Electronic Supplementary Material (ESI) for Sensors & Diagnostics. This journal is © The Royal Society of Chemistry 2023

## **Supporting Information**

## Multi-Target Amyloid Probing and Inhibition Using Basic Orange Fluorescence

Yijing Tang<sup>1</sup>, Dong Zhang<sup>1</sup>, Xiong Gong<sup>2</sup>, and Jie Zheng<sup>1\*</sup>

<sup>1</sup>Department of Chemical, Biomolecular, and Corrosion Engineering The University of Akron, Ohio, USA

> <sup>2</sup>School of Polymer Science and Polymer Engineering The University of Akron, Ohio, USA

\* Corresponding author: <a href="mailto:zhengj@uakron.edu">zhengj@uakron.edu</a>

## Table S1. Comparison of typical amyloid probes and inhibitors in terms of their $K_D$ value and relative inhibited fibrillization rate

		Entry	Detectors/	Target	Binding	Inhibition	Ref.
			Modulators affinity/Sensitivity r		rate (%)		
					(µM)		
Single	Single-	1	PicoGreen	insulin	~38.5	-	1
function-	targeted	2	stryl-11 insulin 20		-	2	
detectors	_	3	Thienoquinoxaline	Αβ	0.077	-	3
		4	styryl-quinoxaline (SQ)	Αβ	0.294	-	3
		5	L1	Αβ	-	-	4
		6	L2	Αβ	-	-	4
		7	L3	Αβ	-	-	4
		8	<u>Mu1</u> <u>Aβ</u> 90.91		-	5	
		$\begin{array}{c c c c c c c c c c c c c c c c c c c $		0.0085	-	6	
		10	ANCA	Αβ	1.4-13.8 (6)	-	7
	Multi-	11	Thioflavin T (ThT)	Αβ,	~0.58	-	8,9
	targeted			hIAPP,			
				hCT, α-			
				syn, etc			
		12	1-anilino-8-naphthalene	Αβ,	-	-	8
			sulfonate (ANS)	hIAPP,			
				hCT, α-			
				syn, etc			
		13	Benzothiazole aniline (BTA)	A $\beta$ , tau	~0.02	-	10
		14	auramine-O (AuO)	BSA,	~1.12	-	11,12
				insulin			
		15	PD-NA	Αβ,	-	-	13
				HEWL			
		16	PD-NA-TEG	Αβ,	-	-	13
				HEWL			
		17	TPE-TPP	Aβ, α-syn,	4.36-8.93	-	14-16
				α-LA,			
				HEWL,			
				κ-casein,			
				$D/6N\beta 2m$ ,			
				αB-			
Circal -	<b>Sim</b> -1 -	10				27	17
Single	Single-	18	<u>meiatonin</u>		-	2/	17
inhibitor	largeled	19			-	100	18
		20	azadıracıtın		-	~100	19
				Ap	-	20-90	20
		22			-	4/	20
		23			-	49	20
		24	troglitazone	niAPP	-	90	20

	Multi-	25	EGCG	$A\beta$ , α-syn,	-	77-100	21-24
	targeted			tau, TTR,			
	C			Htt, hCT,			
				hIAPP			
		26	Genistein	Aβ, hIAPP	-	40-63	25
		27	Resveratrol	Aβ, hIAPP	-	88	26
		28	Vitamin k3	HEWL,	-	77-81	27
				Αβ			
		29	morin	Αβ,	-	~100	28-30
				hIAPP,			
				insulin			
		30	quercetin	Αβ,	-	~100	28,31-
				hIAPP, α-			33
				syn, tau			
		31	myricetin	Αβ,	-	~100	28,34-
				hIAPP, α-			36
				syn, tau			
		32	rutin	Aβ, hIAPP	-	~100	37-39
		33	curcumin	Aβ,hIAPP,	-	~100	23,40-
				α-syn, PrP,			44
				TTR			
		34	silibinin	Aβ, hIAPP	-	49-70	20,45
		35	caffeine	Aβ, hIAPP	-	28	46,47
Dual	Single-	36	BSPOTPE	Αβ	-	~100	48
function-	targeted						
detectors	Multi-	37	Congo red	Αβ,	0.1-1.1	-	49,50
and	targeted			hIAPP,			
modulators				hCT, α-			
				syn, etc			
		38	Basic orange 21 (BO21)	Αβ,	0.015-0.24	~100	51
				hIAPP,			This
				hCT, α-			work
				syn			

