Electronic Supplementary Information

Materials

1-Pyrenecarboxylic acid (97 %) was purchased from Meryer Co. Ltd. (Shanghai). 1-Pyrenebutyric acid (95%), octafluoronaphthalene (96 %), L-alanine methyl ester hydrochloride (98 %), D-alanine methyl ester hydrochloride (98 %) and *tert*-Butyl (3aminopropyl) carbamate (97%) were purchased from Bide Pharmatech Co., Ltd. (Shanghai). Trifluoroacetic acid (TFA, AR) was purchased from Innochem (Beijing). All solvents were purchased from FuYu Chemical Reagent Co. Ltd. (Tianjin). Both chemicals and solvents were used without further purification. Water is deionized (DI) water.

Characterizations

¹H NMR, ¹⁹F NMR, ¹³C NMR, NOESY (nuclear overhauser effect spectroscopy) and DOSY (diffusion ordered spectroscopy) spectra to characterize molecular structures were measured on a Bruker AVANCE III HD 400 MHz spectrometer (Switzerland) at room temperature with tetramethylsilane (TMS) as reference. Dimethyl sulfoxide-d (DMSO- d_6) and CDCl₃ were used. Concentration-dependent ¹H NMR in DMSO- d_6 were recorded via 600 MHz NMR spectrometer (JEOL ECZ600R/S3). High-resolution mass spectrometry (HR-MS) was performed via an Agilent Q-TOF 6510 MS spectrometer (America). UV-Vis spectra were recorded via UV-1900 from SHIMADZU (Japan). Fluorescence and temperature-variation fluorescence spectra were measured by FluoroMax-4 from Horiba (Japan). Circular dichroism (CD) spectra were measured with Applied Photophysics Chirascan V100 (England). Transmission electron microscope (TEM) images were measured by a HITACHI JEM-100CX II electron microscope (Japan). The samples for TEM detection were dropped in the copper grid and air-dried. Atomic force microscopy (AFM) images were recorded by Bioscope Resolve from Bruker (America) with tapping contact mode. The AFM sample was dropped on the mica wafer and dried at room temperature. Powder X-ray diffraction (XRD) patterns were collected on a Rigaku SmartLab 9KW X-ray diffraction (Japan) with Cu K α radiation ($\lambda = 0.15406$ nm, voltage 45 KV, current 200

mA, power 9 KW). The samples were casted onto cover slide glasses ($18 \text{ mm} \times 18 \text{ mm}$) and dried to form thin films. The average diameter of aggregation was recorded by DLS measurement with NanoZS of Malvern Panalytical (UK).

Preparation of aggregates

For CHCl₃/MCH system, we dissolved PMA or PBA in CHCl₃ to prepare concentrated stock solution of 5 mM. Aggregates were prepared through adding 200 µL solution to a screw-neck glass vial *via* pipette and 800 µL MCH was immediately added. The total concentration of sample was 1 mM in the mixture of CHCl₃/MCH. We tightened the vial to minimize solvent volatilization. For DMSO/H₂O system, powder of PMA or PBA was dissolved in DMSO with concentration of 50 mM. We attained 30 µL solution *via* adding 20 µL concentrated solution and 10 µL DMSO into a screw-neck glass. Sample was prepared by further adding 970 µL H₂O to the above solution. The total concentration of sample was 1 mM in the mixture of DMSO/H₂O. And before characterization, samples were incubated for 12 h at room temperature. We did not consider the small amount of volatilization that may occur during samples aging process. The properties of CD spectra, morphology (TEM and AFM images) and XRD pattern are all repeatable with different batch samples.





Scheme S1. The synthesis process of ^{*L*}MA, ^{*L*}BA and PN.

