

Electronic Supplementary Information (ESI):

Promotion of Photocatalytic Hydrogen Production by Utilization of Triplet Excited States of Organic Dyes and Adjustment of π - π Interactions

Siwei Liu, Qian Chen, YanTing Chen, Peixuan Lin, Hangyu Zhuzhang, Mengmeng Han, Zhi-An Lan*, Xiong Chen, Xinchun Wang*, Qianqian Li*, Zhen Li*

Content

Structural characterization of organic dyes.....	7
Scheme S1. Synthetic routes of organic dyes LI-154 ~ LI-158 and LI-161 ~ LI-165.....	7
Figure S1. ¹H NMR spectrum of LI-154.....	15
Figure S2. ¹³C NMR spectrum of LI-154.	16
Figure S3. ¹H NMR spectrum of LI-155.....	16
Figure S4. ¹³C NMR spectrum of LI-155.	17
Figure S5. ¹H NMR spectrum of LI-156.....	17
Figure S6. ¹³C NMR spectrum of LI-156.	18
Figure S7. ¹H NMR spectrum of LI-157.....	18
Figure S8. ¹³C NMR spectrum of LI-157.	19
Figure S9. ¹H NMR spectrum of LI-158.....	19
Figure S10. ¹³C NMR spectrum of LI-158.	20
Figure S11. ¹H NMR spectrum of LI-161.....	20
Figure S12. ¹³C NMR spectrum of LI-161.	21
Figure S13. ¹H NMR spectrum of LI-162.....	21
Figure S14. ¹³C NMR spectrum of LI-162.	22
Figure S15. ¹H NMR spectrum of LI-163.....	22
Figure S16. ¹³C NMR spectrum of LI-163.	23
Figure S17. ¹H NMR spectrum of LI-164.....	23
Figure S18. ¹³C NMR spectrum of LI-164.	24
Figure S19. ¹H NMR spectrum of LI-165.....	24
Figure S20. ¹³C NMR spectrum of LI-165.	25

Figure S21. MALDI-TOF spectrum of dyes.....	26-27
Figure S22. FTIR spectra of organic dyes in the range of 4000-400 cm^{-1}	27
Photophysical property of organic dyes	28
Figure S23. The UV absorption spectra of LI-154 (a), LI-155 (c), LI-156 (e), LI-157 (g), LI-158 (i), LI-161 (k), LI-162 (m), LI-163 (o), LI-164 (q) and LI-165 (s) with different concentrations in CH_2Cl_2 solution. The molar extinction coefficient (504 nm) fitting curve of LI-154 (b), the molar extinction coefficient (490 nm) fitting curve of dyes (d, f, h, j, l, n, p, r, t).	28
Figure S24. Excitation spectra of dyes in CH_2Cl_2 solution at the concentration of $5 \times 10^{-5} \text{ mol L}^{-1}$	29
Figure S25. Photoluminescence spectra of dyes in CH_2Cl_2 solution with different concentrations.	30
Figure S26. Photoluminescence spectra of dyes in 2,5-Dimethyltetrahydrofuran solution at 293K(a) and 77 K(b)	31
Figure S27. Phosphorescence spectra of dyes in 2,5-dimethyltetrahydrofuran solution at 77 K.....	31
Figure S28. (a) UV-vis DRS absorption spectra of LI-154 , LI-155 , LI-156 , LI-157 , LI-158 , LI-161 , LI-162 , LI-163 at solid state. Due to LI-164 and LI-165 oily nature, a thin film UV-vis is used instead. (b, c) the bandgaps of dyes calculated by Tauc-plot.	32
Figure S29. Optimized molecular configurations and frontier orbital distribution of dyes.....	33-34
Table S1 Fluorescence lifetimes of organic dyes.	34
Figure S30. Photographs of organic dyes in thin film under different conditions captured by a camera, and afterglow imaging captured by a chemiluminescence imager (colors only represent phosphorescence intensity) at different time after UV off.	35
Figure S31 The phosphorescence emission wavelengths of organic dyes.	35
Figure S32 Phosphorescence decay of dyes in thin film.	36
Table S2 Phosphorescence lifetimes of organic dyes in thin film.....	36
Characterization of PCN, PCN/Pt and dye/PCN/Pt	37
Figure S33. FTIR spectra of PCN, PCN/Pt and dye/PCN/Pt.	37
Figure S34. XRD patterns of PCN, PCN/Pt and dye/PCN/Pt.	37
Figure S35. UV-vis DRS absorption spectra of dyes, PCN, PCN/Pt and dye/PCN/Pt	38
Figure S36. a) UPS spectra of PCN, HOMO energy levels are calculated according to the following equation: E_{HOMO} (vs. vacuum conditions) = $-(E_{\text{photon}} - \text{width of UPS spectra})$, where E_{photon} denotes the excitation energy of He I (21.22 eV).(b)The bandgaps of PCN calculated by Tauc-plot.....	38
Figure S37. UPS spectra of dyes, HOMO energy levels are calculated according to the following equation: E_{HOMO} (vs. vacuum conditions) = $-(E_{\text{photon}} - \text{width of UPS spectra})$, where E_{photon} denotes the excitation energy of He I (21.22 eV).	39

Figure S38. High-resolution XPS spectra of PCN/Pt and dye/PCN/Pt.	40
Table S3. Relative ratios of two carbon species determined by C 1s spectra of PCN/Pt and dye/PCN/Pt.....	41
Table S4. Relative ratios of four nitrogen species determined by N 1s spectra of PCN/Pt and dye/PCN/Pt.....	42
Figure S39 SEM and TEM images of PCN and PCN/Pt. The Pt morphology on the surface of PCN.	42
Figure S40. SEM and TEM images of dye/PCN/Pt.	43
Figure S41. VB XPS spectra of PCN/Pt and dye/PCN/Pt.....	44
Figure S42. The water contact angles of dye/PCN/Pt electrodes.	44
Figure S43. Electrochemical impedance spectra (EIS) Nyquist plots of dye/PCN/Pt (-1.2 V vs. RHE).	45
Figure S44. The transient open-circuit voltage decay (OCVD) of dye/PCN/Pt.....	45
Figure S45. PL spectra of PCN, PCN/Pt and dye/PCN/Pt TRPL spectra of PCN, PCN/Pt, dye/PCN and dye/PCN/Pt.....	46
Table S5. Fluorescence lifetimes of PCN, PCN/Pt and dye/PCN/Pt.....	46
Figure S46. PL decay of PCN and PCN/Pt.	47
Figure S47. PL spectra of dye, PCN/Pt and dye/PCN/Pt.	48
Figure S48. PL decay of dye/PCN/Pt.	48
Table S6. Fluorescence lifetimes of dye/PCN/Pt..	49
Figure S49. The calculated work function of LI-161 (a), PCN (b) and PCN/Pt (c) by UPS.	49
Figure S50. PL spectra of dyes (thin flim) in H ₂ O (a) and 10% TEOA; TRPL spectra of dyes (thin flim) in H ₂ O (a) and 10% TEOA.....	49
Figure S51. PL spectra of organic dyes (thin film) in water and 10% TEOA aqueous solution.....	50
Figure S52. TRPL spectra of organic dyes (thin film) in water and 10% TEOA aqueous solution..	50
Figure S53. The effective lengths of alkyl chains substituted to thiophene moiety for isolation effect... ..	51

Additional data and spectra

Materials

Tetrahydrofuran (THF) was dried over and distilled from a K–Na alloy under an atmosphere of dry argon. All solvents were of analytical grade and used without further purification. Urea, bis(di-tert-butyl(4-dimethylaminophenyl)phosphine)dichloro palladium(II)(Pd(amphos)Cl₂), tri-*n*-butyltinchloride, 4,7-dibromo-5,6-bis(dodecyloxy)benzo[*c*][1,2,5]thiadiazole, *N*-bromosuccinimide (NBS), *N,N*-diphenyl-4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline, 2-(tributylstannyl)thiophene, 3-ethylthiophene, 3-butylthiophene, 3-hexylthiophene, 3-octylthiophene, 3-(2-ethylhexyl)thiophene, 3-(2-butyloctyl)thiophene, 3-(2-hexyldecyl)thiophene, 3-(2-octyldodecyl)thiophene, 3-(2-decyltetradecyl)thiophene and 4,7-dibromo-5,6-bis(dodecyloxy)benzo[*c*][1,2,5]thiadiazole, were purchased and used directly as received.

Instruments

¹H and ¹³C NMR spectra were recorded on a Bruker Avance III HD 400 MHz using tetramethylsilane (TMS; $\delta = 0$ ppm) as internal standard. Elemental analyses were performed by a Perkin-Elmer microanalyzer. MALDI-TOF spectra were measured on an AB SCIEX MALDI-TOF/TOF 5800. UV-vis spectra were conducted on a Shimadzu UV-2550 spectrometer. Photoluminescence and phosphorescence spectra in solution at 77 K were performed on a Hitachi F-4700 fluorescence spectrophotometer. Photoluminescence spectra and fluorescence lifetimes, and phosphorescence spectra at solid state were determined by a FLS980 spectrometer. Phosphorescence lifetimes were determined with PicoQuant FluoTime 300. The Fourier transform infrared (FTIR) spectra were obtained on Thermo Nicolet IS 10 FT-IR spectrometer with KBr as the diluents in the frequency range of 4000–400 cm⁻¹. UV–Vis diffuse reflectance spectra (UV–Vis DRS) were performed on UV-3600 Ultraviolet-Visible-Near Infrared Spectrophotometer with MPC-3100 Multi-Purpose Large-Sample Compartment. UPS spectra were obtained by ThermoFisher ESCALAB XI+. RTP photographs of organic luminogens at solid state were captured with a Tanon 5200 multi-chemiluminescence imager (cooling temperature of Charge-coupled Device lens is -40 °C)

DFT calculations

DFT calculations were performed on Gaussian 09 program (Revision D09). The ground state (S₀) geometries were optimized with the Becke's three-parameter exchange functional along with the Lee Yang Parr's correlation functional (B3LYP) using 6–31 G(d) basis sets. To simulate the isolation effect of alkyl chains, the lengths of alkyl

chains perpendicular to thiophene planes in the default state were measured by CHEM 3D as efficient lengths.

Synthesis of polymeric carbon nitride (PCN) and preparation of PCN/Pt, dye/PCN and dye/PCN/Pt

In a typical synthesis, 10 g of urea was calcined at 550 °C in air for 2 h at the heating rate of 10 °C min⁻¹. The obtained solid (PCN) was grounded into powder with an agate mortar. Then, PCN powder (500 mg) and H₂PtCl₆·6H₂O (40 mg) was added into a mixture methanol aqueous solution (120 mL, v/v = 5:1) in a beaker, and the solution was irradiated by a Xe lamp (300 W) for 3h, 3.0 wt.% PCN/Pt was obtained by filtration and washed 3 times by water. PCN/Pt (100 mg) and dyes (2 mg) were immersed in a CHCl₃ solution (10 mL) and kept stirring at room-temperature in the dark for 3 hours, then the solvent was removed by rotary evaporation to give dyes/PCN/Pt.

