

Supporting Information

Investigating the Complexation Propensity of Self-Assembling Dipeptides with the Anticancer Peptide-Drug Bortezomib: A Computational Study

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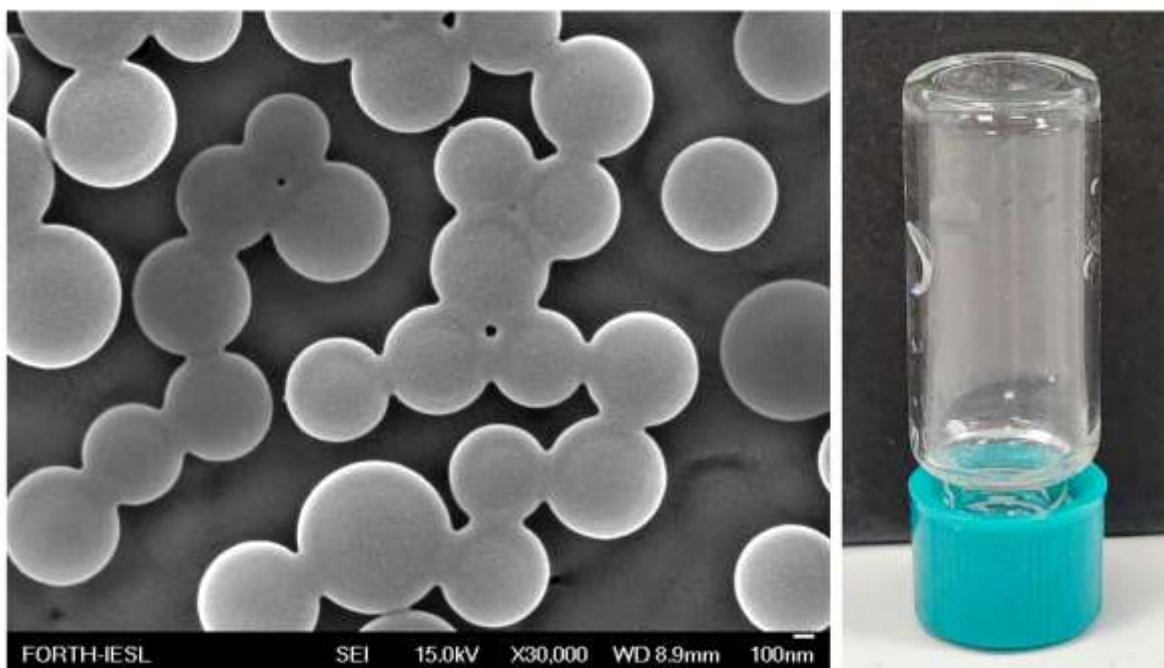
Keywords: dipeptides; Bortezomib; self-assembly; association; molecular dynamics

26 **Experimental materials and methods**

27 The Bortezomib peptide was purchased from Merck in form of lyophilized powder with purity
28 greater than 95 %. The water solvent used was nanopure purified, filtered and sterilized. Sample
29 solutions were prepared by dissolving the Bortezomib powder into water at room temperature
30 following sonication at c=4 mg/ml. Different concentrations were tested as well as briefly heating
31 Bortezomib up to 55 °C, for a couple of seconds, to facilitate dissolution.

32 For FESEM observation, a sample of 10 µL placed in a slide to be dried. After the samples were
33 dried, sputtering was carried out as follows: The samples were placed on a slide with the help of
34 conductive carbon adhesive tape. The slide was subsequently placed on a gold sputtering machine
35 and a 15 nm thick gold (Au) layer was deposited for 64 sec at 40 mA. For the observation of the
36 samples FESEM JEOL 7000 field emission scanning electron microscope was used, operating at
37 15 kV.

