

# SI

## A co-assembly process for a high strength and injectable dual network gel with sustained doxorubicin release performance

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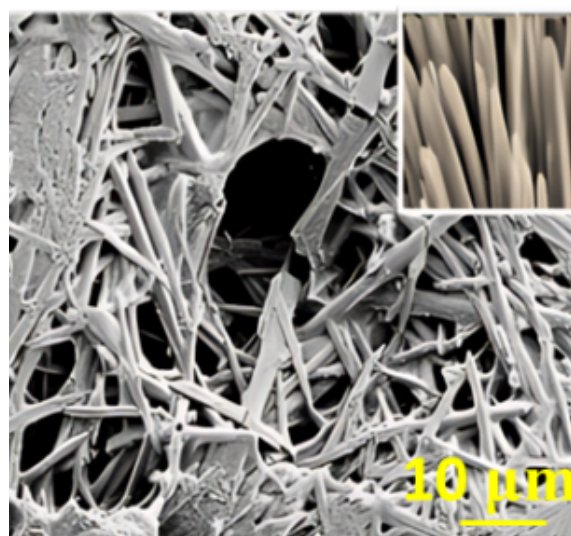
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**Table S1** The collected FTIR peak information of gels

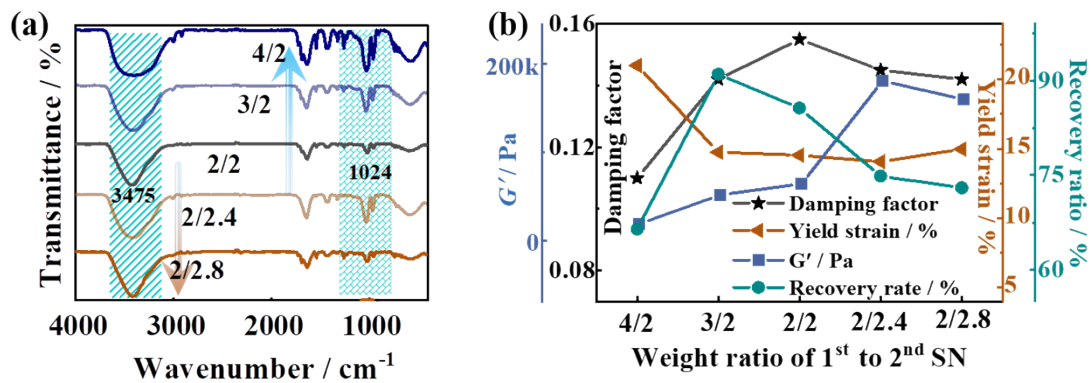
Wavenumber / cm <sup>-1</sup>	Vibration modes
3600	$\nu(-\text{CONH}-)$
3317	$\nu(-\text{COOH}, \text{H-bond})$
3456	$\nu(-\text{OH}, -\text{COOH}, \text{H-bond})$
3063	$\nu(\text{Aryl-H})$
1725	$\nu(-\text{COO-Fmoc}, -\text{CONH}-(\text{Amide I}), -\text{COOH})$
1687	$\gamma(-\text{OH})$
1633	$\nu_{\text{as}}(\text{G-COOH}, \text{M-COOH})$
1610	$\beta\text{-sheet}, \nu_{\text{as}}(-\text{COO}-)$
1538	$\nu(\text{Fmoc}, \text{Bz})$
1450	$\gamma(-\text{CONH}-)$ Amide II
1432	$\nu_{\text{s}}(\text{G-COOH}, \text{M-COOH})$
1395	$\nu(-\text{CONH}-)$ Amide III
1256	$\nu(-\text{OCO-Fmoc})$
1080	$\nu(-\text{CONH}-)$
1024	$\nu(-\text{COOOC}-)$
740	$\beta(\text{Aryl-H})$
563	$\beta(-\text{CH}_2-)$

**Table S2** The collected data of ORCA calculation: the geometry optimization at B97-3C level, the single point energy at RI-B97M-V/def2-TZVP level and the binding energy ( $\Delta E$ ).

