

## Supplementary Material

**Figure S1.**  $^1\text{H}$  NMR spectrum of *B. aegyptiaca* leaf extract in  $\text{CD}_3\text{OD}$

**Figure S2.** A) OPLS-DA score plot. B) Loading S-plots derived from mature fruit modeled against immature fruit sample analyzed by  $^1\text{H}$ -NMR ( $\delta$  5.5-10.0 ppm),  $n = 3$ , showing the covariance  $p$  (1) against the correlation  $p(\text{cor})$  (1) of the variables of the discriminating component of the OPLS-DA model. Cut-off values of  $p = 0.18$  was used. C) Permutation plot. Designated variables are highlighted and identifications are discussed in the text

**Figure S3.** Base peak chromatograms (BPC) of five different organs of *Balanites aegyptiaca* in positive ionization mode.

**Figure S4.** Identification of diosgenin aglycone structure. a) fragmentation pathway scheme of diosgenin aglycone, b) MS/MS mirror match between the diosgenin in *Balanites aegyptiaca* and GNPS library spectrum, and c) Peak areas of diosgenin aglycone in different organs of *Balanites aegyptiaca*.

**Figure S5.** MS/MS fragmentation and abundance of compound **2** (diosgenin-26- hexoside\*) in different organs.

**Figure S6.** MS/MS fragmentation and abundance of compound **3** (diosgenin-3-hexoside).

**Figure S7.** GC-MS-based OPLS-DA score plot (a) derived from modeling silylated primary metabolites of *Balanites aegyptiaca* ripe fruit versus unripe fruit ( $n = 3$ ). (b) Derived from modeling silylated primary metabolites of *B. aegyptiaca* leaf&stem versus other 3 organs( $n = 3$ ). (c) and (d)The respective loading S-plots showing the covariance  $p$  [1] against the correlation  $p(\text{cor})$  [1] of the variables of the discriminating component of the OPLS-DA model. Cut-off values of  $p < 0.519281$  was used. Designated variables are highlighted and identifications are discussed in the text

**Table S1:**  $\text{IC}_{50}$  values ( $\mu\text{g}/\text{mL}$ ) of the plant extracts as determined for the PC-3 prostate cancer and HCT-116 colorectal cancer cell lines by performing MTT and CV assays, respectively.

