

Electronic Supplementary Information for

**Switchover from Singlet Oxygen to Superoxide Radical through a Photoinduced  
Two-Step Sequential Energy Transfer Process**

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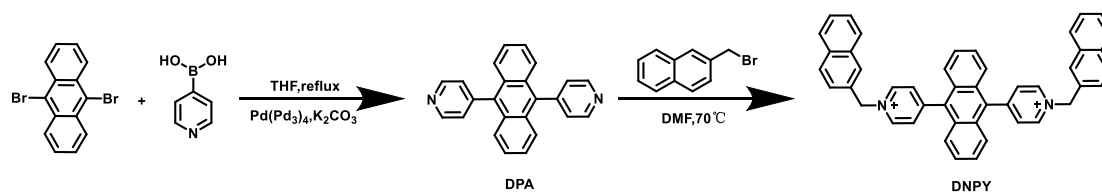
<sup>‡</sup>These authors contributed equally to this work.

## Experimental

Materials: Unless specifically mentioned, all chemicals are commercially available and were used as received.

## Characterizations

<sup>1</sup>H NMR spectra were recorded on a Bruker Avance 400 spectrometer (400 MHz) at 298 K, and the chemical shifts ( $\delta$ ) were expressed in ppm, and  $J$  values were given in Hz. UV-vis spectra were obtained on a Shimadzu UV-1601PC spectrophotometer in a quartz cell (light path 10 mm) at 298 K. Steady-state fluorescence measurements were carried out using a Hitachi 4500 spectrophotometer. Dynamic light scattering (DLS) and zeta potential are measured on Malvern Zetasizer Nano ZS90. Transmission electron microscopy (TEM) images were obtained on a JEM 2100 operating at 120 kV. Samples for TEM measurement was prepared by dropping the mixed aqueous solution on a carbon-coated copper grid (300 mesh) and drying by slow evaporation. Hamamatsu absolute quantum yield measuring instrument Quantaury-QY was used to obtain fluorescence quantum yields. The time-resolved fluorescence decay curve was obtained by the FLS 920 Steady-State/Transient Fluorescence Spectrometer.



**Scheme S1.** Synthetic route of DNPY.

### Synthesis of DPA

9,10-dibromoanthracene (2.07 g, 6 mmol), 4-pyridinyl boronic acid (2.21 g, 18 mmol), tetrakis(triphenylphosphine)palladium (0.14 g, 0.12 mmol) were added into the mixed solution of tetrahydrofuran (15 mL), toluene (3 mL) and 6 mL of 2 mol/L aqueous potassium carbonate. The mixture was refluxed under nitrogen for 3 days, filtered, and the precipitate was collected and washed with  $\text{H}_2\text{O}$  and methanol.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  8.89 (d,  $J = 5.9$  Hz, 4H), 7.57 - 7.53 (m, 8H), 7.51 (d,  $J = 10.3$  Hz, 4H).

### Energy-transfer efficiency calculation

The energy-transfer efficiency ( $\Phi_{ET}$ ) was calculated from excitation fluorescence spectra through the equation S1:

$$\Phi_{ET} = 1 - I_{DA}/I_D \text{ (S1)}$$

Where  $I_{DA}$  and  $I_D$  are the fluorescence intensities of the emission of DNPY-SBE- $\beta$ -CD+RhB, DNPY-SBE- $\beta$ -CD+RhB+SR101, or DNPY-SBE- $\beta$ -CD+SR101 (donor and acceptor) and DNPY-SBE- $\beta$ -CD or DNPY-SBE- $\beta$ -CD+RhB (donor) respectively, when excited at 409 nm. The energy-transfer efficiency ( $\Phi_{ET}$ ) was calculated as 76%, 81% and 84% in an aqueous environment, measured under the condition of  $[DNPY] = 1.0 \times 10^{-5}$  mol/L,  $[SBE-\beta-CD] = 2.0 \times 10^{-6}$  mol/L,  $[RhB] = 1.0 \times 10^{-6}$  mol/L,  $[SR101] = 1.0 \times 10^{-6}$  mol/L.

### Antenna effect calculation

The Antenna effect was calculated based on the excitation spectra using equation S2:

$$\text{Antenna effect} = (I_{DA,409} - I_{D,409}) / I_{DA,550/580} \text{ (S2)}$$

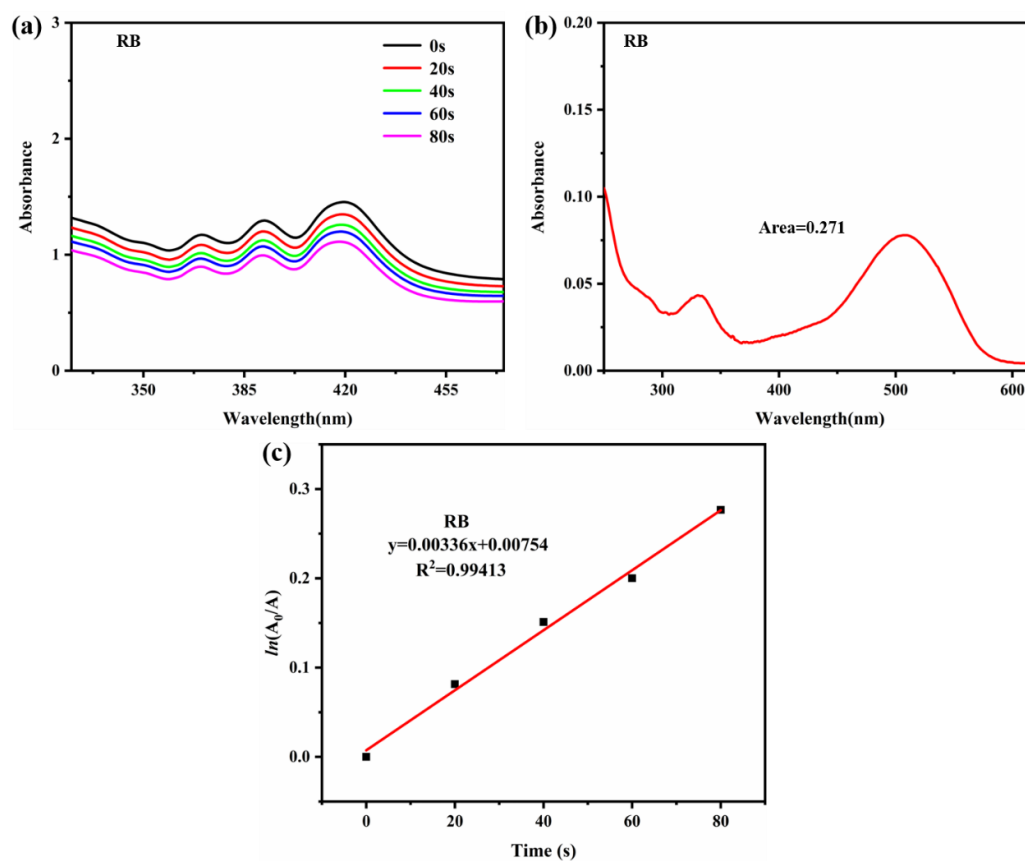
Where  $I_{DA}$  and  $I_D$  are the fluorescence intensities of the emission of DNPY-SBE- $\beta$ -CD+RhB, DNPY-SBE- $\beta$ -CD+RhB+SR101, or DNPY-SBE- $\beta$ -CD+SR101 (donor and acceptor) and DNPY-SBE- $\beta$ -CD or DNPY-SBE- $\beta$ -CD+RhB (donor) respectively, when excited at 409 nm. The antenna effect value was calculated as 15.7, 7.4 and 7.5 in water, measured under the condition of  $[DNPY] = 1.0 \times 10^{-5}$  mol/L,  $[SBE-\beta-CD] = 2.0 \times 10^{-6}$  mol/L,  $[RhB] = 1.0 \times 10^{-6}$  mol/L,  $[SR101] = 1.0 \times 10^{-6}$  mol/L.

### Procedure for $^1O_2$ Quantum Yield Measurement.

The  $^1O_2$  quantum yield was measured using Rose Bengal (RB) as the reference photosensitizer and calculated using the following S3:

$$\Phi_{probe} = \Phi_{RB} \times (K_{probe} A_{RB} / K_{RB} A_{probe}) \quad (S3)$$

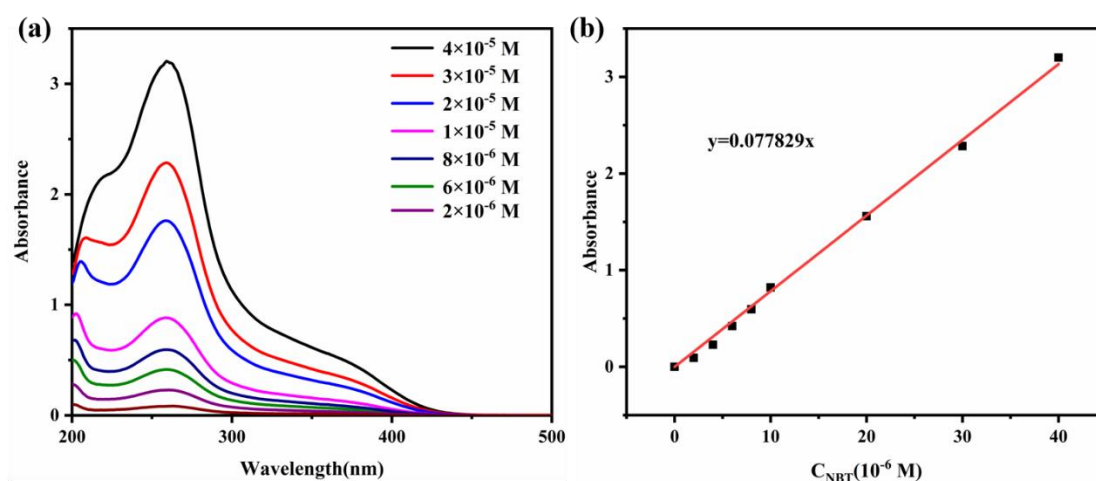
where  $K_{probe}$  and  $K_{RB}$  are the decomposition rate constants of ABDA in the presence of the probe and RB, respectively.  $\Phi_{RB}$  is the  $^1O_2$  quantum yield of RB ( $\Phi_{RB} = 0.75$  in water).  $A_{probe}$  and  $A_{RB}$  represent the integration area of absorption bands ranging from 410 to 415 nm of the probe and RB, respectively. The ABDA ( $1.5 \times 10^{-7}$  mol) in 3 mL of the probe solution was exposed to purple light irradiation (410-415 nm) with a power density of 10W. The natural logarithm of the absorbance ratio ( $A_0/A$ ) of ABDA at 380 nm was plotted against irradiation time and the slope is regarded as the decomposition rate.



**Fig. S1** (a) The absorption spectra of ABDA after irradiation (410-415 nm, 10 W) for different time in the presence of RB; (b) The UV-vis absorption spectra of RB in the aqueous solution; (c) The decomposition rates of ABDA in the presence of RB.

### Procedure for $O_2^{\cdot-}$ Generation Efficiency Measurement.

The amounts of  $O_2^{\cdot-}$  was quantitatively detected by nitroblue tetrazolium (NBT) conversion detection. NBT, which can react with  $O_2^{\cdot-}$  and displays a maximum absorbance at 260 nm, was selected to determine the amounts of  $O_2^{\cdot-}$  generated over the photocatalysts. By recording the concentration of NBT on a UV-vis spectrophotometer, the production of  $O_2^{\cdot-}$  was quantitatively analyzed. First, the photocatalyst ( $3.0 \times 10^{-8}$  mol) and NBT ( $9.0 \times 10^{-8}$  mol) sonication were dispersed into 3 mL of aqueous solution. Then, the mixture was exposed to 410-415nm LED (10W). At appropriate intervals, record the change in absorbance of NBT at 260 nm by UV-vis spectrophotometer, the production of  $O_2^{\cdot-}$  was quantitatively analyzed.



**Fig. S2** (a) The UV-vis absorption spectra of different concentrations of NBT in the aqueous solution; (b) the relation curve of UV-vis absorption intensity of NBT at 260 nm and NBT concentration in aqueous solutions.

### General procedure for the photooxidation reaction of thioanisole and its derivatives

The thioanisole or its derivatives (0.10 mmol) was dissolved in freshly prepared aqueous solution (catalyst total amount: 3 mL, [DNPY] =  $1.67 \times 10^{-4}$  mol/L, [SBE- $\beta$ -CD] =  $3.33 \times 10^{-5}$  mol/L). The mixture was irradiated with 410-415 nm LED (10 W) at room temperature for 2 h. Then,

the mixture was extracted with dichloromethane and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. Then the organic solution was concentrated in a vacuum and purified by rapid column chromatography to obtain the corresponding products.

#### **General procedure for the photocatalytic oxidative hydroxylation of arylboronic acids**

The arylboronic acids (0.10 mmol), *N,N*-diisopropylethylamine (DIPEA) (70 μL, 0.40 mmol) were dissolved freshly prepared aqueous solution (catalyst total amount: 3 mL, [DNPY] =  $1.67 \times 10^{-4}$  mol/L, [SBE-β-CD] =  $3.33 \times 10^{-5}$  mol/L, [RhB] =  $1.67 \times 10^{-5}$  mol/L, [SR101] =  $1.67 \times 10^{-5}$  mol/L. The mixture was irradiated with 410-415 nm LED (10 W) at room temperature for 12 h. Then, the mixture was extracted with dichloromethane and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. Then the organic solution was concentrated in a vacuum and purified by rapid column chromatography to obtain the corresponding products.

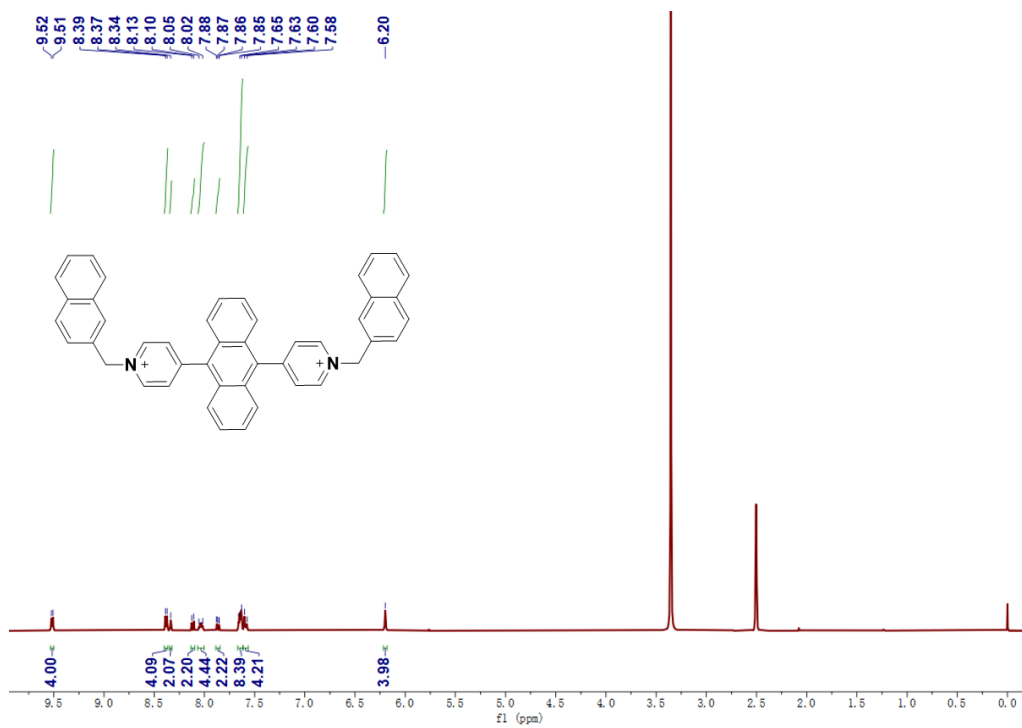
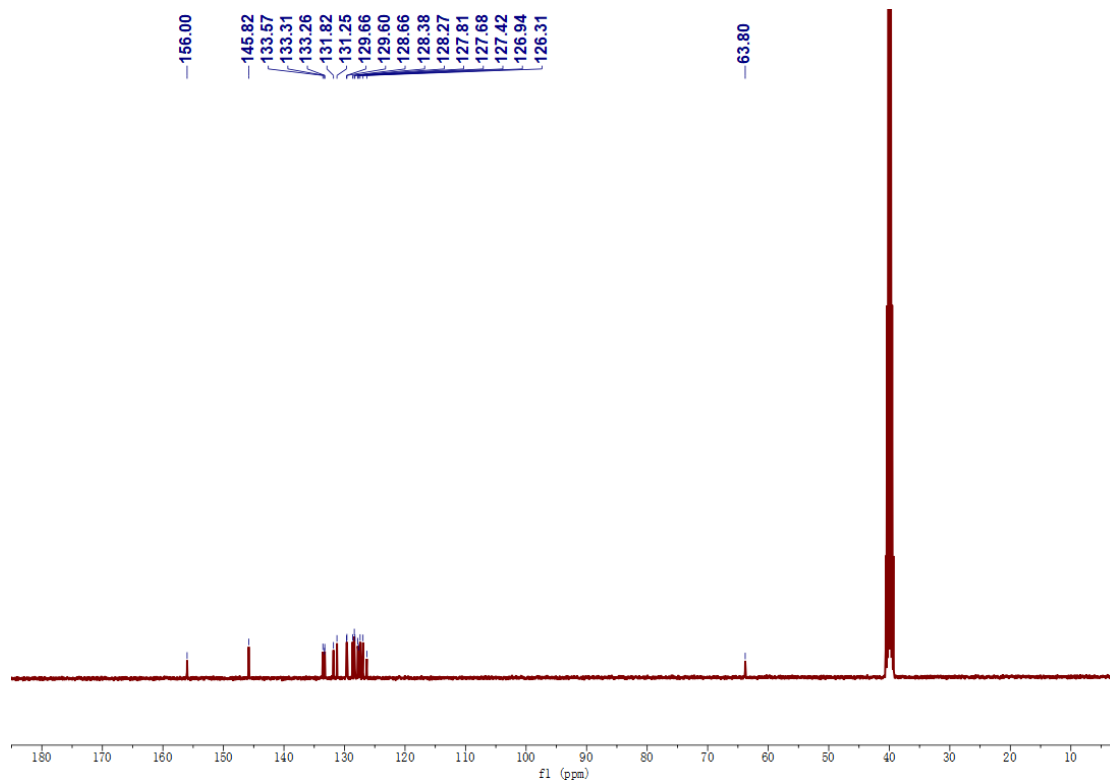
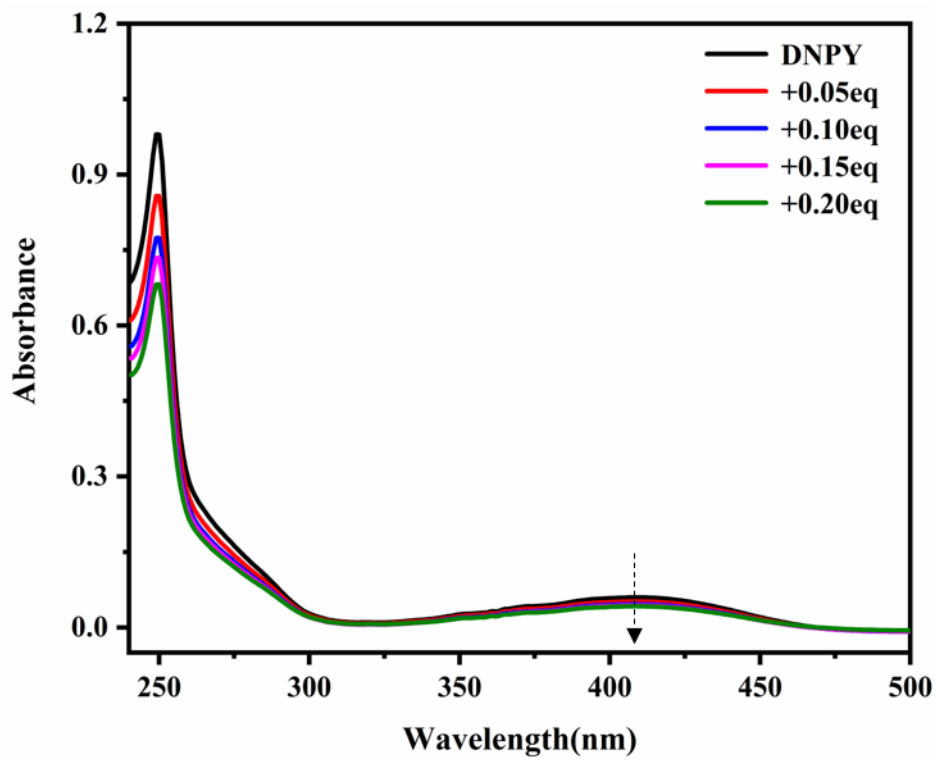


Fig. S3  $^1\text{H}$  NMR spectra of DNPY in  $\text{DMSO-}d_6$ .

