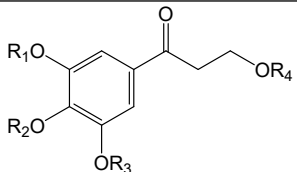
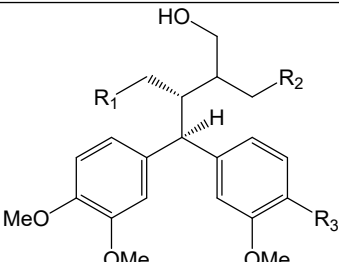
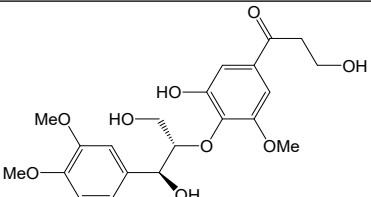
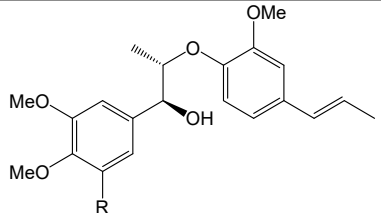
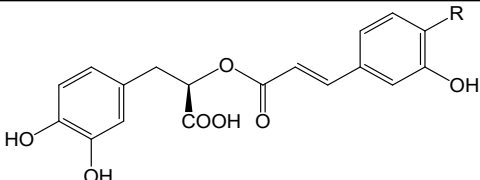
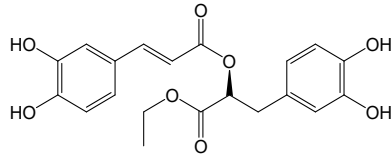
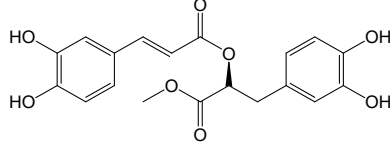


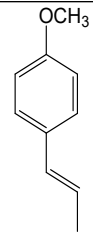
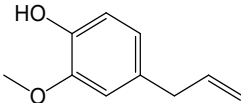
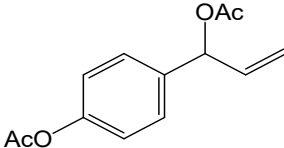
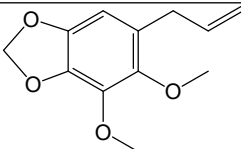
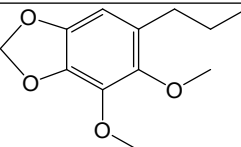
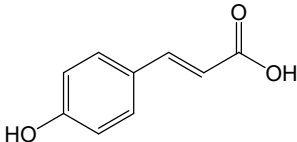
Supplementary Material

Table S1: A list of plant-derived phenylethanoids and phenylpropanoids with anti-viral and anti-protozoal activities

No.	Compound	Source	Structure	Activity	Ref.
					
1	Feddeiketone A	<i>Daphne feddei</i> F. Thymelaeaceae	$R_1 = R_2 = \text{Me}; R_3 = \text{H}; R_4 = \text{Ac}$	Activity against HIV-1 EC ₅₀ 3.96 µg/mL CC ₅₀ 64.5 µg/mL TI 16.3	(Lu et al., 2012)
2	Feddeiketone B	<i>Daphne feddei</i> F. Thymelaeaceae	$R_1 = R_2 = \text{Me}; R_3 = R_4 = \text{H}$	Activity against HIV-1 EC ₅₀ 3.16 µg/mL CC ₅₀ 96.3 µg/mL TI 30.5	(Lu et al., 2012)
3	Feddeiketone C	<i>Daphne feddei</i> F. Thymelaeaceae	$R_1 = \text{Me}; R_2 = R_3 = \text{CH}_2; R_4 = \text{H}$	Activity against HIV-1 EC ₅₀ 2.47 µg/mL CC ₅₀ 43.4 µg/mL TI 17.6	(Lu et al., 2012)
					
4	Feddeiphenol A	<i>Daphne feddei</i> F. Thymelaeaceae	$R_1 = R_2 = \text{OH}; R_3 = \text{OMe}$	Activity against HIV-1 EC ₅₀ 4.08 µg/mL CC ₅₀ 128.2 µg/mL TI 31.4	(Hu et al., 2011)

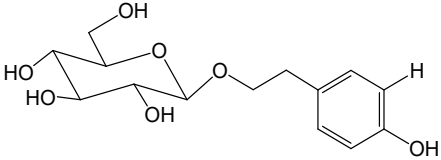
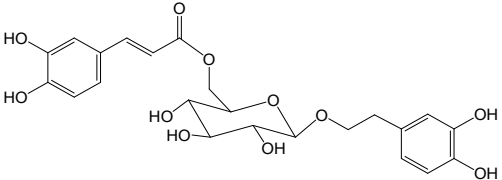
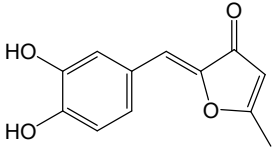
5	Feddeiphenol B	<i>Daphne feddei</i> F. Thymelaeaceae	$R_1 = R_2 = R_3 = \text{OH}$	Activity against HIV-1 EC ₅₀ 3.24 µg/mL CC ₅₀ 132.4 µg/mL TI 40.9	(Hu et al., 2011)
6	Kadangustin J	<i>Daphne feddei</i> F. Thymelaeaceae	$R_1 = R_2 = \text{H}; R_3 = \text{OMe}$	Activity against HIV-1 EC ₅₀ 40.5 µg/mL CC ₅₀ 268.2 µg/mL TI 6.6	(Hu et al., 2011)
7	Feddeiphenol C	<i>Daphne feddei</i> F. Thymelaeaceae		Activity against HIV-1 EC ₅₀ 3.58 µg/mL CC ₅₀ 244.7 µg/mL TI 68.4	(Hu et al., 2011)
					
8	(-)-Surinamensin	<i>Daphne feddei</i> F. Thymelaeaceae	$R = \text{OMe}$	Activity against HIV-1 EC ₅₀ 18.2 µg/mL, CC ₅₀ 186.5 µg/mL TI 10.3	(Hu et al., 2011)
				Activity against <i>Plasmodium falciparum</i>	(Rye and Barker, 2013)
9	Virolin	<i>Daphne feddei</i> F. Thymelaeaceae	$R = \text{H}$	Activity against HIV-1 EC ₅₀ 15.2 µg/mL CC ₅₀ 225.3 µg/mL TI 14.8	(Hu et al., 2011)
				Activity against <i>Leishmania donovani</i> and <i>Plasmodium falciparum</i>	(Rye and Barker, 2013)

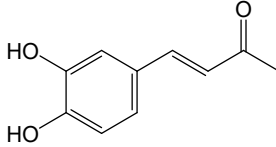
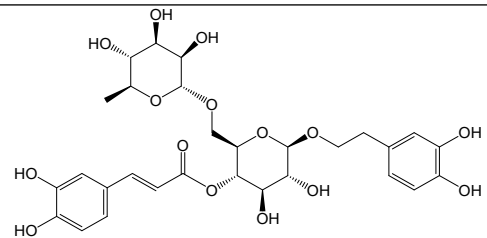
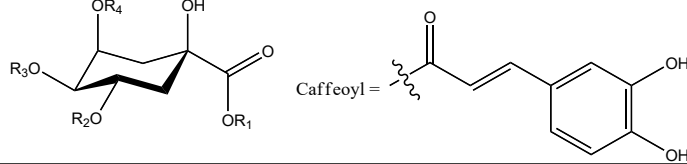
					
10	Rosmarinic acid	<i>Daphne feddei</i> (Thymelaeaceae)	R = OH	Activity against HIV-1 EC ₅₀ 5.82 µg/mL, CC ₅₀ > 300 µg/mL TI > 51.5	(Hu et al., 2011)
		<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV	(Dang et al., 2016);(Tsukamoto et al., 2018); (Liu et al., 2020)
		<i>Rosmarinus officinalis</i> F. Lamiaceae		Activity against DENV	(Panchal et al., 2022)
		<i>Melissa officinalis</i> F. Lamiaceae		Activity against <i>Plasmodium falciparum</i> IC ₅₀ 103.59 µg/ml	(Fordjour et al., 2020)
11	Clinopodic acid A	<i>Daphne feddei</i> F. Thymelaeaceae	R = H	Activity against HIV-1 EC ₅₀ 7.47 µg/mL CC ₅₀ 146.5 µg/mL TI 19.6	(Hu et al., 2011)
12	Ethyl rosmarinate	<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV activity	(Dang et al., 2016)
13	Methyl rosmarinate	<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV	(Dang et al., 2016)
		<i>Salvia plebeia</i> F. Lamiaceae		Activity against H1N1 neuraminidase IC ₅₀ 16.65 ± 0.91 µM	(Omrani et al., 2021)

14	<i>Trans</i> -anethole	<i>Illicium verum</i> F. Illiciaceae <i>Pimpinella anisum</i> F. Apiaceae <i>Foeniculum vulgare</i> F. Apiaceae		Activity against HSV-1 IC ₅₀ 20 ± 1.1 µg/mL	(Astani et al., 2011); (Ilijeva and Buchbauer, 2016)
15	Eugenol (4-allyl-2-methoxyphenol)	<i>Pimenta racemosa</i> F. Myrtaceae, <i>Cinnamomum verum</i> F. Lauraceae & <i>Syzygium aromaticum</i> F. Myrtaceae		Growth inhibition of <i>Trypanosoma cruzi</i> , <i>Giardia lamblia</i> , and <i>Leishmania donovani</i>	(Morais et al., 2014)
16	1'-Acetoxychavicol acetate	<i>Languas galanga</i> (<i>Alipinia galanga</i>) F. Zingiberaceae		Activity against HIV	(Misawa et al., 2008); (Ilijeva and Buchbauer, 2016)
17	Dillapiole	<i>Piper aduncum</i> F. Piperaceae		Inhibitory activity against <i>Leishmania amazonensis</i> and <i>L. brasiliensis</i> IC ₅₀ 69.3 and 59.4 µM, respectively	(Parise-Filho et al., 2012)
18	Dihydrodillapiole	<i>Piper aduncum</i> F. Piperaceae		Inhibitory activity against <i>Leishmania amazonensis</i> and <i>L. brasiliensis</i> IC ₅₀ 99.9 µM for IC ₅₀ 90.5 µM for	(Parise-Filho et al., 2012)
19	<i>p</i> -Coumaric acid	<i>Stereospermum acuminatissimum</i> F. Bignoniaceae		In-vitro and in-vivo activities against <i>Leishmaniasis amazonensis</i> with	(Monzote et al., 2016)

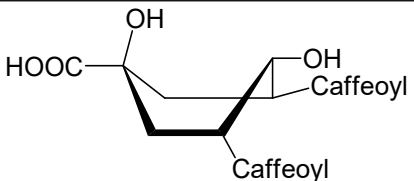
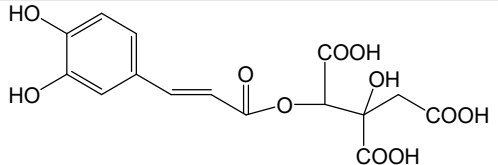
				$IC_{50} \pm SD$ $1.5 \pm 0.5 - 9.0 \pm 1.7$ $\mu\text{g/mL}-\mu\text{M/L}$ for amastigotes and $CC50 \pm SD$ $22.5 \pm 0.7 - 136.8 \pm 0.4$ $\mu\text{g/mL}-\mu\text{M/L}$ for macrophages.	
20	Verbascoside (Acteoside)	<i>Markhamia lutea</i> F. Bignoniaceae	$R_1 = \text{H}; R_2 = \text{Caf}; R_3 = \text{Rha}; R_4 = \text{H}$	Activity against RSV EC_{50} 0.80 $\mu\text{g/mL}$ IC_{50} 76.9 $\mu\text{g/mL}$	(Kernan et al., 1998);(Perez G., 2003); (Jansi et al., 2021)
		<i>Scrophularia scorodonia</i> Scrophulariaceae		Activity against VSV Inhibition percentage of cytopathic effect of 53.6% at 500 $\mu\text{g/mL}$	(Bermejo et al., 2002)
		<i>Tecoma mollis</i> F. Bignoniaceae		In vitro activity against <i>Leishmania donovani</i> with percentage of inhibition 83% (IC_{50} 30.08)	(Abdel-Mageed et al., 2012)
21	Isoverbascoside (Isoacteoside)	<i>Markhamia lutea</i> F. Bignoniaceae	$R_1 = \text{Caf}; R_2 = \text{H}; R_3 = \text{Rha}; R_4 = \text{H}$	Activity against RSV EC_{50} 0.62 $\mu\text{g/mL}$ IC_{50} 51.4 $\mu\text{g/mL}$	(Kernan et al., 1998);(Perez G., 2003);(Jansi et al., 2021)
		<i>Tecoma mollis</i> F. Bignoniaceae		In vitro activity against <i>Leishmania donovani</i> with percentage of inhibition 78% ($IC_{50} > 40$)	(Abdel-Mageed et al., 2012)

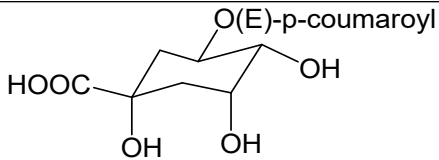
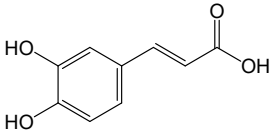
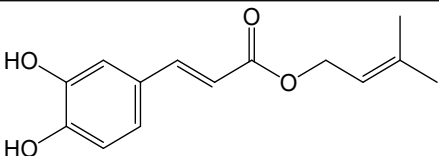
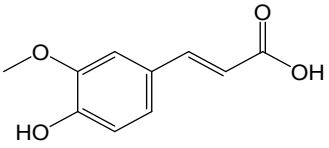
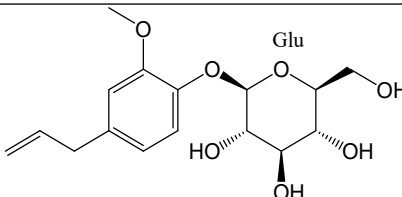
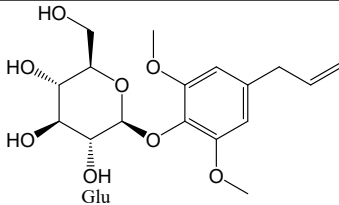
22	Echinacoside	<i>Echinacea purpurea</i> F. Asteraceae	R ₁ = Glu; R ₂ = Caf; R ₃ = Rha; R ₄ = H	Activity against VSV	
23	Luteoside A	<i>Markhamia lutea</i> F. Bignoniaceae	R ₁ = Ac; R ₂ = Caf; R ₃ = Rha; R ₄ = Api	Activity against RSV EC ₅₀ 0.87 µg/mL IC ₅₀ 77 µg/mL	(Kernan et al., 1998);(Perez G., 2003); (Jansi et al., 2021)
		<i>Tecoma mollis</i> F. Bignoniaceae		Moderate antimalarial activity (45% inhibition) against chloroquine sensitive (D6) clones of <i>Plasmodium falciparum</i>	(Abdel-Mageed et al., 2012)
24	Luteoside B	<i>Markhamia lutea</i> F. Bignoniaceae	R ₁ = Caf; R ₂ = H; R ₃ = Rha; R ₄ = Api	Activity against RSV EC ₅₀ 3.4 µg/mL IC ₅₀ > 67µg/mL	(Kernan et al., 1998);(Perez G., 2003); (Jansi et al., 2021)
		<i>Tecoma mollis</i> F. Bignoniaceae		High antileishmanial activity against <i>Leishmania donovani</i> with percentage of inhibition 92% (IC ₅₀ 6.71)	(Abdel-Mageed et al., 2012)
25	Luteoside C	<i>Markhamia lutea</i> F. Bignoniaceae	R ₁ = Fer; R ₂ = H; R ₃ = Rha; R ₄ = Api	Activity against RSV EC ₅₀ 15.5 µg/mL IC ₅₀ 189 µg/mL	(Kernan et al., 1998); (Perez G., 2003)

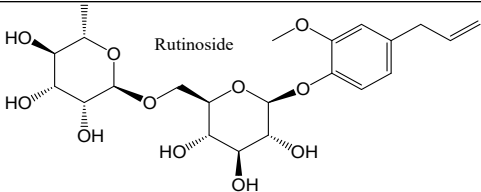
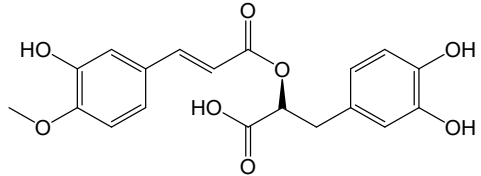
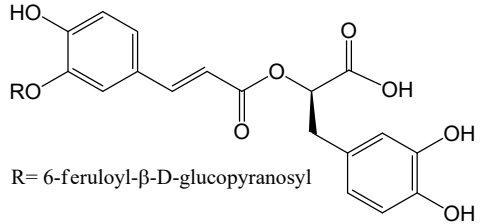
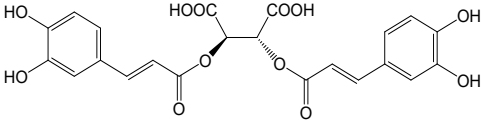
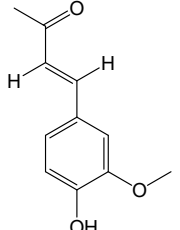
26	Salidroside	<i>Phillyrea latifolia</i> F. Oleaceae		<p>Growth inhibitory effect against the promastigotes of Sodium stibogluconate (SSG) sensitive and resistant strains of <i>L. donovani</i>. IC₅₀ of 15.54 ± 1.53 and 38.16 ± 0.86 µg/mL, respectively.</p>	(Chauhan et al., 2019)
27	Calceolarioside B	<i>Digitalis purpurea</i> F. Scrophulariaceae		<p><i>In vitro</i> antileishmanial activity (IC₅₀ = 20 µg/mL). Its <i>in vivo</i> efficacy was noted at 20 mg/kg body weight when it reduced the hepatic and splenic parasite burden by 79 and 84 %, respectively, in an established model of <i>L. donovani</i> Ag83 infected golden hamster. Furthermore, synergistic potentiations of it at 20 mg/kg body weight and SAG at 5 mg/kg body weight showed a significant reduction of hepatic and splenic parasite burden.</p>	(Poddar et al., 2008)
28	Inotilone	<i>Phellinus linteus</i> F. Hymenochaetaceae		<p>Inhibited H1N1 neuraminidase activity with IC₅₀ value of 29.1 µM and active against IAV/WS/33</p>	(Hwang et al., 2018); (Chen et al., 2019); (Omrani et al., 2021)