Conformation	Single point energy (SPE) / a.u.	$\Delta E$ / (kcal/mol)
<b>Fmoc-F</b>	-1282.960	\
<b>ALG</b>	-1446.194	\
<b>DOX</b>	-1928.674	\
<b>Fmoc-F·Fmoc-F (1)</b>	-2565.940	-12.613
<b>Fmoc-F·Fmoc-F (2)</b>	-2565.943	-14.478
<b>Fmoc-F·H<sub>2</sub>O·Fmoc-F</b>	-2642.395	-28.964
<b>Fmoc-F·2H<sub>2</sub>O·Fmoc-F</b>	-2718.842	-39.275
<b>Fmoc-F·ALG (1)</b>	-2729.175	-13.217
<b>Fmoc-F·ALG (2)</b>	-2729.198	-27.684
<b>ALG·ALG</b>	-2892.429	-25.777
<b>ALG·Ca·ALG</b>	-3570.168	-157.837
<b>Fmoc-F·DOX (1)</b>	-3211.668	-21.847
<b>Fmoc-F·DOX (2)</b>	-3211.669	-22.393
<b>ALG·DOX (1)</b>	-3374.919	-31.661
<b>ALG·DOX (2)</b>	-3374.906	-24.349
<b>DOX·DOX (1)</b>	-3857.393	-28.896
<b>DOX·DOX (2)</b>	-3857.384	-22.750
<b>Fmoc-F·DOX·ALG (1)</b>	-4657.909	-50.982
<b>Fmoc-F·DOX·ALG (2)</b>	-4657.912	-53.021



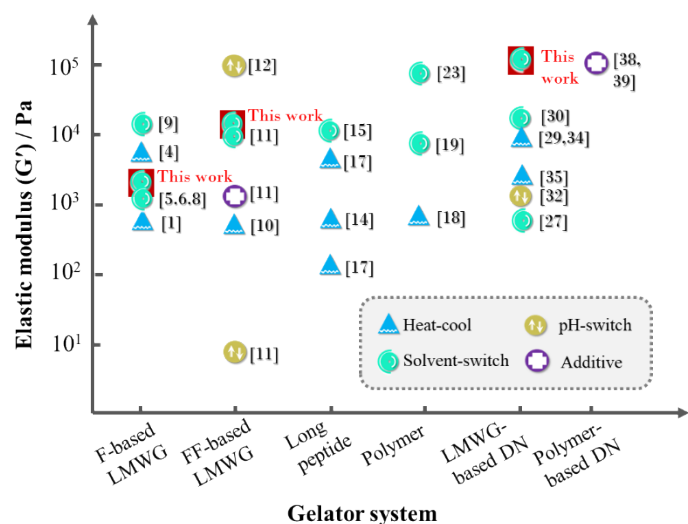
**Figure S1** The network morphology overview of 1<sup>st</sup>SN



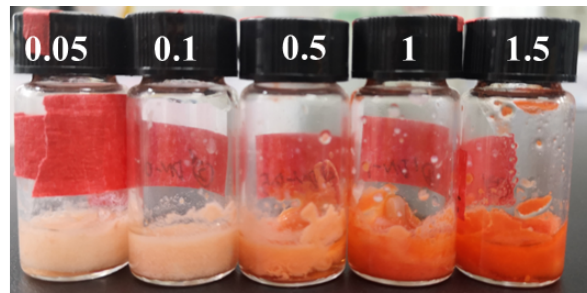
**Figure S2** The dependency of DN gel properties on 1<sup>st</sup>SN/2<sup>nd</sup>SN ratio: (a) FTIR spectra and (b) rheological parameter's comparison.

