

# Supporting Information for

## Development of a green, concise Synthesis of Nicotinamide Derivatives Catalysed by Novozym<sup>®</sup> 435 from *Candida antarctica* in Sustainable Continuous- Flow Microreactors

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### Materials

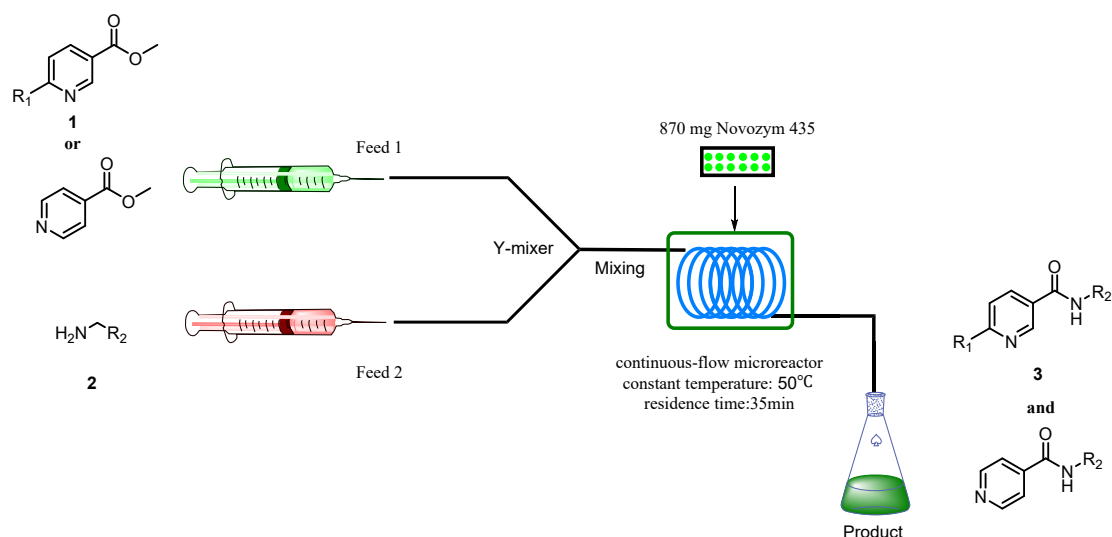
All chemicals in this study were obtained from commercial sources and did not require further purification. Lipozyme TL IM (immobilized *Thermomyces lanuginosus*), Novozym<sup>®</sup> 435 (immobilized *Candida antarctica* lipase B) were purchased from Novo Nordisk Bioindustry (Copenhagen, Denmark). Methyl nicotinate, methyl 6-chloronicotinate, isobutylamine, 4-methoxybenzylamine, methylamine solution, ethylamine were purchased from Aladdin (Shanghai, China). Benzylamine, 4-chlorobenzylamine were purchased from Energy Chemical (Shanghai, China). Methyl isonicotinate was purchased from Macklin (Shanghai, China). Harvard Instrument PHD 2000 syringe pump was purchased from Harvard University (Holliston, Massachusetts, USA). The flow reactor and Y-mixer were purchased from Beijing Haigui Medical Engineering Design Co., Ltd (Beijing China).

### Purification of the product

When the conversion of the nicotinamide derivative reaches a maximum (determined by TLC), the reaction is terminated by filtering the enzyme, and the tert-amyl alcohol solvent is rotary evaporated under reduced pressure. The product is separated by silica gel chromatography (mobile phase petroleum ether / ethyl acetate, 5/1 to 1/1). Purification was monitored by TLC. The graded fractions containing the major product were combined, the solvent evaporated and the residue analyzed by <sup>1</sup>H NMR, <sup>13</sup>C NMR.

## Experimental setup

A continuous-flow microreactor equipment diagram which was used for synthesis of nicotinamide derivatives from methyl nicotinate derivatives and amines catalyzed by Novozym® 435 is described in Figure S1. The experimental setup consists of a syringe pump, two substrate injectors, Y-shaped mixers ( $\phi=1.8$  mm) and a product collector. Syringe pumps (Harvard apparatus PHD 2000) were used to introduce separate feed streams to the flow reactor with  $100\text{cm} \times 2$  mm PFA tubing. Silica gel tubes were filled with Novozym® 435 and immersed in a constant temperature water bath to control the temperature. A total of 5 mmol of methyl nicotinate derivatives were dissolved in 10 mL of tert-amyl alcohol (feed 1), and 10 mmol of amines were dissolved in 10 mL of tert-amyl alcohol (feed 2). Feeds 1 and 2 were placed in separate 10 mL feeders and mixed at a flow rate of  $17.8 \mu\text{L min}^{-1}$  in a Y-mixer at  $50^\circ\text{C}$ . The resulting stream ( $17.8 \mu\text{L min}^{-1}$ ) was connected to a sample vial for collection of the final mixture.



**Figure S1.** The equipment diagram for the synthesis of nicotinamide derivatives in the continuous-flow microreactor catalyzed by lipase Novozym® 435.

## General Procedure for the synthesis of nicotinamide derivatives from methyl nicotinate derivatives and amines catalyzed by Novozym® 435 in Continuous-Flow Microreactors

**Method A:** 5.0 mmol of the methyl nicotinate derivatives was dissolved in 10 mL *tert*-amyl alcohol (feed A, ~0.5 M) and 10.0 mmol amine were dissolved in 10 mL *tert*-amyl alcohol (feed B; ~1.0 M). Novozym® 435 (0.87 g) were filled in PFA reactor coil (inner diameter ID= 2.0 mm, length = 100cm.). Streams A and B were mixed together at a flow rate of 8.9  $\mu\text{L min}^{-1}$  in a Y-mixer at 50 °C and the resulting stream (17.8  $\mu\text{L min}^{-1}$ ) was connected to a sample vial which was used to collect the final mixture. The final mixture was then evaporated, and the residue was submitted to column chromatography on silica gel (200-300 mesh). The crude product was purified by silica gel column chromatography with a petroleum ether / ethyl acetate gradient from 5:1 to 1:1. The purification was monitored by TLC. The fractions containing the main products were pooled, the solvent evaporated, and the residue analyzed by  $^1\text{H NMR}$ , and  $^{13}\text{C NMR}$ .

## General Procedure for the synthesis of nicotinamide derivatives from methyl nicotinate derivatives and amines catalyzed by Novozym® 435 under Shaker Conditions

**Method B:** methyl nicotinate derivatives (5.0 mmol) and amine (10.0 mmol) were added to 20 mL *tert*-amyl alcohol. The biocatalyst Novozym® 435 (0.87 g) was then added and the suspension maintained at 50 °C for 24h under Shaker Conditions (200  $\text{r}\cdot\text{min}^{-1}$ ). The mixture was cooled and filtered. Then evaporated under reduced pressure and the residue was submitted to column chromatography on silica gel (200– 300 mesh). The crude product was purified by silica gel column chromatography with a petroleum ether /ethyl acetate gradient from 5:1 to 1:1. The purification was monitored by TLC. The fractions containing the main products were pooled, the solvent evaporated, and the residue analyzed by  $^1\text{H NMR}$ , and  $^{13}\text{C NMR}$ .

## Calculation of Green Chemistry Metrics.<sup>1-3</sup>

The following formulae were used for evaluating green chemistry metrics such as atom economy (AE), carbon efficiency (CE), atom efficiency, reaction mass efficiency (RME), overall efficiency (OE), process mass intensity (PMI) and E-factor.

1. % *Atom Economy (AE)* = (Mol Wt. of desired product/ Mol Wt. of all reactants)  $\times$  100

2. % *Carbon Efficiency (CE)* = (Amount of carbon in the product)/ (Total no. of carbon present in reactant)  $\times$  100

3. % *Atom efficiency* = (Yield of the product) /Atom economy)  $\times$  100

4. *Reaction Mass Efficiency (%) = (RME)* = (Mass of isolated product)/ Total mass of reactants)  $\times$  100

5. *Overall Efficiency (%) = (OE)* = (RME/AE)  $\times$  100

6. **Process Mass Intensity (%) = (PMI) = (Total mass of input materials in a process) / Mass of product) × 100**

7. **E-Factor = PMI-1 = (Mass of waste) / Mass of product)**

**Table S1.** Evaluation of green chemistry metrics for the synthesized compound **3a**

Reactant 1	Methyl nicotinate	0.686g	5 mmol	FW 137.14
Reactant 2	Isobutylamine	0.731g	10 mmol	FW 73.14
Solvent	<i>Tert</i> -amyl alcohol	16.520g (20 ml)	-----	
Enzyme	Novozym® 435	0.870g	-----	
Product	N-isobutyl-nicotinamide	0.768g	4.31 mmol	FW 178.24

Product yield = 86.2%

E-factor =  $(0.686 + 0.731 + 0.870 + 16.52 - 0.768) / 0.768 \times 100\% = 23.49$  Kg waste/1Kg product.

Atom economy =  $(178.24 / 210.28) \times 100\% = 84.76\%$

Atom efficiency =  $86.2\% \times 84.76\% / 100\% = 73.06\%$

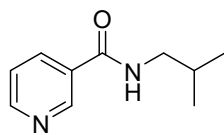
Carbon efficiency =  $10/10 \times 100\% = 100\%$

Reaction mass efficiency =  $0.768 / (0.686 + 0.731) \text{ g} \times 100\% = 54.20\%$

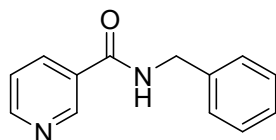
Overall Efficiency =  $54.20\% / 84.76\% \times 100\% = 63.95\%$

Process Mass Intensity =  $(0.686 + 0.731 + 0.870 + 16.52) / 0.768 = 24.49$  kg /kg of the product.

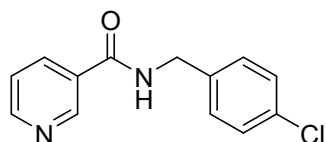
## Experimental data of products



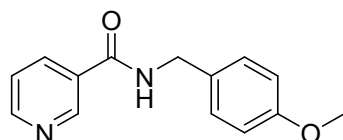
N-isobutyl-nicotinamide (3a). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.00 (d,  $J = 2.2$  Hz, 1H), 8.73 – 8.63 (m, 2H), 8.18 (dt,  $J = 7.9, 2.0$  Hz, 1H), 7.50 (dd,  $J = 7.9, 4.8$  Hz, 1H), 3.10 (dd,  $J = 7.0, 5.8$  Hz, 2H), 1.85 (hept,  $J = 6.7$  Hz, 1H), 0.90 (d,  $J = 6.7$  Hz, 6H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  165.30, 152.16, 148.79, 135.40, 130.65, 123.89, 47.16, 28.54, 20.67.



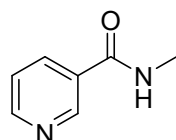
N-benzyl-nicotinamide(3b). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.30 (t,  $J = 6.0$  Hz, 1H), 9.12 (d,  $J = 2.3$  Hz, 1H), 8.73 (dd,  $J = 4.8, 1.7$  Hz, 1H), 8.27 (dt,  $J = 7.9, 2.0$  Hz, 1H), 7.51 (dd,  $J = 8.0, 4.8$  Hz, 1H), 7.40 – 7.30 (m, 4H), 7.30 – 7.21(m, 1H), 4.55 (d,  $J = 6.0$  Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  165.35, 152.36, 148.95, 139.76, 135.50, 130.30, 128.80, 127.76, 127.31, 123.92, 43.17.



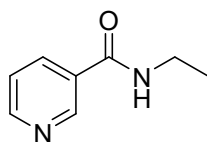
N-[(4-chlorophenyl) methyl] pyridine-3-carboxamide(3c). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.28 (t,  $J = 5.9$  Hz, 1H), 9.05 (dd,  $J = 2.4, 0.9$  Hz, 1H), 8.72 (dd,  $J = 4.8, 1.7$  Hz, 1H), 8.23 (ddd,  $J = 7.9, 2.3, 1.7$  Hz, 1H), 7.53 (ddd,  $J = 8.0, 4.8, 0.9$  Hz, 1H), 7.44 – 7.33 (m, 4H), 4.49 (d,  $J = 5.9$  Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.66, 151.79, 148.22, 138.15, 134.81, 131.19, 129.45, 128.96, 128.08, 123.30, 41.80.



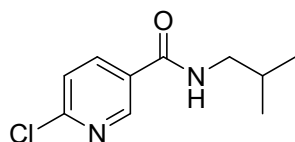
N-[(4-methoxyphenyl) methyl] pyridine-3-carboxamide(3d). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.18 (t,  $J = 5.9$  Hz, 1H), 9.04 (dd,  $J = 2.4, 0.9$  Hz, 1H), 8.71 (dd,  $J = 4.8, 1.7$  Hz, 1H), 8.22 (dt,  $J = 8.0, 1.9$  Hz, 1H), 7.51 (ddd,  $J = 8.0, 4.8, 0.9$  Hz, 1H), 7.31 – 7.23 (m, 2H), 6.94 – 6.86 (m, 2H), 4.44 (d,  $J = 5.9$  Hz, 2H), 3.73 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  165.12, 158.73, 152.34, 148.89, 135.45, 131.71, 130.32, 129.14, 123.93, 114.19, 55.52, 42.57.



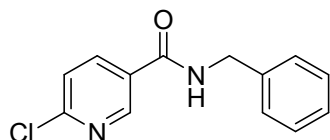
N-methyl nicotinamide(3e). White power.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  8.99 (dd,  $J = 2.3, 0.9$  Hz, 1H), 8.70 (dd,  $J = 4.8, 1.7$  Hz, 1H), 8.64 (s, 1H), 8.17 (dt,  $J = 7.9, 2.0$  Hz, 1H), 7.50 (ddd,  $J = 7.9, 4.8, 1.0$  Hz, 1H), 2.81 (d,  $J = 4.6$  Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  165.62, 152.19, 148.72, 135.25, 130.40, 123.92, 26.65.