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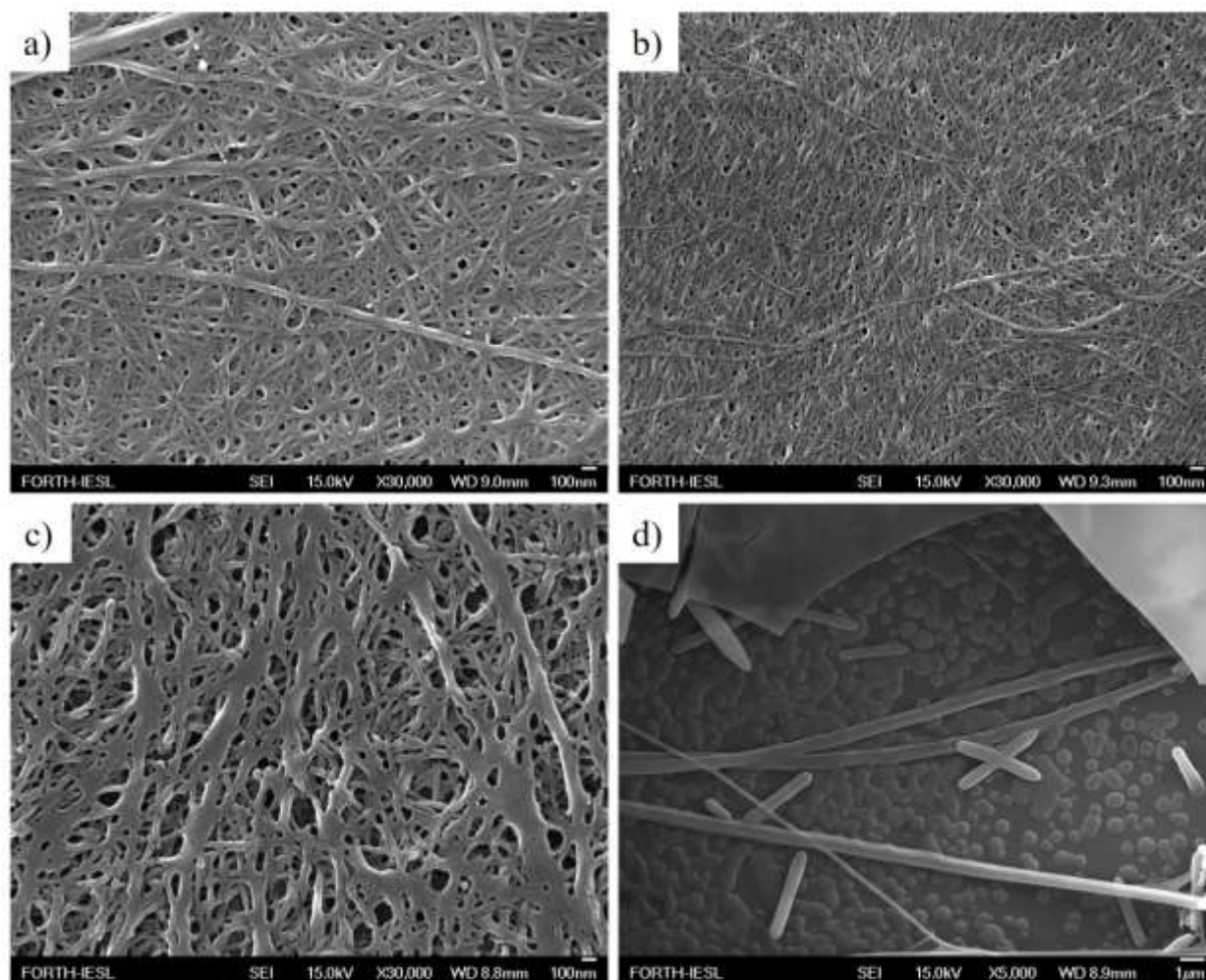
40 **Figure SI-1:** FESEM pictures of bortezomib spheres, following H₂O evaporation after 24h
41 incubation at room temperature, at a concentration of 4 mg/ml, along with an inverted vial showing
42 that the solution flows to the bottom and does not form a self-supporting gel.

