

## Supporting Information

### ***Site-Specific Radiolabeling Using Mushroom Tyrosinase and the Strain-Promoted Oxidation-Controlled 1,2-Quinone Cycloaddition***

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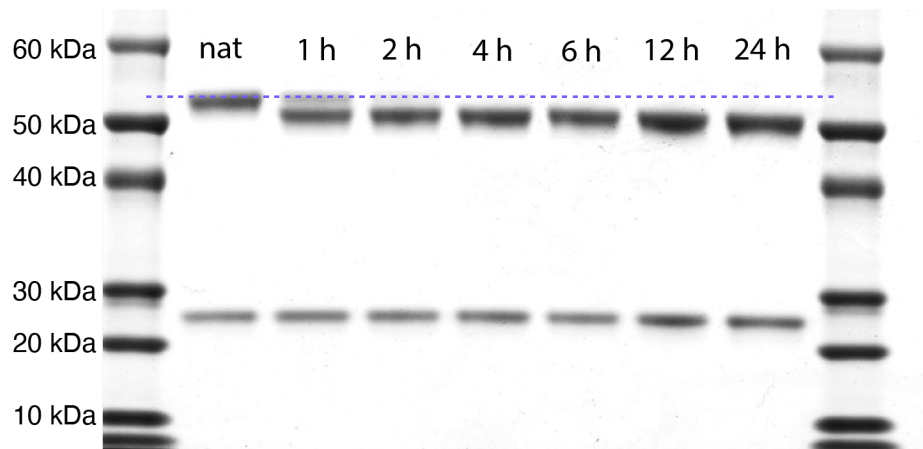
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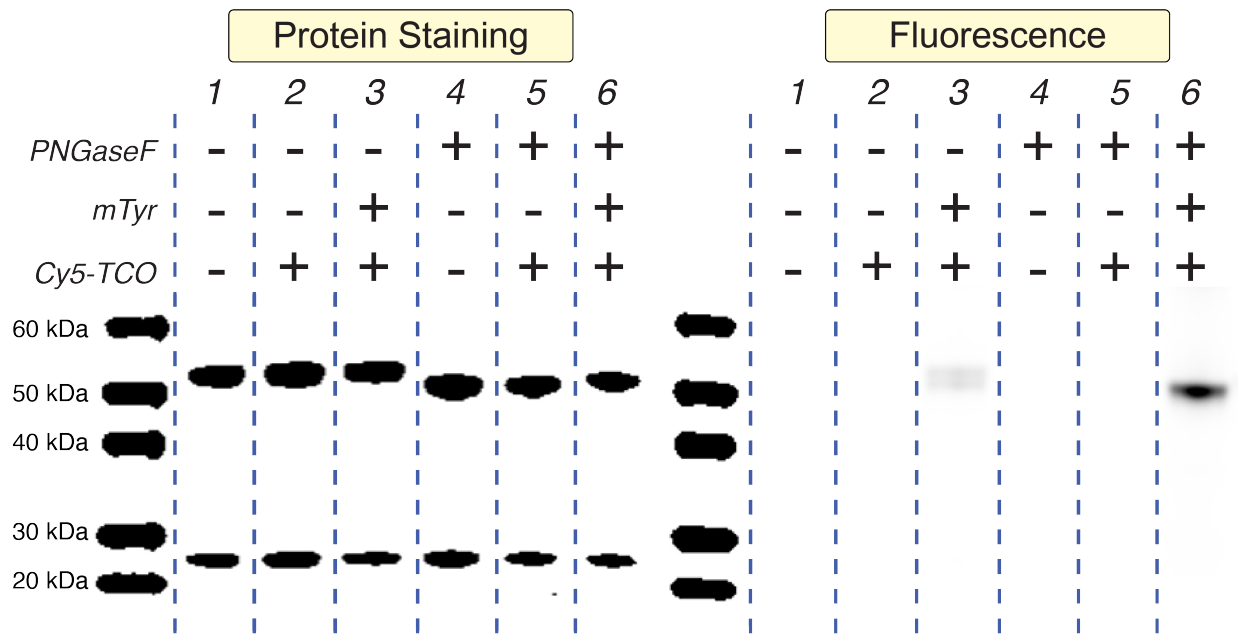
**Running Title:** Site-Specific Radiolabeling Using Mushroom Tyrosinase and the Strain-Promoted Oxidation-Controlled 1,2-Quinone Cycloaddition