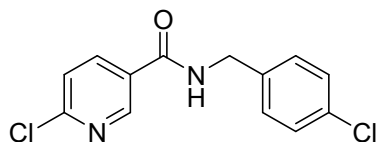
N-ethyl nicotinamide(3f). White powder.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.04 – 8.98 (m, 1H), 8.68 (dt,  $J = 4.8, 2.4$  Hz, 2H), 8.18 (dt,  $J = 7.9, 2.0$  Hz, 1H), 7.49 (ddd,  $J = 7.9, 4.8, 0.9$  Hz, 1H), 3.31 (qd,  $J = 7.2, 5.5$  Hz, 2H), 1.13 (t,  $J = 7.3$  Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.96, 152.13, 148.77, 135.30, 130.53, 123.84, 34.55, 15.09.



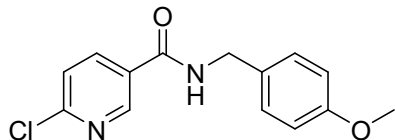
6-chloro-N-isobutyl nicotinamide(3g). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  8.83 (dd,  $J = 2.4, 0.7$  Hz, 1H), 8.72 (t,  $J = 5.9$  Hz, 1H), 8.24 (dd,  $J = 8.3, 2.5$  Hz, 1H), 7.64 (dd,  $J = 8.3, 0.7$  Hz, 1H), 3.10 (dd,  $J = 7.0, 5.8$  Hz, 2H), 1.84 (dq,  $J = 13.5, 6.7$  Hz, 1H), 0.90 (d,  $J = 6.7$  Hz, 6H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.17, 152.83, 149.37, 139.04, 130.04, 124.53, 47.23, 28.50, 20.66.



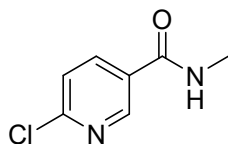
N-benzyl-6-chloro-nicotinamide(3h). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.31 (t,  $J = 6.0$  Hz, 1H), 8.89 (d,  $J = 2.5$  Hz, 1H), 8.29 (dd,  $J = 8.3, 2.5$  Hz, 1H), 7.67 (d,  $J = 8.3$  Hz, 1H), 7.34 (d,  $J = 4.4$  Hz, 4H), 7.31 – 7.22 (m, 1H), 4.51 (d,  $J = 5.9$  Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.18, 153.06, 149.49, 139.53, 139.12, 129.68, 128.83, 127.80, 127.39, 124.63, 43.18.



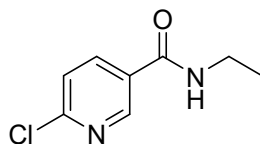
6-Chloro-N-(4-chloro-benzyl)-nicotinamide(3i). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.33 (t,  $J = 5.9$  Hz, 1H), 8.89 (d,  $J = 2.5$  Hz, 1H), 8.28 (dd,  $J = 8.3, 2.5$  Hz, 1H), 7.66 (dd,  $J = 8.3, 1.9$  Hz, 1H), 7.43 – 7.33 (m, 4H), 4.49 (d,  $J = 5.9$  Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.25, 153.14, 149.49, 139.10, 138.59, 131.95, 129.69, 129.54, 128.75, 124.62, 42.56.



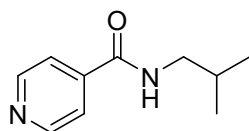
6-Chloro-N-(4-methoxy-benzyl)-nicotinamide(3j). White solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.23 (t,  $J = 5.9$  Hz, 1H), 8.87 (d,  $J = 2.4$  Hz, 1H), 8.27 (dd,  $J = 8.3, 2.5$  Hz, 1H), 7.65 (d,  $J = 8.3$  Hz, 1H), 7.31 – 7.20 (m, 2H), 6.94 – 6.86 (m, 2H), 4.43 (d,  $J = 5.8$  Hz, 2H), 3.73 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.03, 158.78, 153.01, 149.46, 139.09, 131.47, 129.74, 129.22, 124.60, 114.21, 55.53, 42.66.



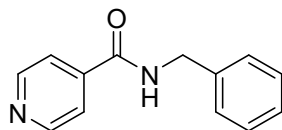
6-Chloro-N-methyl-nicotinamide(3k). White powder,  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.75 (d,  $J$  = 2.4 Hz, 1H), 8.10 (dd,  $J$  = 8.3, 2.5 Hz, 1H), 7.40 (d,  $J$  = 8.3 Hz, 1H), 6.81 (s, 1H), 3.02 (d,  $J$  = 4.7 Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  165.36, 154.08, 147.97, 138.01, 129.23, 124.39, 26.97.



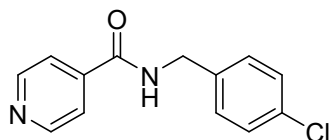
6-chloro-N-ethyl-3-Pyridinecarboxamide(3l). White powder.  
 $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.74 (d,  $J$  = 2.3 Hz, 1H), 8.09 (dd,  $J$  = 8.3, 2.3 Hz, 1H), 7.39 (d,  $J$  = 8.2 Hz, 1H), 6.72 (s, 1H), 3.55 – 3.43 (m, 2H), 1.25 (t,  $J$  = 7.2 Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  164.59, 154.00, 148.01, 138.01, 129.39, 124.33, 35.21, 14.72.



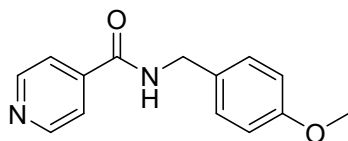
N-isobutyl-isonicotinamide(3m). White solid.  $^1\text{H}$  NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.80 – 8.66 (m, 3H), 7.79 – 7.69 (m, 2H), 3.11 (t,  $J$  = 6.4 Hz, 2H), 1.85 (dq,  $J$  = 13.5, 6.7 Hz, 1H), 0.90 (d,  $J$  = 6.7 Hz, 6H).  $^{13}\text{C}$  NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  164.77, 150.21, 141.69, 121.28, 46.78, 28.05, 20.20.



N-benzyl-isonicotinamide (3n). White solid.  $^1\text{H}$  NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.40 – 9.33 (m, 1H), 8.75 (d,  $J$  = 5.0 Hz, 2H), 7.82 (d,  $J$  = 5.1 Hz, 2H), 7.35 (d,  $J$  = 4.5 Hz, 4H), 7.27 (p,  $J$  = 5.8, 4.4 Hz, 1H), 4.52 (d,  $J$  = 6.1 Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  164.69, 150.29, 141.24, 139.11, 128.36, 127.29, 126.91, 121.28, 42.72.

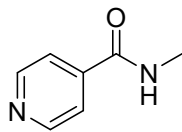


N-[(4-chlorophenyl) methyl]-4-Pyridinecarboxamide(3o). White solid.  $^1\text{H}$  NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.38 (t,  $J$  = 6.0 Hz, 1H), 8.81 – 8.68 (m, 2H), 7.86 – 7.77 (m, 2H), 7.45 – 7.32 (m, 4H), 4.50 (d,  $J$  = 5.9 Hz, 2H).  $^{13}\text{C}$  NMR (101 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  164.74, 150.31, 141.09, 138.16, 131.46, 129.19, 128.31, 121.26, 42.09.

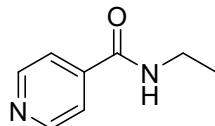


N-[(4-methoxyphenyl) methyl]- 4-Pyridinecarboxamide(3p). White solid.  $^1\text{H}$  NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.29 (t,  $J$  = 6.1 Hz, 1H), 8.74 (d,  $J$  = 5.2 Hz, 2H), 7.80 (d,  $J$  = 5.1 Hz, 2H), 7.27 (d,  $J$  = 8.1 Hz,

2H), 6.91 (d,  $J = 8.0$  Hz, 2H), 4.44 (d,  $J = 5.8$  Hz, 2H), 3.74 (d,  $J = 3.5$  Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.54, 158.31, 150.27, 141.32, 131.06, 128.71, 121.27, 113.75, 55.07, 42.20.



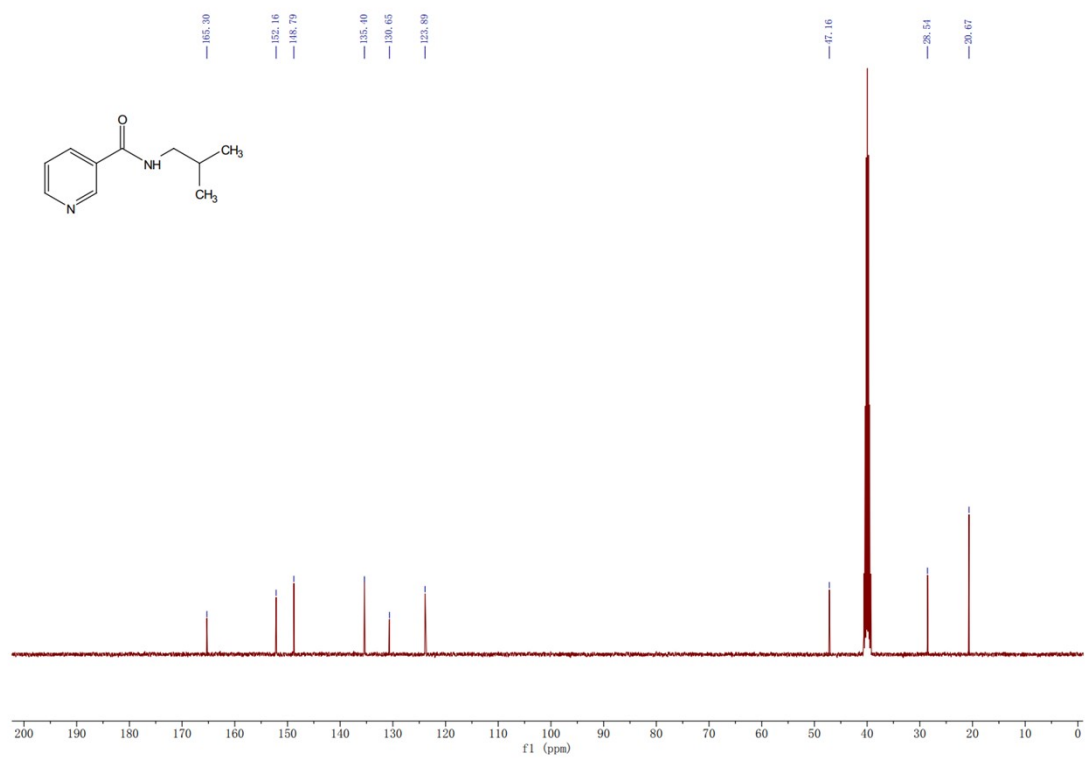
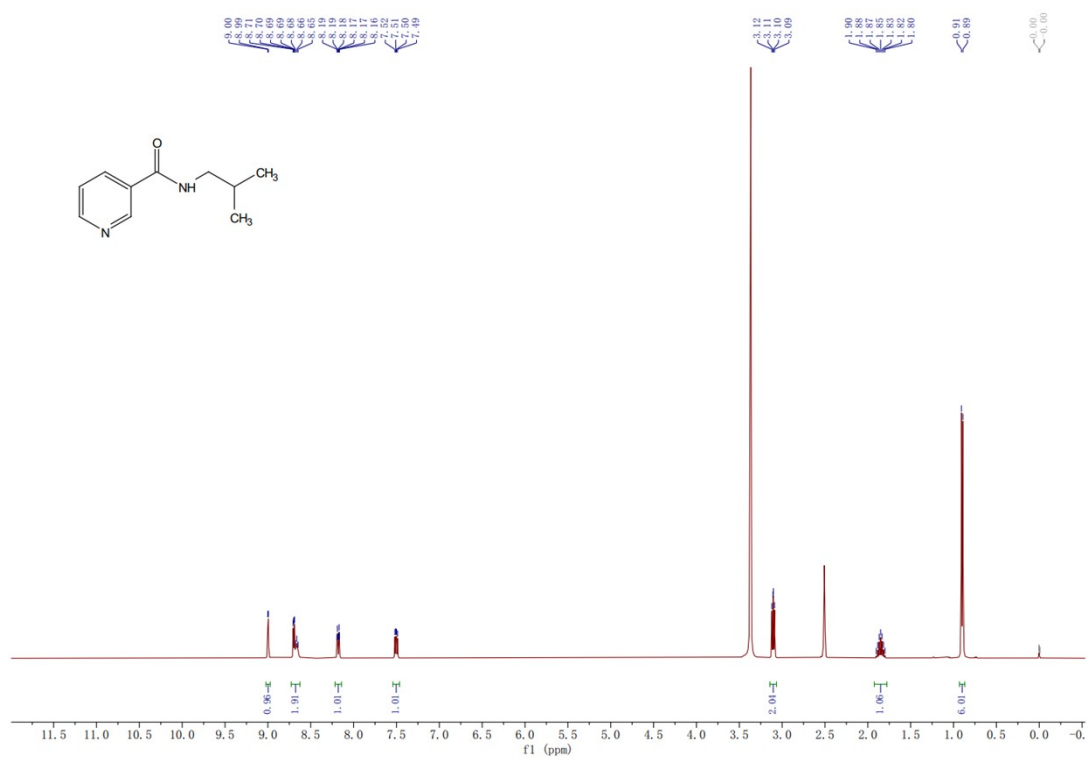
1-methylisonicotinamide(3q). White powder.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  8.77 – 8.69 (m, 3H), 7.77 – 7.71 (m, 2H), 2.81 (d,  $J = 4.6$  Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  165.46, 150.69, 141.83, 121.57, 26.71.