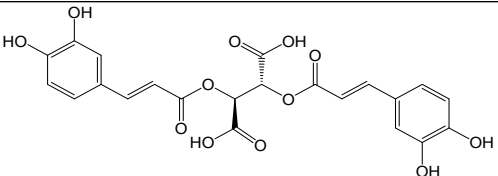
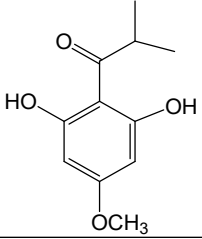
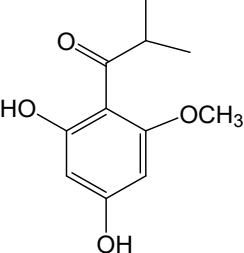
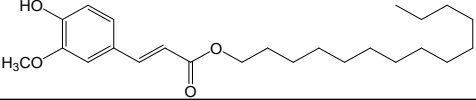
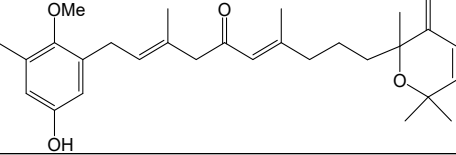
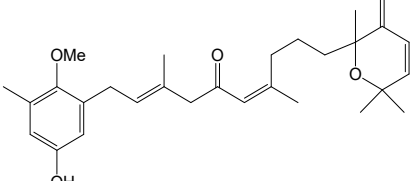
				virus	
29	4-(3,4-Dihydroxyphenyl)-3-buten-2-one	<i>Phellinus linteus</i> F. Hymenochaetaceae		Inhibited H1N1 neuraminidase activity with IC ₅₀ value of 125.6 μM and active against IAV/WS/33 virus	(Hwang et al., 2018); (Chen et al., 2019); (Omrani et al., 2021)
30	Forsythoside A	<i>Forsythia suspensa</i> F. Oleaceae		Activity against IAV	(Omrani et al., 2021)
					
31	Chlorogenic acid (3- <i>O</i> -Caffeoyl-quinic acid)	<i>Coffea arabica</i> F. Rubiaceae	$R_1 = R_3 = R_4 = H, R_2 = \text{Caffeoyl}$	Activity against HBV	(Wu, 2016)
				Activity against poliovirus	(Perez G., 2003); (Jansi et al., 2021)
				Activity against HBV DNA replication IC ₅₀ 5.5 μM (SI >115.0)	(Zhao et al., 2014)
	<i>Artemisia capillaris</i> F. Asteraceae			It inhibits the secretion of HBsAg, HBeAg, and the replication of HBV-DNA on Hep G 2.2.15 cells at the concentration of 100 μg/mL.	(Liu et al., 2020)
	<i>Lonicera japonica</i> F. Caprifoliaceae				

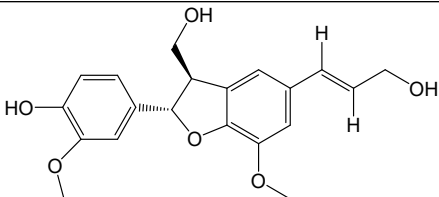
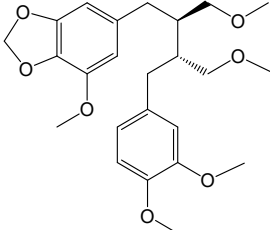
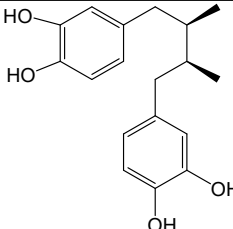
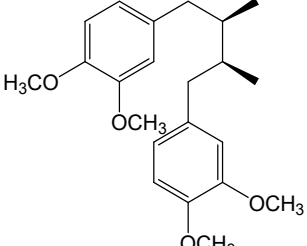
				Moderate activity against <i>Plasmodium falciparum</i> IC ₅₀ 105 µg/ml	(Fordjour et al., 2020)
32	Chlorogenic acid methyl ester	<i>Artemisia capillaris</i> F. Asteraceae	R ₁ = Me; R ₂ = Caffeoyl; R ₃ = R ₄ = H	Activity against HBV DNA replication IC ₅₀ 272.3 µM (SI > 6.2)	(Zhao et al., 2014)
33	Cryptochlorogenic acid	<i>Lonicera japonica</i> F. Caprifoliaceae	R ₁ = R ₂ =R ₄ = H, R ₃ = Caffeoyl	Anti-HBV activity	(Liu et al., 2020)
		<i>Artemisia capillaris</i> F. Asteraceae		Potent activity against HBV DNA replication IC ₅₀ 13.7 µM (SI > 115.0)	
34	Cryptochlorogenic acid methyl ester	<i>Artemisia capillaris</i> F. Asteraceae	R ₁ = Me; R ₂ =R ₄ = H; R ₃ = Caffeoyl	Activity against HBV DNA replication IC ₅₀ 175.3 µM (SI > 11.2)	(Zhao et al., 2014)
35	Neochlorogenic acid	<i>Lonicera japonica</i> F. Caprifoliaceae	R ₁ = R ₂ =R ₃ = H; R ₄ = Caffeoyl	Activity against HBV	(Liu et al., 2020)
		<i>Artemisia capillaris</i> F. Asteraceae		Activity against HBV DNA replication IC ₅₀ 7.3 µM (SI > 249.9)	(Zhao et al., 2014)
36	Neochlorogenic acid methyl ester	<i>Artemisia capillaris</i> F. Asteraceae	R ₁ = Me; R ₂ =R ₃ = H; R ₄ = Caffeoyl	Activity against HBV DNA replication IC ₅₀ 144.7 µM (SI > 20.2)	(Zhao et al., 2014)
37	3,5-Di-O-caffeoylquinic acid	<i>Lactuca indica</i> F. Asteraceae	R ₁ = R ₃ = H; R ₂ =R ₄ = Caffeoyl	Anti-HBV activity	(Wu, 2016)
		<i>Lonicera japonica</i> F. Caprifoliaceae		Activity against the secretions of HBsAg and HBeAg	(Liu et al., 2020)
		<i>Artemisia capillaris</i> F. Asteraceae		Inhibited HBV DNA replication IC ₅₀ 6.4 µM (SI > 256.1). Activity against the secretions of HBsAg and HBeAg	(Zhao et al., 2014)

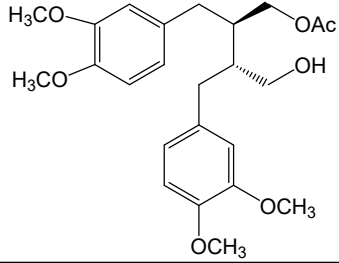
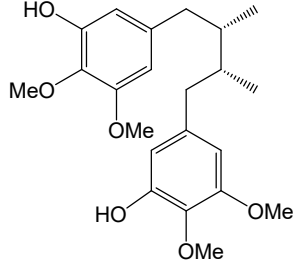
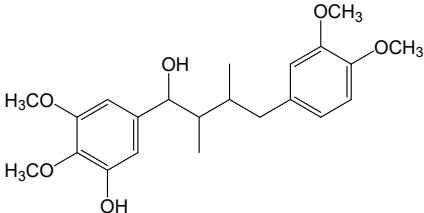
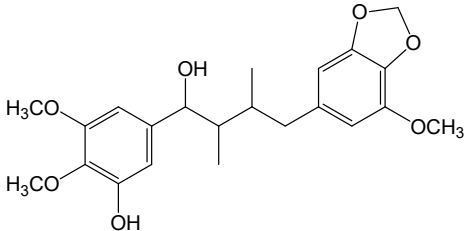
38	4,5- <i>O</i> -Dicaffeoylquinic acid	<i>Lonicera japonica</i> F. Caprifoliaceae	$R_1 = R_2 = H; R_3 = R_4 = \text{Caffeoyl}$	Activity against HBV and secretions of HBsAg and HBeAg	(Liu et al., 2020)
		<i>Artemisia capillaris</i> F. Asteraceae		Inhibited HBV DNA replication IC_{50} 9.8 μM (SI > 184.8). Moderate activity against the secretions of HBsAg and HBeAg	(Zhao et al., 2014)
39	3,4-Di- <i>O</i> -caffeoylquinic acid	<i>Lactuca indica</i> F. Asteraceae	$R_1 = R_4 = H; R_2 = R_3 = \text{Caffeoyl}$	Anti-HBV activity	(Wu, 2016)
		<i>Lonicera japonica</i> F. Caprifoliaceae		Potent activity against HBV-DNA replication IC_{50} 5:5±0:9-13:7±1:3 μM	(Liu et al., 2020)
		<i>Artemisia capillaris</i> F. Asteraceae		inhibited HBV DNA replication IC_{50} 6.1 μM (SI > 184.8). Activity against the secretions of HBsAg and HBeAg	(Zhao et al., 2014)
40	3,4,5-Tri- <i>O</i> -caffeoylquinic acid	<i>Securidata longipedunculata</i> F. Polygalaceae	$R_1 = H; R_2 = R_3 = R_4 = \text{Caffeoyl}$	Anti- HIV activity	(Perez G., 2003)
41	3,5-Di- <i>O</i> -caffeoyl-mucoquinic acid	<i>Lactuca indica</i> F. Asteraceae		Activity against HBV	(Wu, 2016)
42	2- <i>O</i> -Caffeoyl-(+)-allohydroxy-citric acid	<i>Spondias mombin</i> F. Anacardiaceae		Activity against Coxsackie and HSV	(Perez G., 2003); (Jansi et al., 2021)

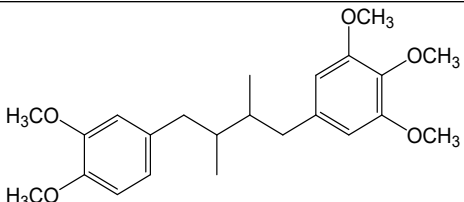
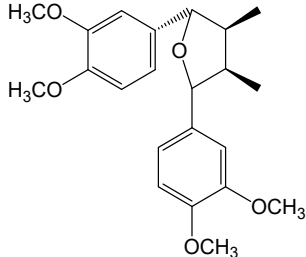
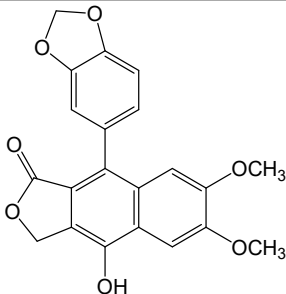
43	5- <i>O</i> -(<i>E</i>)- <i>p</i> -Coumaroylquinic acid	<i>Lactuca indica</i> F. Asteraceae		Anti-HBV activity	(Wu, 2016)
44	Caffeic acid	<i>Coffea arabica</i> F. Rubiaceae		Activity against HBV	(Wu, 2016)
		<i>Salvia prattii</i> F. Lamiaceae		Activity against HSV, influenza, vaccinia, and polio viruses	(Jansi et al., 2021); (Perez G., 2003)
45	3-Methyl-but-2-enyl caffeate	<i>Populus nigra</i> F. Salicaceae		Activity against HSV-1	(Jansi et al., 2021); (Perez G., 2003)
46	Ferulic acid	<i>Eleutherococcus senticosus</i> F. Araliaceae; <i>Populus balsamifera</i> F. Salicaceae		Moderate activity against <i>Plasmodium falciparum</i> IC ₅₀ 93.36 µg/ml	(Fordjour et al., 2020)
47	Eugenyl- <i>O</i> -β-D-glucoside	<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV	(Dang et al., 2016)
48	Erigeside II	<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV	(Dang et al., 2016)

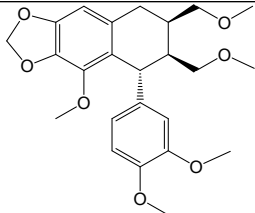
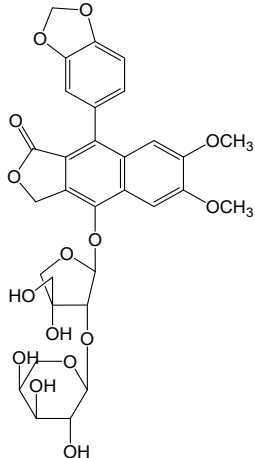
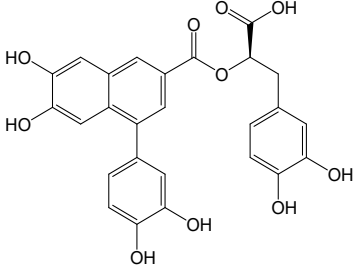
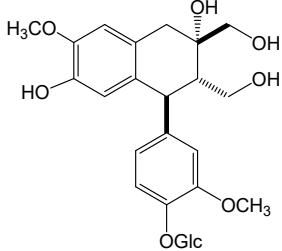
49	Eugenyl- β -rutinoside	<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV	(Dang et al., 2016)
50	Shimobashiric acid B	<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV	(Dang et al., 2016)
51	3-(6-Feruloyl- β -D-glucopyranosyl)-rosmarinic acid	<i>Salvia prattii</i> F. Lamiaceae		Activity against HBV	(Dang et al., 2016)
52	L-Chicoric acid (also known as cichoric acid; dicaffeoyltartaric acid)	<i>Chicorium intybus</i> F. Asteraceae <i>Echinacea purpurea</i> F. Asteraceae		Activity against HIV-1	(Pluymers et al., 2001);(Lee and Scagel, 2013); (Liu et al., 2020)
		<i>Echinacea pallida</i> F. Asteraceae		Anti-VSV activity	(Pan et al., 2003)
53	Dehydrozingerone	<i>Swertia patens</i> F. Gentianaceae		Activity against HBV	(Liu et al., 2020)

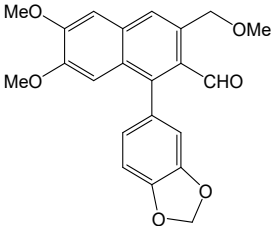
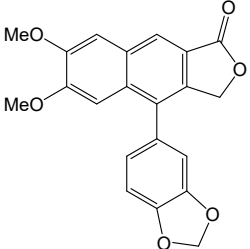
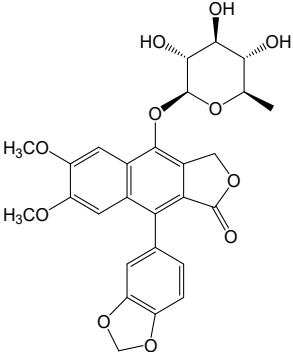
54	Di- <i>E</i> -caffeoyl- <i>meso</i> -tartaric acid	<i>Lactuca indica</i> F. Asteraceae		Activity against HBV	(Kim et al., 2010)
55	2,6-Dihydroxy-methoxyisobutylrophenone	<i>Kunzea sinclairii</i> F. Myrtaceae		Activity against HSV-1 and PV-1	(Bloor, 1992)
56	4,6-Dihydroxymethoxyisobutylrophenone	<i>Kunzea sinclairii</i> F. Myrtaceae		Activity against HSV-1 and PV-1	(Bloor, 1992)
57	Tetradecyl ferulate	<i>Ocimum sanctum</i> F. Lamiaceae		Anti-HIV-1 activity	(Neelam et al., 2020)
58	Usneoidone E	<i>Cystoseira usneoides</i> F. Phaeophyceae		Antiviral activity	(Jansi et al., 2021); (Perez G., 2003)
59	Usneoidone Z	<i>Cystoseira usneoides</i> F. Phaeophyceae		Antiviral activity	(Jansi et al., 2021); (Perez G., 2003)

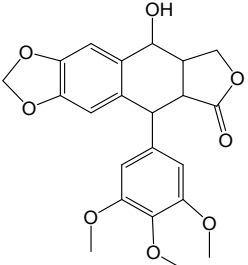
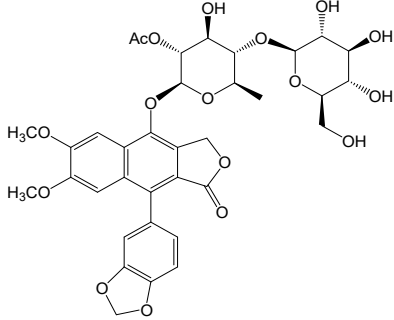
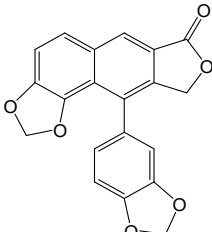
60	(+)-Dehydrodiconiferyl alcohol	<i>Swertia patens</i> F. Gentianaceae		Activity against HBV	(Liu et al., 2020)
61	Niranthin	<i>Phyllanthus niruri</i> F. Euphorbiaceae		Inhibited HBV antigen secretion in HepG2.2.15 cells IC ₅₀ 15.6 µmol/L for HBsAg IC ₅₀ 25.1 µmol/L for HBeAg	(Wu, 2016); (Cui et al., 2020); (Liu et al., 2020)
		<i>Phyllanthus amarus</i> F. Euphorbiaceae		Potent activity against <i>Leishmania donovani</i>	(Chowdhury et al., 2012)
62	Nordihydroguaiaretic Acid (NDGA)	<i>Larrea tridentata</i> F. Zygophyllaceae		Anti- DENV, HCV, WNV and ZIKV, IAV, suppress the replication of IAV	(Cui et al., 2020); (Xu et al., 2022)
				Active against <i>Trypanosoma brucei rhodesiense</i> , <i>T. cruzi</i> , <i>Leishmania donovani</i> and <i>Plasmodium falciparum</i> IC ₅₀ 4.5, 33.1, 12.0 and 7.7 µM, respectively	(Schmidt et al., 2012)
63	Tetra- <i>O</i> -methyl nordihydroguaiaretic acid (TMP)	<i>Larrea tridentata</i> F. Zygophyllaceae		It inhibited the infection of WNV and ZIKA, Poxvirus growth and it has antiviral activity against HSV and HIV.	(Cui et al., 2020)