**Keywords:** Site-specific bioconjugation; site-selective bioconjugation; biorthogonal chemistry; click chemistry; immunoPET

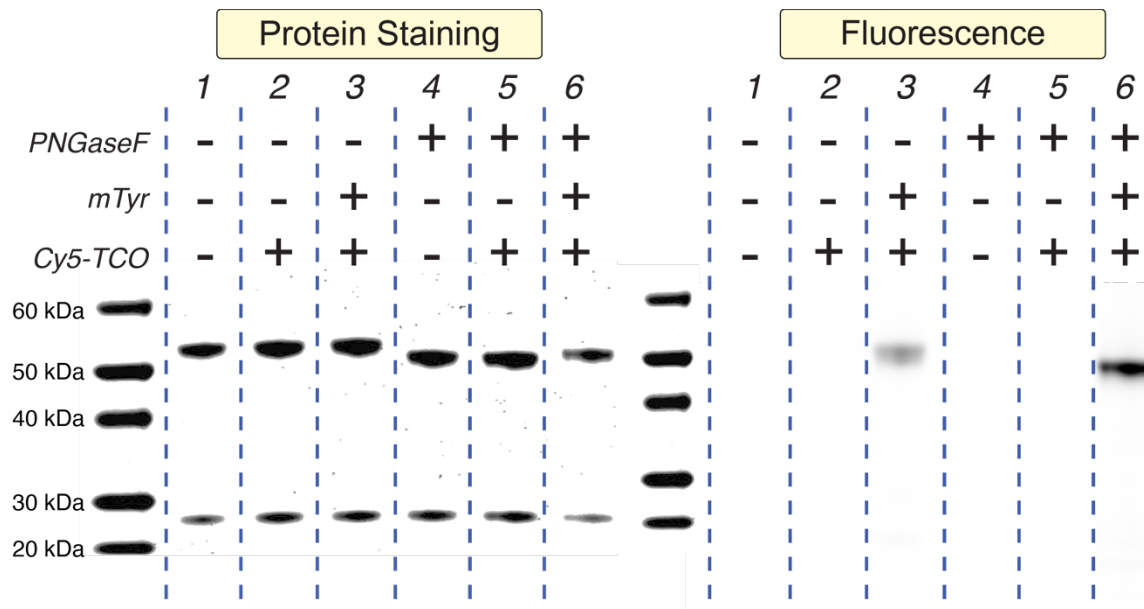
## SUPPLEMENTAL FIGURES



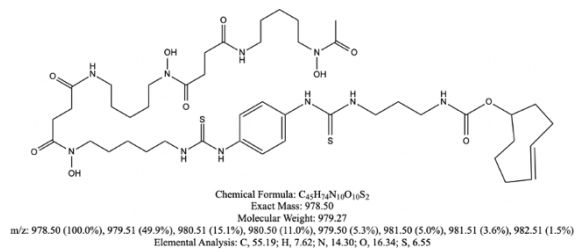
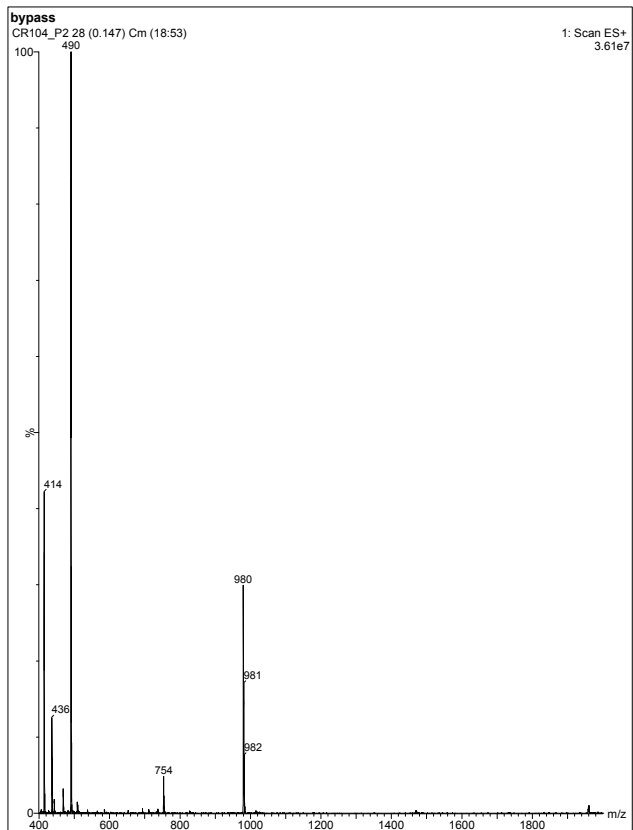
**Figure S1.** Protein-stained SDS-PAGE of huA33 exposed to PNGaseF for various amounts of time. The dotted line denotes the mass of the heavy chain of the parental IgG.