**Table S3** The collected rheological information of SN and DN gels.

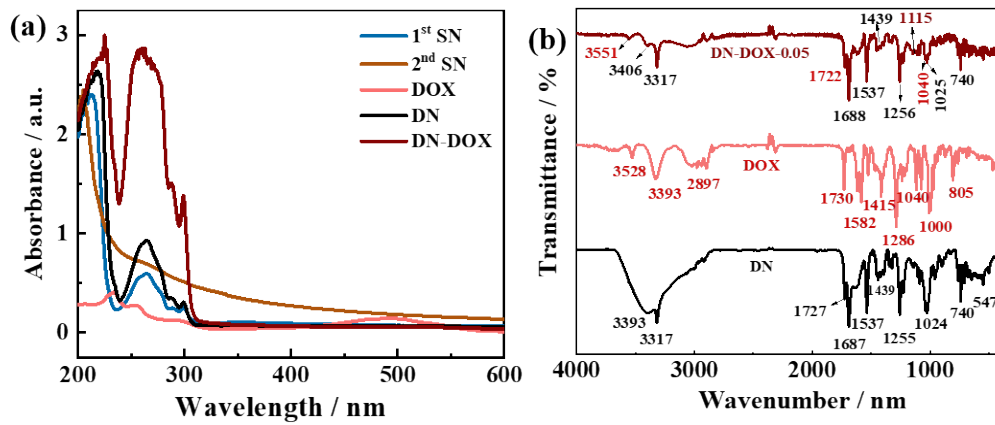
The weight ratio of 1 <sup>st</sup> SN to 2 <sup>nd</sup> SN	Strain sweep				3ITT
	$G' / \text{Pa}$	$G'' / \text{Pa}$	Yield strain / %	$\tan\delta$	Recover ratio within 100 s / %
4/2	18700	2060	21.00	0.110	66.46
3/2	51700	7330	14.75	0.142	91.01
2/2	64400	9960	14.53	0.155	85.63
2/2.4	181000	26200	14.07	0.145	74.87
2/2.8	160000	22700	14.97	0.142	73.01



**Figure S3** The comparison of elastic modulus of gels in this work and other types of gel materials.



**Figure S4** The optical pictures of DN-DOX-X gels with different DOX amounts (X=0.05, 0.1, 0.5, 1 and 1.5).



**Figure S5** The structural change before and after DOX loading: (a) the UV-vis spectra, (b) FTIR spectra.

**Table S4** The collecting rheological data of DN-DOX gels.

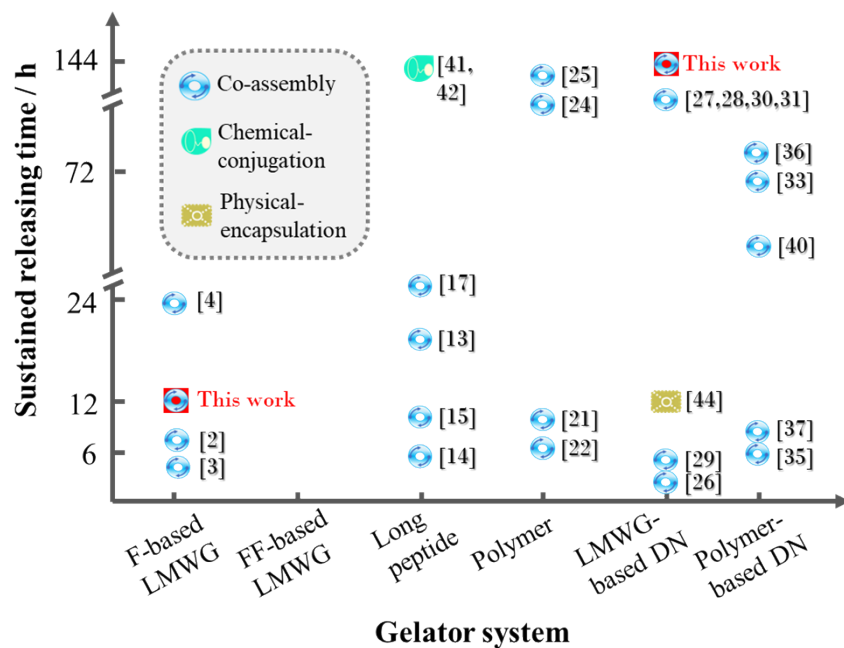
Sample	Strain sweep		3ITT	
	$G'$ / Pa	$G''$ / Pa	Yield strain / %	Recover ratio / %
DN-DOX-0.05	35600	6420	8.46	84.10
DN-DOX-0.1	57400	7190	14.29	89.60
DN-DOX-0.5	108000	13300	17.68	84.88
DN-DOX-1	60100	8350	16.88	88.53
DN-DOX-1.5	51800	8210	21.45	87.55