	Linear range (µM)	Linear function	R <sup>2</sup>	$K_D(nM)$
mAβ-BO21	0.625-40	y = 0.3853x + 0.013	0.9502	34
οΑβ-ΒΟ21	2.5-40	y = 0.4546x + 0.0067	0.9868	15
fAβ-BO21	0.625-40	y = 0.0682x + 0.0095	0.9555	139
mhIAPP-BO21	0.625-40	y = 2.029x + 0.094	0.9786	46
ohIAPP-BO21	1.25-40	y = 0.1771x + 0.0032	0.9994	18
fhIAPP-BO21	2.5-40	y = 0.0539x + 0.0062	0.9742	115
mhCT-BO21	0.625-40	y = 0.1786x + 0.0051	0.9945	29
ohCT-BO21	5-40	y = 0.0832x + 0.003	0.9935	36
fhCT-BO21	2.5-40	y = 0.4642x + 0.1113	0.9810	240

 Table S2. Summary of fluorescence spectroscopic titration

	Αβ-ΒΟ21	hIAPP-BO21	hCT-BO21	Aβ-ThT	hIAPP-ThT	hCT-ThT
δ	5.512	9.247	9.851	32.94	77.18	116.8
$K(\mu M^{-1})$	111.8	142.2	110	185.3	577.5	127.5
$DL(\mu M)$	0.1479	0.1951	0.2687	0.5333	0.4009	2.7482
$R^2$	0.9852	0.9745	0.9528	0.8365	0.9005	0.1616

Table S3. Detection limits of BO21 and ThT for different amyloid peptides

The detection limits (DL) were estimated based on the  $3\delta/k$  method.



**Figure S1.** Linear fitting analysis of fluorescence spectroscopic titration shown in **Fig. 3a** using  $1/(Ft-F0)=1/(Fmax-F0) + 1/((Fmax-F0)\cdot KD\cdot [X])$ . 1 µM BO21 was added as the increased concentrations (0-40 µM) of monomeric (1<sup>st</sup> row), oligomeric (2<sup>nd</sup> row) and fibrillar (3<sup>rd</sup> row) aggregates of A $\beta$  (1<sup>st</sup> column), hIAPP (2<sup>nd</sup> column), and hCT (3<sup>rd</sup> column) under excitation of 470 nm.



Figure S2. Fluorescence image of amyloid monomers (1st row) and oligomers (2nd row) stained by BO21.



**Figure S3.** Fluorescence spectra of 1  $\mu$ M BO21 (1st column) and ThT (2nd column) in the presence (colored) and absence (black) of fibrillar aggregates of A $\beta$  (1st raw), hIAPP (2nd raw), and hCT (3rd raw).



**Figure S4.** Linear fitting analysis of fluorescence intensity and correasponding signal/noise ratio of using 1  $\mu$ M BO21 (1<sup>st</sup> column) and ThT (2<sup>nd</sup> column) as probe to detect increasing concentrations (0-40  $\mu$ M) of fibrillar aggregates of A $\beta$  (1<sup>st</sup> raw), hIAPP (2<sup>nd</sup> raw), and hCT (3<sup>rd</sup> raw) under excitation of 470 nm.



**Figure S5.** The fluorescence spectrum (1st column) and corresponding linear fitting analysis (2nd column) of 20  $\mu$ M A $\beta$  (1st column), hIAPP (2nd column), and hCT (3rd column) by stepwise addition of BO21 (0.01 to 1  $\mu$ M) in PBS buffer (pH = 7.4).  $\lambda$ ex = 470 nm.



**Figure S6.** The fluorescence spectrum (1st column) and corresponding linear fitting analysis (2nd column) of 20  $\mu$ M A $\beta$  (1st column), hIAPP (2nd column), and hCT (3rd column) by stepwise addition of ThT (0.01 to 1  $\mu$ M) in PBS buffer (pH = 7.4).  $\lambda$ ex = 450 nm.