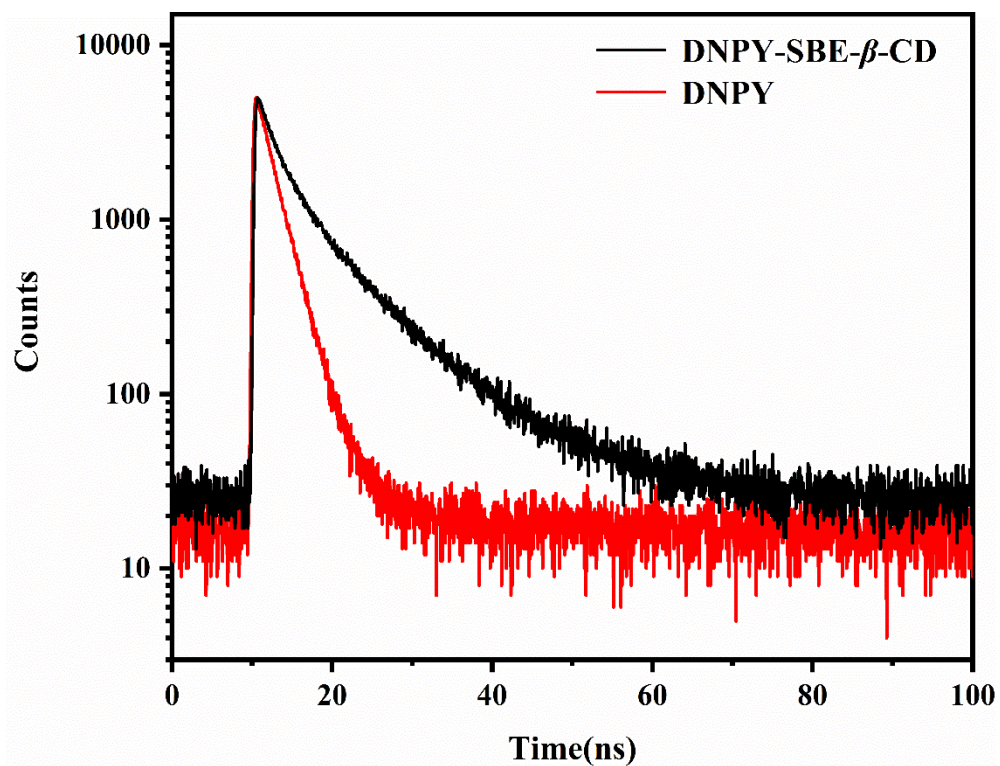




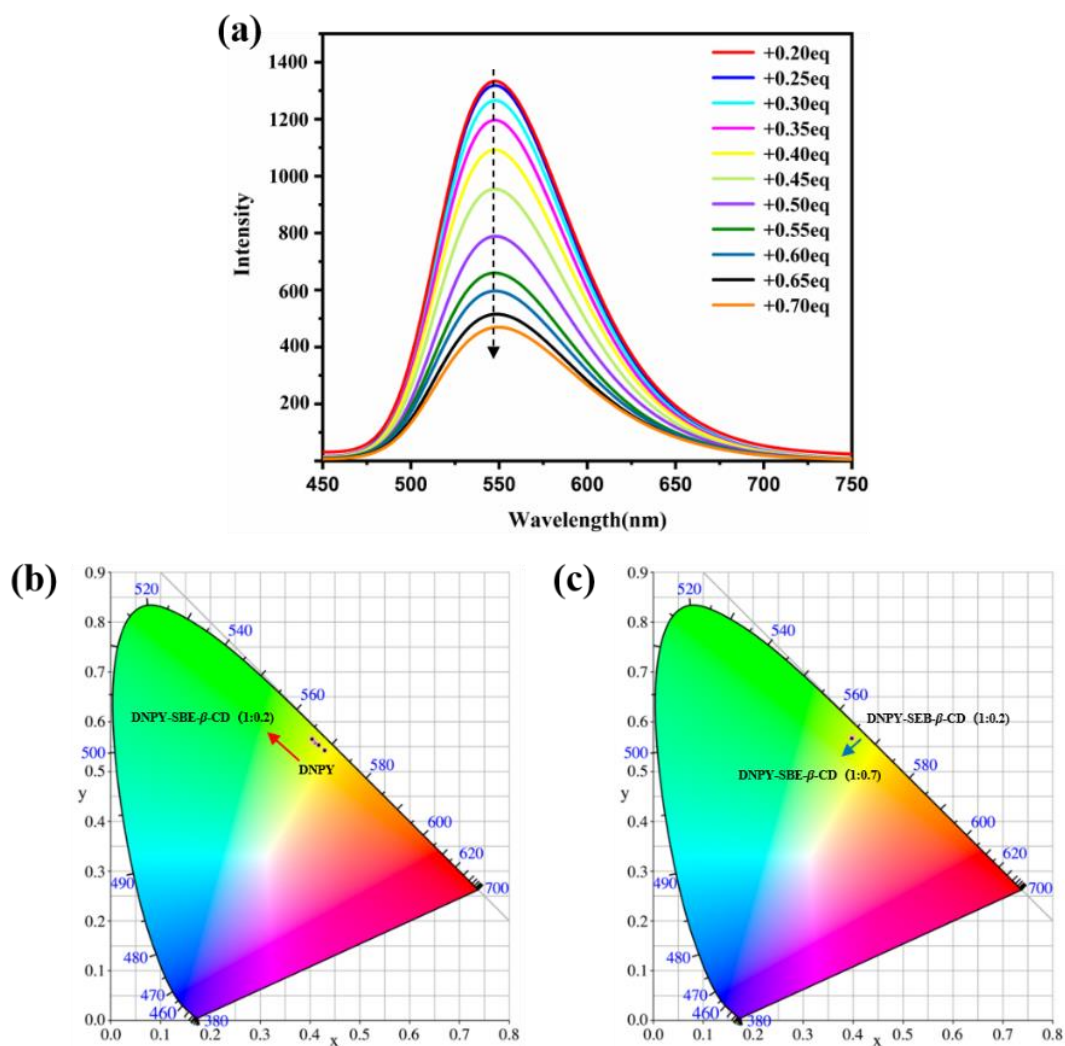
**Fig. S4**  $^{13}\text{C}$  NMR spectra of DNPY in  $\text{DMSO-}d_6$ .



**Fig. S5** The UV-vis absorption spectra of DNPY with gradual addition of SBE-β-CD in the aqueous solution.  $[\text{DNPY}] = 1.0 \times 10^{-5}$  mol/L.



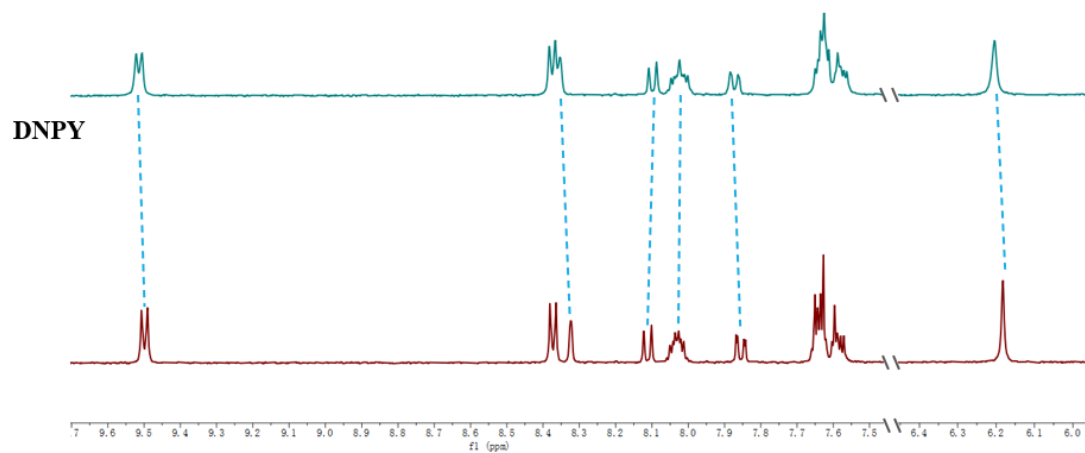
**Fig. S6** Time-resolved fluorescence decay curves of DNPY and DNPY-SBE- $\beta$ -CD. [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE- $\beta$ -CD] =  $2.0 \times 10^{-6}$  mol/L.



**Fig. S7** (a) Fluorescence emission spectra of DNPY in aqueous solutions with different concentrations of SBE- $\beta$ -CD (from 0.20 equiv. to 0.70 equiv.); (b) CIE chromaticity coordinates of DNPY at different concentrations of SBE- $\beta$ -CD (from 0 to 0.20 equiv.); (c) CIE chromaticity coordinates of DNPY at different concentrations of SBE- $\beta$ -CD (from 0.20 equiv. to 0.70 equiv.).

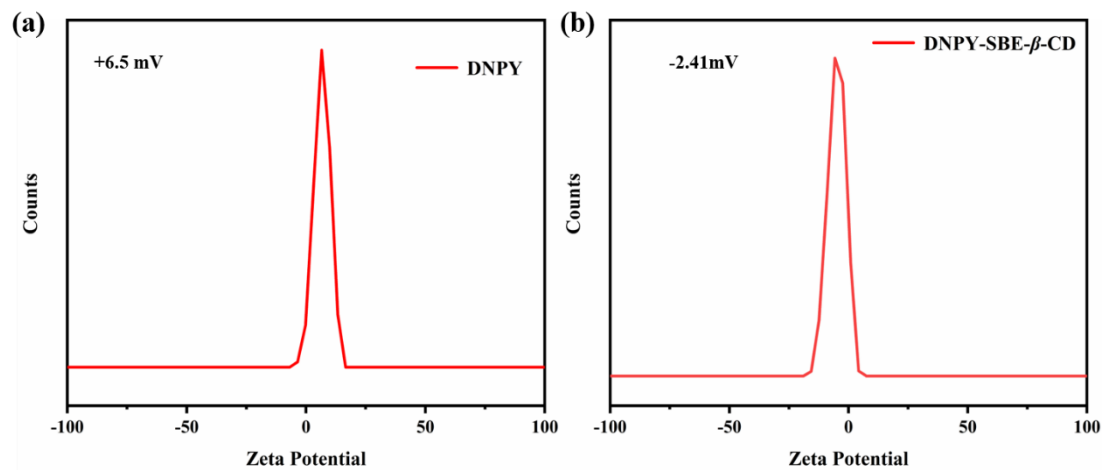
$[\text{DNPY}] = 1.0 \times 10^{-5} \text{ mol/L}$ ,  $[\text{SBE-}\beta\text{-CD}] = 2.0 \times 10^{-6} \text{ mol/L}$ .

**DNPY + 0.20 eq. SBE- $\beta$ -CD**

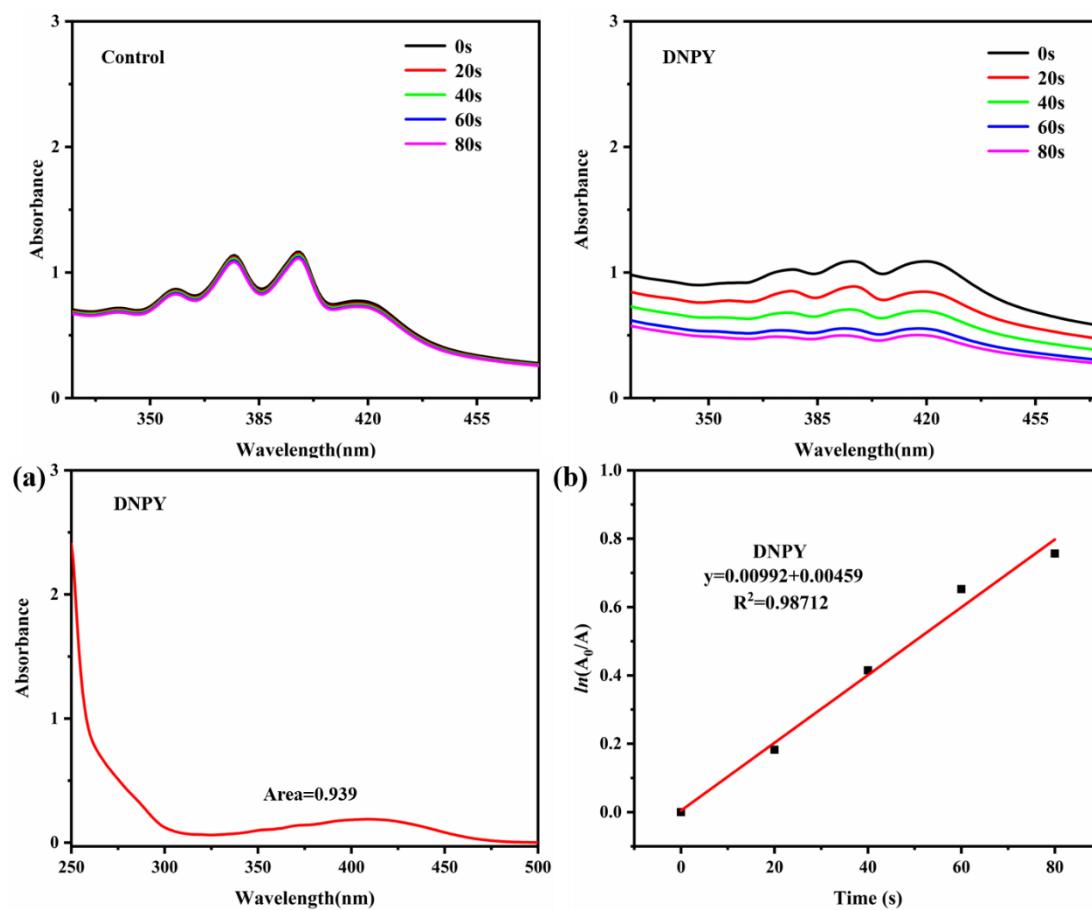


**Fig. S8**  $^1\text{H}$  NMR spectra of DNPY in the presence of 0.20 equiv. SBE- $\beta$ -CD in  $\text{DMSO-}d_6$ . [DNPY]

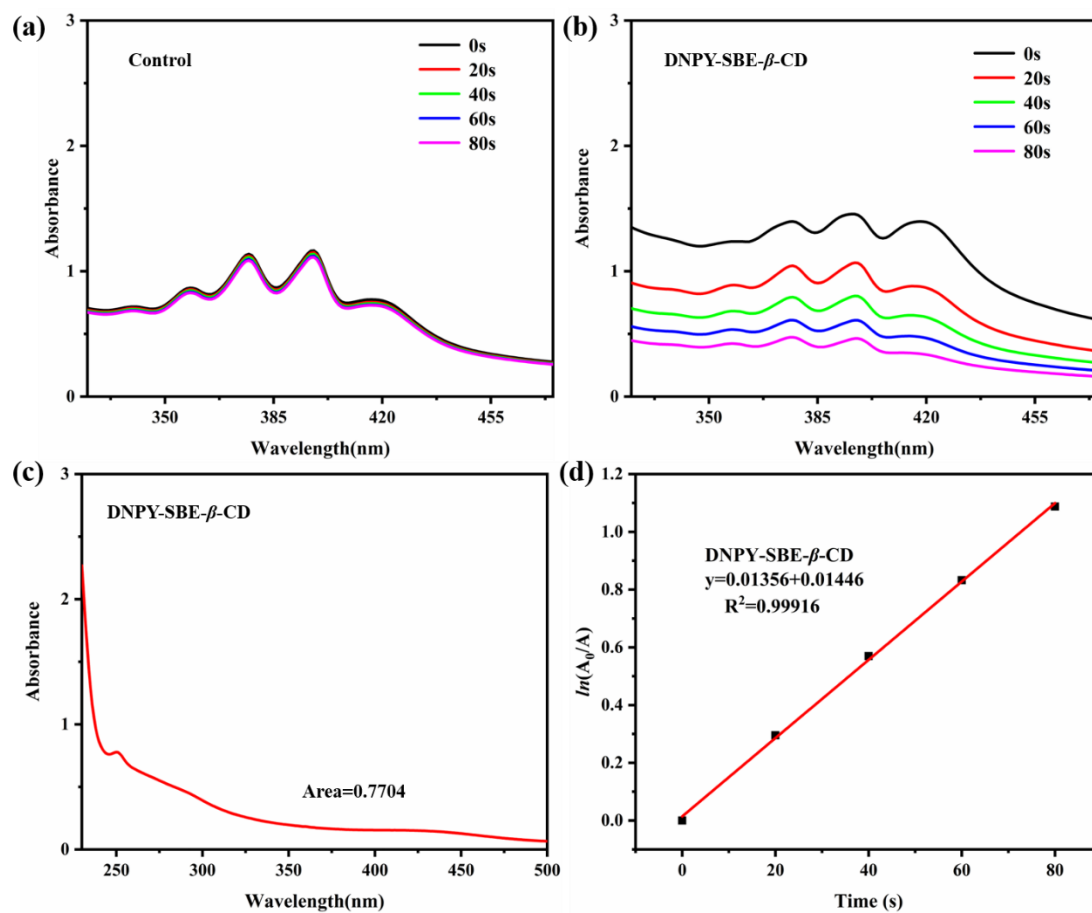
$= 1.0 \times 10^{-5}$  mol/L, [SBE- $\beta$ -CD] =  $2.0 \times 10^{-6}$  mol/L.



**Fig. S9** Zeta potential of DNPY before and after the addition of 0.20 equiv. SBE-β-CD. [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE-β-CD] =  $2.0 \times 10^{-6}$  mol/L.

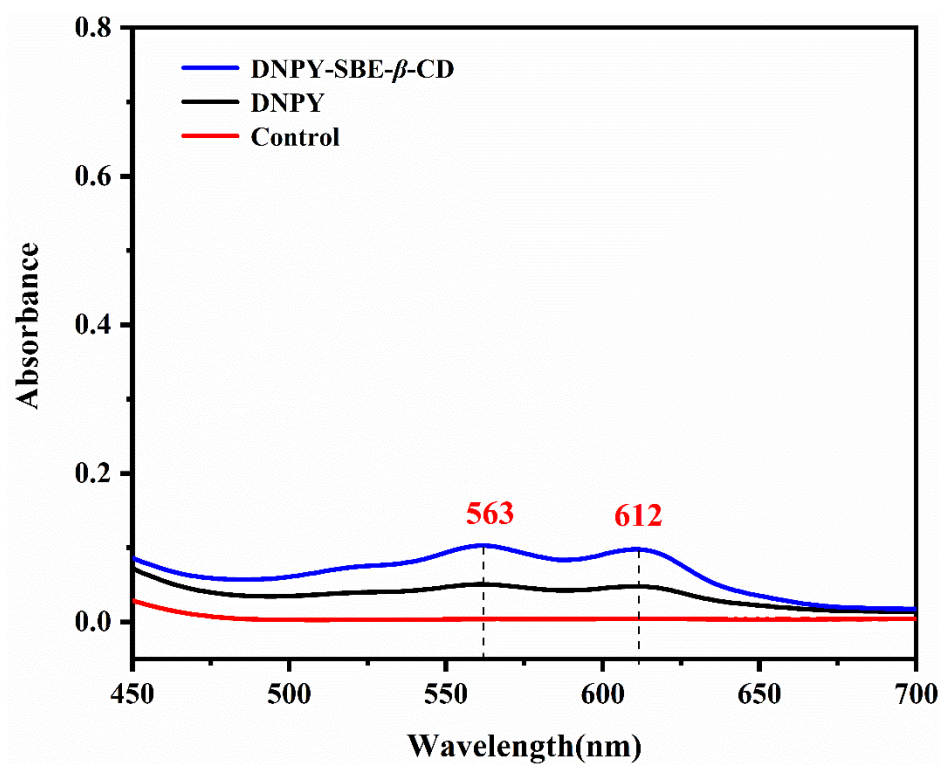


**Fig. S10** The absorption spectra of ABDA after irradiation (410-415 nm, 10 W) for different time in the presence of (a) Control: ABDA without any additive; (b) DNPY; (c) The UV-vis absorption spectra of DNPY in the aqueous solution; (d) The decomposition rates of ABDA in the presence of DNPY.

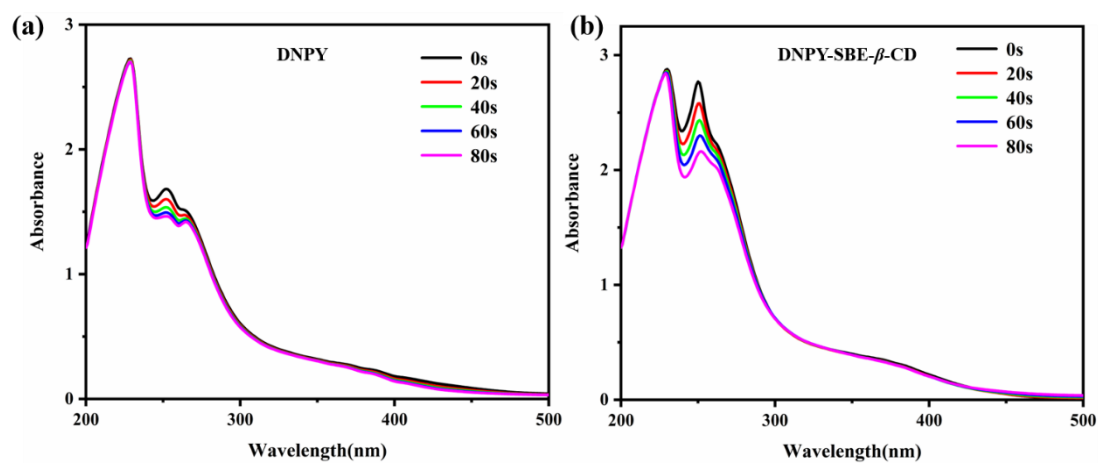


**Fig. S11** The absorption spectra of ABDA after irradiation (410-415 nm, 10 W) for different time in the presence of (a) Control: ABDA without any additive; (b) DNPY-SBE- $\beta$ -CD; (c) The UV-vis absorption spectra of DNPY-SBE- $\beta$ -CD in the aqueous solution; (d) The decomposition rates of ABDA in the presence of DNPY-SBE- $\beta$ -CD.





**Fig. S12** UV-vis absorption spectra for cationic radicals of TMPD generated by indicated samples under the same conditions (Control: TMPD without any additive).



**Fig. S13** The absorption spectra of NBT after irradiation (410-415 nm, 10 W) for different times in the presence of (a) DNPY; (b) DNPY-SBE- $\beta$ -CD.