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45 In the case of Cyclo-LF, Z-FF and Fmoc-FF with and without Bortezomib the two-solvent
46 ethanol/water (3:7) approach was used. Following ethanol addition, the mixture was vortexed and
47 placed in a water bath thermostat with a temperature of $T=50\text{ }^{\circ}\text{C}$. Sonication was performed for 4
48 seconds with 20 seconds intervals for 15 minutes until complete dissolution was achieved, after
49 which water addition took place to induce self-assembly. When mixed with Bortezomib, it was
50 done in a 1:1 peptide/BTZ ratio. The final concentration in all cases was $c=4\text{ mg/ml}$, although
51 lower concentrations were also tested. The above was also tested with Bortezomib's addition
52 without heating, without observing discernible alterations in the resultant self-assembled
53 structures. Bortezomib on its own was also examined in the EtOH/H₂O 3:7 system, at $c=4\text{mg/ml}$,
54 without any apparent deviation in the spherical conformations it adopted.

55 For field emission scanning electron microscopy (FESEM) observations, once more, 10 μL of the
56 sample was placed on a glass slide and allowed to dry. Subsequently, sputtering was conducted as
57 follows: The samples were affixed to the slide using conductive carbon adhesive tape and
58 subjected to a gold sputtering procedure, depositing a 15 nm thick gold (Au) layer for 64 seconds
59 at 40 mA. FESEM analysis was performed using a JEOL 7000 field emission scanning electron
60 microscope, operating at 15 kV, to examine the structural characteristics of the samples.



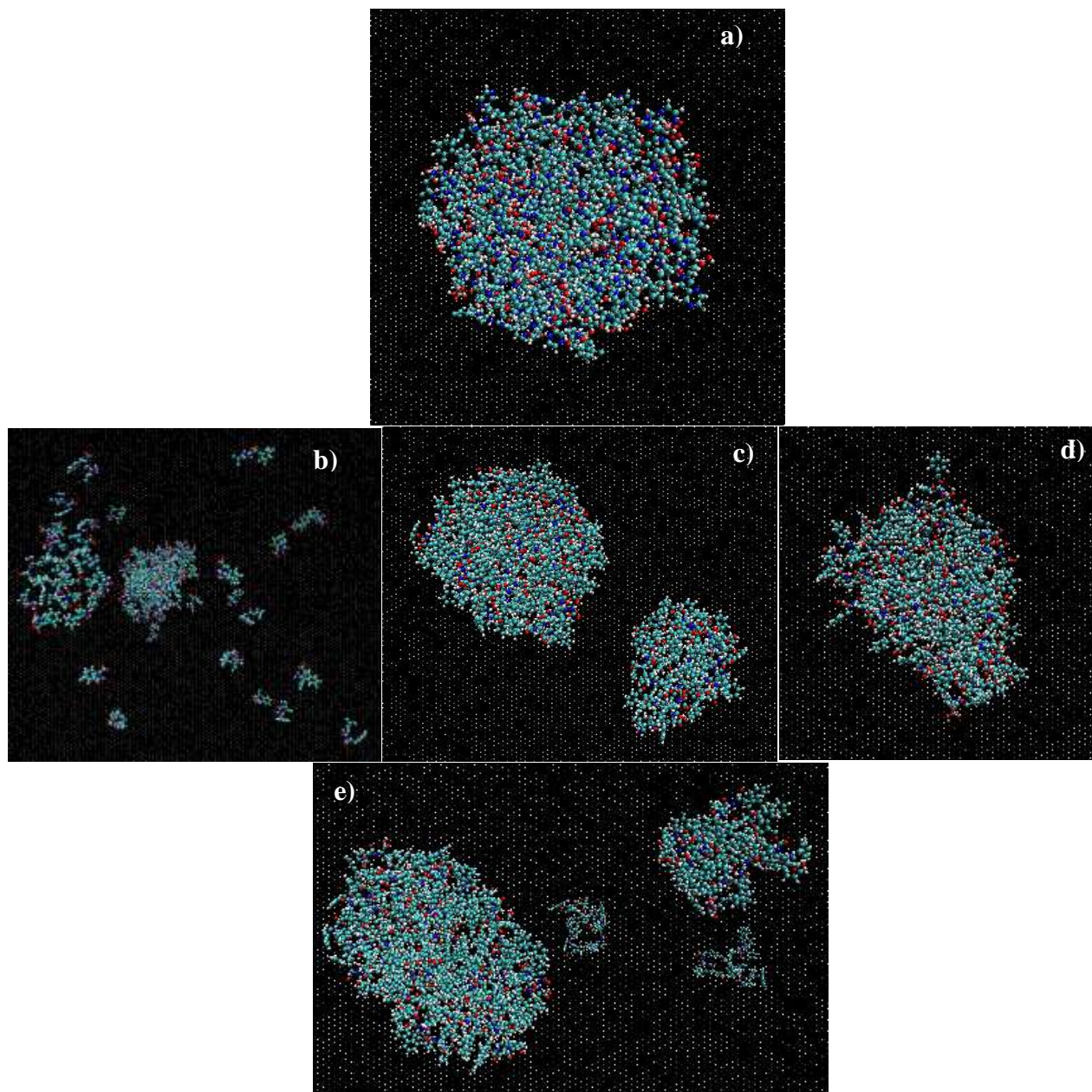
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 62 **Figure SI-2:** FESEM images peptides in (EtOH/H₂O 3:7) with BTZ at a 1:1 ratio at a concentration
 63 of $c = 4$ mg/ml after 24h at room temperature. Scale bars are 1 μ m. a) Z-FF with BTZ b) Fmoc-FF
 64 with BTZ c) Cyclo-LF with BTZ and d) LF with BTZ.