**Figure S7**. Time-dependent ThT fluorescence curves to monitor the aggregation kinetics of 20  $\mu$ M pure amyloid peptides of A $\beta$ , hIAPP, and hCT with and without 1  $\mu$ M BO21.



Figure S8. Time-dependent ThT fluorescence curves to monitor the aggregation kinetics of pure BO21 at different concentrations of 0.1-80  $\mu$ M.



Figure S9. Time-dependent circular dichroism spectra for monitoring the secondary structure changes by adding 5-20  $\mu$ M BO21 to freshly prepared A $\beta$ .



Figure S10. Time-dependent circular dichroism spectra for monitoring the secondary structure changes by adding 20-80  $\mu$ M BO21 to freshly prepared hIAPP.



Figure S11. Time-dependent circular dichroism spectra for monitoring the secondary structure changes by adding 20-80  $\mu$ M BO21 to freshly prepared hCT.



**Figure S12.** MTT reduction assay for cell viability (1st column) and LDH activity assay for cytotoxicity (2nd column) in the presence of different concentrations of BO21 from 0.1 to 80  $\mu$ M. t-test was used for data analysis for cells treated with amyloids relative to untreated cell groups (n>3). (°p < 0.05, °°p < 0.01, °°°p < 0.005, °°°p < 0.001).

## Reference

- (1) Mora, A. K.; Singh, P. K.; Patro, B. S.; Nath, S. PicoGreen: a better amyloid probe than Thioflavin-T. *Chemical Communications* **2016**, *52* (82), 12163.
- (2) Mudliar, N. H.; Singh, P. K. A molecular rotor-based turn-on sensor probe for amyloid fibrils in the extreme near-infrared region. *Chemical Communications* **2019**, *55* (27), 3907.
- (3) Benzeid, H.; Mothes, E.; Essassi, E. M.; Faller, P.; Pratviel, G. A thienoquinoxaline and a styryl-quinoxaline as new fluorescent probes for amyloid-β fibrils. *Comptes Rendus Chimie* 2012, 15 (1), 79.
- (4) Dyrager, C.; Vieira, R. P.; Nyström, S.; Nilsson, K. P. R.; Storr, T. Synthesis and evaluation of benzothiazole-triazole and benzothiadiazole-triazole scaffolds as potential molecular probes for amyloid-β aggregation. *New Journal of Chemistry* 2017, *41* (4), 1566.
- (5) Mu, X.; Wu, F.; Wang, R.; Huang, Z.; Lv, T.; Lu, Y.; Liu, B.; Zhou, X. A cyaninederived NIR molecular rotor for ratiometric imaging of amyloid-β aggregates. *Sensors Actuators B: Chemical* 2021, 338, 129842.
- (6) Yue, N.; Fu, H.; Chen, Y.; Gao, X.; Dai, J.; Cui, M. Rational design of molecular rotorbased fluorescent probes with bi-aromatic rings for efficient in vivo detection of amyloidβ plaques in Alzheimer's disease. *European Journal of Medicinal Chemistry* 2022, 243, 114715.
- (7) Chang, W. M.; Dakanali, M.; Capule, C. C.; Sigurdson, C. J.; Yang, J.; Theodorakis, E. A. ANCA: a family of fluorescent probes that bind and stain amyloid plaques in human tissue. *ACS chemical neuroscience* **2011**, *2* (5), 249.
- (8) Sulatsky, M.; Sulatskaya, A.; Povarova, O.; Antifeeva, I. A.; Kuznetsova, I.; Turoverov, K. Effect of the fluorescent probes ThT and ANS on the mature amyloid fibrils. *Prion* 2020, *14* (1), 67.
- (9) Ghadami, S. A.; Ahadi-Amandi, K.; Khodarahmi, R.; Ghanbari, S.; Adibi, H. Synthesis of benzylidene-indandione derivatives as quantification of amyloid fibrils. *Biophysical chemistry* **2023**, *296*, 106982.
- (10) Klunk, W. E.; Wang, Y.; Huang, G.-f.; Debnath, M. L.; Holt, D. P.; Mathis, C. A. Uncharged thioflavin-T derivatives bind to amyloid-beta protein with high affinity and readily enter the brain. *Life sciences* **2001**, *69* (13), 1471.
- (11) Mudliar, N. H.; Sadhu, B.; Pettiwala, A. M.; Singh, P. K. Evaluation of an ultrafast molecular rotor, auramine O, as a fluorescent amyloid marker. *The Journal of Physical Chemistry B* **2016**, *120* (40), 10496.
- (12) Amdursky, N.; Huppert, D. Auramine-O as a fluorescence marker for the detection of amyloid fibrils. *The Journal of Physical Chemistry B* **2012**, *116* (45), 13389.
- (13) Wang, Y.-L.; Fan, C.; Xin, B.; Zhang, J.-P.; Luo, T.; Chen, Z.-Q.; Zhou, Q.-Y.; Yu, Q.; Li, X.-N.; Huang, Z.-L.et al. AIE-based super-resolution imaging probes for β-amyloid plaques in mouse brains. *Materials Chemistry Frontiers* **2018**, *2* (8), 1554.
- (14) Leung, C. W. T.; Guo, F.; Hong, Y.; Zhao, E.; Kwok, R. T. K.; Leung, N. L. C.; Chen, S.; Vaikath, N. N.; El-Agnaf, O. M.; Tang, Y. Detection of oligomers and fibrils of αsynuclein by AIEgen with strong fluorescence. *Chemical Communications* **2015**, *51* (10), 1866.

