

## **Protein Oxidation of Fucose Environments (POFE) Reveals Fucose-protein Interactions**

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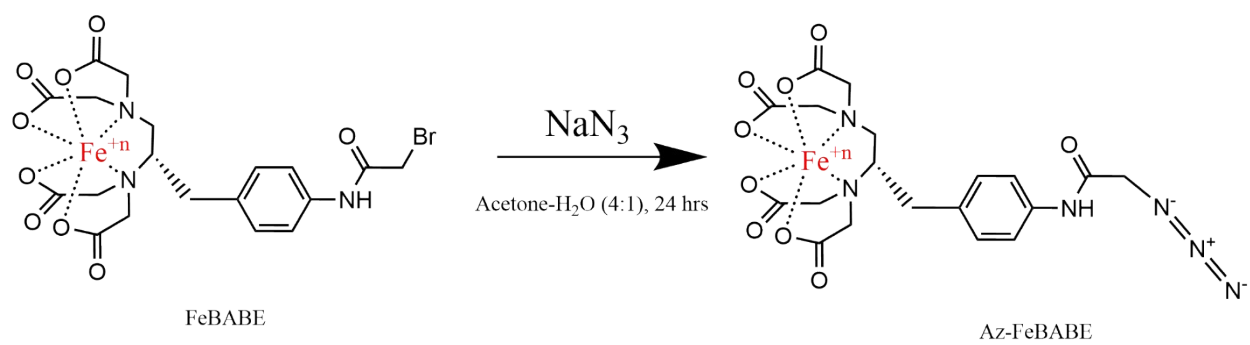
**Figure S19.** Modeling of glycan structures with or without the addition fucose.

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## Methods

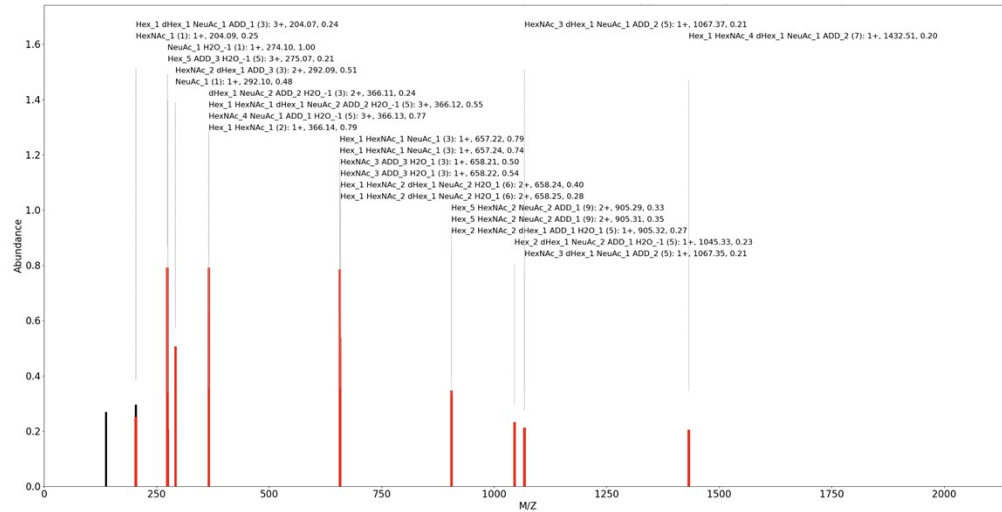
**Cell culture.** Human immortalized prostate epithelium PNT2 and human colon adenocarcinoma Caco-2 cells were obtained from American Type Culture Collection (ATCC, Manassas, VA). The PNT2 cells were cultured in Roswell Park Memorial Institute (RPMI) 1640 medium, and Caco-2 cells were grown in Eagle's Minimum Essential Medium (EMEM). The medium were supplemented with 10% (v/v) fetal bovine serum (FBS), 0.1% (v/v) penicillin and streptomycin, and 1% (v/v) GlutaMAX. Cells were maintained in a humidified incubator at 37°C with 5% CO<sub>2</sub> and subcultured at 80% confluency. For the fucose reporter treatment, cells were treated with 100 μM of 6AlkFuc, 6AzFuc, or 7AlkFuc for 72 hours.

**Confocal image analysis of fucose-labeled glycoproteins.** The cells were seeded onto FluoroDish cell culture dishes (WPI, FL) coated with Poly-D-Lysine with appropriate density. At 40% confluency, cells were treated with 100 μM of fucose reporters supplemented growing media for 72 hours. Afterward, the treated cells were rinsed with phosphate-buffered saline (PBS) and treated with 50 μM of coumarin azide (for labeling alkynyl groups) or DBCO-Cy3 (for labeling azido groups) in PBS at room temperature for 1 hour, followed by the fixation with 4% paraformaldehyde at room temperature for 1 hour. The plasma membrane and the nucleus were stained with CellMask Deep Red plasma membrane stain and Hoechst 33342, respectively, at 37°C for 10 minutes. Fluorescent images were captured by Leica TCS SP8 STED 3X Super-Resolution Confocal Microscope (Wetzlar, Germany).

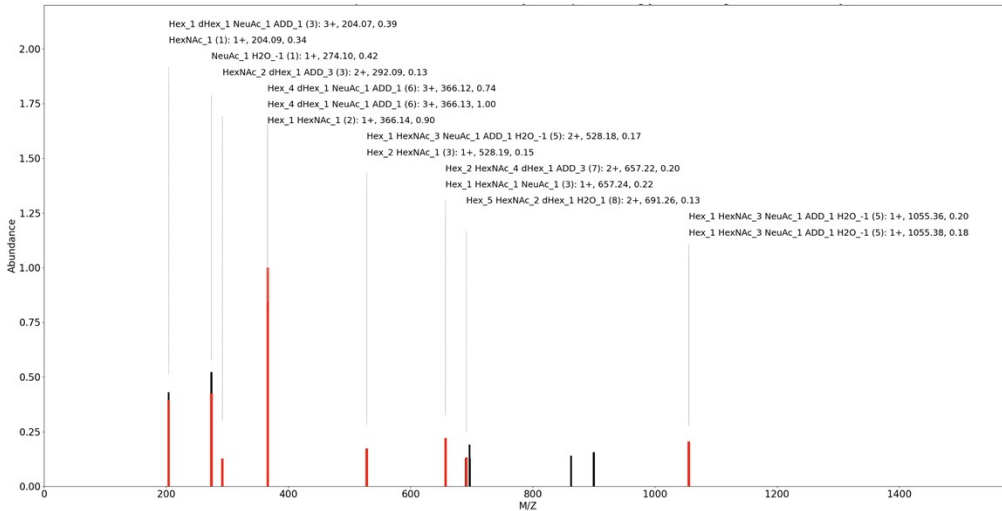


**Schematic 1.** Synthesis of the Az-FeBABA probe.

(a)