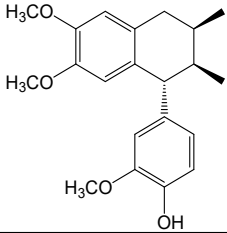
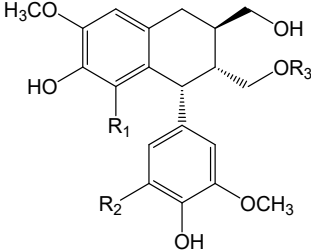
		<i>Schisandra propinqua</i> F. Schisandraceae		Anti-HIV-1 with EC ₅₀ 14.8 μM	(Xu et al., 2022)
64	Secoisolariciresinol dimethyl ether acetate	<i>Justicia patentiflora</i> F. Acanthaceae		Anti-HIV-1 activity with an IC ₅₀ value of 5.27 μmol/L and SI value of 2.2	(Xu et al., 2019)
65	3,3'-Dihydroxy-4,4',5,5'-tetramethoxylignan	<i>Schisandra rubriflora</i> F. Schisandraceae		Anti- HIV-1IIB induced syncytium formation with an EC ₅₀ 5.8 μM TI 4.46	(Xu et al., 2022)
66	Kadangustin H	<i>Kadsura angustifolia</i> F. Schisandraceae		Anti-HIV-1 EC ₅₀ 27.0 μM	(Gao et al., 2008); (Xu et al., 2022)
67	Kadangustin I	<i>Kadsura angustifolia</i> F. Schisandraceae		Anti-HIV-1 EC ₅₀ 21.5 μM	(Gao et al., 2008); (Xu et al., 2022)

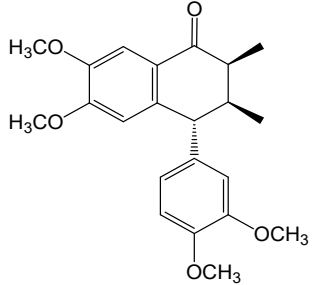
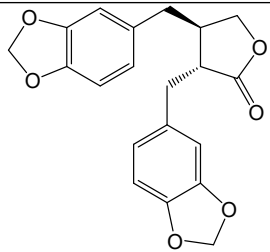
68	Tiegusanin N	<i>Schisandra propinqua</i> F. Schisandraceae		Anti-HIV-1 EC ₅₀ 9.4 μM	(Li et al., 2009); (Xu et al., 2022)
69	Ganschisdandrine	<i>Schisandra sphenanthera</i> F. Schisandraceae		Active against HSV-2 and adenovirus (SI = 16.93)	(Song et al., 2013)
70	Diphyllin	<i>Justicia patentiflora</i> F. Acanthaceae		Active against VSV, influenza virus and HIV-1 IC ₅₀ 0.38 μmol/L (SI = 5.3)	(Xu et al., 2019) (Xu et al., 2022)
		<i>Haplophyllum alberti-regelii</i> and <i>H. perforatum</i> F. Rutaceae		It has broad-spectrum antiviral activity e.g. it blocked ZIKAV infection in HT1080 cells IC ₅₀ ~0.06 μM	(Cui et al., 2020)
		<i>Haplophyllum bucharicum</i> F. Rutaceae		Antiviral activity with MIC 0.66 μM against VSV and v-ATPase blocker against influenza virus with IC ₅₀ 0.04–0.49 μM	(Xu et al., 2022)
				In vitro activity against <i>Leishmania infantum</i> activity IC ₅₀ = 14.4 μM	(Di Giorgio et al., 2005)

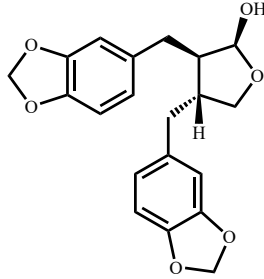
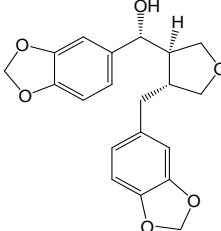
71	Nirtetralin B	<i>Phyllanthus niruri</i> F. Euphorbiaceae		Inhibited HBV antigen secretion in HepG2.2.15 cells with IC ₅₀ 17.4 and 63.9 μmol/L for HBsAg and HBeAg, respectively	(Wu, 2016)
72	Procumbenoside A	<i>Justicia patentiflora</i> F. Acanthaceae		Inhibitory activity against HIV-1 with IC ₅₀ values of 4.95 0.38 μmol/L (SI > 6.2)	(Xu et al., 2019)
73	Globoidnan A	<i>Eucalyptus globoidea</i> F. Myrtaceae		Active against HIV integrase EC ₅₀ 0.64 μM	(Xu et al., 2022)
74	(+)-Cycloolivil-4'-O-β-D-glucopyranoside	<i>Swertia chirayita</i> F. Gentianaceae		Anti-HBV	(Liu et al., 2020)
				Inhibitory effect on the HBsAg and HBeAg secretion IC ₅₀ 0.31 ± 0.045 mmol/L (SI = 4.29) and 0.77 ± 0.076 mmol/L	(Wu, 2016)

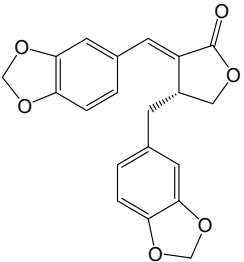
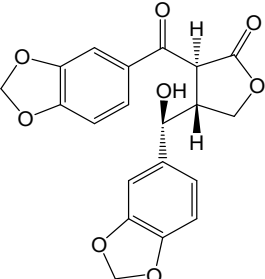
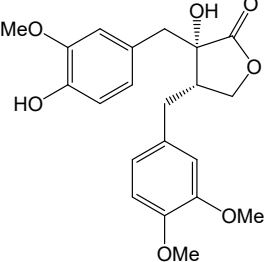
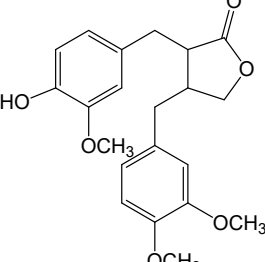
				(SI = 1.75), respectively. Also, it inhibits the replication of HBV DNA IC ₅₀ 0.29 ± 0.034 mmol/L (SI = 4.66)	
75	Phyllamycin B	<i>Phyllanthus myrtifolius</i> F. Euphorbiaceae		Active against HIV-1 EC ₅₀ 3.5 μM	(Xu et al., 2022)
76	Retrojusticidin B	<i>Phyllanthus myrtifolius</i> F. Euphorbiaceae		Active against HIV-1 EC ₅₀ 5.5 μM	(Xu et al., 2022)
77	6-Deoxyglucose-diphyllin (DGP) also known as Patentiflorin A	<i>Justicia gendarussa</i> F. Acanthaceae		Potently inhibits a broad spectrum of HIV-1 strains including some resistant strains EC ₅₀ 15–37 nM (AZT: 77–95 nM) and mosquito-borne flavivirus such as ZIKV, DENV1, TBEV, WNV, JEV and EBV EC ₅₀ 0.12–1.0 nM	(Zhang et al., 2017b); (Cui et al., 2020); (Xu et al., 2022)

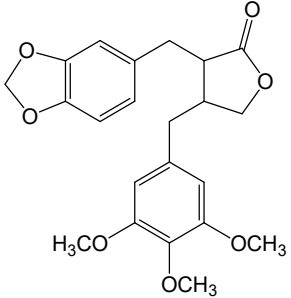
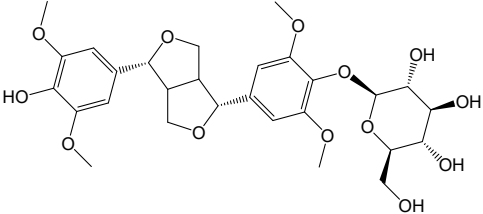
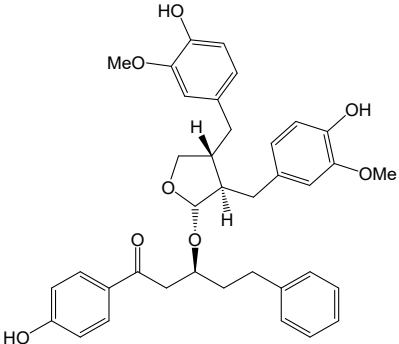
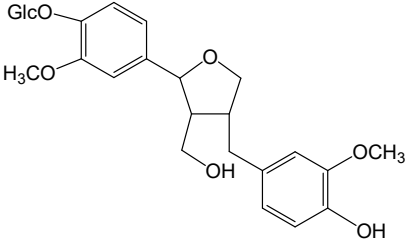
78	Podophyllotoxin	<i>Podophyllum versipelle</i> & <i>P. peltatum</i> F. Berberidaceae		It is used as topical treatment for venereal warts (<i>Condyloma acuminatum</i>), which is caused by <i>Papilloma</i> virus.	(Cui et al., 2020)
79	Justiprocumin B	<i>Justicia gendarussa</i> F. Acanthaceae		<p>Potent activity against a broad spectrum of HIV strains IC_{50} 15-21 nM (AZT, IC_{50} 77-95 nM). Also, it displayed potent inhibitory activity against the NRTI (nucleoside reverse transcriptase inhibitor)-resistant isolate (HIV-1₁₆₁₇₋₁) of the analogue (AZT) as well as the NNRTI (non-nucleoside reverse transcriptase inhibitor)-resistant isolate (HIV-1_{N119}) of the analogue (nevaripine)</p>	(Zhang et al., 2017a)
80	Helioxanthin (HE-145)	<i>Taiwania cryptomerioides</i> F. Taxodiaceae		Inhibits the gene expression and replication of HBV in HCC cells	(Xu et al., 2022) (Wu, 2016)

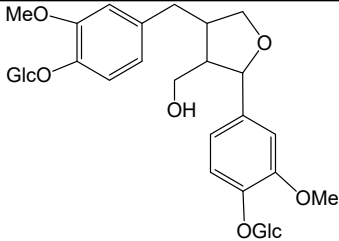
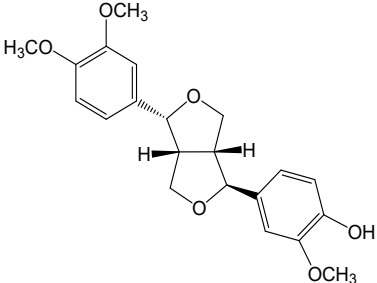
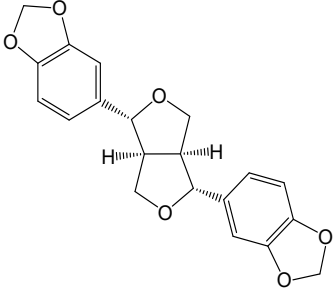
				Active against HBV EC ₅₀ 1 μM	(Xu et al., 2022)
81	Sinensisin A	<i>Schisandra propinqua</i> F. Schisandraceae		Active against HIV-1 EC ₅₀ 4.5 μM TI 6.7	(Xu et al., 2022)
					
82	(-)-Isolariciresinol	<i>Streblus asper</i> F. Moraceae	R ₁ = R ₂ = R ₃ = H	Active against HBV IC ₅₀ 3.67 and 14.67 μM, for HBsAg and HBeAg, respectively with no cytotoxicity	(Li et al., 2012) ;(Wu, 2016) ;(Liu et al., 2020); (Xu et al., 2022)
83	9-β- Xylopyranosyl(-)- isolariciresinol	<i>Streblus asper</i> F. Moraceae	R ₁ = R ₂ = H; R ₃ = Xyl	Anti-HBsAg EC ₅₀ 6.98 μM, anti-HBeAg EC ₅₀ 26.74 μM. It inhibited the expression of HBsAg and HBeAg with EC ₅₀ values of 6.58 μM for HBsAg (TI >2) and 24.86 μM (TI = 8.06) for HBeAg	(Wu, 2016) (Xu et al., 2022)

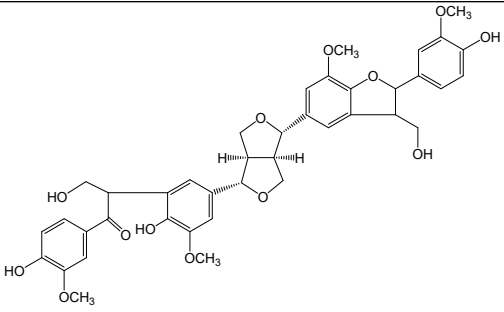
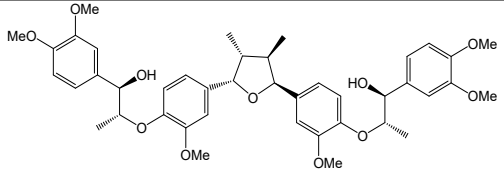
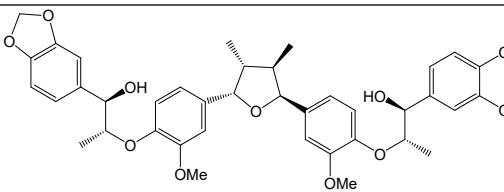
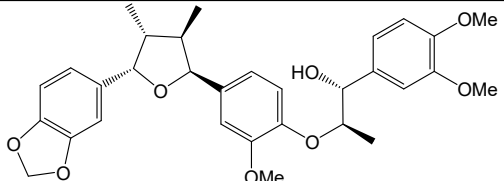
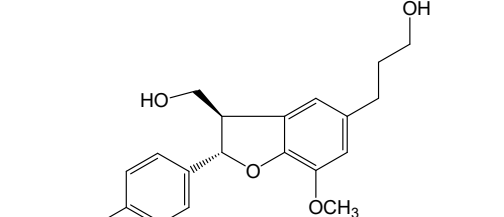
84	5-methoxy-9- β -xylopyranosyl(-)-isolariciresinol	<i>Streblus asper</i> F. Moraceae	$R_1 = H$; $R_2 = OCH_3$; $R_3 = Xyl$	It inhibited the expression of HBsAg and HBeAg with EC_{50} values of 39.56 μ M for HBsAg (TI >2) and 61.23 μ M (TI = 1.75) for HBeAg	(Xu et al., 2022)
85	(-)-8 ϵ -aristoligone (7'R,8S,8'S)-3',4,4',5-Tetramethoxy-2,7'-cyclo lignan-7-one	<i>Schisandra sphenanthera</i> F. Schisandraceae		It has moderate inhibition of HSV-2 and adenovirus	(Song et al., 2013)
		<i>Holostylis reniformis</i> (Aristolochiaceae)		Activity against <i>Plasmodium falciparum</i> parasites (isolate BHZ 26/86; chloroquine IC_{50} (μ M) 0.63 ± 0.20 C_{90} (μ M) 2.61 ± 0.06)	(Andrade-neto et al., 2007)
86	Hinokinin	<i>Chamecyparis obtuse</i> F. Cupressaceae, <i>Phyllanthus</i> spp. F. Eupobiaceae, <i>Aristolochia</i> F. Aristolochiaceae, <i>Piper</i> spp. F. Piperaceae, <i>Viola</i> spp. F. Myristicaceae F. Linaceae		Antiviral Anti-HBV, HIV, SARS-virus and (HCMV). Anti-SARS-CoV replication with $EC_{50} > 10 \mu$ M	(Cui et al., 2020) (Xu et al., 2022)
		<i>Piper cubeba</i> F. Piperaceae		Activity against <i>Trypanosoma cruzi</i>	(Esperandim et al., 2013)
		In vitro anthelmintic activity against gastrointestinal nematodes in sheep EC_{50} 68.38 μ g/mL for ovicidal activity		(Carlis et al., 2019)	

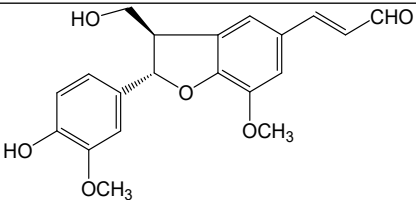
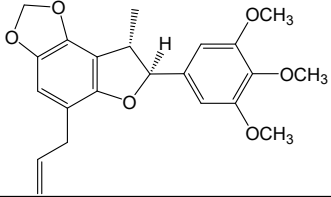
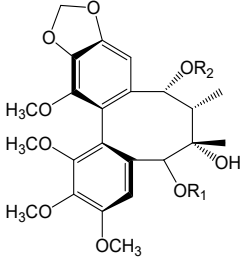
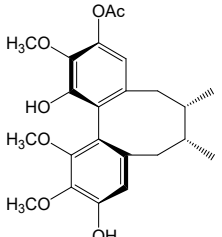
				In the LDT, it inhibited 100% the larval development at all concentrations EC ₅₀ 0.89 µg/mL for L3 migration inhibition test (LMT)	
87	Cubebin	<i>Piper cubeba</i> F. Piperaceae		Activity against <i>Trypanosoma cruzi</i>	(Esperandim et al., 2013)
				In vitro anthelmintic activity against gastrointestinal nematodes in sheep EC ₅₀ 150.00 µg/mL for ovicidal activity EC ₅₀ of 14.89 µg/ mL for larval development test (LDT) EC ₅₀ 0.34 µg/mL for L3 migration inhibition test (LMT)	(Carlis et al., 2019)
88	Isocubebin	<i>Daphne feddei</i> F. Thymelaeaceae		Activity against HIV-1 EC ₅₀ 8.27 µg/ mL CC ₅₀ 173.4 µg/mL TI 21.0	(Hu et al., 2011)