- (15) Das, A.; Gupta, A.; Hong, Y.; Carver, J. A.; Maiti, S. A Spectroscopic Marker for Structural Transitions Associated with Amyloid-β Aggregation. *Biochemistry* 2020, 59 (19), 1813.
- (16) Kumar, M.; Hong, Y.; Thorn, D. C.; Ecroyd, H.; Carver, J. A. Monitoring early-stage protein aggregation by an aggregation-induced emission fluorogen. *Analytical chemistry* 2017, 89 (17), 9322.
- (17) Aarabi, M. H.; Mirhashemi, S. M. To estimate effective antiamyloidogenic property of melatonin and fisetin and their actions to destabilize amyloid fibrils. *Pak J Pharm Sci* 2017, 30 (5), 1589.
- (18) Dubey, R.; Patil, K.; Dantu, S. C.; Sardesai, D. M.; Bhatia, P.; Malik, N.; Acharya, J. D.; Sarkar, S.; Ghosh, S.; Chakrabarti, R. Azadirachtin inhibits amyloid formation, disaggregates pre-formed fibrils and protects pancreatic β-cells from human islet amyloid polypeptide/amylin-induced cytotoxicity. *Biochemical Journal* **2019**, *476* (5), 889.
- (19) Török, M.; Abid, M.; Mhadgut, S. C.; Török, B. Organofluorine inhibitors of amyloid fibrillogenesis. *Biochemistry* **2006**, *45* (16), 5377.
- (20) Fortin, J. S.; Benoit-Biancamano, M.-O. In vitro evaluation of hypoglycemic agents to target human islet amyloid polypeptide: a key protein involved in amyloid deposition and beta-cell loss. *Canadian journal of diabetes* **2015**, *39* (5), 373.
- (21) Xu, Z.-X.; Zhang, Q.; Ma, G.-L.; Chen, C.-H.; He, Y.-M.; Xu, L.-H.; Zhang, Y.; Zhou, G.-R.; Li, Z.-H.; Yang, H.-J. Influence of aluminium and EGCG on fibrillation and aggregation of human islet amyloid polypeptide. *Journal of diabetes research* 2016, *2016*.
- (22) Ehrnhoefer, D. E.; Duennwald, M.; Markovic, P.; Wacker, J. L.; Engemann, S.; Roark, M.; Legleiter, J.; Marsh, J. L.; Thompson, L. M.; Lindquist, S. Green tea (-)-epigallocatechin-gallate modulates early events in huntingtin misfolding and reduces toxicity in Huntington's disease models. *Human molecular genetics* 2006, *15* (18), 2743.
- (23) Ferreira, N.; Saraiva, M. J.; Almeida, M. R. Natural polyphenols inhibit different steps of the process of transthyretin (TTR) amyloid fibril formation. *FEBS letters* **2011**, *585* (15), 2424.
- (24) Wobst, H. J.; Sharma, A.; Diamond, M. I.; Wanker, E. E.; Bieschke, J. The green tea polyphenol (–)-epigallocatechin gallate prevents the aggregation of tau protein into toxic oligomers at substoichiometric ratios. *FEBS letters* **2015**, *589* (1), 77.
- (25) Ren, B.; Liu, Y.; Zhang, Y.; Cai, Y.; Gong, X.; Chang, Y.; Xu, L.; Zheng, J. Genistein: a dual inhibitor of both amyloid β and human islet amylin peptides. ACS chemical neuroscience 2018, 9 (5), 1215.
- (26) Mishra, R.; Sellin, D.; Radovan, D.; Gohlke, A.; Winter, R. Inhibiting islet amyloid polypeptide fibril formation by the red wine compound resveratrol. *ChemBioChem* 2009, 10 (3), 445.
- (27) Alam, P.; Chaturvedi, S. K.; Siddiqi, M. K.; Rajpoot, R. K.; Ajmal, M. R.; Zaman, M.; Khan, R. H. Vitamin k3 inhibits protein aggregation: implication in the treatment of amyloid diseases. *Scientific reports* **2016**, *6* (1), 1.
- (28) Ono, K.; Yoshiike, Y.; Takashima, A.; Hasegawa, K.; Naiki, H.; Yamada, M. Potent anti-amyloidogenic and fibril-destabilizing effects of polyphenols in vitro: implications for the prevention and therapeutics of Alzheimer's disease. *Journal of neurochemistry* 2003, 87 (1), 172.
- (29) Noor, H.; Cao, P.; Raleigh, D. P. Morin hydrate inhibits amyloid formation by islet amyloid polypeptide and disaggregates amyloid fibers. *Protein Science* **2012**, *21* (3), 373.