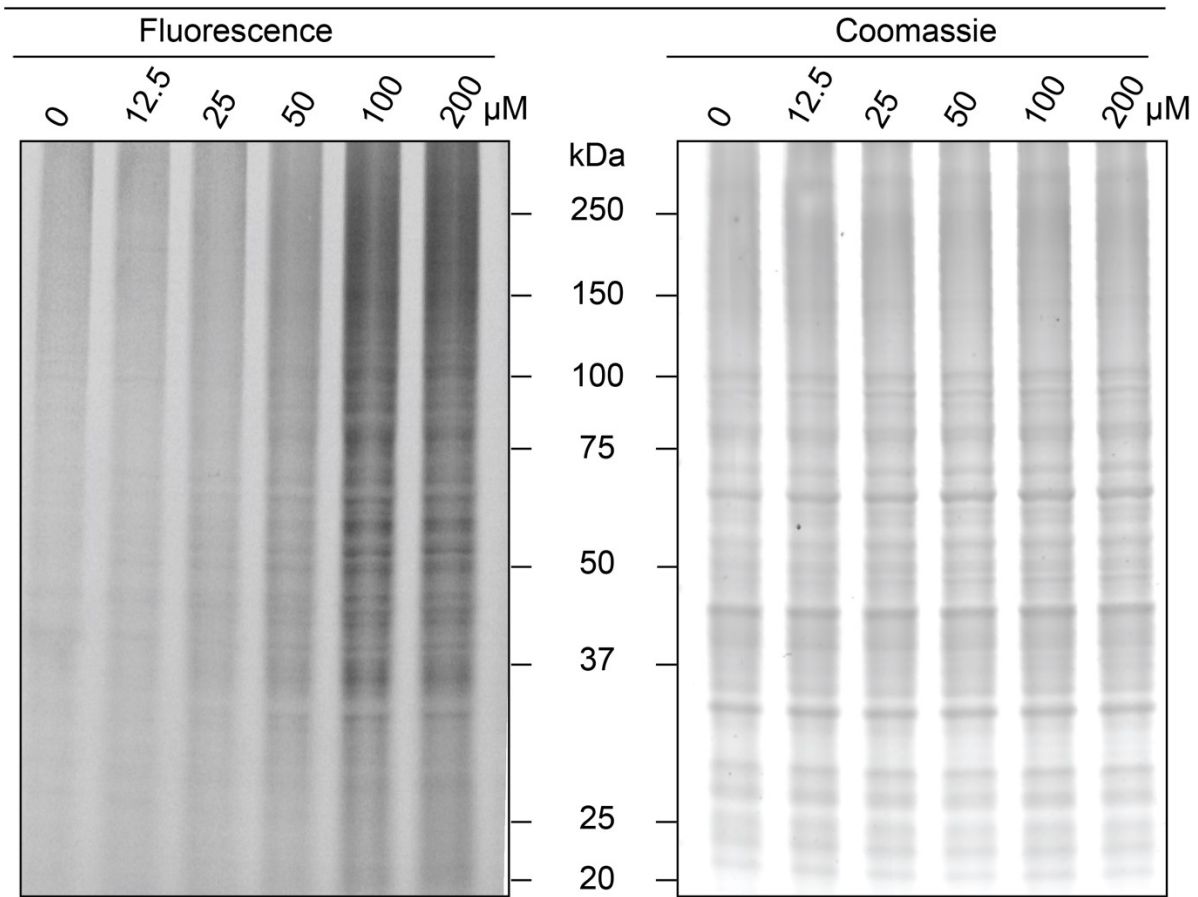


(b)

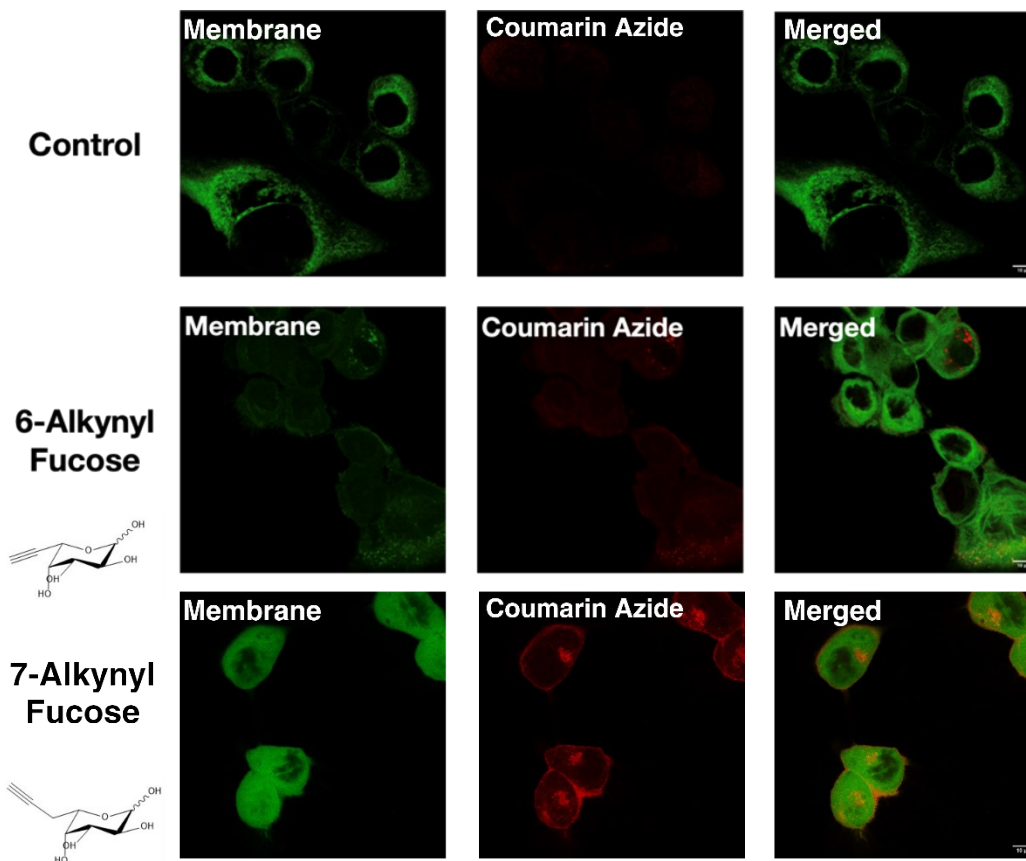


**Figure S1. Tandem mass spectra of selected compounds (a) Hex<sub>(5)</sub>HexNAC<sub>(4)</sub>7AlkFuc<sub>(1)</sub>Sia<sub>(2)</sub> and (b) Hex<sub>(5)</sub>HexNAC<sub>(4)</sub>7AlkFuc<sub>(1)</sub>Sia<sub>(1)</sub>. The glycan structures and fragments were annotated using GlycoNote.**

### 7-Alkynl Fucose

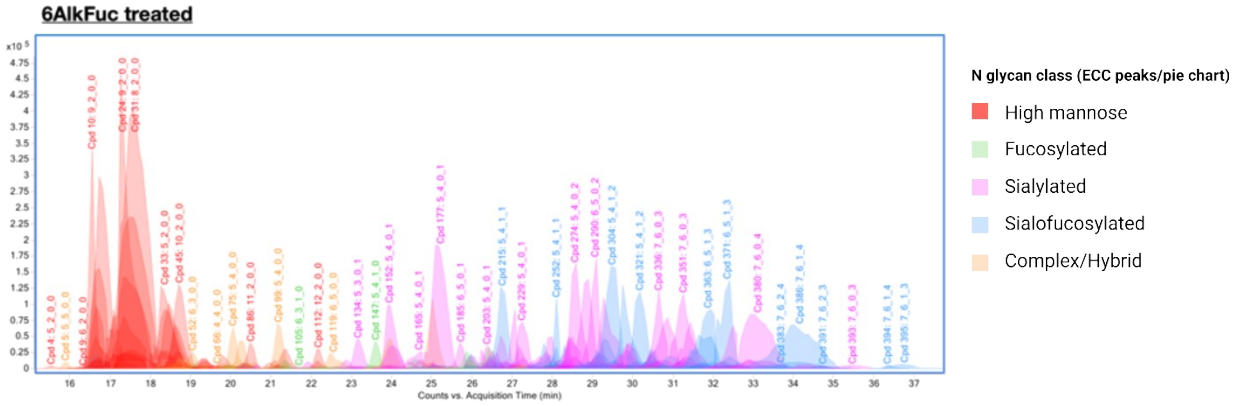


**Figure S2. Dose-dependent gel-based profiles of PNT2 cells treated by 7AlkFuc.** The cells were incubated for varying amounts of probe (0, 12.5, 25, 50, 100, and 200  $\mu\text{M}$ ). Corresponding expression profiles are shown after Coomassie staining. Concentrations and molecular weight markers are indicated.