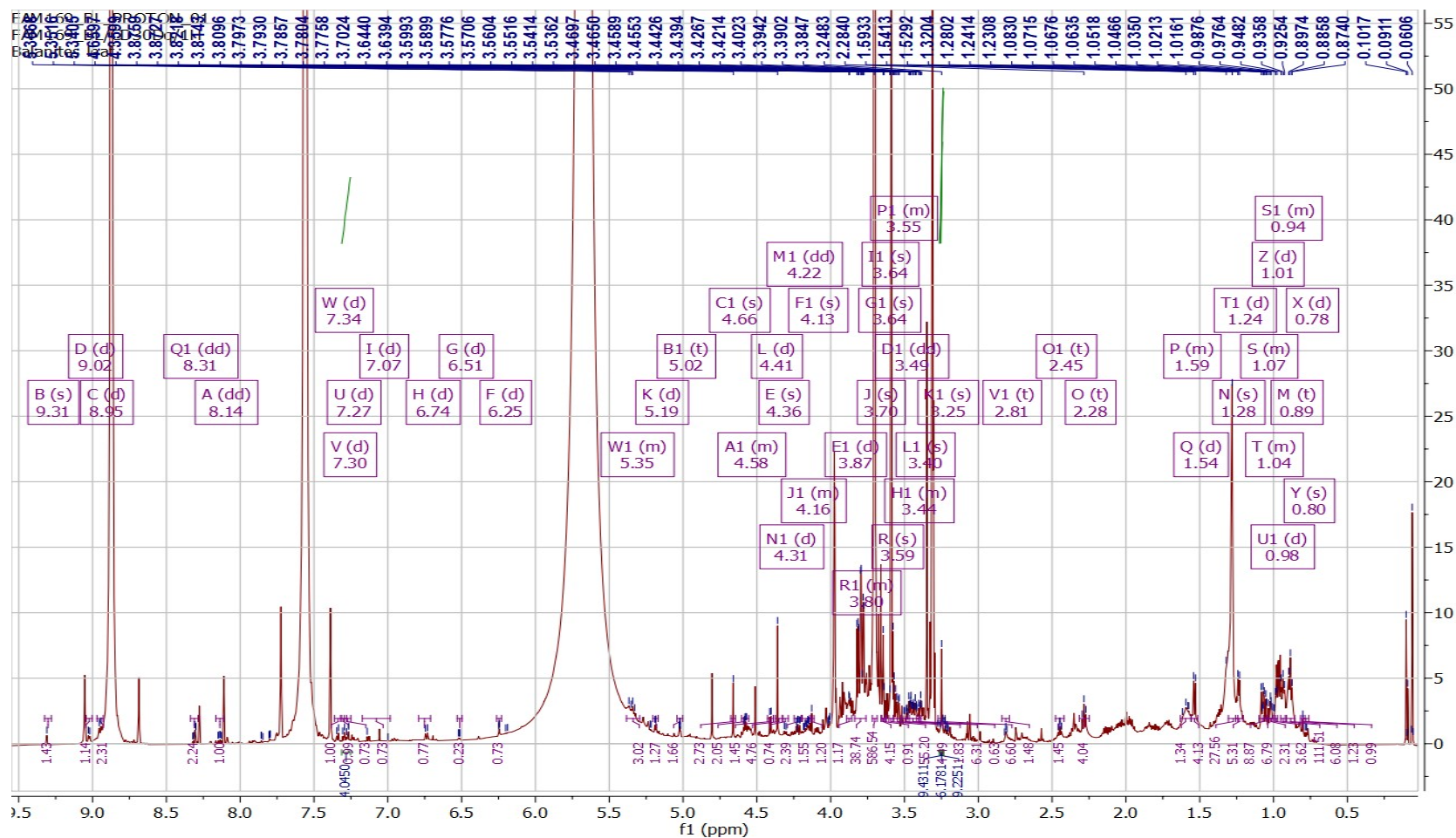
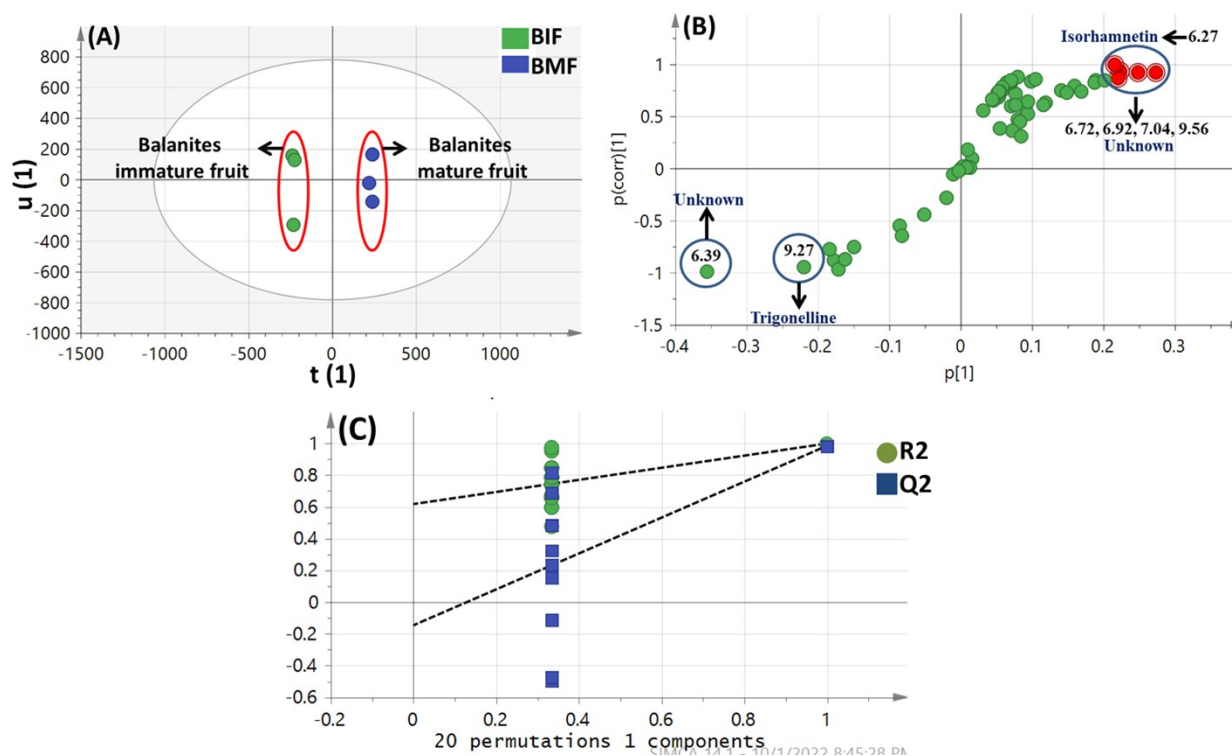
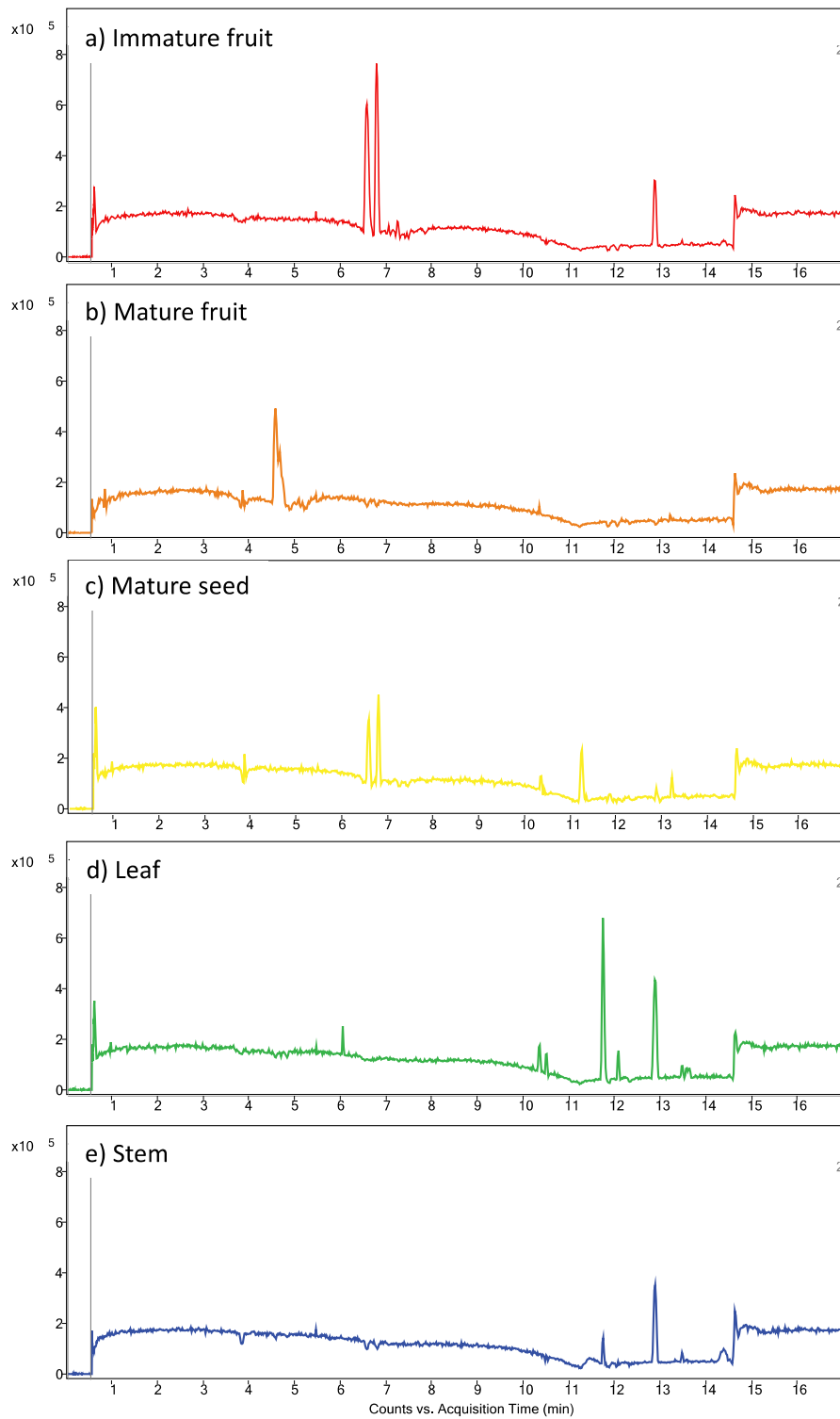