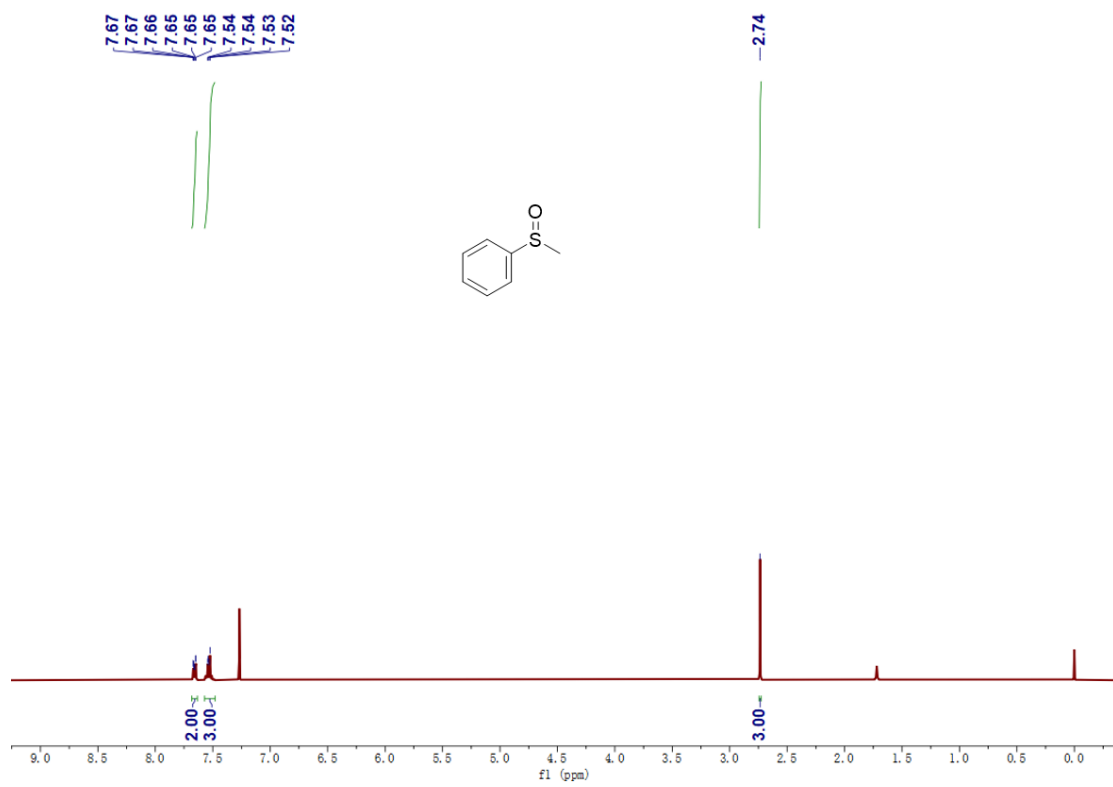
**Table S1** Comparison of  $^1\text{O}_2$  production efficiencies.

| Entry | Systems  | $\Phi_{\Delta}$ ( $^1\text{O}_2$ ) | literatures |
|-------|--|------------------------------------|-------------|
| 1     | NI-S   | 0.32                               | S1          |
| 2     | MONI-S   | 0.74                               |             |
| 3     | MANI-S   | ~1.00                              |             |
| 4     | mCN-2I-BODIPY  | 0.526                              | S2          |
| 5     | TPP  | 0.576                              |             |
| 6     | mTz-2I-BODIPY  | 0.217                              |             |
| 7     | pNH-Tz-2I-BODIPY   | 0.440                              |             |
| 8     | pNH-Tz-TPP   | 0.591                              |             |
| 9     | (mTz-Nor)-2I-BODIPY  | 0.505                              |             |
| 10    | pNH-(Tz-Nor)-2I-BODIPY   | 0.473                              |             |
| 11    | pNH-(Tz-Nor)-TPP   | 0.581                              |             |
| 12    | P <sub>2</sub>   | 0.14                               | S3          |
| 13    | P <sub>2</sub> -NMeI   | 0.50                               |             |
| 14    | P <sub>2</sub> C <sub>2</sub> -NMeI                            | 0.25                               |             |
| 15    | P <sub>2</sub> -NMeOAc   | 0.36                               |             |
| 16    | P <sub>2</sub> -SO <sub>3</sub> NH <sub>4</sub>                | 0.59                               |             |
| 17    | P <sub>2</sub> C <sub>2</sub> -CO <sub>2</sub> NH <sub>4</sub> | 0.24                               |             |
| 18    | P <sub>2</sub> -Suc  | 0.43                               |             |
| 19    | H <sub>2</sub> TCPP  | 0.53                               | S4          |
| 20    | PCN-222/MOF545(FB)   | 0.35                               | S5          |
| 21    | (R)-DTP-COF-QA   | 0.57                               | S6          |
| 22    | TfR/TPETH-2T7  | 0.92                               | S7          |
| 23    | TPCI   | 0.986                              | S8          |
| 24    | 1•4Cl <sup>-</sup>   | 1.30                               |             |
| 25    | 1•2Cl <sup>-</sup>   | 0.67                               | S9          |
| 26    | TTDPzMg(H <sub>2</sub> O)                                      | 0.30                               |             |
| 27    | TTDPzGaCl  | 0.69                               |             |
| 28    | TTDPzAlCl  | 0.35                               |             |
| 29    | TTDPzCd  | ≤0.2                               |             |
| 30    | TTDPzCu  | 0.08                               |             |
| 31    | TTDPzZn  | 0.52                               |             |
| 32    | 3,4-TPyPzZn  | 0.56                               | S10         |
| 33    | 2,3-TPyPzZn  | 0.16                               | S11         |
| 34    | TPyzPzZn   | 0.487                              | S12         |
| 35    | ZnPc   | 0.56                               | S13         |
| 36    | ZnPc 6   | 0.47                               | S14         |
| 37    | 1  | 0.23                               | S15         |
| 38    | 4  | 0.196                              | S16         |
| 39    | [(PtCl <sub>2</sub> )LMg(H <sub>2</sub> O)]                    | 0.40                               | S17         |

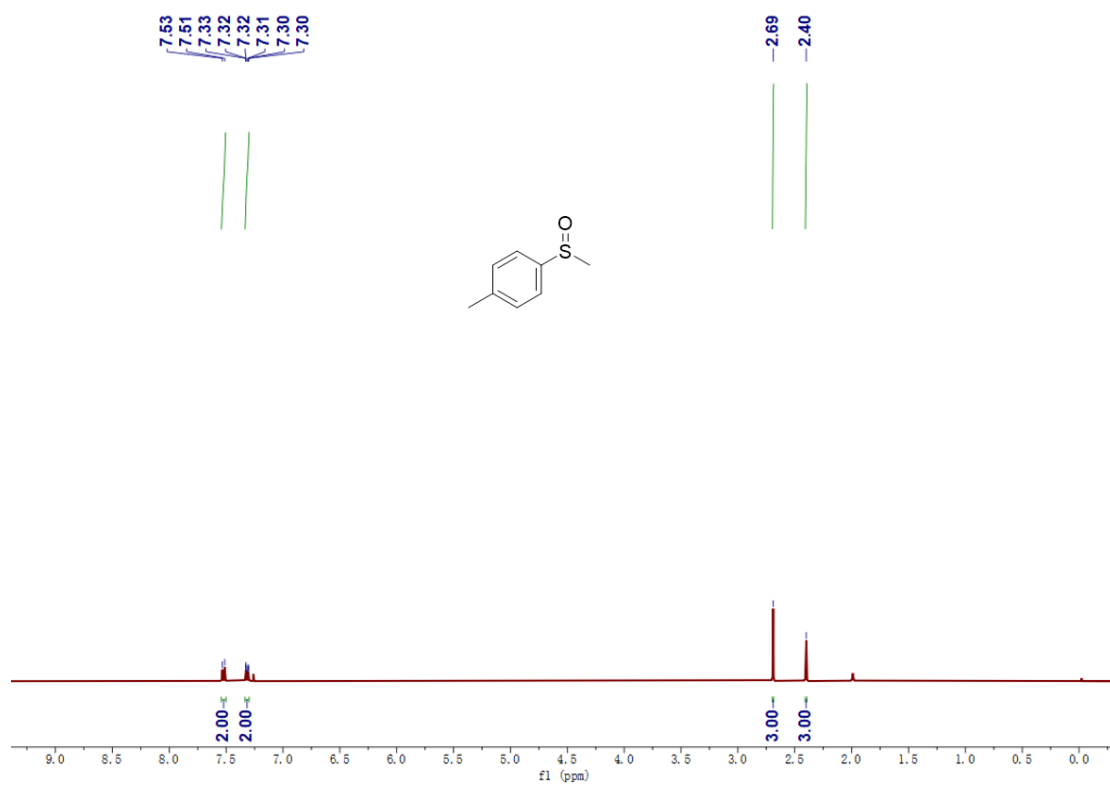
|    |  |        |           |
|----|--|--------|-----------|
| 40 | 16   | 0.137  | S18       |
| 41 | 19   | 0.0073 | S19       |
| 42 | 10   | 0.42   | S20       |
| 43 | [{Pd(OAc) <sub>2</sub> } <sub>4</sub> LZn] | 0.43   | S21       |
| 44 | 4b   | 0.54   | S22       |
| 45 | ZnTM <sub>2,3</sub> PyPz                   | 0.65   | S23       |
| 46 | ZnPc 2                                     | 0.50   | S24       |
| 47 | ZnAPc <sup>4+</sup>                        | 0.50   | S25       |
| 48 | ZnPc 1                                     | 0.50   | S26       |
| 49 | 3  | 0.72   | S27       |
| 50 | DNPY                                       | 0.597  | This work |
|    | DNPY-SBE-β-CD                              | 0.994  |           |
|    | DNPY-SBE-β-CD+RhB,                         | 0.069  |           |
|    | DNPY-SBE-β-CD+SR101                        | 0.042  |           |
|    | DNPY-SBE-β-CD+RhB+SR101                    | 0.054  |           |

**Table S2** Comparison of  $O_2^{\cdot-}$  production efficiencies.

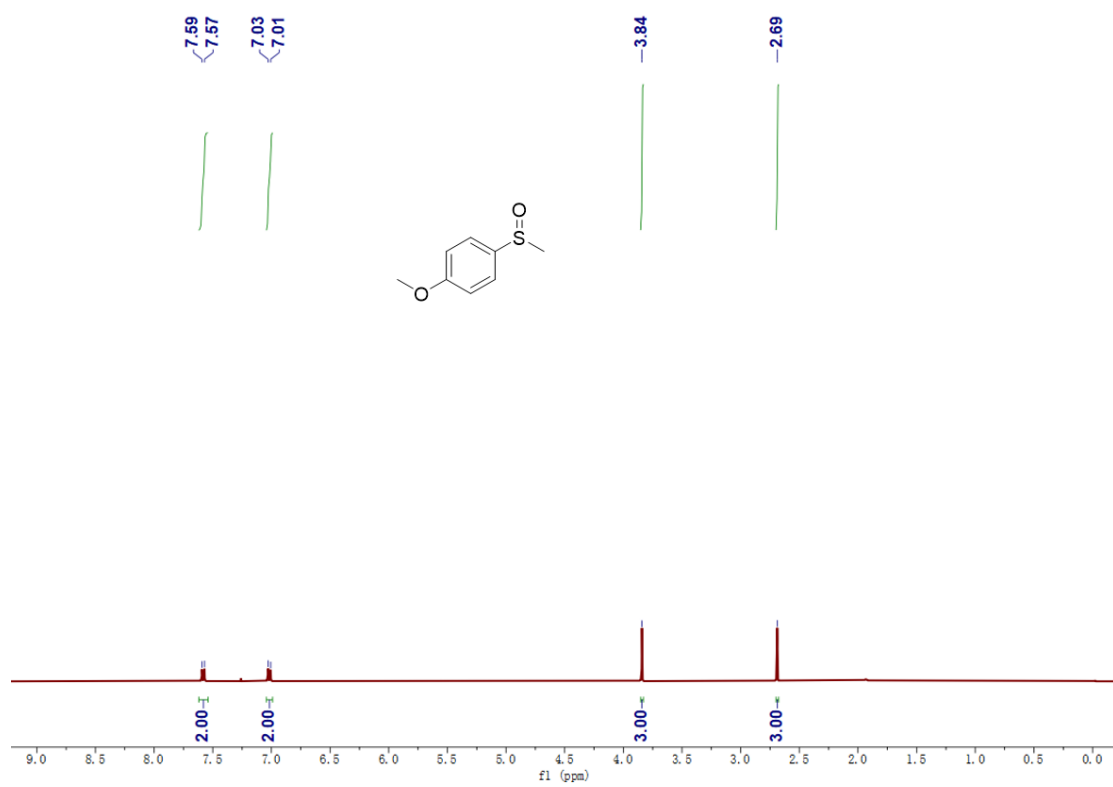
| Entry | Systems  | $\Phi_{\Delta}$ ( $O_2^{\cdot-}$ )                | literatures |
|-------|--|---|-------------|
| 1     | 20%BI  | 67% (NBT)   | S28         |
| 2     | BiOBr  | 10.9% (NBT)                                       | S29         |
| 3     | TiO <sub>2</sub>   | 8.0 $\mu$ M                                       | S30         |
| 4     | CeO <sub>2</sub>   | 8.4 $\mu$ M                                       |             |
| 5     | SiO <sub>2</sub>   | -   |             |
| 6     | Al <sub>2</sub> O <sub>3</sub>   | -   |             |
| 7     | ZnO  | 167 $\mu$ M                                       |             |
| 8     | CuO  | -   |             |
| 9     | Fe <sub>2</sub> O <sub>3</sub>   | 18.1 $\mu$ M                                      |             |
| 10    | Disrupted NanoMANI-S   | 3.0-fold greater amount of $O_2^{\cdot-}$ than MB | S1          |
| 11    | DNPY<br>DNPY-SBE- $\beta$ -CD<br>DNPY-SBE- $\beta$ -CD+RhB<br>DNPY-SBE- $\beta$ -CD+SR101<br>DNPY-SBE- $\beta$ -CD+RhB+SR101 | 6.3%<br>9.2%<br>19.7%<br>24.9%<br>44.1%           | This work   |



**Fig. S14** <sup>1</sup>H NMR spectra of **2a** in CDCl<sub>3</sub>.



**Fig. S15**  $^1\text{H}$  NMR spectra of **2b** in  $\text{CDCl}_3$ .



**Fig. S16** <sup>1</sup>H NMR spectra of **2c** in CDCl<sub>3</sub>.



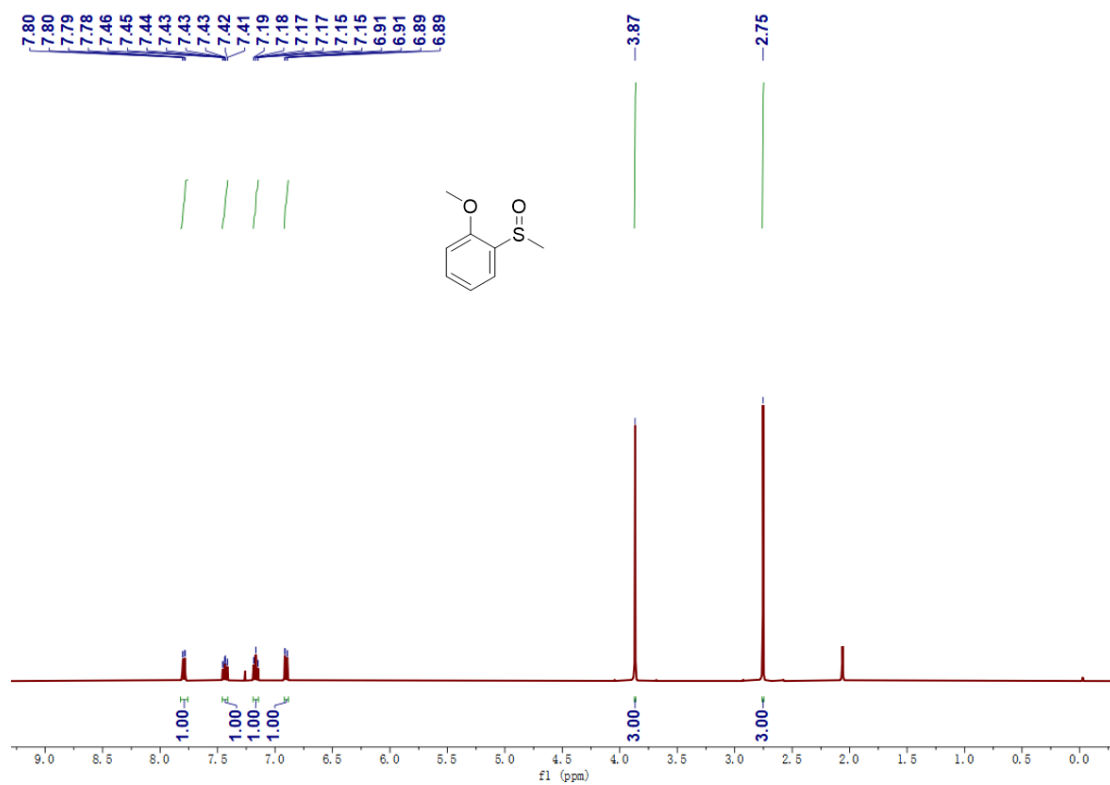
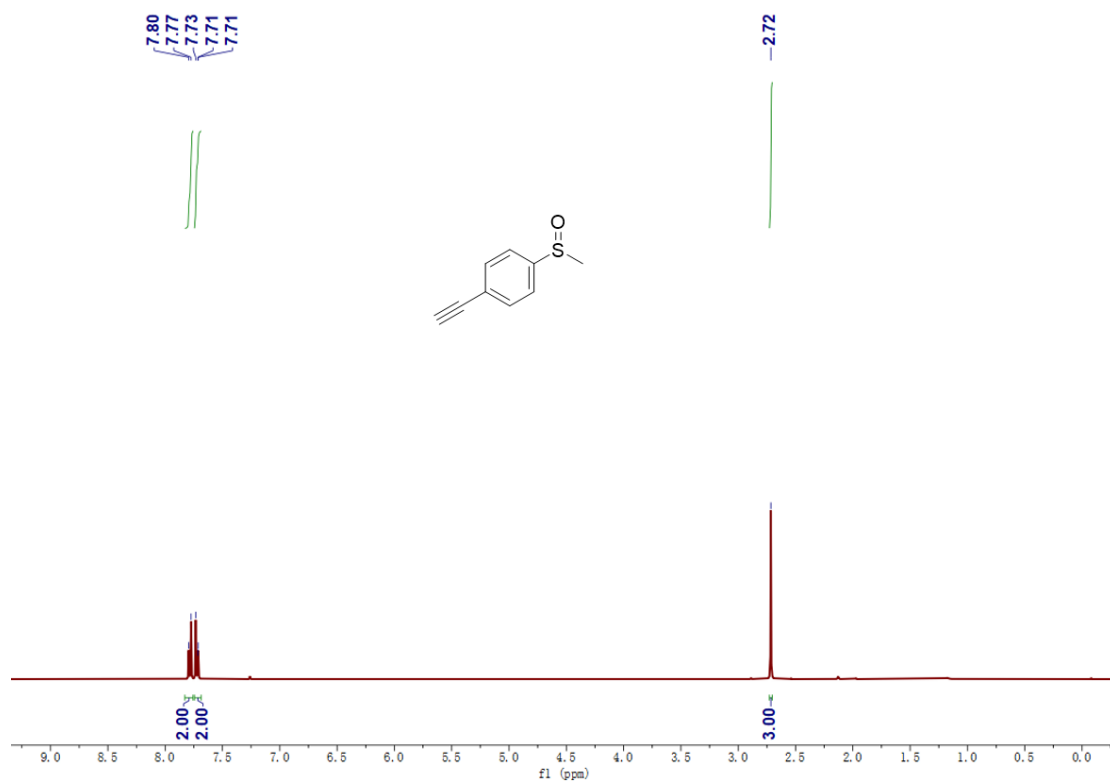
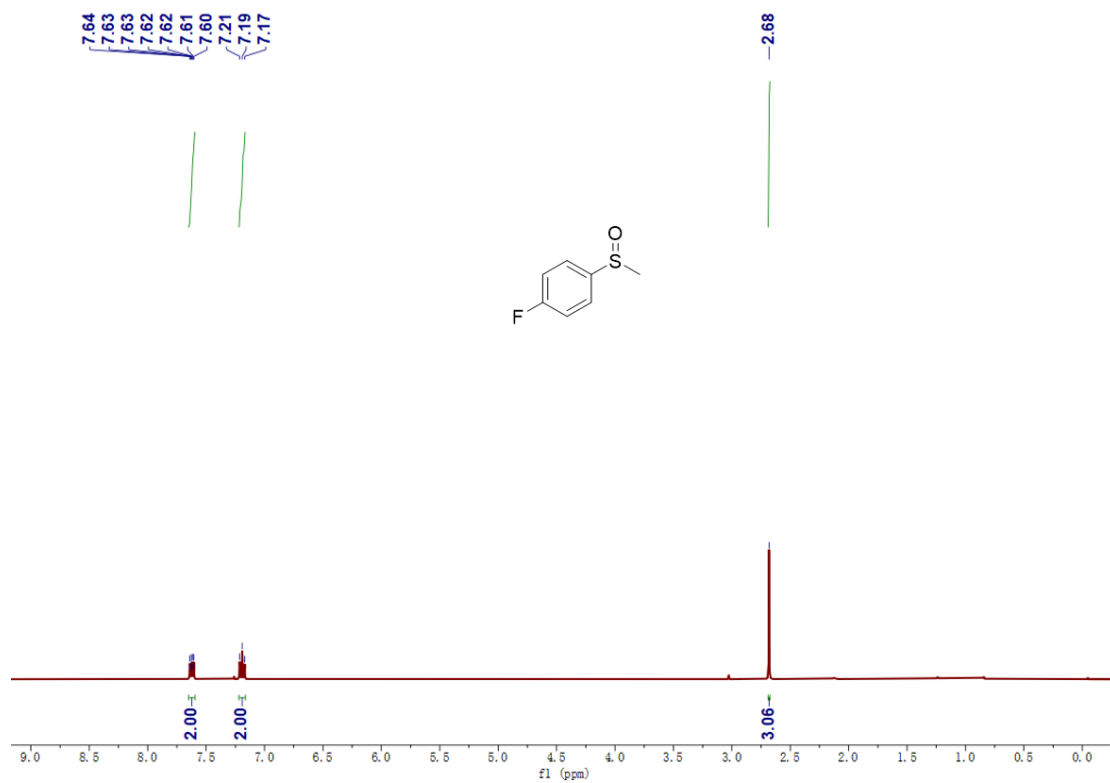


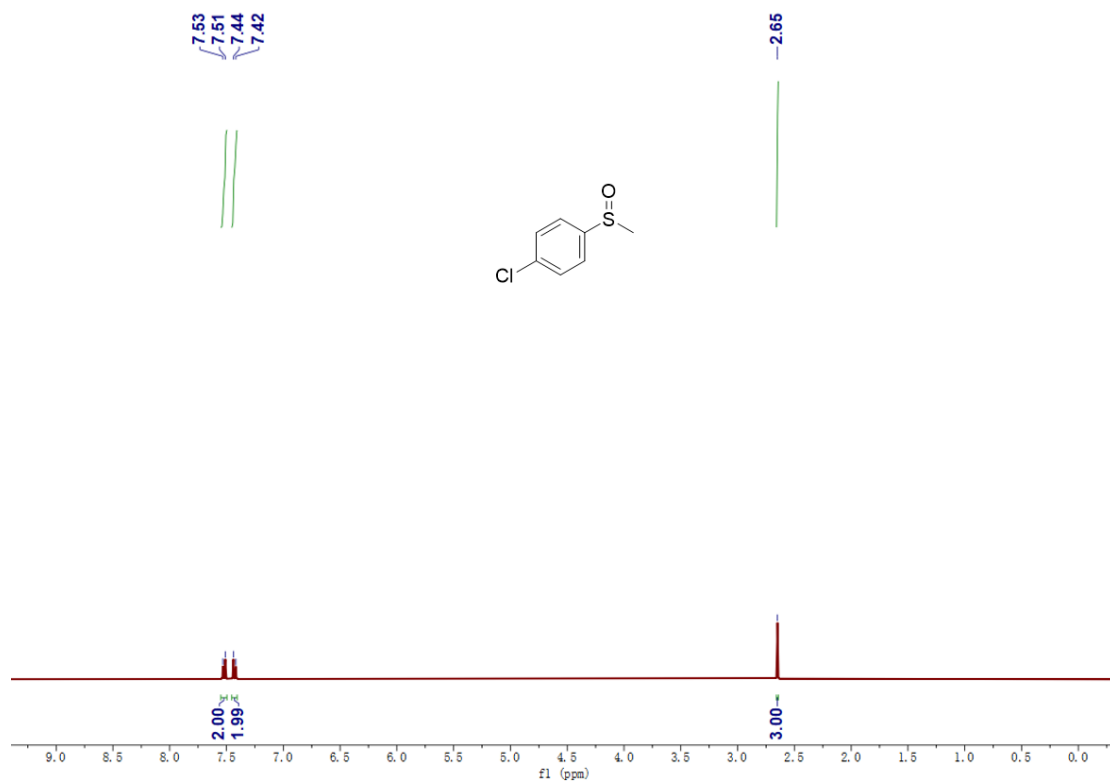
Fig. S17  $^1\text{H}$  NMR spectra of **2d** in  $\text{CDCl}_3$ .