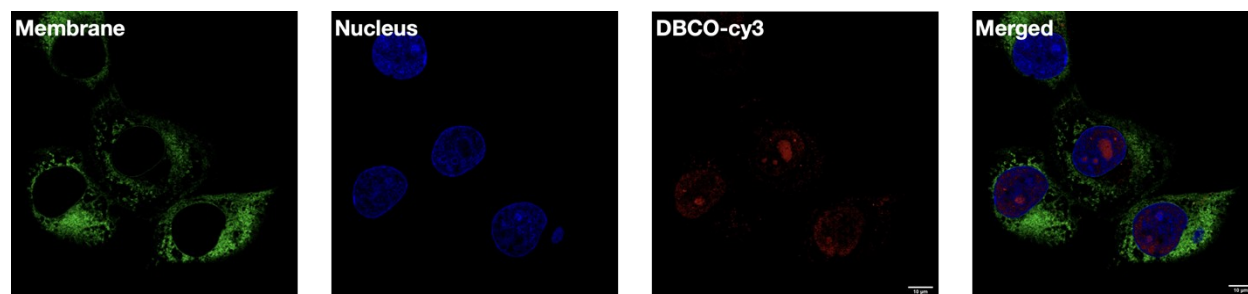


**Figure S3. Validation of the labeling of the fucose probes using confocal microscopy.** PNT2 Cells were treated with DMSO (**top**), 6AlkFuc (**middle**), and 7AlkFuc (**bottom**). The cell membrane was stained with CellMask™ Deep Red Plasma membrane Stain.





**Figure S4. LC-MS profile of N-Glycans released from 6AlkFuc-treated PNT2 cells.** Annotated structures are putative based on mass and composition. LC-MS peaks were color coded to assign glycan subtypes.



**Figure S5. Validation of the labeling of the 6AzFuc probe using confocal microscopy.** The incorporated azido group was labeled using DBCO-cy3. And the cell membrane and nucleus were stained with CellMask™ Deep Red Plasma membrane Stain and Hoechst 33342, respectively.

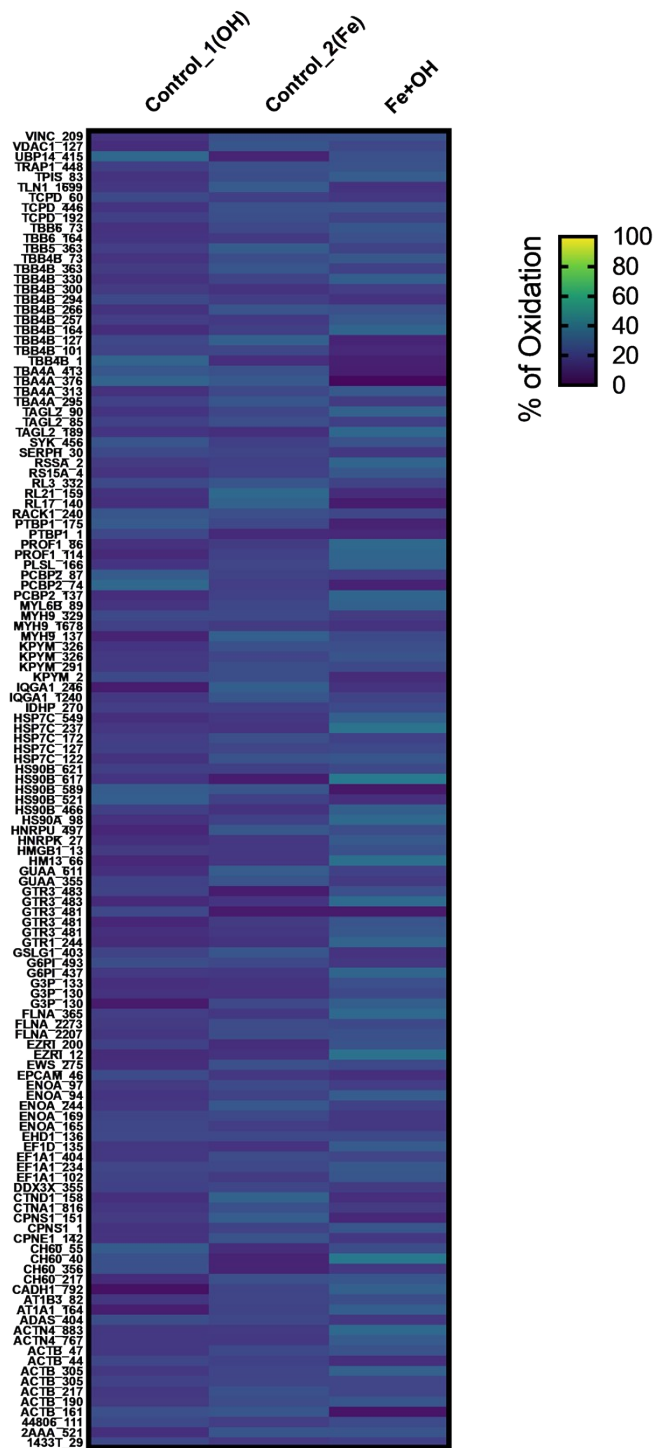
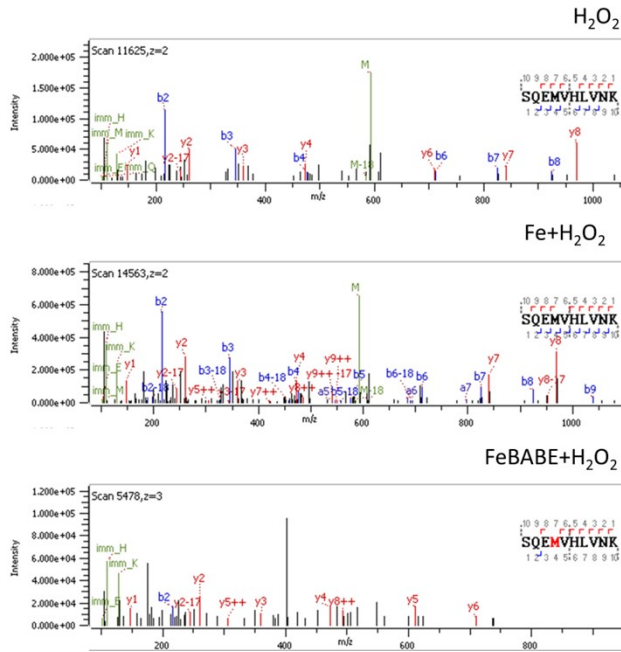
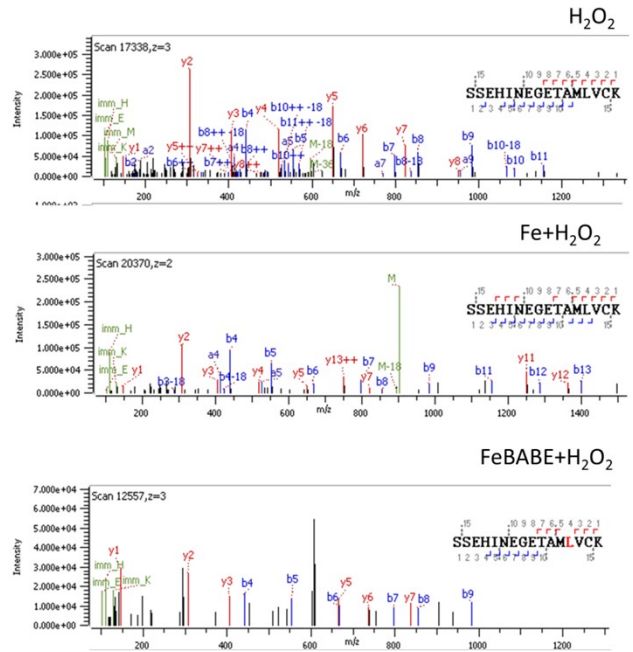


Figure S6. The extent of background oxidation quantified by Biologic. All experiments (including controls) were conducted in copper-environments to account for background oxidation by copper.