Characterization of PCN, PCN/Pt, dye/PCN and dye/PCN/Pt

The powder X-ray diffraction patterns were recorded by Rigaku MiniFlex 600 using Cu-K α radiation from 5° to 80°. Transmission electron microscopy (TEM) were obtained in JEM-F200 transmission electron microscope. X-ray photoelectron spectroscopy (XPS) data were obtained on Thermo Fisher Nexsa instrument with a monochromatized Al K α line source. UV–Vis diffuse reflectance spectra (UV–Vis DRS) were performed on Perkinelmer LAMBDA 850 UV/Vis Spectrophotometer. The Fourier transform infrared (FTIR) spectra Thermo Nicolet IS 50 FT-IR spectrometer with KBr as the diluents in the frequency range of 4000-400 cm⁻¹. Fluorescence spectra and lifetimes were determined with FLS980 spectrometer. Water contact angle were performed at POWERREACH JC2000D contact angle measuring instrument.

Photoelectrochemical Measurements

Photocurrent density curves and Electrochemical Impedance Spectroscopy (EIS) were obtained in a conventional three electrode cell by ModuLab XM ECS, using a Pt plate as the counter electrode and a Ag/AgCl (3 M KCl) electrode as the reference electrode. The working electrode was prepared on indium-tin oxide (ITO) glass, that was cleaned by sonication in ethanol and acetone for 30 min and dried at 353 K. The boundary of ITO glass was protected using Scotch tape. The 5 mg of sample (PCN, PCN/Pt, dye/PCN/Pt) was dispersed in a 0.5 mL solution (480 μ L of absolute ethanol and 20 μ L of Nafion solution) by sonication to get a slurry. The slurry was spread onto pretreated ITO glass. After air-drying, the Scotch tape was unstuck, and the uncoated part of the electrode was isolated with epoxy resin to get a 1 cm \times 1 cm film electrode. A Na₂SO₄ solution (0.5 M) was used as the electrolyte. Before testing, the electrolyte was purged with N₂ to remove dissolved oxygen. EIS was determined over the frequency

range of 10^{-2} - 10^6 Hz with an ac amplitude of 10 mV at the open circuit voltage and -1.2 V (vs. RHE), respectively.

Photocatalytic test

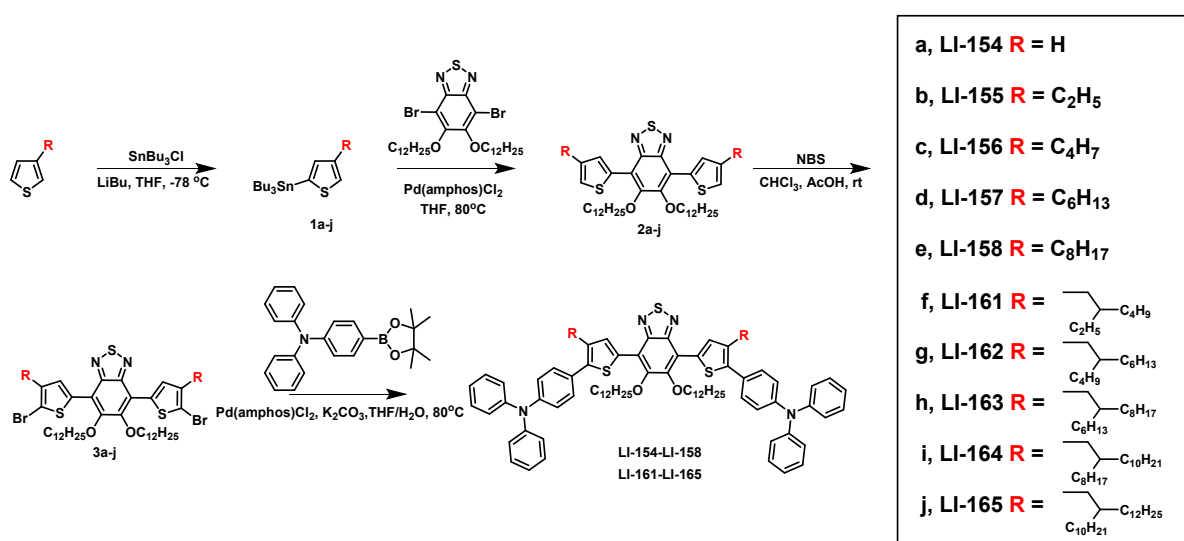
Photocatalytic hydrogen evolution arrays were performed in a Pyrex top-irradiation reaction vessel linked to a glass closed gas system. H₂ production was carried out by dispersing 50 mg of photocatalyst powder in an aqueous solution (100 mL) containing triethanolamine (10 vol.%) as sacrificial electron donor. The mixture was evacuated several times to remove air completely before irradiation under a 300 W Xe-lamp and a water-cooling filter. The mixture was kept at room temperature by a flow of cooling water. The generated gases were analyzed by gas chromatography equipped with a thermal conductive detector (TCD) with Argon as the carrier gas.

The apparent quantum yield (AQY) for H₂ evolution was acquired with monochromatic LED lamps with a band-pass filter of 420, 450, 490 and 520 nm. The intensities were 17.1, 13.2, 13.0 and 8.5 mW cm⁻², respectively (Newport 2936-R optical power meter). The irradiation area was 9 cm² for the monochromatic light. Depending on the amount of hydrogen produced by the photocatalytic reaction in 1 h, the AQY value was calculated by the following Equation:

$$AQY\% = \frac{2 \times \text{Number of } H_2}{\text{Number of incident photons}} \times 100\% = \frac{2 \times R_{H_2} \times N_A}{S \times P \times t \times \frac{\lambda}{hc}} \times 100\%$$

Where R_{H_2} denotes the evolved hydrogen amount (μmol), N_A is the Avogadro constant, S is the illumination area, P and λ are the intensity (W cm^{-2}) and wavelength (nm) of the monochromatic light, respectively. t is the irradiation time, h is the Plank constant, and c is the speed of light.

Structural characterization of organic dyes



Scheme S1. Synthetic routes of organic dyes **LI-154 ~ LI-158** and **LI-161 ~ LI-165**

The general synthesis of compound **1b-j**:

Under nitrogen atmosphere, thiophene derivatives (1.0 equiv.) in THF (40 mL) was placed in Schlenk tube and stirred at $-78\text{ }^{\circ}\text{C}$. *n*-Butyllithium (1.0 equiv.) was added dropwise, and the solution was stirred for 1 h under the same temperature. Then tributyltin chloride (1.0 equiv.) was added, and the reaction mixture was recovered to room temperature and stirred for overnight. The mixture was poured into water and extracted with dichloromethane for three times. The organic layer was combined and dried with anhydrous sodium sulfate. After the solvent was evaporated, the desired product was used directly in the next step without further purification.

The general synthesis of compound **2a-j**:

Under nitrogen atmosphere, a mixture of 2-(Tributylstannyl)thiophene or compound **1b-j** (2.2 equiv.), 4,7-dibromo-5,6-bis(dodecyloxy) benzo [*c*][1,2,5] thiadiazole (1.0 equiv.) and Pd(amphos)Cl₂ (0.05 equiv.) in THF (40 mL) was placed in Schlenk tube and stirred at $80\text{ }^{\circ}\text{C}$ for 12 h. After being cooled to room temperature, the mixture was poured into water and extracted with dichloromethane for three times. The combined organic layer was dried with anhydrous sodium sulfate. After the solvent was evaporated, the crude products were purified by column chromatography.

2a: 2-(Tributylstannyl)thiophene (1.05 g, 3.32 mmol) and orange solid (0.83 g, 82%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.47 (dd, *J* = 3.8, 1.2 Hz, 2H, ArH), 7.50 (dd, *J* = 5.2, 1.2 Hz, 2H, ArH), 7.23 (dd, *J* = 5.2, 3.8 Hz, 2H, ArH), 4.11 (t, *J* = 7.1 Hz, 4H, OCH₂), 1.92 (p, *J* = 7.2 Hz, 4H, CH₂), 1.50-1.38 (m, 4H, CH₂), 1.35-1.25 (m, 32H, CH₂), 0.96 – 0.84 (m, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.94, 150.98, 134.10, 130.54, 127.31, 126.77, 117.61, 74.36, 31.95, 30.35, 29.72, 29.69, 29.65, 29.57, 29.40, 25.96, 22.72, 14.15.

2b: 3-Ethylthiophene (0.37 g, 3.32 mmol) and orange solid (0.78 g, 71%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.31 (d, *J* = 1.4 Hz, 2H, ArH), 7.08 (d, *J* = 1.4 Hz, 2H, ArH), 4.09 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.71 (t, *J* = 7.7 Hz, 4H, CH₂), 1.92 (p, *J* = 7.2 Hz, 4H, CH₂), 1.77-1.65 (m, 4H, CH₂), 1.48-1.41 (m, 4H, CH₂), 1.35-1.27 (m, 32H, CH₂), 0.95-0.85 (m, 12H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.97, 151.01, 142.93, 133.81, 131.96, 122.30, 117.61, 74.27, 31.97, 31.95, 30.67, 30.64, 30.41, 29.76, 29.72, 29.70, 29.65, 29.53, 29.49, 29.42, 29.36, 27.87, 26.87, 26.80, 26.04, 22.73, 22.72, 17.54, 17.30, 14.15, 13.63.

2c: 3-Butylthiophene (0.47 g, 3.32 mmol) and orange solid (0.80 g, 68%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.31 (s, 2H, ArH), 7.09 (s, 2H, ArH), 4.09 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.72 (t, *J* = 7.7 Hz, 4H, CH₂), 1.92 (p, *J* = 7.2 Hz, 4H, CH₂), 1.71 (p, *J* = 7.7 Hz, 4H, CH₂), 1.48 – 1.40 (m, 8H, CH₂), 1.31-1.27 (m, 32H, CH₂), 0.97 (t, *J* = 7.4 Hz, 6H, CH₃), 0.90-0.87 (m, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.97, 151.00, 142.88, 133.80, 131.95, 122.30, 117.60, 74.28, 32.76, 31.95, 30.39, 30.34, 29.73, 29.69, 29.67, 29.63, 29.40, 26.02, 22.72, 22.50, 14.14, 13.99.

2d: 3-Hexylthiophene (0.56 g, 3.32 mmol) and orange solid (1.05 g, 83%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.31 (d, *J* = 1.4 Hz, 2H, ArH), 7.09 (q, *J* = 0.9 Hz, 2H, ArH), 4.09 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.71 (t, *J* = 7.6 Hz, 4H, CH₂), 1.92 (p, *J* = 7.1 Hz, 4H, CH₂), 1.77-1.65 (m, 4H, CH₂), 1.46-1.39 (m, 8H, CH₂), 1.35-1.25 (m, 36H, CH₂), 0.95-0.84 (m, 12H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.95, 150.99, 142.94, 133.79, 131.95, 122.30, 117.59, 77.36, 77.04, 76.73, 74.27, 31.98, 31.79, 31.63, 30.67, 30.60, 30.42, 29.76, 29.72, 29.70, 29.65, 29.46, 29.43, 29.16, 29.09, 27.87, 26.89, 26.04, 22.83, 22.74, 22.70, 22.66, 17.54, 14.17, 13.66.

2e: 3-Octylthiophene (0.65 g, 3.32 mmol) and orange solid (0.94 g, 70%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.32 (s, 2H, ArH), 7.09 (s, 2H, ArH), 4.10 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.72 (t, *J* = 7.7 Hz, 4H, CH₂), 1.92 (p, *J* = 7.2 Hz, 4H, CH₂), 1.77-1.65 (m, 4H, CH₂), 1.47-1.28 (m, 56H, CH₂), 0.93-0.87 (m, 12H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.94, 150.98, 142.91, 133.78, 131.94, 122.28, 117.58, 77.34, 77.03,

76.71, 74.25, 31.96, 31.93, 30.65, 30.63, 30.40, 29.74, 29.70, 29.68, 29.63, 29.52, 29.47, 29.41, 29.34, 28.28, 27.86, 26.79, 26.03, 22.72, 17.52, 17.29, 14.14, 13.61.

