 equiv) and the benzyl bromine (1.0 equiv) in CH₃CN (30 mL), was added K₂CO₃ (2.0 equiv). The suspension was stirred for 24 h at 24 °C. After the reaction was complete, the reaction mixture was filtered by Celite and washed with EtOAc. The combined organic layers were concentrated under vacuum. The residue was purified

directly by flash silica-gel chromatography to provide the desired product **1a** with 56% yield. Compounds **1a–1z** were also synthesized according to the similar method.

3. Typical Procedure for the Preparation of N-Aryl-tetrahydroisoquinolines

The substrates **4a-4j** were synthesized according to literature procedures.² A typical procedure is described as following for the synthesis of **4a**: Copper (I) iodide (0.2 g, 1.0 mmol) and potassium phosphate (4.3 g, 20.0 mmol) were added to a Schlenk tube. The tube was evacuated and back filled with nitrogen. 2-propanol (10.0 mL), ethylene glycol (1.1 mL, 20.0 mmol), 1,2,3,4-tetrahydroisoquinoline (2.0 g, 15.0 mmol) and iodobenzene (1.1 mL, 10.0 mmol) were added successively via a micro-syringe at room temperature. The reaction mixture was heated at 85–90 °C for 24 h and then allowed to cool to room temperature. Diethyl ether (20.0 mL) and water (20.0 mL) were then added. The aqueous layer was extracted by diethyl ether (3×2.0 mL). The combined organic phases was washed with brine and dried over magnesium sulfate. The solvent was removed via rotary evaporation, and the residue was subjected to a short plug of silica gel for purification using EtOAc/ Hexane (1:30 \rightarrow 1:20) as eluent to give the desired product **4a** with 93% yield. Compounds **4b–4j** were also synthesized according to the similar method.

4. General Procedures



To a stirring solution of UO₂(NO₃)₂·6H₂O (1.0 mg, 0.002 mmol) in 1.0 mL MeCN were added trifluoroacetic acid (11.4 mg, 0.1 mmol), 4-methoxyanilines (0.05 mmol) and indole derivatives (0.1 mmol). The solution was irradiated by blue LEDs (456 nm 40 W) with cooling fan to keep the reaction temperature near 25 °C under air for 3 h. The mixture was concentrated in vacuo. Dichloromethane (2.0 mL) and water (1.0 mL) were then added. The mixture was extracted with dichloromethane (2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The water phase was

recovered. The crude product was purified by column chromatography on silica gel to give the desired products.



To a stirring solution of UO₂(NO₃)₂·6H₂O (2.0 mg, 0.004 mmol) in 1.0 mL H₂O were added *N*-aryltetrahydroisoquinoline derivatives (0.1 mmol, **4b**, **4c** need to be dispersed into water by 50 μ L MeCN) and indole derivatives (0.05 mmol). The solution was irradiated by blue LEDs (456 nm 40 W) with cooling fan to keep the reaction temperature near 25 °C for 18 h. The mixture was extracted with dichloromethane (3 × 2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The water phase was recovered. The crude product was purified by column chromatography on silica gel to give the desired products.

5. Recovery of the Catalyst and Recyclability Test

For the cross-coupling reaction of secondary anilines and indoles:

To a stirring solution of $UO_2(NO_3)_2$ ·6H₂O (1.0 mg, 0.002 mmol) in 1.0 mL MeCN were added trifluoroacetic acid (11.4 mg, 0.1 mmol), ethyl (4-methoxyphenyl)glycinate **1a** (10.5 mg, 0.05 mmol) and 1*H*-indole **2a** (12.3 mg, 0.105 mmol). The solution was irradiated by blue LEDs (456 nm 40W) with cooling fan to keep the reaction temperature near 25 °C under air for 3 h. The mixture was concentrated in vacuo. Dichloromethane (2.0 mL) and water (1.0 mL) were then added. The mixture was extracted with dichloromethane (2.0 mL). The combined organic phases were purified by column chromatography on silica gel to give the desired product dimer product **3a**. The recovered water phase was dried by lyophilization then the residue used for the next catalytic cycle. The catalytic activity diminished after 4th cycle, the yield was still more than 75% (Figure 3).

For the cross-coupling reaction of tetrahydroisoquinolines and indoles:

To a stirring solution of UO₂(NO₃)₂·6H₂O (2.0 mg, 0.004 mmol) in 1.0 mL H₂O were added *N*-phenyltetrahydroisoquinoline **4a** (20.9 mg, 0.1 mmol) and 1,2-dimethyl-1*H*-indole **2d** (7.3 mg, 0.05mmol). The solution was irradiated by blue LEDs (456 nm) with cooling fan to keep the reaction temperature near 25 °C for 18 h. The mixture was extracted with dichloromethane (3×2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography on silica gel to give the desired product dimer product **5a**. The recovered water phase was used for the next catalytic cycle. The catalytic activity diminished after 4th cycle, the yield was still more than 73% (Figure 3).

6. Photocatalyzed "Window Ledge" Reactions

To a solution of $UO_2(NO_3)_2$ ·6H₂O (1.0 mg, 0.002 mmol) in 1.0 mL MeCN were added trifluoroacetic acid (11.4 mg, 0.1 mmol), ethyl (4-methoxyphenyl)glycinate (10.5 mg, 0.05 mmol) and 1*H*-indole (11.7 mg, 0.1 mmol). The solution was irradiated by sunlight under air for 24 h. The mixture was concentrated in vacuo. Dichloromethane (2.0 mL) and water (1.0 mL) were then added. The mixture was extracted with dichloromethane (3 × 2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The water phase was recovered. The crude product was purified by column chromatography on silica gel to give the desired products **3a** with 95% yield.





Scheme S7. The "Window Ledge" reaction of 3a.

To a solution of $UO_2(NO_3)_2 \cdot 6H_2O$ (2.0 mg, 0.004 mmol) in 1.0 mL H₂O were added 2-phenyl-1,2,3,4tetrahydroisoquinoline (11.4 mg, 0.1 mmol) and 1,2-dimethyl-1*H*-indole (7.3 mg, 0.05 mmol). The solution was irradiated by sunlight under air for 24 h. The mixture was extracted with dichloromethane (3 × 2.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The water phase was recovered. The crude product was purified by column chromatography on silica gel to give the desired products **5a** with 94% yield.





Scheme S8. The "Window Ledge" reaction of 5a.

7. Gram-scale Synthesis of 30 and 5a.

To a stirring solution of UO₂(NO₃)₂·6H₂O (36.4 mg, 0.073 mmol) in 37 mL MeCN were added trifluoroacetic acid (415.1 mg, 3.64 mmol), cholesteryl ester **10** (1g, 1.82 mmol) and 1*H*-indole **2a** (446.9 mg, 3.82 mmol). The solution was irradiated by blue LEDs (456 nm 40W) with cooling fan to keep the reaction temperature near 25 °C under air for 3 h. The mixture was concentrated in vacuo. Dichloromethane (70.0 mL) and water (35.0 mL) were then added. The mixture was extracted with dichloromethane (3 × 70.0 mL). The water phase was recovered. The combined organic phases were purified by column chromatography on silica gel to give the desired product dimer product **30** with 88% yield.