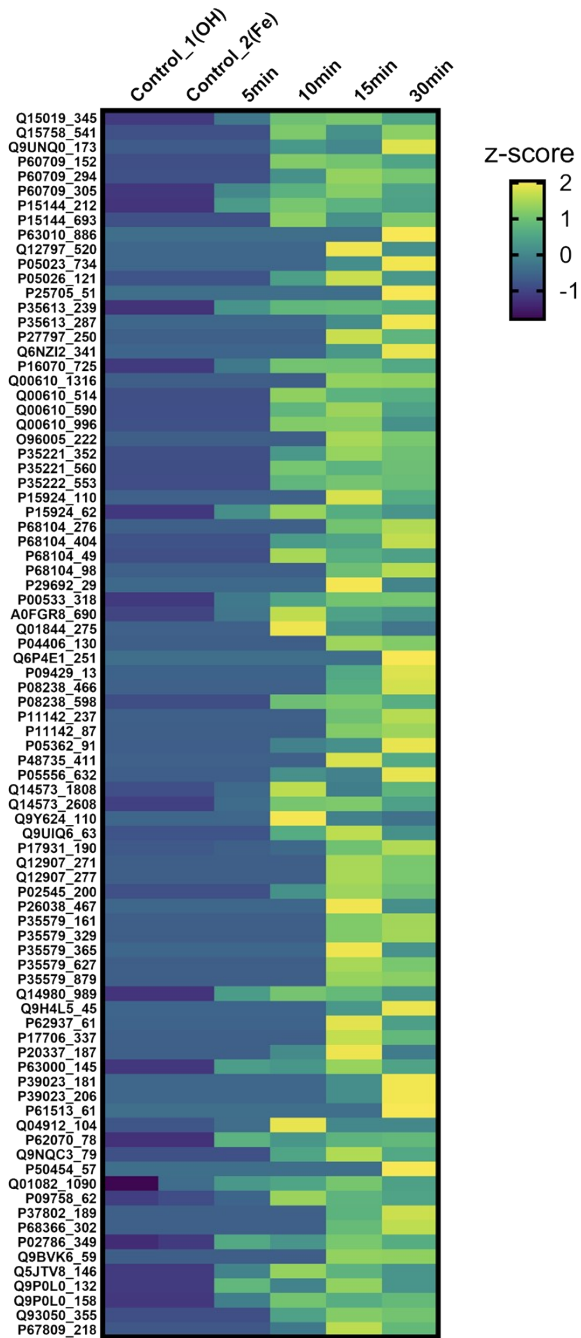
CD44: SQEM(+16)VHLVNK



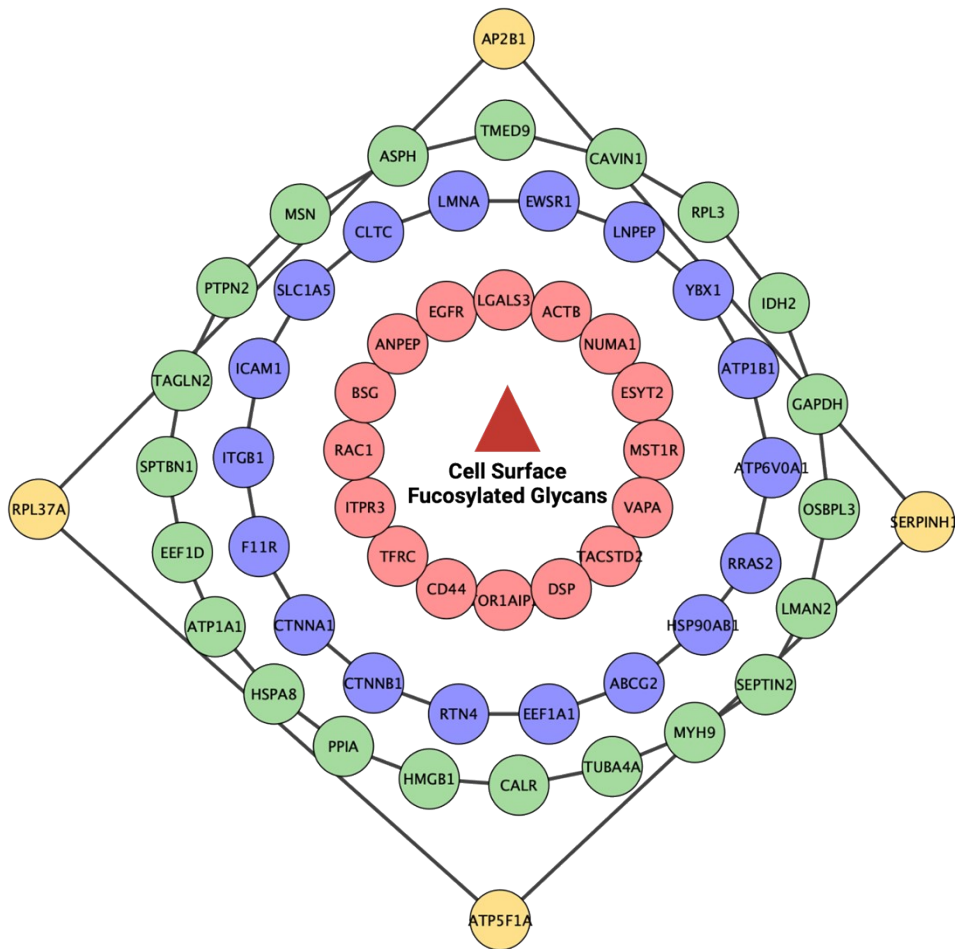
BASI: SSEHINEGETAML(+16)VCK



**Figure S7.** Annotated spectra of CD44 and BASI proteins in the control, H<sub>2</sub>O<sub>2</sub>, and FeBABE-H<sub>2</sub>O<sub>2</sub> treatments.



**Figure S8.** The extent of oxidation quantified by Byologic for selected proteins oxidized in PNT2 cells. Each column represents one treatment condition, and each row represents one oxidation site of a selected protein.