^{*L*}MA and ^{*L*}BA were prepared via condensation reaction. In details, 1-pyrenecarboxylic acid (1.7 g, 7 mmol) was mixed with L-alanine methyl ester hydrochloride (2 g, 10

dichloromethane (DCM, 50 mL). 1-(3-dimethylaminopropyl)-3mmol) in ethylcarbodiimide (EDC, 2 g, 10 mmol), 1-hydroxybenzotriazole (HOBT, 135 mg, 1 mmol), 4-dimethylaminopyridine (DMAP, 122 mg, 1 mmol) and triethylamine (Et₃N, 2 mL) were added into the mixture. After 12 h reaction at room temperature, crude products were washed by saturated sodium chloride (NaCl) aqueous solution. The DCM phase was dried with anhydrous sodium sulfate (Na₂SO₄), and evaporated solvent via rotary evaporator. Column chromatography was used to purify crude produce (ethyl acetate: petroleum ether = 1:3). The obtained white powder was dissolved via tetrahydrofuran (THF, 20 mL). Sodium hydroxide (NaOH, 1 g, 25 mmol) and water (20 mL) mixed with the solution, which was stirred at room temperature for 1 h. Hydrochloric acid was used adjusting the solution to acidity. The mixture was extracted by DCM and dried with anhydrous Na₂SO₄. ^LMA powder was obtained by evaporating solvent. Yield: 67 %. ¹H NMR (400 MHz, DMSO- d_6 , 298 K) δ = 12.69 (s, 1H), 9.03 (d, 1 H), 8.59 (d, 1H), 8.37-8.34 (m, 3H), 8.28-8.22 (m, 3H), 8.16-8.11 (m, 2H), 4.62-4.55 (m, 1H), 1.47 (d, 3H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 174.83, 169.42, 132.10, 131.20, 130.72, 128.79, 128.54, 128.40, 127.71, 127.06, 126.28, 126.10, 125.83, 125.29, 124.85, 124.24, 124.12, 67.48, 48.88, 25.58, 17.32. HRMS (TOF) m/z [M+H]+, calcd for C₂₀H₁₅NO₃, 317.1052; found, 318.1109.



Figure S2. ¹³C NMR of ^{*L*}MA



Figure S3. HRMS (TOF) of ^{*L*}MA

^{*L*}BA was prepared via the similar method. Pyrene-1-butyric acid (1.7 g, 6 mmol) was mixed with L-alanine methyl ester hydrochloride diethylmalonate (2 g, 10 mmol), EDC (2 g, 10 mmol), HOBT (135 mg, 1 mmol), DMAP (122 mg, 1 mmol) and Et₃N (2 mL) in DCM. After 12 h reaction at room temperature, crude products were washed by saturated NaCl aqueous solution and dried via anhydrous Na₂SO₄, which was subjected to column purification to obtain product. Hydrolysis reaction was occurred in LiOH (1 g, 25 mM) with THF (20 mL), water (20 mL) at room temperature for 1 h. Hydrochloric acid was used adjusting the solution to acidity. The mixture was extracted by DCM and dried with anhydrous Na₂SO₄. ^{*L*}MA powder was obtained by evaporating solvent. Yield: 79 %. ¹H NMR (400 MHz, DMSO-*d*₆, 298 K) δ = 12.39 (s, 1H), 8.40 (d, 1 H), 8.28 (t, 2H), 8.21 (m, 3H), 8.14 (m, 2H), 8.07 (t, 1H), 7.96 (d, 1H), 4.24 (t, 1H), 3.32 (m, 5 H, β-H mixed with H₂O-H from DMSO-*d*₆), 2.29 (m, 2H), 2.01 (m, 2H), 1.27 (d, 3H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 175.05, 172.32, 137.14, 131.37, 130.92, 129.78,

128.99, 128.07,127.95, 127.71, 126.98, 126.62, 125.44, 125.27, 124.71, 124.06, 47.92, 35.14, 32.62, 28.03, 17.58. HRMS (TOF) m/z $[M+H]^+$, calcd for $C_{23}H_{21}NO_3$, 359.1521; found, 360.1575.