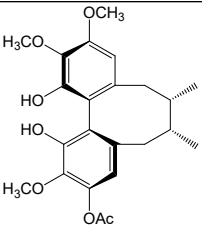
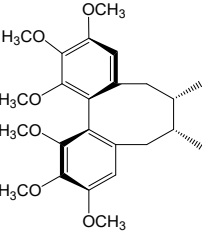
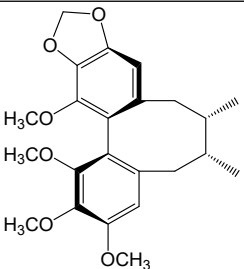
89	Savinin	<i>Chamaecyparis obtusa</i> F. Cupressaceae		Active against SARS-CoV replication EC ₅₀ 1.13 μM	(Xu et al., 2022)
90	Sanguinolignan A	<i>Piper sanguineispicum</i> F. Piperaceae		Active against axenic amastigote forms of <i>Leishmania amazonensis</i> IC ₅₀ 36.7±3.08 μg cm ⁻³	(Teponno et al., 2016)
91	Trachelogenin	<i>Trachelospermum jasminoides</i> F. Apocynaceae		Anti-HCV EC ₅₀ 0.87 μM and 0.69 μM for HCVcc model and HCVpp model, respectively	(Xu et al., 2022)
92	Arctigenin (ATG)	<i>Arctium lappa</i> F. Asteraceae		It induces the production of interferon and inhibits the expression of protein P17 and P24 of the HIV-1 virus	(Cui et al., 2020)

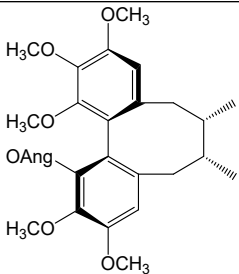
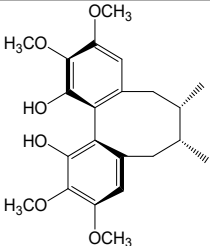
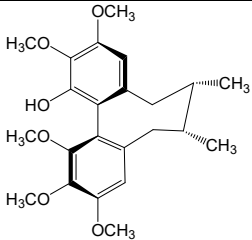
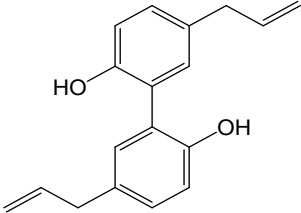
93	Yatein	<i>Chamaecyparis obtusa</i> F. Cupressaceae		It inhibits HSV-1 alpha gene expression, including expression of the ICP0 and ICP4 genes, and by arresting HSV-1 DNA synthesis and structural protein expression in HeLa cells	(Cui et al., 2020)
94	Syringaresinol 4''-O-β-D-glucopyranoside	<i>Swertia chirayita</i> F. Gentianaceae		Active against HBV	(Liu et al., 2020)
95	Daphnenin	<i>Daphne acutiloba</i> F. Thymelaeaceae		Active against HIV EC ₅₀ 0.64 μM	(Xu et al., 2022)
96	Lariciresinol-4-O-β-D-glucopyranoside	<i>Isatis indigotica</i> F. Cruciferae		It inhibits the IAV-induced pro-inflammatory response	(Cui et al., 2020)

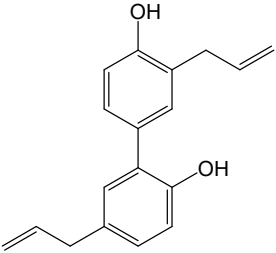
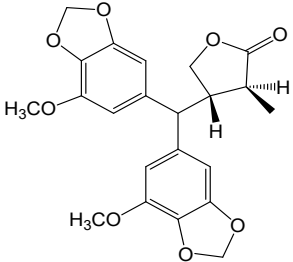
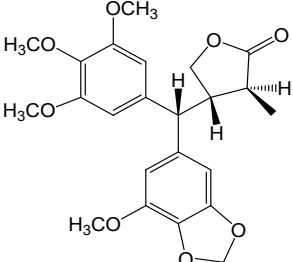
97	Clemastanin B (7S,8R,8'R-(-)-lariciresinol-4,4' -bis-O-β-D-glucopyranoside)	<i>Isatis indigotica</i> F. Cruciferae		Active against subtypes of human IAVs (H1N1, H3N2, and influenza B)	(Cui et al., 2020)
98	Phillygenin	<i>Forsythia suspensa</i> F. Oleaceae		Good protector against influenza A virus (IAV)	(Cui et al., 2020)
99	Sesamin	<i>Sesamum indicum</i> F. Pedaliaceae		It inhibits influenza type A H1N1 infection <i>via</i> reduced IL-1β and TNF-α levels leading to inhibition of the cytokine storm and increased IL-2 levels leading to a stimulation of immune response	(Fanhchaksai et al., 2016); (Cui et al., 2020)
		<i>Artemisia gorgonum</i> F. Asteraceae			Active against chloroquine-resistant <i>Plasmodium falciparum</i> (FcB1 strain) IC ₅₀ 3.37 μg/ml

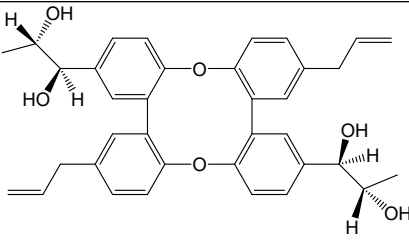
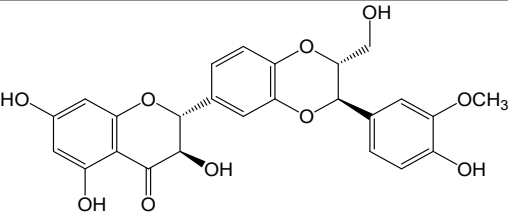
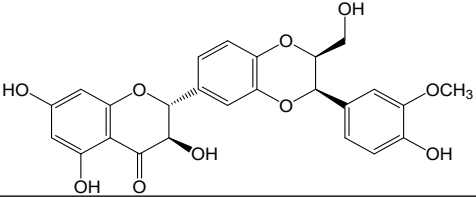
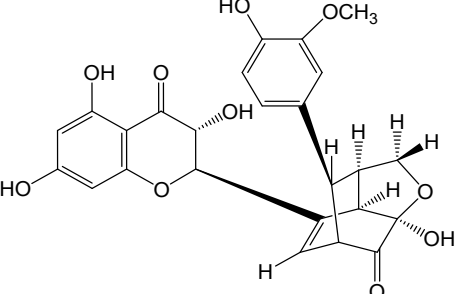
100	Herpetetrone	<i>Herpetospermum caudigerum</i> F. Cucurbitaceae		Inhibitory effects on HBsAg secretion with IC ₅₀ values of 4.89 μmol/L, and on HBeAg secretion with IC ₅₀ values of 8.02 μmol/L	(Wu, 2016)
101	Manassantin A	<i>Saururus chinensis</i> F. Saururaceae		HIV-1-induced cytopathic effects with EC ₁₀₀ 1.0 μM. It inhibits EBV lytic DNA replication in P3HR-1 cells with EC ₅₀ 3.42 μM	(Xu et al., 2022)
102	Manassantin B	<i>Saururus chinensis</i> F. Saururaceae		HIV-1-induced cytopathic effects with EC ₁₀₀ 1.0 μM. It inhibits EBV lytic DNA replication in P3HR-1 cells with EC ₅₀ 1.72 μM	(Xu et al., 2022)
103	Saucerneol D	<i>Saururus chinensis</i> F. Saururaceae		It inhibits EBV lytic DNA replication in P3HR-1 cells with EC ₅₀ 1.09 μM	(Xu et al., 2022)
104	Vladinol F	<i>Daphne feddei</i> F. Thymelaeaceae		Anti-HIV-1 activity with EC ₅₀ 3.28 μg/mL, CC ₅₀ 102.5 μg/mL and TI 31.3	(Hu et al., 2011)
		<i>Schisandra micrantha</i> F. Schisandraceae		Anti-HIV-1 activity with an EC ₅₀ value of 9.75 μM and TI value of 27.45	(Xu et al., 2022)

		<i>Drybalanops oblongifolia</i> F. Dipterocarpaceae		Active against <i>Plasmodium falciparum</i> 3D7 IC ₅₀ 3.51 µg/mL.	(Indriani et al., 2021)
105	Balaphonin	<i>Schisandra lancifolia</i> F. Schisandraceae		Anti-HIV-1 effects with an EC ₅₀ value of 8.43 µM and TI value of 4.3	(Xu et al., 2022)
106	Ococymosin	<i>Ocotea cymosa</i> F. Lauraceae		Antiparasitic activity against the Dd2 strain of <i>Plasmodium falciparum</i> IC ₅₀ 0.45 µM	(Teponno et al., 2016)
107	Tiegusanin G	<i>Schisandra propinqua</i> F. Schisandraceae	 R ₁ = Ang, R ₂ = t-Cin	Anti-HIV-1 activity with an EC ₅₀ value of 8 µM and TI of more than 25	(Teponno et al., 2016)
108	Neglectalignan A	<i>Schisandra neglecta</i> F. Schisandraceae		It showed moderate anti-HIV-1 activities with TI value above 62	(Teponno et al., 2016)

109	Neglectalignan B	<i>Schisandra neglecta</i> F. Schisandraceae		It showed moderate anti-HIV-1 activity with TI value above 23	(Teponno et al., 2016)
110	Deoxyschizandrin (Schisandrin A)	<i>Schisandra Chinensis</i> F. Schisandraceae		It can inhibit the replication of DENV in a concentration- and time-dependent manner, with 50% (EC ₅₀) value of 28.1 ± 0.42 μM against DENV serotype type 2 without significant cytotoxicity	(Yu et al., 2017)
		<i>Schisandra wilsoniana</i> F. Schisandraceae		Anti-HBV effects. It inhibited the RT-associated RDDP activity in the micromolar range IC ₅₀ ~30 μM	(Wu, 2016); (Xu et al., 2015)
111	Schisandrin B	<i>Schisandra Chinensis</i> F. Schisandraceae		It inhibited the HIV-1 RT-associated DNA polymerase activity. It inhibited the RT-associated RDDP activity in the micromolar range (IC ₅₀ ~30 μM	(Xu et al., 2015)

112	Schinlignan G	<i>Schisandra chinensis</i> F. Schisandraceae		Active against HBV DNA replication IC ₅₀ 5 μg/mL	(Teponno et al., 2016)
113	Rubrifloralignan A	<i>Schisandra rubriflora</i> F. Schisandraceae		Active against HIV EC ₅₀ 11.3 μM TI 5.7	(Chen et al., 2006); (Cui et al., 2020)
114	Schisanhenol	<i>Schisandra rubriflora</i> F. Schisandraceae		Active against HIV EC ₅₀ 5.7 μM TI 7.4	(Chen et al., 2006)
115	Magnolol	<i>Magnolia officinalis</i> F. Magnoliaceae		Inhibitor of Epstein-Barr virus early antigen (EBV-EA)	(Perez G., 2003); (Jansi et al., 2021)
		<i>Streblus asper</i> F. Moraceae		Active against HBV IC ₅₀ 2.03 and 3.76 μM, for HBsAg and HBeAg with no cytotoxicity. It inhibits the replication of HBV DNA IC ₅₀ 8.67 μM	(Li et al., 2012); (Wu, 2016); (Liu et al., 2020)
		<i>Magnolia grandiflora</i> F.		Antiplasmodial activity IC ₅₀ 3.4 ± 0.08 μM	(Latif et al., 2017)

		Magnoliaceae			
116	Honokiol	<i>Magnolia officinalis</i> F. Magnoliaceae		Inhibitor of Epstein-Barr virus early antigen (EBV-EA)	(Perez G., 2003); (Jansi et al., 2021)
		<i>Streblus asper</i> F. Moraceae		Active against HBV IC ₅₀ 3.14 μmol/L (SI = 21.47) and 4.74 μmol/L (SI = 14.22) for HBsAg and HbeAg, respectively	(Wu, 2016); (Liu et al., 2020)
		<i>Magnolia grandiflora</i> F. Magnoliaceae		Activity against <i>Plasmodium falciparum</i> IC ₅₀ 16.5 ± 0.2 μM	(Latif et al., 2017)
117	Peperomin A	<i>Peperomia pellucida</i> F. Piperaceae		It has inhibitory effects on HIV-1 IIIB growth in C8166 cells EC ₅₀ ~5 μM	(Cui et al., 2020)
118	Peperomin B	<i>Peperomia pellucida</i> F. Piperaceae		It has inhibitory effect on HIV-1 IIIB growth in C8166 cells, EC ₅₀ ~5 μM	(Cui et al., 2020)

119	(7'R,8'S,7''R,8''S)- erythro- Strebluslignan G	<i>Streblus asper</i> F. Moraceae		It exhibited significant anti-HBV activity in the secretion of HBsAg and HBeAg IC ₅₀ 1.58 and 3.24 μM, respectively. Also, it showed significant anti-HBV activity to inhibit the replication of HBV DNA IC ₅₀ 9.02 μM	(Li et al., 2012); (Cui et al., 2020)
120	Silybin A	<i>Silybum marianum</i> F. Asteraceae		Anti-HCV and other viruses; it inhibited the replication of IAV and other viruses, such as DENV, Chikungunya virus, Mayaro virus, HIV, and HBV	(Cui et al., 2020); (Antika and Dewi, 2021)
121	Silybin B	<i>Silybum marianum</i> F. Asteraceae		Antiviral activity against HCV, HBV, DENV, enterovirus family, Mayaro virus, and chikungunya virus	(Antika and Dewi, 2021)
122	Silydianin	<i>Silybum marianum</i> F. Asteraceae		Antiviral activity against HCV, HBV, DENV, enterovirus family, Mayaro virus, and chikungunya virus	(Antika and Dewi, 2021)

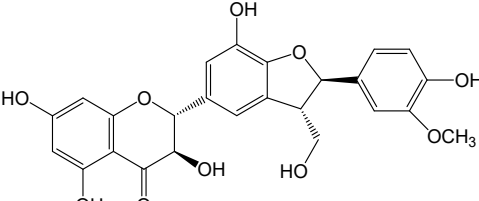
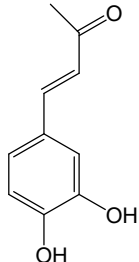
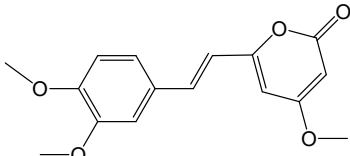
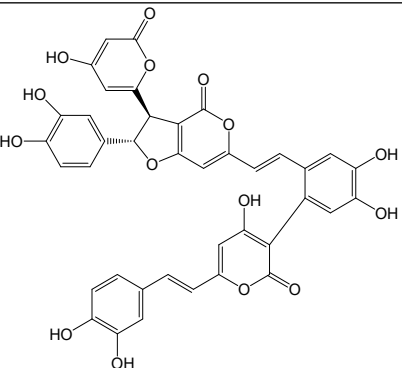
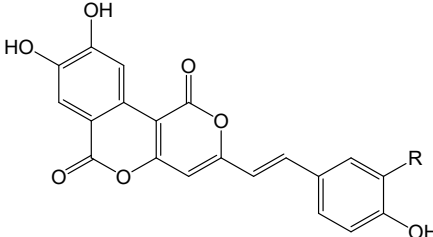
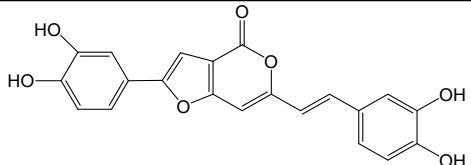
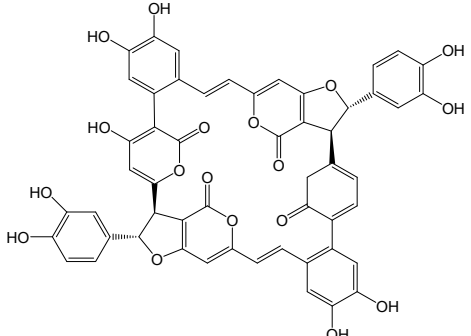
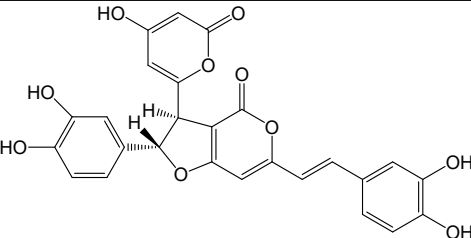
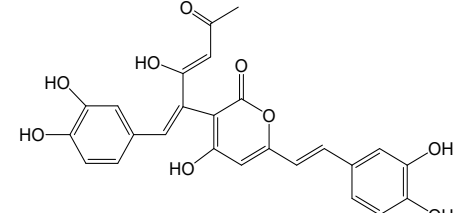
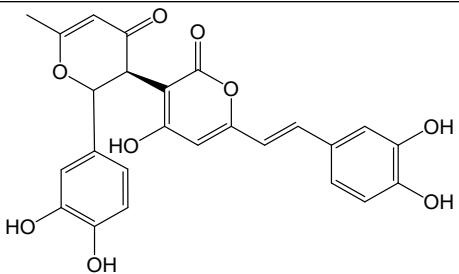
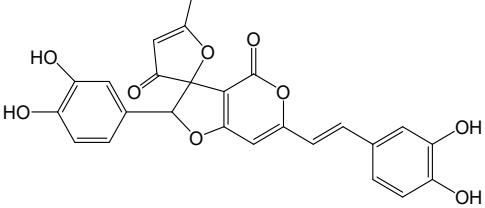
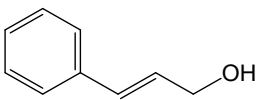
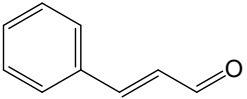
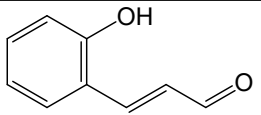
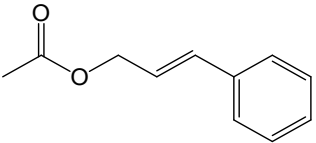
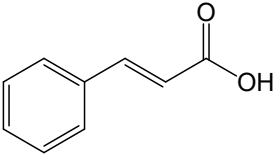
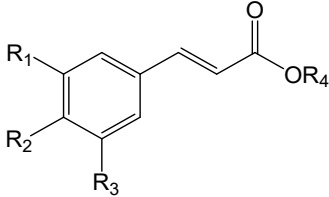
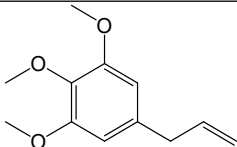
123	Silychristin	<i>Silybum marianum</i> F. Asteraceae		Antiviral activity against HCV, HBV, DENV, enterovirus family, Mayaro virus, and chikungunya virus	(Antika and Dewi, 2021)
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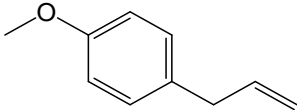
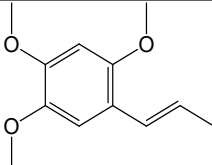
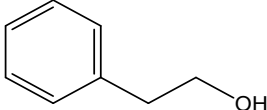
Table S2: A list of plant-derived phenylethanoids and phenylpropanoids with other biological activities