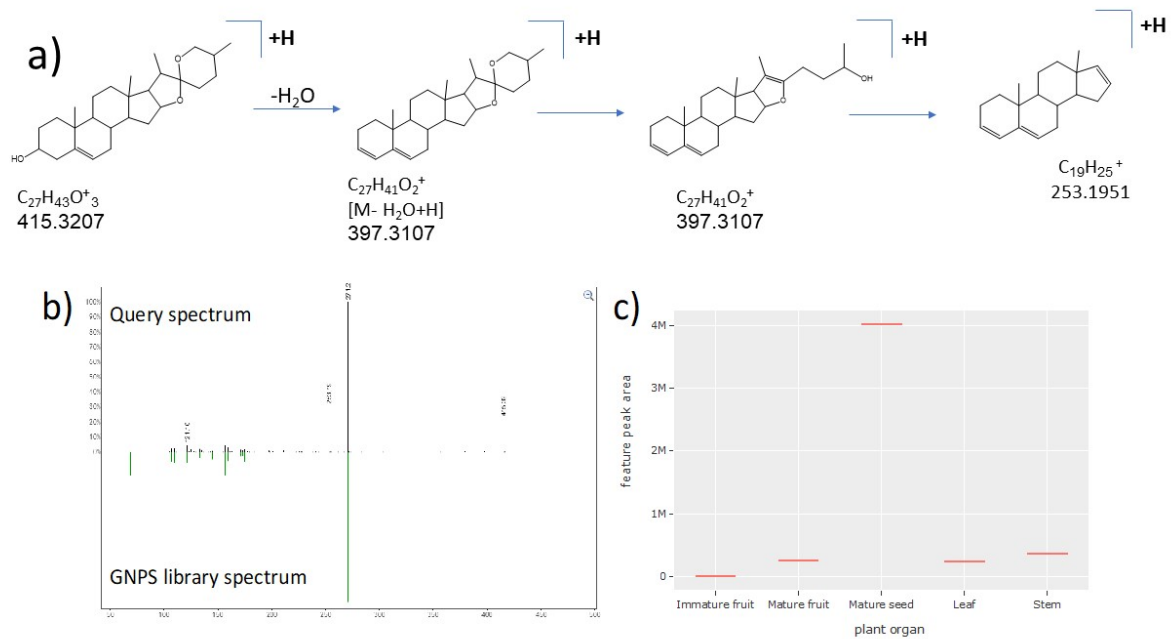
Fig. S1. <sup>1</sup>H NMR spectrum of *Balanites aegyptiaca* leaf extract