Figure S4. ¹H NMR of ^{*L*}BA



Figure S6. HRMS (TOF) of ^LBA

FN was synthesis via octoflonaphthalene (OFN) and N-boc-1,3-diaminopropane. OFN (2.7 g, 10 mmol) was dissolved with isopropanol. Excess N-boc-1,3-diaminopropane (2 mL) was added to the solution. The mixture was stirred at room temperature for a week. After evaporating solvent, the crude product was subjected to column purification (DCM) to obtain purity product 1.50 g. Yield: 35 %. ¹H NMR (400 MHz, CDCl₃, 298 K) δ = 4.6 (s, 2H), 3.54 (m, 2 H), 3.27 (m, 2H), 1.78 (m, 2H), 1.46 (s, 6H). ¹⁹F NMR (376 MHz, CDCl₃, 298 K) δ = 144.60 (d, 1F), 147.62 (m, 1F), 148.63 (m, 1F), 149.62 (s, 2F), 156.96 (s, 1F), 150.99 (s, 1F). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.19, 142.27, 139.79, 139.26, 137.32, 136.89, 127.62, 127.50, 127.37, 108.22, 100.99, 77.97, 42.59, 42.53, 37.51, 30.99, 28.63. HRMS (TOF) m/z [M+H]⁺, calcd for C₁₈H₁₇F₇N₂O₂, 426.1178; found, 427.1230.



Figure S7. ¹H NMR of FN





Figure S9. ¹³C NMR of ^{*L*}FN



Figure S10. HRMS (TOF) of ^LFN

Preparation of compound PMA and PBA



Scheme S2. Preparation of ^{*L*}PMA and ^{*L*}PBA.

FN (852 mg, 2 mM) was mixed with TFA 5 mL) for ten minutes to deprotect boc. After evaporated solvent, ^LMA (476 mg, 1.5 mM), HOBT (27 mg, 0.2 mM), DMAP (24 mg, 0.2 mM), EDC (400 mg, 2 mM), Et₃N (1 mL) and DCM (40 mL) as solvent were added.

The mixture was stirred at room temperature for 12 h. After evaporating solvent, the crude product was subjected to column purification to obtain pure product 0.65 g. Yield: 69 %. ¹H NMR (400 MHz, DMSO- d_6 , 298 K) δ 8.81 (d, 1H), 8.56 (d, 1H), 8.31 (m, 3H), 8.24 (m, 2H), 8.17 (m, 2H), 8.10 (m, 2H), 6.25 (s, 1H), 4.58 (m, 1H), 3.47 (m, 2H), 3.25 (m, 2H), 1.79 (m, 2H), 1.40 (d, 3H). ¹⁹F NMR (376 MHz, DMSO- d_6 , 298 K) δ -145.50 (m, 1F), -148.39 (s, 1F), -148.81 (m, 1F), -149.84 (m, 1F), -150.97 (m, 1F), -157.79 (m, 1H), -162.69 (t, 1F). ¹³C NMR (101 MHz, DMSO- d_6) δ 173.20, 169.18, 132.08, 131.74, 131.10, 130.58, 128.69, 128.39, 128.26, 127.62, 126.93, 126.15, 125.98, 125.93, 125.28, 124.66, 124.10, 123.95, 49.93, 42.63, 36.34, 30.65, 18.24. HRMS (TOF) m/z [M+H]⁺, calcd for C₃₃H₂₂F₇N₃O₂, 625.1600; found, 626.1699.