**Proteins colored by H<sub>2</sub>O<sub>2</sub> treatment time:**

5 min

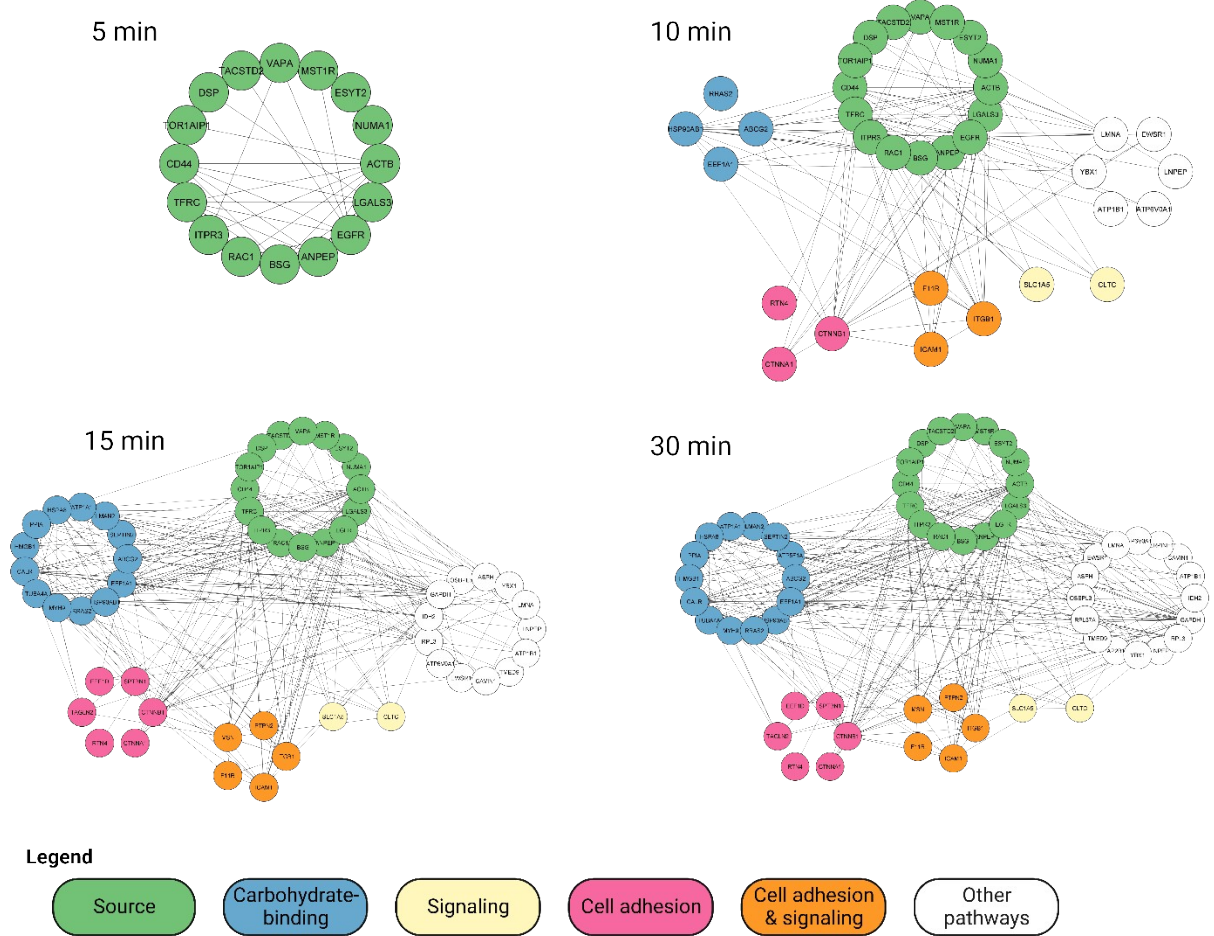
10 min

15 min

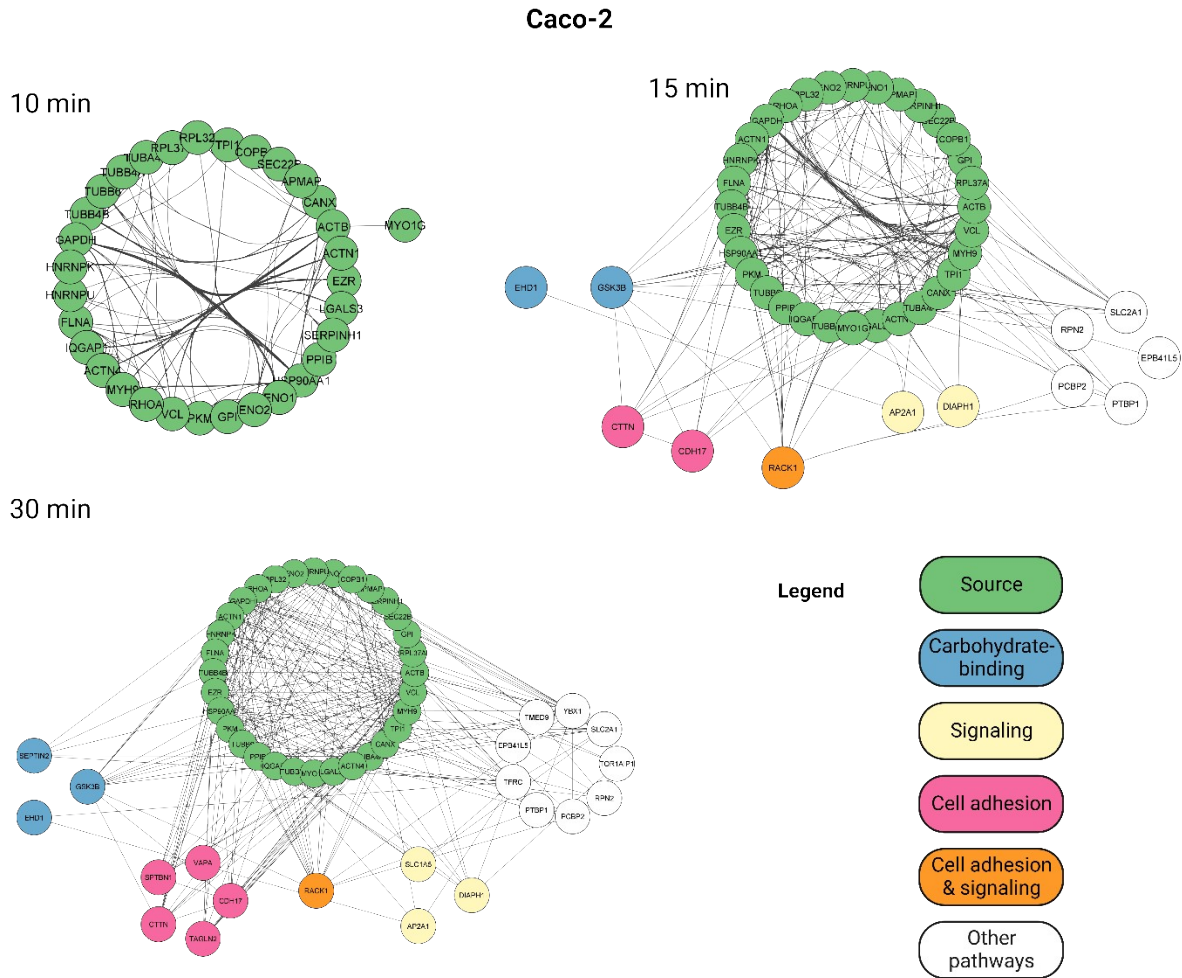
30 min

**Figure S9. Proteins from PNT2 within the spatial environment of cell-surface fucose identified by POFE.** The spatial information was revealed by different H<sub>2</sub>O<sub>2</sub> treatment time. Proteins with different spatial distances (reported as unit of time) relative to the fucose were labeled with different colors. Colors correspond to incubation time: 5 minutes (red), 10 minutes (blue), 15 minutes (green), and 30 minutes (orange).

PNT2



**Figure S10. Interaction networks of PNT2.** Proteins are categorized and color-coded into different biological pathways based on being the Source protein (green) or as Molecular Functions: Carbohydrate binding (blue), Signaling (yellow), Cell adhesion (pink), Cell adhesion and signaling (orange), and Other pathways (white).



**Figure S11. Interaction networks of Caco-2.** Proteins are categorized and color-coded into different biological pathways based on being the Source protein (green) or as Molecular Functions: Carbohydrate binding (blue), Signaling (yellow), Cell adhesion (pink), Cell adhesion and signaling (orange), and Other pathways (white).