Figure S5. Gram-scale synthesis of 30.

To a stirring solution of $UO_2(NO_3)_2 \cdot 6H_2O$ (120.0 mg, 0.24 mmol) in 60.0 mL H₂O were added *N*-phenyl-tetrahydroisoquinoline **4a** (1.25 g, 6.0 mmol) and 1,2-dimethyl-1*H*-indole **2d** (438 mg, 3.0 mmol). The solution was irradiated by blue LEDs (456 nm 40W) with cooling fan to keep the reaction temperature near 25 °C for 36 h. The mixture was extracted with dichloromethane (3 × 100.0 mL). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuo. The water phase was recovered. The crude product was purified by column chromatography on silica gel to give the desired product dimer product **5a** with 73% yield.



Figure S6. Gram-scale synthesis of 5a.

8. Spectroscopic Data of Compounds

Ethyl 2,2-di(1*H*-indol-3-yl)acetate (3a)



Following the general procedure, was obtained as a red powder (15.3 mg, 96%), m.p. 57 \sim 59 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 8.02 (s, 2 H), 7.65 (d, *J* = 8.0 Hz, 2 H), 7.34 (d, *J* = 8.1 Hz, 2 H), 7.19 (t, *J* = 7.1 Hz, 2 H), 7.12–7.05 (m, 4 H), 5.51 (s, 1 H), 4.23 (q, *J* = 7.1 Hz, 2 H), 1.27 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 173.6, 136.5, 126.8, 123.5, 122.3, 119.7, 119.5, 113.9, 111.3, 61.2, 40.8, 14.4; HRMS (*m*/*z*): [M + Na]⁺ calcd for C₂₀H₁₈N₂NaO₂⁺ 341.1260, found 341.1275.



Following the general procedure, was obtained as a red powder (18.5 mg, 98%), m.p. $131 \sim 133$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.02$ (s, 2 H), 7.65 (d, J = 8.0 Hz, 2 H), 7.34 (d, J = 8.1 Hz, 2 H), 7.19 (t, J = 7.1 Hz, 2 H), 7.12–7.05 (m, 2 H), 5.51 (s, 1 H), 4.23 (q, J = 7.1 Hz, 2 H), 3.95 (s, 6 H),1.27 (t, J = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 173.5$, 146.2, 128.2, 127.0, 123.0, 120.1, 114.4, 112.3, 102.1, 61.1, 55.4, 41.1, 14.4; HRMS (m/z): [M + H]⁺ calcd for C₂₅H₂₃N₂O₄⁺ 379.1640, found 379.1653.



Following the general procedure, was obtained as a red powder (16.9 mg, 98%), m.p. $128 \sim 130$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.88$ (s, 2 H), 7.51 (d, J = 8.1 Hz, 2 H), 7.11 (s, 2 H), 6.99 (d, J = 2.3 Hz, 2 H), 6.93 (d, J = 9.1 Hz, 2 H), 5.45 (s, 1 H), 4.21 (q, J = 7.1 Hz, 2 H), 2.45 (s, 6 H), 1.26 (t, J = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 173.6$, 136.9, 132.0, 124.7, 122.8, 121.4, 119.2, 113.7, 111.3, 61.2, 40.9, 21.8, 14.4; HRMS (m/z): [M + H]⁺ calcd for C₂₂H₂₃N₂O₂⁺ 347.1754, found 347.1739.



Following the general procedure, was obtained as a red powder (18.2 mg, 97%), m.p. 144 \sim 146 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.42 (d, *J* = 8.0 Hz, 2 H), 7.25 (d, *J* = 8.2 Hz, 2 H), 7.14–7.11 (m, 2 H), 7.10–6.97 (m, 2 H), 5.53 (s, 1 H), 4.27 (q, *J* = 7.1 Hz, 2 H), 3.64 (s, 6 H), 2.26 (s, 6 H), 1.25 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 174.0, 136.6, 134.2, 127.4, 120.5, 119.1, 118.9, 108.7, 108.5, 61.1, 40.7, 29.6, 14.4, 10.9; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₄H₂₇N₂O₂⁺ 375.2067, found 375.2050.



Following the general procedure, was obtained as a red powder (16.6 mg, 94%), m.p. 65 \sim 67 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.11$ (s, 2 H), 7.50 (dd, J = 8.7, 5.3 Hz, 2 H), 7.03 (d, J = 2.4 Hz, 2 H), 6.99 (dd, J = 9.5, 2.2 Hz, 2 H), 6.91–6.80 (m, 2 H), 5.42 (s, 1 H), 4.23 (q, J = 7.1 Hz, 2 H), 1.26 (t, J = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 173.4$, 160.1 (d, J = 238.0 Hz), 136.4 (d, J = 12.4 Hz), 123.7 (d, J = 3.5 Hz), 123.3, 120.2 (d, J = 10.1 Hz), 113.7, 108.6 (d, J = 24.4 Hz), 97.7 (d, J = 26.1 Hz), 61.4, 40.9, 14.3; HRMS (m/z): [M + H]⁺ calcd for C₂₀H₁₇F₂N₂O₂⁺ 355.1253, found 355.1293.



Following the general procedure, was obtained as a red powder (18.4 mg, 95%), m.p. 68 \sim 70 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 8.13 (s, 2 H), 7.48 (d, *J* = 8.5 Hz, 2 H), 7.29 (d, *J* = 1.5 Hz, 2 H), 7.04 (dd, *J* = 8.7, 1.9 Hz, 2 H), 7.01 (d, *J* = 1.9 Hz, 2 H), 5.41 (s, 1 H), 4.23 (q, *J* = 7.1 Hz, 2 H), 1.26 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 173.3, 136.8, 128.3, 125.2, 124.1, 120.5, 120.3, 113.6, 111.4, 61.5, 40.7, 29.8, 14.4; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₀H₁₇Cl₂N₂O₂⁺ 387.0662, found 387.0667.



Following the general procedure, was obtained as a red powder (17.6 mg, 74%), m.p. 63 \sim 65 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 8.24 (s, 2 H), 7.23 (dd, *J* = 7.6, 4.9 Hz, 4 H), 6.97 (t, *J* = 7.9 Hz, 2 H), 6.82 (s, 1 H), 6.76 (d, *J* = 2.3 Hz, 2 H), 4.27 (q, *J* = 7.1 Hz, 2 H), 1.29 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 174.9, 138.0, 125.3, 124.7, 124.3, 123.1, 115.7, 114.3, 110.8, 61.4, 40.9, 14.4; HRMS (*m*/*z*): [M + Na]⁺ calcd for C₂₀H₁₆Br₂N₂NaO₂⁺ 496.9471, found 496.9475.



Following the general procedure, was obtained as a red powder (26.8 mg, 94%), m.p. 74 \sim 76 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.16$ (s, 2 H), 7.95 (s, 2 H), 7.42 (d, J = 10.1 Hz, 2 H), 7.08 (d, J = 8.5 Hz, 2 H), 6.98 (s, 2 H), 5.34 (s, 1 H), 4.24 (q, J = 7.1 Hz, 2 H), 1.30 (t, J = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 173.2$, 135.6, 130.7, 129.1, 128.3, 124.3, 113.5, 112.7, 83.3, 61.6, 40.6, 14.4; HRMS (m/z): [M + Na]⁺ calcd for C₂₀H₁₆I₂N₂NaO₂⁺ 592.9193, found 592.9209.