2f: 3-(2-Ethylhexyl)thiophene (0.65 g, 3.32 mmol) and orange solid (0.97 g, 72%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.28 (d, *J* = 1.4 Hz, 2H, ArH), 7.06 (d, *J* = 1.3 Hz, 2H, ArH), 4.09 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.65 (d, *J* = 6.8 Hz, 4H, CH₂), 1.92 (p, *J* = 7.2 Hz, 4H, CH₂), 1.68-1.60 (m, 2H, CH), 1.48-1.41 (m, 4H, CH₂), 1.37-1.25 (m, 48H, CH₂), 0.96-0.90 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.94, 151.02, 141.60, 133.62, 132.48, 123.22, 117.59, 74.23, 40.40, 34.67, 32.56, 31.96, 29.68, 29.40, 28.94, 27.86, 26.87, 26.02, 25.69, 23.13, 22.72, 17.54, 14.14, 13.62.

2g: 3-(2-Butyloctyl)thiophene (0.84 g, 3.32 mmol) and orange solid (0.95 g, 63%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.29 (s, 2H, ArH), 7.06 (s, 2H, ArH), 4.10 (t, *J* = 7.2 Hz, 4H, OCH₂), 2.66 (d, *J* = 6.9 Hz, 4H, CH₂), 1.96-1.88 (m, 4H, CH₂), 1.72-1.68 (m, 2H, CH), 1.47-1.43 (m, 4H, CH₂), 1.37-1.27 (m, 64H, CH₂), 0.95-0.86 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.93, 151.03, 141.58, 133.59, 132.56, 123.23, 117.60, 74.22, 39.00, 35.04, 33.40, 31.97, 31.96, 30.42, 30.12, 29.69, 29.66, 29.41, 28.42, 26.55, 26.01, 23.13, 22.72, 14.14.

2h: 3-(2-Hexyldecyl)thiophene (0.93 g, 3.32 mmol) and orange solid (0.97 g, 59%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.29 (s, 2H, ArH), 7.05 (s, 2H, ArH), 4.09 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.65 (d, *J* = 6.7 Hz, 4H, CH₂), 1.92 (p, *J* = 7.2 Hz, 4H, CH₂), 1.74-1.67 (m, 2H, CH), 1.46-1.41 (m, 4H, CH₂), 1.37-1.27 (m, 80H, CH₂), 0.91-0.82 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.91, 151.03, 141.58, 133.59, 132.54, 123.23, 117.59, 74.22, 39.00, 35.03, 33.42, 33.40, 31.97, 31.95, 30.41, 30.12, 29.79, 29.75, 29.71, 29.66, 29.41, 28.40, 28.29, 27.10, 26.79, 26.68, 26.66, 26.04, 22.73, 22.71, 17.30, 14.14, 13.61.

2i: 3-(2-Octyldodecyl)thiophene (1.21 g, 3.32 mmol) and orange solid (0.94 g, 52%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.28 (s, 2H, ArH), 7.05 (s, 2H, ArH), 4.09 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.65 (d, *J* = 6.8 Hz, 4H, CH₂), 1.92 (p, *J* = 7.2 Hz, 4H, CH₂), 1.70 (t, *J* = 5.9 Hz, 2H, CH), 1.46-1.42 (m, 4H, CH₂), 1.35-1.25 (m, 96H, CH₂), 0.90-0.85 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.91, 151.03, 141.58, 133.59, 132.55, 123.23, 117.60, 74.22, 38.99, 35.04, 33.40, 31.97, 31.95, 30.41, 30.12, 29.76, 29.71, 29.69, 29.67, 29.41, 29.39, 26.69, 26.05, 22.73, 22.71, 14.14.

2j: 3-(2-Decyltetradecyl)thiophene (1.40 g, 3.32 mmol) and orange oil (1.07 g, 54%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.29 (s, 2H, ArH), 7.05 (s, 2H, ArH), 4.09 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.65 (d, *J* = 6.8 Hz, 4H, CH₂), 1.92 (p, *J* = 7.3 Hz, 4H, CH₂), 1.73-1.67 (m, 2H, CH), 1.46-1.43 (m, 4H, CH₂), 1.34-1.23 (m, 112H, CH₂), 0.90-0.85 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): δ 151.91, 151.03, 141.58,

133.60, 132.55, 123.23, 117.60, 74.22, 39.00, 35.05, 33.41, 31.98, 31.96, 30.42, 30.13, 29.77, 29.75, 29.73, 29.70, 29.68, 29.43, 29.40, 26.70, 26.06, 22.74, 22.72, 14.15, 13.72.

The general synthesis of compound 3a-j:

Compound **2a-j** (1.0 equiv.) was dissolved in 60 mL mixture solvent (CHCl₃/AcOH = 1/1). Then NBS (2.2 equiv.) was added in batch and reacted at room temperature in dark for 3 h. After that, the mixture was poured into water and extracted with dichloromethane for three times. The combined organic layer was dried with anhydrous sodium sulfate. After the solvent was evaporated, the crude products were purified by column chromatography.

3a: Compound **2a** (0.67 g, 1.0 mmol) and orange solid (0.55 g, 67%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.37 (d, *J* = 4.1 Hz, 2H, ArH), 7.17 (d, *J* = 4.1 Hz, 2H, ArH), 4.12 (t, *J* = 7.2 Hz, OCH₂), 1.94 (p, *J* = 7.2 Hz, CH₂), 1.48-1.42(m, 4H, CH₂), 1.38-1.25(m, 32H, CH₂), 0.92-0.84 (m, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): δ 151.91, 151.02, 141.58, 133.59, 132.54, 123.24, 117.59, 74.23, 38.97, 35.03, 33.41, 33.09, 31.98, 30.41, 29.80, 29.76, 29.71, 29.67, 29.42, 28.93, 26.66, 26.04, 23.14, 22.73, 14.22, 14.16.

3b: Compound **2b** (0.72 g, 1.0 mmol) and orange solid (0.55 g, 62%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.33 (s, 2H, ArH), 4.11 (t, *J* = 7.2 Hz, 4H, OCH₂) 2.69 (t, *J* = 7.7 Hz, 4H, CH₂), 1.95 (p, *J* = 7.2 Hz, 4H, CH₂), 1.55-1.24 (m, 42H, CH₂, CH₃), 0.91-0.85 (m, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.54, 150.45, 142.96, 133.90, 131.18, 116.94, 112.21, 74.50, 31.98, 30.37, 29.76, 29.72, 29.69, 29.60, 29.42, 26.01, 23.10, 22.74, 14.32, 14.16.

3c: Compound **2c** (0.78 g, 1.0 mmol) and orange solid (0.59 g, 63%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.31 (s, 2H, ArH), 4.11 (t, *J* = 7.2 Hz, 4H, OCH₂), 2.67 (t, *J* = 7.7 Hz, 4H, CH₂), 1.94 (p, *J* = 7.2 Hz, 4H, CH₂), 1.71-1.63 (m, 4H, CH₂), 1.48-1.41 (m, 8H, CH₂), 1.38-1.25 (m, 32H, CH₂), 0.98 (t, *J* = 7.3 Hz, 6H, CH₂), 0.90-0.87 (m, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.56, 150.48, 141.73, 133.76, 131.61, 116.94, 112.59, 74.49, 32.04, 31.95, 30.34, 29.73, 29.69, 29.67, 29.58, 29.40, 25.98, 22.72, 22.44, 14.14.

3d: Compound **2d** (0.84 g, 1.0 mmol) and orange solid (0.71 g, 71%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.31 (s, 2H, ArH), 4.11 (t, *J* = 7.2 Hz, 4H, OCH₂), 2.66 (t, *J* = 7.6 Hz, 4H, CH₂), 1.94 (p, *J* = 7.2 Hz, 4H, CH₂), 1.72-1.64 (m, 4H, CH₂), 1.49-1.27 (m, 48H, CH₂), 0.92-0.87 (m, 12H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.52, 150.46, 141.78, 133.75, 131.60, 116.93, 112.60, 74.48, 31.98, 31.72, 30.37, 29.89, 29.77, 29.73, 29.71, 29.62, 29.44, 29.07, 26.01, 22.75, 22.69, 14.18.

3e: Compound **2e** (0.89 g, 1.0 mmol) and orange solid (0.64 g, 61%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.32 (s, 2H, ArH), 4.12 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.67 (t, *J* = 7.6 Hz, 4H, CH₂), 1.96 (p, *J* = 7.2 Hz, 4H, CH₂), 1.70 (p, *J* = 7.3 Hz, 4H, CH₂), 1.51-1.25 (m, 56H, CH₂), 0.92-0.87 (m, 12H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.52, 150.44, 141.75, 133.77, 131.61, 116.92, 112.62, 74.46, 31.99, 31.96, 30.38, 29.92, 29.78, 29.74, 29.72, 29.69, 29.63, 29.49, 29.44, 29.41, 29.36, 26.03, 22.75, 22.74, 14.16.

3f: Compound **2f** (0.89 g, 1.0 mmol) and orange solid (0.77 g, 73%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.30 (s, 2H, ArH), 4.13 (t, *J* = 7.2 Hz, 4H, OCH₂), 2.61 (d, *J* = 7.2 Hz, 4H, CH₂), 1.96 (p, *J* = 7.3 Hz, 4H, CH₂), 1.81-1.69 (m, 2H, CH), 1.51-1.44 (m, 4H, CH₂), 1.42-1.26 (m, 48H, CH₂), 0.98-0.85 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.54, 150.49, 140.91, 133.59, 132.18, 116.94, 113.27, 74.45, 40.04, 33.87, 32.55, 31.98, 30.36, 29.76, 29.72, 29.70, 29.62, 29.43, 28.83, 26.00, 25.77, 23.15, 22.74, 14.20, 14.16.

3g: Compound **2g** (1.00 g, 1.0 mmol) and orange solid (0.70 g, 60%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.29 (s, 2H, ArH), 4.11 (t, *J* = 7.2 Hz, 4H, OCH₂), 2.60 (d, *J* = 7.1 Hz, 4H, CH₂), 1.94 (p, *J* = 7.3 Hz, 4H, CH₂), 1.77 (s, 2H, CH), 1.49-1.42 (m, 4H, CH₂), 1.36-1.23 (m, 64H, CH₂), 0.91-0.85 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.53, 150.52, 140.93, 133.53, 132.24, 116.95, 113.23, 74.47, 38.60, 34.19, 33.40, 33.12, 31.96, 30.33, 29.77, 29.75, 29.69, 29.60, 29.41, 28.82, 26.55, 25.99, 23.13, 22.72, 14.19, 14.14.

3h: Compound **2h** (1.09 g, 1.0 mmol) and orange solid (0.73 g, 57%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.29 (s, 2H, ArH), 4.12 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.60 (d, *J* = 7.0 Hz, 4H, CH₂), 1.94 (p, *J* = 7.2 Hz, 4H, CH₂), 1.77 (s, 2H, CH), 1.48-1.42 (m, 4H, CH₂), 1.39-1.24 (m, 80H, CH₂), 0.90-0.84 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.53, 150.52, 140.93, 133.53, 132.25, 116.95, 113.24, 74.46, 38.60, 34.19, 33.44, 33.41, 31.96, 31.94, 31.61, 30.34, 30.09, 29.76, 29.69, 29.61, 29.41, 29.40, 26.59, 26.56, 26.00, 22.73, 22.70, 22.67, 14.14.