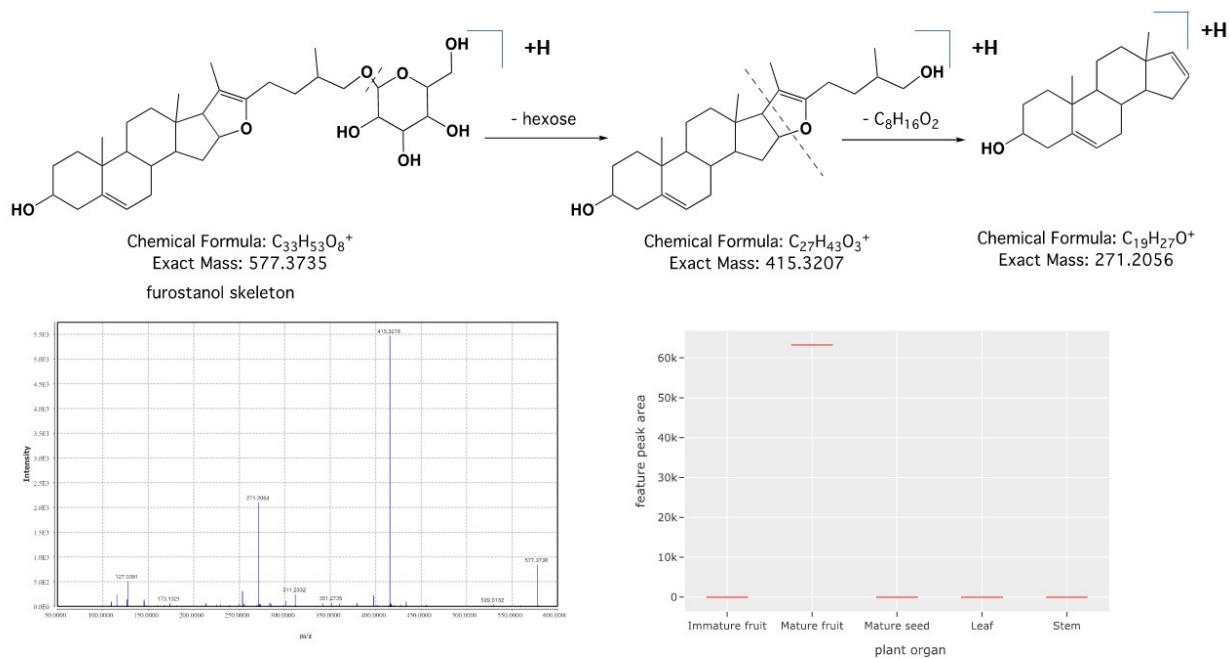
**Figure S2.** A) OPLS-DA score plot. B) Loading S-plots derived from mature fruit modeled against immature fruit sample analyzed by  $^1\text{H-NMR}$  ( $\delta$  5.5-10.0 ppm),  $n = 3$ , showing the covariance  $p(1)$  against the correlation  $p(\text{cor})(1)$  of the variables of the discriminating component of the OPLS-DA model. Cut-off values of  $p = 0.18$  was used. C) Permutation plot. Designated variables are highlighted and identifications are discussed in the text.