Following the general procedure, was obtained as a red powder (12.8 mg, 63%), m.p. 95~97 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.76$ (s, 2 H), 8.34 (s, 2 H), 7.98 (d, J = 9.3 Hz, 2 H), 7.62 (d, J = 8.9 Hz, 2 H), 7.45 (s, 2 H), 5.51 (s, 1 H), 4.26 (q, J = 7.1 Hz, 2 H), 1.28 (t, J = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 172.4$, 143.8, 135.0, 131.1, 129.1, 119.4, 115.5, 114.3, 108.5, 61.9, 40.5, 14.4; HRMS (*m/z*): [M + Na]⁺ calcd for C₂₀H₁₆N₄NaO₆⁺ 431.0962, found 431.0968.



Following the general procedure, was obtained as a red powder (9.8 mg, 53%), m.p. $133 \sim 135$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.70$ (s, 2 H), 7.92 (s, 2 H), 7.42–7.41 (m, 4 H), 7.33 (d, J = 2.3 Hz, 2 H), 5.44 (s, 1 H), 4.25 (q, J = 7.1 Hz, 2 H), 1.28 (t, J = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 172.31$, 138.31, 126.36, 125.54, 125.40, 125.39, 120.78, 113.91, 112.52, 103.01, 61.82, 40.85, 29.83, 14.33; HRMS (m/z): [M + Na]⁺ calcd for C₂₂H₁₆N₄NaO₂⁺ 391.1165, found 391.1175.



Following the general procedure, was obtained as a red powder (20.4 mg, 94%), m.p. 208~210 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.62$ (s, 2 H), 8.41 (s, 2 H), 7.86 (dd, J = 8.6, 1.6 Hz, 2 H), 7.29 (d, J = 8.6 Hz, 2 H), 7.11 (d, J = 1.8 Hz, 2 H), 5.55 (s, 1 H), 4.24 (q, J = 7.1 Hz, 2 H), 3.88 (s, 6 H), 1.29 (t, J = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 173.4$, 168.4, 139.2, 126.3, 124.9, 123.7, 122.4, 121.8, 114.7, 111.2, 61.6, 52.0, 40.6, 14.3; HRMS (m/z): [M + Na]⁺ calcd for C₂₄H₂₂N₂NaO₆⁺ 457.1370, found 457.1392.



Following the general procedure, was obtained as a red powder (16.1 mg, 93%), m.p. 69 \sim 71 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 8.02 (s, 2 H), 7.64 (d, *J* = 7.9 Hz, 2 H), 7.33 (d, *J* = 8.1 Hz, 2 H), 7.18 (t, *J* = 7.1 Hz, 2 H), 7.09 (t, *J* = 7.9 Hz, 2 H), 7.04 (d, *J* = 1.9 Hz, 2 H), 5.41 (s, 1 H), 1.46 (s, 9 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 172.9, 136.5, 126.9, 123.3, 122.1, 119.6, 119.5, 114.3, 111.3, 81.2, 41.8, 28.2; HRMS (*m*/*z*): [M + Na]⁺ calcd for C₂₂H₂₂N₂NaO₂⁺ 369.1573, found 369.1576.



Following the general procedure, was obtained as a colorless crystal (17.4 mg, 92%); ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.02$ (s, 2 H), 7.59 (d, J = 7.9 Hz, 2 H), 7.34 (d, J = 8.2 Hz, 2 H), 7.32–7.25 (m, 5 H), 7.18 (t, J = 7.6 Hz, 2 H), 7.11–6.98 (m, 4 H), 5.57 (s, 1 H), 5.20 (s, 2 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 173.3$, 136.5, 136.0, 128.6, 128.4, 128.3, 126.8, 123.5, 122.3, 119.8, 119.5, 113.7, 111.3, 67.0, 40.8; HRMS (*m*/*z*): [M + Na]⁺ calcd for C₂₅H₂₀N₂NaO₂⁺ 403.1417, found 403.1416; CCDC 2220117 contains the supplementary crystallographic data of **3m** [m.p.: 167–168 °C (Hexane/EtOAc, 3:1)]. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Following the general procedure, was obtained as a red powder (23.3 mg, 93%), m.p. 86~88 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.16$ (s, 2 H), 7.62 (dd, J = 7.8, 5.2 Hz, 2 H), 7.32 (d, J = 8.1 Hz, 2 H), 7.17 (t, J = 7.6 Hz, 2 H), 7.12–7.02 (m, 4 H), 5.50 (s, 1 H), 4.73 (q, J = 6.2 Hz, 1 H), 4.37 (s, 1 H), 4.19–4.10 (m, 2 H), 4.00 (s, 1 H), 2.74 (t, J = 12.7 Hz, 1 H), 2.05 (s, 1 H), 1.92–1.83 (m, 1 H), 1.77–1.68 (m, 1 H), 1.61–1.37 (m, 5 H), 1.17 (d, J = 6.2 Hz, 3 H), 0.85 (t, J = 7.4 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 173.59$, 155.73, 155.71, 136.49, 136.47, 126.80, 126.78, 123.60, 122.19, 119.64, 119.39, 119.37, 113.69, 113.59, 111.37, 111.36, 73.23, 73.18, 62.99, 48.19, 40.74, 29.19, 29.16, 19.91, 19.86, 19.05, 9.85, 9.81; HRMS (m/z): [M + Na]⁺ calcd for C₃₀H₃₅N₃NaO4⁺ 524.2520, found 524.2532.

(3S,8S,9S,10R,13R,14S,17R)-10,13-Dimethyl-17-((R)-6-methylheptan-2-yl)-

2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthren-3-yl 2,2-di(1H-

indol-3-yl)acetate (30)



Following the general procedure, was obtained as a white powder (29.9 mg, 91%), m.p. 202 \sim 204 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 8.01 (s, 2 H), 7.65 (d, *J* = 7.9 Hz, 2 H), 7.32 (d, *J* = 8.1 Hz, 2 H), 7.18 (t, *J* = 7.6 Hz, 2 H), 7.13–7.06 (m, 2 H), 7.06–7.01 (m, 2 H), 5.48 (s, 1 H), 5.33 (dd, *J* = 5.0, 2.0 Hz, 1 H), 4.75–4.68 (m, 1 H), 2.34 (d, *J* = 8.1 Hz, 2 H), 1.98 (ddt, *J* = 32.3, 17.3, 4.4 Hz, 2 H), 1.84 (ddt, *J* = 16.9, 9.7, 3.5 Hz, 3 H), 1.65–1.22 (m, 13 H), 1.18–1.03 (m, 8 H), 1.00 (s, 3 H), 0.92 (d, *J* = 6.5 Hz, 3 H), 0.88 (dd, *J* = 6.6, 2.3 Hz, 6 H), 0.68 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 173.02, 139.77, 136.48, 126.84, 123.48, 122.79, 122.20, 119.61, 119.55, 113.91, 111.34, 74.91, 56.81, 56.27, 50.12, 42.44, 41.05, 39.86, 39.66, 38.16, 37.09, 36.72, 36.32, 35.93, 32.03, 31.98, 28.37, 28.15, 27.87, 24.42, 23.97, 22.96, 22.71, 21.15, 19.46, 18.85, 11.99; HRMS (*m*/*z*): [M + Na]⁺ calcd for C₄₅H₅₈N₂NaO₂⁺ 681.4390, found 681.4401.