3i: Compound **2i** (1.20 g, 1.0 mmol) and orange oil (0.97 g, 70%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.29 (s, 2H, ArH), 4.11 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.59 (d, *J* = 7.1 Hz, 4H, CH₂), 1.94 (p, *J* = 7.2 Hz, 4H, CH₂), 1.79-1.75 (m, 2H, CH), 1.48-1.43 (m, 4H, CH₂), 1.38-1.21 (m, 96H, CH₂), 0.90-0.84 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.53, 150.52, 140.94, 133.53, 132.25, 116.95, 113.24, 74.46, 38.59, 34.21, 33.42, 31.97, 31.94, 30.34, 30.09, 29.76, 29.74, 29.71, 29.69, 29.62, 29.51, 29.42, 29.40, 29.38, 26.59, 26.00, 22.73, 22.70, 14.13.

3j: Compound **2j** (1.31 g, 1.0 mmol) and orange oil (0.96 g, 64%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.29 (s, 2H, ArH), 4.11 (t, *J* = 7.2 Hz, 4H, OCH₂), 2.59 (d, *J* = 7.1 Hz, 4H, CH₂), 1.94 (p, *J* = 7.2 Hz, 4H,

CH₂), 1.77 (m, 2H, CH), 1.48-1.43 (m, 4H, CH₂), 1.36-1.21 (m, 112H, CH₂), 0.91-0.84 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.53, 150.52, 140.94, 133.53, 132.25, 116.95, 113.24, 74.46, 38.59, 34.21, 33.41, 31.97, 31.94, 30.35, 30.09, 29.76, 29.73, 29.71, 29.68, 29.62, 29.42, 29.38, 26.59, 26.00, 22.73, 22.71, 14.14.

The general synthesis of LI-154-158 and LI-161-LI-165

Under atmosphere of nitrogen, a mixture of the compound **3a-j** (1.0 equiv.), *N,N*-diphenyl-4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline(2.2 equiv.), Pd(amphos)Cl₂ (0.1 equiv.) and potassium carbonate (10.0 equiv.) in 10 mL mixed solvent (THF/H₂O = 10/1) was placed in a 100 mL dry Schlenk tube and stirred at 80 °C for 24 hours. The mixture was poured into water and extracted with chloroform three times. The combined organic layers were dried with anhydrous sodium sulfate. After the solvent was evaporated, the crude products were purified by column chromatography.

LI-154: Compound **3a** (165 mg, 0.2 mmol) and red solid (200 mg, 87%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.43 (d, *J* = 4.0 Hz, 2H, ArH), 7.56-7.47 (m, 4H, ArH), 7.28 (d, *J* = 4.0 Hz, 2H, ArH), 7.26-7.17 (m, 8H, ArH), 7.11-6.94 (m, 16H, ArH), 4.09 (t, *J* = 7.0 Hz, 4H, OCH₂), 1.90 (p, *J* = 7.2 Hz, 4H, CH₂), 1.48-1.25 (m, 36H, CH₂), 0.79 (t, *J* = 6.8 Hz, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.59, 150.96, 147.53, 147.43, 145.49, 132.97, 131.97, 129.39, 128.53, 126.66, 124.63, 123.68, 123.19, 122.27, 117.43, 74.46, 32.00, 30.55, 29.79, 29.74, 29.43, 26.20, 22.77, 14.22. MS (MALDI-TOF, *m/z*): [M]⁺ calcd for C₇₄H₈₂N₄O₂S₃: 1154.560, found 1154.566; Elemental Analysis Calcd for C₇₄H₈₂N₄O₂S₃: C, 76.91; H, 7.15; N, 4.85; S, 8.32. Found: C, 76.72; H, 7.32; N, 4.50 S, 8.53.

LI-155: Compound **3b** (177 mg, 0.2 mmol) and red solid (202 mg, 83%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.34 (s, 2H, ArH), 7.35-7.32 (m, 4H, ArH), 7.24 -7.19 (m, 8H, ArH), 7.11-7.03 (m, 12H, ArH), 7.00-6.96 (m, 4H, ArH), 4.08 (t, 4H, *J* = 7.2 Hz, OCH₂), 2.75 (dd, 4H, *J* = 5.6 Hz, CH₂), 1.88 (p, 4H, *J* = 7.2 Hz, CH₂), 1.42-1.36 (m, 4H, CH₂), 1.28 (t, 6H, *J* = 7.6 Hz, CH₃), 1.22-1.17 (m, 32H, CH₂), 0.79 (t, 6H, *J* = 6.8 Hz, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.78, 151.02, 147.59, 147.08, 139.52, 139.33, 133.10, 131.86, 129.83, 129.34, 128.50, 124.68, 123.14, 117.31, 74.37, 31.94, 30.45, 29.71, 29.67, 29.37, 26.14, 22.71, 22.26, 15.60, 14.14. MS (MALDI-TOF, *m/z*): [M]⁺ calcd for C₇₈H₉₀N₄O₂S₃: 1210.623, found 1210.636; Elemental Analysis Calcd for C₇₈H₉₀N₄O₂S₃: C, 77.31; H, 7.49; N, 4.62; S, 7.94. Found: C, 77.40; H, 7.25; N, 4.34; S, 7.51.

LI-156: Compound **3c** (188 mg, 0.2 mmol) and A red solid (188 mg, 74%). ¹H NMR (400 MHz, CDCl₃) δ

(ppm): 8.31 (s, 2H, ArH), 7.36-7.30 (d, $J = 8.2$ Hz, 4H, ArH), 7.26-7.17 (m, 8H, ArH), 7.11-6.94 (m, 16H, ArH), 4.07 (t, $J = 7.2$ Hz, 4H, OCH₂), 2.71 (t, $J = 7.8$ Hz, 4H, CH₂), 1.88 (p, $J = 7.0$ Hz, 4H, CH₂), 1.69-1.61 (m, 4H, CH₂), 1.42 -1.17 (m, 40H, CH₂), 0.86 (t, $J = 7.4$ Hz, 6H, CH₃), 0.79 (t, $J = 6.8$ Hz, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.78, 151.03, 147.61, 147.06, 139.82, 138.07, 133.49, 131.74, 129.90, 129.35, 128.59, 124.68, 123.13, 117.31, 74.36, 33.36, 31.95, 30.46, 29.72, 29.69, 29.38, 28.74, 26.15, 22.76, 22.72, 14.15. MS (MALDI-TOF, m/z): [M]⁺ calcd for C₈₂H₉₈N₄O₂S₃: 1266.685, found 1266.711; Elemental Analysis Calcd for C₈₂H₉₈N₄O₂S₃: C, 77.68; H, 7.79; N, 4.42; S, 7.59. Found: C, 78.01; H, 7.47; N, 4.17. S, 7.34

LI-157: Compound **3d** (200 mg, 0.2 mmol) and red solid (225 mg, 87%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.31 (s, 2H, ArH), 7.36- 7.30 (m, 4H, ArH), 7.26-7.16 (m, 8H, ArH), 7.13-6.94 (m, 16H, ArH), 4.08 (t, $J = 7.1$ Hz, 4H, OCH₂), 2.70 (t, $J = 7.9$ Hz, 4H, CH₂), 1.88 (p, $J = 7.2$ Hz, 4H, CH₂), 1.72-1.60 (m, 4H, CH₂), 1.46-1.17 (m, 48H, CH₂), 0.83-0.86 (m, 12H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.77, 151.02, 147.60, 147.04, 139.80, 138.11, 133.48, 131.72, 129.88, 129.33, 128.57, 124.68, 123.12, 117.30, 77.34, 77.03, 76.71, 74.34, 31.94, 31.16, 30.45, 29.72, 29.68, 29.49, 29.38, 29.35, 28.98, 26.15, 22.71, 14.14. MS (MALDI-TOF, m/z): [M]⁺ calcd for C₈₆H₁₀₆N₄O₂S₃: 1322.748, found 1322.794; Elemental Analysis Calcd for C₈₆H₁₀₆N₄O₂S₃: C, 78.02; H, 8.07; N, 4.23; S, 7.26. Found: C, 78.15; H, 7.94; N, 4.16; S, 7.28.

LI-158: Compound **3e** (210 mg, 0.2 mmol) and red solid (184 mg, 67%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.38 (s, 2H, ArH), 7.44- 7.36 (m, 4H, ArH), 7.33-7.26 (m, 4H, ArH), 7.20-7.09 (m, 12H, ArH), 7.08-7.03 (m, 4H, ArH), 4.15 (t, $J = 7.0$ Hz, 4H, OCH₂), 2.78 (t, $J = 7.7$ Hz, 4H, CH₂), 1.95 (p, $J = 7.2$ Hz, 4H, CH₂), 1.78-1.66 (m, 4H, CH₂), 1.51-1.24 (m, 56H, CH₂), 0.94 (t, $J = 7.4$ Hz, 6H, CH₃), 0.89-0.85 (m, 6H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.77, 151.03, 147.60, 147.05, 139.80, 138.11, 133.49, 131.73, 129.89, 129.34, 128.58, 124.68, 123.13, 123.11, 117.31, 74.35, 31.95, 31.17, 30.46, 29.73, 29.69, 29.67, 29.50, 29.38, 29.35, 28.99, 26.16, 22.71, 14.15. MS (MALDI-TOF, m/z): [M]⁺ calcd for C₉₀H₁₁₄N₄O₂S₃: 1378.810, found 1378.788; Elemental Analysis Calcd for C₉₀H₁₁₄N₄O₂S₃: C, 78.33; H, 8.33; N, 4.06; S, 6.97. Found: C, 78.26; H, 8.16; N, 3.97; S, 6.94.

LI-161: Compound **3f** (210 mg, 0.2 mmol) and red solid (198 mg, 72%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.34 (s, 2H, ArH), 7.44- 7.37 (m, 4H, ArH), 7.32-7.25 (m, 8H, ArH), 7.18-7.10 (m, 12H, ArH), 7.07-7.02 (m, 4H, ArH), 4.15 (t, $J = 7.1$ Hz, 4H, OCH₂), 2.72 (d, $J = 7.1$ Hz, 4H, CH₂), 1.95 (p, $J = 7.2$ Hz, 4H, CH₂), 1.67 (h, $J = 6.3$ Hz, 2H, CH), 1.53-1.41 (m, 4H, CH₂), 1.37-1.21 (m, 48H, CH₂), 0.91-0.78 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.75, 151.06, 147.64, 147.05, 140.53, 137.31, 133.91, 13

131.57, 130.23, 129.32, 128.80, 124.60, 123.20, 123.07, 117.33, 74.32, 40.61, 32.79, 32.62, 31.95, 30.43, 29.73, 29.69, 29.38, 28.73, 26.13, 25.80, 23.12, 22.71, 14.20, 14.15, 10.83. MS (MALDI-TOF, m/z): [M]⁺ calcd for C₉₀H₁₁₄N₄O₂S₃: 1378.810, found 1378.815; Elemental Analysis Calcd for C₉₀H₁₁₄N₄O₂S₃: C, 78.33; H, 8.33; N, 4.06; S, 6.97. Found: C, 77.94; H, 8.17; N, 3.82; S, 6.77.