- (30) Patel, P.; Parmar, K.; Das, M. Inhibition of insulin amyloid fibrillation by Morin hydrate. *International journal of biological macromolecules* **2018**, *108*, 225.
- (31) López, L. C.; Varea, O.; Navarro, S.; Carrodeguas, J. A.; Sanchez de Groot, N.; Ventura, S.; Sancho, J. Benzbromarone, quercetin, and folic acid inhibit amylin aggregation. *International journal of molecular sciences* 2016, *17* (6), 964.
- (32) Zhu, M.; Han, S.; Fink, A. L. Oxidized quercetin inhibits α-synuclein fibrillization. *Biochimica et Biophysica Acta (BBA)-General Subjects* **2013**, *1830* (4), 2872.
- (33) Kumar, S.; Krishnakumar, V. G.; Morya, V.; Gupta, S.; Datta, B. Nanobiocatalyst facilitated aglycosidic quercetin as a potent inhibitor of tau protein aggregation. *International journal of biological macromolecules* **2019**, *138*, 168.
- (34) Aarabi, M.-H.; Mirhashemi, S. M. The role of two natural flavonoids on human amylin aggregation. *African Journal of Pharmacy and Pharmacology* **2012**, *6* (31), 2374.
- (35) Ono, K.; Yamada, M. Antioxidant compounds have potent anti-fibrillogenic and fibril-destabilizing effects for  $\alpha$ -synuclein fibrils in vitro. *Journal of neurochemistry* **2006**, *97* (1), 105.
- (36) Taniguchi, S.; Suzuki, N.; Masuda, M.; Hisanaga, S.-i.; Iwatsubo, T.; Goedert, M.; Hasegawa, M. Inhibition of heparin-induced tau filament formation by phenothiazines, polyphenols, and porphyrins. *Journal of Biological Chemistry* **2005**, *280* (9), 7614.
- (37) Aitken, J. F.; Loomes, K. M.; Riba-Garcia, I.; Unwin, R. D.; Prijic, G.; Phillips, A. S.; Phillips, A. R.; Wu, D.; Poppitt, S. D.; Ding, K. Rutin suppresses human-amylin/hIAPP misfolding and oligomer formation in-vitro, and ameliorates diabetes and its impacts in human-amylin/hIAPP transgenic mice. *Biochemical and biophysical research communications* 2017, 482 (4), 625.
- (38) Yu, X.-L.; Li, Y.-N.; Zhang, H.; Su, Y.-J.; Zhou, W.-W.; Zhang, Z.-P.; Wang, S.-W.; Xu, P.-X.; Wang, Y.-J.; Liu, R.-T. Rutin inhibits amylin-induced neurocytotoxicity and oxidative stress. *Food & Function* **2015**, *6* (10), 3296.
- (39) Wang, S.-w.; Wang, Y.-J.; Su, Y.-j.; Zhou, W.-w.; Yang, S.-g.; Zhang, R.; Zhao, M.; Li, Y.-n.; Zhang, Z.-p.; Zhan, D.-w. Rutin inhibits β-amyloid aggregation and cytotoxicity, attenuates oxidative stress, and decreases the production of nitric oxide and proinflammatory cytokines. *Neurotoxicology* **2012**, *33* (3), 482.
- (40) Daval, M.; Bedrood, S.; Gurlo, T.; Huang, C.-J.; Costes, S.; Butler, P. C.; Langen, R. The effect of curcumin on human islet amyloid polypeptide misfolding and toxicity. *Amyloid* **2010**, *17* (3-4), 118.
- (41) Sparks, S.; Liu, G.; Robbins, K. J.; Lazo, N. D. Curcumin modulates the self-assembly of the islet amyloid polypeptide by disassembling α-helix. *Biochemical and biophysical research communications* 2012, 422 (4), 551.
- (42) Yang, F.; Lim, G. P.; Begum, A. N.; Ubeda, O. J.; Simmons, M. R.; Ambegaokar, S. S.; Chen, P. P.; Kayed, R.; Glabe, C. G.; Frautschy, S. A. Curcumin inhibits formation of amyloid β oligomers and fibrils, binds plaques, and reduces amyloid in vivo. *Journal of Biological Chemistry* 2005, 280 (7), 5892.
- (43) Pandey, N.; Strider, J.; Nolan, W. C.; Yan, S. X.; Galvin, J. E. Curcumin inhibits aggregation of α-synuclein. *Acta neuropathologica* **2008**, *115* (4), 479.
- (44) Caughey, B.; Raymond, L. D.; Raymond, G. J.; Maxson, L.; Silveira, J.; Baron, G. S. Inhibition of protease-resistant prion protein accumulation in vitro by curcumin. *Journal* of virology **2003**, 77 (9), 5499.