Following the general procedure, was obtained as a red powder (11.2 mg, 71%), m.p. $163 \sim 165 \,^{\circ}$ C; ¹H NMR (500 MHz, Dimethyl sulfoxide- d_6) : $\delta = 11.05$ (s, 2 H), 7.36 (t, J = 7.9 Hz, 4 H), 7.30 (d, J = 2.6 Hz, 2 H), 7.02 (t, J = 8.1 Hz, 2 H), 6.84 (t, J = 7.0 Hz, 2 H), 4.44 (t, J = 6.6 Hz, 2 H), 3.07 (t, J = 6.7 Hz, 2 H); ¹³C NMR (126 MHz, Dimethyl sulfoxide- d_6): $\delta = 177.1$, 137.0, 125.4, 123.4, 121.0, 120.1, 118.3, 113.6, 111.6, 65.8, 45.9, 36.3; HRMS (m/z): [M + Na]⁺ calcd for C₂₀H₁₆N₂NaO₂⁺ 339.1104, found 339.1118.



Following the general procedure, was obtained as a red powder (8.7 mg, 54%), m.p. $124 \sim 126$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.92$ (s, 2 H), 7.39 (d, J = 8.0 Hz, 2 H), 7.37–7.34 (m, 4 H), 7.28 (t, J = 7.6 Hz, 2 H), 7.19 (dt, J = 20.5, 7.2 Hz, 3 H), 7.00 (t, J = 7.5 Hz, 2 H), 6.67 (dd, J = 2.5, 1.0 Hz, 2 H), 5.89 (s, 1 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 144.1$, 136.8, 128.9, 128.4, 127.2, 126.3, 123.7, 122.1, 120.1, 119.9, 119.4, 111.1, 40.3; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₃H₁₉N₂⁺ 323.1543, found 323.1563.



Following the general procedure, was obtained as a red powder (10.3 mg, 59%), m.p. 187 \sim 189 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.90 (s, 2 H), 7.39 (d, *J* = 7.9 Hz, 2 H), 7.35 (d, *J* = 8.2 Hz, 2 H), 7.29–7.22 (m, 2 H), 7.17 (t, *J* = 7.6 Hz, 2 H), 7.00 (t, *J* = 7.9 Hz, 2 H), 6.82 (d, *J* = 8.7 Hz, 2 H), 6.67–6.63 (m, 2 H), 5.84 (s, 1 H), 3.78 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 158.0, 136.9, 136.4, 129.7, 127.2, 123.6, 122.0, 120.2, 120.1, 119.3, 113.7, 111.1, 55.4, 39.5; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₄H₂₁N₂O⁺ 353.1649, found 353.1654.



Following the general procedure, was obtained as a red powder (11.9 mg, 63%), m.p. 126 \sim 128 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.92 (s, 2 H), 7.41 (d, *J* = 8.0 Hz, 2 H), 7.35 (d, *J* = 8.2 Hz, 2 H), 7.27–7.27 (m, 3 H), 7.16 (t, *J* = 8.1 Hz, 2 H), 7.00 (t, *J* = 7.9 Hz, 2 H), 6.70 (d, *J* = 3.0 Hz, 2 H), 5.86 (s, 1 H), 1.29 (s, 9 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 148.86, 140.95, 136.84, 128.36, 127.31, 125.18, 123.66, 122.00, 120.18, 120.15, 119.28, 111.10, 39.79, 34.51, 31.59; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₇H₂₇N₂⁺ 379.2169, found 379.2154.


Following the general procedure, was obtained as a red powder (14.4 mg, 66%), m.p. 188 \sim 190 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.82 (s, 2 H), 7.28 (d, *J* = 3.4 Hz, 4 H), 7.25–7.23 (m, 2 H), 6.84–6.78 (m, 4 H), 6.72–6.69 (m, 2 H), 5.74 (s, 1 H), 3.68 (s, 6 H), 1.30 (s, 9 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 153.8, 148.9, 140.9, 132.0, 128.4, 127.7, 125.2, 124.5, 119.7, 112.0, 111.8, 102.2, 56.0, 39.9, 34.5, 31.6; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₉H₃₁N₂O₂⁺ 439.2380, found 439.2380.

3,3'-((4-(Trifluoromethyl)phenyl)methylene)bis(1*H*-indole) (3u)



Following the general procedure, was obtained as a red powder (8.0 mg, 41%), m.p. $135 \sim 137$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.97$ (s, 2 H), 7.53 (d, J = 8.2 Hz, 2 H), 7.46 (d, J = 8.1 Hz, 2 H), 7.37 (t, J = 7.1 Hz, 4 H), 7.19 (t, J = 8.1 Hz, 2 H), 7.02 (t, J = 7.5 Hz, 2 H), 6.66 (d, J = 1.5 Hz, 2 H), 5.95 (s, 1 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 148.3$, 136.8, 129.2, 128.7, 128.5, 127.0, 125.4 (q, J = 3.6 Hz), 123.8, 122.3, 119.9, 119.6, 118.9, 111.3, 40.2; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₄H₁₈F₃N₂⁺ 391.1417, found 391.1428.



Following the general procedure, was obtained as a red powder (13.8 mg, 74%), m.p. 125 \sim 127 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.75$ (d, J = 4.5 Hz, 1 H), 8.16 (t, J = 8.1 Hz, 2 H), 8.06 (s, 2 H), 7.68 (t, J = 8.3 Hz, 1 H), 7.49–7.42 (m, 1 H), 7.38 (t, J = 7.3 Hz, 4 H), 7.24–7.14 (m, 3 H), 7.03 (t, J = 7.9 Hz, 2 H), 6.66 (s, 1 H), 6.59 (d, J = 1.8 Hz, 2 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 150.4$, 136.9, 130.0, 129.2, 127.5, 126.9, 126.7, 124.4, 124.3, 122.4, 121.1, 119.7, 119.6, 117.8, 111.4, 35.7; HRMS (m/z): [M + H]⁺ calcd for C₂₆H₂₀N₃⁺ 374.1652, found 374.1658.



Following the general procedure, was obtained as a red powder (11.7 mg, 65%), m.p. $168 \sim 170$ °C; ¹H NMR (500 MHz, Acetone-*d*₆): $\delta = 9.92$ (s, 3 H), 7.47 (d, J = 7.9 Hz, 3 H), 7.37 (d, J = 8.1 Hz, 3 H), 7.05-6.87 (m, 3 H), 6.93 (d, J = 1.3 Hz, 3 H), 6.88 (m, 3 H), 6.18 (s, 1 H); ¹³C NMR (126 MHz, Acetone-*d*₆): $\delta = 138.1$, 128.3, 124.2, 121.8, 120.4, 120.0, 119.1, 112.1, 32.2; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₅H₂₀N₃⁺ 362.1652, found 362.1665.