**Fig. S18**  $^1\text{H}$  NMR spectra of **2e** in  $\text{CDCl}_3$ .



**Fig. S19**  $^1\text{H}$  NMR spectra of **2f** in  $\text{CDCl}_3$ .



**Fig. S20**  $^1\text{H}$  NMR spectra of **2g** in  $\text{CDCl}_3$ .

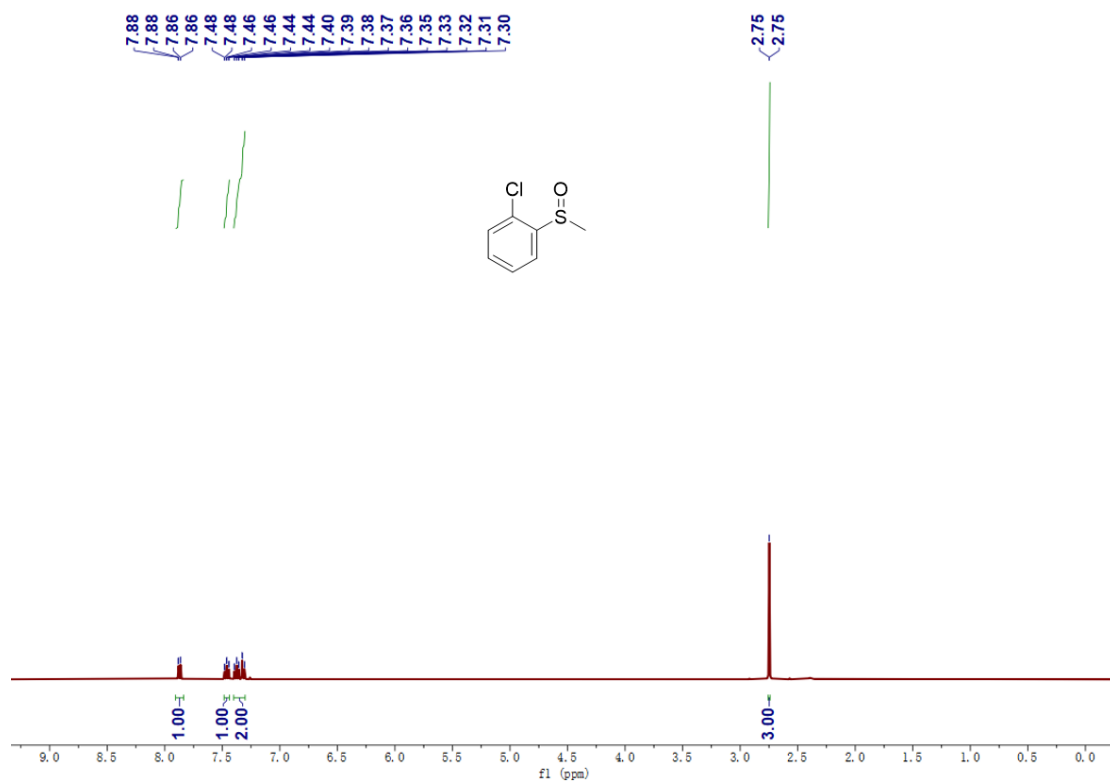
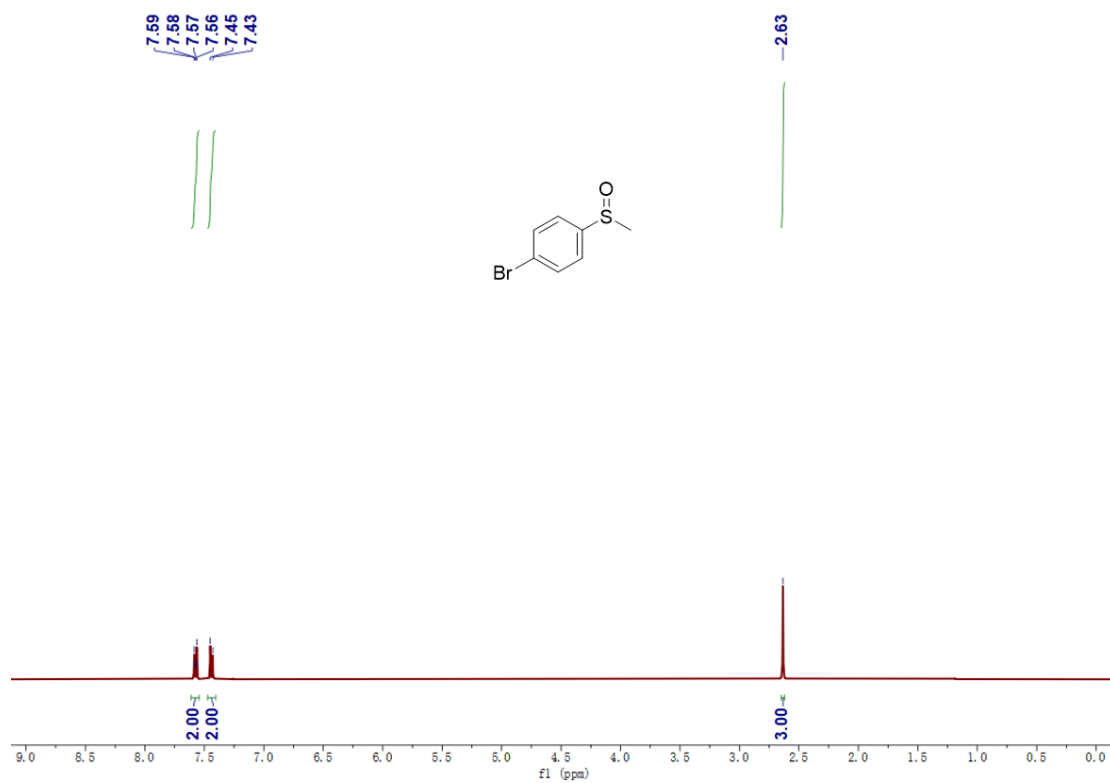


Fig. S21  $^1\text{H}$  NMR spectra of **2h** in  $\text{CDCl}_3$ .



**Fig. S22** <sup>1</sup>H NMR spectra of **2i** in CDCl<sub>3</sub>.

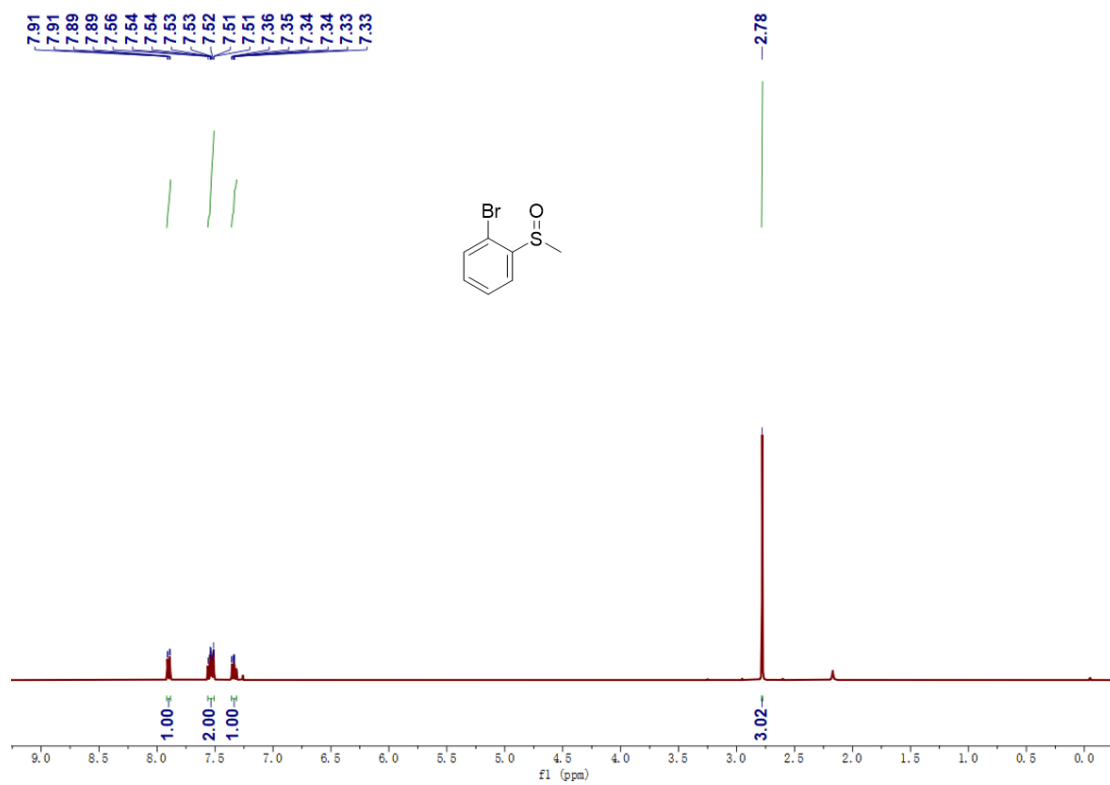
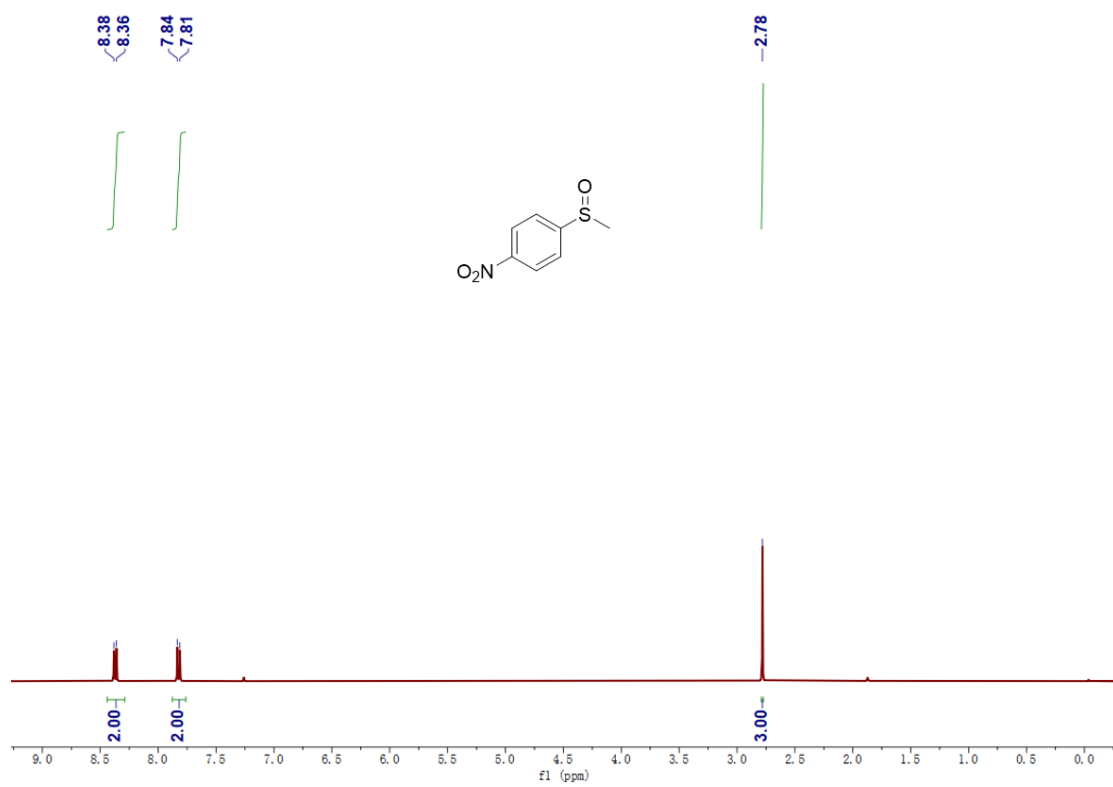
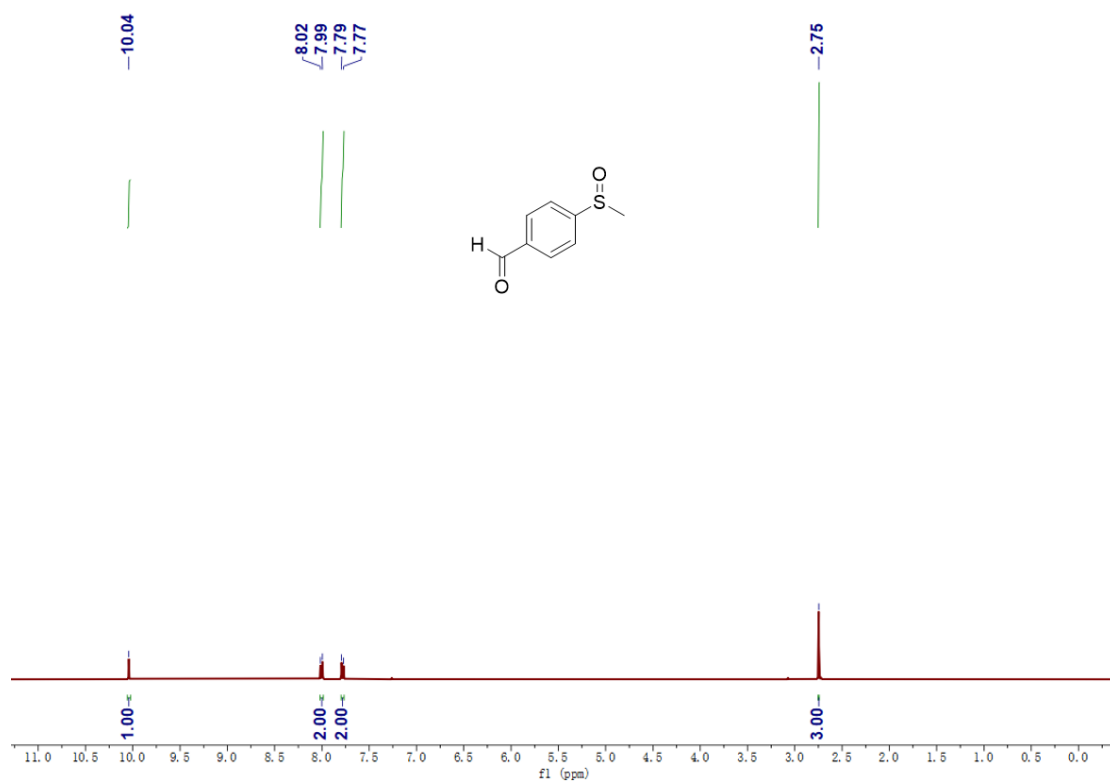


Fig. S23 <sup>1</sup>H NMR spectra of **2j** in CDCl<sub>3</sub>.

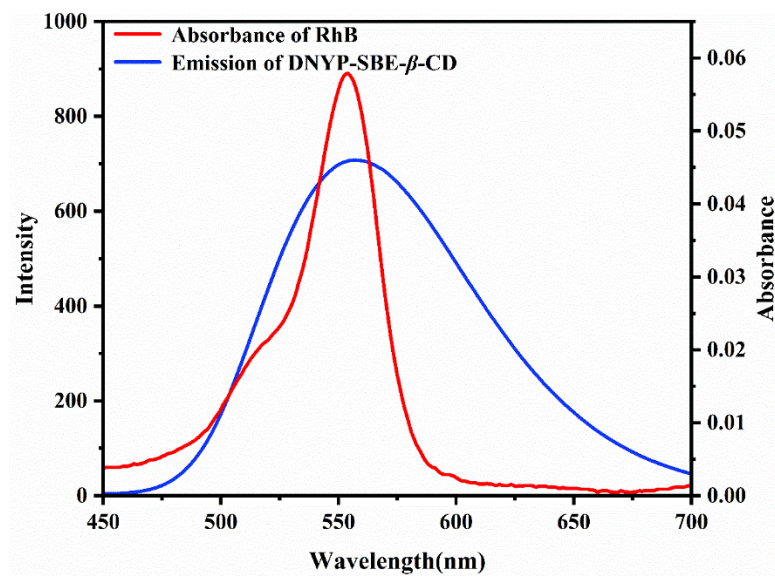


**Fig. S24** <sup>1</sup>H NMR spectra of **2k** in CDCl<sub>3</sub>.

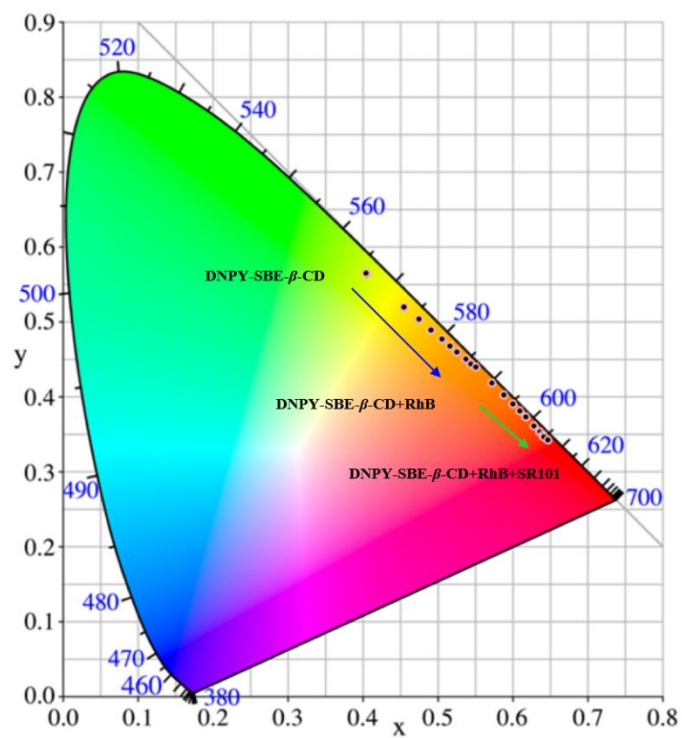




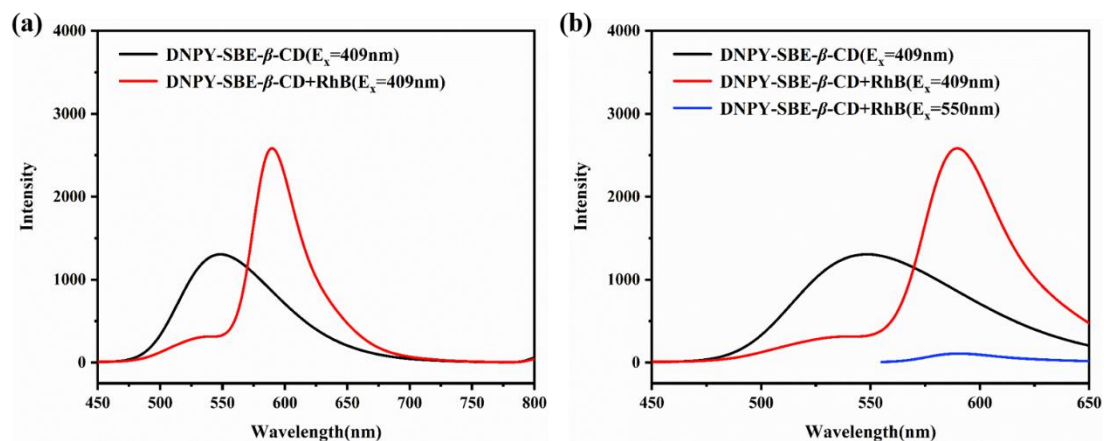
**Fig. S25** <sup>1</sup>H NMR spectra of **21** in CDCl<sub>3</sub>.



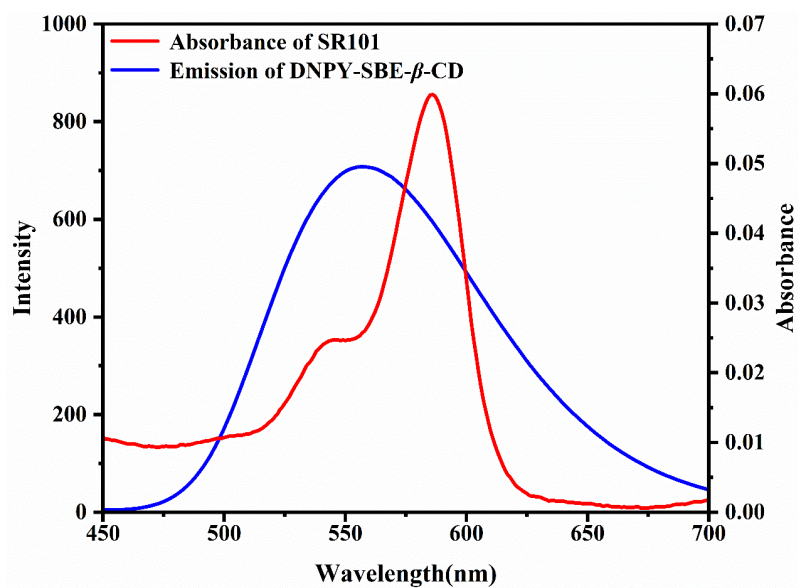
**Fig. S26** The UV-vis absorption spectra of RhB and the fluorescence emission spectra of DNPY-SBE-β-CD. [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE-β-CD] =  $2.0 \times 10^{-6}$  mol/L, [RhB] =  $1.0 \times 10^{-6}$  mol/L.