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 66 As depicted in the Field Emission Scanning Electron Microscopy (FESEM) images, it is evident that all
 67 examined peptides, with the exception of LF (Figure SI-2d), exhibit the capacity to yield hydrogels, even
 68 upon the introduction of BTZ (Bortezomib). Among these, Fmoc-FF demonstrates the highest degree of
 69 hydrogel stability in the presence of BTZ (Figure SI-2b), forming intricate and densely interwoven fibril
 70 networks. Z-FF, in conjunction with BTZ (Figure SI-2a), constitutes the second most robust hydrogel,
 71 characterized by substantial fibril networks. In contrast, CycloLF exhibits the lowest hydrogel stability in
 72 the presence of BTZ (Figure SI-2c) among the three peptides, and a notably slower rate of hydrogel
 73 formation.

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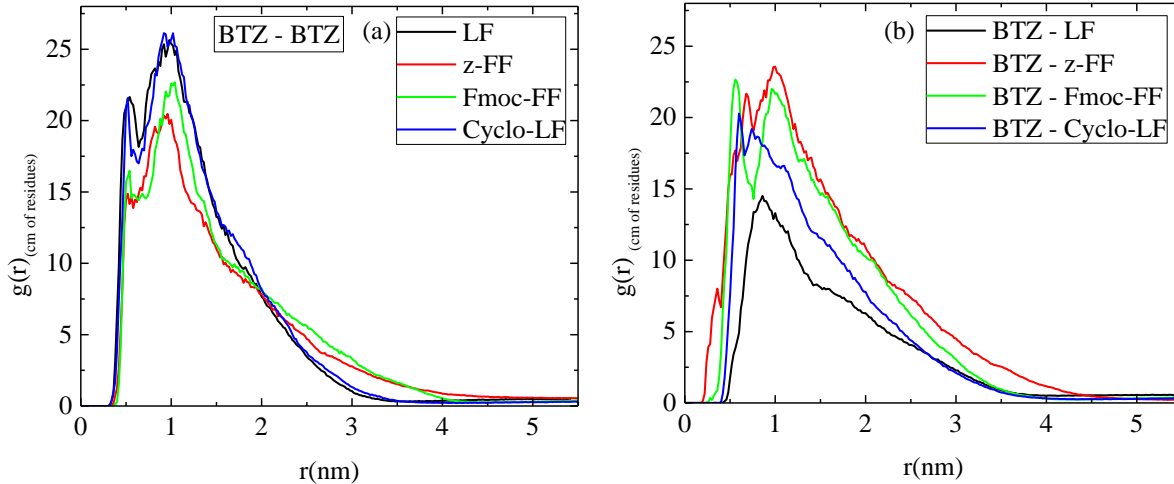
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82 **Figure SI-3:** Characteristic snapshots of clusters (a) BTZ; (b) LF; (c) z-FF; (d) Cyclo-FF; (e)
83 Fmoc-FF. Water molecules are presented as ghost for clarity.