Following the general procedure, was obtained as a red powder (2.2 mg, 17%), m.p. 89 \sim 91 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.91 (s, 2 H), 7.57 (d, *J* = 7.9 Hz, 2 H), 7.35 (d, *J* = 8.1 Hz, 2 H), 7.16 (t, *J* = 8.1 Hz, 2 H), 7.04 (t, *J* = 7.5 Hz, 2 H), 6.94 (d, *J* = 2.2 Hz, 2 H), 4.68 (q, *J* = 7.1 Hz, 1 H), 1.81 (d, *J* = 7.1 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 136.81, 127.07, 121.93, 121.88, 121.31, 119.89, 119.17, 111.18, 28.33, 21.87; HRMS (*m*/*z*): [M + Na]⁺ calcd for C₁₈H₁₆N₂Na⁺ 283.1205, found 283.1211.



Following the general procedure, was obtained as a red powder (4.6 mg, 32%), m.p. $95 \sim 97$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.91$ (s, 2 H), 7.60 (d, J = 7.9 Hz, 2 H), 7.34 (d, J = 8.1 Hz, 2 H), 7.14 (t, J = 7.6 Hz, 2 H), 7.06–6.95 (m, 4 H), 4.50 (t, J = 7.5 Hz, 1 H), 2.20 (q, J = 7.5 Hz, 2 H), 1.48–1.38 (m, 2 H), 0.95 (t, J = 7.4 Hz, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 136.7$, 127.4, 121.9, 121.5, 120.8, 119.8, 119.1, 111.2, 77.4, 77.2, 76.9, 38.2, 33.8, 21.6, 14.4; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₀H₂₁N₂⁺ 289.1699, found 289.1716.



Following the general procedure, was obtained as a red powder (2.4 mg, 15%), m.p. 96~98 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.00$ (s, 2 H), 7.63 (d, J = 8.0 Hz, 2 H), 7.36 (d, J = 8.1 Hz, 2 H), 7.18 (t, J = 7.6 Hz, 2 H), 7.07 (t, J = 7.9 Hz, 2 H), 7.02 (d, J = 2.0 Hz, 2 H), 4.95 (d, J = 7.1 Hz, 1 H), 4.74 (d, J = 7.1 Hz, 2 H), 1.99 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 171.4$, 136.6, 127.2, 122.3, 122.2, 119.7, 119.5, 116.5, 111.3, 67.5, 33.7, 21.3; HRMS (*m*/*z*): [M + Na]⁺ calcd for C₂₀H₁₈N₂NaO₂⁺ 341.1260, found 341.1265.

1-(1,2-Dimethyl-1*H*-indol-3-yl)-2-phenyl-1,2,3,4-tetrahydroisoquinoline (5a)



Following the general procedure, was obtained as a colorless crystal (16.8 mg, 96%), m.p. $113 \sim 115$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.22-7.18$ (m, 5 H), 7.10–7.03 (m, 5 H), 6.89–6.83 (m, 3 H), 6.02 (s, 1 H), 3.74–3.61 (m, 2 H), 3.60 (s, 3 H), 3.13–2.99 (m, 2 H), 2.13 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 151.1$, 138.3, 136.6, 135.4, 135.3, 128.9, 128.8, 128.4, 127.9, 126.3, 126.2, 120.4, 120.2, 119.6, 119.2, 119.2, 112.9, 108.6, 57.2, 46.1, 29.5, 27.9, 10.7; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₅H₂₅N₂⁺ 353.2012, found 353.2015; CCDC 2124830 contains the supplementary crystallographic data of **5a** [m.p.: 113–115 °C (Hexane/EtOAc, 10:1)]. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

1-(1,2-Dimethyl-1*H*-indol-3-yl)-6-methoxy-2-phenyl-1,2,3,4-tetrahydroisoquinoline (5b)



Following the general procedure, was obtained as light yellow powder (12.8 mg, 67%), m.p. 135 \sim 137 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.22–7.14 (m, 3 H), 7.09–7.06 (m, 1 H), 7.03–6.93 (m, 4 H), 6.91–6.87 (m, 1 H), 6.84–6.81 (m, 1 H), 6.72 (d, *J* = 2.7 Hz, 1 H), 6.66 (dd, *J* = 8.5, 2.7 Hz, 1 H), 5.96 (s, 1 H), 3.80 (s, 3 H), 3.72–3.60 (m, 2 H), 3.59 (s, 3 H), 3.13–2.89 (m, 2 H), 2.12 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 157.99, 151.07, 136.65, 136.56, 135.27, 130.51, 129.42, 128.91, 127.91, 120.34, 120.15, 119.50, 119.26, 119.20, 113.20, 113.04, 112.62, 108.56, 56.68, 55.34, 45.84, 29.52, 28.08, 10.70; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₆H₂₇N₂O⁺ 383.2118, found 383.2126.

1-(1,2-Dimethyl-1*H*-indol-3-yl)-6,7-dimethoxy-2-phenyl-1,2,3,4-tetrahydroisoquinoline (5c)



Following the general procedure, was obtained as a light yellow powder (16.7 mg, 81%), m.p. 156~ 158 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.23-7.16$ (m, 3 H), 7.10–7.01 (m, 3 H), 6.92–6.78 (m, 3 H), 6.65 (s, 1 H), 6.59 (s, 1 H), 5.98 (s, 1 H), 3.89 (s, 3 H), 3.68 (s, 3 H), 3.65–3.62 (m, 2 H), 3.61 (s, 3 H), 2.99 (dt, J = 16.2, 8.0 Hz, 1 H), 2.75 (dt, J = 16.2, 3.7 Hz, 1 H), 2.10 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 150.81$, 147.81, 147.70, 136.50, 135.37, 129.75, 129.05, 128.07, 127.57, 120.39, 119.85, 119.32, 119.22, 118.98, 112.75, 111.35, 111.11, 108.53, 56.42, 56.04, 55.98, 44.92, 29.54, 26.46, 10.72, 10.72; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₇H₂₉N₂O₂⁺ 413.2224, found 413.2230.



Following the general procedure, was obtained as a light yellow powder (18.1 mg, 84%), m.p. $107 \sim 109 \,^{\circ}$ C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.35 \,(\text{s}, 1 \,\text{H}), 7.22-7.13 \,(\text{m}, 4 \,\text{H}), 7.08 \,(\text{t}, J = 7.0 \,\text{Hz}, 1 \,\text{H}), 6.99-6.82 \,(\text{m}, 6 \,\text{H}), 5.82 \,(\text{s}, 1 \,\text{H}), 3.66-3.61 \,(\text{m}, 1 \,\text{H}), 3.58 \,(\text{s}, 3 \,\text{H}), 3.57-3.53 \,(\text{m}, 1 \,\text{H}), 3.08-2.88 \,(\text{m}, 2 \,\text{H}), 2.09 \,(\text{s}, 3 \,\text{H}); {}^{13}$ C NMR (126 MHz, Chloroform-*d*): $\delta = 151.0, 137.8, 137.6, 136.6, 135.5, 131.5, 130.2, 129.3, 128.9, 127.7, 120.9, 120.5, 120.1, 120.0, 119.4, 119.1, 112.2, 108.7, 57.3, 46.2, 29.6, 28.0, 10.7; HRMS <math>(m/z)$: $[\text{M} + \text{H}]^+$ calcd for C₂₅H₂₄BrN₂⁺ 431.1117, found 431.1115.