Figure S11. ¹H NMR of ^{*L*}PMA





Figure S13. ¹³C NMR of ^{*L*}PMA



Figure S14. HRMS (TOF) of ^LPMA

^{*L*}PBA was prepared successfully with the similar method of ^{*L*}PMA. FN (852 mg, 2 mM) was mixed with TFA (5 mL) for ten minutes. After evaporated solvent, ^{*L*}BA (653 mg, 1.5 mM), HOBT (27 mg, 0.2 mM), DMAP (24 mg, 0.2 mM), EDC (400 mg, 2 mM), Et₃N (1 mL) and DCM (40 mL) were added. The mixture was stirred at room temperature for 12 h. After evaporating solvent, the crude product was subjected to column purification to obtain white powder 0.80 g. Yield: 72 %. ¹H NMR (400 MHz, DMSO-*d*₆, 298 K) δ 8.26 (d, 1H), 8.19 (m, 3H), 8.08 (t, 3H), 8.00 (t, 3H), 7.85 (d, 1H), 6.11 (s, 1H), 4.31 (m, 1H), 3.42 (m, 2H), 3.28 (m, 2H), 3.11 (m, 2H), 2.35 (m, 2H), 1.98 (m, 2H), 1.70 (m, 2H), 1.25 (d, 3H). ¹⁹F NMR (376 MHz, DMSO-*d*₆, 298 K) δ - 145.50 (d, 1F), -148.39 (s, 1F), -148.81 (m, 1F), -149.84 (m, 1F), -150.97 (m, 1F), -157.79 (t, 1F), -162.69 (t. 1F). ¹³C NMR (101 MHz, DMSO-*d*₆, 298 K) δ 173.83, 172.48, 131.03, 130.59, 129.40, 128.21, 127.61, 127.52, 127.15, 126.53, 126.27, 125.03, 124.94, 124.15, 124.11, 123.82, 35.92. HRMS (TOF) m/z [M+H]⁺, calcd for C₃₆H₂₈F₇N₃O₂, 667.2070; found, 668.4451.



Figure S16. ¹⁹F NMR of ^{*L*}PBA









^DPMA and ^DPBA were prepared via the same method of ^LPMA and ^LPBA,

respectively. ¹H NMR, ¹⁹F NMR and ¹³C NMR of ^DPMA and ^DPBA were shown in Figure S13-S18.





Figure S21. ¹³C NMR of ^DPMA





Figure S24. ¹³C NMR of ^DPBA

X-ray crystallography

The single crystals of ^{*L*}PMA, ^{*L*}PBA, ^{*D/L*}PMA and ^{*D/L*}PBA were cultured via solvent evaporation. Single crystal with appropriate dimensions were chosen to record on a Rigaku Oxford Diffraction XtaLAB Synergy-S diffractometer at 173 K using Cu K α (λ = 1.54184 Å) from PhotonJet micro-focus X-ray Source. The crystal data were showed in below table.

Deposition Number	2246067
Formula	C33H22F7N3O2
Temperature (K)	173
Wavelength	1.54184Å
Crystal system	triclinic
Space group	C 2 2 21
a,b,c/Å	7.1815(3) 17.3109(7) 46.2197(17)
V, Å ³	5745.95
Cell angles	90 90 90
Ζ, Ζ'	Z: 8 Z': 0
R-factor (%)	5.22

Table S2	2. Crysta	l data c	of <i>D/L</i> PMA
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Deposition Number	2246095
Formula	C33H22F7N3O2
Temperature (K)	173
Wavelength	1.54184Å
Crystal system	orthorhombic
Space group	P -1
a,b,c/Å	8.3043(2) 8.6910(2) 20.6746(4)

V, Å ³	1356.82
Cell angles	80.978(2) 81.965(2) 67.556(3)
Ζ, Ζ'	Z: 2 Z': 0
R-factor (%)	5.74

Table S3. Crystal data of ^LPBA

Deposition Number	2246071
Formula	2(C36H28F7N3O2)
Temperature (K)	173
Wavelength	1.54184Å
Crystal system	monoclinic
Space group	P 2 ₁
a,b,c/Å	17.8394(3) 8.2183(2) 22.8962(5)
V, Å ³	3272.29
Cell angles	90 102.884(2) 90
Ζ, Ζ'	Z: 2 Z': 0
R-factor (%)	11.07

Table S4. Crystal data of ^{D/L}PBA

Deposition Number	2246072
Formula	2(C36H28F7N3O2)
Temperature (K)	173
Wavelength	1.54184Å
Crystal system	monoclinic
Space group	P 2 ₁
a,b,c/Å	22.8294(3) 9.44580(10) 31.1842(5)
$V, Å^3$	6724.53
Cell angles	90 90.301(2) 90
Ζ, Ζ'	Z: 4 Z': 0