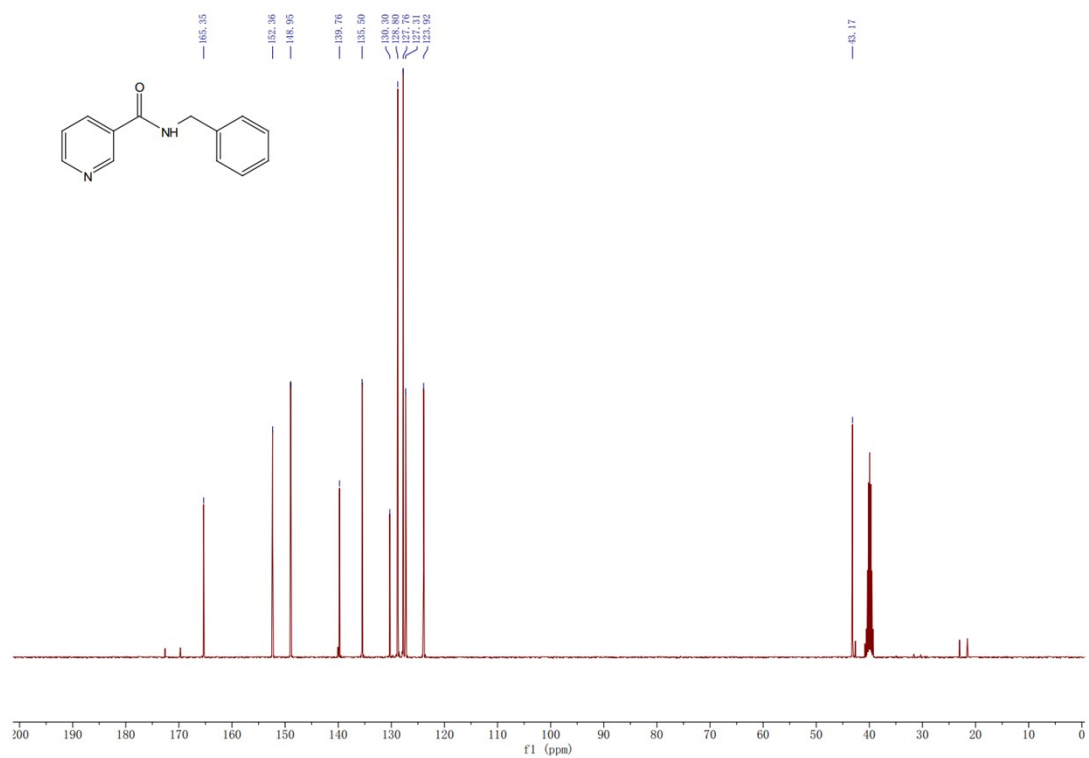
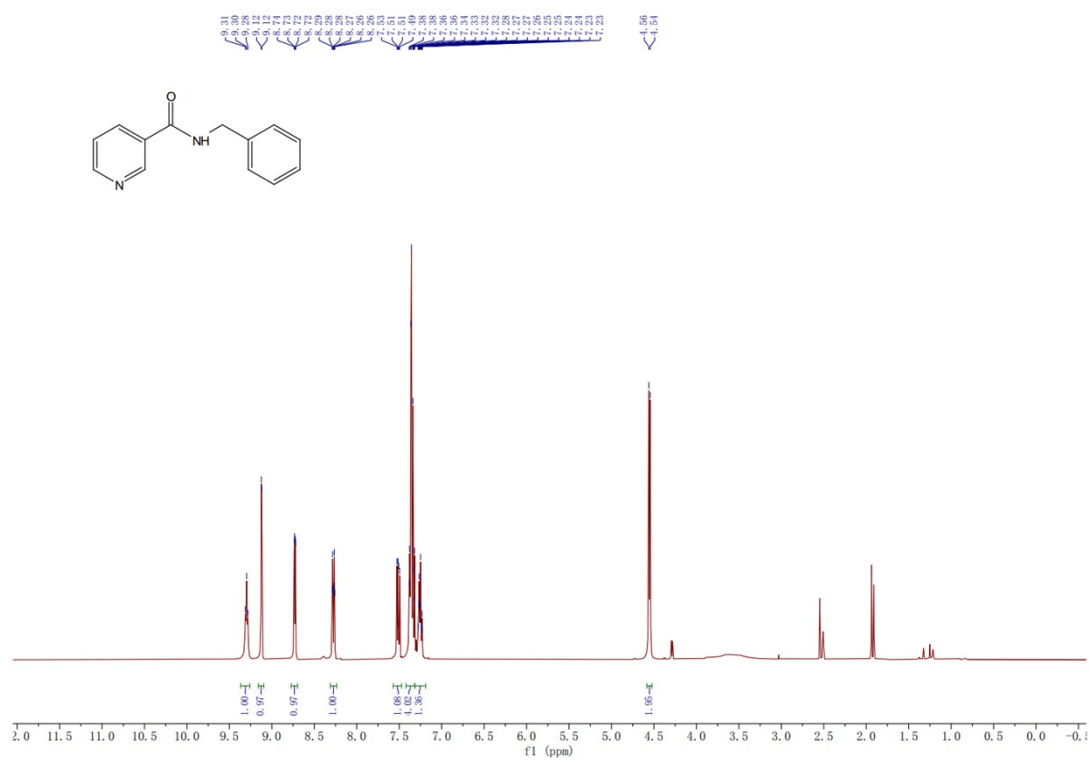
N-Ethylisonicotinamide: (3r). White powder.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  8.77 (t,  $J = 5.5$  Hz, 1H), 8.75 – 8.69 (m, 2H), 7.78 – 7.72 (m, 2H), 3.35 – 3.25 (m, 2H), 1.13 (t,  $J = 7.2$  Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.78, 150.66, 141.99, 121.64, 34.63, 15.04.



3a



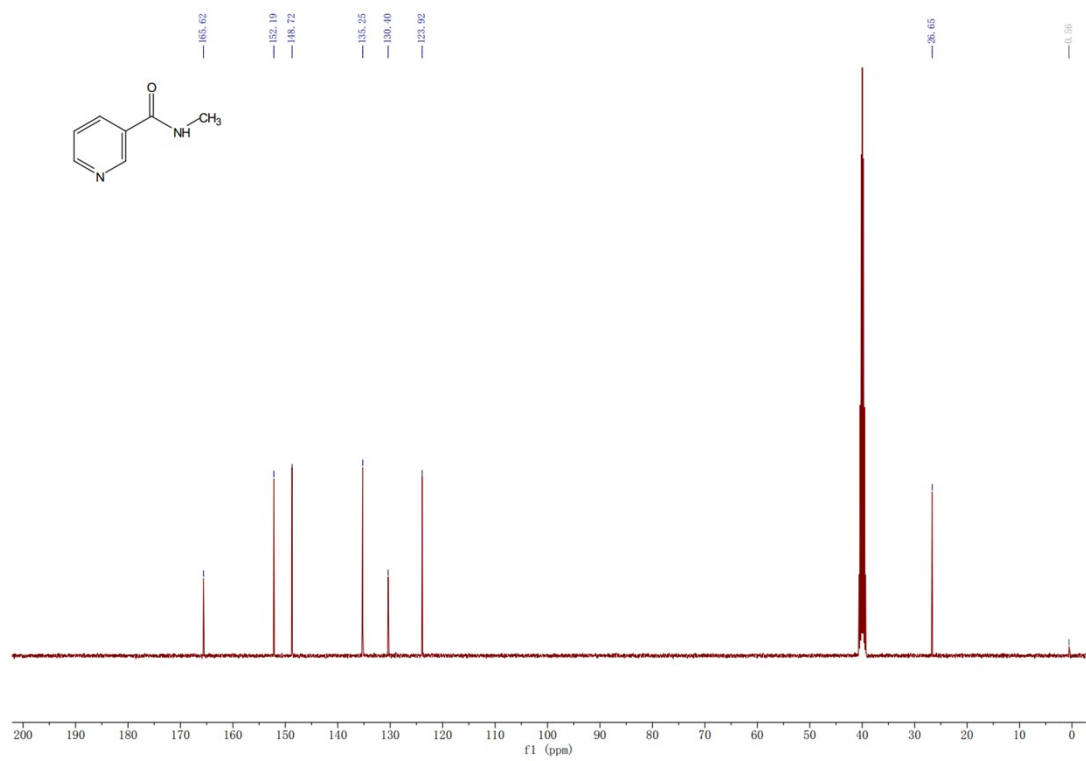
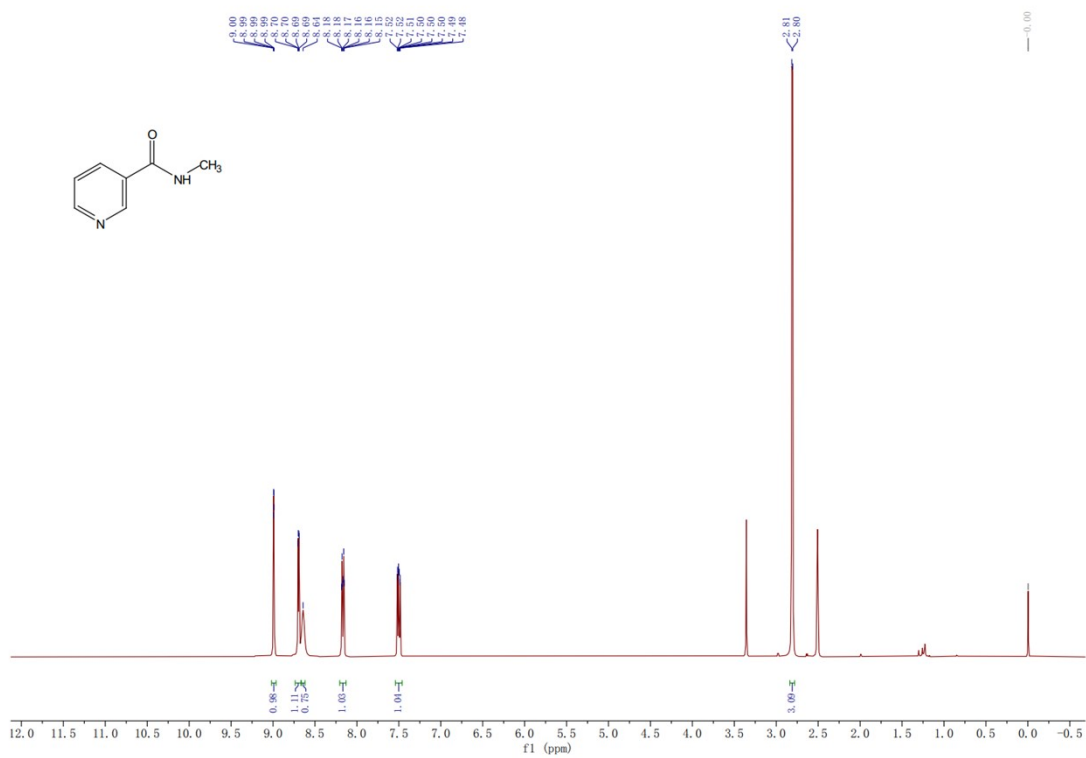
3b