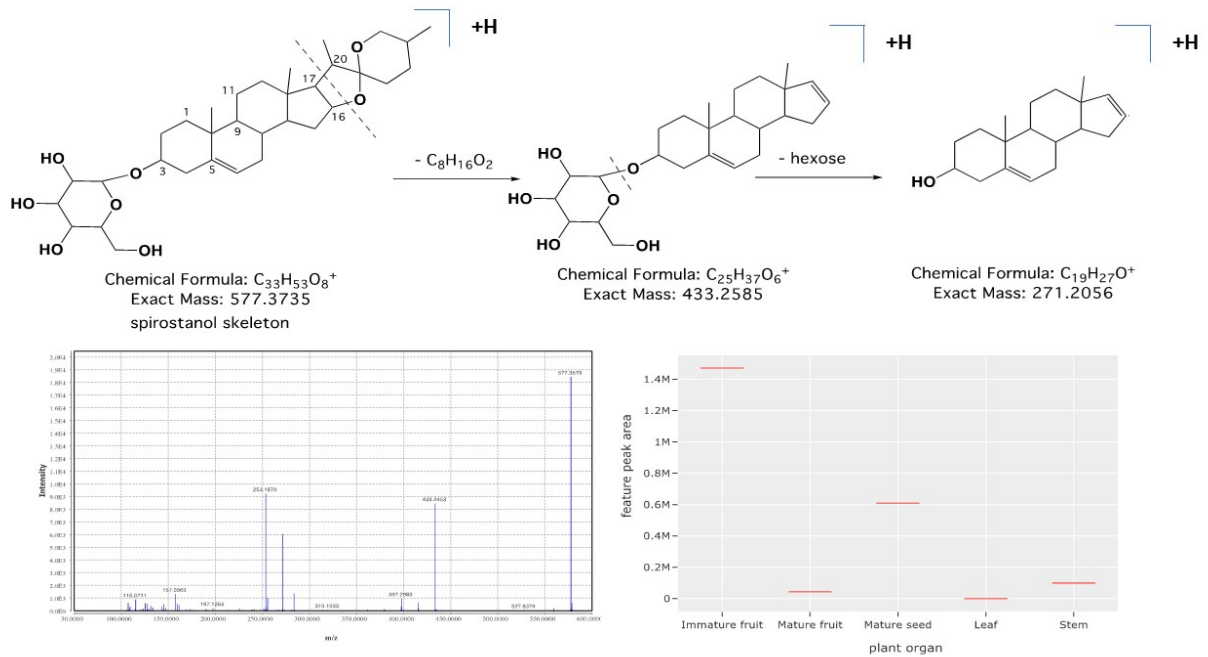
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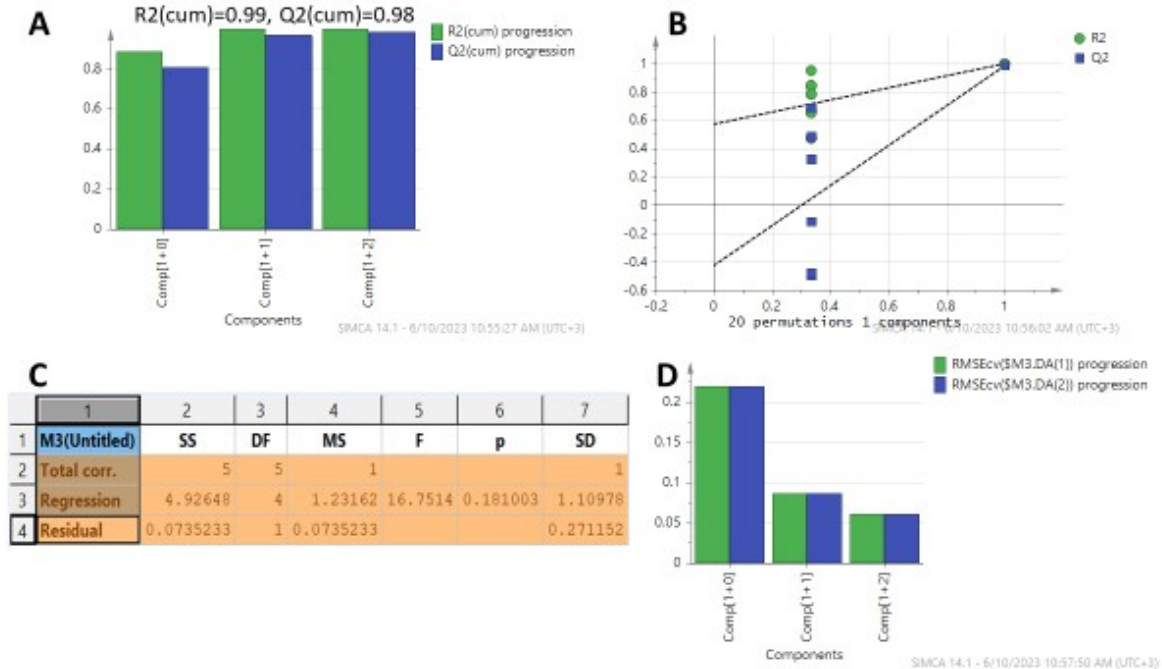
**Figure S4.** Identification of diosgenin aglycone structure. a) fragmentation pathway scheme of diosgenin aglycone, b) MS/MS mirror match between the diosgenin in *Balanites aegyptica* and GNPS library spectrum, and c) Peak areas of diosgenin aglycone in different organs of *Balanites aegyptica*.