R-factor (%)	12.82
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Molecular dynamics simulation

The native structures of model compound were built from the GaussView6.0 program. The obtained configuration was optimized and the electrostatic potential (ESP) was simultaneously calculated by Hartree-Fork method. The 6-31G(d) basis set was employed in Gaussian 16 program. The Antechamber program was used to fit the restrained electrostatic potential (RESP) charge, and then the general Amber force field (GAFF) was adopted to parameterize the bonded interaction for subsequent MD simulations. The MD simulation was implemented with the GROMACS 2020 program. The long-range electrostatic interaction was calculated by the particle mesh Ewald (PME) method. The cut-off distance for non-bonded interactions was set to 1 nm. The total simulation time is 30 ns. Energy minimization was conducted using the steepest descent algorithm before performing dynamic simulation. V-rescale and Parrinello-Rahman were respectively employed to control temperature and pressure. All results were visualized using VMD program.

Theoretical calculation

Non-covalent interaction (NCI)/reduced density gradient (RDF) analysis were manipulated via Multiwfn 3.8.^{1, 2} The original structures from crystal without further treatment. Input file for Multiwfn 3.8 were achieved from Gaussian 16 program³. Hirshfeld surfaces were calculated directly form crystals through CrystalExplorer.⁴ The dimerization interaction energies were estimated via the calculation of single point energy with the B3LYP/def2svp-D3(BJ) basis set of the Gaussian 16 program. The original structures of ^{*L*}PMA mono, ^{*L*}PMA dimer and ^{*D/L*}PMA dimer are directly form crystal structures.



Figure S25. Scatter points images of NCI analysis including a) ^LPMA (dimer), b)

^{*L*}PBA (dimer).



Figure S26. Surface area of reciprocal contacts from ^{*L*}PMA(dimer) crystal including a) F…H; b) H…H; c) C…C; d) O…H; e) C…F; f) C…H and g) the corresponding pie chart.



Figure S27. Surface area of reciprocal contacts from ^{*L*}PBA(dimer) crystal including a) F…H; b) H…H; c) C…C; d) O…H; e) C…F; f) C…H and g) the corresponding pie chart.



Figure S28. Scatter points images of NCI analysis including ^{D/L}PMA(dimer),

^{*L*}PBA(folder) from ^{*D*/*L*}PBA crystal.



Figure S29. Noncovalent interaction analysis of ^{D/L}PMA dimer and ^LPBA folder.



Figure S30. Surface area of reciprocal contacts from ^{*D/L*}PMA(dimer) crystal including a) F···H; b) H···H; c) C···C; d) O···H; e) C···F; f) C···H and g) the corresponding pie

chart.



Figure S31. Surface area of reciprocal contacts of ^{*L*}PBA folder from ^{*D/L*}PBA crystal including a) $F \cdots H$; b) $H \cdots H$; c) $C \cdots C$; d) $O \cdots H$; e) $C \cdots F$; f) $C \cdots H$ and g) the corresponding pie chart.



Figure S32. a) ^{*L*}PBA folded structure from ^{*D/L*}PBA crystal; b) Simulated model after replacing naphthalene-F7 with methyl; c) Optimizing the structure with b3lyp-D3(BJ)/6-311g.



Figure S33. a) ^{*L*}PBA folded structure from ^{*D/L*}PBA crystal; b) Simulating model and c) optimizing structure with b3lyp-D3(BJ)/6-311g after replacing N1 to O; d) Simulating model and e) optimizing structure with b3lyp-D3(BJ)/6-311g after replacing N2 to O.



Figure S34. The chemical structure of model compounds.



Figure S35. Molecular dynamic simulation. (a) Structure evolution process with time. Variation of (b) energy (c, d). Distance between C1 and C2 and (e) number of H-bond for simulation system.



