

1 Supplementary Information for

2 **Removal of Trichloroethene by Glucose Oxidase Immobilized on Magnetite**

3 **Nanoparticles**

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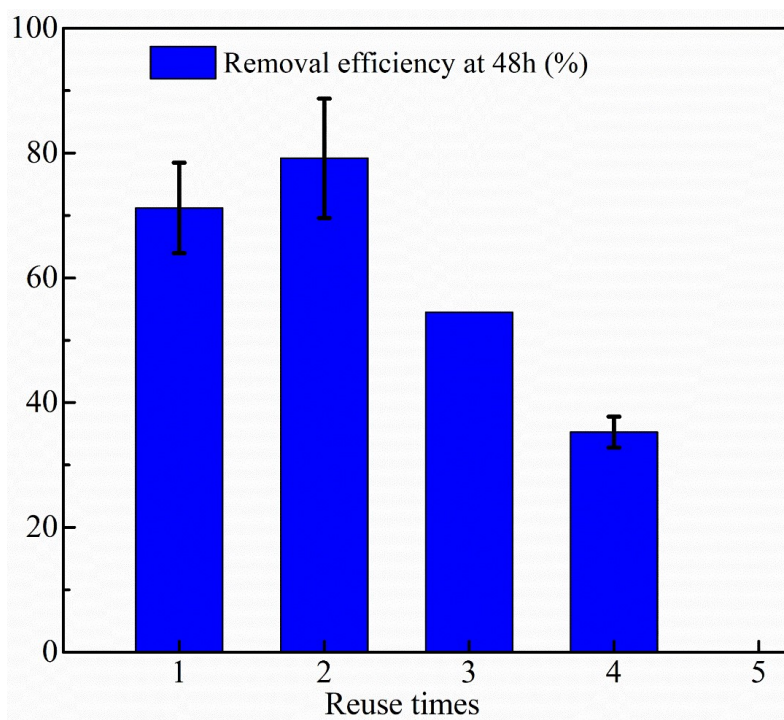
21 **Preparation of Magnetite nanoparticles and MIG**

22 Preparation of magnetite nanoparticles: 8.68 mmol of ferric ammonium sulfate
23 and 4.34 mmol of ferrous ammonium sulfate were dissolved in a 250 ml three-necked
24 flask containing 100 ml of ultrapure anaerobic water and then mechanically stirred at
25 800 rpm in a water bath at 85°C for 20 minutes under the protection of N₂. Then 25%
26 (v/v) ammonia solution was added dropwise and continued dropwise when the color of
27 the solution changed from orange-red to black until the pH of the mixture reached 10.
28 The mixture was transferred to an anaerobic flask and the magnetite particles were
29 separated with neodymium magnets and rinsed several times using deionized water to
30 remove excess ammonia and salt to pH neutral. The magnetite was dried by freeze dryer
31 and stored in a vacuum bag.

32 Preparation of MIG: First, the magnetite particles (1g) were dispersed in 1 wt%
33 chitosan solution (in 0.05M acetic acid) with a solid-liquid ratio of 10g/L. Then stable
34 chitosan-stabilized magnetic nanoparticles (Fe₃O₄-CS) were obtained by dispersing
35 ultrasonic treatment (40 kHz) for 25 min at room temperature, followed by rinsing with
36 deionized water to pH-neutral, and lyophilization after separation by a magnet. 200 mg
37 Fe₃O₄-CS was added in 30 ml anaerobic water and sonicated at 25°C for 5 min, then
38 0.4 ml 25% glutaraldehyde (0.2%) was added drop by drop and vortexed for 2 min,
39 then 20 ml solution containing 20 mg of enzyme was added and vortexed for 2 min;
40 finally shaken in a shaker at 150 rpm for 3 h at 30°C (designed enzyme/magnetite
41 content of 10U/mg). The solid was separated with a neodymium magnet and rinsed
42 three times with 50 mL of deionized water to obtain magnetite particles loaded with
43 glucose oxidase (MIG).

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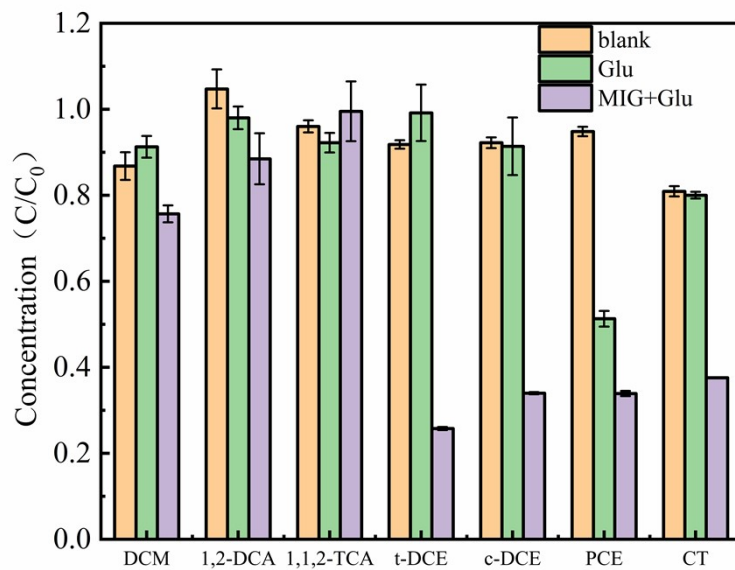


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47 Fig. S1 Effect of recycling numbers of MIG on TCE removal efficiency. Experimental conditions:

48 200 mg MIG, 10 mg/L TCE, , 2.5 mM glucose, 25°C, 150 rpm, 48 h.

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51 Fig. S2 Removal efficiencies of different chlorinated pollutants by MIG-glucose system.

52 Experimental conditions: 200 mg MIG, 10 mg/L chlorinated pollutant, 2.5 mM glucose, 25°C, 150

53 rpm, 48 h.

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55 Table S1 Kinetic fitting results of TCE degradation.

Conditions							Pseudo-first order fitting		
[TCE] (mg/L)	Temp (°C)	pH Initial	pH Buffered	DO (mg/L)	Salt (2mM)	[HA] (mg/L)	$C_t=C_0*e^{-kt}$		
							$k*10^3$ (h ⁻¹)	$t_{1/2}$ (h ⁻¹)	R ²
5							26.26±7.36	26.40	0.7925
10	25	6.1	N/A	6.01	N/A	N/A	62.62±16.31	11.07	0.5868
20							52.29±4.94	13.26	0.9598
50							60.44±8.38	11.47	0.9090
							50.34±8.56	13.77	0.8262
							71.54±6.06	9.69	0.9731
							82.4±12.29	8.41	0.9097
							58.42±4.75	11.86	0.9736
							35.62±7.09	19.46	0.8699
							90.41±20.16	7.67	0.7788
							51.88±11.08	13.36	0.7052
	71.75±9.83	9.66	0.9209						
	79.01±9.82	8.77	0.9459						
				0			20.97±5.55	33.05	0.2711
				3.45			27.62±8.55	25.10	0.1399
10	25	6.1	N/A	6.01	N/A	N/A	21.52±2.23	32.21	0.9189
							25.51±5.62	27.17	0.5420
							26.72±4.48	25.94	0.7615
							24.75±6.56	28.01	0.3085
							23.28±6.01	29.77	0.3226
							23.12±5.14	29.98	0.5130
							26.92±6.04	25.75	0.5345
							22.19±5.49	31.24	0.7896
							30.2±6.41	22.95	0.8470
							18.05±3.28	38.40	0.8448
	17.39±1.12	39.86	0.9750						