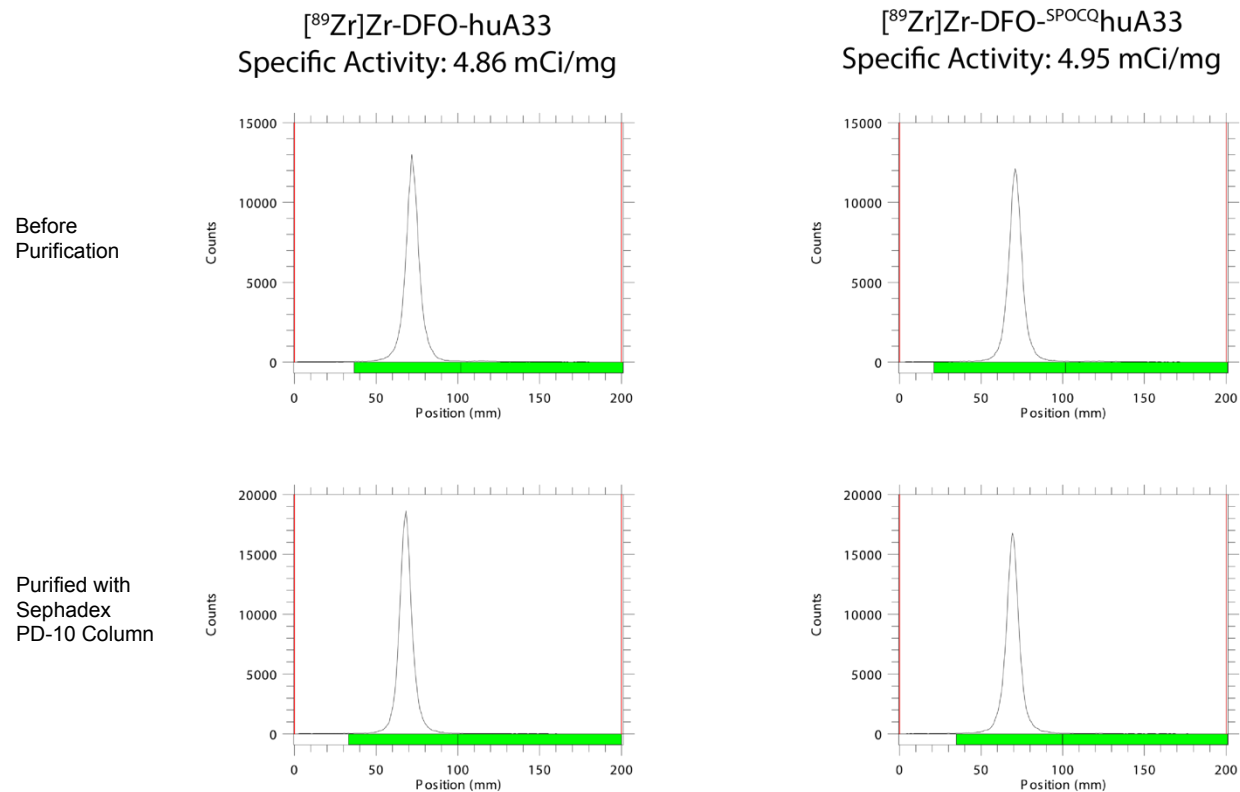
**Figure S2.** Protein-stained and fluorescent SDS-PAGE of pertuzumab modified under various conditions.