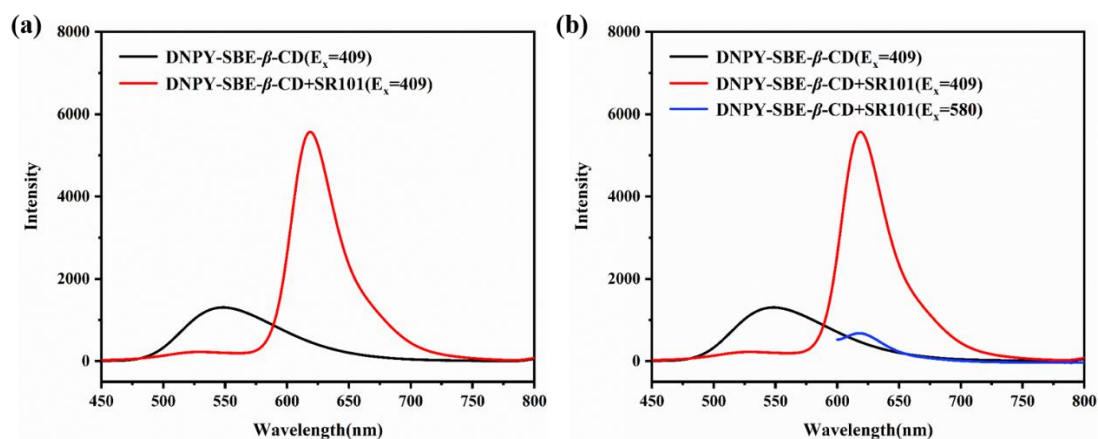
**Fig. S27** CIE chromaticity coordinates of DNPY-SBE- $\beta$ -CD at different concentrations of RhB (from 0 to 0.1 equiv.) and SR101 (from 0 to 0.10 equiv.).  $[\text{DNPY}] = 1.0 \times 10^{-5} \text{ mol/L}$ ,  $[\text{SBE-}\beta\text{-CD}] = 2.0 \times 10^{-6} \text{ mol/L}$ ,  $[\text{RhB}] = 1.0 \times 10^{-6} \text{ mol/L}$ ,  $[\text{SR101}] = 1.0 \times 10^{-6} \text{ mol/L}$ .



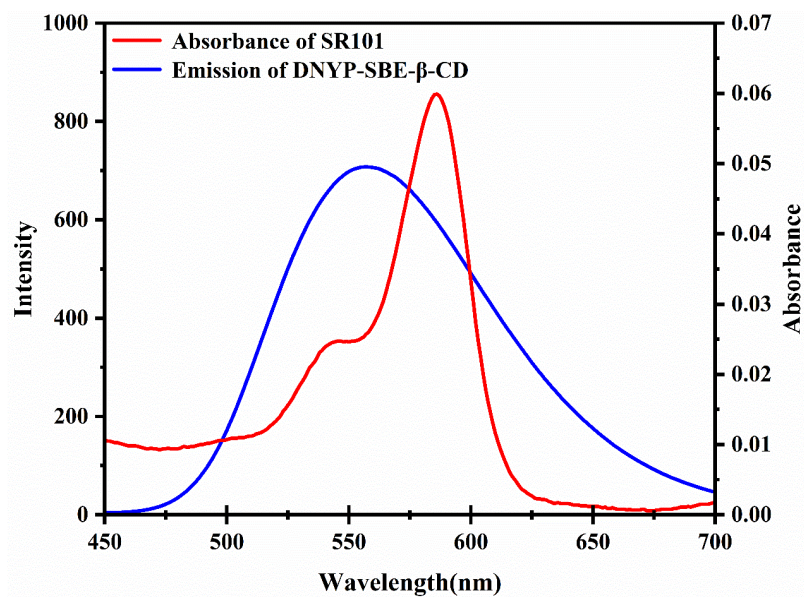
**Fig. S28** (a) Fluorescence emission spectra of DNPY-SBE- $\beta$ -CD and DNPY-SBE- $\beta$ -CD+RhB; (b) Fluorescence emission spectra of DNPY-SBE- $\beta$ -CD+RhB (the red line), DNPY-SBE- $\beta$ -CD+RhB (the blue line), DNPY-SBE- $\beta$ -CD (the black line).  $[\text{DNPY}] = 1.0 \times 10^{-5} \text{ mol/L}$ ,  $[\text{SBE-}\beta\text{-CD}] = 2.0 \times 10^{-6} \text{ mol/L}$ ,  $[\text{RhB}] = 1.0 \times 10^{-6} \text{ mol/L}$ ,  $[\text{SR101}] = 1.0 \times 10^{-6} \text{ mol/L}$ .



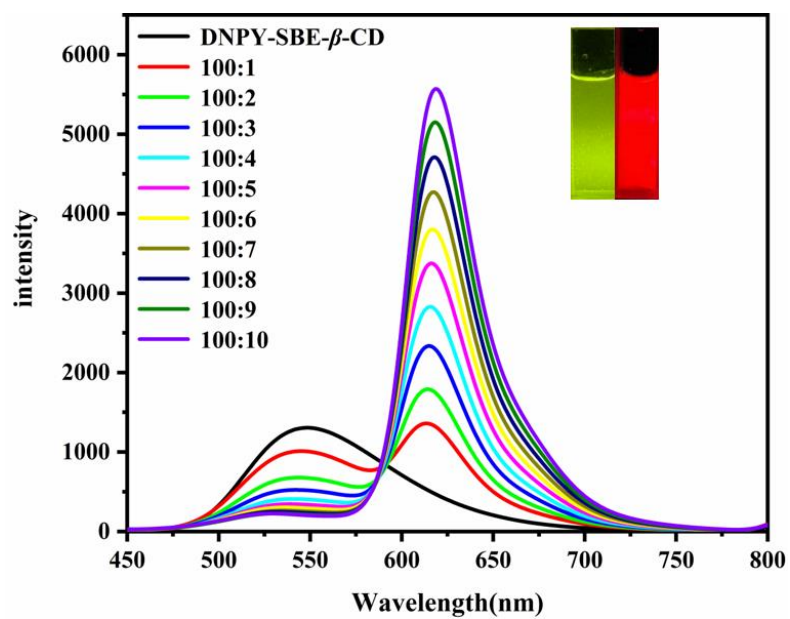
**Fig. S29** The UV-vis absorption spectra of SR101 and the fluorescence emission spectra of DNPY-SBE-β-CD+RhB. [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE-β-CD] =  $2.0 \times 10^{-6}$  mol/L, [RhB] =  $1.0 \times 10^{-6}$  mol/L, [SR101] =  $1.0 \times 10^{-6}$  mol/L.



**Fig. S30** (a) Fluorescence emission spectra of DNPY-SBE-β-CD+RhB and DNPY-SBE-β-CD+RhB+SR101; (b) Fluorescence emission spectra of DNPY-SBE-β-CD+RhB+SR101 (the red line), DNPY-SBE-β-CD+RhB+SR101 (the blue line), DNPY-SBE-β-CD+RhB (the black line).  $[DNPY] = 1.0 \times 10^{-5}$  mol/L,  $[SBE-\beta-CD] = 2.0 \times 10^{-6}$  mol/L,  $[RhB] = 1.0 \times 10^{-6}$  mol/L,  $[SR101] = 1.0 \times 10^{-6}$  mol/L.

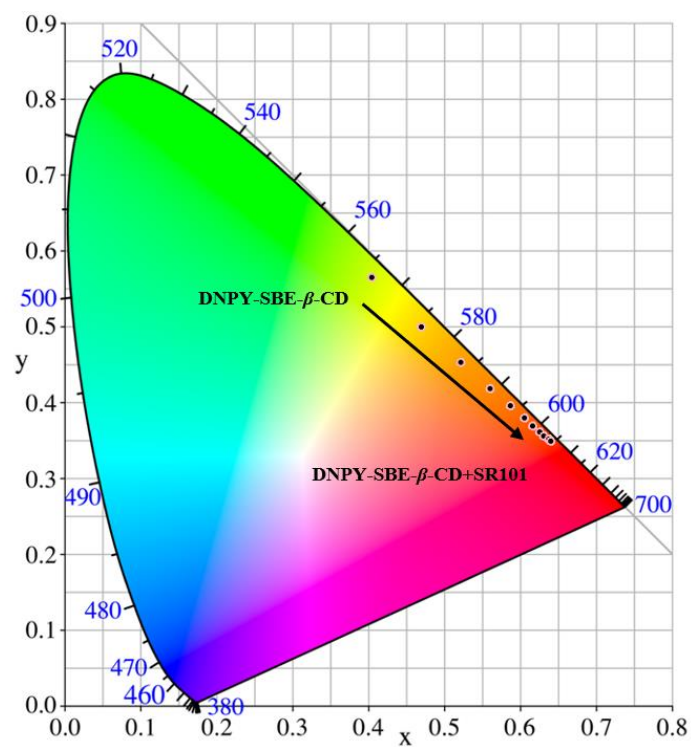


**Fig. S31** The UV-vis absorption spectra of SR101 and the fluorescence emission spectra ( $\lambda_{\text{ex}} = 409$  nm) of DNPY-SBE- $\beta$ -CD.  $[\text{DNPY}] = 1.0 \times 10^{-5}$  mol/L,  $[\text{SBE-}\beta\text{-CD}] = 2.0 \times 10^{-6}$  mol/L,  $[\text{SR101}] = 1.0 \times 10^{-6}$  mol/L.

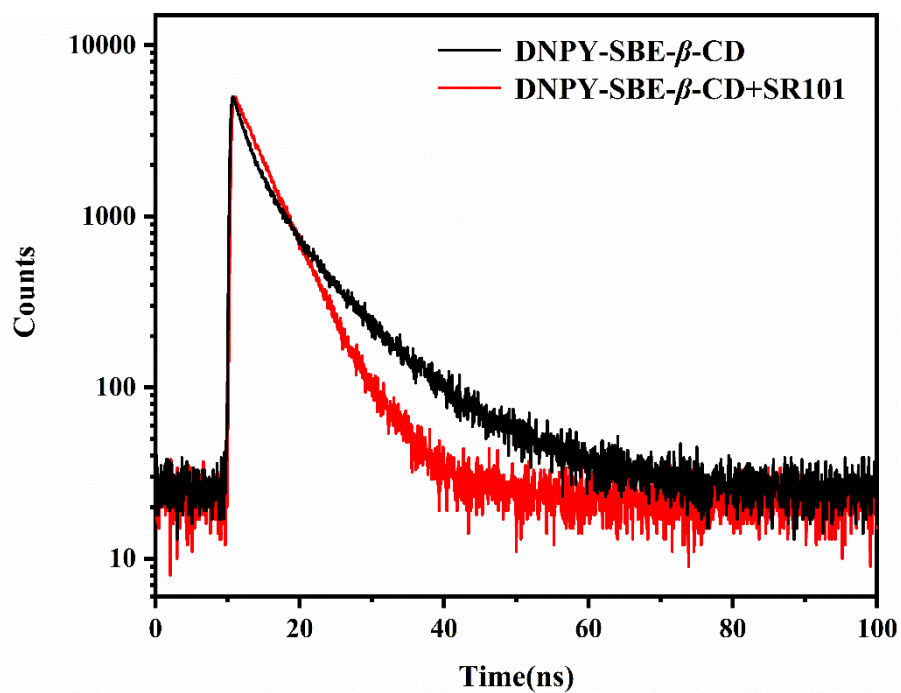


**Fig. S32** Fluorescence emission spectra of DNPY-SBE-β-CD with addition of SR101 in aqueous solution. (Inset: Fluorescence emission colour of DNPY-SBE-β-CD before and after addition of SR101).  $[DNPY] = 1.0 \times 10^{-5}$  mol/L,  $[SBE-\beta-CD] = 2.0 \times 10^{-6}$  mol/L,  $[SR101] = 1.0 \times 10^{-6}$  mol/L.

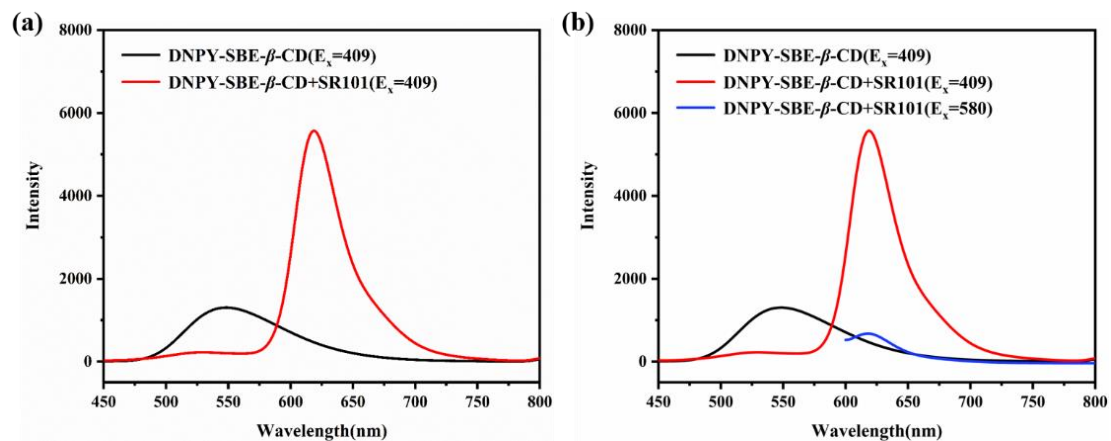




**Fig. S33** CIE chromaticity coordinates of DNPY-SBE-β-CD at different concentrations of SR101 (from 0 to 0.10 equiv.). [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE-β-CD] =  $2.0 \times 10^{-6}$  mol/L, [SR101] =  $1.0 \times 10^{-6}$  mol/L.

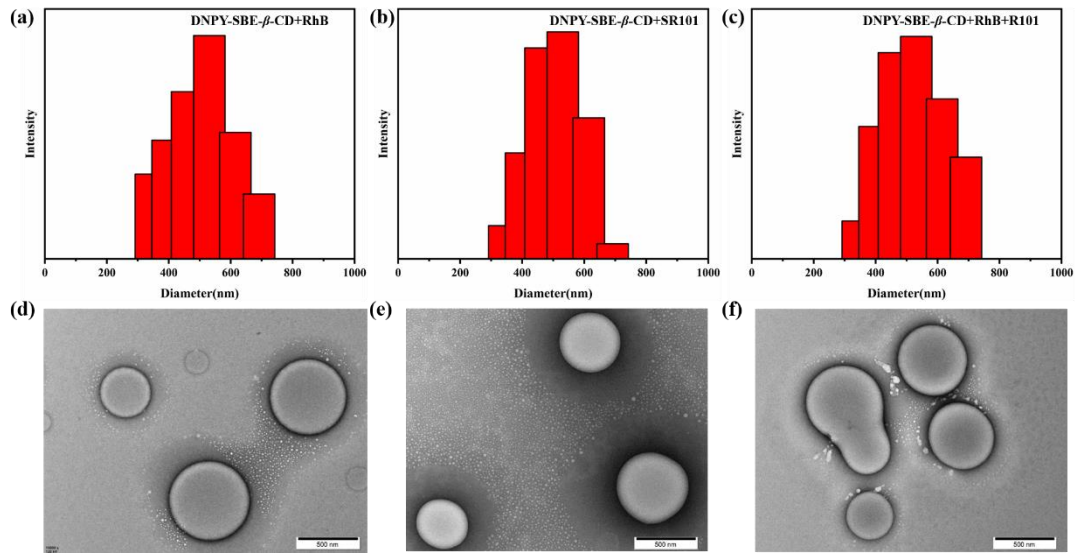


**Fig. S34** Time-resolved fluorescence decay curves of DNPY-SBE- $\beta$ -CD and DNPY-SBE- $\beta$ -CD +SR101. [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE- $\beta$ -CD] =  $2.0 \times 10^{-6}$  mol/L, [SR101] =  $1.0 \times 10^{-6}$  mol/L.

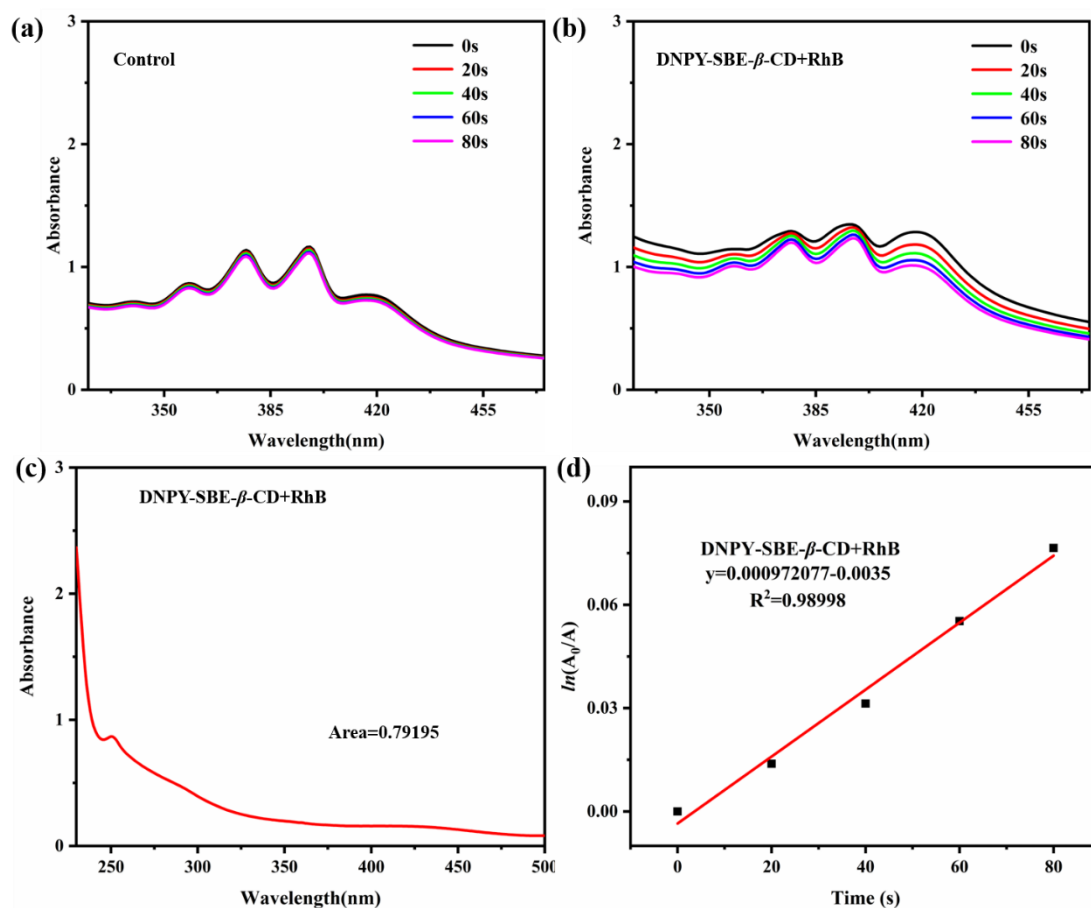


**Fig. S35** (a) Fluorescence emission spectra of DNPY-SBE- $\beta$ -CD and DNPY-SBE- $\beta$ -CD+SR101; (b)

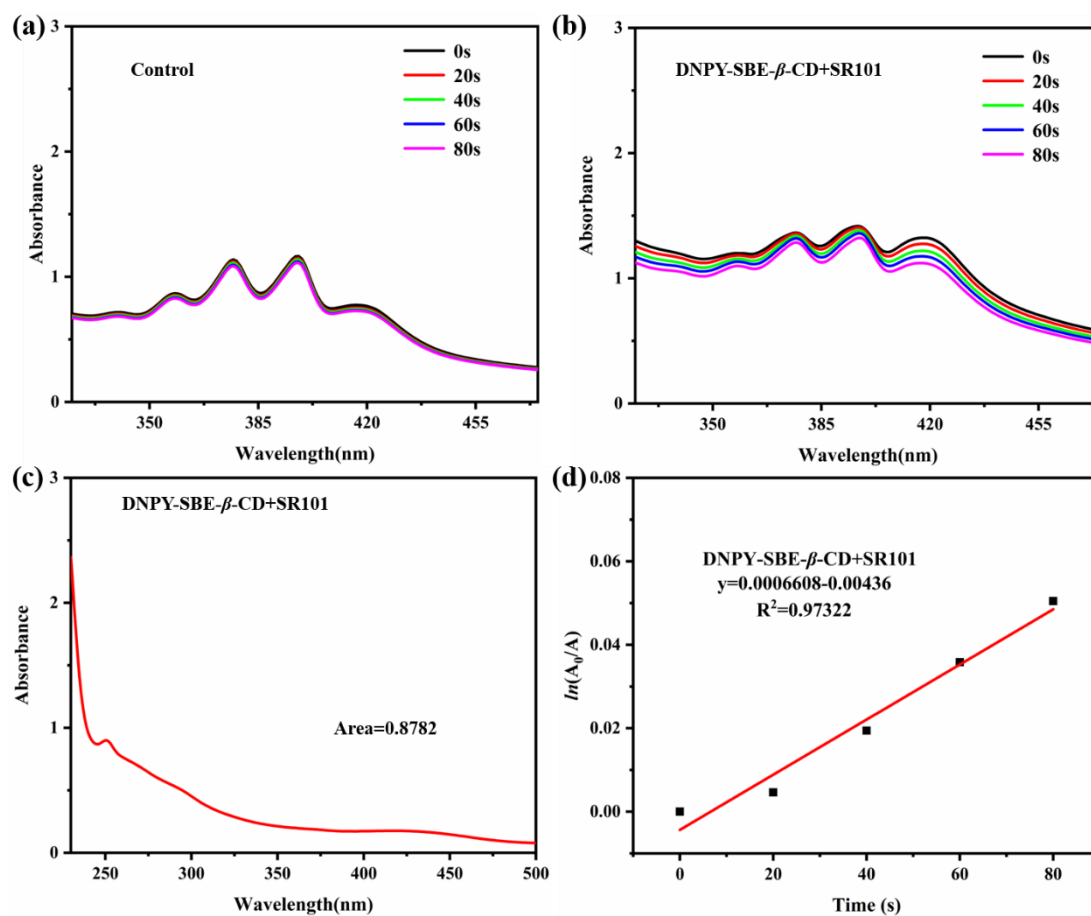
Fluorescence emission spectra of DNPY-SBE- $\beta$ -CD+SR101 (the red line), DNPY-SBE- $\beta$ -CD+SR101 (the blue line), DNPY-SBE- $\beta$ -CD (the black line). [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE- $\beta$ -CD] =  $2.0 \times 10^{-6}$  mol/L, [SR101] =  $1.0 \times 10^{-6}$  mol/L.