**Table S5** The DOX loading efficiency, encapsulation efficiency and releasing in DN-DOX gels.

Drug delivery efficiency		1 <sup>st</sup> SN-DOX-0.05	2 <sup>nd</sup> SN-DOX-0.05	DN-DOX-x					
				0.05	0.1	0.5	1	1.5	
Loading capacity	$E_l / \%$	0.10	0.07	0.12	0.24	1.20	2.22	3.26	
	$E_e / \%$	77.06	59.11	96.71	95.48	97.47	90.65	89.77	
Releasing profile	$E_r$ at 30 min / %	16.57	11.65	2.34	3.35	5.48	11.88	10.00	
	$E_r$ at 24 h / %	/	/	52.01	37.89	30.31	44.71	50.45	
	t for $E_r > 50\% / h$	2	2	24	30	36	32	24	
	t for $E_r > 90\% / h$	10	10	57	82	144	168	72	
	Whole range	$a$	36.04	35.47	12.28	11.16	9.94	18.40	26.58
Fitting result of Er on a Rigter-Peppas model	$n$	<b>0.48</b>	<b>0.41</b>	<b>0.48</b>	<b>0.47</b>	<b>0.46</b>	<b>0.33</b>	<b>0.27</b>	
	$R^2$	0.97	0.90	0.98	0.96	0.95	0.96	0.91	
	0~24h	$a_1$			11.46	9.25	8.18	20.11	21.42
		$n_1$		\	<b>0.49</b>	<b>0.45</b>	<b>0.41</b>	<b>0.26</b>	<b>0.29</b>
		$R_1^2$			0.97	0.96	0.99	0.98	0.96
	24~48h	$a_2$			6.07	4.29	1.61	5.27	7.88
		$n_2$			<b>0.67</b>	<b>0.74</b>	<b>0.96</b>	<b>0.56</b>	<b>0.62</b>
		$R_2^2$			0.96	0.97	0.98	0.99	0.90
	48~End	$a_3$			42.60	34.19	23.34	38.25	69.58
$n_2$				<b>0.19</b>	<b>0.22</b>	<b>0.28</b>	<b>0.17</b>	<b>0.06</b>	
$R_3^2$				0.72	0.82	0.97	0.88	0.84	