LI-162: Compound **3g** (232mg, 0.2 mmol) and red solid (152 mg, 51%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.35 (s, 2H, ArH), 7.43-7.36 (m, 4H, ArH), 7.31-7.26 (m, 8H, ArH), 7.19-7.14 (m, 8H, ArH), 7.13-7.09 (m, 4H, ArH), 7.07-7.03 (m, 4H, ArH), 4.15 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.72 (d, *J* = 7.0 Hz, 4H, CH₂), 1.95 (p, *J* = 7.2 Hz, 4H, CH₂), 1.77-1.69 (m, 2H, CH), 1.51-1.43 (m, 4H, CH₂), 1.39-1.23 (m, 64H, CH₂), 0.89-0.82 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.73, 151.07, 147.63, 147.03, 140.50, 137.28, 133.98, 131.55, 130.19, 129.32, 128.78, 124.63, 123.11, 123.08, 117.33, 74.32, 39.16, 33.47, 33.18, 31.97, 31.95, 30.44, 29.80, 29.74, 29.69, 29.38, 28.74, 26.48, 26.15, 23.13, 22.75, 22.71, 14.20, 14.16, 14.14. MS (MALDI-TOF, m/z): [M]⁺ calcd for C₉₈H₁₃₀N₄O₂S₃: 1492.324, found 1492.948; Elemental Analysis Calcd for C₉₈H₁₃₀N₄O₂S₃: C, 78.88; H, 8.78; N, 3.75; S, 6.44. Found: C, 78.87; H, 8.70; N, 3.68; S, 6.36.

LI-163: Compound **3h** (255 mg, 0.2 mmol) and red solid (156 mg, 48%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.35 (s, 2H, ArH), 7.40-7.38 (m, 4H, ArH), 7.30-7.26 (m, 8H, ArH), 7.18-7.13 (m, 8H, ArH), 7.12-7.10 (m, 4H, ArH), 7.07-7.03 (m, 4H, ArH), 4.15 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.72 (d, *J* = 7.0 Hz, 4H, CH₂), 1.95 (p, *J* = 7.2 Hz, 4H, CH₂), 1.75-1.71 (m, 2H, CH), 1.49-1.43 (m, 4H, CH₂), 1.39-1.22 (m, 80H, CH₂), 0.88-0.81 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.72, 151.07, 147.63, 147.03, 140.51, 137.28, 133.97, 131.54, 130.18, 129.32, 128.76, 124.65, 123.09, 123.07, 117.32, 74.32, 39.19, 33.50, 33.16, 31.97, 31.95, 30.45, 30.14, 29.80, 29.75, 29.71, 29.43, 29.39, 26.54, 26.48, 26.16, 22.75, 22.71, 14.16, 14.14. MS (MALDI-TOF, m/z): [M]⁺ calcd for C₁₀₆H₁₄₆N₄O₂S₃: 1603.061, found 1603.075; Elemental Analysis Calcd for C₁₀₆H₁₄₆N₄O₂S₃: C, 79.35; H, 9.17; N, 3.49; S, 5.99. Found: C, 79.26; H, 8.96; N, 3.37; S, 5.77.

LI-164: Compound **3i** (276 mg, 0.2 mmol) and red oil (180 mg, 56%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.35 (s, 2H, ArH), 7.41-7.37 (m, 4H, ArH), 7.30-7.26 (m, 8H, ArH), 7.16-7.14 (m, 8H, ArH), 7.13-7.09 (m, 4H, ArH), 7.07-7.03 (m, 4H, ArH), 4.15 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.72 (d, *J* = 7.1 Hz, 4H, CH₂), 1.95 (p, *J* = 7.2 Hz, 4H, CH), 1.77-1.71 (m, 2H, CH₂), 1.51-1.43 (m, 4H, CH₂), 1.39-1.22 (m, 96H, CH₂), 0.88-0.83 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.71, 151.06, 147.61, 147.01, 140.48, 137.27, 133.94, 131.53, 130.17, 129.30, 128.75, 124.63, 123.07, 117.31, 74.29, 39.17, 33.48, 33.16, 31.93, 30.44, 30.13, 29.77, 29.74, 29.69, 29.42, 29.38, 26.52, 26.15, 22.69, 14.13. MS (MALDI-TOF, m/z): [M]⁺ calcd for C₁₁₄H₁₆₂N₄O₂S₃: 1715.186, found 1715.167; Elemental Analysis Calcd for C₁₁₄H₁₆₂N₄O₂S₃: C, 79.76; H,

9.51; N, 3.26; S, 5.60. Found: C, 79.45; H, 9.17; N, 3.03; S, 5.42.

LI-165: Compound **3j** (300 mg, 0.2 mmol) and red oil (189 mg, 57%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.34 (s, 2H, ArH), 7.40-7.38 (m, 4H, ArH), 7.30-7.26 (m, 8H, ArH), 7.16-7.14 (m, 8H, ArH), 7.12-7.10 (m, 4H, ArH), 7.07-7.03 (m, 4H, ArH), 4.15 (t, *J* = 7.1 Hz, 4H, OCH₂), 2.77-2.65 (m, 4H, CH₂), 1.98-1.92 (m, 4H, CH₂), 1.75-1.71 (m, 2H, CH), 1.50-1.44 (m, 4H, CH₂), 1.39-1.22 (m, 112H, CH₂), 0.88-0.84 (m, 18H, CH₂). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 151.71, 151.06, 147.01, 137.27, 133.94, 130.17, 129.30, 128.76, 124.63, 123.08, 77.33, 77.02, 76.70, 74.29, 39.17, 33.47, 33.15, 31.93, 30.44, 30.13, 29.77, 29.74, 29.69, 29.38, 26.52, 26.14, 22.70, 14.13. MS (MALDI-TOF, *m/z*): [M]⁺ calcd for C₁₂₂H₁₇₈N₄O₂S₃: 1827.311, found 1827.291; Elemental Analysis Calcd for C₁₂₂H₁₇₈N₄O₂S₃: C, 80.12; H, 9.81; N, 3.06; S, 5.26. Found: C, 79.88; H, 9.59; N, 2.83; S, 4.95.

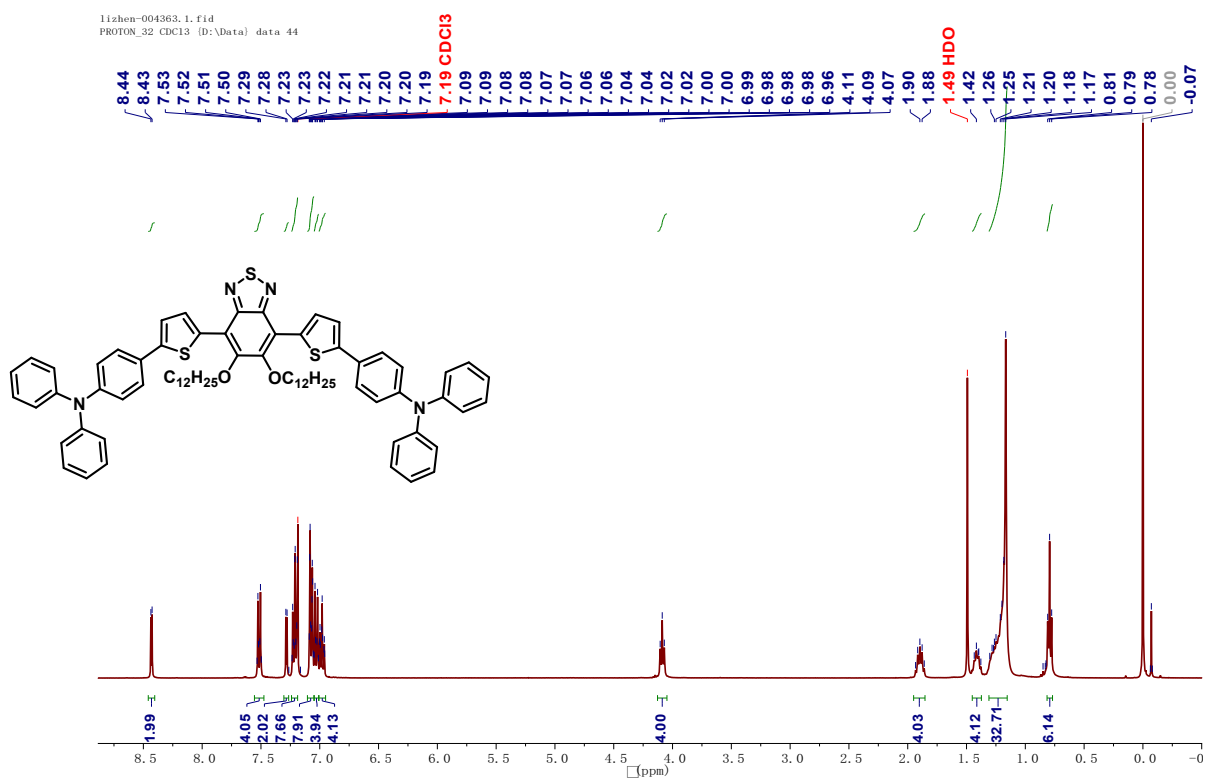


Figure S1. ¹H NMR spectrum of LI-154.

核磁.3.fid

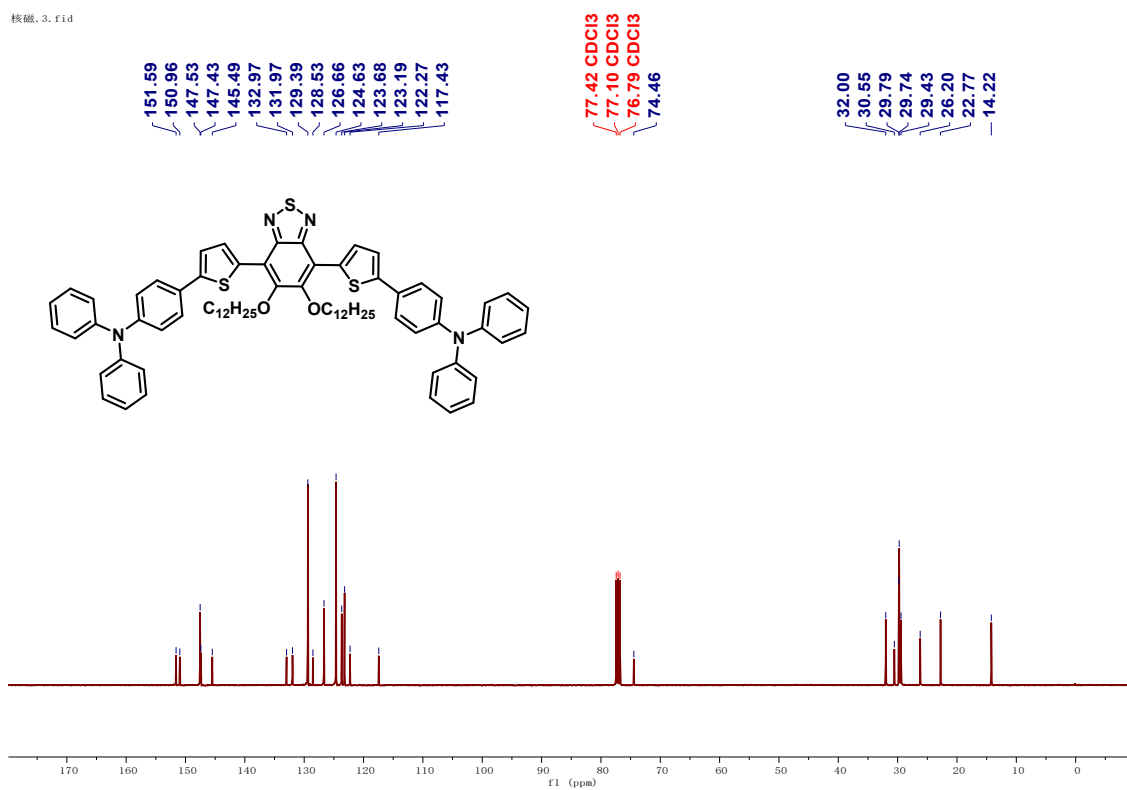


Figure S2. ¹³C NMR spectrum of LI-154.