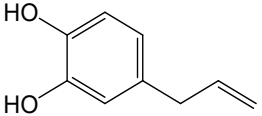
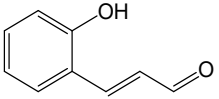
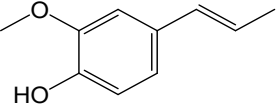
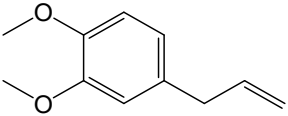
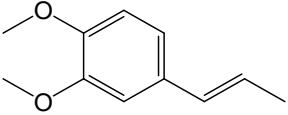
No.	Compound	Source	Structure	Biological activity	Ref.
1	3,4-Dihydroxybenzalacetone	<i>Phellinus linteus</i> F. Hymenochaetaceae		Anti-inflammatory, antitumor	(Chen et al., 2019)
2	Hispidin	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antioxidative, antitumor, antidiabetic, cardioprotective	(Chen et al., 2019)
3	Phellinstatin	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antimicrobial	(Chen et al., 2019)
					
4	Meshimakobnol A	<i>Phellinus linteus</i> F. Hymenochaetaceae	R = OH	Antitumor	(Chen et al., 2019)

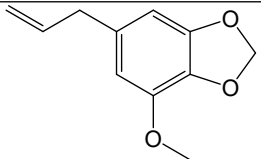
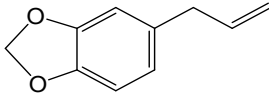
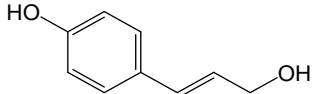
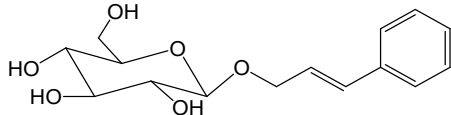
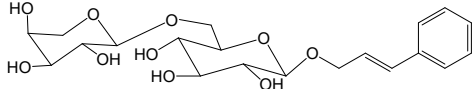
5	Meshimakobnol B	<i>Phellinus linteus</i> F. Hymenochaetaceae	R = H	Antitumor	(Chen et al., 2019)
6	Phellifuropyranone A	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antitumor	(Chen et al., 2019)
7	Phelligridimer A	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antidiabetic	(Chen et al., 2019)
8	Hypholomine B	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antidiabetic	(Chen et al., 2019)
9	Interfungin A	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antidiabetic	(Chen et al., 2019)

10	Davallialactone	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antidiabetic	(Chen et al., 2019)
11	Inoscavin A	<i>Phellinus linteus</i> F. Hymenochaetaceae		Antidiabetic	(Chen et al., 2019)
12	Cinnamyl alcohol	<i>Rhodiola rosea</i> & <i>R. arctica</i> F. Crassulaceae		Weak tonic	(Kurkin, 2003)
				suppresses 3T3-L1 cell differentiation by inhibiting anti-adipogenesis-related proteins	(Hwang et al., 2017)
13	Cinnamaldehyde (3-phenyl-2-propenal)	<i>Cinnamomum cassia</i> & <i>C. zeylanicum</i> F. Lauraceae		Anti-inflammatory	(Ilijeva and Buchbauer, 2016); (Sá et al., 2014)
14	2'-hydroxycinnamaldehyde	<i>Cinnamomum cassia</i> F. Lauraceae		Anti-inflammatory	(Sá et al., 2014)
15	Cinnamyl acetate	<i>Cinnamomum zeylanicum</i> & other <i>Cinnamomum</i> spp. F. Lauraceae		Anti-inflammatory	(Sá et al., 2014)

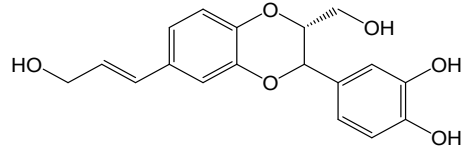
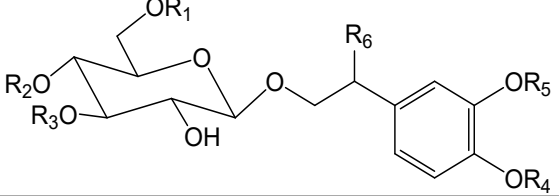
16	Cinnamic acid (3-phenylprop-2-enoic acid)	<i>Cinnamomum zeylanicum</i> F. Lauraceae, <i>Coffea arabica</i> F. Rubiaceae <i>Thea sinensis</i> F. Theaceae <i>Theobroma cocoa</i> F. Malvaceae <i>Ilexpara guariensis</i> F. Aquifoliaceae <i>Spinacia oleracea</i> F. Amaranthaceae <i>Solanum tuberosum</i> F. Solanaceae <i>Solanum lycopersicum</i> F. Solanaceae		Antibacterial, antitubercular, antifungal, anticancer and anti-inflammatory activity	(Sá et al., 2014)
					
17	<i>p</i> -Hydroxycinnamic acid	<i>Vepris glomerata</i> F. Rutaceae	$R_1 = R_2 = R_3 = R_4 = H$; $R_2 = OH$	Antibacterial agent	(Kiplimo and Koorbanally, 2012)
18	Methyl cinnamate	<i>Vepris glomerata</i> F. Rutaceae	$R_1 = R_2 = OH$; $R_3 = H$; $R_4 = CH_3$	Antibacterial agent	(Kiplimo and Koorbanally, 2012)
19	Elemicin (5-allyl-1,2,3-trimethoxybenzene)	<i>Myristica fragrans</i> F. Myristicaceae & <i>Asiasarum sieboldii</i> F. Aristolochiaceae		Anti-inflammatory	(Ilijeva and Buchbauer, 2016); (Sá et al., 2014)

20	Estragole (1-allyl-4-methoxy- benzene)	<i>Artemisia dracunculus</i> F. Asteraceae <i>Leonotis ocyimifolia</i> F. Lamiaceae, <i>Ocimum basilicum</i> F. Lamiaceae, <i>Croton zehntneri</i> F. Euphorbiaceae, <i>Pimpinella anisum</i> F. Apiaceae <i>Foeniculum vulgare</i> F. Apiaceae; <i>Illicium anisatum</i> F. Schisandraceae, <i>Boswellia serrata</i> F. Asparagaceae		Anti-inflammatory and anti-platelet activity. It is involved in immunological processes, including the inhibition of all effects on TNF α	(Ilijeva and Buchbauer, 2016)
21	Asarone (1,2,4-trimethoxy-5-[(E)- prop-1-enyl]benzene)	<i>Daucus carota</i> F. Apiaceae		Anti-inflammatory	(Sá et al., 2014)
22	Phenethyl alcohol	<i>Oils obtained from the blossoms of Rose spp.</i> F. Rosaceae <i>Citrus aurantium</i> subsp. <i>amara</i> F. Rutaceae <i>Cananga odorata</i> F. Annonaceae, <i>Dianthus caryophyllus</i> F. Caryophyllaceae and <i>Pelargonium</i> spp. F. Geraniaceae		Anti-inflammatory	(Sá et al., 2014)

23	Hydroxychavicol (4-prop-2-enylbenzene-1,2-diol)	<i>Piper betle</i> F. Piperaceae		Antimicrobial, antioxidant, anti-inflammatory, cytotoxic and antiplatelet activities	(Ilijeva and Buchbauer, 2016)
24	2'-Hydroxycinnamaldehyde [3-(2-hydroxyphenyl)-2-propenal]	<i>Cinnamomum cassia</i> F. Lauraceae		Immune-modulatory and anti-inflammatory activity	(Ilijeva and Buchbauer, 2016); (Sá et al., 2014)
25	Isoeugenol (4-propenyl-2-methoxyphenol)	<i>Eugenia caryophyllata</i> F. Myrtaceae		Inhibitory effect against some metabolic enzymes such as α -amylase, α -glycosidase and acetylcholinesterase enzymes IC ₅₀ 411.5, 19.25 and 77.00 nM, respectively Ki 21 ± 9 nM and 16 ± 3 nM, respectively	(Topal, 2019)
26	Methyleugenol (4-allyl-1,2-dimethoxybenzene)	<i>Cinnamomum cordatum</i> F. Lauraceae & <i>Croton malambo</i> F. Euphorbiaceae		Antibacterial, antifungal, anticancer, anti-inflammatory and antiplatelet activities	(Ilijeva and Buchbauer, 2016); (Sá et al., 2014)
27	Methylisoeugenol (1,2-dimethoxy-4-propenylbenzene)	<i>Asarum arifolium</i> F. Aristolochiaceae, <i>Cymbopogon javanensis</i> F. Poaceae & <i>Pimenta pseudocaryophyllus</i> F.		Hypotensive, vasorelaxant, anxiolytic and antidepressant activities	(Ilijeva and Buchbauer, 2016)

		Myrtaceae			
28	Myristicin (1-allyl-3,4-methylenedioxy-5-methoxybenzene)	<i>Petroselinum crispum</i> F. Apiaceae <i>Daucus crota</i> F. Apiaceae <i>Ocimum basilicum</i> F. Lamiaceae <i>Cinnamomum zeylanicum</i> F. Lauraceae <i>Myristica fragrans</i> F. Myristicaceae		Used in traditional medicine for the treatment of anxiety, diarrhea and stomach aches Besides its carminative effect it has antibacterial, anti-inflammatory, anticancer and hepatoprotective activity	(Ilijeva and Buchbauer, 2016); (Sá et al., 2014)
29	Safrole (4-allyl-1,2-methylenedioxybenzen)	<i>Sassafras albidum</i> F. Lauraceae <i>Ocimum basilicum</i> F. Lamiaceae <i>Pimpinella anisum</i> F. Apiaceae <i>Myristica fragrans</i> F. Myristicaceae and <i>Piper nigrum</i> F. Piperaceae		It is a well-known mutagenic and carcinogenic agent. It has anticancer, anti-inflammatory and analgesic effects	(Ilijeva and Buchbauer, 2016); (Sá et al., 2014)
30	<i>p</i> -Coumaryl alcohol	<i>Rhodiola rosea</i> F. Crassulaceae		Tonic	(Kurkin, 2003)
31	Rosin	<i>Rhodiola rosea</i> & <i>R. arctica</i> F. Crassulaceae		Tonic	(Kurkin, 2003)
32	Rosavin	<i>Rhodiola rosea</i> ; <i>R. arctica</i> F. Crassulaceae		Immunomodulating activity	(Zapesochnaya et al., 1995)

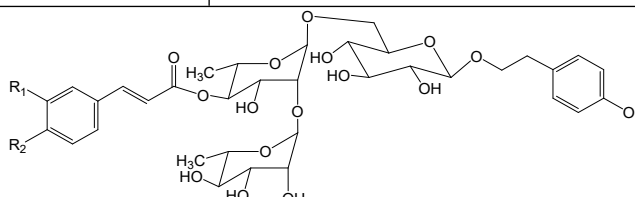
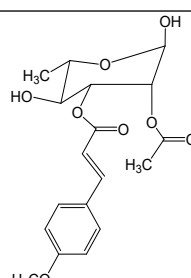
33	Rosarin	<i>Rhodiola rosea</i> ; <i>R. arctica</i> F. Crassulaceae		Tonic	(Kurkin, 2003)
34	Triandrin	<i>Rhodiola rosea</i> F. Crassulaceae; <i>Salix triandra</i> F. Salicaceae; <i>Lilium cordatum</i> F. Liliaceae		Immunomodulating activity	(Zapesochnaya et al., 1995)
35	Glomerol	<i>Vepris glomerata</i> F. Rutaceae		Antibacterial agent	(Kiplimo and Koorbanally, 2012)
36	1,3-Dicaffeoylquinic acid (cynarin)	<i>Helichrysum italicum</i> F. Asteraceae		Cholagogue	(Kurkin, 2003)
37	Forsythoside C	<i>Forsythia suspensa</i> F. Oleaceae		Antimicrobial	(Kurkin, 2003)
38	Plantamajoside	<i>Plantago major</i> F. Plantaginaceae		Antimicrobial	(Kurkin, 2003)

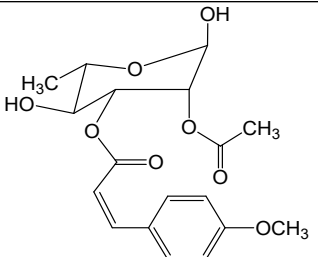
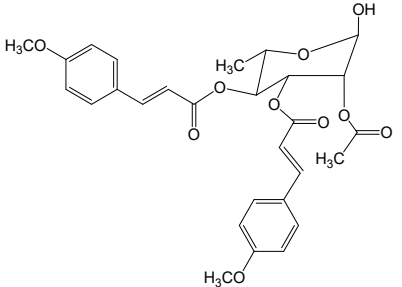
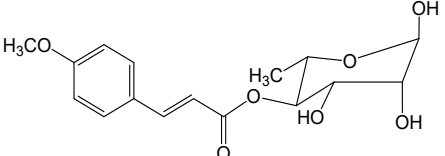
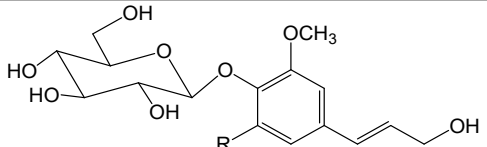
39	Americanol A	<i>Phytolacca americana</i> F. Phytolaccaceae		Hepatoprotector and neutropic activity	(Kurkin, 2003)
					
40	Jinoside A	<i>Rehmannia glutinosa</i> F. Orobanchaceae	R ₁ = β-galactosyl; R ₂ = feruloyl; R ₃ = α-rhamnosyl; R ₄ = R ₅ = R ₆ = H	immunosuppressive action	(Pan et al., 2003)
41	Jinoside B	<i>Rehmannia glutinosa</i> F. Orobanchaceae	R ₁ = β-galactosyl; R ₂ = feruloyl; R ₃ = α-rhamnosyl; R ₄ = CH ₃ ; R ₅ = R ₆ = H	immunosuppressive action	(Pan et al., 2003)
42	3,4-Dihydroxy-β-phenethyl-O-β-D-glucopyranosyl-(1→3)-O-α-L-rhamopyranosyl-(1→6)-4-O-caffeoyl-β-D-glucopyranoside	<i>Rehmannia glutinosa</i> F. Orobanchaceae	R ₁ = H; R ₂ = caffeoyl; R ₃ = β-glucosyl; R ₄ = R ₅ = R ₆ = H	Active against <i>Pseudomonas capacia</i> and <i>P. maltophilia</i>	(Pan et al., 2003)
43	3,4-Dihydroxy-β-phenethyl-O-β-D-glucopyranosyl-(1→3)-4-O-caffeoyl-β-D-glucopyranoside	<i>Rehmannia glutinosa</i> F. Orobanchaceae	R ₁ = α-rhamnosyl; R ₂ = caffeoyl; R ₃ = β-glucosyl; R ₄ = R ₅ = R ₆ = H	Active against <i>Pseudomonas capacia</i> and <i>P. maltophilia</i> at concentrations of 0.2–0.5 mg/disk	(Pan et al., 2003)
44	Incanoside	<i>Caryopteris incana</i> F. Lamiaceae	R ₁ = caffeoyl; R ₂ = H; R ₃ = β-glucosyl(1→2)α-rhamnosyl; R ₄ = R ₅ = R ₆ = H	Antioxidant and antimicrobial activity against Gram +ve and -ve bacteria	(Pan et al., 2003)
45	Incanoside C	<i>Caryopteris incana</i> F.	R ₁ = H; R ₂ = feruloyl; R ₃ = β-	Antioxidant	(Pan et al.,

		Lamiaceae	glucosyl(1→2) α -rhamnosyl; R ₄ = R ₅ = R ₆ = H		2003)
46	Incanoside D	<i>Caryopteris incana</i> F. Lamiaceae	R ₁ = H; R ₂ = feruloyl; R ₃ = β -glucosyl(1→2) α -rhamnosyl; R ₄ = CH ₃ ; R ₅ = R ₆ = H	Antioxidant	(Pan et al., 2003)
47	Incanoside E	<i>Caryopteris incana</i> F. Lamiaceae	R ₁ = H; R ₂ = feruloyl; R ₃ = β -glucosyl(1→2) α -rhamnosyl; R ₄ = H; R ₅ = CH ₃ ; R ₆ = H	Antioxidant	(Pan et al., 2003)
48	Arenarioside	<i>Ballota nigra</i> F. Lamiaceae	R ₁ = β -xylosyl; R ₂ = caffeoyl; R ₃ = α -rhamnosyl; R ₄ = R ₅ = R ₆ = H	Antioxidative and free radical scavenging activities	(Pan et al., 2003)
49	Balloletroside	<i>Ballota nigra</i> F. Lamiaceae	R ₁ = β -apiosyl; R ₂ = caffeoyl; R ₃ = α -arabinosyl(1→2) α -rhamnosyl; R ₄ = R ₅ = R ₆ = H	Antioxidative and free radical scavenging activities	(Pan et al., 2003)
50	Campneoside I	<i>Paulownia tomentosa</i> F. <i>Paulowniaceae</i>	R ₁ = H; R ₂ = caffeoyl; R ₃ = α -rhamnosyl; R ₄ = R ₅ = H; R ₆ = OCH ₃	Active against <i>Streptococcus faecium</i> MD8b, <i>S. aureus</i> (SG 511, 285 and 503) and <i>S. pyogenes</i> (A308 and A77)	(Pan et al., 2003)
51	Pedicularioside A	<i>Pedicularis striata</i> F. Scrophulariaceae	R ₁ = R ₂ = R ₃ = H	Antitumor, anti-hemolysis and	(Pan et al., 2003); (Frezza