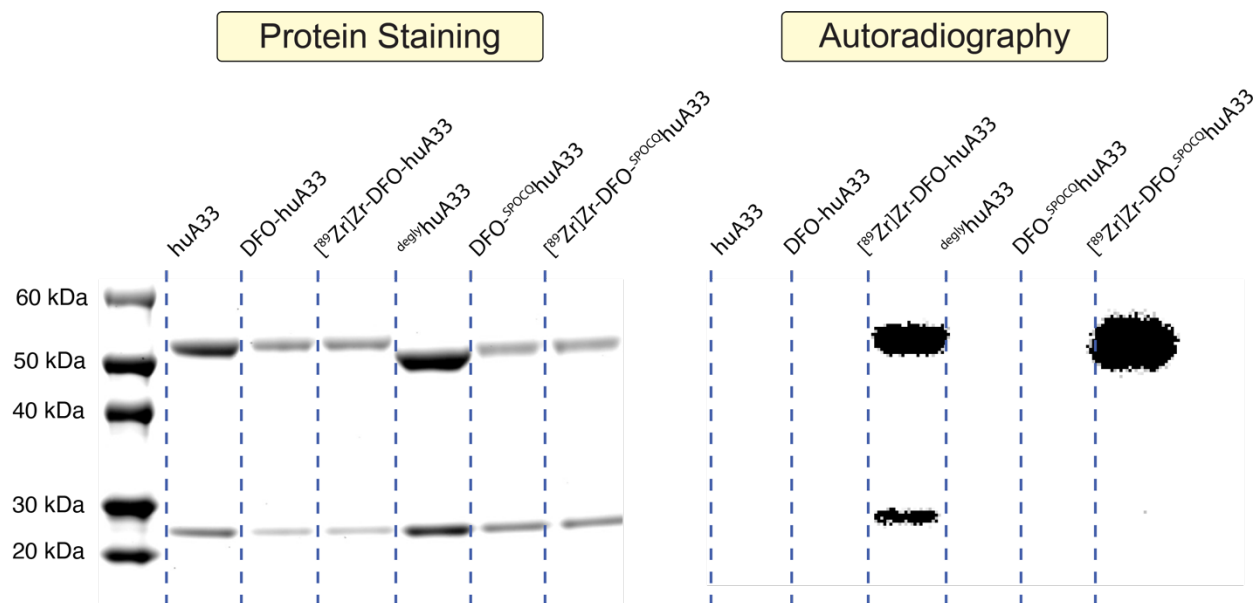
**Figure S3.** Protein-stained and fluorescent SDS-PAGE of trastuzumab modified under various conditions.



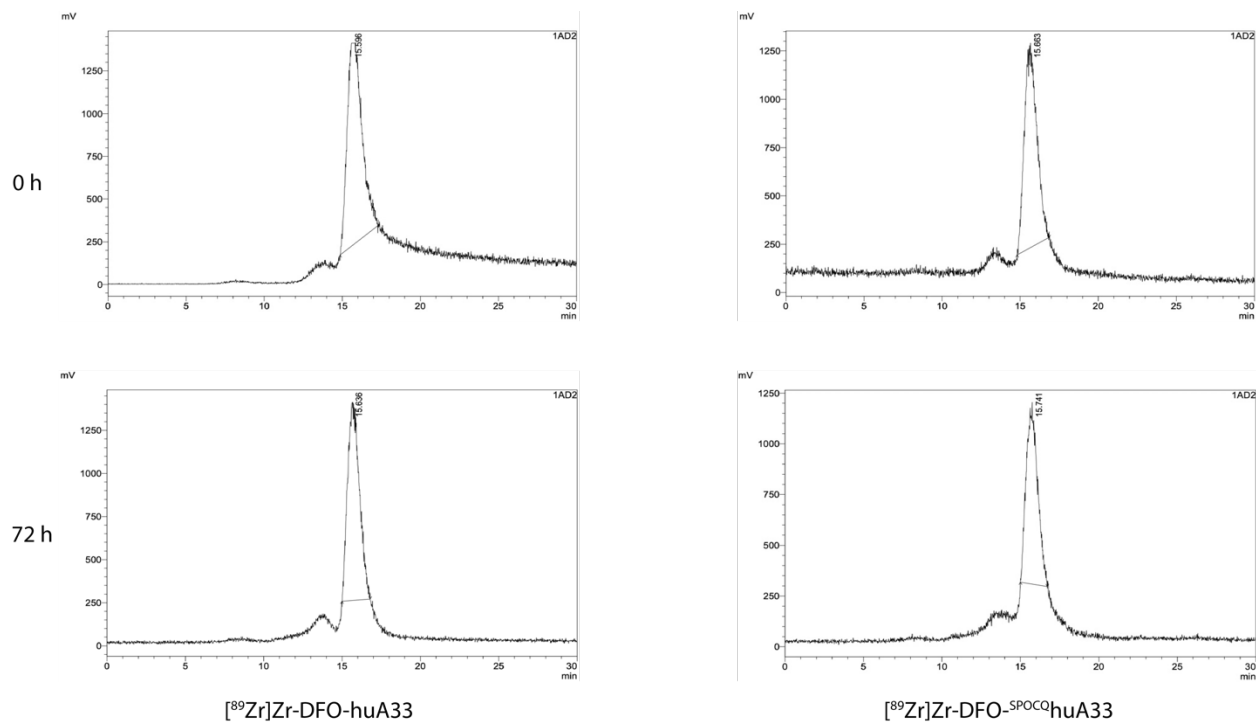
**Figure S4.** ESI-MS of purified DFO-TCO.