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88 **Figure SI-4:** Pair radial distribution functions between (a) Bortezomib molecules and (b)
89 Bortezomib-dipeptide calculated for the cm of molecules in the aqueous solutions at T=300K.

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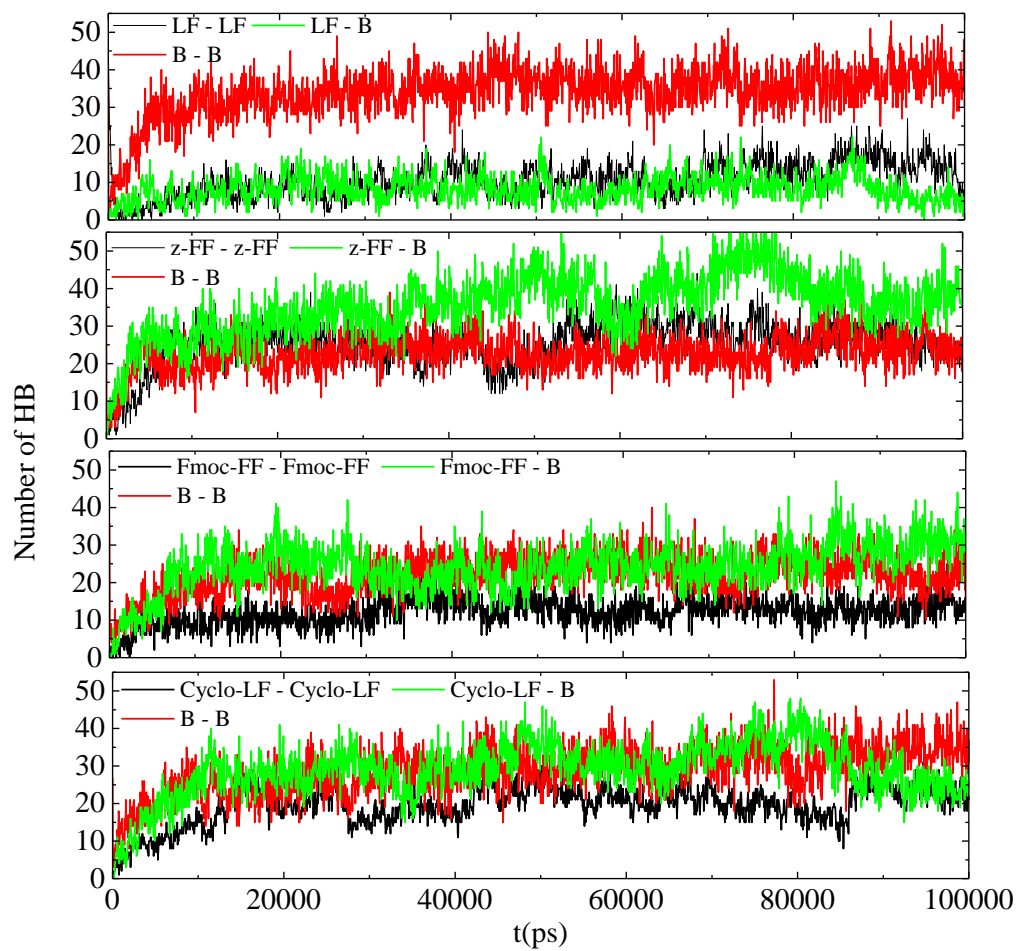
91 **Table SI-1:** Average number of hydrogen bonds between the three groups of hydrogen bonding
92 active sites of BTZ and dipeptide molecules.

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Systems	(OH-DIPEP.)/BTZ	(NH-DIPEP.)/BTZ	(CO-DIPEP.)/BTZ
LF	0.08±0.05	0.02±0.01	0.06±0.03
Cyclo-LF	0.12±0.01	0.29±0.01	0.15±0.01
Fmoc-FF	0.17±0.06	0.35±0.06	0.14±0.05
Z-FF	0.16±0.05	0.33±0.06	0.27±0.05