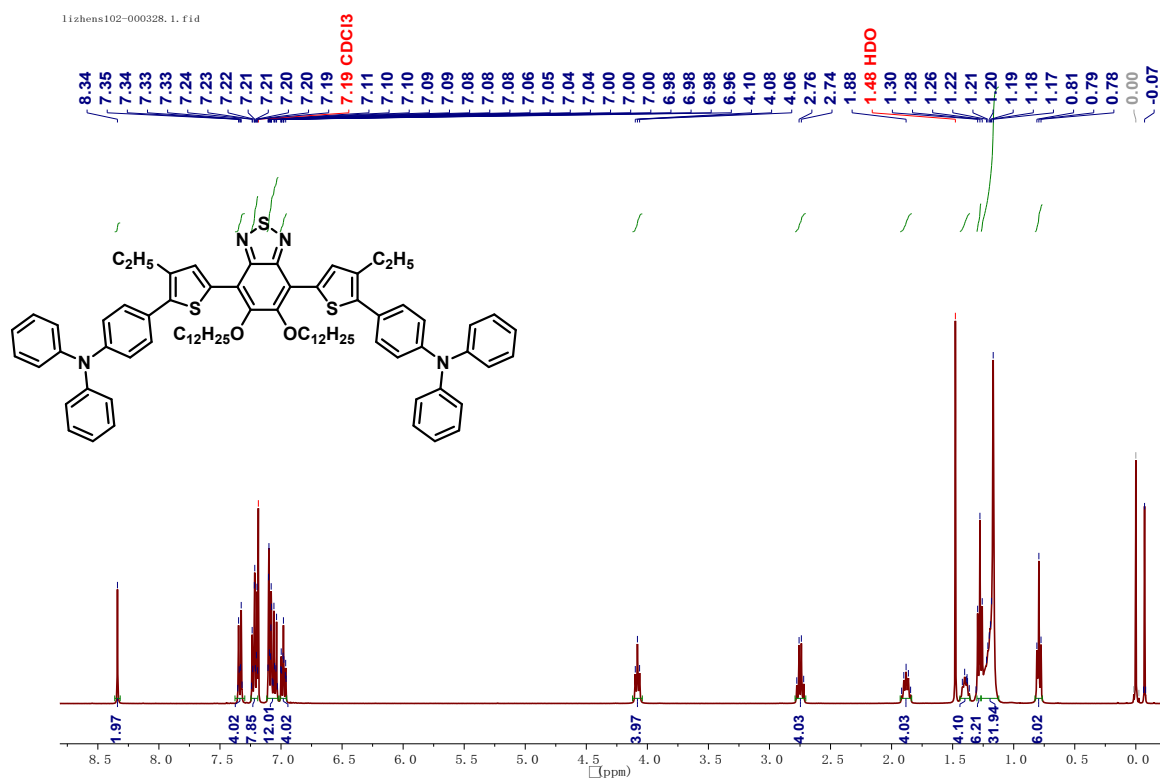
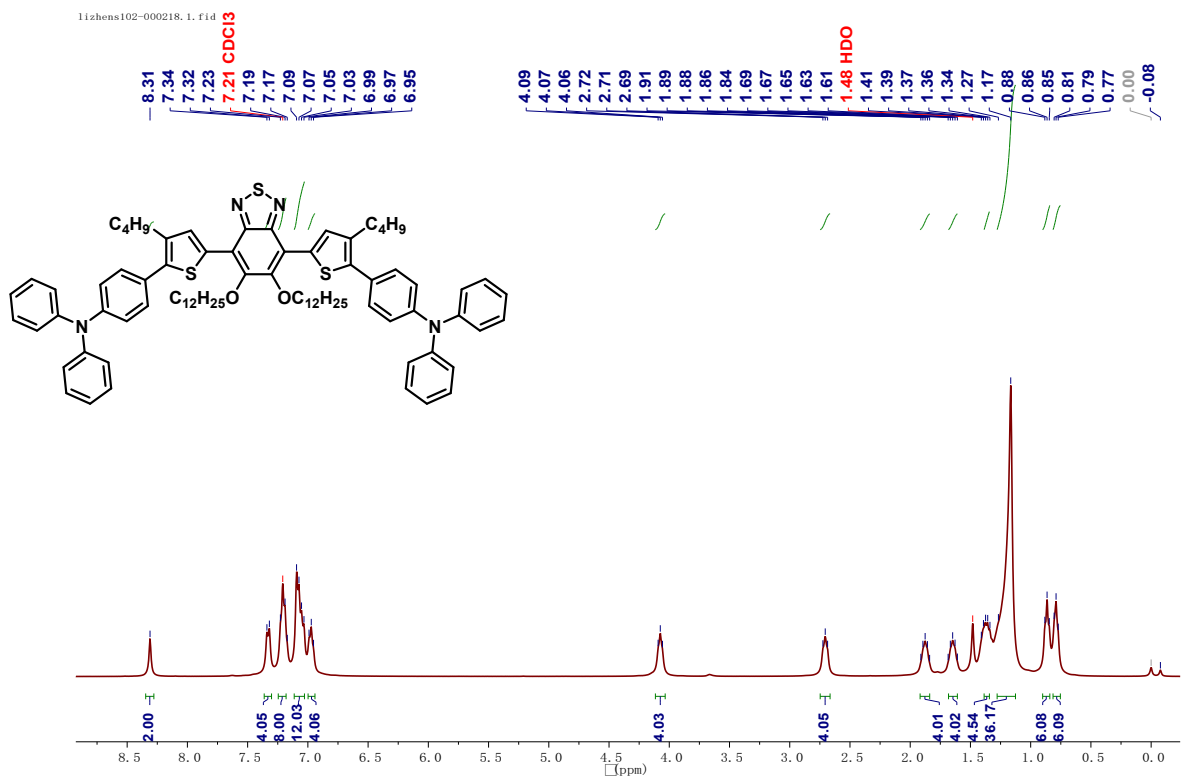
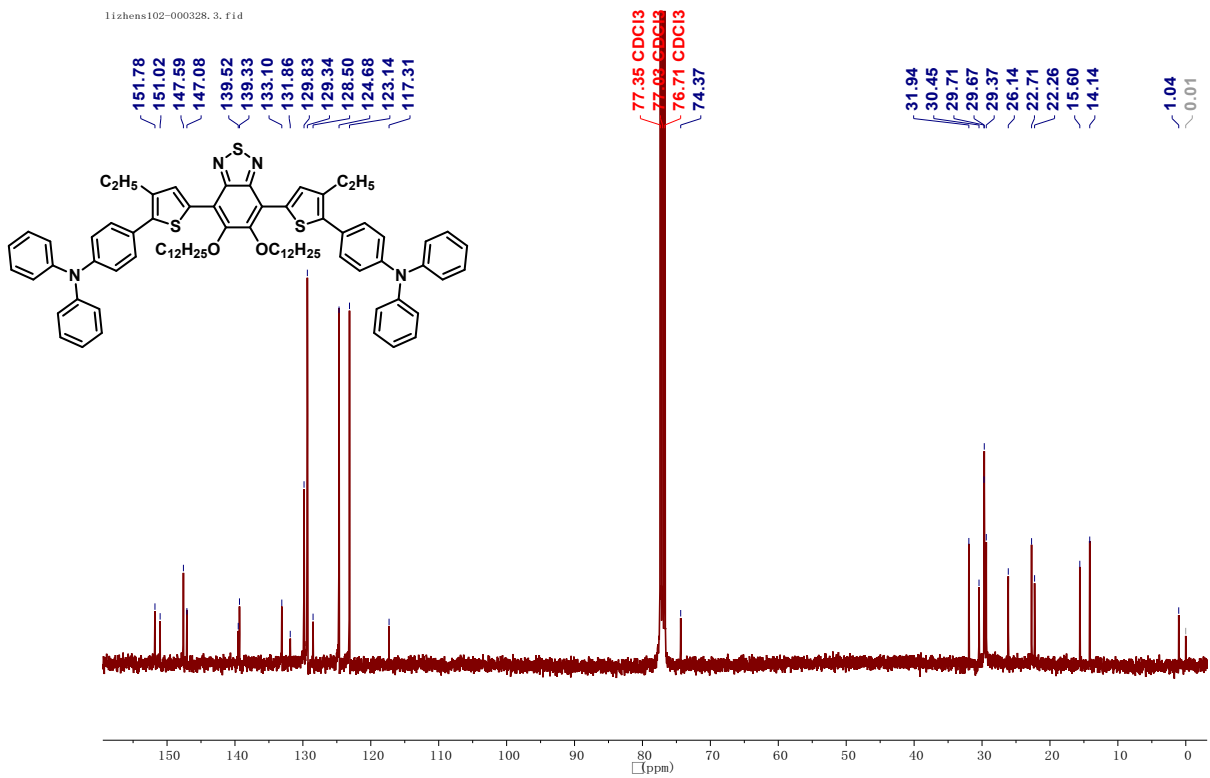


Figure S3. ¹H NMR spectrum of LI-155.



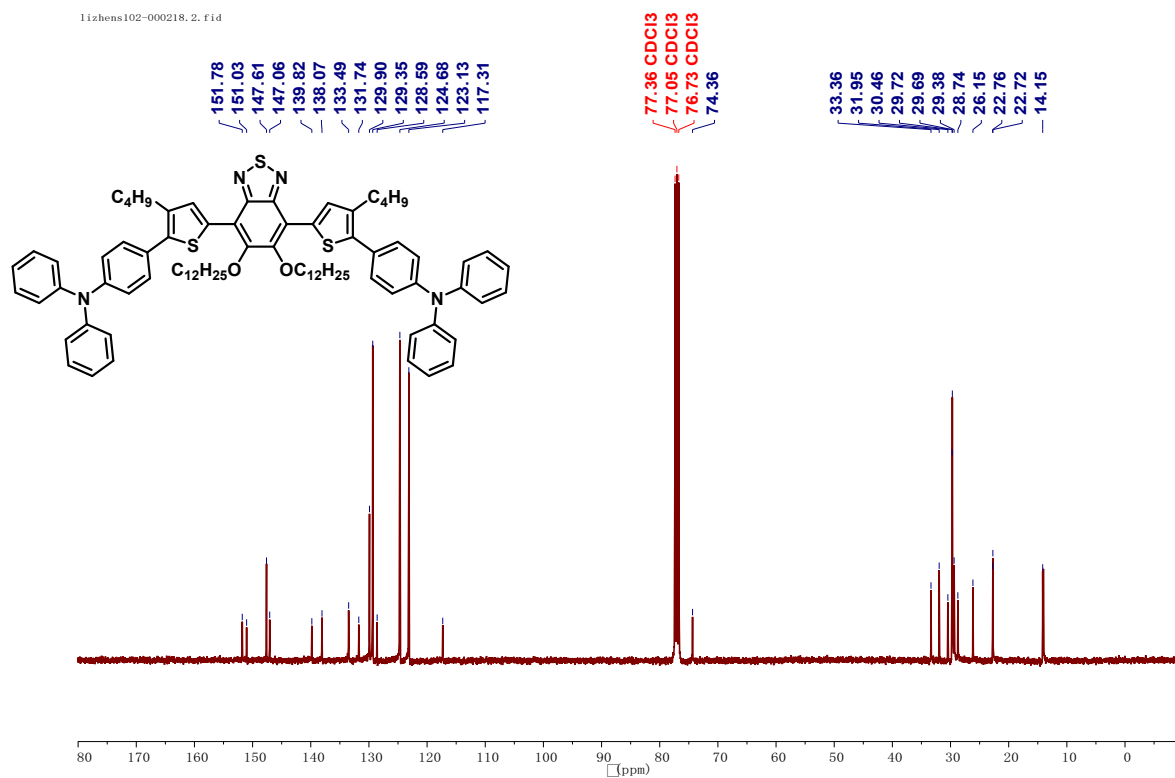


Figure S6. ¹³C NMR spectrum of LI-156.