**Figure S5. Representative radio-iTLC chromatograms.** The radiolabeling of [<sup>89</sup>Zr]Zr-DFO-huA33 and [<sup>89</sup>Zr]Zr-DFO-<sup>SPOCQ</sup>huA33 was followed via radio-iTLC using EDTA (50 mM, pH 5.0) as the eluent. Both constructs typically displayed >98% radiochemical purity both before and after purification via gel filtration. The peak at ~75 mm corresponds to the radiolabeled antibody; any free [<sup>89</sup>Zr]Zr-EDTA would be seen at ~150 mm. All radiolabeling studies and iTLC scans were performed in triplicate.

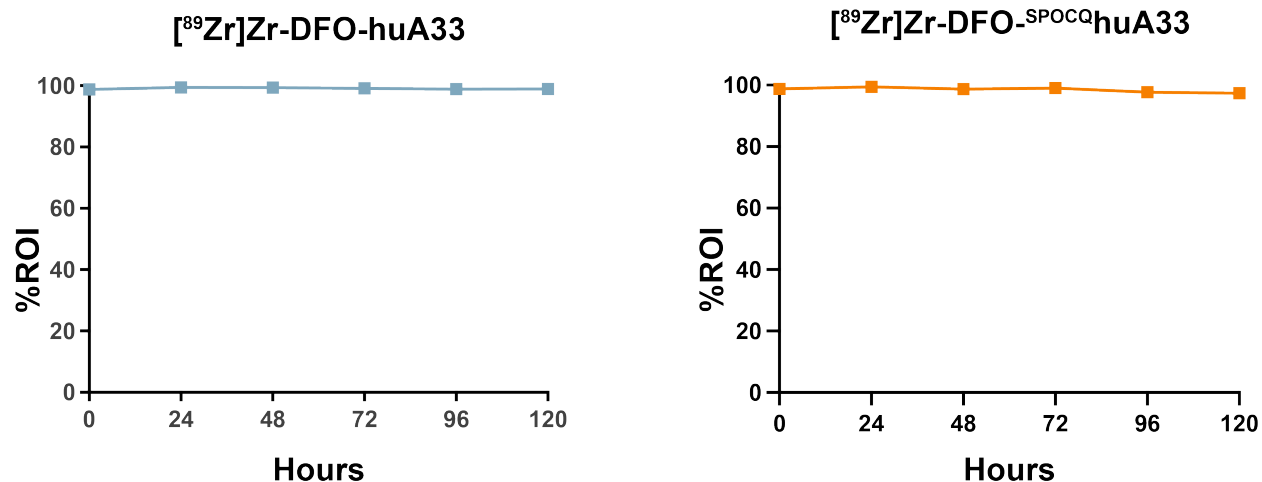


**Figure S6. SDS-PAGE analysis of  $[^{89}\text{Zr}]\text{Zr-DFO-huA33}$  and  $[^{89}\text{Zr}]\text{Zr-DFO-SPOCQhuA33}$ .** Protein-stained reducing SDS-PAGE of the radioimmunoconjugates as well as autoradiography of the stained gel.