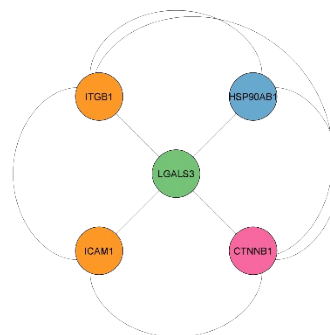


## PNT2

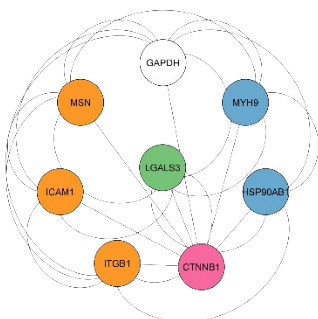
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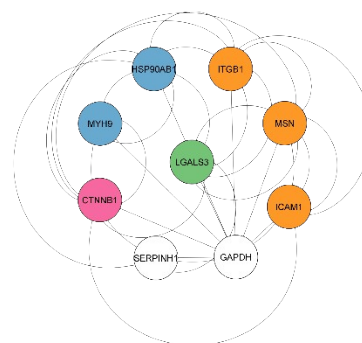
10 min



15 min



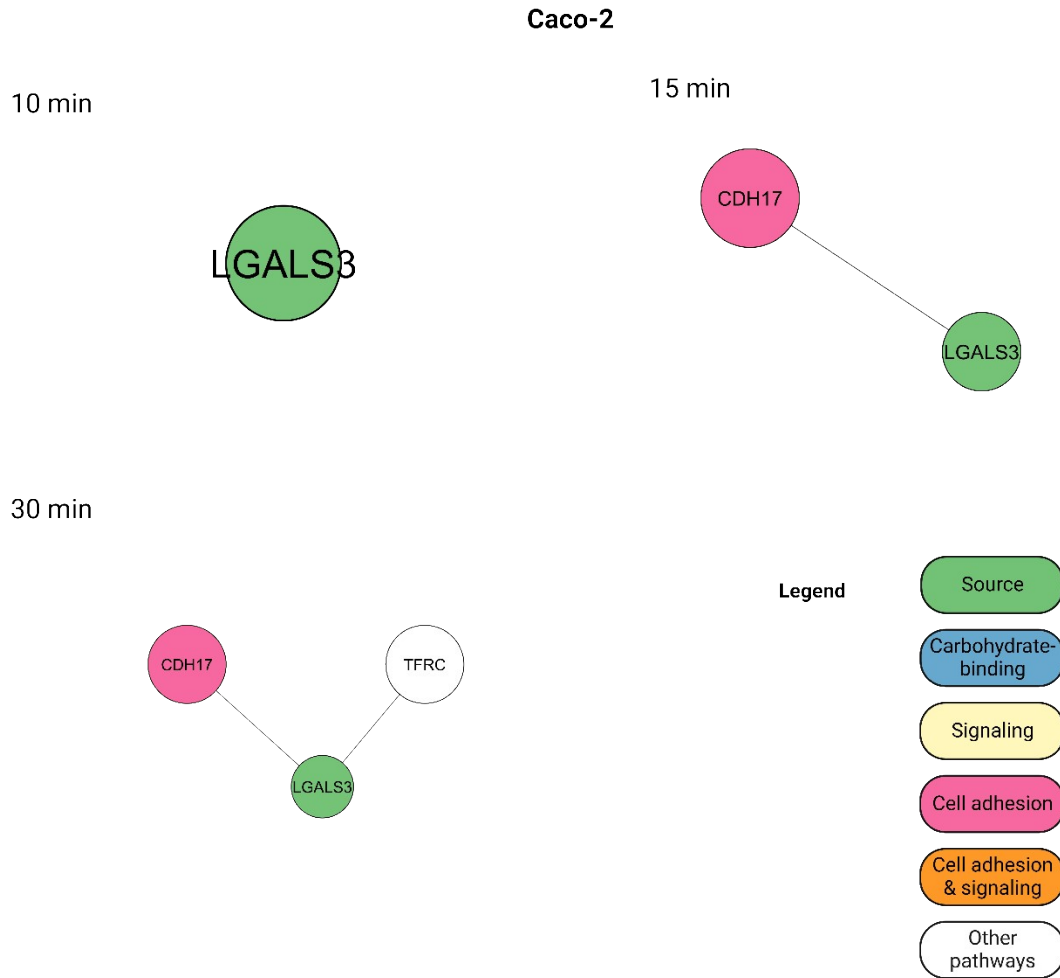
30 min



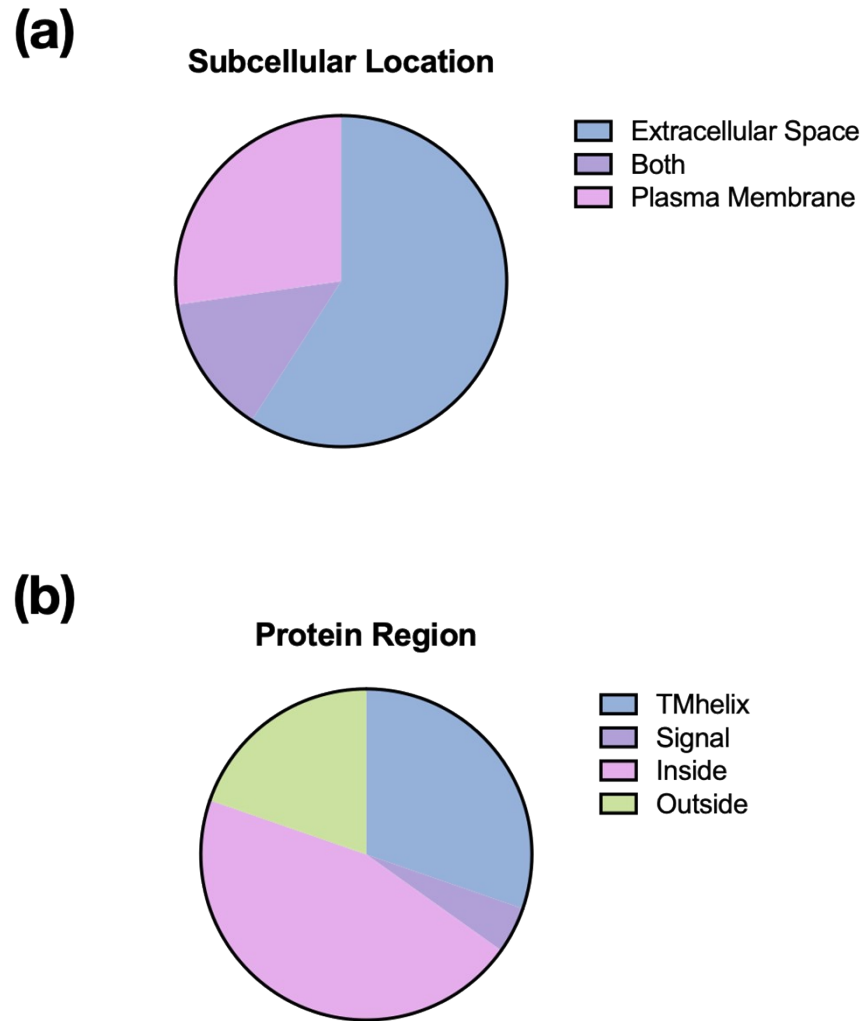
### Legend



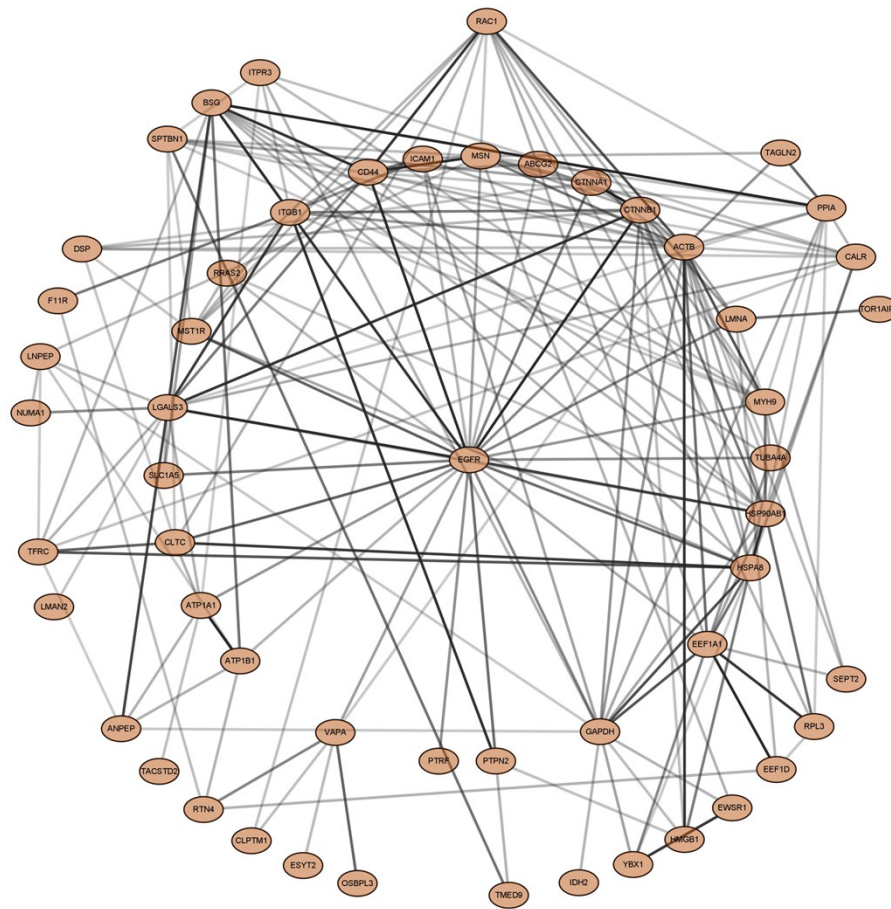
**Figure S12. Interaction networks of LGALS3 in PNT2.** Proteins are categorized and color-coded into different biological pathways based on being the Source protein (green) or as Molecular Functions: Carbohydrate binding (blue), Signaling (yellow), Cell adhesion (pink), Cell adhesion and signaling (orange), and Other pathways (white).



**Figure S13. Interaction networks of LGALS3 in Caco-2.