**Table S6** The release kinetics fitting by other models

Sample	Zero-order	First-order	Higuchi
1 <sup>st</sup> SN-DOX-0.05	y=20.63+9.26x, R <sup>2</sup> =0.83	y=98.68 $\beta$ (1-e <sup>-0.35x</sup> ), R <sup>2</sup> =0.99	y=33.27 $\beta$ x <sup>1/2</sup> -0.03, R <sup>2</sup> =0.97
2 <sup>nd</sup> SN-DOX-0.05	y=26.51+1.09x, R <sup>2</sup> =0.68	y=94.44 $\beta$ (1-e <sup>-0.36x</sup> ), R <sup>2</sup> =0.99	y=27.72 $\beta$ x <sup>1/2</sup> +4.16, R <sup>2</sup> =0.89
0.05	y=21.61+1.09x, R <sup>2</sup> =0.87	y=103.09 $\beta$ (1-e <sup>-0.03x</sup> ), R <sup>2</sup> =0.97	y=11.33 $\beta$ x <sup>1/2</sup> +0.08, R <sup>2</sup> =0.97
	y=24.20+0.77x, R <sup>2</sup> =0.79	y=102.77 $\beta$ (1-e <sup>-0.03x</sup> ), R <sup>2</sup> =0.98	y=9.82 $\beta$ x <sup>1/2</sup> -0.05, R <sup>2</sup> =0.95
DN-DOX-x	y=24.84+0.53x, R <sup>2</sup> =0.77	y=98.36 $\beta$ (1-e <sup>-0.02x</sup> ), R <sup>2</sup> =0.98	y=8.10 $\beta$ x <sup>1/2</sup> +1.17, R <sup>2</sup> =0.94
	y=35.65+0.44x, R <sup>2</sup> =0.71	y=90.02 $\beta$ (1-e <sup>-0.03x</sup> ), R <sup>2</sup> =0.87	y=6.82 $\beta$ x <sup>1/2</sup> +15.17, R <sup>2</sup> =0.92
1.5	y=46.32+0.41x, R <sup>2</sup> =0.52	y=90.91 $\beta$ (1-e <sup>-0.05x</sup> ), R <sup>2</sup> =0.91	y=7.04 $\beta$ x <sup>1/2</sup> +23.33, R <sup>2</sup> =0.81



**Figure S6** The comparison of sustained releasing time of gels in this work and other types of gel materials.

**Reference:**

- [1] Murali DM, Shanmugam G. The aromaticity of the phenyl ring imparts thermal stability to a supramolecular hydrogel obtained from low molecular mass compound. *New J Chem*, 2019, 43 (31): 12396-12409.
- [2] Gahane AY, Ranjan P, Singh V, et al. Fmoc-phenylalanine displays antibacterial activity against Gram-positive bacteria in gel and solution phases. *Soft Matt*, 2018, 14 (12): 2234-2244.
- [3] Singh V, Snigdha K, Singh C, et al. Understanding the self-assembly of Fmoc-phenylalanine to hydrogel formation. *Soft Matt*, 2015, 11 (26): 5353-5364.
- [4] Snigdha K, Singh BK, Mehta AS, et al. Self-assembling N -(9-Fluorenylmethoxycarbonyl)-l-Phenylalanine hydrogel as novel drug carrier. *Int J Biol Macromol*, 2016, 93: 1639-1646.
- [5] Abraham JN, Joseph S, Trivedi R, et al. Injectable dextran-fluorenylmethoxycarbonyl phenylalanine composite hydrogels with improved mechanical properties. *Polym Int*, 2021, 70 (2): 222-229.
- [6] Ryan DM, Anderson SB, Nilsson BL. The influence of side-chain halogenation on the self-assembly and hydrogelation of Fmoc-phenylalanine derivatives. *Soft Matt*, 2010, 6 (14): 3220.
- [7] Ryan DM, Doran TM, Nilsson BL. Stabilizing self-assembled Fmoc-F5-Phe hydrogels by co-assembly with PEG-functionalized monomers. *Chem Commun (Camb)*, 2011, 47 (1): 475-477.
- [8] Hashemnejad SM, Huda MM, Rai N, et al. Molecular Insights into Gelation of Di-Fmoc-l-Lysine in Organic Solvent-Water Mixtures. *ACS Omega*, 2017, 2 (5): 1864-1874.
- [9] Zhang Y, Li S, Ma M, et al. Tuning of gel morphology with supramolecular chirality amplification using a solvent strategy based on an Fmoc-amino acid building block. *New J Chem*, 2016, 40 (6): 5568-5576.