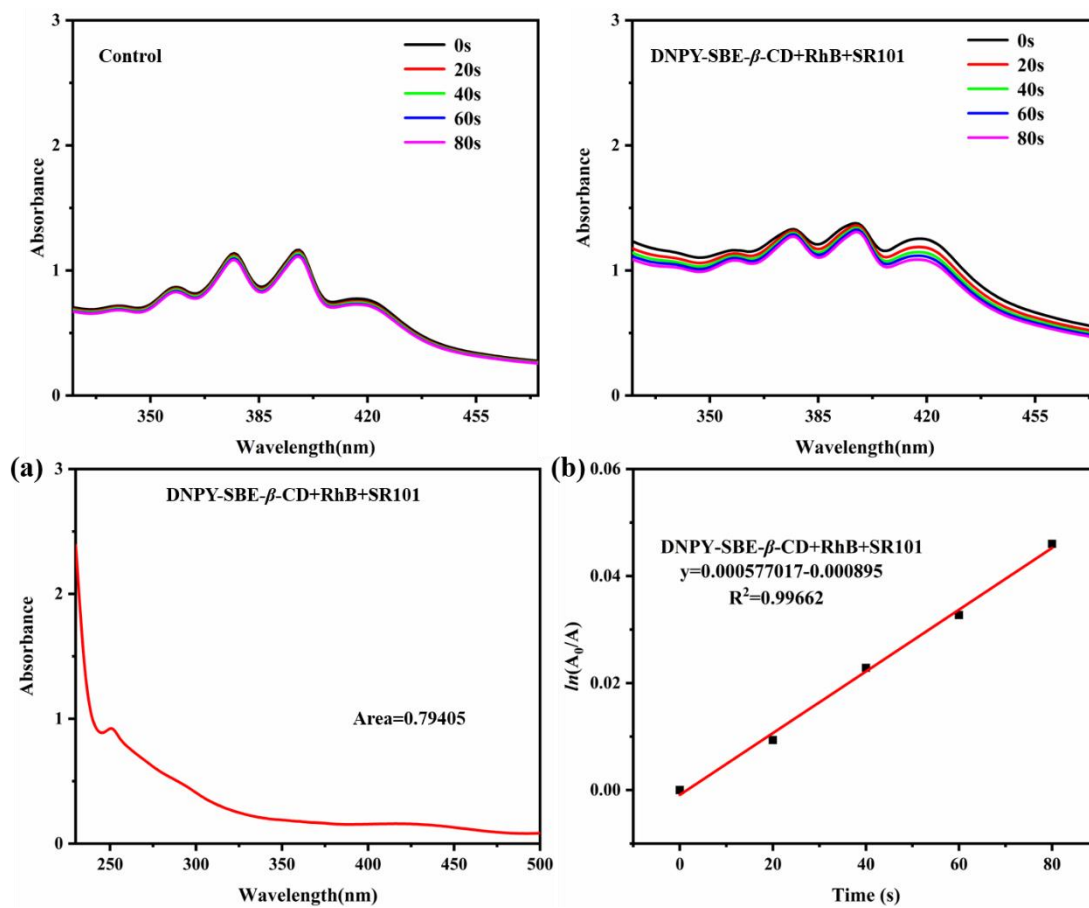
**Fig. S36** DLS and TEM images of (a), (d) DNPY-SBE- $\beta$ -CD+RhB and (b), (e) DNPY-SBE- $\beta$ -CD+SR101 and (c), (f) DNPY-SBE- $\beta$ -CD+RhB+SR101. [DNPY] =  $1.0 \times 10^{-5}$  mol/L, [SBE- $\beta$ -CD] =  $2.0 \times 10^{-6}$  mol/L, [RhB] =  $1.0 \times 10^{-6}$  mol/L, [SR101] =  $1.0 \times 10^{-6}$  mol/L.



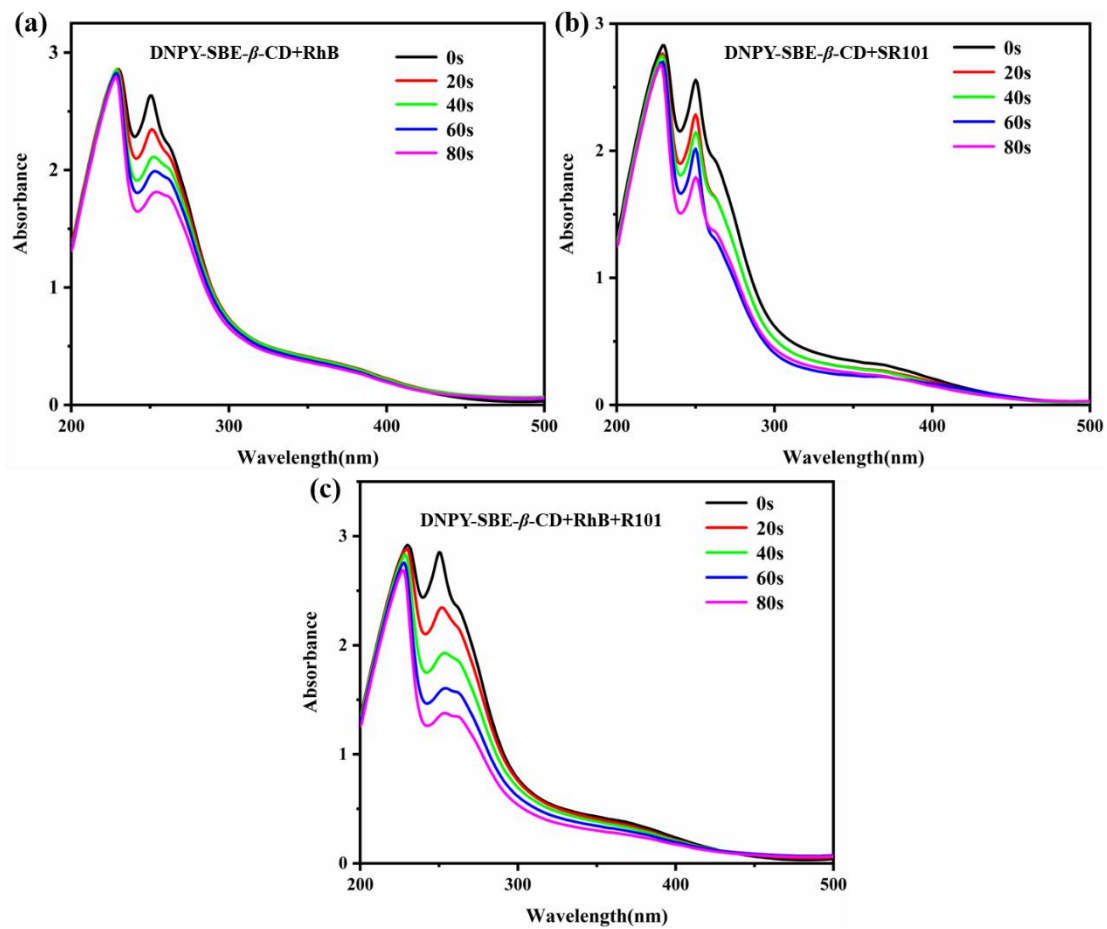
**Fig. S37** The absorption spectra of ABDA after irradiation (410-415 nm, 10 W) for different time in the presence of (a) Control: ABDA without any additive; (b) DNPY-SBE- $\beta$ -CD+RhB; (c) The UV-vis absorption spectra of DNPY-SBE- $\beta$ -CD+RhB in the aqueous solution; (d) The decomposition rates of ABDA in the presence of DNPY-SBE- $\beta$ -CD+RhB.



**Fig. S38** The absorption spectra of ABDA after irradiation (410-415 nm, 10 W) for different time in the presence of (a) Control: ABDA without any additive; (b) DNPY-SBE- $\beta$ -CD+SR101; (c) The UV-vis absorption spectra of DNPY-SBE- $\beta$ -CD+SR101 in the aqueous solution; (d) The decomposition rates of ABDA in the presence of DNPY-SBE- $\beta$ -CD+SR101.

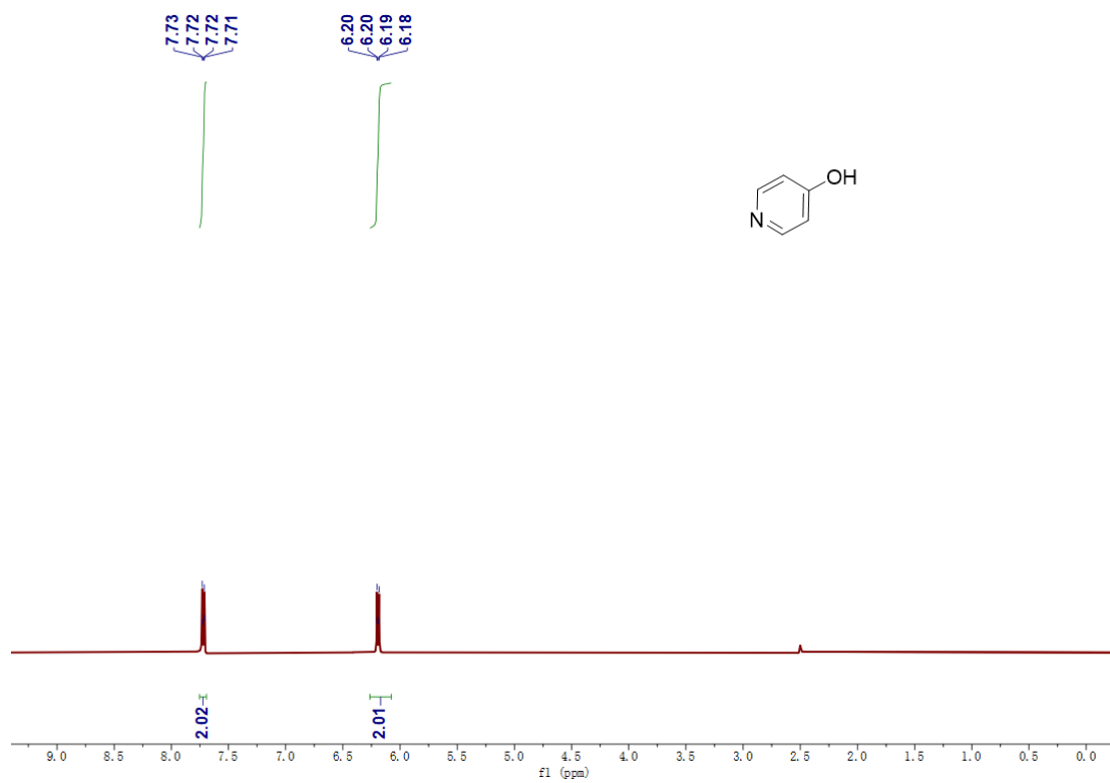


**Fig. S39** The absorption spectra of ABDA after irradiation (410-415 nm, 10 W) for different time in the presence of (a) Control: ABDA without any additive; (b) DNPY-SBE- $\beta$ -CD+RhB+SR101; (c) The UV-vis absorption spectra of DNPY-SBE- $\beta$ -CD+RhB+SR101 in the aqueous solution; (d) The decomposition rates of ABDA in the presence of DNPY-SBE- $\beta$ -CD+RhB+SR101.



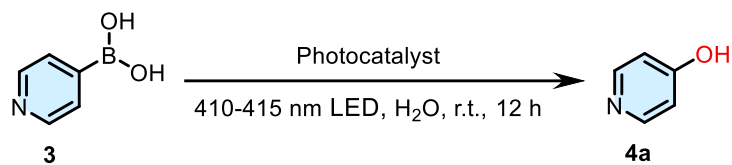
**Fig. S40** The absorption spectra of NBT after irradiation (410-415 nm, 10 W) for different time in the presence of (a) DNPY-SBE- $\beta$ -CD+RhB; (b) DNPY-SBE- $\beta$ -CD+SR101; (c) DNPY-SBE- $\beta$ -CD+RhB +SR101.





**Fig. S41**  $^1\text{H}$  NMR spectra of **4a** in DMSO- $d_6$ .

**Table S3** Oxidative hydroxylation of arylboronic acids with different base.<sup>a, b</sup>



| Entry | Variation from standard conditions <sup>a</sup>       | Yield <sup>b</sup> [%] |
|-------|---|------------------------|
| 1     | None  | 93                     |
| 2     | DIPEA (0.2 mmol) instead of DIPEA (0.4 mmol)          | 75                     |
| 3     | Triethylamine (0.4 mmol) instead of DIPEA (0.4 mmol)  | 88                     |
| 4     | Trimethylamine (0.4 mmol) instead of DIPEA (0.4 mmol) | 72                     |

<sup>a</sup>Reaction conditions: 4-pyridylboronic acid (0.1 mmol), DIPEA (0.4 mmol), DNPY-SBE- $\beta$ -CD+RhB+SR101 aqueous solution (0.5 mmol%, 3 mL), 410-415 nm LED, room temperature, 12 h; <sup>b</sup>Isolated yields.

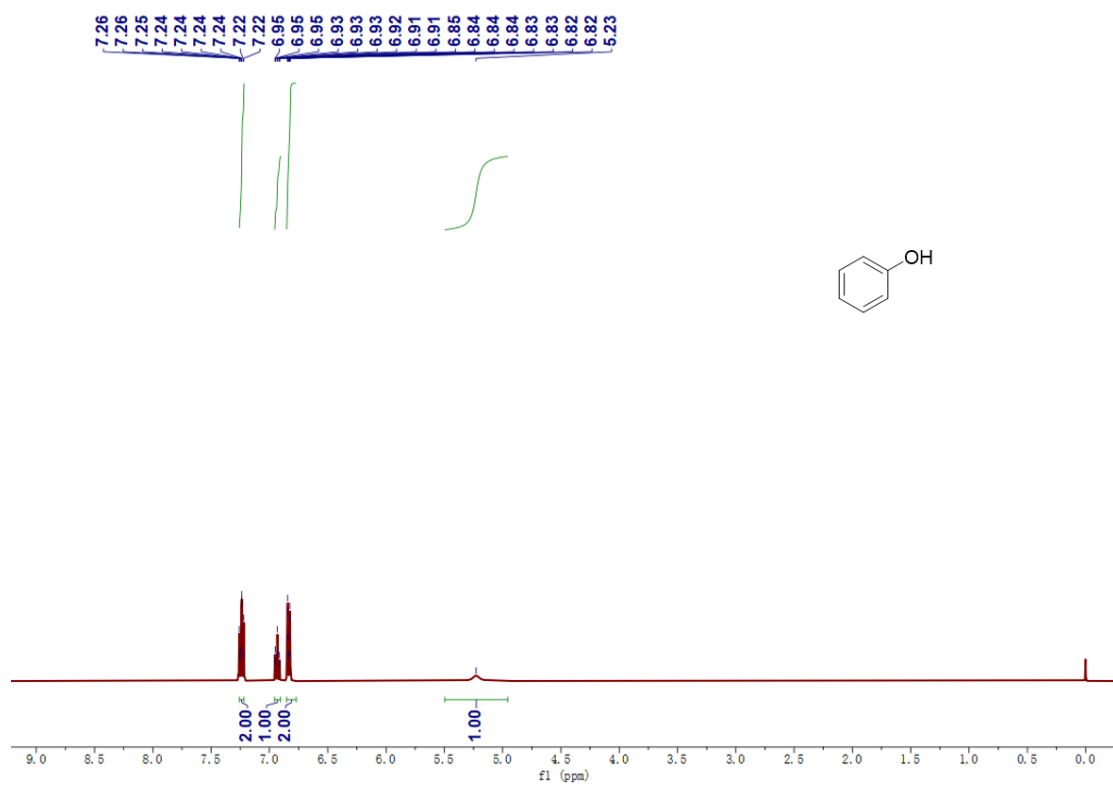
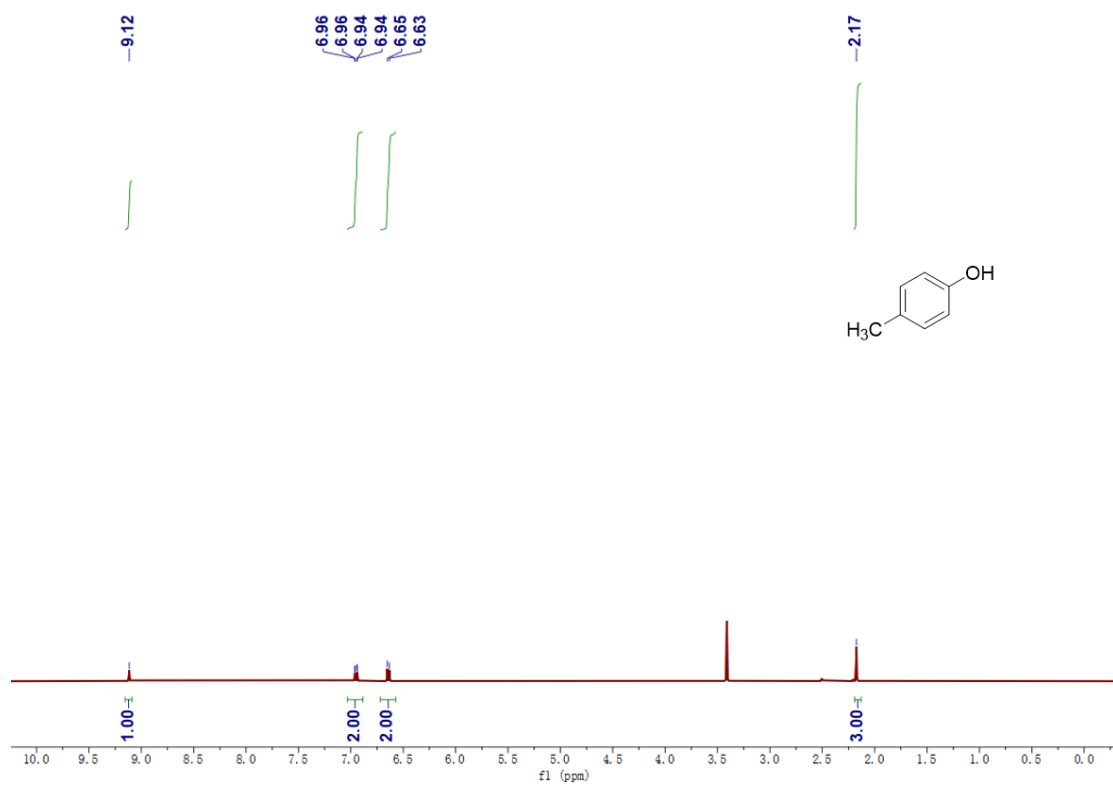


Fig. S42 <sup>1</sup>H NMR spectra of **4b** in CDCl<sub>3</sub>.



**Fig. S43**  $^1\text{H}$  NMR spectra of **4c** in  $\text{DMSO-}d_6$ .

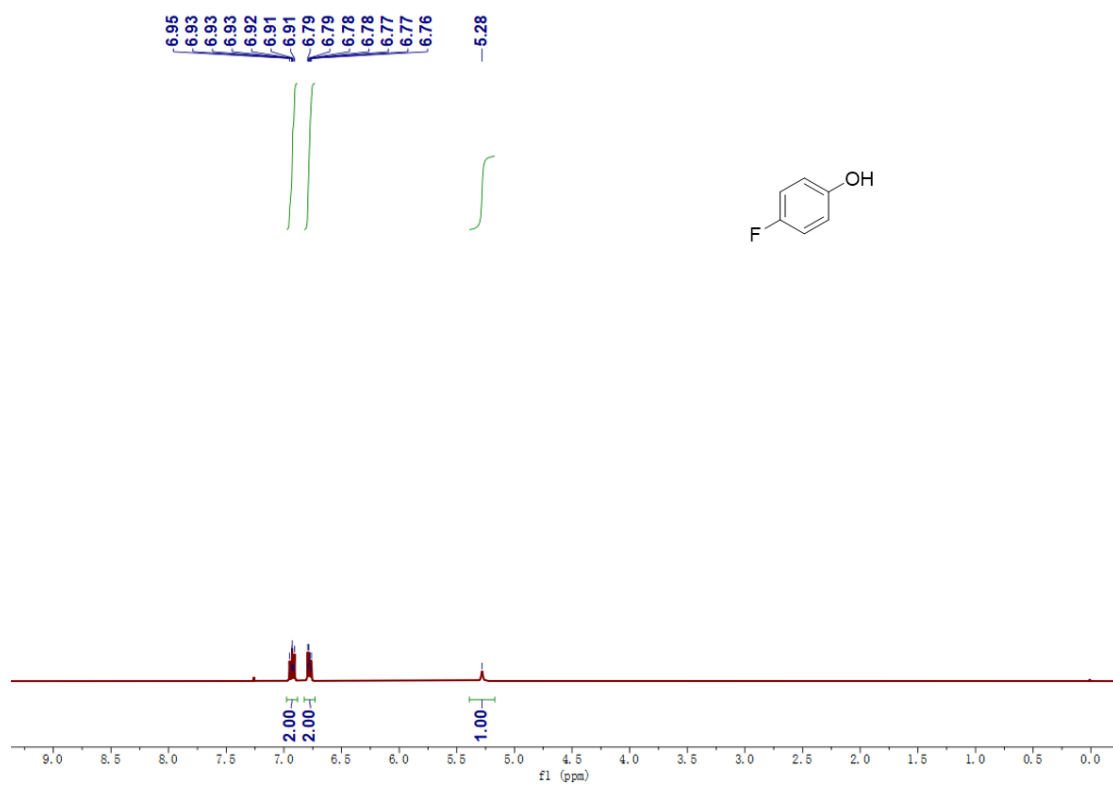
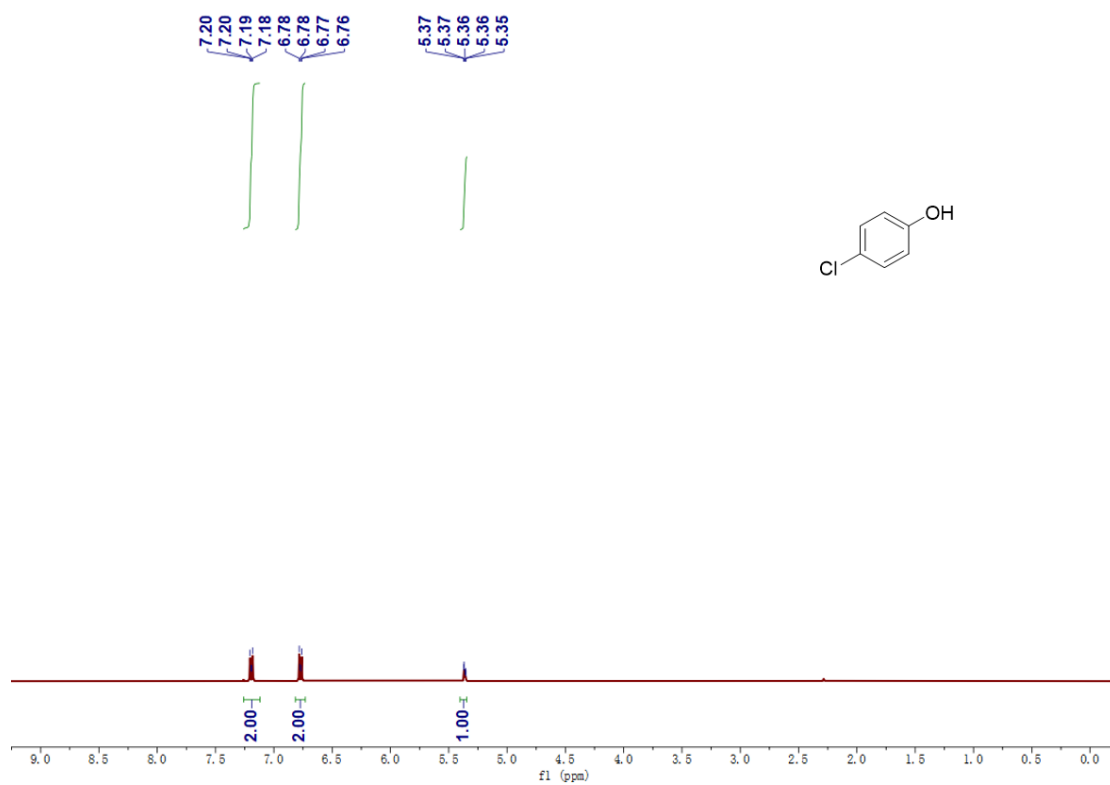
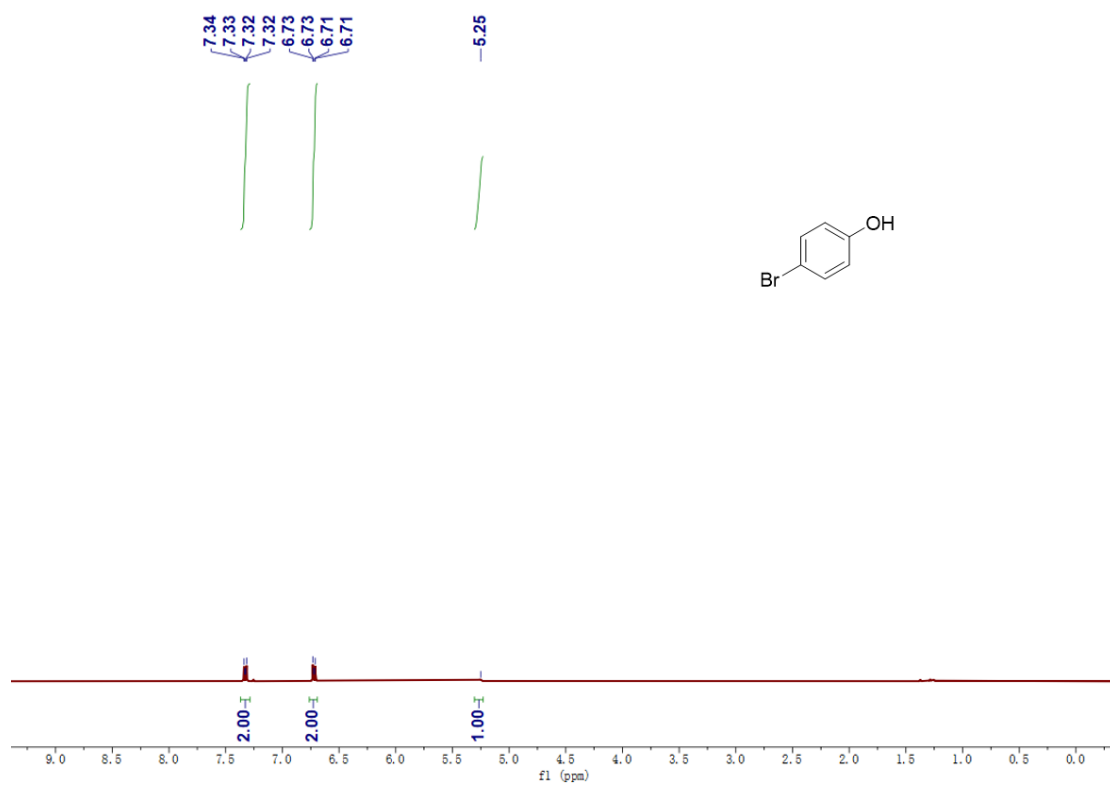