Figure S36. Concentration dependent ¹H NMR spectra of ^{*L*}PMA form 0.05 mM to 2.0



Figure S37. Concentration dependent ¹H NMR spectra of ^{*L*}PBA form 0.05 mM to 2.0 mM in CDCl₃



Figure S38. Concentration dependent ¹H NMR spectra of ^{*L*}PMA including 0.1 mM, 1 mM, 3 mM, 5 mM, 7 mM and 10 mM in DMSO-*d*₆.



Figure S39. Concentration dependent ¹H NMR spectra of ^{*L*}PMA including 0.1 mM, 1 mM, 3 mM, 5 mM, 7 mM and 10 mM in DMSO-*d*₆.



Figure S40. Partial concentration-dependent ¹H NMR spectra of ^{*L*}PMA and ^{*L*}PBA in DMSO- d_6



Figure S41. Concentration dependent ¹H NMR spectra of ^{*L*}PBA including 0.1 mM, 1 mM, 3 mM, 5 mM, 7 mM and 10 mM in DMSO-*d*₆.



Figure S42. Concentration dependent ¹H NMR spectra of ^{*L*}PBA including 0.1 mM, 1 mM, 3 mM, 5 mM, 7 mM and 10 mM in DMSO-*d*₆.



Figure S43. Temperature-variable ¹H NMR spectra of ^{*L*}PMA in DMSO- d_6 (10 mM)



Figure S44. Temperature-variable ¹H NMR spectra of ^{*L*}PBA in DMSO- d_6 (10 mM).



Figure S45. 2D NOESY spectra of ^{*L*}PBA in CDCl₃/CH₃OD (9/1, 10 mM).



Figure S46. 2D NOESY spectra of ^{*L*}PBA in CDCl₃ (saturated) (cyan regions are possible corrections to prove existing of dimer).

Fitting procedures to derive size parameters form log D⁵⁻⁷

Stocks-Einstein equation build a relationship between diffusion coefficient and hydrodynamic radius with sphere model:

$$D = \frac{k_b T}{6\pi\eta r_h(sp)}$$

Where k_B is the *Boltzmann* constant ($k_B = 1.38 \times 10^{-23}$), T is temperature (K), η is viscosity of the solvent, and $r_h(sp)$ is the hydrodynamic radius.

The equation can be modified for oblate spheroid model:

$$D = \frac{k_b T}{c f_s \pi \eta r_h(ob)}$$

Where

$$c = \frac{6}{1 + 0.695 \left(\frac{r_{vdW}}{r_h(ob)}\right)^{2.234}}$$

 $c \approx 6$, because r_{vdW} (van der Waals radius of the solvent) is far less than the

hydrodynamic radius of the molecules which are investigated. Thus:

$$r_{h}(ob) = \frac{r_{h}(sp)}{f_{s}}$$
$$f_{s} = \frac{\sqrt{(\frac{b}{a})^{2} - 1}}{(\frac{b}{a})^{\frac{2}{3}} \tan^{-1} \sqrt{(\frac{b}{a})^{2} - 1}}$$

a and b are the semiminor and semimajor axis of the oblate spheroid.

The following figures shows the result for hydrodynamic radii of PMA and PBA in DMSO and $CHCl_3$ with various temperature. Viscosity information get from fitting curves of DMSO and $CHCl_3$ upon various temperature. r(cal) is the radius of a sphere with an equivalent volume as the spheroid generated by a and b; spheroid volume (

$$V = \frac{4}{3}\pi ab^{2}_{\);} r(cal) = \sqrt[3]{\frac{3V}{4\pi}} = \sqrt[3]{ab^{2}}_{\ }.$$
 Ratio of the hydrodynamic radius generated from the oblate model (*r*h(ob)) and the equivalent radius generated by the volume of the spheroid

(*r*(cal)) was used to guide the adjustment of a and b until the volume generated by them matches the volume generated by $rh(ob) (rh(ob)/r(cal) \approx 1)$



Figure S47. Fitting of viscosity versue temperature data for DMSO based on values found in the literature⁸.



Figure S48. Fitting of viscosity versue temperature data for CHCl₃ based on values found in the literature⁹.