** Proteins are categorized and color-coded into different biological pathways based on being the Source protein (green) or as Molecular Functions: Carbohydrate binding (blue), Signaling (yellow), Cell adhesion (pink), Cell adhesion and signaling (orange), and Other pathways (white).



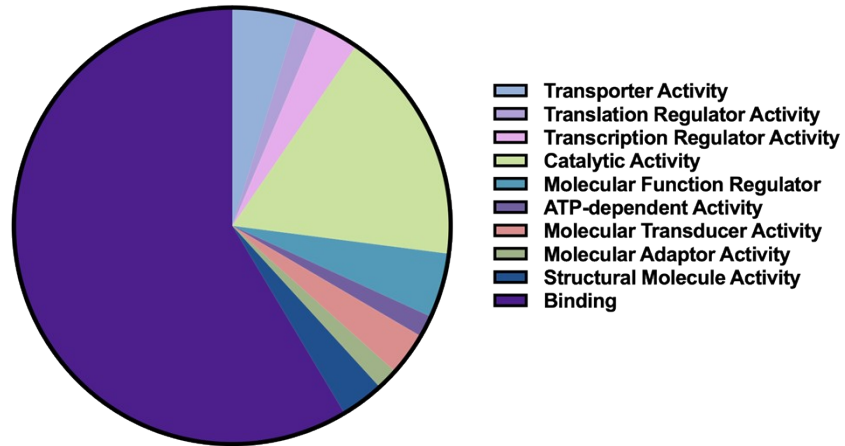
**Figure S14. Annotated Location of identified proteins from PNT2.** **(a)** The annotated subcellular location of oxidized proteins (level 3) by Gene Ontology (GO). **(b)** The predicted protein region distribution of oxidized proteins (level 3) by DeepTMHMM.



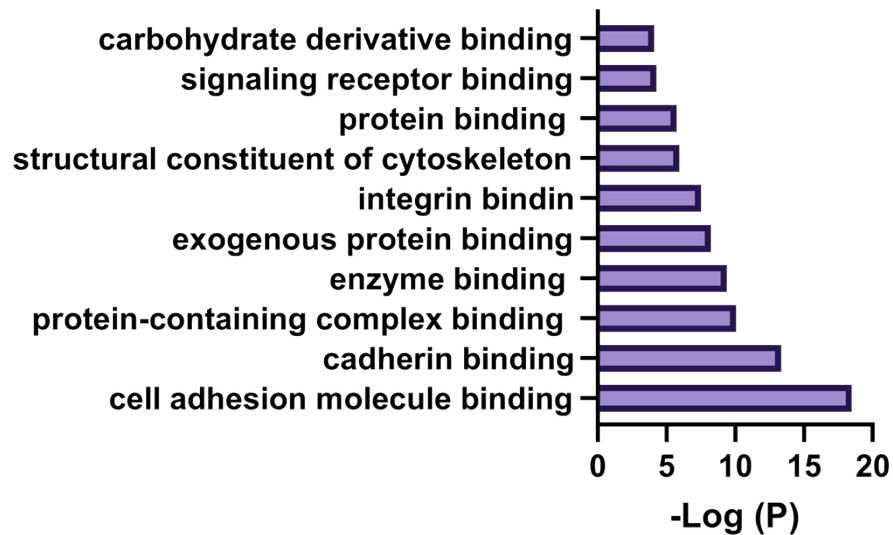
**Figure S15. Interaction network of the identified proteins in proximity to the cell-surface fucose.** All the interactions were identified by STRING and visualized by Cytoscape.