Fig. S44  $^1\text{H}$  NMR spectra of **4d** in  $\text{CDCl}_3$ .



**Fig. S45**  $^1\text{H}$  NMR spectra of **4e** in  $\text{CDCl}_3$ .



**Fig. S46**  $^1\text{H}$  NMR spectra of **4f** in  $\text{CDCl}_3$ .

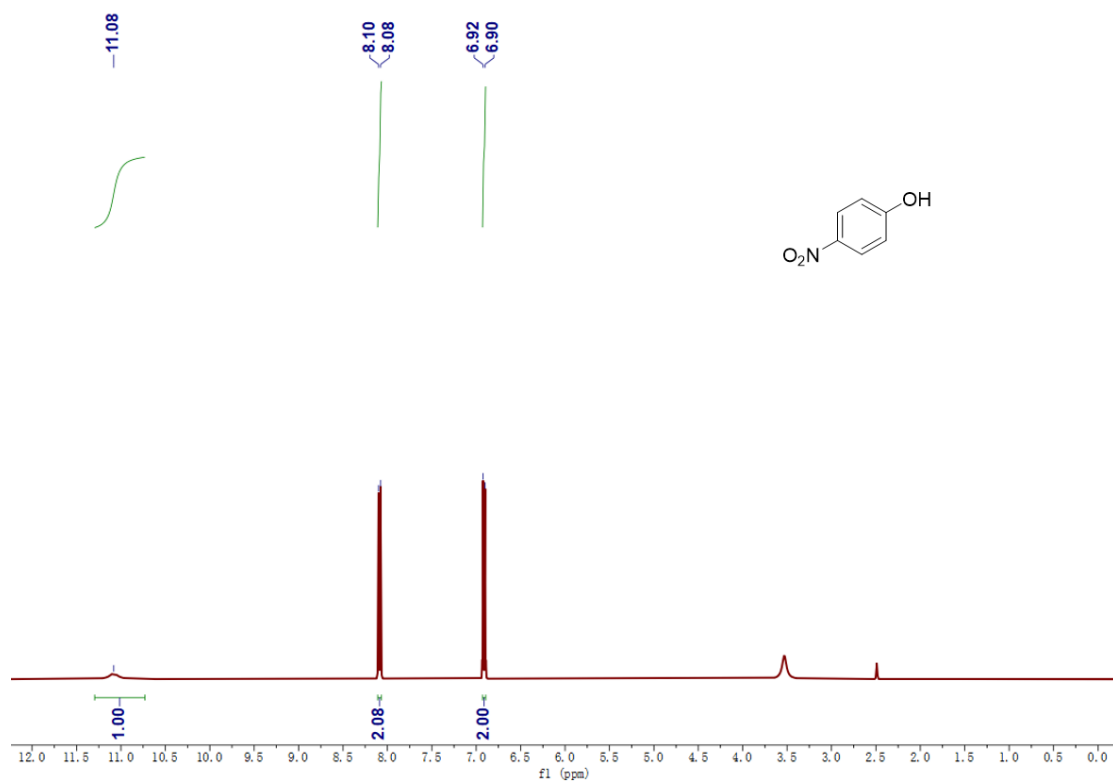


Fig. S47  $^1\text{H}$  NMR spectra of **4g** in  $\text{DMSO-}d_6$ .



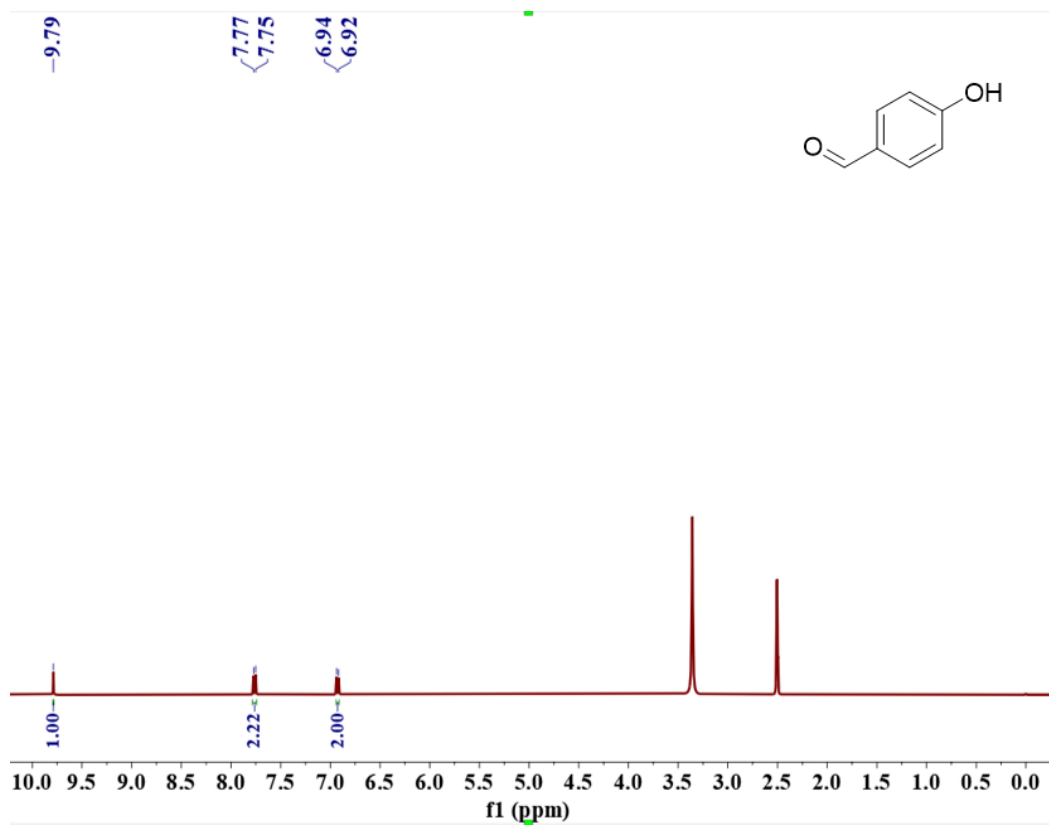
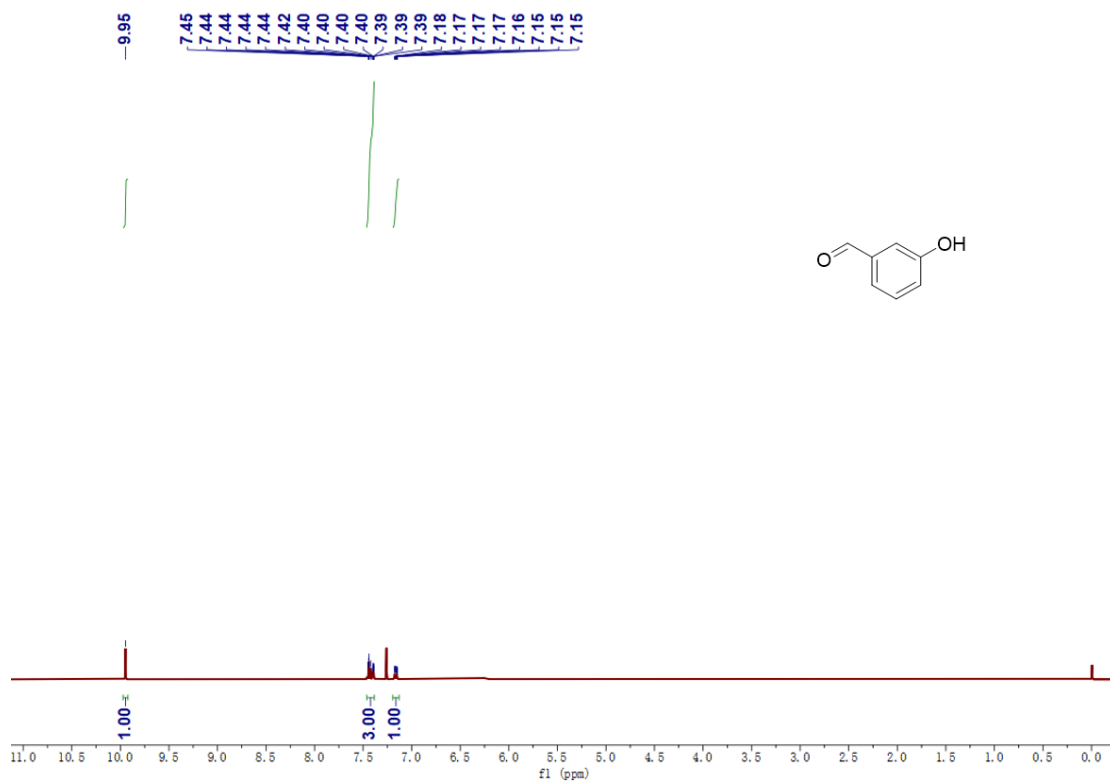


Fig. S48  $^1\text{H}$  NMR spectra of **4h** in  $\text{DMSO-}d_6$ .



**Fig. S49**  $^1\text{H}$  NMR spectra of **4i** in  $\text{CDCl}_3$ .

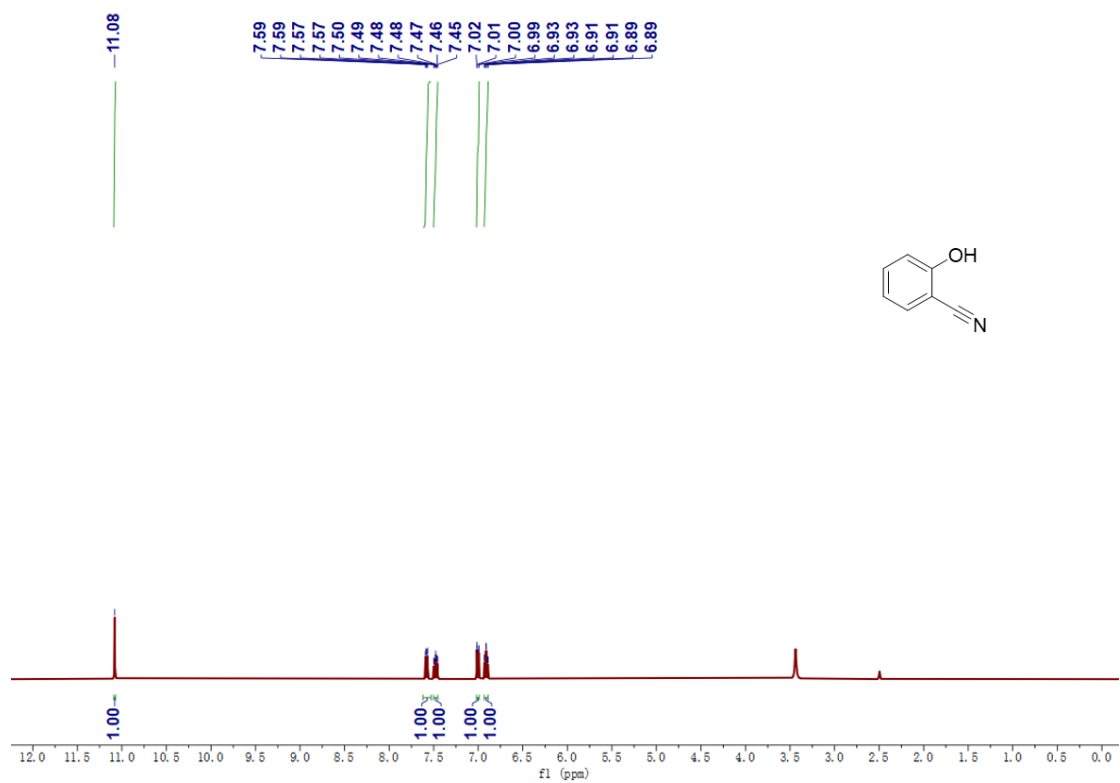
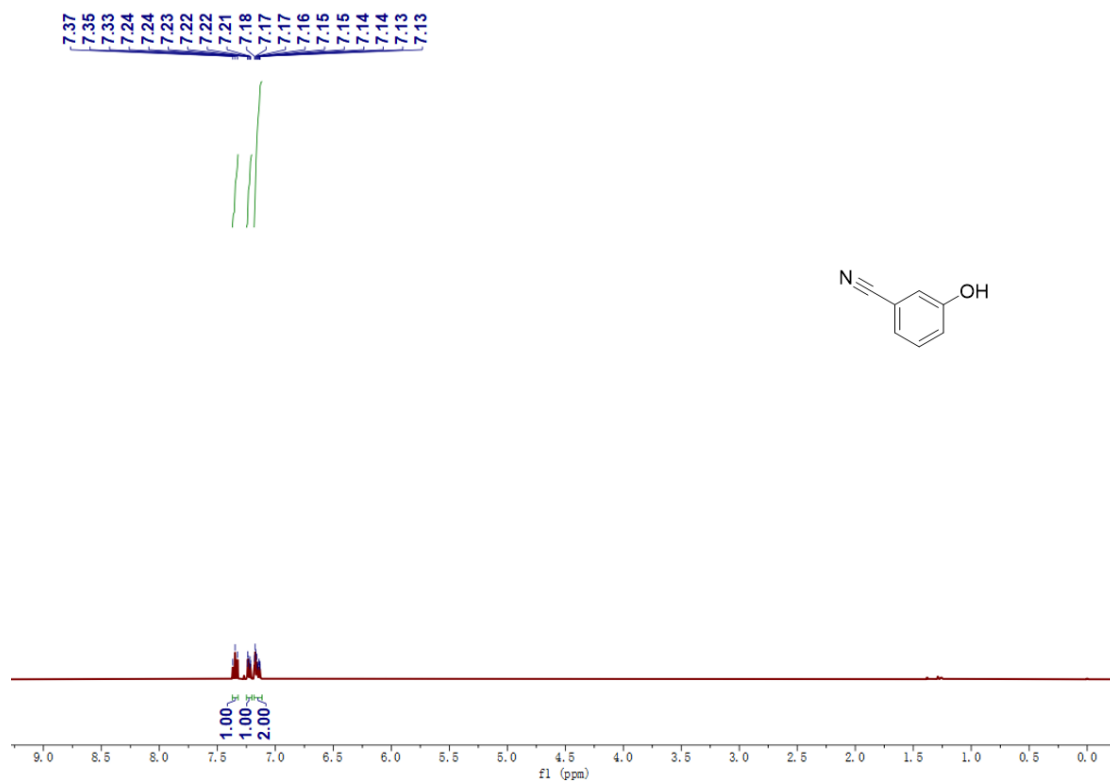
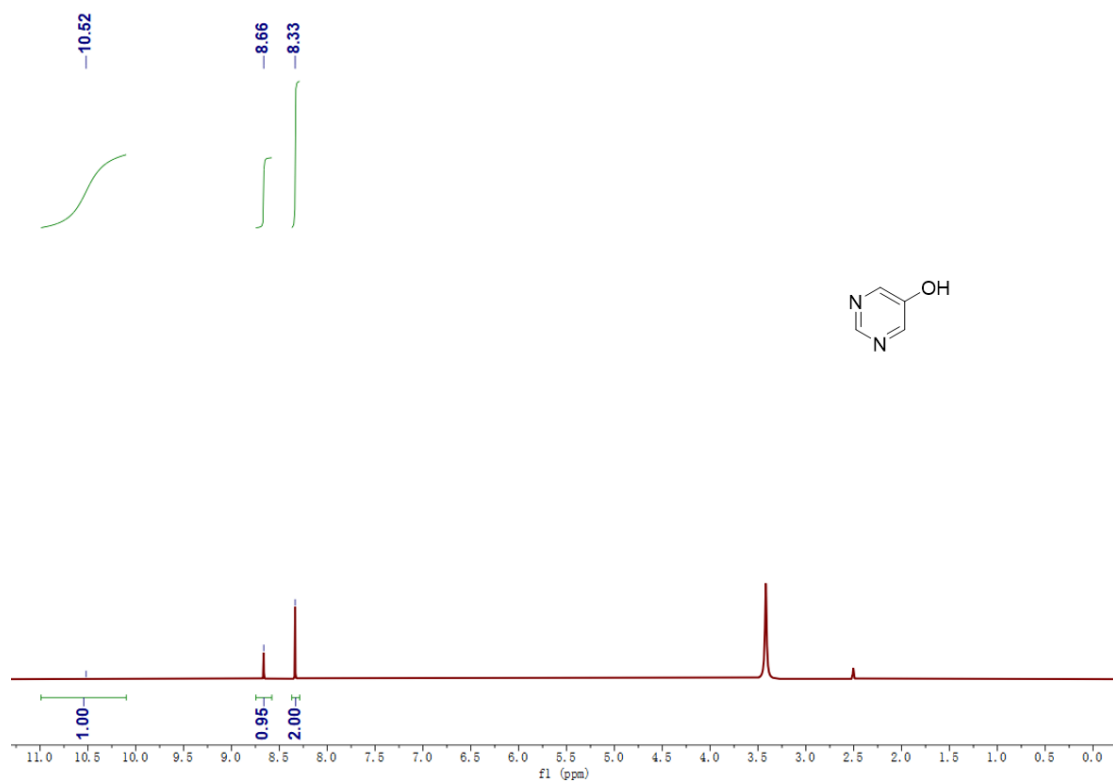


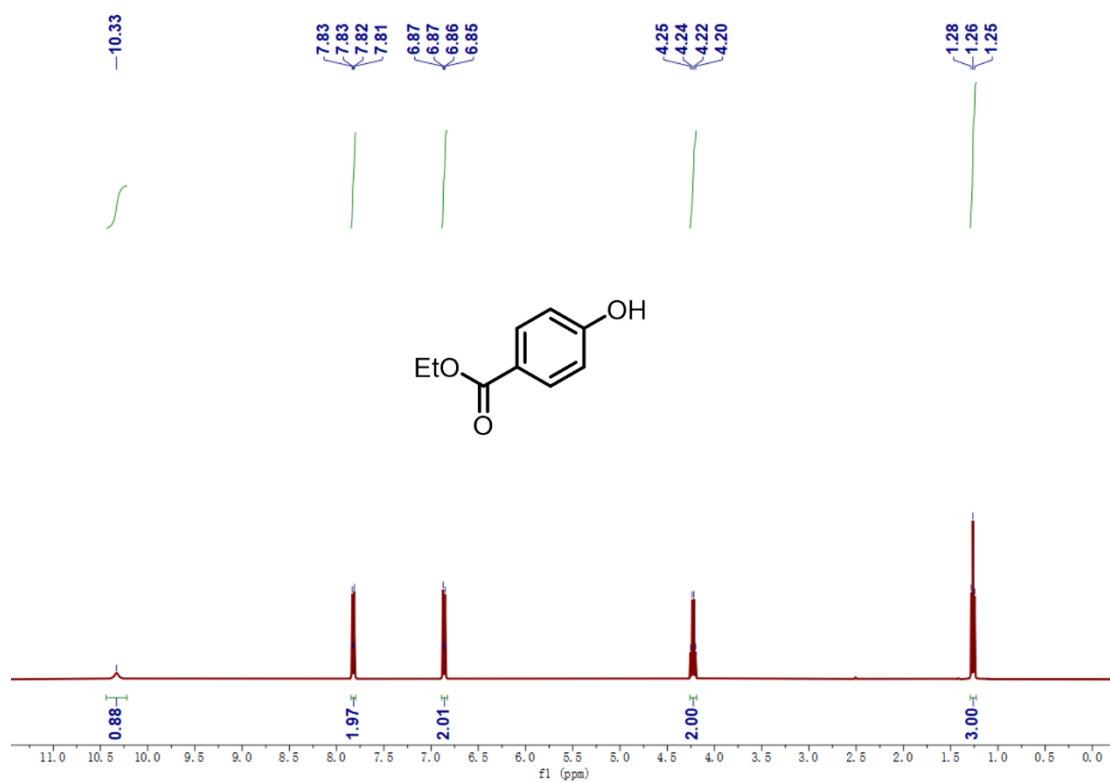
Fig. S50  $^1\text{H}$  NMR spectra of **4j** in  $\text{DMSO-}d_6$ .