Figure S49. DOSY NMR spectra of ^LPMA in CDCl₃ with 298.15 K.



Figure S50. DOSY NMR spectra of ^{*L*}PMA in CDCl₃ with 318.15 K.



Figure S51. DOSY NMR spectra of ^{*L*}PBA in CDCl₃ with 298.15 K.



Figure S52. DOSY NMR spectra of ^{*L*}PBA in CDCl₃ with 318.15 K.



Figure S53. DOSY NMR spectra of ^LPMA in DMSO- d_6 with 298.15 K.



Figure S54. DOSY NMR spectra of ^LPMA in DMSO- d_6 with 323.15 K.



Figure S55. DOSY NMR spectra of ^{*L*}PBA in DMSO- d_6 with 298.15 K.



Figure S56. DOSY NMR spectra of ^LPBA in DMSO- d_6 with 323.15 K.

PMA												
CHCI ₃		K _B										
Oblate	b>a	p>1										
			log D									
D	1.04E-09		-8.982967									
T(K)	298.15											
η(Pa*s	0.000536											
rh(sp)=	kbT/6πDη	3.92E-10										
	b	a	rh(sp)	rh(ob)	fs	r(cal)	p	1/p	r(cal)/rh(ob	fs1	fs2	fs3
	5.50E-10	1.30E-10	3.92E-10	3.32E-10	1.18	3.40061E-10	4.23E+00	0.236	1.02E+00	4.111	2.616	1.332

Figure S57. Screenshot of Microsoft Excel spreadsheet used to fit ^LPMA in CDCl₃

with 298.15 K using oblate spheroid model.

^L PMA									
CHCI ₃		K _B							
Oblate	b>a	p>1							
			log D						
D	1.05E-10		-9.978811						
T(K)	318.15								
η(Pa*s)	0.000494								
rh(sp)=	kbT/6πDη	4.49E-09							
	b	а	rh(sp)	rh(ob)	fs	r(cal)	p	1/p	r(cal)/rh(ob)f
	7.00E-10	3.00E-11	4.49E-09	2.40E-09	1.868	2.44966E-10	2.33E+01	0.043	1.02E-01

Figure S58. Screenshot of Microsoft Excel spreadsheet used to fit ^LPMA in CDCl₃

with 318.15 K using oblate spheroid model.

^L PBA									-
CHCI ₃		K _B							
Oblate	b>a	p>1							
			log D						
D	6.91E-10		-9.160522						
T(K)	298.15								
η(Pa*s	0.000536								
rh(sp)=	kbT/6πDη	5.90E-10							
b		а	rh(sp)	rh(ob)	fs	r(cal)	p	1/p	r(cal)/rh(ob)
	8.00E-10	2.50E-10	5.90E-10	5.28E-10	1.117	5.42884E-10	3.20E+00	0.313	1.03E+00

Figure S59. Screenshot of Microsoft Excel spreadsheet used to fit ^LPBA in CDCl₃

PBA CHCl₃ K_B Oblate b>a p>1 log D D 9.74E-10 -9.01144 T(K) 318.15 n(Pa*s) 0.000494 rh(sp)=kbT/6πDη 4.84E-10 rh(ob) r(cal)/rh(ob) rh(sp) fs r(cal) 1/p b a p 5.70E-10 3.30E-10 4.84E-10 4.72E-10 1.026162 4.75067E-10 1.73E+00 0.578947 1.006529

with 298.15 K using oblate spheroid model.

Figure S60. Screenshot of Microsoft Excel spreadsheet used to fit ^LPBA in CDCl₃

with 318.15 K using oblate spheroid model.

PMA													
DMSO		K _B											
Oblate	b>a	p>1											
			log D										
D	1.61E-10		-9.793174										
T(K)	298.15												
η(Pa*s	0.001967												
rh(sp)=	kbT/6πDη	6.90E-10											
	b	а	rh(sp)	rh(ob)	fs	r(cal)	p	1/p	r(cal)/rh(ob	fs1	fs2	fs3	
	8.90E-10	3.00E-10	6.90E-10	6.25E-10	1.103	6.19394E-10	2.97E+00	0.337	9.90E-01	2.793	2.065	1.227	

Figure S61. Screenshot of Microsoft Excel spreadsheet used to fit ^LPMA in DMSO-

 d_6 with 298.15 K using oblate spheroid model.