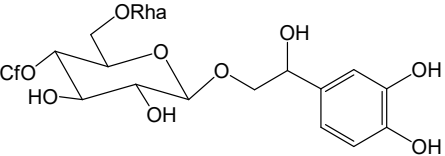
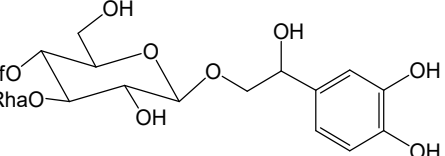
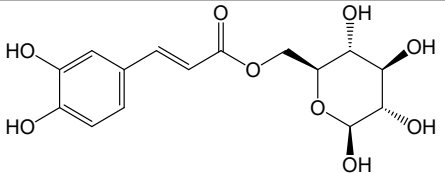
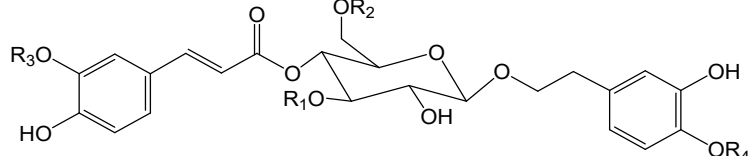
				antioxidant activities	et al., 2019)
52	Pedicularioside N	<i>Pedicularis striata</i> F. Scrophulariaceae	$R_1 = R_3 = \text{Me}; R_2 = \text{H}$	Antitumor and antioxidant activity	(Pan et al., 2003); (Frezza et al., 2019)
53	Pedicularioside M	<i>Pedicularis striata</i> F. Scrophulariaceae	$R_1 = \text{Me}; R_2 = R_3 = \text{H}$	Antioxidant activity	(Pan et al., 2003); (Frezza et al., 2019)
54	Forsythoside B	<i>Phlomis armeniaca</i> F. Lamiaceae & <i>Scutellaria salviifolia</i> F. Lamiaceae	$R_1 = R_2 = \text{H}; R_3 = \beta\text{-apiosyl}; R_4 = \text{H}$	Active against several kinds of cancer cells and antioxidant activity $\text{IC}_{50} 0.08 \mu\text{M}$	(Saracoglu et al., 1995); (Pan et al., 2003)
55	Leucosceptoside A	<i>Phlomis armeniaca</i> F. Lamiaceae & <i>Scutellaria salviifolia</i> F. Lamiaceae	$R_1 = \text{H}; R_2 = \text{CH}_3; R_3 = R_4 = \text{H}$	Active against several kinds of cancer cells and antioxidant activity $\text{IC}_{50} 0.18 \mu\text{M}$	(Saracoglu et al., 1995); (Pan et al., 2003)
		<i>Penstemon linarioides</i> F. Scrophulariaceae		inhibitory activity against PKCR $\text{IC}_{50} 19.0 \mu\text{M}$	(Zhou et al., 1998)
56	Martynoside	<i>Phlomis armeniaca</i> F. Lamiaceae <i>Scutellaria salviifolia</i> F. Lamiaceae	$R_1 = R_2 = \text{CH}_3; R_3 = R_4 = \text{H}$	Active against several kinds of cancer cells and antioxidant activity $\text{IC}_{50} 0.17 \mu\text{M}$	(Saracoglu et al., 1995); (Pan et al., 2003)

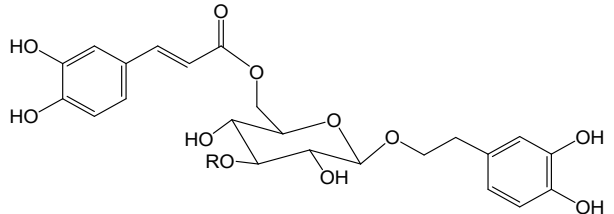
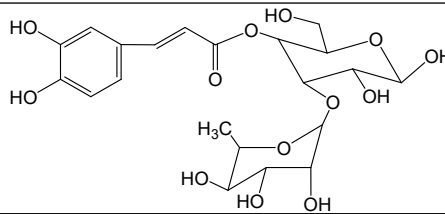
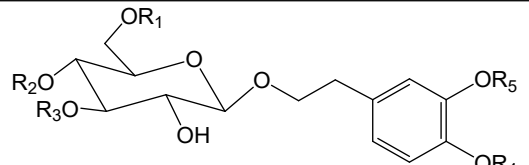
57	Phlinoside A	<i>Caryopteris incana</i> F. Lamiaceae	$R_1 = R_2 = R_3 = H$; $R_4 = \beta$ -glucosyl	Antioxidant	(Pan et al., 2003)
58	Phlinoside B	<i>Phlomis armeniaca</i> F. Lamiaceae <i>Scutellaria salviifolia</i> F. Lamiaceae	$R_1 = R_2 = R_3 = H$; $R_4 = \beta$ -xylosyl	Active against several kinds of cancer cells	(Saracoglu et al., 1995); (Pan et al., 2003)
59	Phlinoside C	<i>Phlomis armeniaca</i> F. Lamiaceae <i>Scutellaria salviifolia</i> F. Lamiaceae	$R_1 = R_2 = R_3 = H$; $R_4 = \alpha$ -rhamnosyl	Active against several kinds of cancer cells	(Saracoglu et al., 1995); (Pan et al., 2003)
60	Tecurioside	<i>Phlomis armeniaca</i> F. Lamiaceae <i>Scutellaria salviifolia</i> F. Lamiaceae	$R_1 = R_2 = R_3 = H$; $R_4 = \alpha$ -lyxosyl	Active against several kinds of cancer cells	(Saracoglu et al., 1995); (Pan et al., 2003)
61	2'-Acetylacteoside	<i>Cistanche deserticola</i> F. Orobanchaceae	$R_1 = H$; $R_2 =$ caffeoyl; $R_3 = \alpha$ -rhamnosyl; $R_4 = Ac$; $R_5 = H$	Hepatoprotective and anti-hemolysis activities Antioxidant activity	(Pan et al., 2003)
62	Tubuloside B	<i>Cistanche deserticola</i> F. Orobanchaceae	$R_1 =$ caffeoyl; $R_2 = H$; $R_3 = \alpha$ -rhamnosyl; $R_4 = Ac$; $R_5 = H$	Hepatoprotective activity Antioxidant activity	(Pan et al., 2003)
63	Cistanoside C	<i>Cistanches</i> spp. F. Orobanchaceae	$R_1 = H$; $R_2 =$ caffeoyl; $R_3 = \alpha$ -rhamnosyl; $R_4 = H$; $R_5 = CH_3$	Protective effects on both sex and learning behaviors in the hanging stress loaded mice	(Pan et al., 2003)

64	Cistanoside D	<i>Pedicularis striata</i> F. Scrophulariaceae	R ₁ = H; R ₂ = feruloyl; R ₃ = α-rhamnosyl; R ₄ = H; R ₅ = CH ₃	Anti-hemolysis and antioxidant activities	(Pan et al., 2003)
65	Poliumoside	<i>Brandisia hancei</i> F. Orobanchaceae	R ₁ = α-rhamnosyl; R ₂ = caffeoyl; R ₃ = α-rhamnosyl; R ₄ = R ₅ = H	Anti-hemolysis effect	(Pan et al., 2003)
		<i>Penstemon linarioides</i> F. Scrophulariaceae		Inhibitory activity against PKCR IC ₅₀ 24.4 μM	(Zhou et al., 1998)
66	Brandioside	<i>Brandisia hancei</i> F. Orobanchaceae	R ₁ = α-rhamnosyl; R ₂ = caffeoyl; R ₃ = α-rhamnosyl; R ₄ = R ₅ = H	Anti-hemolysis effect	(Pan et al., 2003)
					
67	4-Cinnamoyl-mussatioside	<i>Mussatia</i> species F. Bignoniaceae	R ₁ = R ₂ = H	Antiaggregation effect on platelets	(Pan et al., 2003)
68	4-Di-methylcaffeoyl-mussatioside	<i>Mussatia hyacinthina</i> F. Bignoniaceae	R ₁ = R ₂ = OMe	Antiaggregation effect on platelets	(Pan et al., 2003)
69	4- <i>p</i> -Methoxycinnamoyl-mussatioside	<i>Mussatia hyacinthina</i> F. Bignoniaceae	R ₁ = H; R ₂ = OMe	Antiaggregation effect on platelets	(Pan et al., 2003)
70	2- <i>O</i> -Acetyl-3- <i>O</i> -(<i>E</i>)- <i>p</i> -methoxycinnamoyl-α-L-rhamnopyranoside	<i>Scrophularia buergeriana</i> F. Scrophulariaceae		Anti-neurodegeneration effect	(Kim and Kim, 2000); (Pan et al., 2003)

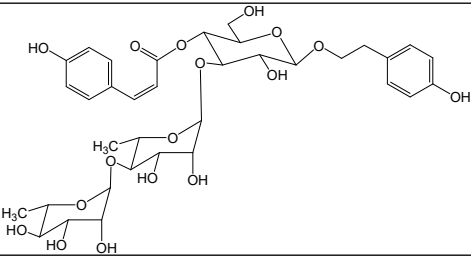
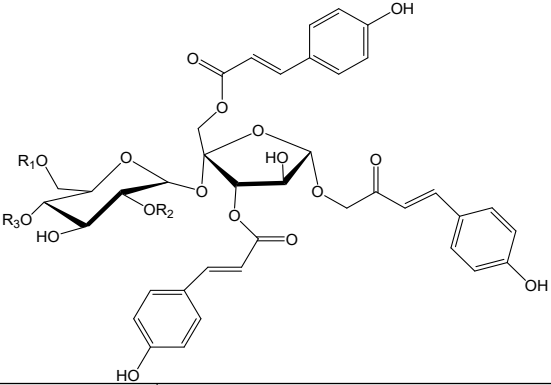
71	2- <i>O</i> -Acetyl-3- <i>O</i> - (<i>Z</i>)- <i>p</i> -methoxycinnamoyl- α -L-rhamnopyranoside	<i>Scrophularia buergeriana</i> F. Scrophulariaceae		Anti-neurodegeneration effect	(Kim and Kim, 2000); (Pan et al., 2003)
72	2- <i>O</i> -Acetyl-3,4-di- <i>O</i> -(<i>E</i>)- <i>p</i> -methoxycinnamoyl- α -L-rhamnopyranoside	<i>Scrophularia buergeriana</i> F. Scrophulariaceae		Anti-neurodegeneration effect	(Kim and Kim, 2000); (Pan et al., 2003)
73	4- <i>O</i> -(<i>E</i>)- <i>p</i> -Methoxycinnamoyl- α -L-rhamnopyranoside	<i>Scrophularia buergeriana</i> F. Scrophulariaceae		Anti-neurodegeneration effect	(Kim and Kim, 2000); (Pan et al., 2003)
					
74	Coneferin	<i>Phillyrea latifolia</i> F. Oleaceae	R = H	Change the activity of some enzymes; antioxidant and anti-inflammatory activities	(Pan et al., 2003); (Lanza et al., 2001)
75	Syringin	<i>Phillyrea latifolia</i> F. Oleaceae	R = OCH ₃	Change the activity of some enzymes; antioxidant and anti-inflammatory activities	(Pan et al., 2003); (Lanza et al., 2001)

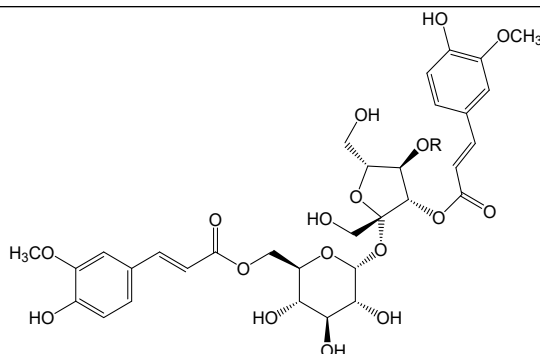
		<i>Syringa vulgaris</i> , <i>S. amurensis</i> F. Oleaceae; <i>Lilium cordatum</i> F. Liliaceae; <i>Codonopsis tangshen</i> F. Campanulaceae; <i>Eleutherococcus senticosus</i> F. Araliaceae		Immunomodulating activity	(Zapesochnaya et al., 1995)
76	Cistanoside A	<i>Cistanches</i> species F. Orobanchaceae	R ₁ = H; R ₂ = α-rhamnosyl; R ₃ = caffeoyl; R ₄ = β-glucosyl; R ₅ = OH; R ₆ = OCH ₃	Protective effects on both sex and learning behaviors in the hanging stress loaded mice	(Pan et al., 2003)
		<i>Cistanche deserticola</i> F. Orobanchaceae		Antioxidative and free radical scavenging activities	(Pan et al., 2003)
77	Tubuloside A	<i>Cistanche deserticola</i> F. Orobanchaceae	R ₁ = Ac; R ₂ = α-rhamnosyl; R ₃ = caffeoyl; R ₄ = β-glucosyl; R ₅ = R ₆ = OH	Antioxidative and free radical scavenging activities	(Pan et al., 2003)
78	Syringalide A 3'-α-rhamnopyranoside	<i>Cistanche deserticola</i> F. Orobanchaceae	R ₁ = H; R ₂ = α-rhamnosyl; R ₃ = caffeoyl; R ₄ = R ₅ = H; R ₆ = OH	Antioxidative and free radical scavenging activities	(Pan et al., 2003)

79	Suspensaside	<i>Forsythiae suspensa</i> & <i>F. viridisscima</i> F. Oleaceae		Change the activity of some enzymes and antioxidant activity	(Pan et al., 2003)
80	β -Hydroxyacteoside	<i>Forsythiae suspensa</i> & <i>F. viridisscima</i> F. Oleaceae		Change the activity of some enzymes and antioxidant activity	(Pan et al., 2003)
81	6-O-Caffeoyl- β -O-glucose	<i>Caryopteris incana</i> F. Lamiaceae		Antioxidative and free radical scavenging activities	(Pan et al., 2003)
					
82	2-(3-Hydroxy-4-methoxy-phenyl)-ethyl-O-(α -L-rhamnosyl)-(1 \rightarrow 3)-O-(α -L-rhamnosyl)-(1 \rightarrow 6)-4-O-E-feruloyl- β -D-glucopyranoside	<i>Digitalis purpurea</i> F. Scrophulariaceae	$R_1 = R_2 = \alpha$ -rhamnosyl; $R_3 = R_4 = \text{CH}_3$	Inhibitory activity against PKCR IC_{50} 125 μM	(Zhou et al., 1998)
83	Calceolarioside A	<i>Digitalis purpurea</i> F. Scrophulariaceae	$R_1 = R_2 = R_3 = R_4 = \text{H}$	Inhibitory activity against PKCR IC_{50} 0.6 μM	(Zhou et al., 1998)
		<i>Globularia orientalis</i> F. Plantaginaceae		Antioxidant activity IC_{50} 0.04 μM	(Heilmann et al., 2000); (Pan et al., 2003)

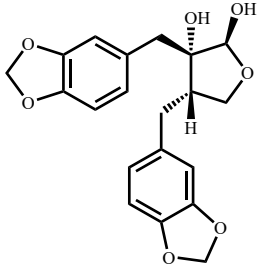
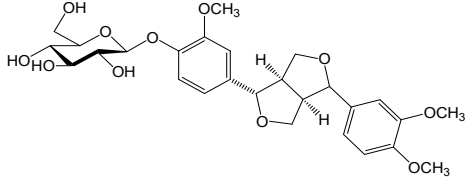
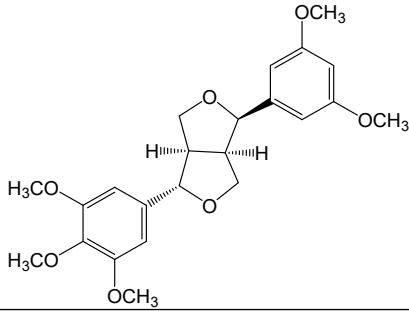
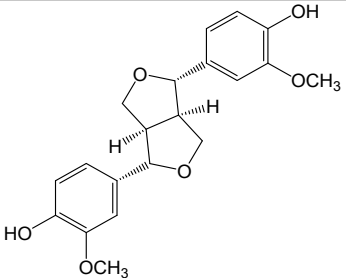
					
84	Plantainoside D	<i>Digitalis purpurea</i> F. Scrophulariaceae	R = β -glucosyl	Inhibitory activity against PKCR IC ₅₀ 14.8 μ M	(Zhou et al., 1998)
					
85	Cistanoside F	<i>Cistanche deserticola</i> F. Orobanchaceae		Antioxidative and free radical scavenging activity	(Pan et al., 2003)
					
86	Isomartynoside	<i>Galeopsis pubescens</i> F. Lamiaceae	R ₁ = feruloyl; R ₂ = H; R ₃ = α - rhamnosyl; R ₄ = CH ₃ = R ₅ = H	Antioxidant activity IC ₅₀ 0.17 μ M	(Heilmann et al., 2000); (Pan et al., 2003)
87	Lugrandoside	<i>Digitalis ferruginea</i> subsp. <i>ferruginea</i> F. Scrophulariaceae	R ₁ = β -glucosyl; R ₂ = caffeoyl; R ₃ = R ₄ = R ₅ = H	Antioxidant activity IC ₅₀ 0.04 μ M	(Heilmann et al., 2000); (Pan et al., 2003)
88	Ferruginoside B	<i>Digitalis ferruginea</i> subsp. <i>ferruginea</i> F. Scrophulariaceae	R ₁ = β -glucosyl; R ₂ = R ₃ = R ₄ = R ₅ = H	Antioxidant activity IC ₅₀ 1.9 μ M	(Heilmann et al., 2000); (Pan et al., 2003)
89	Darendoside B	<i>Scutellaria orientalis</i> subsp. <i>pinnatifida</i> F. Lamiaceae	R ₁ = R ₂ = H; R ₃ = α -rhamnosyl; R ₄ = CH ₃ ; R ₅ = H	Antioxidant activity IC ₅₀ 3.6 μ M	(Heilmann et al., 2000); (Pan et al., 2003)
90	Deacylacteoside dimethyl ether	<i>Incarvillea sinensis</i> var. <i>przewalskii</i> F.	R ₁ = R ₂ = H; R ₃ = α -rhamnosyl; R ₄ = R ₅ = CH ₃	Antioxidant activity	(Heilmann et al., 2000); (Pan