Following the general procedure, was obtained as a light yellow powder (13.6. mg, 76%), m.p. 110 \sim 112 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.18 (dd, *J* = 15.6, 7.7 Hz, 3 H), 7.09 (t, *J* = 7.5 Hz, 1 H), 7.05–6.96 (m, 4 H), 6.93 (t, *J* = 7.4 Hz, 1 H), 6.87 (t, *J* = 7.2 Hz, 1 H), 6.63 (d, *J* = 5.1 Hz, 1 H), 5.92 (s, 1 H), 3.79–3.60 (m, 2 H), 3.58 (s, 3 H), 3.14–2.88 (m, 2 H), 2.11 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 151.1, 137.1, 136.6, 135.2, 134.2, 128.9, 127.8, 126.9, 122.4, 120.9, 120.5, 120.4, 119.3, 119.0, 111.5, 108.6, 55.6, 47.3, 29.5, 24.5, 10.5; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₃H₂₃N₂S⁺ 359.1732, found 359.1737.

1-(1,2-Dimethyl-1*H*-indol-3-yl)-2-(*p*-tolyl)-1,2,3,4-tetrahydroisoquinoline (5f)



Following the general procedure, was obtained as a white powder (14.4mg, 79%), m.p. $123 \sim 125$ °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.16$ (dt, J = 14.8, 7.6 Hz, 3 H), 7.09–6.99 (m, 3 H), 6.95 (t, J = 7.7 Hz, 3 H), 6.93–6.83 (m, 3 H), 5.89 (s, 1 H), 3.67-3.62 (m, 2 H), 3.58 (s, 3 H), 3.56–3.52 (m, 1 H), 3.13–2.83 (m, 1 H), 2.23 (s, 3 H), 2.10 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 149.03$, 138.54, 136.59, 135.42, 135.37, 130.01, 129.43, 128.72, 128.43, 127.93, 120.31, 119.26, 119.15, 113.03, 108.53, 57.64, 47.08, 29.55, 28.09, 20.71, 10.67; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₆H₂₇N₂⁺ 367.2169, found 367.2174.

2-(4-(*tert*-Butyl)phenyl)-1-(1,2-dimethyl-1*H*-indol-3-yl)-1,2,3,4-tetrahydroisoquinoline (5g)



Following the general procedure, was obtained as a light yellow powder (15.7 mg, 77%), m.p. 122 \sim 124 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.22–7.15 (m, 5 H), 7.09–7.02 (m, 3 H), 6.96–6.83 (m, 4 H), 5.94 (s, 1 H), 3.72–3.63 (m, 1 H), 3.59 (s, 3 H), 3.58–3.54 (m, 1 H), 3.15–3.03 (m, 1 H), 2.97 (dt, *J* = 16.3, 4.4 Hz, 1 H), 2.09 (s, 3 H), 1.26 (s, 9 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 148.59, 142.96, 138.53, 136.57, 135.55, 135.43, 128.76, 128.45, 127.96, 126.28, 126.15, 125.67, 120.30, 119.30, 119.22, 119.17, 113.02, 108.53, 57.57, 46.01, 34.10, 31.62, 29.53, 27.82, 10.68; HRMS (*m/z*): [M + H]⁺ calcd for C₂₉H₃₃N₂⁺ 409.2638, found 409.2637.

1-(1,2-Dimethyl-1*H*-indol-3-yl)-2-(4-methoxyphenyl)-1,2,3,4-tetrahydroisoquinoline (5h)



Following the general procedure, was obtained as a white powder (13.9 mg, 73%), m.p. 130 \sim 132 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.22–7.13 (m, 3 H), 7.11–7.04 (m, 4 H), 6.89 (dt, *J* = 14.7, 7.5 Hz, 2 H), 6.66 (dd, *J* = 8.2, 1.6 Hz, 1 H), 6.54 (s, 1 H), 6.38 (dd, *J* = 8.1, 1.9 Hz, 1 H), 6.01 (s, 1 H), 3.71–3.66 (m, 1 H), 3.65 (s, 3 H), 3.64–3.61 (m, 1 H), 3.60 (s, 3 H), 3.13–3.03 (m, 1 H), 2.96 (dt, *J* = 16.4, 4.4 Hz, 1 H), 2.14 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 160.4, 152.3, 138.2, 136.6, 135.5, 135.3, 129.6, 128.8, 128.4, 127.9, 126.4, 126.2, 120.4, 119.3, 119.1, 113.0, 111.7, 108.6, 105.2, 105.1, 57.0, 55.2, 45.8, 29.5, 27.7, 10.8; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₆H₂₇N₂O⁺ 383.2118, found 383.2118.

2-(4-Bromophenyl)-1-(1,2-dimethyl-1*H*-indol-3-yl)-1,2,3,4-tetrahydroisoquinoline (5i)



Following the general procedure, was obtained as a light yellow powder (16.8 mg, 96%), m.p. 115 \sim 117 °C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.25-7.15$ (m, 6 H), 7.11–7.01 (m, 4 H), 6.94–6.86 (m, 5 H), 5.93 (s, 1 H), 3.67–3.64 (m, 1 H), 3.60 (s, 1 H), 3.58–3.54 (m, 1 H), 3.09–3.01 (m, 1 H), 2.13 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 150.1$, 138.0, 136.6, 135.3, 135.2, 131.7, 128.7, 128.4, 127.7, 126.5, 126.3, 121.2, 120.5, 119.3, 119.0, 112.6, 112.5, 108.7, 57.2, 46.4, 29.6, 28.0, 10.8; HRMS (*m/z*): [M + H]⁺ calcd for C₂₅H₂₄BrN₂⁺ 431.1117, found 431.1112.

2-(4-Chlorophenyl)-1-(1,2-dimethyl-1*H*-indol-3-yl)-1,2,3,4-tetrahydroisoquinoline (5j)



Following the general procedure, was obtained as a light yellow powder (16.2 mg, 84%), m.p. 112 \sim 114 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.20–7.17 (m, 3 H), 7.10–7.01 (m, 5 H), 6.95–6.86 (m, 4 H), 5.90 (s, 1 H), 3.66–3.61(m, 1 H), 3.59 (s, 3 H), 3.57–3.53 (m, 1 H), 3.07–3.01 (m, 2 H), 2.11 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 149.8, 138.0, 136.6, 135.3, 135.2, 128.8, 128.7, 128.4, 127.7, 126.5, 126.3, 125.3, 121.1, 120.5, 119.3, 119.1, 112.6, 108.7, 57.5, 46.7, 29.6, 28.1, 10.8; HRMS (*m/z*): [M + H]⁺ calcd for C₂₅H₂₄ClN2⁺ 387.1623, found 387.1625.