**Figure S7. Radio-size-exclusion HPLC for human serum stability study.** Following the radiosynthesis of  $[^{89}\text{Zr}]\text{Zr-DFO-huA33}$  and  $[^{89}\text{Zr}]\text{Zr-DFO-SPOCQhuA33}$ , the radioimmunoconjugates were incubated in 0.5 mL human serum and placed on a thermomixer at 37 °C, 400 rpm (n = 3). At 0 and 72 h after the start of the study, an aliquot of each radioimmunoconjugate was injected into a size-exclusion HPLC with a radioactivity detector. The peak seen at ~15 minutes is the radiolabeled antibody. Any free  $[^{89}\text{Zr}]\text{Zr}^{4+}$  would be seen as a peak at >20 minutes.





**Figure S8. Human serum stability study.** Following the radiosynthesis of  $[^{89}\text{Zr}]\text{Zr-DFO-huA33}$  and  $[^{89}\text{Zr}]\text{Zr-DFO-SPOCQ-huA33}$ , the radioimmunoconjugates were incubated in 0.5 mL human serum and placed on a thermomixer at 37 °C, 400 rpm. At 0, 24, 48, 72, 96, and 120 h after the start of the study, each radioimmunoconjugate was assayed via radio-iTLC with EDTA (50 mM, pH 5.0) as an eluent. The % radioactivity associated with the antibody at each timepoint was plotted using GraphPad Prism 8.0 (n = 3). This data corresponds with the data shown in Table S8.

## SUPPLEMENTAL TABLES

	<b>Cy5-<sup>SPOCQ</sup>huA33 (One-Pot Trial)</b>	<b>Cy5-<sup>SPOCQ</sup>huA33 (High Equivalents)</b>
<b>DOL (Cy5/mAb)</b>	0.2 ± 0.02	1.1 ± 0.1

**Table S1.** Degree of labeling results of the three-reagent-one-pot approach and the stepwise strategy using a 4 h incubation and quadruple the normal number of equivalents of mTyr and Cy5-TCO.

	Cy5-SPOCQ trastuzumab	Cy5-SPOCQ pertuzumab
DOL (Cy5/mAb)	1.6 ± 0.2	1.7 ± 0.2

**Table S2.** Degree of labeling results for trastuzumab and pertuzumab site-specifically modified with Cy5 using the mTyr- and the SPOCQ cycloaddition-based strategy.

	<b>DFO-huA33</b>	<b>DFO-<sup>SPOCQ</sup>huA33</b>
<b>DOL (DFO/mAb)</b>	1.7 ± 0.1	1.6 ± 0.1

**Table S3.** Degree of labeling results for huA33 (a) randomly modified with DFO via the stochastic modification of lysines and (b) site-specifically modified with DFO using our approach based on mTyr and the SPOCQ cycloaddition.

<b>Organs</b>	<b>[<sup>89</sup>Zr]Zr-DFO-huA33</b>	<b>[<sup>89</sup>Zr]Zr-DFO-<sup>SPOCQ</sup>huA33</b>
<b>Blood</b>	2.1 ± 1.6	3.0 ± 0.8
<b>Tumor</b>	70.0 ± 33.8	109.8 ± 17.6
<b>Heart</b>	2.0 ± 0.5	1.5 ± 0.2
<b>Lungs</b>	1.4 ± 0.8	1.7 ± 0.2
<b>Liver</b>	4.0 ± 1.4	2.7 ± 0.8
<b>Spleen</b>	2.0 ± 0.8	2.5 ± 1.1
<b>Pancreas</b>	0.7 ± 0.3	0.6 ± 0.2
<b>Stomach</b>	0.6 ± 0.2	0.5 ± 0.2
<b>S. Intestine</b>	1.2 ± 0.4	0.6 ± 0.1
<b>L. Intestine</b>	0.9 ± 0.0	0.5 ± 0.1
<b>Kidneys</b>	1.7 ± 0.8	4.9 ± 0.5
<b>Ovaries</b>	1.4 ± 0.6	1.2 ± 1.1
<b>Muscle</b>	0.4 ± 0.3	0.5 ± 0.2
<b>Bone</b>	4.6 ± 0.5	7.0 ± 5.3
<b>Skin</b>	2.1 ± 0.9	1.8 ± 1.2
<b>Tail</b>	3.1 ± 2.3	1.5 ± 0.1