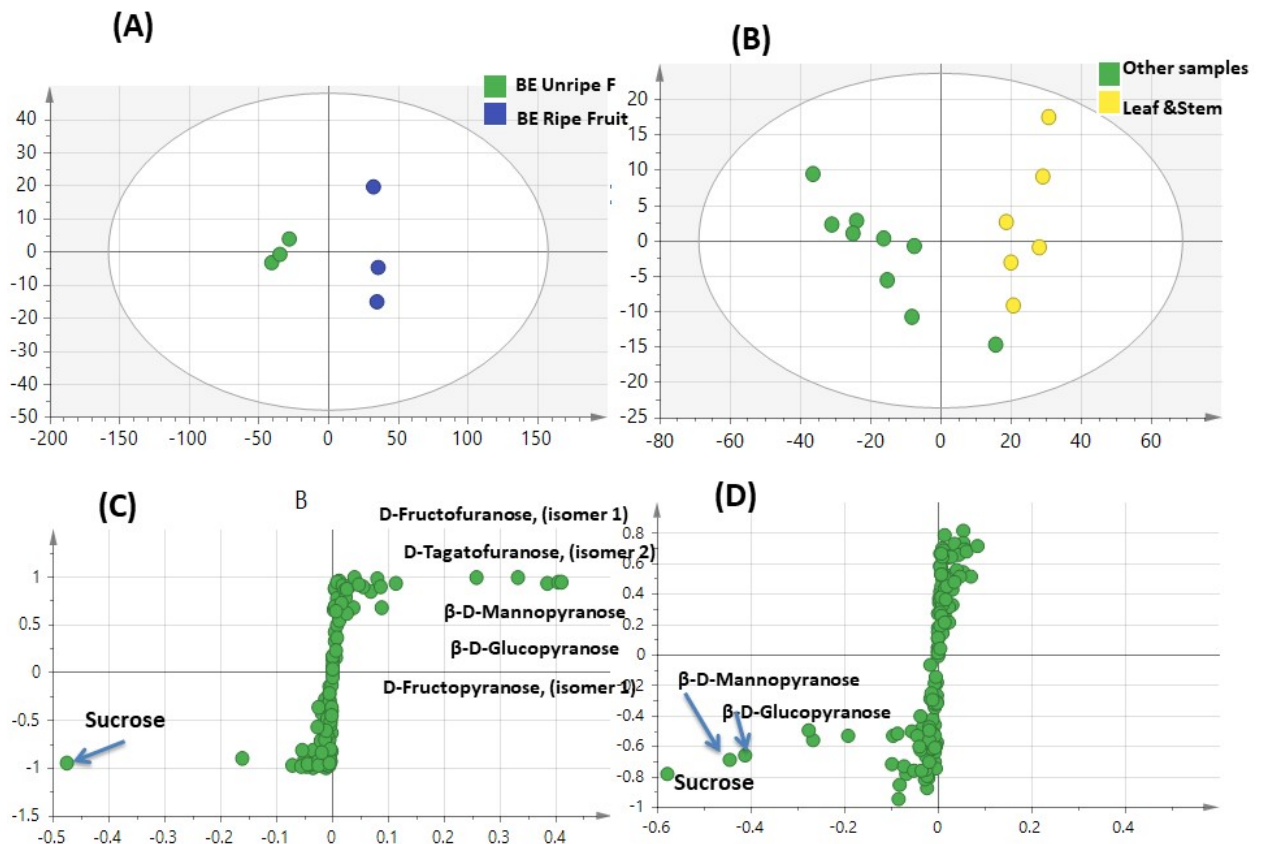
**Figure S5.** MS/MS fragmentation and abundance of compound **2** (diosgenin-26- hexoside\*) in different organs.