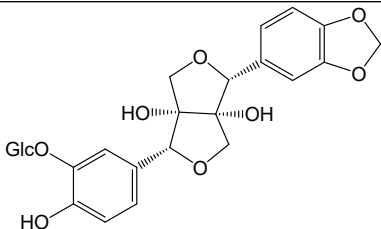
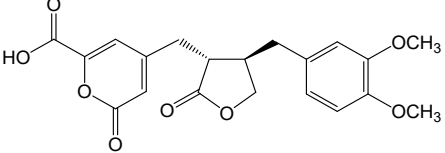
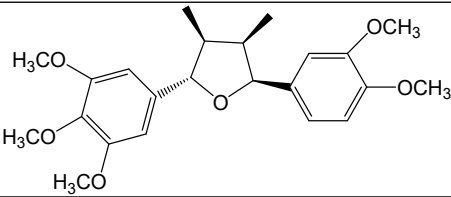
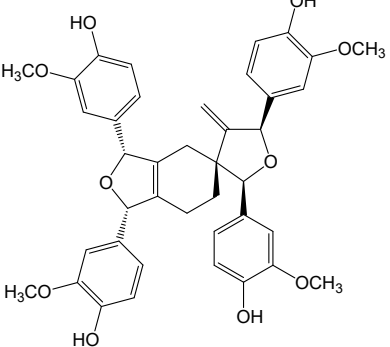
		Bignoniaceae		IC ₅₀ 8.5 μM	et al., 2003)
91	Leucosceptoside B	<i>Marrubium alysson</i> F. Lamiaceae	R ₁ = β-apiosyl; R ₂ = feruloyl; R ₃ = α-rhamnosyl; R ₄ = CH ₃ ; R ₅ = H	Antioxidant activity IC ₅₀ 0.17 μM	(Heilmann et al., 2000); (Pan et al., 2003)
92	Alyssonoside	<i>Phlomis grandiflora</i> var. <i>fimbrilligera</i> F. Lamiaceae	R ₁ = β-apiosyl; R ₂ = feruloyl; R ₃ = α-rhamnosyl; R ₄ = R ₅ = H	Antioxidant activity IC ₅₀ 0.15 μM	(Heilmann et al., 2000); (Pan et al., 2003)
93	Hattushoside	<i>Phlomis grandiflora</i> var. <i>fimbrilligera</i> F. Lamiaceae	R = Syringyl	Antioxidant activity IC ₅₀ 0.35 μM	(Heilmann et al., 2000); (Pan et al., 2003)
94	Phlomisethanoside	<i>Phlomis grandiflora</i> var. <i>fimbrilligera</i> F. Lamiaceae	R = Vanilloyl	Antioxidant activity IC ₅₀ 0.04 μM	(Heilmann et al., 2000); (Pan et al., 2003)
95	Ligupurpurososide A	<i>Ligustrum purpurascens</i> F. Oleaceae	R ₁ = R ₂ = OH; R ₃ = H	Antioxidative and free radical scavenging activities	(Pan et al., 2003)
96	<i>Trans</i> -Ligupurpurososide B	<i>Ligustrum purpurascens</i> F. Oleaceae	R ₁ = R ₂ = H; R ₃ = α-rhamnosyl	Antioxidative and free radical scavenging activities	(Pan et al., 2003)

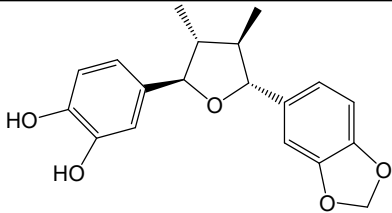
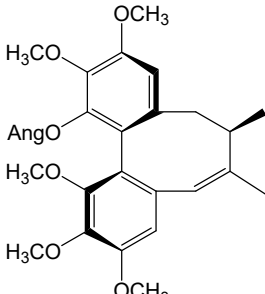
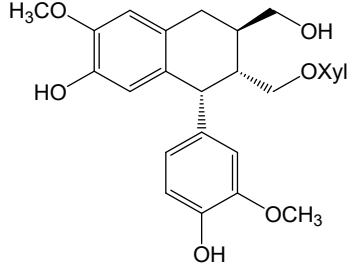
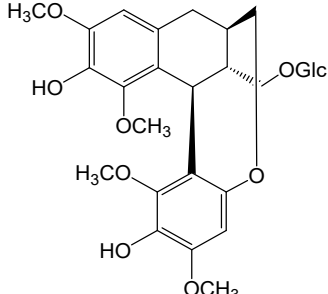
97	Osmanthuside B	<i>Ligustrum purpurascens</i> F. Oleaceae	$R_1 = R_2 = R_3 = H$	Antioxidative and free radical scavenging activities	(Pan et al., 2003)
98	<i>Cis</i> -Ligupurpuroside B	<i>Ligustrum purpurascens</i> F. Oleaceae		Antioxidative and free radical scavenging activities	(Pan et al., 2003)
					
99	Hydropiperoside	<i>Persicaria orientalis</i> F. Polygonaceae	$R_1 = R_2 = R_3 = H$	Inhibitor of melanogenesis	(Masum et al., 2019)
100	Vanicoside A	<i>Persicaria orientalis</i> F. Polygonaceae	$R_1 = \text{feruloyl}; R_2 = \text{Ac}; R_3 = H$	Inhibitor of melanogenesis	(Masum et al., 2019)
101	Vanicoside B	<i>Persicaria orientalis</i> F. Polygonaceae	$R_1 = \text{feruloyl}; R_2 = R_3 = H$	Inhibitor of melanogenesis	(Masum et al., 2019)
102	Vanicoside C	<i>Persicaria orientalis</i> F. Polygonaceae	$R_1 = R_3 = H; R_2 = \text{Ac}$	Inhibitor of melanogenesis	(Masum et al., 2019)
103	Vanicoside E	<i>Persicaria orientalis</i> F. Polygonaceae	$R_1 = \text{feruloyl}; R_2 = R_3 = \text{Ac}$	Inhibitor of melanogenesis	(Masum et al., 2019)

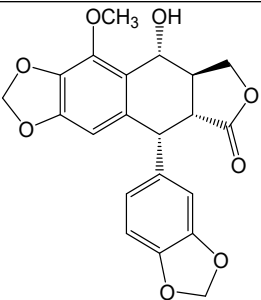
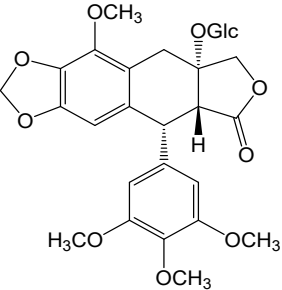
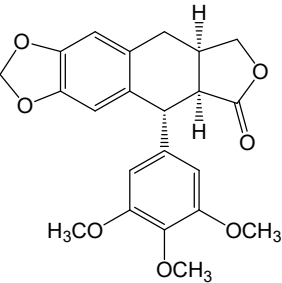
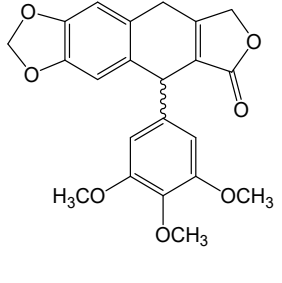


104	3,6'- <i>O</i> -Diferuloyl-sucrose	<i>Lilium longiflorum</i> F. Liliaceae	R = H	DPP-IV inhibitory activity IC ₅₀ 46.19	(Kim et al., 2020)
105	4- <i>O</i> -Acetyl-3,6'- <i>O</i> -diferuloyl-sucrose	<i>Lilium longiflorum</i> F. Liliaceae	R = Ac	DPP-IV inhibitory activity IC ₅₀ 63.26 μM	(Kim et al., 2020)
106	(+)-Liriodenol	<i>Liriodendron hybrid</i> F. Magnoliaceae		Cytotoxic effects against NIC-H460, BGC-823, SGC-7901 and MDA-MB-231 IC ₅₀ 31, 45, 34 and 29 mg/mL, respectively	(Yang et al., 2015); (Teponno et al., 2016)
107	(+)-Lariciresinol	<i>Wikstroemia elliptica</i> F. Thymelaeaceae		Anticancer activity	(Saarinen et al., 2008); (Cui et al., 2020)

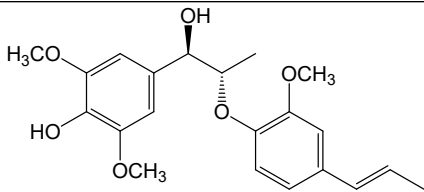
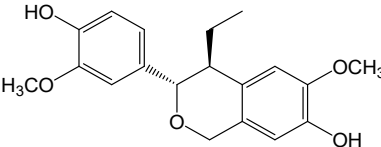
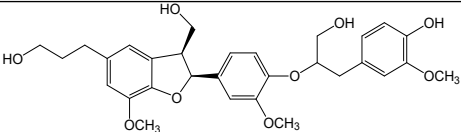
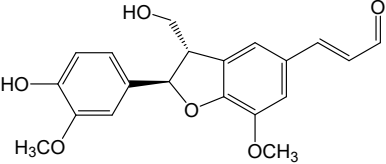
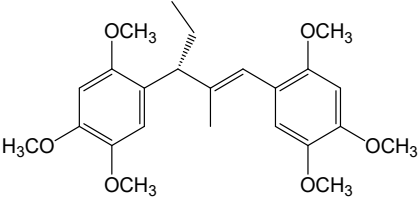
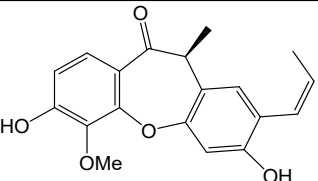
108	Glandularin	<i>Glandularia</i> × <i>hybrida</i> F. Verbenaceae		Moderate cytotoxic activity against SK-MEL and VERO cells with IC ₅₀ 45.6 and 37.6 μM, respectively. Moderate inhibition activity on iNOS IC ₅₀ 49.7 μM	(Mohamed et al., 2022)
109	Phillyrin	<i>Phillyrea latifolia</i> F. Oleaceae		<i>In vitro</i> anti-inflammatory activity. It inhibits the release of the cyclo-oxygenase metabolites prostaglandin E ₂ IC ₅₀ 45.6 μM	(Lanza et al., 2001)
110	Epimagnolin B	<i>Magnolia fargesii</i> F. Magnoliaceae		Anti-inflammatory activity by inhibition of the production of NO and PGE ₂ and the expression of respective enzyme iNOS and COX-2	(Teponno et al., 2016)
111	(+)-Pinoresinol	<i>Melia toosendan</i> F. Meliaceae		Potent ABTS free radical scavenging activity IC ₅₀ 45.1 μM	(Wang et al., 2014)

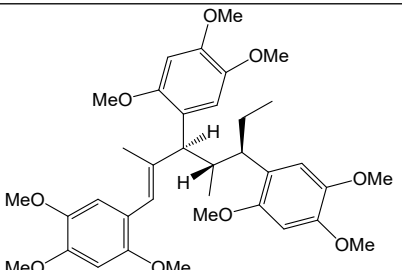
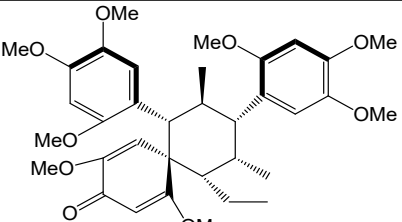
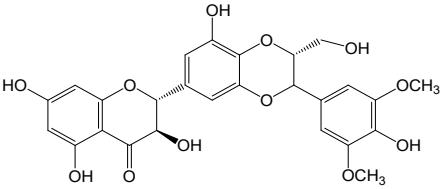
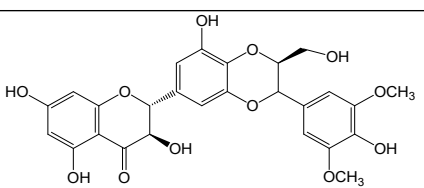
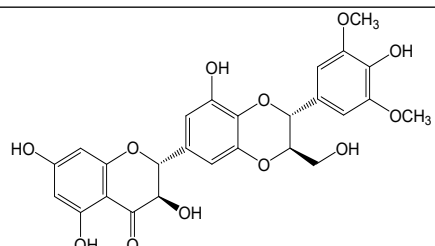
112	Khainaoside A	<i>Vitex glabrata</i> F. Verbenaceae		Potent inhibitory effects on estrogen-enhanced cell proliferation	(Teponno et al., 2016)
113	Arctiidilactone	<i>Arctium lappa</i> F. Asteraceae		Potent anti-inflammatory and exhibited 75.51, 70.72, and 61.17% inhibition at 10, 1, and 0.1 μM, respectively	(Yang et al., 2015); (Teponno et al., 2016)
114	Ligraminol A	<i>Acorus gramineus</i> F. Araceae		Active against the SK-MEL-2 cell line IC ₅₀ 4.5 μM	(Teponno et al., 2016)
115	Ramonanin A	<i>Guaiacum officinale</i> F. Zygophyllaceae		Moderate <i>in vitro</i> cytotoxic activities against human breast cancer cell lines and induced cell death <i>via</i> apoptosis	(Teponno et al., 2016)

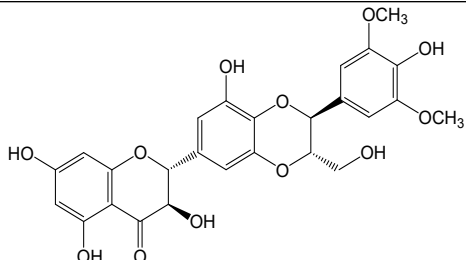
116	(7, 8-Trans-8, 8'-trans-7', 8'-trans)-7- (3,4-methylenedioxyphenyl)-8-methyl-8'-methyl-7'-(3',4'- dihydroxyphenyl)-tetrahydrofuran	<i>Schisandra sphenanthera</i> F. Schisandraceae		Anti-oxidative haemolysis of human red blood cells (RBCs) activity IC ₅₀ 35 µg/mL	(Jiang et al., 2015); (Teponno et al., 2016)
117	Neglschisandrin F	<i>Schisandra neglecta</i> F. Schisandraceae		It exhibited marginal cytotoxicity against the human lung carcinoma A549 cell line with EC ₅₀ values of 12 µg/mL	(Teponno et al., 2016)
118	Tiliamuroside B	<i>Tilia amurensis</i> F. Tiliaceae		Significant cytotoxicity against A549, SK-OV-3, SK-MEL-2, and HCT-15 cell lines IC ₅₀ 7.3, 8.9, 7.8 and 6.2 µM, respectively.	(Kim et al., 2012)
119	(+)-Ovafolinin B-9'-O-β-D-glucopyranoside	<i>Eurya japonica</i> F. Theaceae		Potent antioxidant activity ED ₅₀ 23	(Teponno et al., 2016)

120	Cleistantoxin	<i>Cleistanthus indochnensis</i> F. Euphorbiaceae		Strong cytotoxic activity against KB, MCF-7, HT29 and MCF-7R cancer cell lines IC ₅₀ 0.02, 0.04, 0.01 and 0.03 μM, respectively	(Teponno et al., 2016); (Thanh et al., 2012)
121	3α-O-(β-D - Glucopyranosyl)desoxy-podophyllotoxin	<i>Cleistanthus boivinianus</i> F. Euphorbiaceae		Active against A2780 ovarian cancer and HCT-116 human colon carcinoma cell lines. IC ₅₀ 33.0 ± 3.6 nM and 20.5 nM, respectively	(Liu et al., 2015)
122	Deoxypicropodophyllotoxin	<i>Cleistanthus boivinianus</i> F. Euphorbiaceae		Active against A2780 ovarian cancer cell line IC ₅₀ >10 μM	(Liu et al., 2015)
123	(±)-β-Apopicropodophyllin	<i>Cleistanthus boivinianus</i> F. Euphorbiaceae		Potent activity against the A2780 ovarian cancer cell line IC ₅₀ 63.1 ± 6.7nM. Modest activity against the A2058 human caucasian	(Liu et al., 2015)

				metastatic melanoma and MES-SA human uterine sarcoma cell lines IC ₅₀ 4.6 and 4.0 μM, respectively	
124	(-)-Desoxypodophyllotoxin	<i>Cleistanthus boivinianus</i> F. Euphorbiaceae	R = H	Potent activity against the A2780 ovarian cancer cell line IC ₅₀ 230 ± 1 nM	(Liu et al., 2015)
125	β-Peltatin-5-O-β-D-glucopyranoside	<i>Cleistanthus boivinianus</i> F. Euphorbiaceae	R = O-β-D-glucopyranose	Moderate activity against the A2780 IC ₅₀ 4.9 ± 0.1 μM	(Liu et al., 2015)
126	Conidendrin	<i>Abies holophylla</i> F. Pinaceae		Moderately inhibited NO production IC ₅₀ 28.5 μM	(Kim et al., 2013)

127	Myrifralignan C	<i>Myristica fragrans</i> F. Myristicaceae		Potent inhibitory activity against the production of nitric oxide (NO) in the RAW264.7 cell line stimulated by lipopolysaccharide	(Teponno et al., 2016)
128	Meliasendanin A	<i>Melia toosendan</i> F. Meliaceae		Stronger ABTS free radical scavenging activity IC ₅₀ 62.8 μM	(Wang et al., 2014)
129	Euryalin B	<i>Euryale ferox</i> F. Nymphaeaceae		Strong effect against DPPH assay SC ₅₀ 6.8 μM	(Teponno et al., 2016)
130	7 <i>R</i> ,8 <i>S</i> -Balanophonin	<i>Crataegus pinnatifida</i> F. Rosaceae		Potent cytotoxic activity against the HT-1080 cell line IC ₅₀ = 8.86 μM	(Huang et al., 2013)
131	Tatarinan S	<i>Acorus tatarinowii</i> F. Araceae		Potential protective effect against β-amyloid toxicity, its value of PT50 extended up to 62.3% at 100 μM	(Luo et al., 2016)
132	Penthorin B	<i>Penthorum chinense</i> F. Saxifragaceae		<i>In vitro</i> protective activities against acetaminophen-induced hepatocyte injury, the inhibition was calculated as 15.0%	(He et al., 2015)