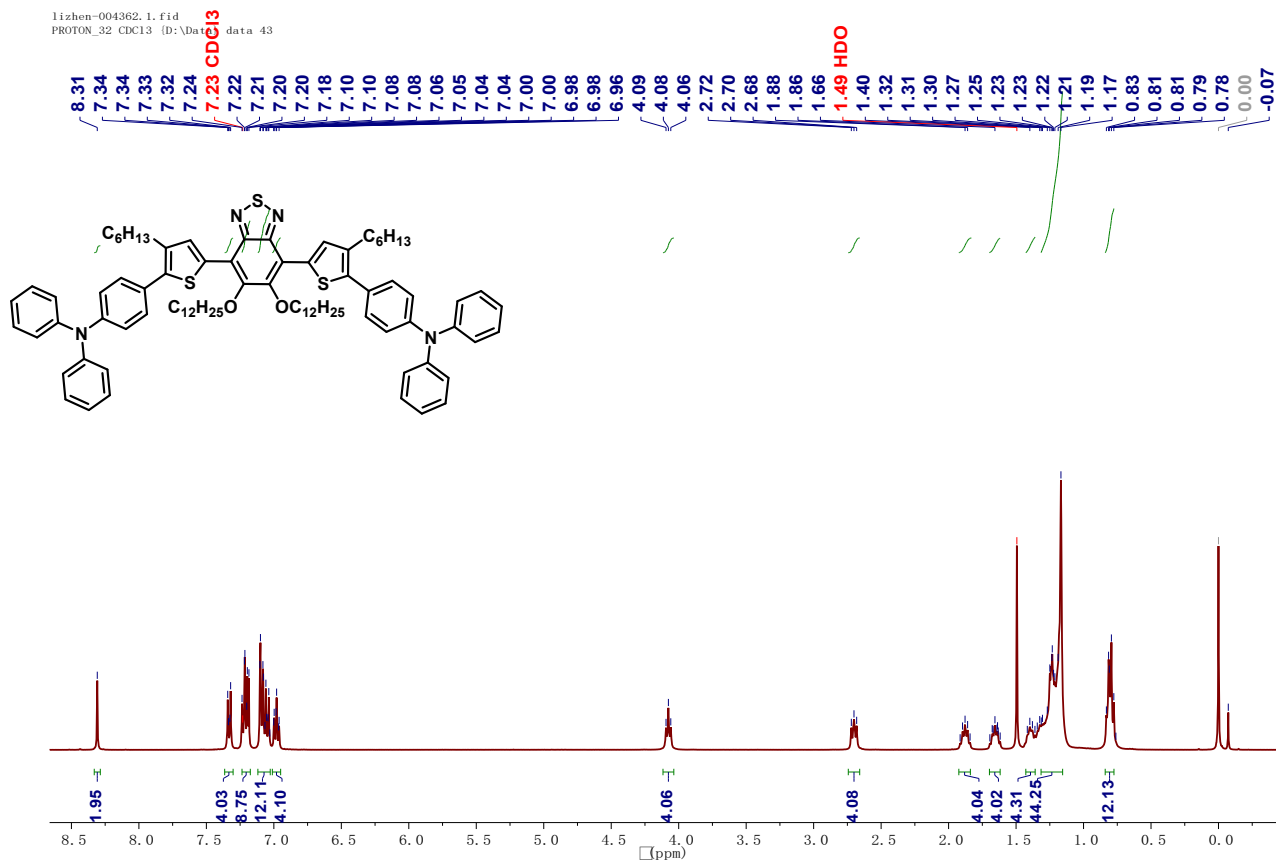


Figure S7. ¹H NMR spectrum of LI-157.

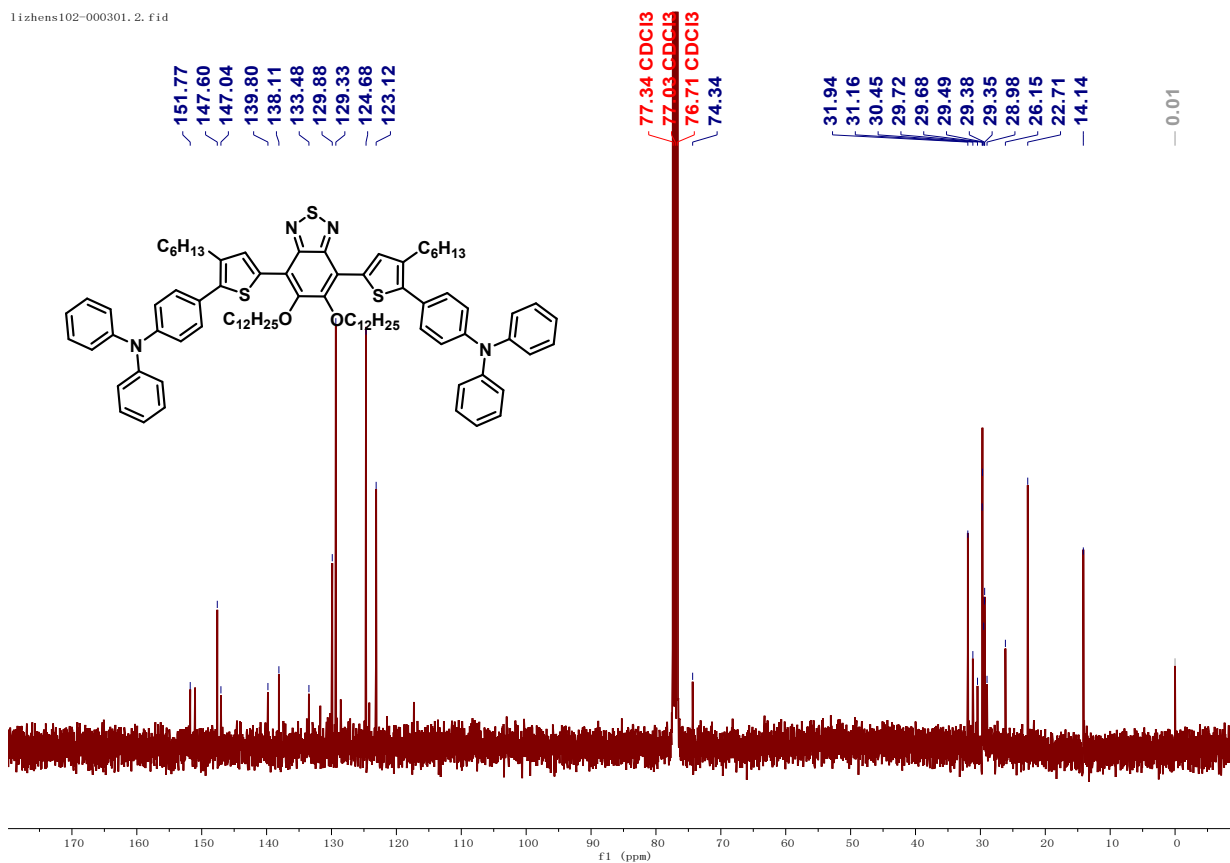


Figure S8. ¹³C NMR spectrum of LI-157.

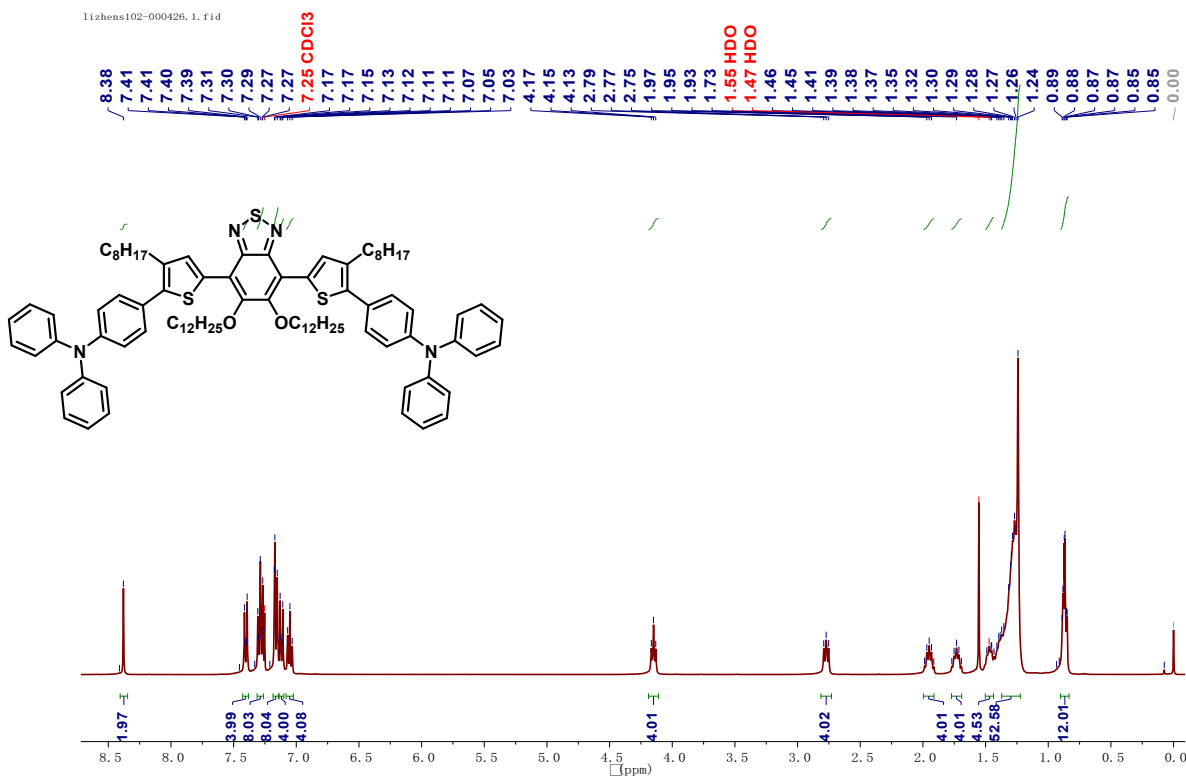


Figure S9. ¹H NMR spectrum of LI-158.