Following the general procedure, was obtained as a white powder (5.3 mg, 33%), m.p. $167 \sim 169$ °C; ¹H NMR (500 MHz, Dimethyl sulfoxide- d_6): $\delta = 10.88$ (s, 1 H), 7.48 (d, J = 7.9 Hz, 1 H), 7.41–7.28 (m, 2 H), 7.26–7.12 (m, 5 H), 7.08–6.96 (m, 3 H), 6.92–6.89 (m, 1 H), 6.79 (d, J = 2.5 Hz, 1 H), 6.65 (t, J = 7.2 Hz, 1 H), 6.24 (s, 1 H), 3.59-3.57 (m, 2 H), 3.04-2.98 (m, 1 H), 2.93-2.87 (m, 1 H); ¹³C NMR (126 MHz, Dimethyl sulfoxide- d_6): $\delta = 149.1$, 137.9, 136.5, 134.9, 129.0, 128.4, 127.7, 126.4, 125.9, 125.6, 124.3, 121.0, 119.2, 118.5, 117.4, 117.0, 114.5, 111.5, 55.4, 26.5; HRMS (m/z): [M + H]⁺ calcd for C_{23H21N2⁺} 325.1699, found 325.1704



Following the general procedure, was obtained as a colorless oil (6.1 mg, 45%); ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 8.12$ (s, 1 H), 7.33–7.14 (m, 6 H), 6.99 (d, J = 8.1 Hz, 2 H), 6.83 (t, J = 7.2 Hz, 1 H), 6.66 (q, J = 2.3 Hz, 1 H), 6.08 (q, J = 2.9 Hz, 1 H), 5.90 (s, 1 H), 5.70 (d, J = 3.4 Hz, 1 H), 3.61–3.56 (m, 1 H), 3.51–3.47 (m, 1 H), 3.03–2.97 (m, 1 H), 2.79–2.74 (m, 1 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 150.0, 136.0, 135.5, 133.6, 129.6, 128.9, 127.9, 127.3, 126.1, 118.9, 117.2, 115.7, 108.4, 107.7, 57.7, 43.0, 27.2; HRMS ($ *m*/*z*): [M + H]⁺ calcd for C₁₉H₁₉N₂⁺ 275.1619, found 275.1614.

1-(1-Methyl-1*H*-indol-3-yl)-2-phenyl-1,2,3,4-tetrahydroisoquinoline (5m)



Following the general procedure, was obtained as a white powder (11.1 mg, 66%), m.p. 126 \sim 128 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.55 (d, *J* = 8.0 Hz, 1 H), 7.31–7.28 (m, 1 H), 7.27–7.13 (m, 7 H), 7.02 (dd, *J* = 8.0, 5.4 Hz, 3 H), 6.77 (t, *J* = 7.3 Hz, 1 H), 6.51 (s, 1 H), 6.18 (s, 1 H), 3.66 (s, 3 H), 3.65–3.58 (m, 2 H), 3.07 (dt, *J* = 15.9, 8.0 Hz, 1 H), 2.82 (d, *J* = 16.1 Hz, 1 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 149.9, 137.8, 137.5, 135.7, 129.3, 128.9, 128.2, 127.0, 126.8, 125.8, 121.8, 120.3, 119.3, 118.1, 117.8, 115.8, 109.3, 56.7, 42.3, 32.9, 26.8; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₄H₂₃N₂⁺ 339.1856, found 339.1848.

1-(1,6-Dimethyl-1*H*-indol-3-yl)-2-phenyl-1,2,3,4-tetrahydroisoquinoline (5n)



Following the general procedure, was obtained as a white powder (13.9 mg, 79%), m.p. 120 \sim 122 °C; ¹H NMR (500 MHz, Chloroform-*d*): δ = 7.41 (d, *J* = 8.1 Hz, 1 H), 7.31–7.28 (m, 1 H), 7.26–7.22 (m, 2 H), 7.20–7.13 (m, 3 H), 7.06–6.99 (m, 3 H), 6.86 (dd, *J* = 8.2, 1.5 Hz, 1 H), 6.77 (t, *J* = 7.3 Hz, 1 H), 6.43 (s, 1 H), 6.15 (s, 1 H), 3.69–3.58 (m, 5 H), 3.07 (ddd, *J* = 15.6, 9.3, 5.9 Hz, 1 H), 2.82 (dt, *J* = 16.1, 4.6 Hz, 1 H), 2.46 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): δ = 149.94, 137.90, 137.85, 135.68, 131.62, 129.32, 128.89, 128.34, 128.14, 126.72, 125.80, 124.87, 120.99, 119.96, 118.00, 117.61, 115.72, 109.28, 56.80, 42.31, 32.76, 26.83, 21.95; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₅H₂₅N₂⁺ 353.2012, found 353.2017.

1-(7-Methoxy-1-methyl-1*H*-indol-3-yl)-2-phenyl-1,2,3,4-tetrahydroisoquinoline (50)



Following the general procedure, was obtained as a light yellow powder (14.0 mg, 76%), m.p. $131 \sim 133 \,^{\circ}$ C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.27 \,(d, J = 8.5 \text{ Hz}, 1 \text{ H})$, $7.25-7.21 \,(m, 2 \text{ H})$, $7.19-7.11 \,(m, 4 \text{ H})$, $7.02 \,(d, J = 8.1 \text{ Hz}, 2 \text{ H})$, $6.90 \,(t, J = 7.9 \text{ Hz}, 1 \text{ H})$, $6.75 \,(d, J = 7.6 \text{ Hz}, 1 \text{ H})$, $6.58 \,(d, J = 7.7 \text{ Hz}, 1 \text{ H})$, $6.36 \,(s, 1 \text{ H})$, $6.12 \,(s, 1 \text{ H})$, $3.91 \,(s, 3 \text{ H})$, $3.89 \,(s, 3 \text{ H})$, $3.64 \,(q, J = 7.2, 5.6 \text{ Hz}, 2 \text{ H})$, $3.06 \,(dt, J = 15.8, 7.7 \text{ Hz}, 1 \text{ H})$, $2.80 \,(d, J = 16.3 \text{ Hz}, 1 \text{ H})$; ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 149.88$, 147.84, 137.78, 135.66, 130.02, 129.32, 129.29, 128.92, 128.20, 127.15, 126.70, 125.80, 119.76, 118.02, 117.80, 115.75, 113.03, 102.62, 56.58, 55.52, 42.28, 36.61, 26.69; HRMS <math>(m/z): [M + H]⁺ calcd for C₂₅H₂₅N₂O⁺ 369.1961, found 369.1957.