- [10] Li J, Kuang Y, Shi J, et al. The conjugation of nonsteroidal anti-inflammatory drugs (NSAID) to small peptides for generating multifunctional supramolecular nanofibers/hydrogels. *Beilstein J Org Chem*, 2013, 9: 908-917.
- [11] Raeburn J, Zamith Cardoso A, Adams DJ. The importance of the self-assembly process to control mechanical properties of low molecular weight hydrogels. *Chem Soc Rev*, 2013, 42 (12): 5143-5156.
- [12] Najafi H, Abolmaali SS, Heidari R, et al. Nitric oxide releasing nanofibrous Fmoc-dipeptide hydrogels for amelioration of renal ischemia/reperfusion injury. *J Control Release*, 2021, 337: 1-13.
- [13] Howe EJ, Okesola BO, Smith DK. Self-assembled sorbitol-derived supramolecular hydrogels for the controlled encapsulation and release of active pharmaceutical ingredients. *Chem Commun (Camb)*, 2015, 51 (35): 7451-7454.
- [14] Yu A, Hu Y, Ma X, et al. Sequential drug release of co-assembled supramolecular hydrogel as synergistic therapy against *Staphylococcus aureus* endophthalmitis. *Chem Eng J*, 2022, 427: 130979.
- [15] Valls A, Isabel Burguete M, Kuret L, et al. Open chain pseudopeptides as hydrogelators with reversible and dynamic responsiveness to pH, temperature and sonication as vehicles for controlled drug delivery. *J Mol Liq*, 2021, 348: 118051.
- [16] Dadfar SMR, Pourmahdian S, Tehrani MM, et al. Novel dual-responsive semi-interpenetrating polymer network hydrogels for controlled release of anticancer drugs. *J Biomed Mater Res A*, 2019, 107 (10): 2327-2339.
- [17] Tao N, Li G, Liu M, et al. Preparation of dual responsive low-molecular-weight hydrogel for long-lasting drug delivery. *Tetrahedron*, 2017, 73 (22): 3173-3180.
- [18] Mo G, Zhang R, Wang Y, et al. Rheological and optical investigation of the gelation with and without phase separation in PAN/DMSO/H<sub>2</sub>O ternary blends. *Polymer*, 2016, 84: 243-253.
- [19] Appaw C, Gilbert RD, Khan SA, et al. Viscoelastic behavior of cellulose acetate in a mixed solvent system. *Biomacromolecules*, 2007, 8 (5): 1541-1547.
- [20] Gonzalez MV, Tang Y, Phillips GJ, et al. Doxorubicin eluting beads-2: methods for evaluating drug elution and in-vitro:in-vivo correlation. *J Mater Sci Mater Med*, 2008, 19 (2): 767-775.
- [21] Mao J, Qiu L, Ge L, et al. Overcoming multidrug resistance by intracellular drug release and inhibiting p-glycoprotein efflux in breast cancer. *Biomed Pharmacother*, 2021, 134: 111108.
- [22] Maciel D, Figueira P, Xiao S, et al. Redox-responsive alginate nanogels with enhanced anticancer cytotoxicity. *Biomacromolecules*, 2013, 14 (9): 3140-3146.
- [23] Ma M, Feng Z, Zhao M, et al. Fabrication of macrocyclic organogel utilizing solvent balance and its application in vascular supporting materials. *Colloid Surf*, 2020, 589: 124432.
- [24] Zhou Q, Li C, Guo J, et al. Self-assembled biocompatible heparin-based supramolecular hydrogel for doxorubicin delivery. *Carbohydr Res*, 2022, 511: 108464.
- [25] Wang M, Chen M, Niu W, et al. Injectable biodegradation-visual self-healing citrate hydrogel with high tissue penetration for microenvironment-responsive degradation and local tumor therapy. *Biomaterials*, 2020, 261: 120301.
- [26] Dou XQ, Zhao CL, Mehwish N, et al. Photoresponsive supramolecular hydrogel co-assembled from Fmoc-Phe-OH and 4,4'-azopyridine for controllable dye release. *Chinese J Polym Sci*, 2019, 37 (5): 437-443.
- [27] Chen J, Tao N, Fang S, et al. Incorporation of Fmoc-Y nanofibers into Ca-alginate hydrogels