**Table S4. Biodistribution data from mice bearing SW1222 xenografts.** Biodistribution data collected 120 h after the intravenous administration of [<sup>89</sup>Zr]Zr-DFO-huA33 or [<sup>89</sup>Zr]Zr-DFO-<sup>SPOCQ</sup>huA33 [3.7 -4.0 MBq (100-110 µCi), 20-22 µg, in 100 µL of PBS] to athymic nude mice bearing A33-expressing SW1222 colorectal carcinoma xenografts (n = 4). Values are in units of %ID/g and are expressed as mean ± standard deviation.

Organs	[ <sup>89</sup> Zr]Zr-DFO-huA33	[ <sup>89</sup> Zr]Zr-DFO- <sup>SPOCQ</sup> huA33
Blood	0.3 ± 0.1	5.7 ± 0.4
Tumor	33.1 ± 10.6	76.2 ± 13.0
Heart	1.8 ± 0.6	1.1 ± 0.6
Lungs	1.6 ± 0.2	3.3 ± 1.8
Liver	16.2 ± 0.6	5.9 ± 2.1
Spleen	42.1 ± 8.6	9.7 ± 4.0
Pancreas	1.1 ± 0.8	0.9 ± 0.6
Stomach	1.1 ± 0.4	1.2 ± 0.1
S. Intestine	5.1 ± 0.4	1.7 ± 0.2
L. Intestine	1.2 ± 0.6	2.9 ± 3.7
Kidneys	1.4 ± 0.3	6.2 ± 5.7
Ovaries	3.1 ± 1.4	1.0 ± 0.2
Muscle	0.3 ± 0.1	0.3 ± 0.1
Bone	8.2 ± 5.1	6.6 ± 0.5
Skin	2.2 ± 0.2	3.8 ± 0.4
Tail	2.7 ± 1.3	2.0 ± 0.2

**Table S5. Biodistribution data from mice bearing SW1222 xenografts.** Biodistribution data collected 120 h after the intravenous administration of [<sup>89</sup>Zr]Zr-DFO-huA33 or [<sup>89</sup>Zr]Zr-DFO-<sup>SPOCQ</sup>huA33 [3.7 -4.0 MBq (100-110 μCi), 20-22 μg, in 100 μL of PBS] to NSG mice bearing A33-expressing SW1222 colorectal carcinoma xenografts (n = 4). Values are in units of %ID/g and are expressed as mean ± standard deviation.

Immunoconjugate Concentration ( $\mu\text{g/ml}$ )	Absorbance @ 450 nm			
	huA33	<sup>degly</sup> huA33	DFO-huA33	DFO- <sup>SPOCQ</sup> huA33
<b>1000</b>	0.44 $\pm$ 0.03	0.45 $\pm$ 0.01	0.41 $\pm$ 0.02	0.43 $\pm$ 0.004
<b>333.3</b>	0.41 $\pm$ 0.02	0.44 $\pm$ 0.01	0.38 $\pm$ 0.02	0.41 $\pm$ 0.01
<b>111.1</b>	0.38 $\pm$ 0.31	0.41 $\pm$ 0.002	0.34 $\pm$ 0.02	0.38 $\pm$ 0.01
<b>37.03</b>	0.31 $\pm$ 0.01	0.35 $\pm$ 0.01	0.26 $\pm$ 0.02	0.33 $\pm$ 0.003
<b>12.34</b>	0.2 $\pm$ 0.01	0.25 $\pm$ 0.01	0.17 $\pm$ 0.002	0.25 $\pm$ 0.004
<b>4.115</b>	0.12 $\pm$ 0.01	0.16 $\pm$ 0.02	0.10 $\pm$ 0.001	0.18 $\pm$ 0.01
<b>1.371</b>	0.07 $\pm$ 0.002	0.11 $\pm$ 0.01	0.07 $\pm$ 0.003	0.15 $\pm$ 0.03
<b>0.475</b>	0.06 $\pm$ 0.002	0.10 $\pm$ 0.02	0.06 $\pm$ 0.01	0.14 $\pm$ 0.01