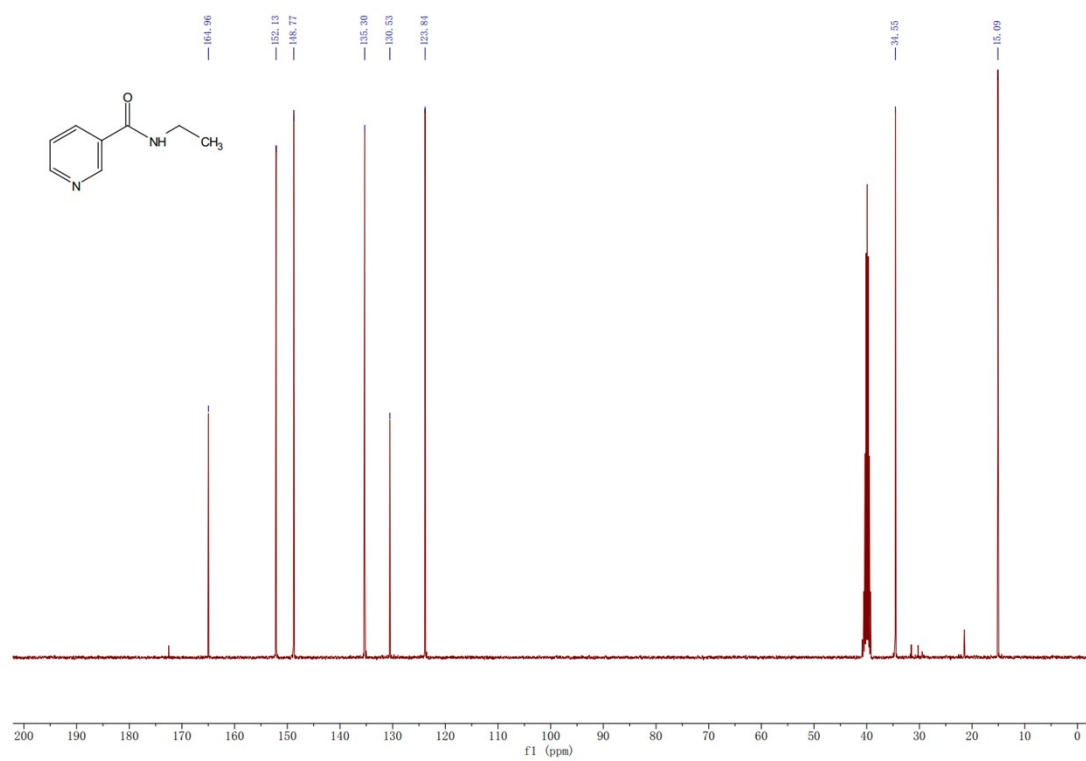
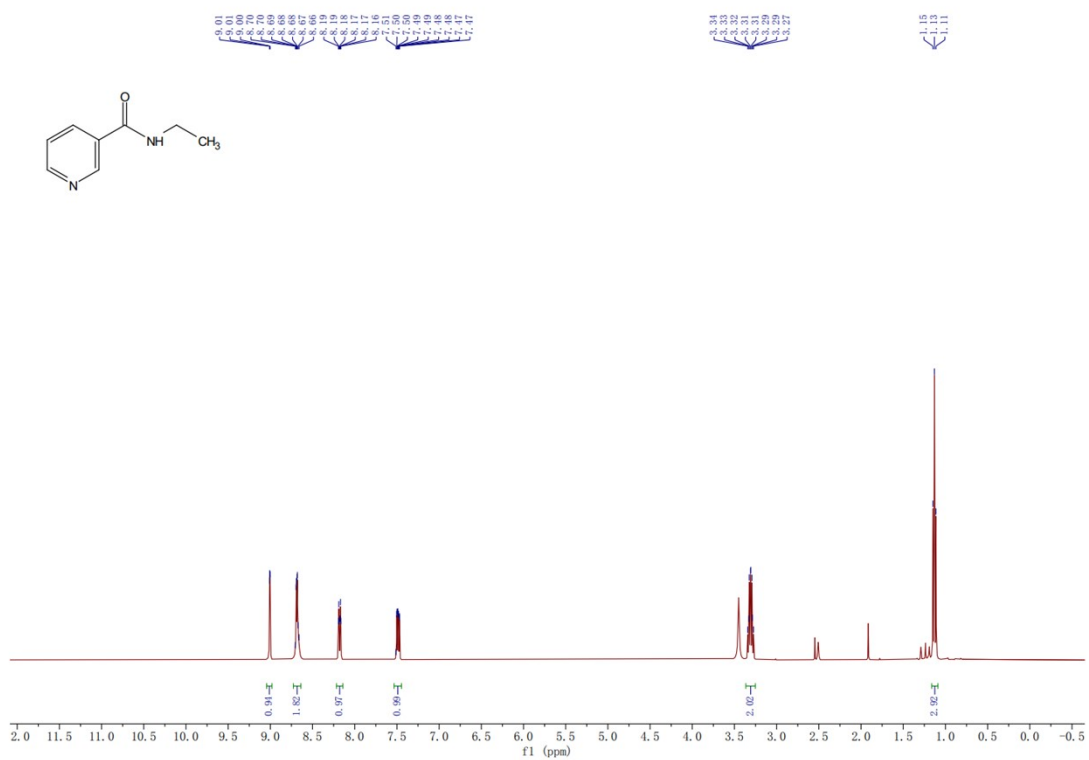




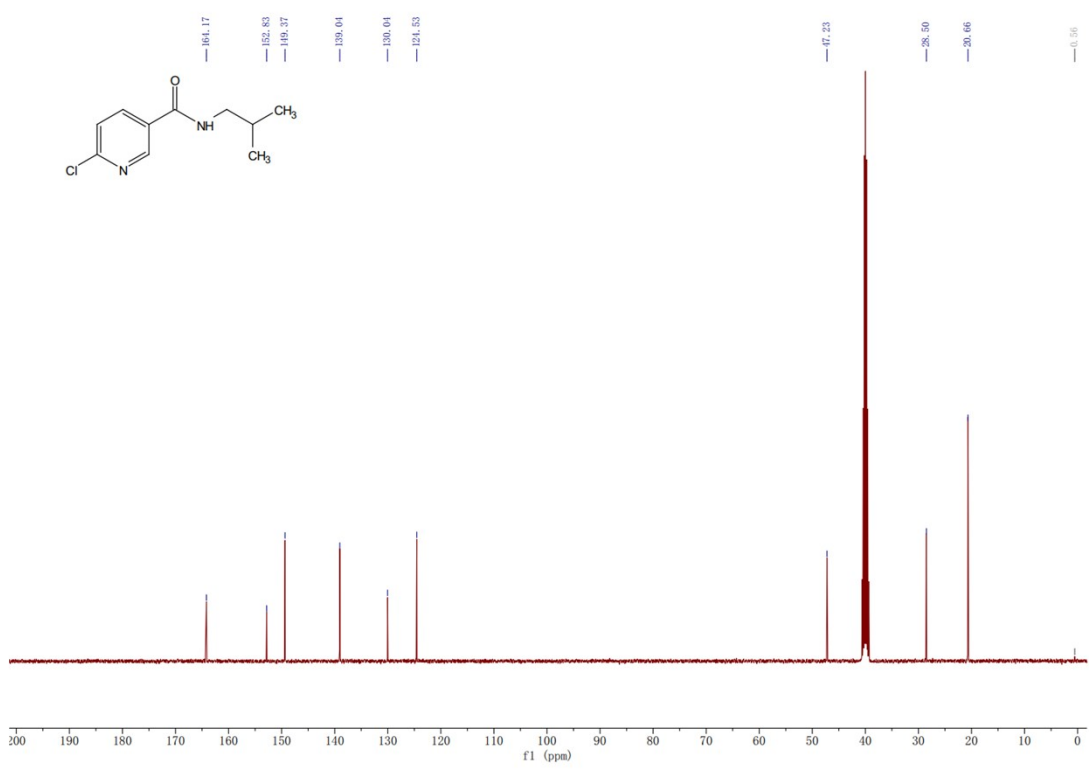
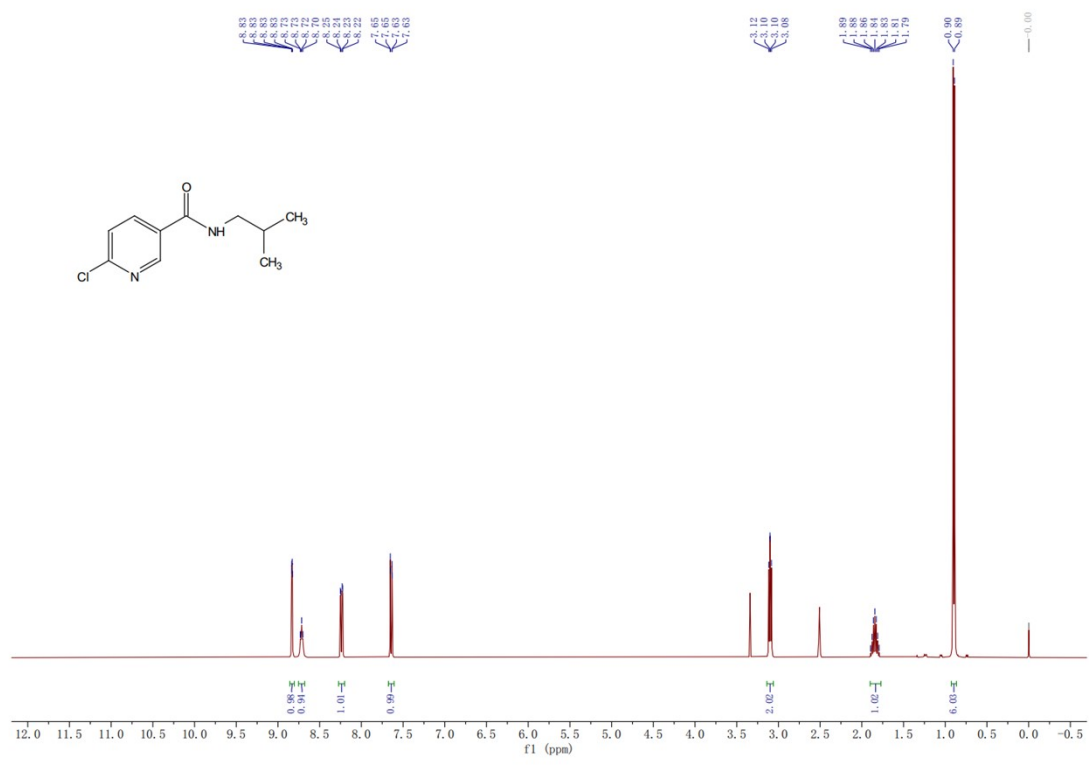
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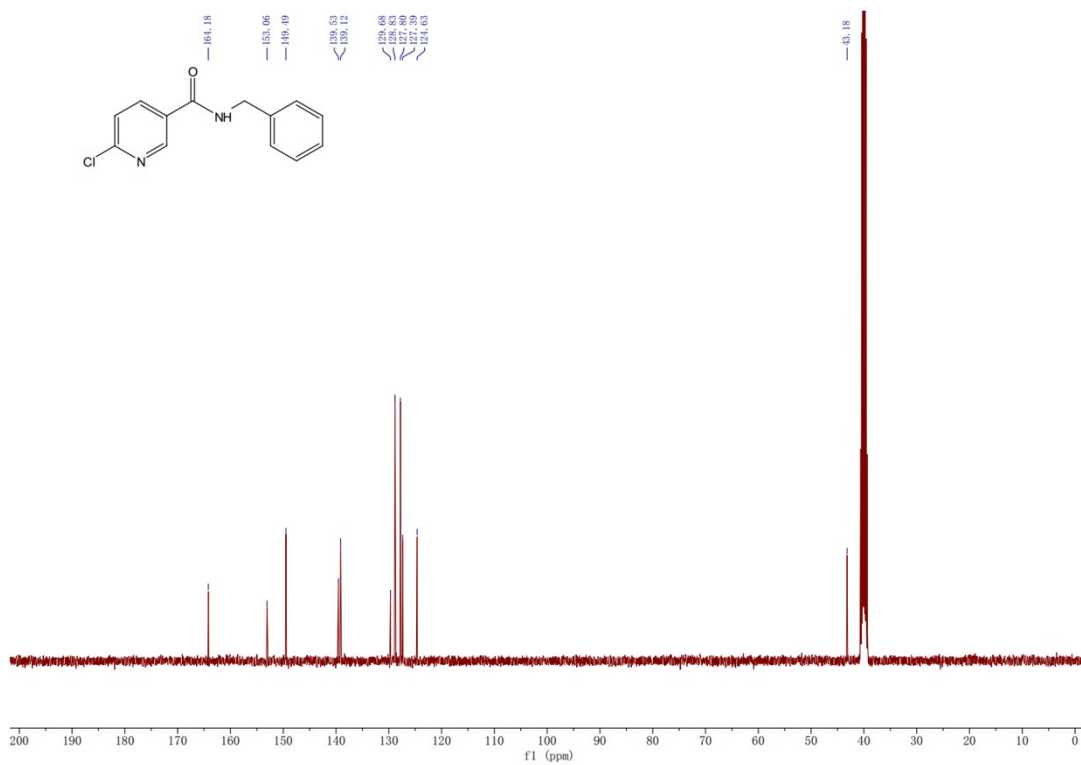
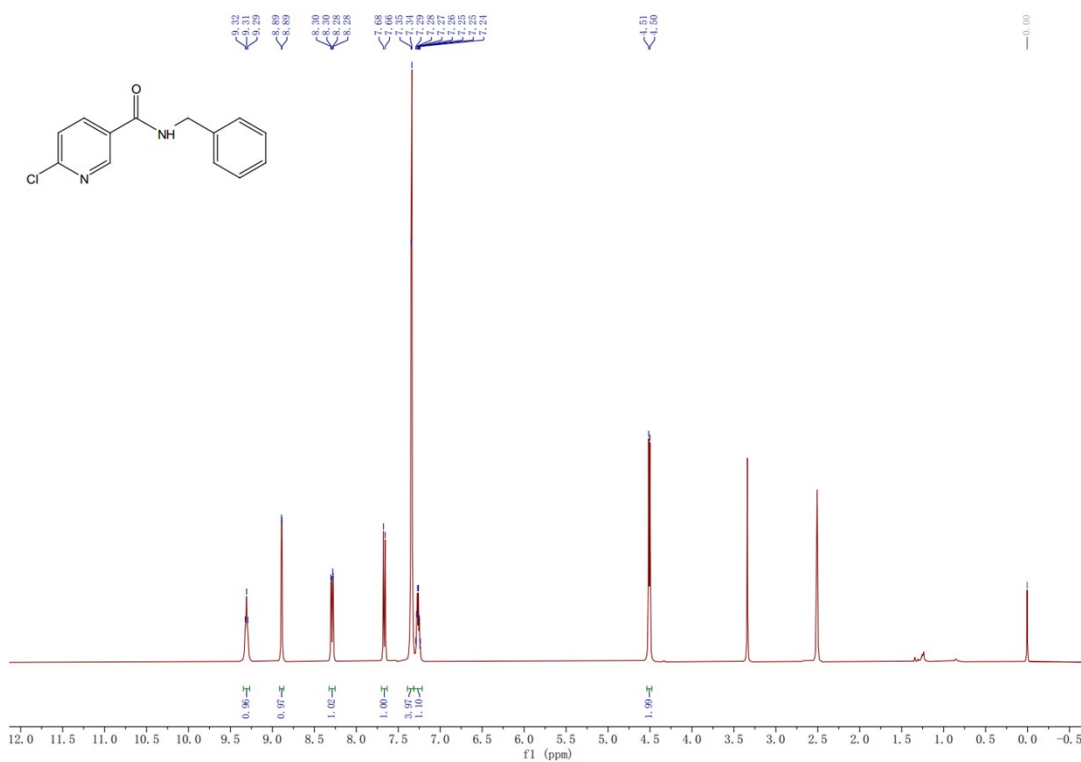
3f