- for improving their mechanical properties and the controlled release of small molecules. *New J Chem*, 2018, 42 (12): 9651-9657.
- [28] Shulman LP. Effectiveness of maternal influenza immunization in mothers and infants. *yearbook of obstetrics, gynecology and women's health. N Engl J Med*, 2009, 359: 115-117.
- [29] Grijalvo S, Puras G, Zárate J, et al. Nioplexes encapsulated in supramolecular hybrid biohydrogels as versatile delivery platforms for nucleic acids. *RSC Adv*, 2016, 6 (46): 39688-39699.
- [30] Huang R, Qi W, Feng L, et al. Self-assembling peptide–polysaccharide hybrid hydrogel as a potential carrier for drug delivery. *Soft Matt*, 2011, 7 (13): 6222.
- [31] Gallo E, Diaferia C, Rosa E, et al. Peptide-based hydrogels and nanogels for delivery of doxorubicin. *Int J Nanomedicine*, 2021, 16: 1617-1630.
- [32] Giuri D, Barbalinardo M, Zanna N, et al. Tuning mechanical properties of pseudopeptide supramolecular hydrogels by graphene doping. *Molecules*, 2019, 24 (23): 4345.
- [33] Bai J, Liu Y, Jiang X. Multifunctional PEG-GO/CuS nanocomposites for near-infrared chemophotothermal therapy. *Biomaterials*, 2014, 35 (22): 5805-5813.
- [34] Piras CC, Mahon CS, Smith DK. Self-assembled supramolecular hybrid hydrogel beads loaded with silver nanoparticles for antimicrobial applications. *Chem A Europ J*, 2020, 26 (38): 8452-8457.
- [35] Patterson AK, Smith DK. Two-component supramolecular hydrogel for controlled drug release. *Chem Commun (Camb)*, 2020, 56 (75): 11046-11049.
- [36] Vieira VMP, Hay LL, Smith DK. Multi-component hybrid hydrogels-understanding the extent of orthogonal assembly and its impact on controlled release. *Chem Sci*, 2017, 8 (10): 6981-6990.
- [37] Nanda J, Biswas A, Banerjee A. Single amino acid based thixotropic hydrogel formation and pH-dependent morphological change of gel nanofibers. *Soft Matt*, 2013, 9 (16): 4198.
- [38] Sun JY, Zhao X, Illeperuma WR, et al. Highly stretchable and tough hydrogels. *Nature*, 2012, 489 (7414): 133-136.
- [39] Chen Q, Zhu L, Chen H, et al. A novel design strategy for fully physically linked double network hydrogels with tough, fatigue resistant, and self-healing properties. *Adv Func Mater*, 2015, 25 (10): 1598-1607.
- [40] Zhang R, Xing R, Jiao T, et al. Carrier-free, chemophotodynamic dual nanodrugs via self-assembly for synergistic antitumor therapy. *ACS Appl Mater Interfaces*, 2016, 8 (21): 13262-13269.
- [41] Zhao D, Zhao X, Zu Y, et al. Preparation, characterization, and in vitro targeted delivery of folate-decorated paclitaxel-loaded bovine serum albumin nanoparticles. *Int J Nanomedicine*, 2010, 5: 669-677.
- [42] Yue Y, Liu ZW. Conjugating doxorubicin to supramolecular organic frameworks: polymeric prodrugs with enhanced therapeutic efficacy and safety. *J Mater Chem B*, 2022, 10(22): 4163-4171.
- [43] Gao Y, Chen Y, Ji X, et al. Controlled intracellular release of Doxorubicin in multidrug-resistant cancer cells by tuning the shell-pore sizes of mesoporous silica nanoparticles. *ACS Nano*, 2011, 5 (12): 9788-9798.
- [44] Ye W, Zhang G, Liu X, et al. Fabrication of polysaccharide-stabilized zein nanoparticles by flash nanoprecipitation for doxorubicin sustained release. 2022, 70: 103183.