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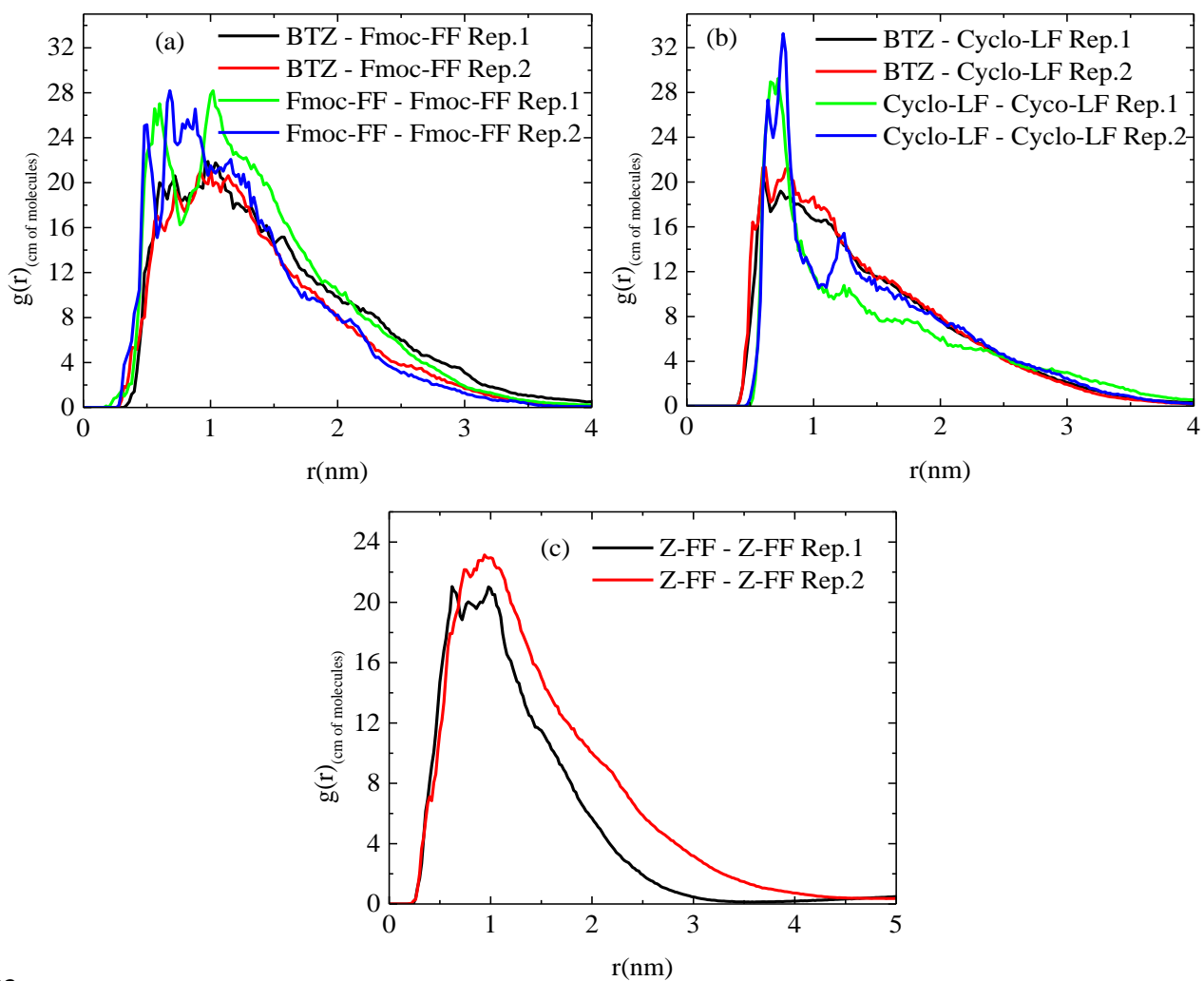
97 **Figure SI-5:** Hydrogen bonds, between the BTZ and dipeptide molecules, as a function of time.

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104 **Figure SI-6:** Pair radial distribution functions between dipeptide molecules and Bortezomib-
 105 dipeptide molecules calculated for the cm of molecules in the aqueous solutions (a) Fmoc-FF with
 106 BTZ; (b) Cyclo-LF with BTZ; (c) Z-FF bulk system.

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