3g

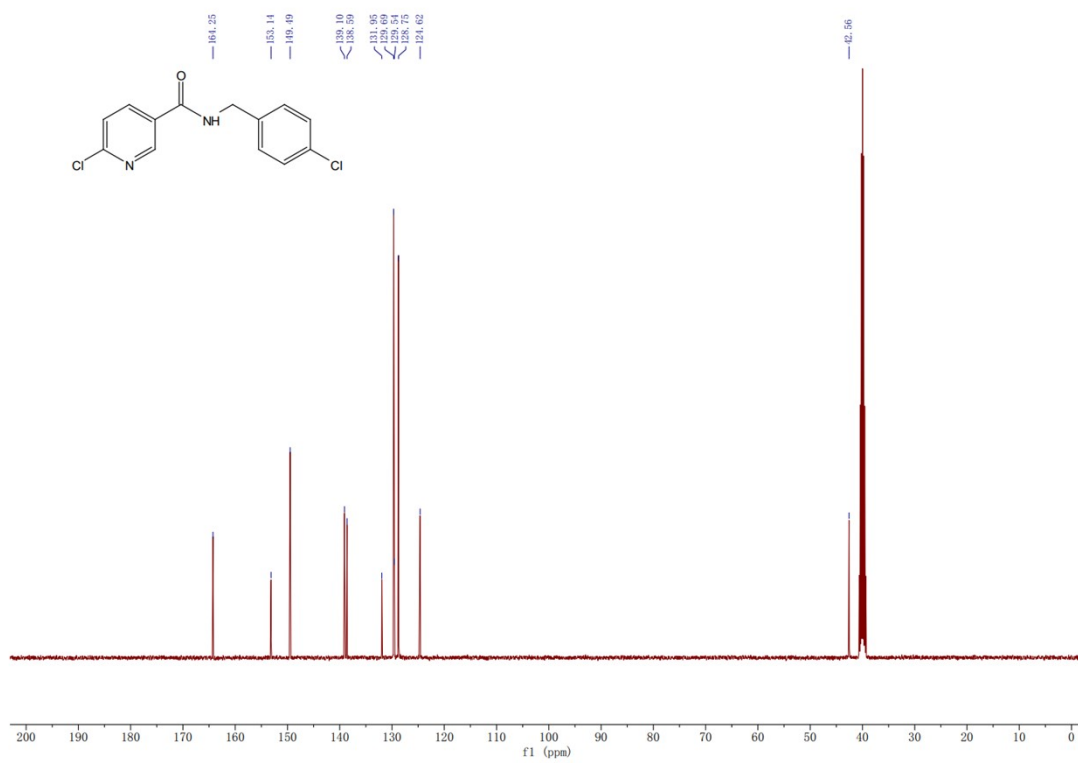
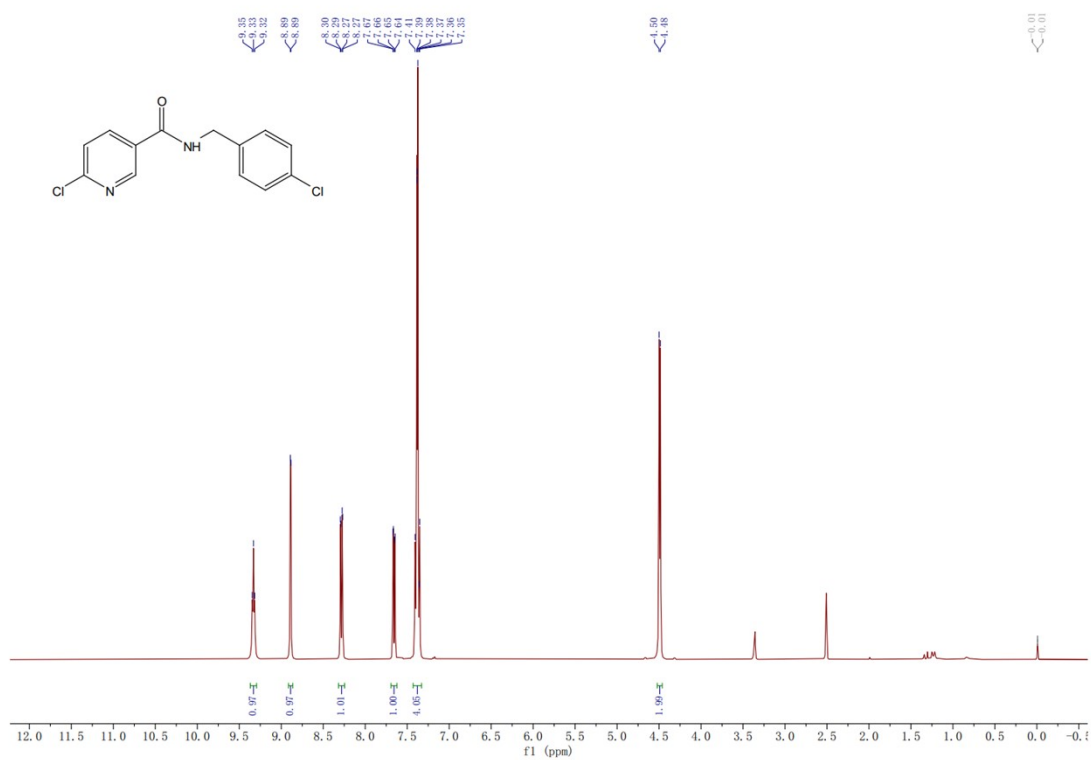


3h

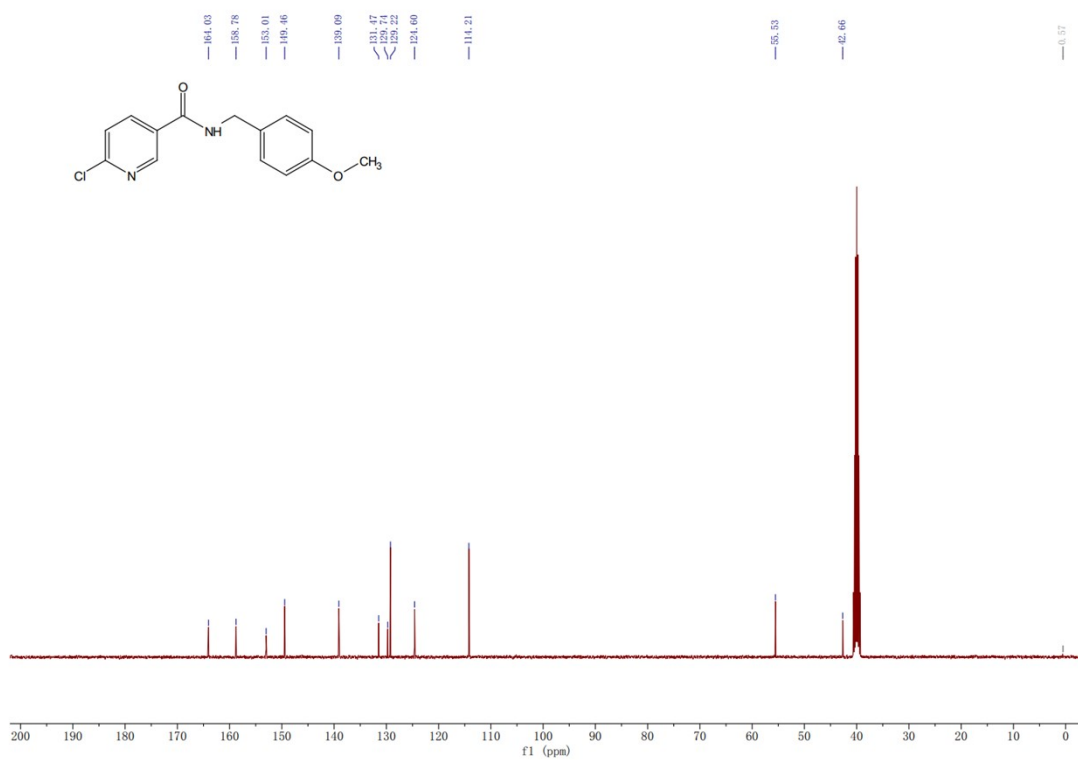
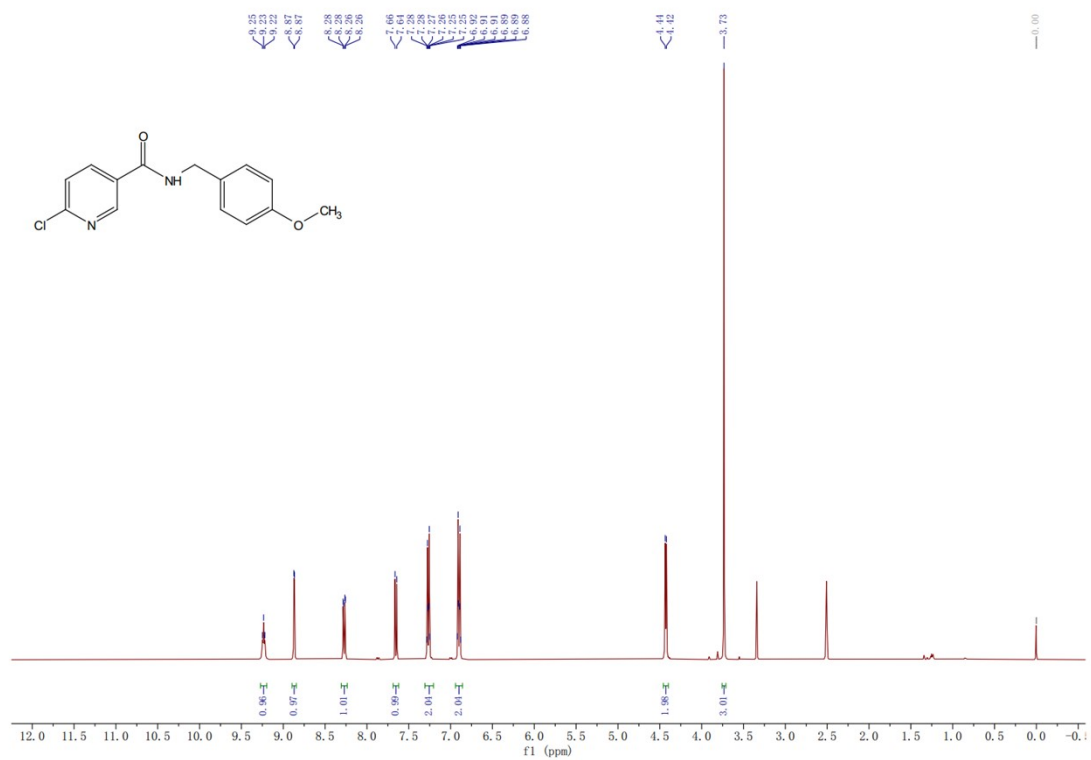




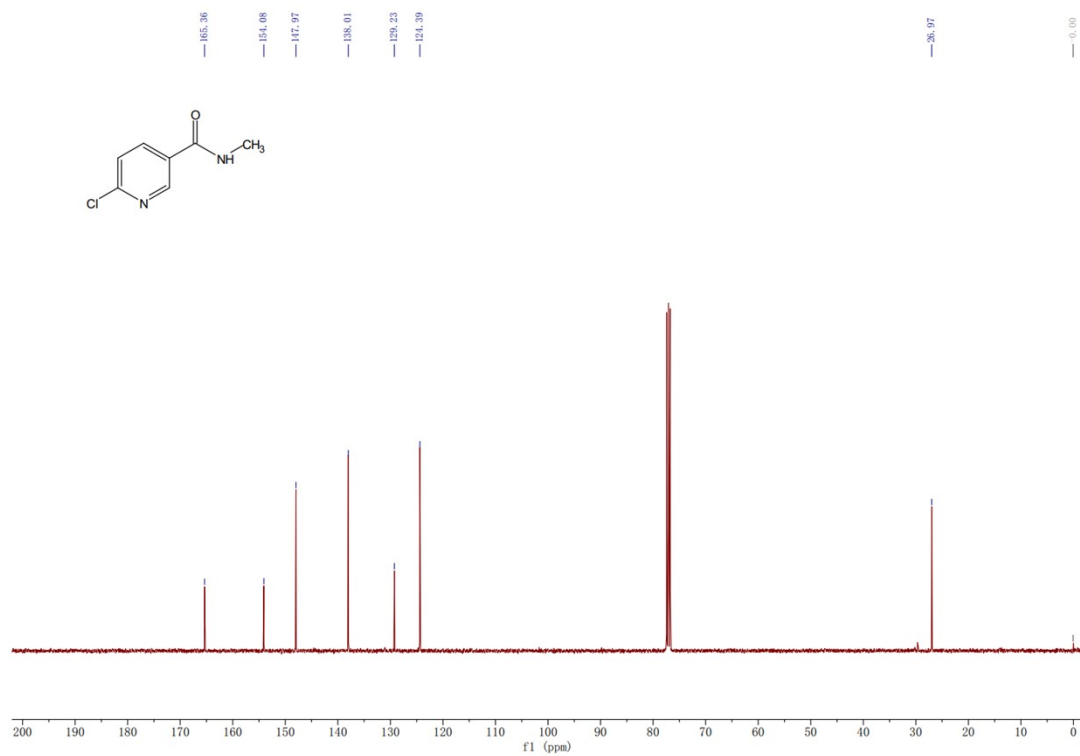
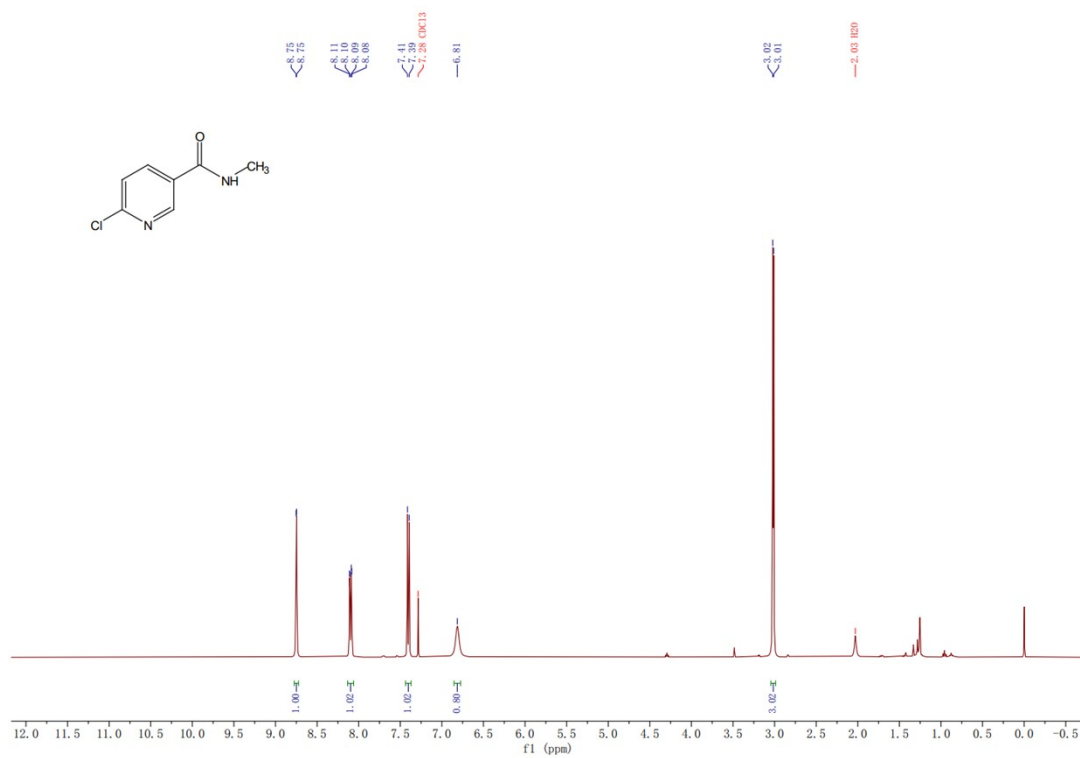
3i