133	Tatanan A	<i>Acorus tatarinowii</i> F. Araceae		at 5 μ M Potently increase Glucokinase (GK) enzymatic activity EC _{1,5} 0.16–1.85 μ M	(Ni et al., 2011)
134	Tatanan B	<i>Acorus tatarinowii</i> F. Araceae		Potently increase Glucokinase (GK) enzymatic activity EC _{1,5} 0.16–1.85 μ M	(Ni et al., 2011)
135	Hovenin A	<i>Hovenia acerba</i> F. Rhamnaceae		Moderate inhibition on the production of NO and IL-6 in LPS-stimulated RAW264.7 cells IC ₅₀ 45 – 63 μ M	(Teponno et al., 2016)
136	Hovenin B	<i>Hovenia acerba</i> F. Rhamnaceae		Moderate inhibition on the production of NO and IL-6 in LPS-stimulated RAW264.7 cells IC ₅₀ 45 – 63 μ M	(Teponno et al., 2016)
137	Hovenin C	<i>Hovenia acerba</i> F. Rhamnaceae		Moderate inhibition on the production of NO and IL-6 in LPS-stimulated RAW264.7 cells IC ₅₀ 45 – 63 μ M	(Teponno et al., 2016)

138	Hovenin D	<i>Hovenia acerba</i> F. Rhamnaceae		Moderate inhibition on the production of NO and IL-6 in LPS-stimulated RAW264.7 cells IC ₅₀ 45– 63 μM	(Teponno et al., 2016)
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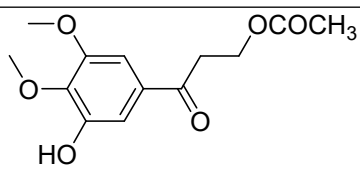
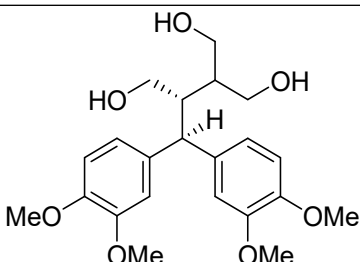
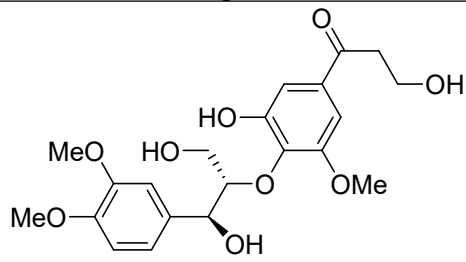
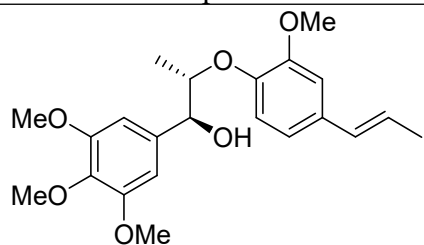
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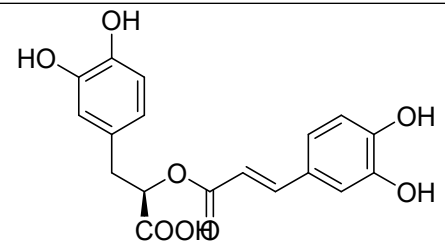
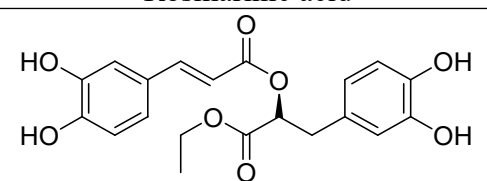
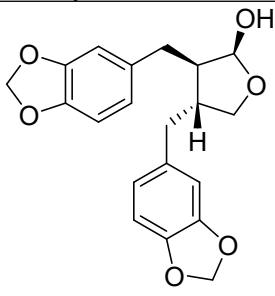
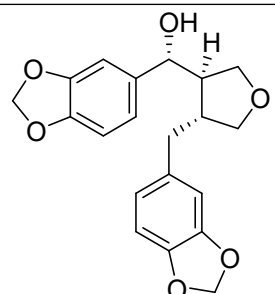
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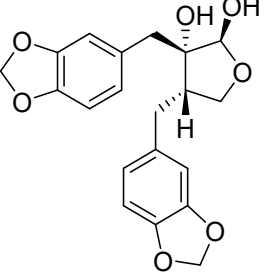
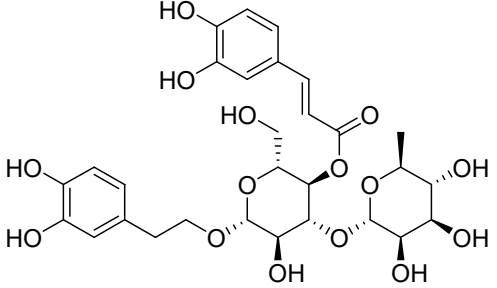
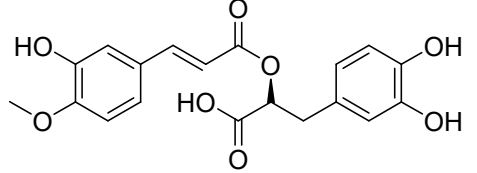
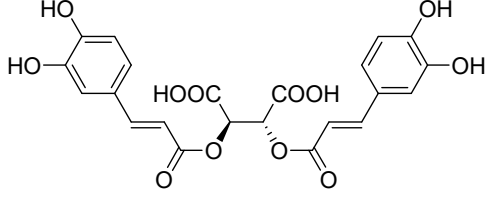
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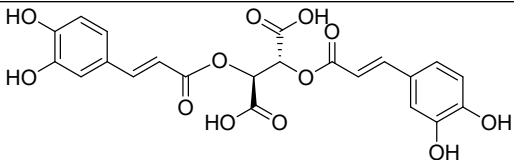
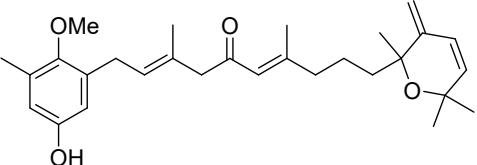
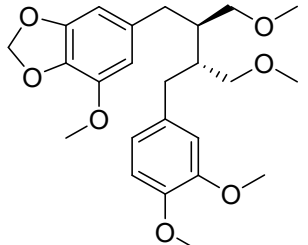
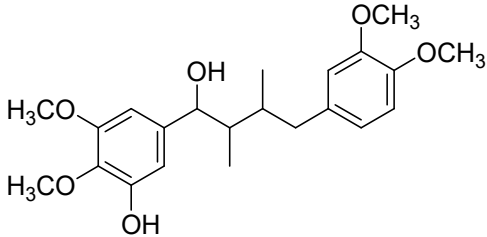
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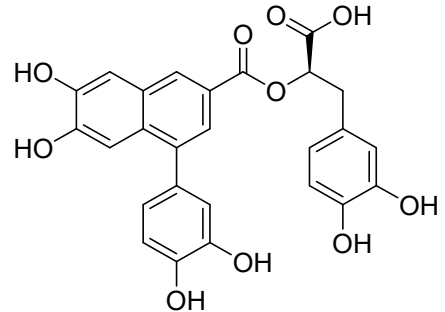
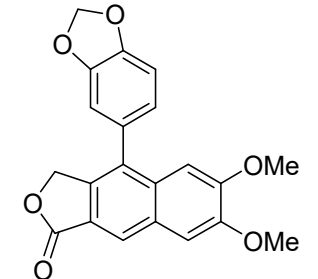
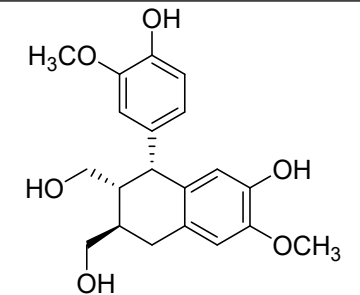
Table S3. Docking Scores of selected phenylpropanoids & phenylethanoids within three proteins of *P. falciparum*.

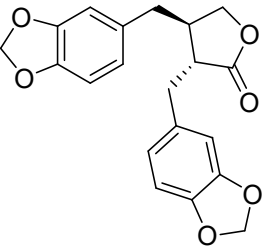
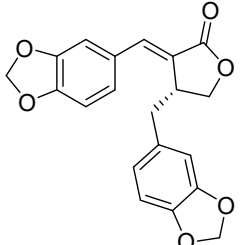
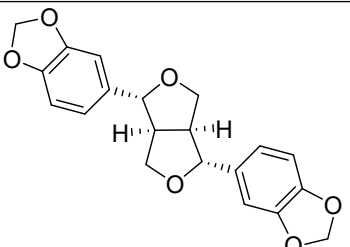
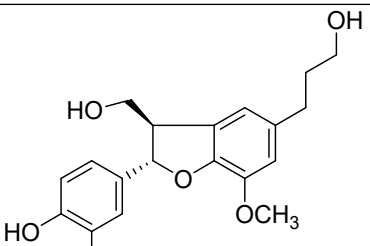
ID	Molecule Structure	1V0P ^a		4PD4 ^b		6AGT ^c	
		S _g	RMSD _h	S	RMSD	S	RMSD
1	 Feddeiketone A	-5.48	1.68	-6.37	2.29	-7.33	1.24
2	 Feddeiphenol A	-7.10	1.61	-7.04	1.51	-6.48	1.58
3	 Feddeiphenol C	-7.22	1.69	-7.47	1.71	-8.87	1.83
4	 Feddeiphenol D	-6.75	2.48	-8.20	1.49	-8.68	1.47

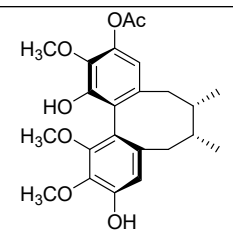
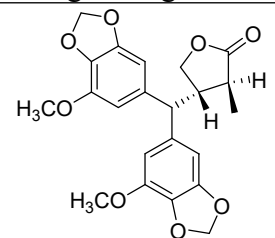
	Surinamensin						
5	 <p style="text-align: center;">Rosmarinic acid</p>	-6.26	1.79	-7.73	1.19	-7.34	1.35
6	 <p style="text-align: center;">Ethyl rosmarinate</p>	-6.94	1.65	-7.33	1.55	-7.34	1.02
7	 <p style="text-align: center;">Cubebin</p>	-6.2	1.56	-7.02	1.72	-8.24	0.82
8	 <p style="text-align: center;">Isocubebin</p>	-6.13	1.26	-7.39	1.55	-7.88	1.89

9	 <p>Glandularin</p>	-6.43	1.84	-7.67	1.89	-7.89	1.78
10	 <p>Forsythoside A</p>	-8.31	2.47	-8.41	1.44	-7.65	2.16
11	 <p>Shimobashiric acid B</p>	-6.33	1.52	-7.48	1.57	-7.80	1.10
12	 <p>L-Chicoric acid</p>	-7.65	1.29	-7.56	1.69	-8.38	1.49

13	 <p>Di-E-caffeoyl-meso-tartaric acid</p>	-6.32	1.82	-8.50	1.32	-7.29	1.12
14	 <p>Usneidone E</p>	-6.94	2.64	-8.10	1.28	-5.98	1.59
15	 <p>Niranthin</p>	-7.07	1.71	-7.13	1.21	-7.04	1.33
16	 <p>Kadangustin H</p>	-6.52	1.97	-9.35	1.62	-8.87	0.93

17	 <p style="text-align: center;">Globoidnan A</p>	-7.71	1.51	-7.70	1.78	-9.20	1.44
18	 <p style="text-align: center;">Retrojusticidin B</p>	-6.53	2.52	-5.75	1.03	-7.66	1.62
19	 <p style="text-align: center;">(-)-Isolariciresinol</p>	-7.01	1.55	-5.00	1.48	-6.84	1.06

20	 <p>Hinokinin</p>	-5.99	1.21	-7.12	1.68	-7.55	1.97
21	 <p>Savinin</p>	-6.58	1.84	-6.15	2.15	-6.78	0.98
22	 <p>Sesamin</p>	-6.01	2.09	-7.61	1.57	-7.73	1.72
23	 <p>Vladinol F</p>	-6.59	2.09	-7.03	1.54	-7.88	0.93

24	 Neglectalignan A	-6.47	2.15	-5.84	0.81	-7.26	1.05
25	 Peperomin A	-7.06	1.76	-7.69	0.87	-6.39	1.4
#	Co-crystallized Ligand	-7.32 ^d	2.10	-6.91 ^e	1.63	-7.98 ^f	0.63

^a1V0P: *P. falciparum* kinase; ^b4PD4: cytochrome bc1; and ^c6AGT: lysyl-tRNA synthetase (PfKRS1) active sites; ^dPurvalanol B (PVB); ^e2-[trans-4-(4-chlorophenyl)cyclohexyl]-3-hydroxynaphthalene-1,4-dione (AOQ); ^fN-(cyclohexylmethyl)-4-oxo-4H-1-benzopyran-2-carboxamide (9X0); ^gDocking Score (S; Kcal/mol); ^hRoot-Mean Square Deviation (Å)

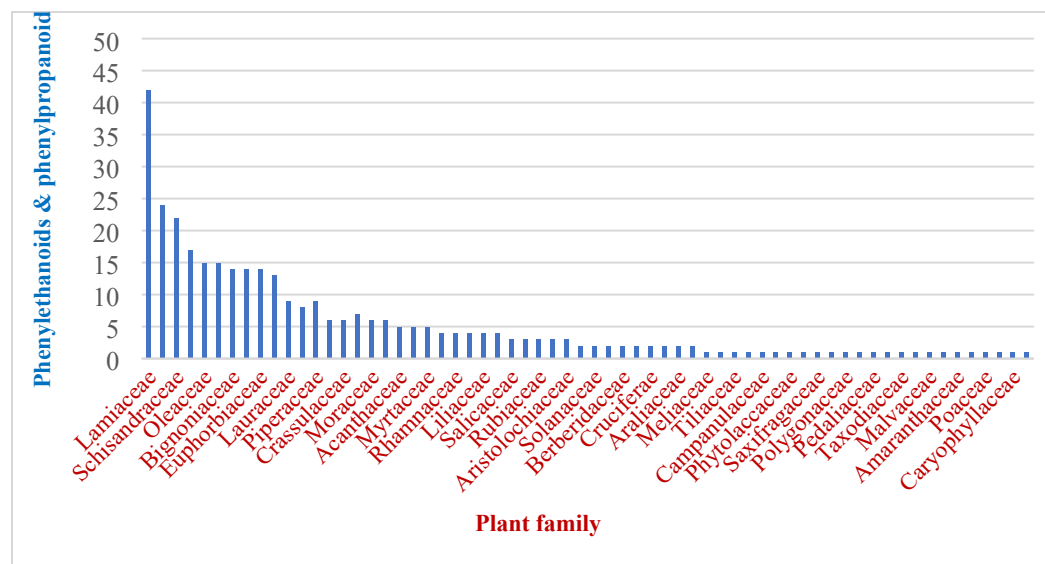


Fig. S1. Distribution of the investigated 261 compounds in different plant families

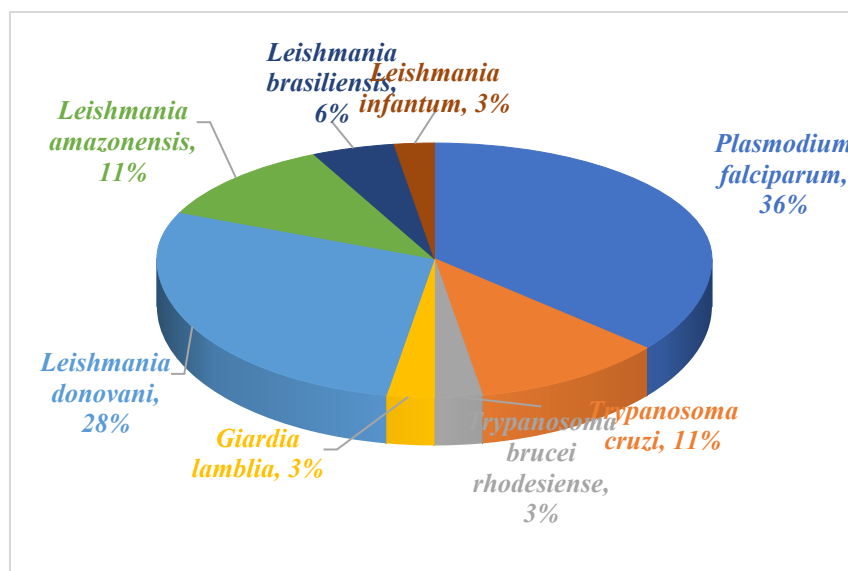


Fig. S2. Different antiprotozoal activity of 27 compounds out of the investigated 261 compounds

Feddeiphenol A

Feddeiphenol C

Forsythoside A

L-Chicoric acid

Niranthin

Globoidnan A

(-)-Isolariciresinol

Peperomin A

PVB

Fig. S3. Presumptive mode of interaction of test molecules and PVB within active site of *P. falciparum* kinase (PDB ID: 1V0P)

Rosmarinic acid

Forsythoside A

Di-E-caffeoyl-meso-tartaric acid

Globoidnan A

Sesamin

AOQ

Fig S4. Presumptive mode of interaction of test molecules and AOQ within active site of *P. falciparum* mitochondrial cytochrome bc1 complex (PDB ID: 4PD4).

Feddeiphenol C

Surinamensin

Kadangustin H

Globoidnan A

9X0

Fig. S5. Presumptive mode of interaction of test molecules and 9X0 within active site of *P. falciparum* lysyl-tRNA synthetase (PfKRS1; PDB ID: 6AGT)