- (45) Yin, F.; Liu, J.; Ji, X.; Wang, Y.; Zidichouski, J.; Zhang, J. Silibinin: A novel inhibitor of Aβ aggregation. *Neurochemistry international* **2011**, *58* (3), 399.
- (46) Cheng, B.; Liu, X.; Gong, H.; Huang, L.; Chen, H.; Zhang, X.; Li, C.; Yang, M.; Ma, B.; Jiao, L. Coffee components inhibit amyloid formation of human islet amyloid polypeptide in vitro: possible link between coffee consumption and diabetes mellitus. *Journal of agricultural food chemistry* 2011, 59 (24), 13147.
- (47) Sharma, B.; Paul, S. Action of caffeine as an amyloid inhibitor in the aggregation of Aβ16–22 peptides. *The Journal of Physical Chemistry B* 2016, *120* (34), 9019.
- (48) Hong, Y.; Meng, L.; Chen, S.; Leung, C. W. T.; Da, L.-T.; Faisal, M.; Silva, D.-A.; Liu, J.; Lam, J. W. Y.; Huang, X. Monitoring and inhibition of insulin fibrillation by a small organic fluorogen with aggregation-induced emission characteristics. *Journal of the American Chemical Society* 2012, *134* (3), 1680.
- (49) Groenning, M. Binding mode of Thioflavin T and other molecular probes in the context of amyloid fibrils—current status. *Journal of chemical biology* **2010**, *3*, 1.
- (50) Lorenzo, A.; Yankner, B. A. Beta-amyloid neurotoxicity requires fibril formation and is inhibited by congo red. *Proceedings of the National Academy of Sciences* **1994**, *91* (25), 12243.
- (51) Pandey, S. P.; Singh, P. K. Basic Orange 21: a molecular rotor probe for fluorescence turn-on sensing of amyloid fibrils. *Journal of Molecular Liquids* **2020**, *303*, 112618.