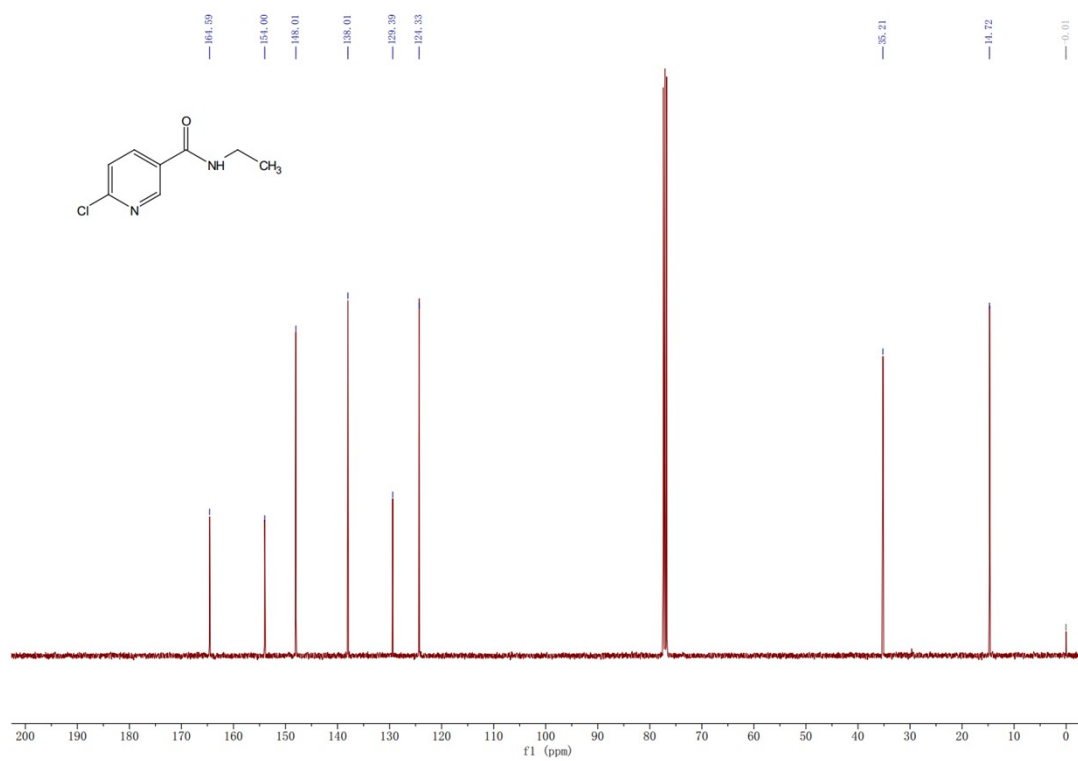
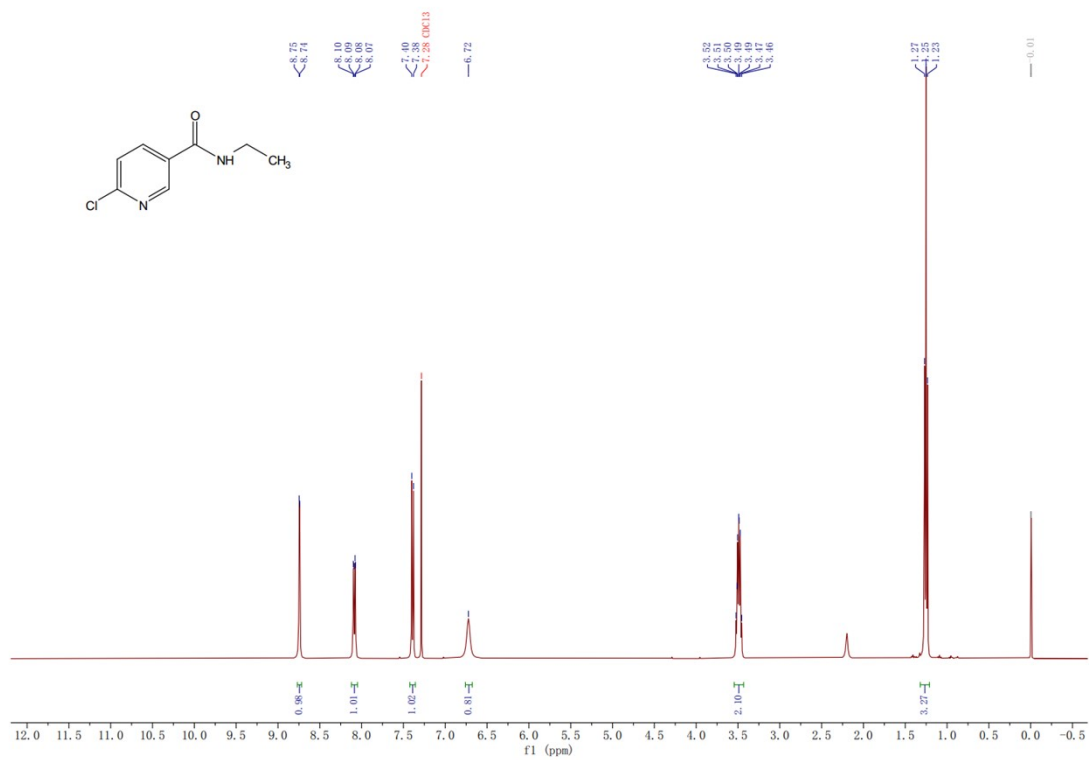


3j

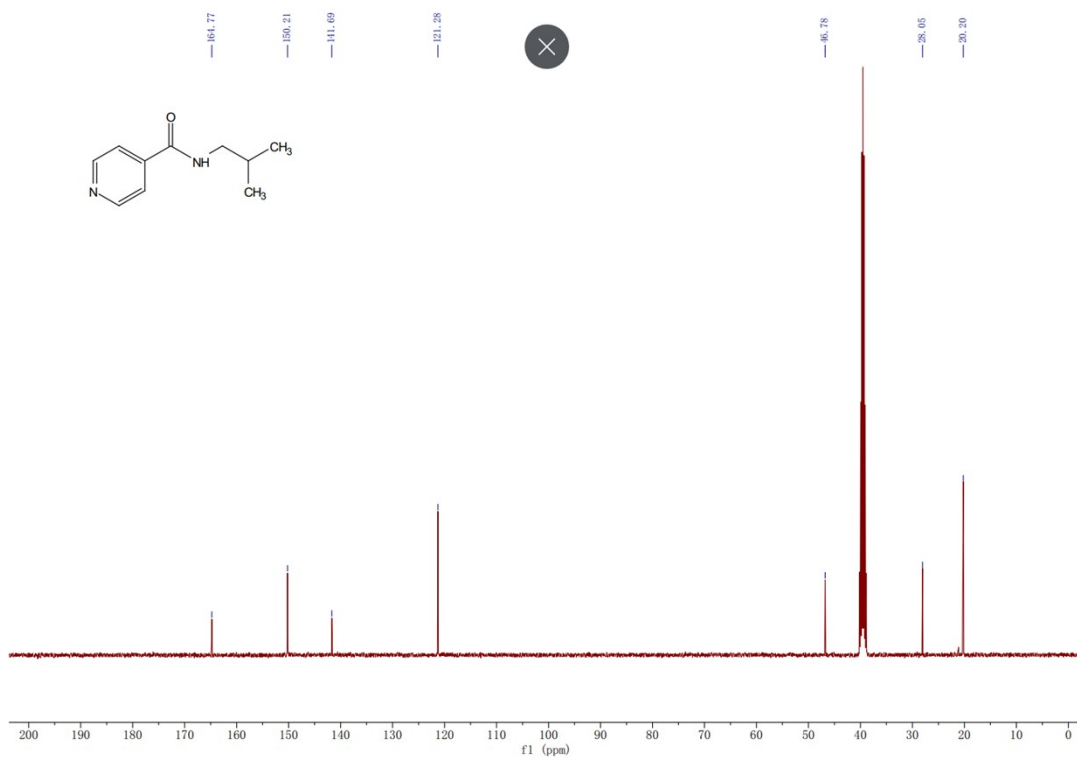
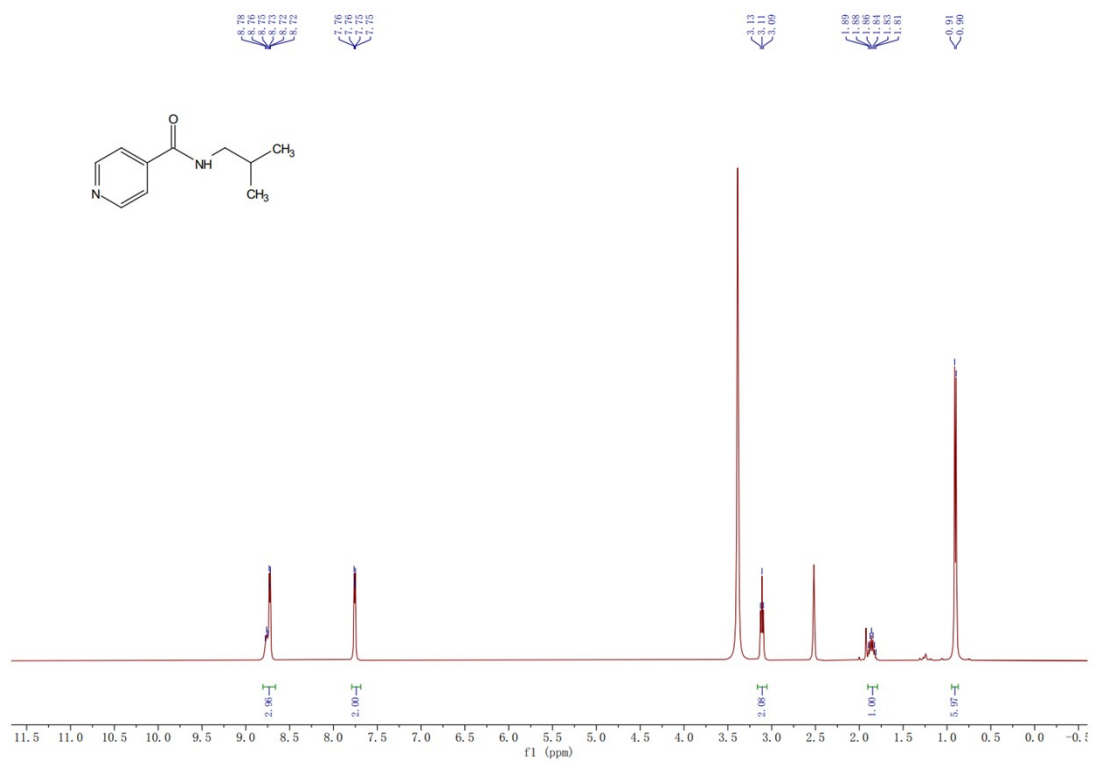