**Figure S6.** MS/MS fragmentation and abundance of compound 3 (diosgenin-3-hexoside).



**Fig. S7.** OPLS-DA model validation for modelling *B. aegyptiaca* samples based on LCMS in positive mode **A.** the diagnostic metrics R2 and Q2 **B.** permutation testing. n=20, and **C.** CV-ANOVA to assess for model statistical significance. **D.** SECV residuals.



**Figure S8.** GC-MS-based OPLS-DA score plot (a) derived from modeling silylated primary metabolites of *Balanites aegyptiaca* ripe fruit versus unripe fruit (n = 3). (b) Derived from modeling silylated primary metabolites of *B.aegyptiaca* leaf&stem versus other 3 organs(n = 3). (c) and (d)The respective loading S-plots showing the covariance  $p[1]$  against the correlation  $p(\text{cor})[1]$  of the variables of the discriminating component of the OPLS-DA model. Cut-off values of  $p < 0.519281$  was used. Designated variables are highlighted and identifications are discussed in the text



**Table S1:** IC<sub>50</sub> values (μg/mL) of the plant extracts as determined for the PC-3 prostate cancer and HCT-116 colorectal cancer cell lines by performing MTT and CV assays, respectively. The data represent the mean of three independent biological replicates with at least technical duplicates ± SD.

Relative IC <sub>50</sub> (μg/mL)	<b>PC-3</b>		<b>HCT-116</b>	
	MTT assay	CV assay	MTT assay	CV assay
<b>BS</b>	4.8 ± 1.2	5.6 ± 0.6	6.8 ± 0.5	7.8 ± 1.2
<b>BMF</b>	98.4 ± 19.5	112.6 ± 19.1	> 200	> 200
<b>BST</b>	38.3 ± 7.3	45.6 ± 2.7	47.7 ± 22.0	48.2 ± 18.5
<b>BL</b>	> 200	> 200	> 200	> 200
<b>BIF</b>	2.8 ± 0.3	2.8 ± 0.3	3.4 ± 0.7	3.5 ± 0.4