LDMA	-				-	-			
FINA									
DMSO		KB							
Oblate	b>a	p>1							
			log D						
D	3.10E-10		-9.508638						
T(K)	323.15								
η(Pa*s)	0.001269								
rh(sp)=	kbT/6πDη	6.02E-10							
	b	a	rh(sp)	rh(ob)	fs	r(cal)	p	1/p	r(cal)/rh(ob
	8.50E-10	2.80E-10	6.02E-10	5.44E-10	1.107	5.87037E-10	3.04E+00	0.329	1.08E+00

Figure S62. Screenshot of Microsoft Excel spreadsheet used to fit ^LPMA in DMSO-

 d_6 with 323.15 K using oblate spheroid model.

PBA									
DMSO		K _B							
Oblate	b>a	p>1							
			log D						
D	1.57E-10		-9.8041						
T(K)	298.15								
η(Pa*s	0.001967								
rh(sp)=	kbT/6πDη	7.07E-10							
	b	а	rh(sp)	rh(ob)	fs	r(cal)	p	1/p	r(cal)/rh(ob)
	8.20E-10	4.90E-10	7.07E-10	6.91E-10	1.023	6.90676E-10	1.67E+00	0.598	9.99E-01

Figure S63. Screenshot of Microsoft Excel spreadsheet used to fit ^LPBA in DMSO- d_6

with 298.15 K using oblate spheroid model.

PBA									
DMSO		K _B							
Oblate	b>a	p>1							
			log D						
D	2.84E-10		-9.546682						
T(K)	323.15								
η(Pa*s)	0.001269								
rh(sp)=	kbT/6πDη	6.57E-10							
	b	а	rh(sp)	rh(ob)	fs	r(cal)	p	1/p	r(cal)/rh(ob
	8.70E-10	2.90E-10	6.57E-10	5.95E-10	1.105	6.03224E-10	3.00E+00	0.333	1.01E+00

Figure S64. Screenshot of Microsoft Excel spreadsheet used to fit ^LPBA in DMSO- d_6

with 323.15 K using oblate spheroid model.



Figure S65. CD and corresponding UV-Vis absorption spectra of PMA in a) CHCl₃

and b) DMSO (0.3 mM).



Figure S66. CD and corresponding UV-Vis absorption spectra of PBA in a) CHCl₃



Figure S67. Temperature-variable CD spectra of ^LPMA and corresponding g_{Abs} (0.4 mM) in (a, d) DMSO and (b, e, c, f) CHCl₃ (g_{Abs} = CD/ Abs*32980)

and b) DMSO (0.3 mM)



Figure S68. Temperature-variable CD spectra of ^LPBA (0.3 mM) in (a) DMSO and (b,





Figure S69. CD and corresponding UV-Vis absorption spectra of (a) PMA and (b) PBA in CHCl₃/MCH (2/8 by volume, 1 mM)



Figure S70. CD and corresponding UV-Vis absorption spectra of PMA and PBA crystals (1.8 mg solid with 540 mg KBr)



Figure S71. TEM images of self-assembled ^{*L*}PMA and ^{*L*}PBA in the mixture of DMSO/H₂O (30/970 by volume, 1 mM).



Figure S72. AFM images of self-assembled ^{*L*}PMA and ^{*L*}PBA in the mixture of DMSO/H₂O (30/970 by volume, 1 mM).



Figure S73. DLS of self-assembled L PMA and L PBA in the mixture of DMSO/H₂O (30/970 by volume, 1 mM)



Figure S74. CD and corresponding UV-Vis absorption spectra of PMA and PBA assemblies in DMSO/H₂O (30/970 by volume, 1 mM).

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