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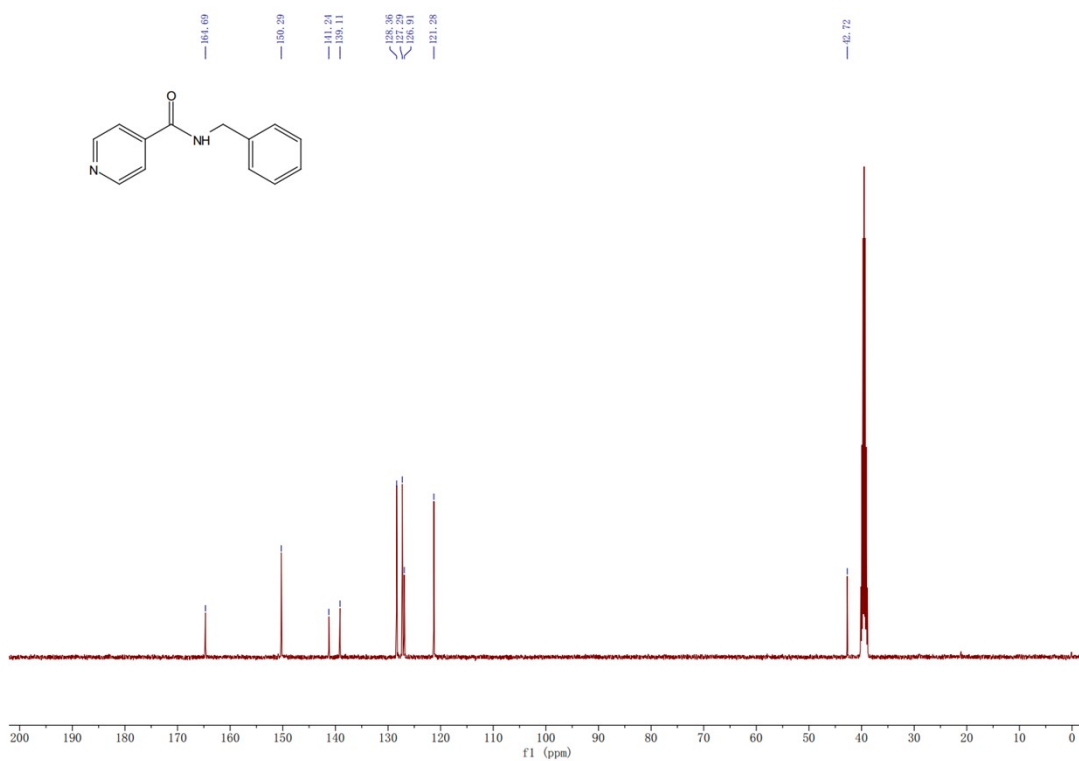
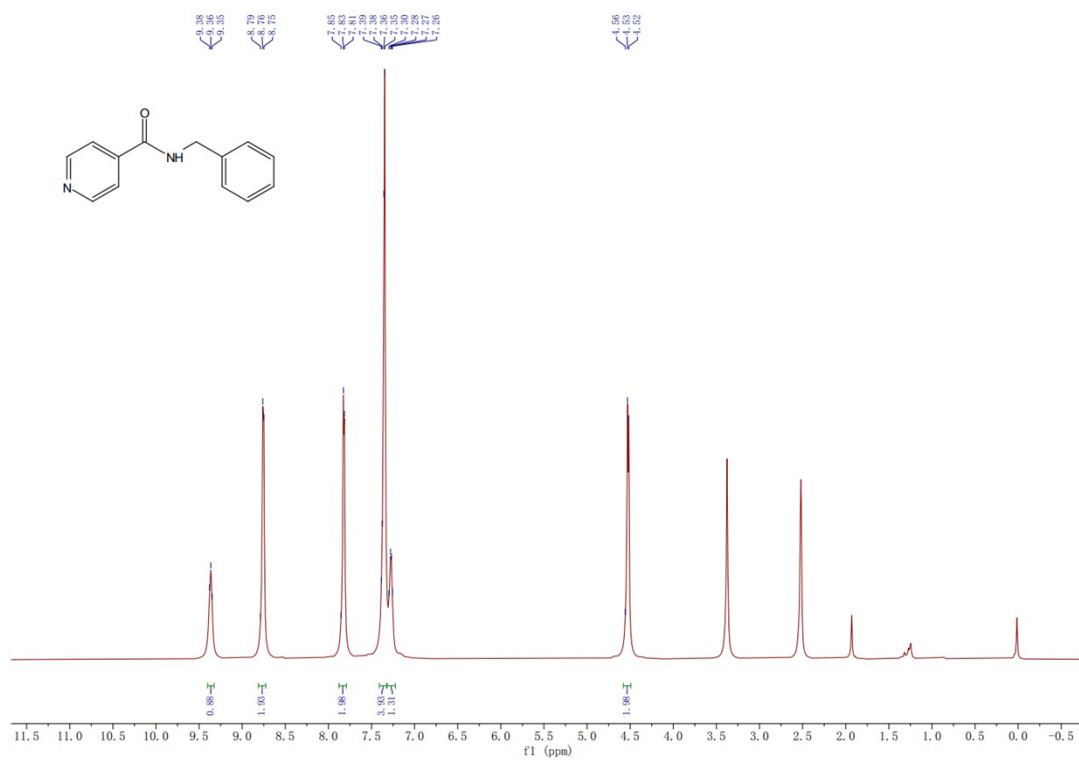




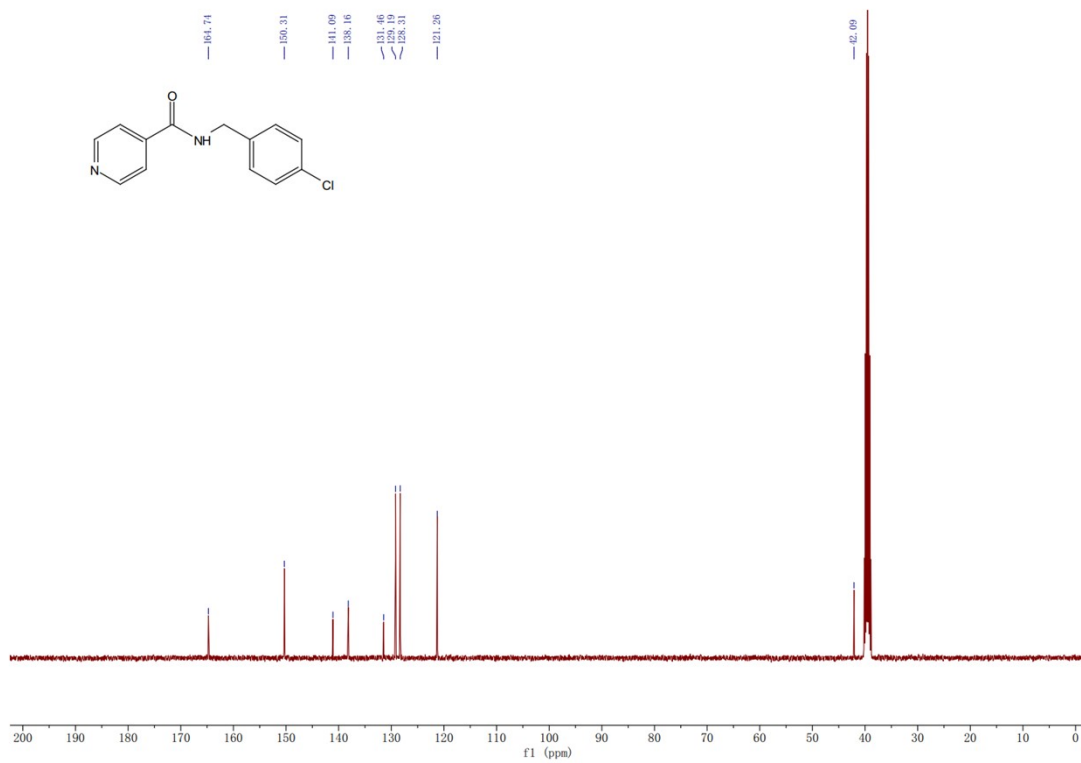
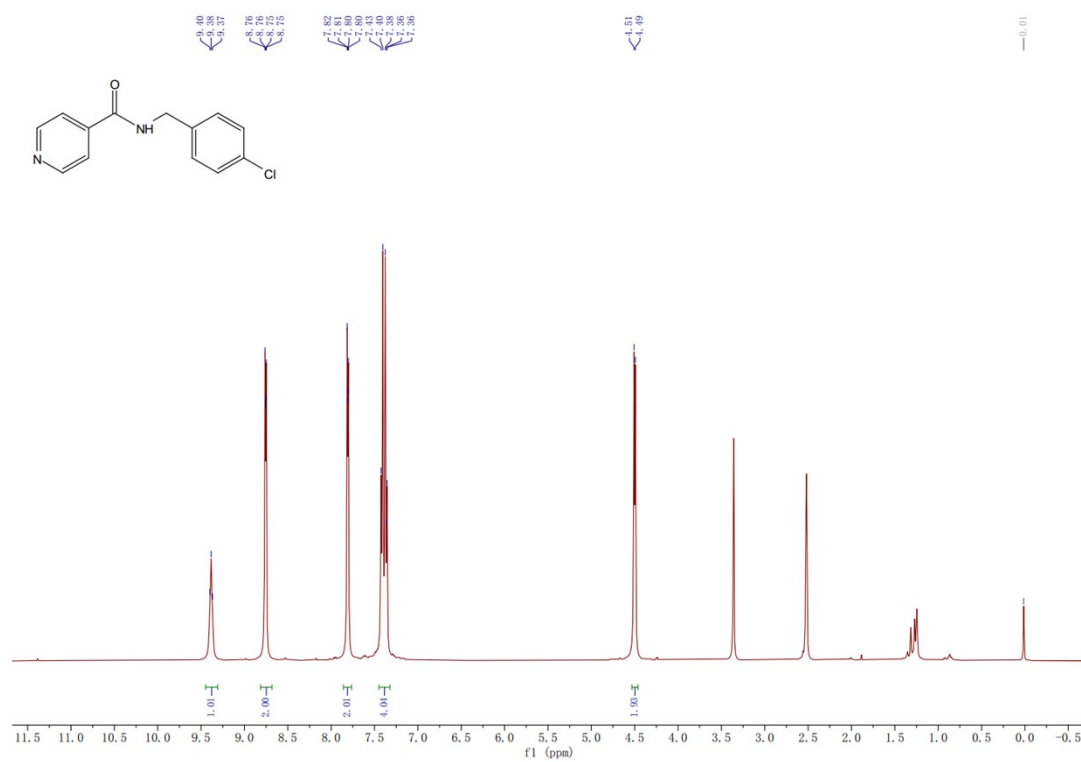
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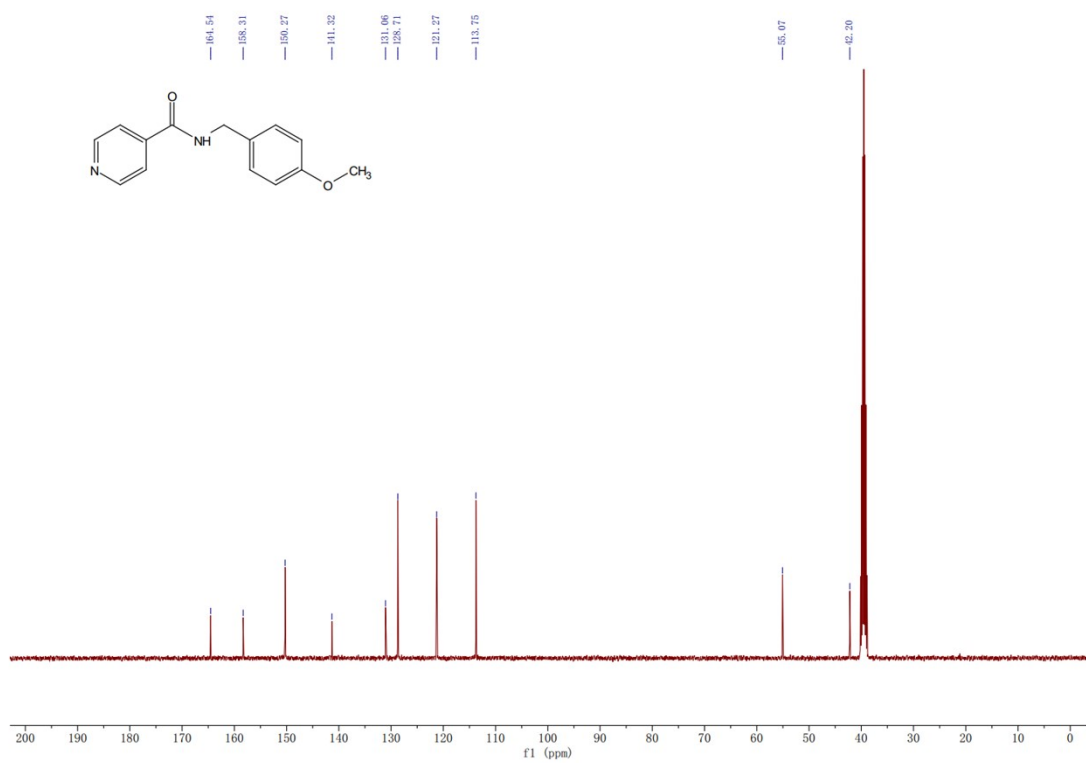
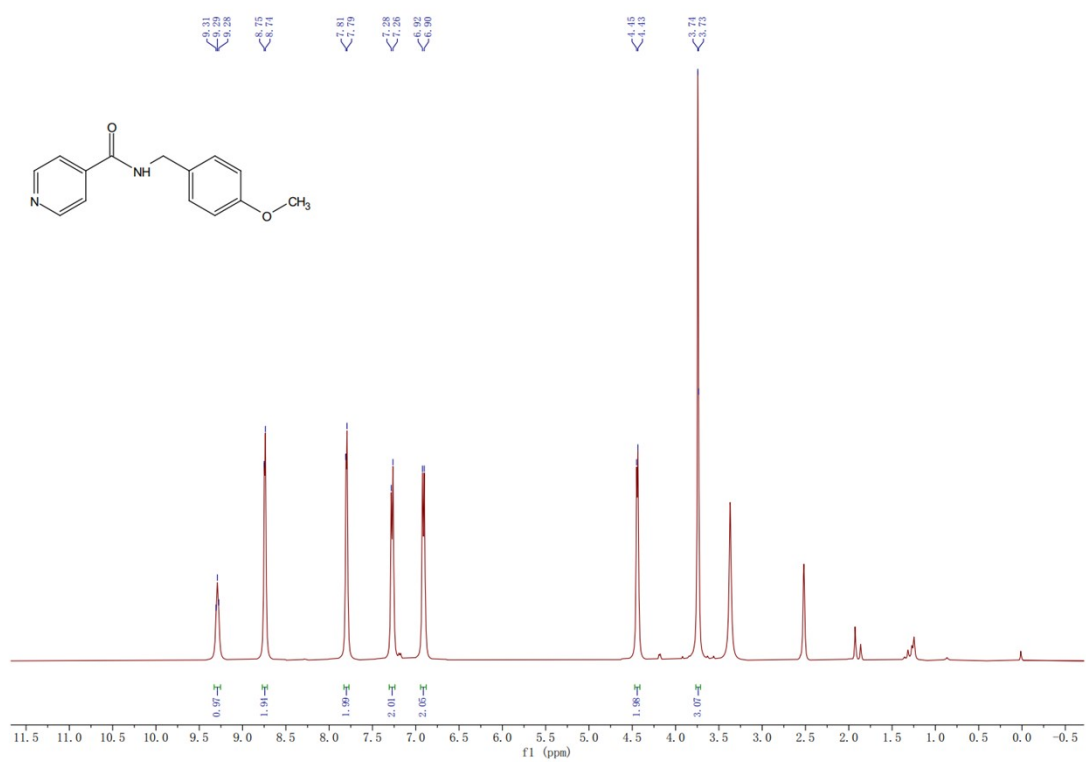
3n