**Fig. S51**  $^1\text{H}$  NMR spectra of **4k** in  $\text{CDCl}_3$ .



**Fig. S52**  $^1\text{H}$  NMR spectra of **4l** in  $\text{DMSO-}d_6$ .



**Fig. S53** <sup>1</sup>H NMR spectra of **4m** in DMSO-*d*<sub>6</sub>.

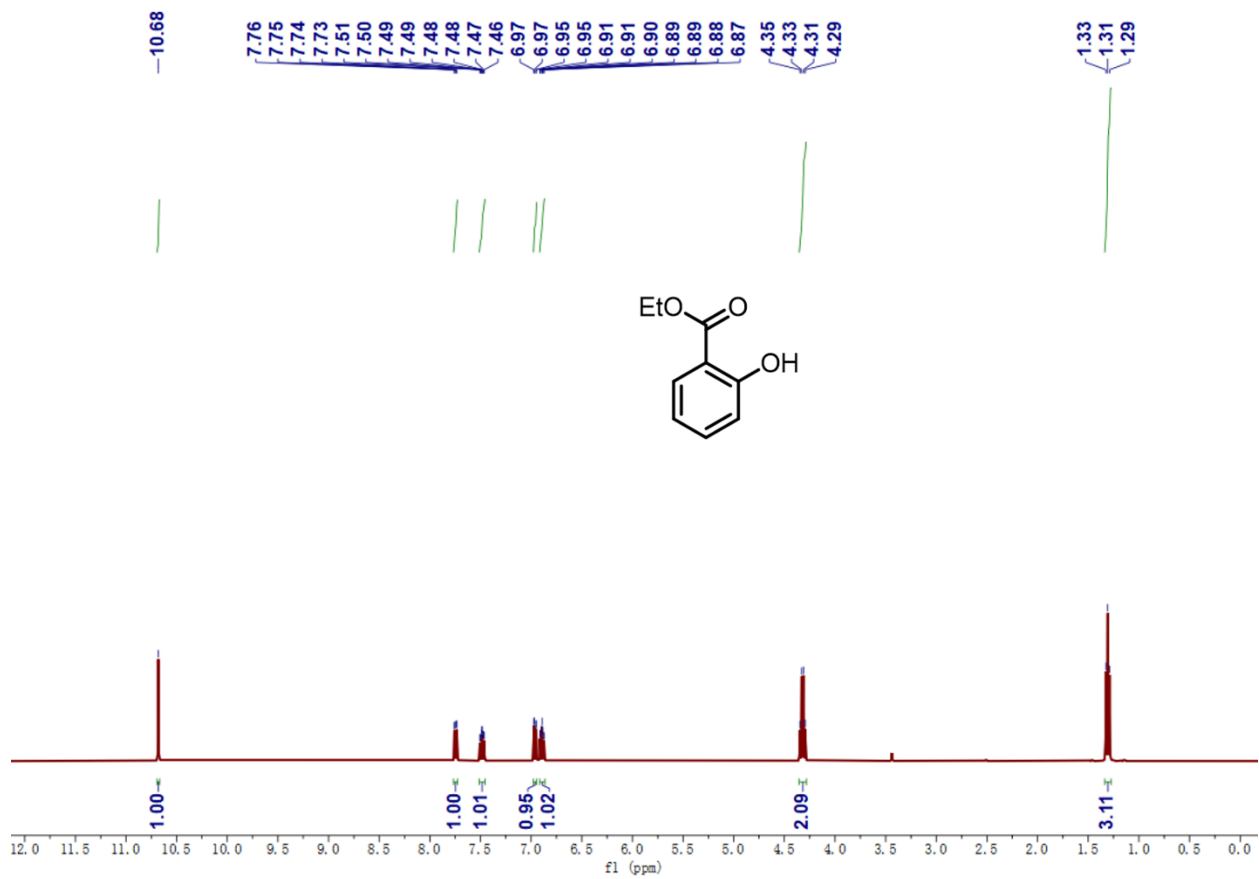


Fig. S54 <sup>1</sup>H NMR spectra of **4n** in DMSO-*d*<sub>6</sub>.

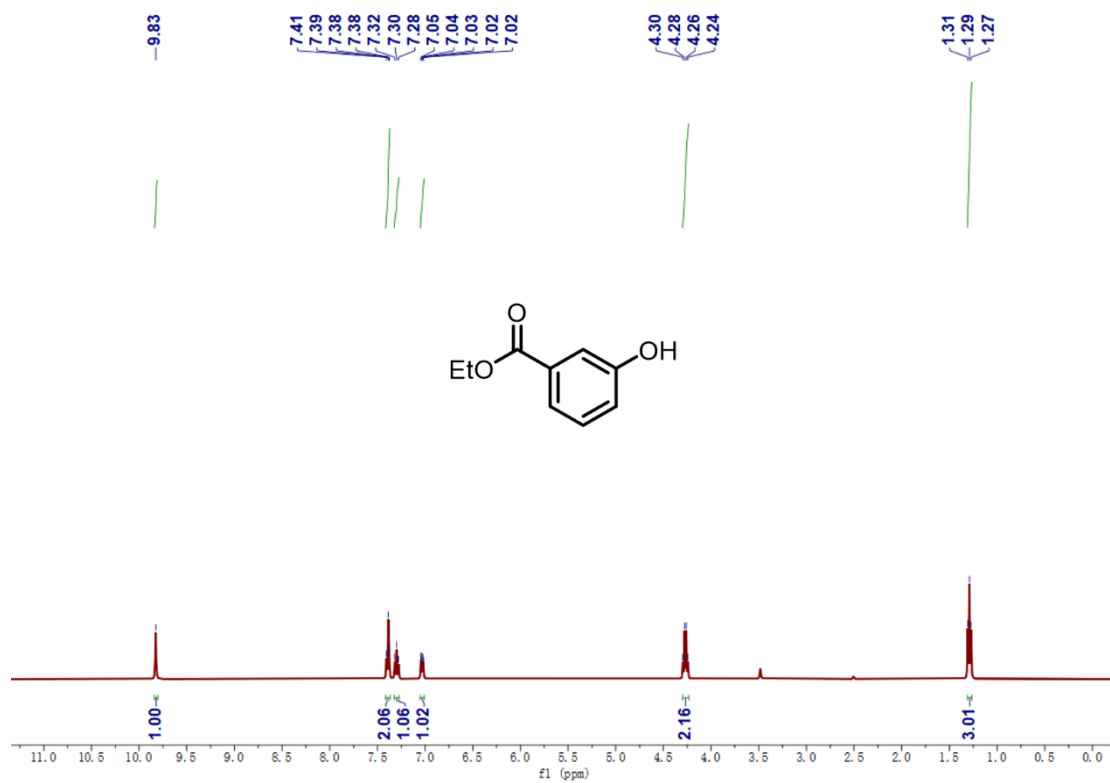
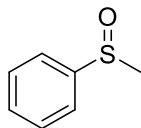


Fig. S55 <sup>1</sup>H NMR spectra of **4o** in DMSO-*d*<sub>6</sub>.



## <sup>1</sup>H NMR data of 2a-2l

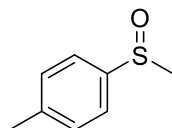
### 2a. (Methylsulfinyl)benzene



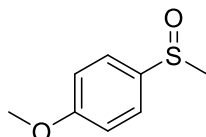
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.66 (dd, *J* = 8.0, 1.6 Hz, 2H), 7.53 (d, *J* = 7.5 Hz, 3H), 2.74 (s, 3H).

### 2b. 1-Methyl-4-(methylsulfinyl)benzene

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.37 (d, *J* = 8.8 Hz, 2H), 7.82 (d, *J* = 8.8 Hz, 2H), 2.78 (s, 3H).

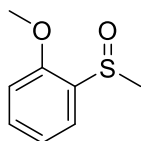


### 2c. 1-Methoxy-4-(methylsulfinyl)benzene



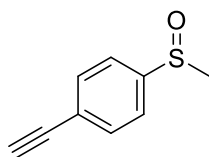
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.58 (d, *J* = 8.8 Hz, 2H), 7.02 (d, *J* = 8.8 Hz, 2H), 3.84 (s, 3H), 2.69 (s, 3H).

### 2d. 1-Methoxy-2-(methylsulphinyl)benzene



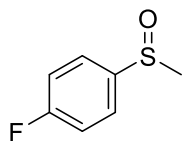
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.79 (dd, *J* = 7.7, 1.7 Hz, 1H), 7.43 (ddd, *J* = 8.2, 7.4, 1.7 Hz, 1H), 7.17 (td, *J* = 7.6, 1.0 Hz, 1H), 6.90 (dd, *J* = 8.2, 0.9 Hz, 1H), 3.87 (s, 3H), 2.75 (s, 3H).

### 2e. 1-Ethynyl-4-(methylsulfinyl)benzene



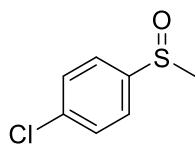
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.83-7.76 (m, 2H), 7.75-7.69 (m, 2H), 2.72 (s, 3H).

**2f. 1-Fluoro-4-(methylsulfinyl)benzene**



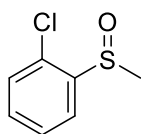
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.68-7.56 (m, 2H), 7.23-7.14 (m, 2H), 2.68 (s, 3H).

**2g. 1-Chloro-4-(methylsulfinyl)benzene**



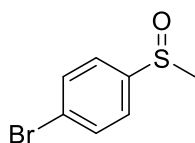
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.55-7.49 (m, 2H), 7.46-7.40 (m, 2H), 2.65 (s, 3H).

**2h. 1-Chloro-2-(methylsulphiny)benzene**



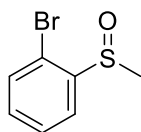
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.87 (dd,  $J = 7.8, 1.7$  Hz, 1H), 7.46 (td,  $J = 7.5, 1.3$  Hz, 1H), 7.41-7.28 (m, 2H), 2.75 (s, 3H).

**2i. 1-Bromo-4-(methylsulfinyl)benzene**



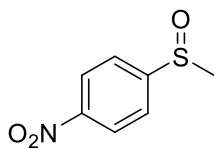
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.61-7.54 (m, 2H), 7.48-7.40 (m, 2H), 2.63 (s, 3H).

**2j. 1-Bromo-2-(methylsulfinyl)benzene**



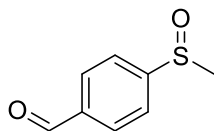
$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.90 (dd,  $J = 7.8, 1.6$  Hz, 1H), 7.57-7.50 (m, 2H), 7.34 (ddd,  $J = 8.0, 7.3, 1.7$  Hz, 1H), 2.78 (s, 3H).

**2k. 1-Methanesulfinyl-4-nitrobenzene**



$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.52 (d,  $J = 8.2$  Hz, 2H), 7.34-7.29 (m, 2H), 2.69 (s, 3H), 2.40 (s, 3H).

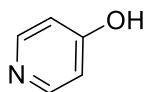
**2l. 4-(Methylsulfinyl)benzaldehyde**



$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.37 (d,  $J = 8.8$  Hz, 2H), 7.82 (d,  $J = 8.8$  Hz, 2H), 2.78 (s, 3H).

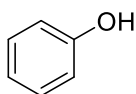
## <sup>1</sup>H NMR data of 4a-4o

### 4a. 4-Hydroxypyridine



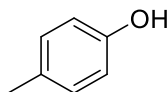
<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.27-7.21 (m, 2H), 6.93 (tt, *J* = 7.4, 1.1 Hz, 1H), 6.89-6.77 (m, 2H), 5.23 (s, 1H).

### 4b. Phenylboronic acid



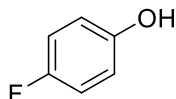
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.27-7.22 (m, 2H), 6.95-6.90 (m, 1H), 6.85-6.81 (m, 2H), 5.23 (s, 1H).

### 4c. 4-Methylphenol



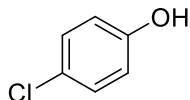
<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 9.12 (s, 1H), 7.03-6.87 (m, 2H), 6.64 (d, *J* = 8.4 Hz, 2H), 2.17 (s, 3H).

### 4d. 4-Fluorophenol



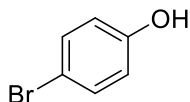
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 6.98-6.88 (m, 2H), 6.82-6.73 (m, 2H), 5.28 (s, 1H).

### 4e. 4-Chlorophenol



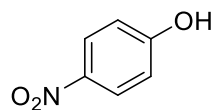
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.55-7.49 (m, 2H), 7.46-7.40 (m, 2H), 2.65 (s, 3H).

### 4f. 4-Bromophenol



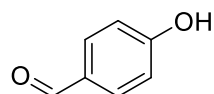
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.37-7.28 (m, 2H), 6.76-6.66 (m, 2H), 5.25 (s, 1H).

**4g. 4-Nitrophenol**



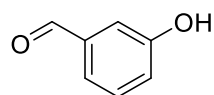
$^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  11.08 (s, 1H), 8.09 (d,  $J = 9.2$  Hz, 2H), 6.91 (d,  $J = 9.2$  Hz, 2H).

**4h. 4-Hydroxybenzaldehyde**



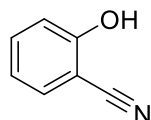
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.79 (s, 1H), 7.76 (d,  $J = 8.6$  Hz, 2H), 6.93 (d,  $J = 8.6$  Hz, 2H).

**4i. 3-Hydroxybenzaldehyde**



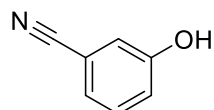
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.95 (s, 1H), 7.47-7.39 (m, 3H), 7.16 (ddd,  $J = 7.1, 2.6, 1.9$  Hz, 1H).

**4j. 2-Hydroxybenzonitrile**



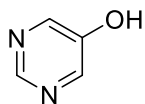
$^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  11.08 (s, 1H), 7.58 (dd,  $J = 7.8, 1.7$  Hz, 1H), 7.48 (ddd,  $J = 8.9, 7.4, 1.7$  Hz, 1H), 7.00 (dd,  $J = 8.5, 1.0$  Hz, 1H), 6.91 (td,  $J = 7.5, 1.0$  Hz, 1H).

**4k. 3-Hydroxybenzonitrile**



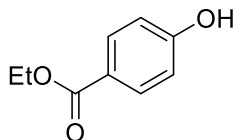
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.35 (t,  $J = 7.9$  Hz, 1H), 7.23 (dt,  $J = 7.7, 1.3$  Hz, 1H), 7.19-7.12 (m, 2H).

**4l. 5-Hydroxypyrimidine**



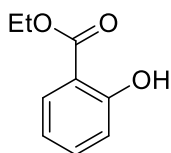
$^1\text{H NMR}$  (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  10.52 (s, 1H), 8.66 (s, 1H), 8.33 (s, 2H).

#### 4m. Ethyl 4-hydroxybenzoate



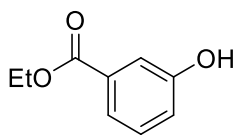
$^1\text{H NMR}$  (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  10.33 (s, 1H), 7.85-7.80 (m, 2H), 6.89-6.83 (m, 2H), 4.23 (q,  $J = 7.1$  Hz, 2H), 1.26 (t,  $J = 7.1$  Hz, 3H).

#### 4n. Ethyl 2-hydroxybenzoate



$^1\text{H NMR}$  (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  10.68 (s, 1H), 7.75 (dd,  $J = 8.0, 1.8$  Hz, 1H), 7.48 (ddd,  $J = 8.4, 7.2, 1.8$  Hz, 1H), 6.96 (dd,  $J = 8.4, 1.2$  Hz, 1H), 6.89 (ddd,  $J = 8.2, 7.2, 1.2$  Hz, 1H), 4.32 (q,  $J = 7.1$  Hz, 2H), 1.31 (t,  $J = 7.2$  Hz, 3H).

#### 4o. Ethyl 3-hydroxybenzoate



$^1\text{H NMR}$  (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.83 (s, 1H), 7.41-7.37 (m, 2H), 7.30 (t,  $J = 7.8$  Hz, 1H), 7.03 (dd,  $J = 9.0, 2.5$  Hz, 1H), 4.27 (q,  $J = 7.1$  Hz, 2H), 1.29 (t,  $J = 7.1$  Hz, 3H).

## References:

- S1. V.-N. Nguyen, S. Qi, S. Kim, N. Kwon, G. Kim, Y. Yim, S. Park, and J. Yoon, *J. Am. Chem. Soc.*, 2019, **141**, 16243–16248.
- S2. G. Linden, L. Zhang, F. Pieck, U. Linne, D. Kosenkov, R. Tonner, and O. Vázquez, *Angew. Chem. Int. Ed.*, 2019, **58**, 12868–12873.
- S3. M. K. Kuimova, H. A. Collins, M. Balaz, E. Dahlstedt, J. A. Levitt, N. Sergent, K. Suhling, M. Drobizhev, N. S. Makarov, A. Rebane, H. L. Anderson and D. Phillips, *Org. Biomol. Chem.*, 2009, **7**, 889–896.
- S4. Y. Hao, B. M. Liu, T. F. Bennett, C. G. Monsour, M. Selke, Y. Liu, *J. Phys. Chem. C*, 2021, **125**, 7392–7400.
- S5. X. Kan, J.-C. Wang, Z. Chen, J.-Q. Du, J.-L. Kan, W.-Y. Li, and Y.-B. Dong, *J. Am. Chem. Soc.*, 2022, **144**, 6681–6686.
- S6. R. Zhang, G. Feng, C.-J. Zhang, X. Cai, X. Cheng, and B. Liu, *Anal. Chem.*, 2016, **88**, 4841–4848.
- S7. Y. Gao, X. Wang, X. He, Z. He, X. Yang, S. Tian, F. Meng, D. Ding, L. Luo, and B. Z. Tang, *Adv. Funct. Mater.*, 2019, **29**, 1902673
- S8. X. Dong, X. Dai, G. Li, Y.-M. Zhang, X. Xu, and Y. Liu, *Adv. Sci.*, 2022, 2201962.
- S9. M. P. Donzello, E. Viola, M. Giustini, C. Ercolani and F. Monacelli, *Dalton Trans.*, 2012, **41**, 6112-6121.
- S10. E. A. Dupouy, D. Lazzeri and E. N. Durantini, *Photochem. Photobiol. Sci.*, 2004, **3**, 992-998.
- S11. G. Schnurpfeil, A. K. Sobbi, W. Spiller, H. Kleisch and D. Wöhrle, *J. Porphyrins Phthalocyanines*, 1997, 01, 159-167.
- S12. E. H. Mørkved, T. Andreassen, V. Novakova and P. Zimcik, *Dyes Pigm.*, 2009, **82**, 276-285.
- S13. U. Michelsen, H. Kliesch, G. Schnurpfeil, A. K. Sobbi and D. Wöhrle, *Photochem. Photobiol.*, 1996, **64**, 694-701.
- S14. T. C. Tempesti, M. G. Alvarez, E. N. Durantini, *Dyes Pigm.*, 2011, **91**, 6-12.

- S15. M. Machacek, J. Kollár, M. Miletin, R. Kučera, P. Kubát, T. Simunek, V. Novakova and P. Zimcik, *RSC Adv.*, 2016, **6**, 10064-10077.
- S16. P. Zimcik, M. Miletin, H. Radilova, V. Novakova, K. Kopecky, J. Svec, E. Rudolf, *Photochem. Photobiol.*, 2010, **86**, 168-175.
- S17. G. De Mori, Z. Fu, E. Viola, X. H. Cai, C. Ercolani, M. P. Donzello and K. M. Kadish, *Inorg. Chem.*, 2011, **50**, 8225-8237.
- S18. B. Ghazal, M. Machacek, M. A. Shalaby, V. Novakova, P. Zimcik and S. Makhseed, *J. Med. Chem.*, 2017, **60**, 6060-6076.
- S19. V. Novakova, R. Z. Uslu Kobak, R. Kucera, K. Kopecky, M. Miletin, V. Krepsova, J. Ivincova and P. Zimcik, *Dalton Trans.*, 2012, **41**, 10596-10604.
- S20. S. Makhseed, A. Tuhl, J. Samuel, P. Zimcik, N. Al-Awadi and V. Novakova, *Dyes Pigm.*, 2012, **95**, 351-357.
- S21. M. P. Donzello, D. Vittori, E. Viola, L. H. Zeng, Y. Cui, K. M. Kadish, L. Mannina and C. Ercolani, *J. Porphyrins Phthalocyanines*, 2015, **19**, 903-919.
- S22. C. J. Song, J. M. Park, W. Yao, C. Y. Jung and J. Y. Jaung, *J. Porphyrins Phthalocyanines*, 2015, **19**, 967-972.
- S23. T. C. Tempesti, J. C. Stockert and E. N. Durantini, *J. Phys. Chem. B*, 2008, **112**, 15701-15707.
- S24. E. A. Dupouy, D. Lazzeri and E. N. Durantini, *Photochem. Photobiol. Sci.*, 2004, **3**, 992-998.
- S25. M. B. Spesia, M. Rovera and E. N. Durantini, *Eur. J. Med. Chem.*, 2010, **45**, 2198-2205.
- S26. C. A. Suchetti and E. N. Durantini, *Dyes Pigm.*, 2007, **74**, 630-635.
- S27. M. Machacek, J. Demuth, P. Cermak, M. Vavreckova, L. Hruby, A. Jedlickova, P. Kubat, T. Simunek, V. Novakova and P. Zimcik, *J. Med. Chem.*, 2016, **59**, 9443-9456.
- S28. H. Huang, X. Han, X. Li, S. Wang, P. K. Chu, Y. Zhang, *ACS Appl. Mater. Interfaces*, 2015, **7**, 482-492.
- S29. L. Ye, J. Liua, Z. Jiang, T. Peng, L. Zan, *Appl. Catal. B Environ.*, 2013, **142-143**, 1-7.



S30. Y. Li, W. Zhang, J. Niu, Y. Chen, *ACS Nano*, 2012, **6**, 5164-5173.