**(a)**

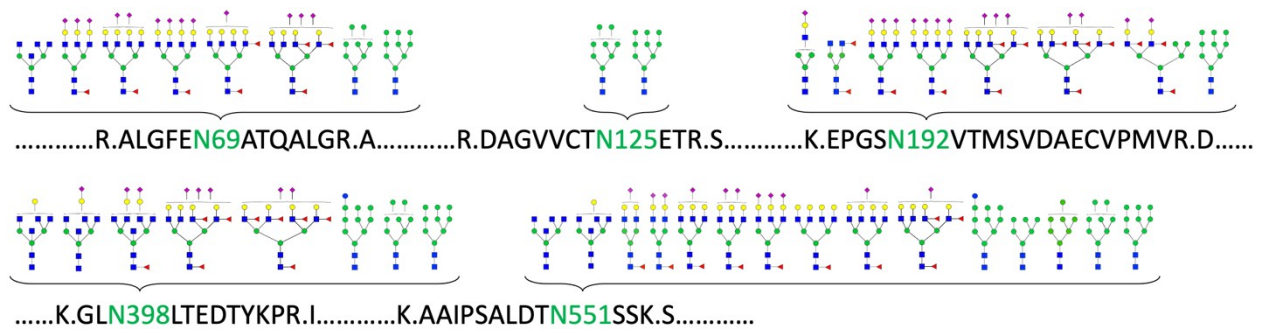
### Molecular Function



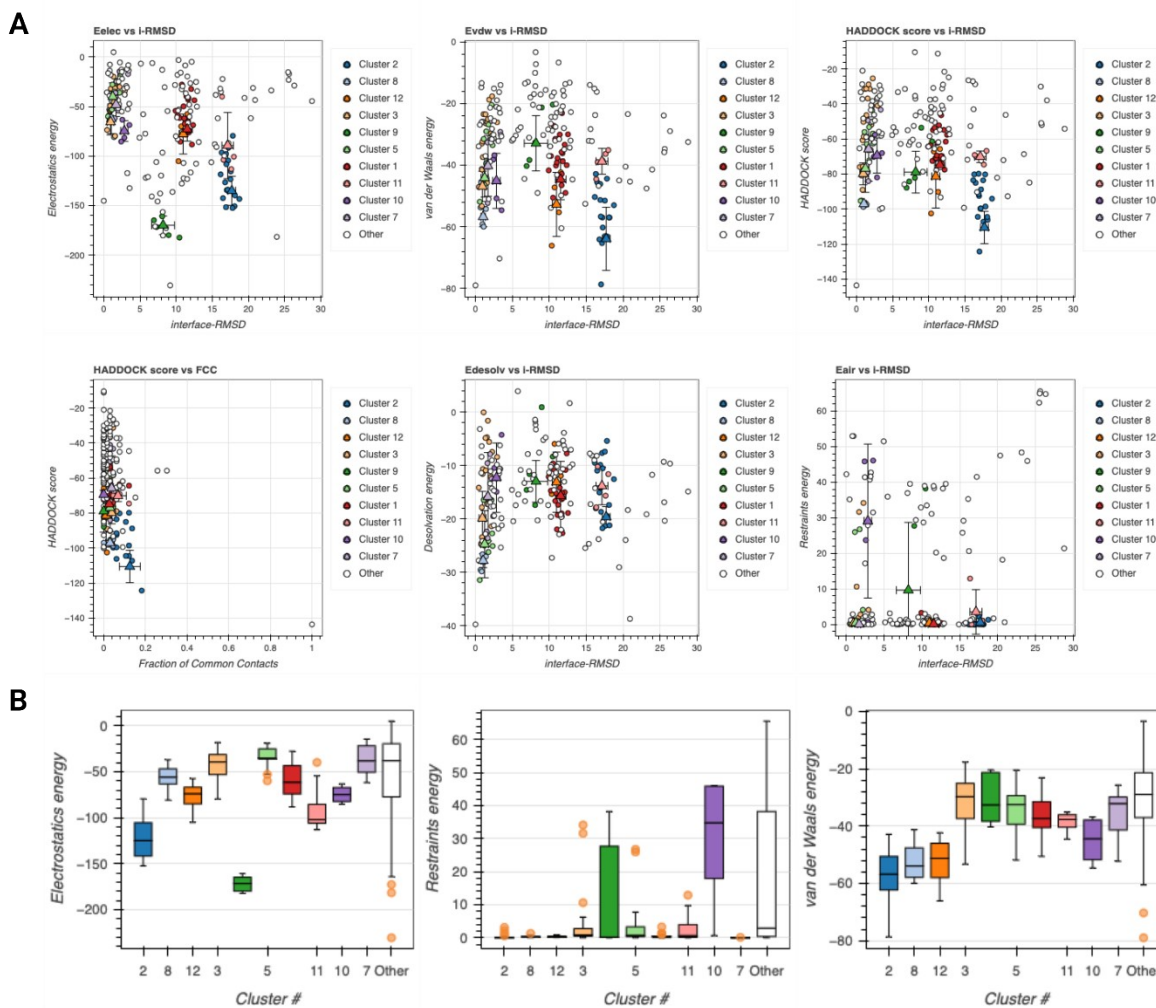
**(b)**



**Figure S16. Functional annotation of identified proteins from PNT2 by POFE. (a)** Molecular functions of the identified proteins. **(b)** Binding function of the identified proteins.

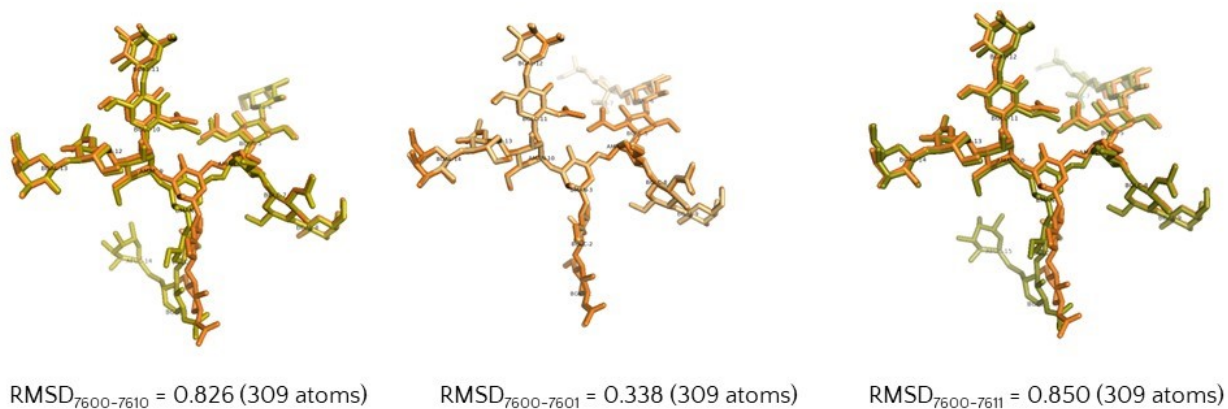


**Figure S17. The N-glycans associated with the five glycosylation sites of protein LG3BP identified from PNT2 cells.** Putative glycan structures were assigned for each site.



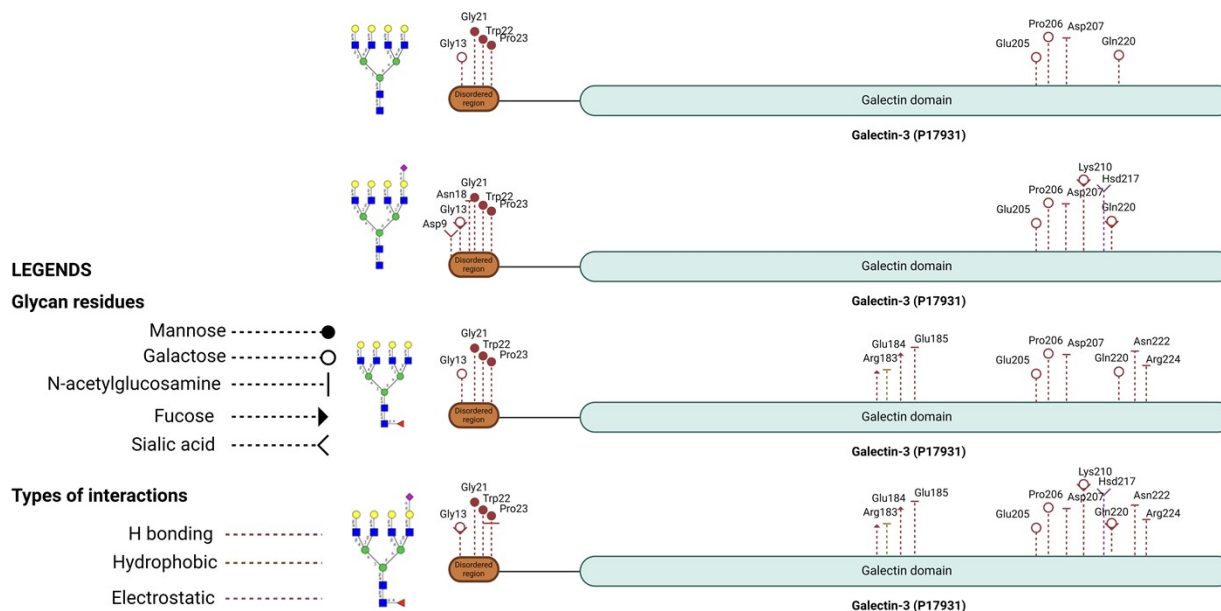
**Figure S18. Modeling statistics of LEG3 and LG3BP from HADDOCK.** HADDOCK clustered (a) 115 structures in (b) 12 cluster(s), which represents 57 % of the water-refined models HADDOCK generated.

**Fucose changes conformation of N-glycan core**  
**[Man- $\beta$ -(1-4)-GlcNAc- $\beta$ -(1-4)-GlcNAc- $\beta$ -(1-4)]**



**Figure S19. Modeling of glycan structures with or without the addition fucose.** The conformation of fucosylated N-glycan structures significantly changed compared to the undecorated N-glycan.





**Figure S20. Summary of the glycan-protein interaction maps between LEG3 and LG3BP-Asn511 N-glycans.** The domain information for LEG3 was obtained from Uniprot. The N-glycans were found to interact primarily with the Galectin domain of LEG3.