1-(5-Chloro-1,2-dimethyl-1*H*-indol-3-yl)-2-phenyl-1,2,3,4-tetrahydroisoquinoline (5p)



Following the general procedure, was obtained as a light yellow powder (8.9 mg, 46%), m.p. $105 \sim 107 \,^{\circ}$ C; ¹H NMR (500 MHz, Chloroform-*d*): $\delta = 7.21-7.12$ (m, 4 H), 7.08 (d, J = 8.7 Hz, 2 H), 6.99 (td, J = 6.0, 5.5, 2.7 Hz, 4 H), 6.88–6.82 (m, 2 H), 5.89 (s, 1 H), 3.70–3.58 (m, 2 H), 3.56 (s, 3 H), 3.05 (s, 2 H), 2.11 (s, 3 H); ¹³C NMR (126 MHz, Chloroform-*d*): $\delta = 151.16$, 135.02, 128.92, 128.85, 128.76, 128.22, 126.56, 126.30, 124.88, 120.77, 120.71, 120.59, 120.10, 119.12, 118.68, 109.74, 109.52, 99.45, 57.28, 46.95, 29.74, 28.14, 10.80; HRMS (*m*/*z*): [M + H]⁺ calcd for C₂₅H₂₄ClN₂⁺ 387.1623, found 387.1629.

III References

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- 2. J. Zhang, B. Tiwari, C. Xing, X. Chen and Y. R. Chi, Angew. Chem. Int. Ed., 2012, 51, 3649-3652.

IV NMR Spectra ¹H NMR Spectrum of 2a' (500 MHz, CDCl₃)



¹³C NMR Spectrum of 2a' (126 MHz, CDCl₃)



¹H NMR Spectrum of 4a' (500 MHz, CDCl₃)



¹³C NMR Spectrum of 4a' (126 MHz, CDCl₃)



¹H NMR Spectrum of 4ab and 4ab' (500 MHz, CDCl₃)



¹³C NMR Spectrum of 4ab and 4ab' (126 MHz, CDCl₃)



¹H NMR Spectrum of 2d' (500 MHz, DMSO)



¹³C NMR Spectrum of 2d' (126 MHz, DMSO)



¹H NMR Spectrum of 3a (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3a (126 MHz, CDCl₃)



¹H NMR Spectrum of 3b (500 MHz, CDCl₃)





¹³C NMR Spectrum of 3b (126 MHz, CDCl₃)
¹H NMR Spectrum of 3c (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3c (126 MHz, CDCl₃)



¹H NMR Spectrum of 3d (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3d (126 MHz, CDCl₃)

	Me	O OEt Me Me 3d	t N Me	— 174.04			- 136.55	104.20 127.35	120.51 119.13 118.03	108.70 108.48			77.41	L 76.91	— 61.15		— 40.68	— 29.62	— 14.43	— 10.90	
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210	200	190	180	••••••••••••••••••••••••••••••••••••••	160	150	140	130	120		100		, אוויהאווייי 1008			50	40	30		10	0

¹H NMR Spectrum of 3e (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3e (126 MHz, CDCl₃)



¹H NMR Spectrum of 3f (500 MHz, CDCl₃)

	8 7 7 7 7 7 7 7 7 7 7 7 7 7	1.27 1.25 1.25
	1.90 [±] 1.90 [±] 1.90 [±] 1.90 [±] 1.90 [±]	3.26-
16 15 14 13 12 11 10	9 8 7 6 5 4 3 f1 (ppm)	2 1 0 -1

¹³C NMR Spectrum of 3f (126 MHz, CDCl₃)



¹H NMR Spectrum of 3g (500 MHz, CDCl₃)

$ \begin{array}{c} $		1.29
16 15 14 13 12 11 10	9 8 7 6 5 4 3 f1 (ppm)	

¹³C NMR Spectrum of 3g (126 MHz, CDCl₃)



¹H NMR Spectrum of 3h (500 MHz, CDCl₃)



¹H NMR Spectrum of 3i (500 MHz, CDCl₃)





¹³C NMR Spectrum of 3i (126 MHz, CDCl₃)



¹H NMR Spectrum of 3j (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3j (126 MHz, CDCl₃)



¹H NMR Spectrum of 3k (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3k (126 MHz, CDCl₃)



¹H NMR Spectrum of 3l (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3l (126 MHz, CDCl₃)



¹H NMR Spectrum of 3m (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3m (126 MHz, CDCl₃)



¹H NMR Spectrum of 3n (500 MHz, CDCl₃)







¹H NMR Spectrum of 30 (500 MHz, CDCl₃)



¹³C NMR Spectrum of 30 (126 MHz, CDCl₃)



¹H NMR Spectrum of 3p (500 MHz, DMSO)



¹³C NMR Spectrum of 3p (126 MHz, DMSO)



¹H NMR Spectrum of 3q (500 MHz, CDCl₃)







¹H NMR Spectrum of 3r (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3r (126 MHz, CDCl₃)



¹H NMR Spectrum of 3s (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3s (126 MHz, CDCl₃)



¹H NMR Spectrum of 3t (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3t (126 MHz, CDCl₃)


¹H NMR Spectrum of 3u (500 MHz, CDCl₃)







¹H NMR Spectrum of 3v (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3v (126 MHz, CDCl₃)



¹H NMR Spectrum of 3w (500 MHz, Acetone-*d*₆)



¹³C NMR Spectrum of 3w (126 MHz, Acetone-*d*₆)



¹H NMR Spectrum of 3x (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3x (126 MHz, CDCl₃)



¹H NMR Spectrum of 3y (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3y (126 MHz, CDCl₃)



¹H NMR Spectrum of 3z (500 MHz, CDCl₃)



¹³C NMR Spectrum of 3z (126 MHz, CDCl₃)



¹H NMR Spectrum of 5a (500 MHz, CDCl₃)



 138.32

 136.56

 135.45

 135.45

 135.45

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 135.45

 135.45

 135.45

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151.09 29.52
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¹³C NMR Spectrum of 5a (126 MHz, CDCl₃)

¹H NMR Spectrum of 5b (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5b (126 MHz, CDCl₃)



¹H NMR Spectrum of 5c (500 MHz, CDCl₃)







¹H NMR Spectrum of 5d (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5d (126 MHz, CDCl₃)



¹H NMR Spectrum of 5e (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5e (126 MHz, CDCl₃)



¹H NMR Spectrum of 5f (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5f (126 MHz, CDCl₃)



¹H NMR Spectrum of 5g (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5g (126 MHz, CDCl₃)



¹H NMR Spectrum of 5h (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5h (126 MHz, CDCl₃)



¹H NMR Spectrum of 5i (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5i (126 MHz, CDCl₃)



¹H NMR Spectrum of 5j (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5j (126 MHz, CDCl₃)



¹H NMR Spectrum of 5k (500 MHz, DMSO)



¹³C NMR Spectrum of 5k (126 MHz, DMSO)



¹H NMR Spectrum of 5l (500 MHz, CDCl₃)






¹H NMR Spectrum of 5m (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5m (126 MHz, CDCl₃)



¹H NMR Spectrum of 5n (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5n (126 MHz, CDCl₃)



¹H NMR Spectrum of 50 (500 MHz, CDCl₃)



¹³C NMR Spectrum of 50 (126 MHz, CDCl₃)



¹H NMR Spectrum of 5p (500 MHz, CDCl₃)



¹³C NMR Spectrum of 5p (126 MHz, CDCl₃)