**Table S6. ELISA results for A33 binding.** A33-coated plates were incubated with varying concentrations of huA33, <sup>degly</sup>huA33, DFO-huA33, or DFO-<sup>SPOCQ</sup>huA33 (n = 3). After a 2 h incubation, HRP-labeled goat anti-human secondary antibody was incubated in each well for 1 h followed by TMB for 10 minutes. The absorbance of each well was detected on a microplate reader, and the results were analyzed using GraphPad Prism 8.0 software. This data corresponds with the data shown in Figure 2.

Immunoconjugate Concentration ( $\mu\text{g/ml}$ )	Absorbance @ 450 nm			
	huA33	<sup>degly</sup> huA33	DFO-huA33	DFO- <sup>SPOCQ</sup> huA33
<b>1000</b>	0.49 $\pm$ 0.01	0.14 $\pm$ 0.01	0.46 $\pm$ 0.01	0.18 $\pm$ 0.01
<b>333.3</b>	0.44 $\pm$ 0.01	0.10 $\pm$ 0.002	0.41 $\pm$ 0.01	0.11 $\pm$ 0.002
<b>111.1</b>	0.38 $\pm$ 0.01	0.07 $\pm$ 0.00	0.35 $\pm$ 0.01	0.07 $\pm$ 0.001
<b>37.03</b>	0.28 $\pm$ 0.01	0.05 $\pm$ 0.003	0.24 $\pm$ 0.01	0.06 $\pm$ 0.002
<b>12.34</b>	0.16 $\pm$ 0.002	0.04 $\pm$ 0.001	0.13 $\pm$ 0.004	0.05 $\pm$ 0.001
<b>4.115</b>	0.09 $\pm$ 0.002	0.04 $\pm$ 0.00	0.08 $\pm$ 0.003	0.05 $\pm$ 0.003
<b>1.371</b>	0.06 $\pm$ 0.001	0.04 $\pm$ 0.001	0.06 $\pm$ 0.004	0.05 $\pm$ 0.01
<b>0.475</b>	0.07 $\pm$ 0.003	0.04 $\pm$ 0.001	0.05 $\pm$ 0.001	0.04 $\pm$ 0.001

**Table S7. ELISA results for hFc $\gamma$ RI binding.** hFc $\gamma$ RI- coated plates were incubated with varying concentrations of huA33, <sup>degly</sup>huA33, DFO-huA33, or DFO-<sup>SPOCQ</sup>huA33 (n = 3). After a 2 h incubation, HRP-labeled goat anti-human secondary antibody was incubated in each well for 1 h followed by TMB for 10 minutes. The absorbance of each well was detected on a microplate reader, and the results were analyzed using GraphPad Prism 8.0 software. This data corresponds with the data shown in Figure 2.



Constructs	%ROI of radio-iTLC					
	0 h	24 h	48 h	72 h	96 h	120 h
$[^{89}\text{Zr}]\text{Zr-DFO-huA33}$	98.8 ± 0.3	99.4 ± 0.2	99.4 ± 0.2	99.1 ± 0.2	98.9 ± 0.1	99.0 ± 0.2
$[^{89}\text{Zr}]\text{Zr-DFO-}^{\text{SPOCQ}}\text{huA33}$	98.8 ± 0.2	99.4 ± 0.1	98.7 ± 0.3	99.0 ± 0.0	97.7 ± 0.3	97.4 ± 0.6

**Table S8. Human serum stability study results.** Following the radiosynthesis of  $[^{89}\text{Zr}]\text{Zr-DFO-huA33}$  and  $[^{89}\text{Zr}]\text{Zr-DFO-}^{\text{SPOCQ}}\text{huA33}$ , the radioimmunoconjugates were incubated in 0.5 mL human serum and placed on a thermomixer at 37 °C, 400 rpm. At 0, 24, 48, 72, 96, and 120 h after the start of the study, each radioimmunoconjugate was assayed via radio-iTLC using EDTA (50 mM, pH 5.0) as the eluent (n = 3). This data corresponds with the data shown in Figure S8.