3o

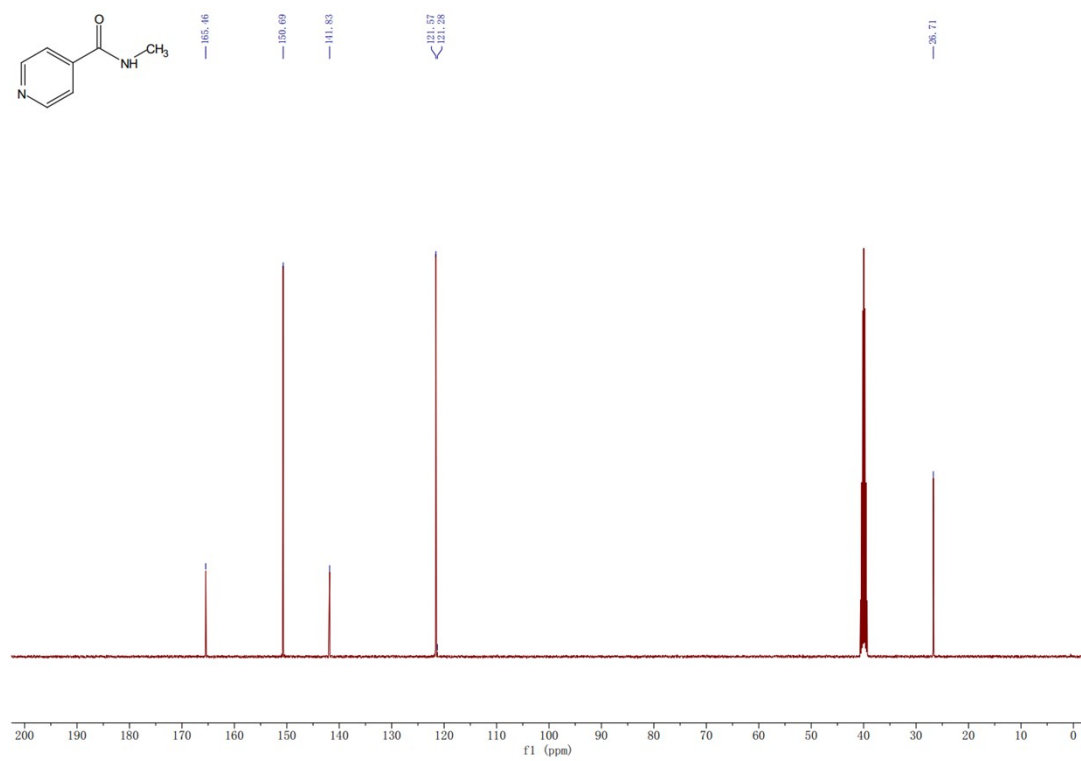
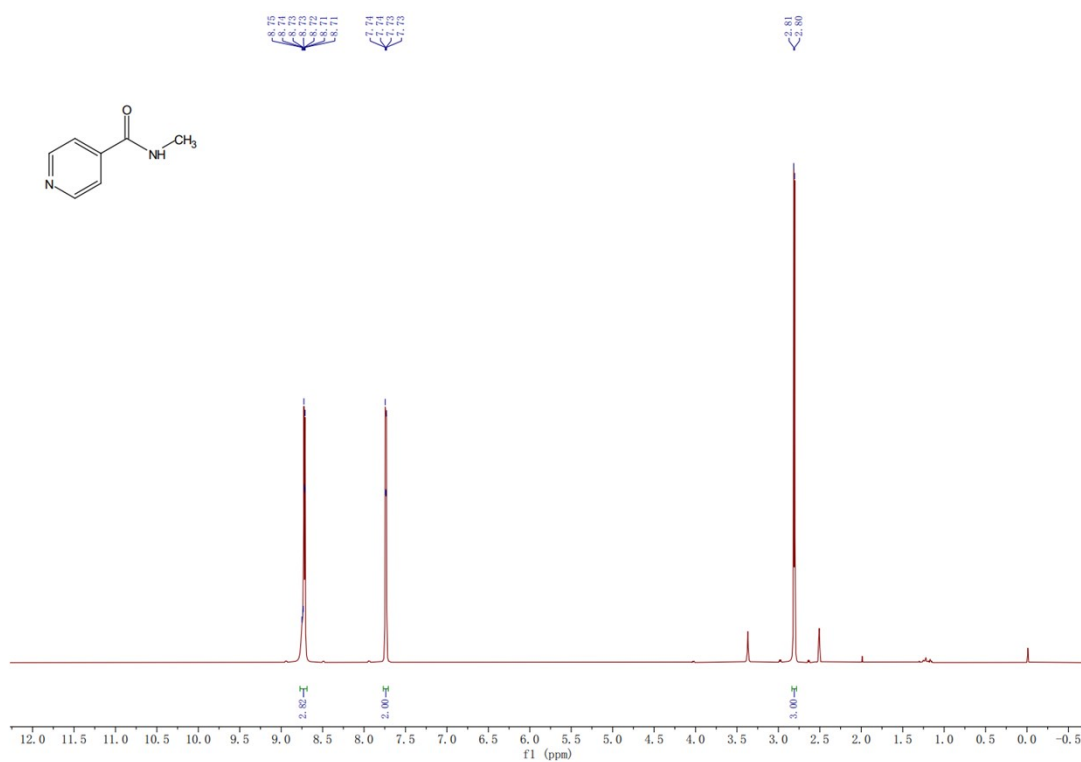


3p

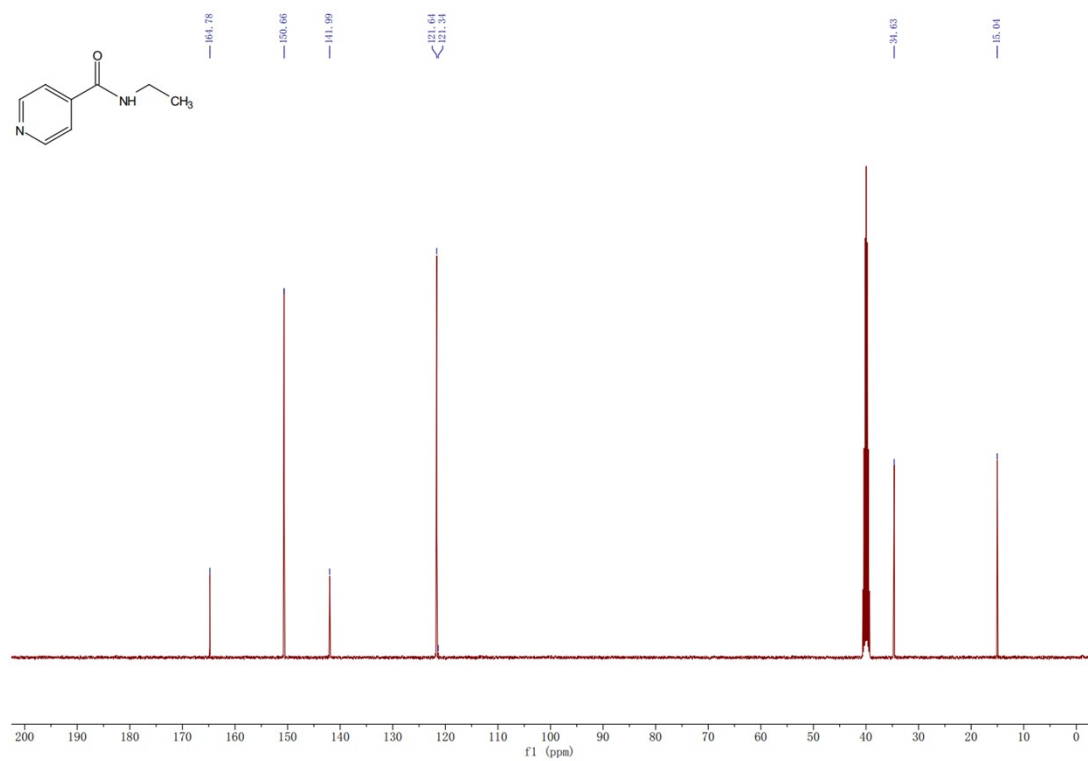
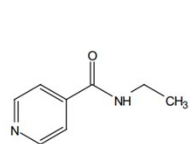
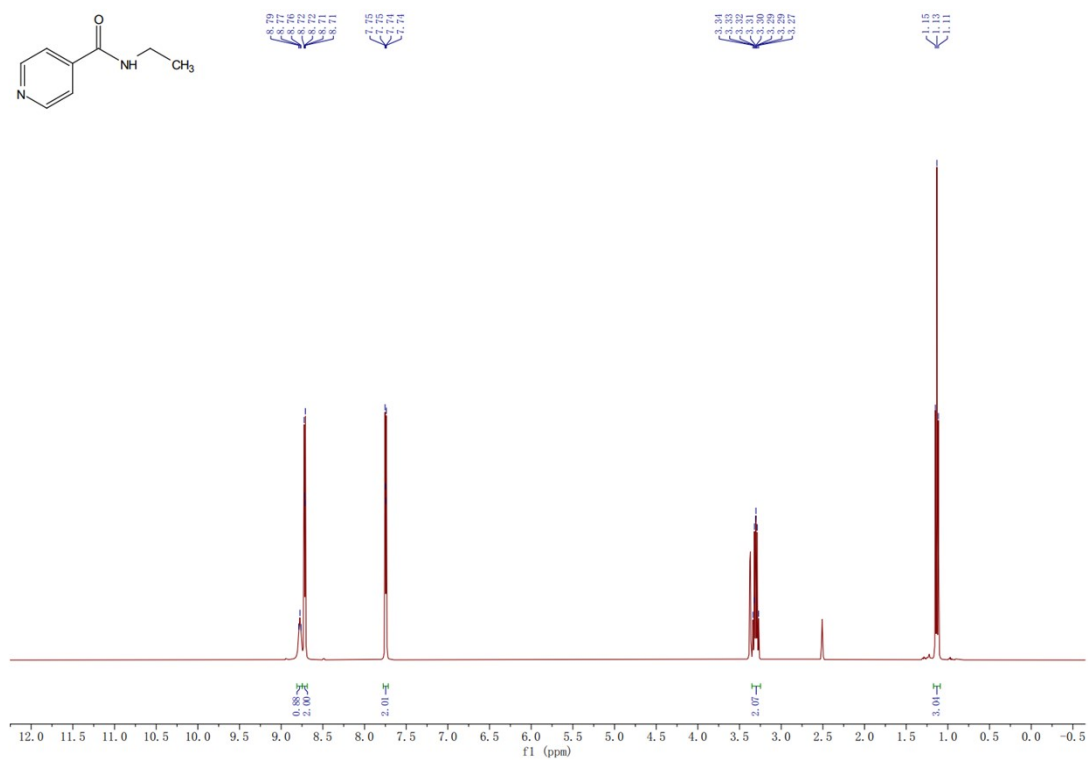
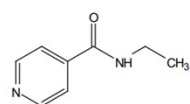




3q



3r



### Notes and references

1. M. M. Khan, Saigal, S. Khan, S. Shareef and S. C. Sahoo, *Rsc Advances*, 2018, **8**, 41892-41903.
2. S. Singh, S. Mondal, N. Vodnala and C. K. Hazra, *Green Chemistry*, 2023, **25**, 1014-1022.
3. B. Dam, A. K. Sahoo and B. K. Patel, *Green Chemistry*, 2022, **24**, 7122-7130.