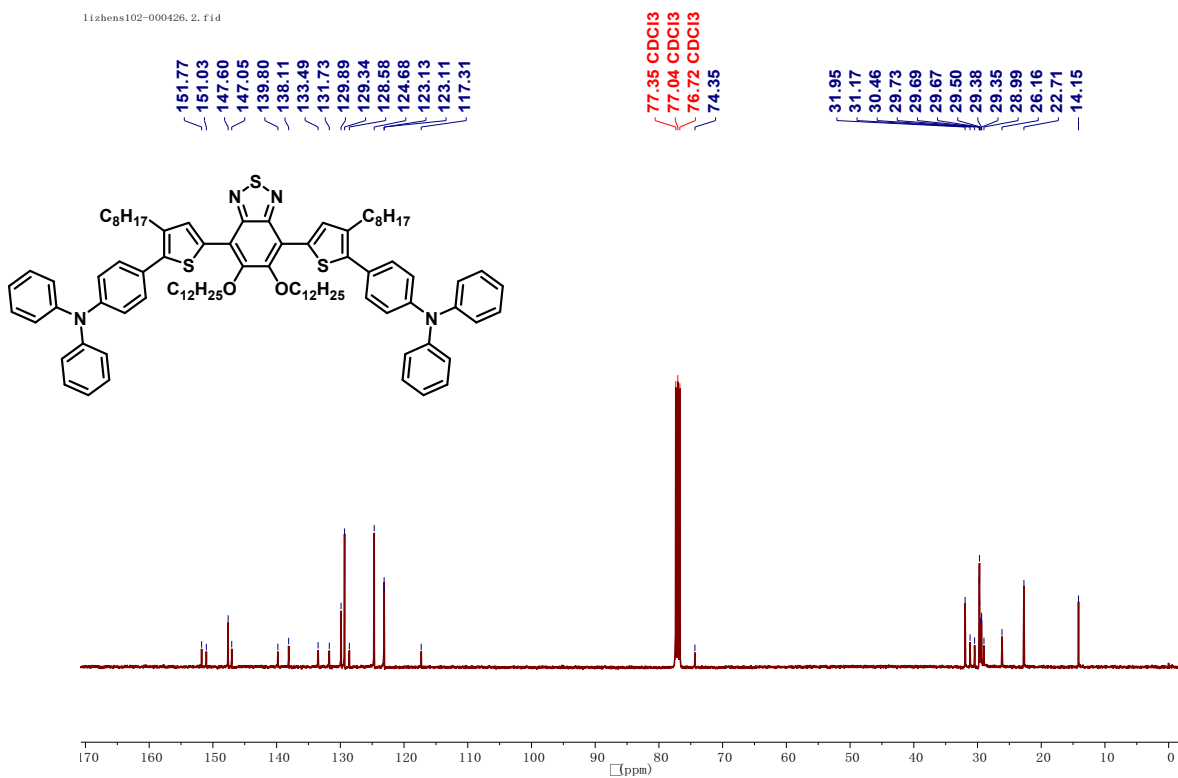


Figure S10. ^{13}C NMR spectrum of LI-158.

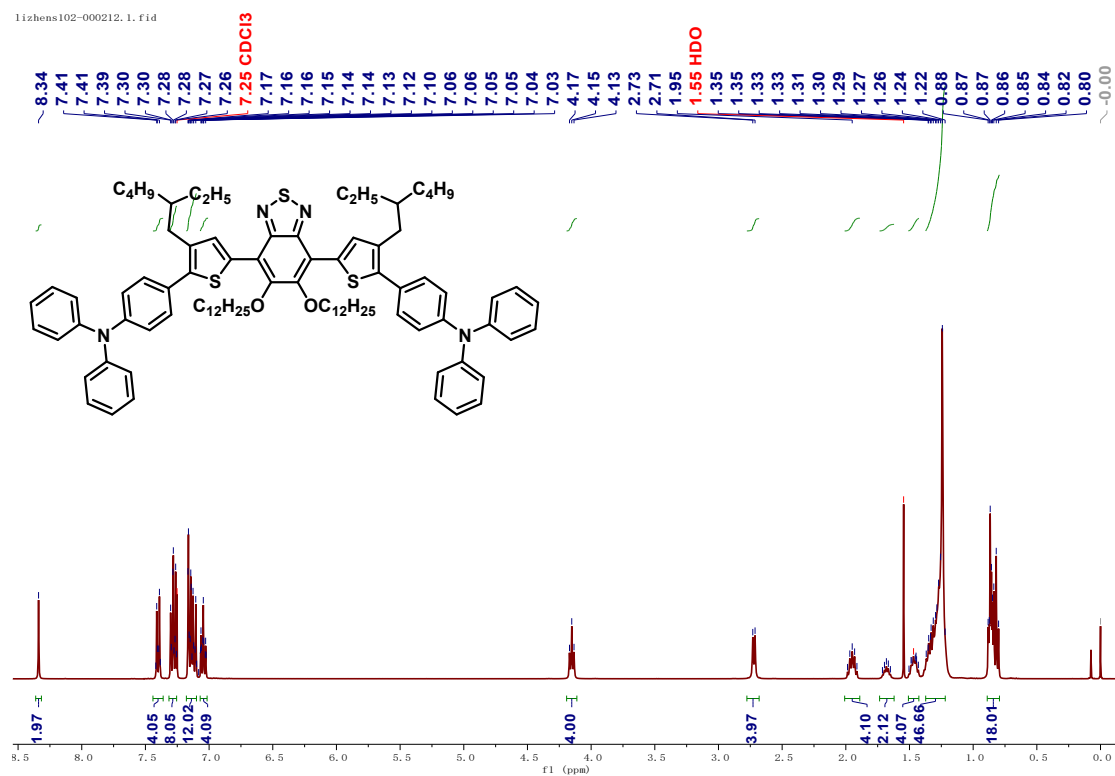


Figure S11. ^1H NMR spectrum of LI-161.

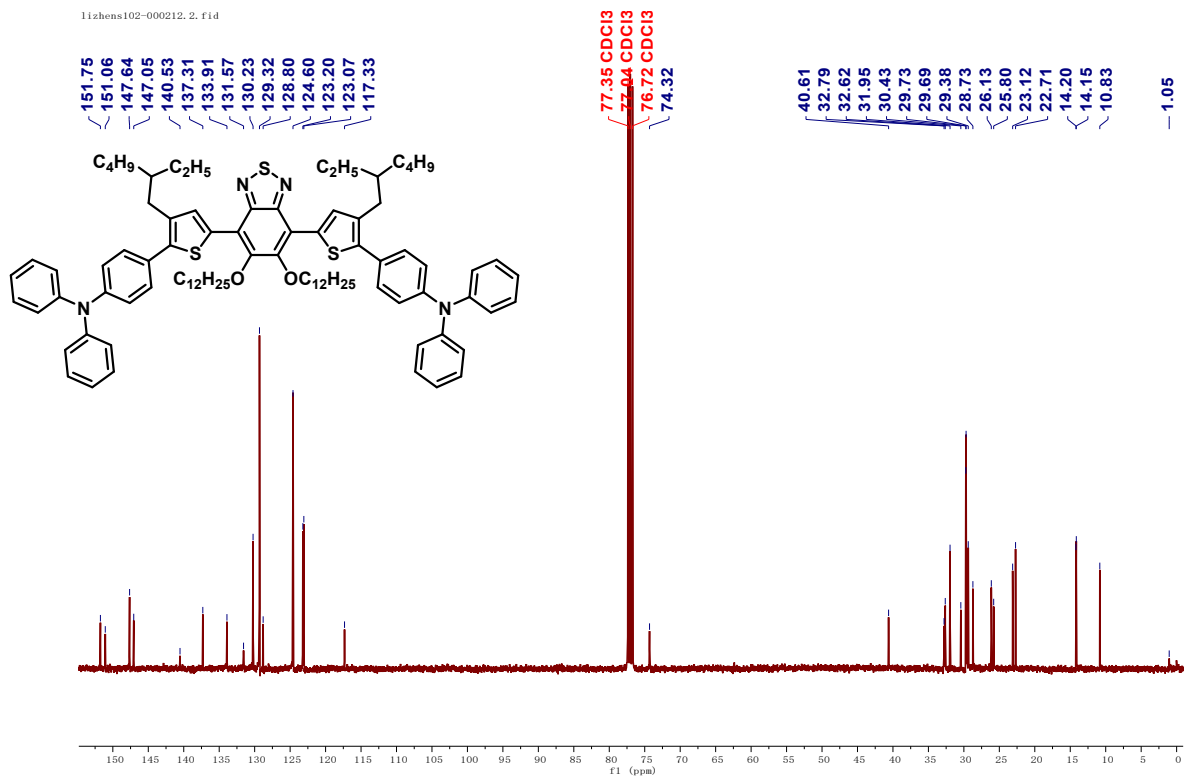


Figure S12. ^{13}C NMR spectrum of LI-161.

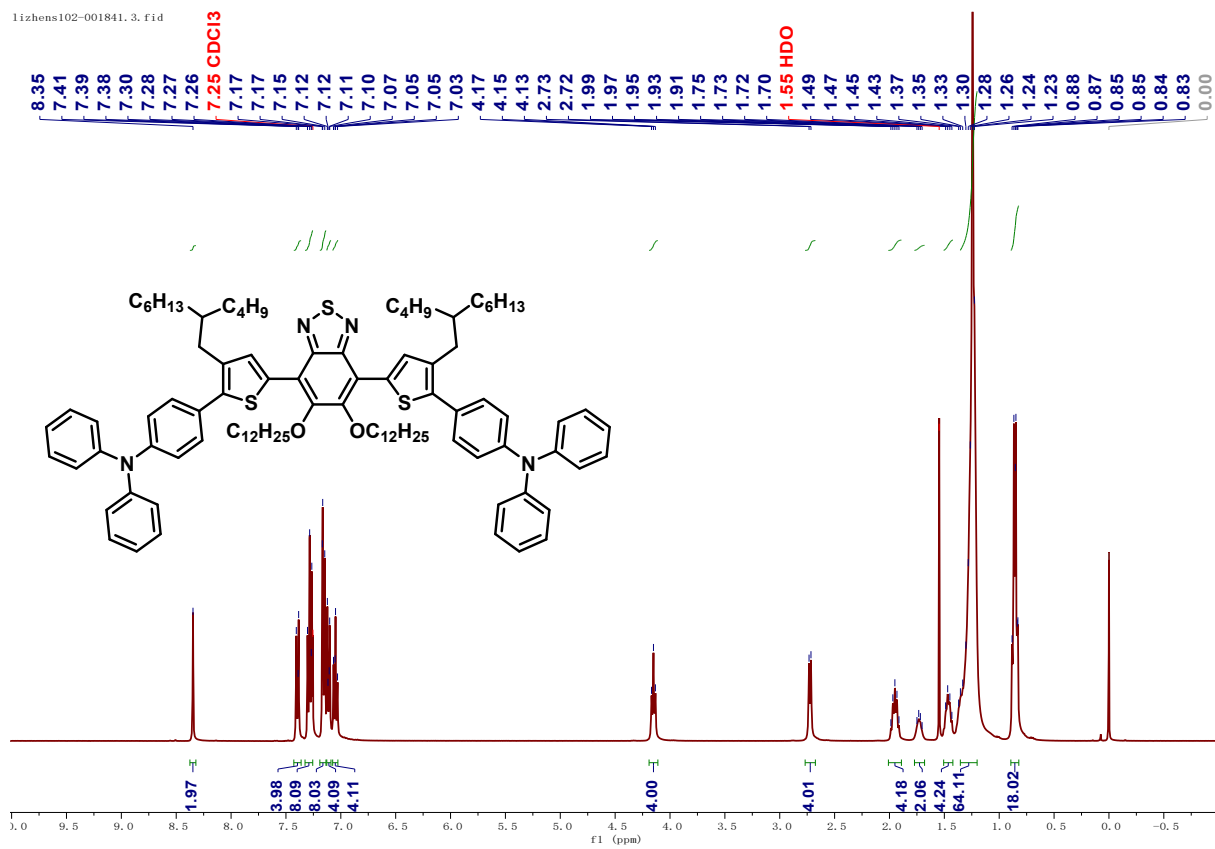


Figure S13. 1H NMR spectrum of LI-162.

lizhens102-001841.4.fid

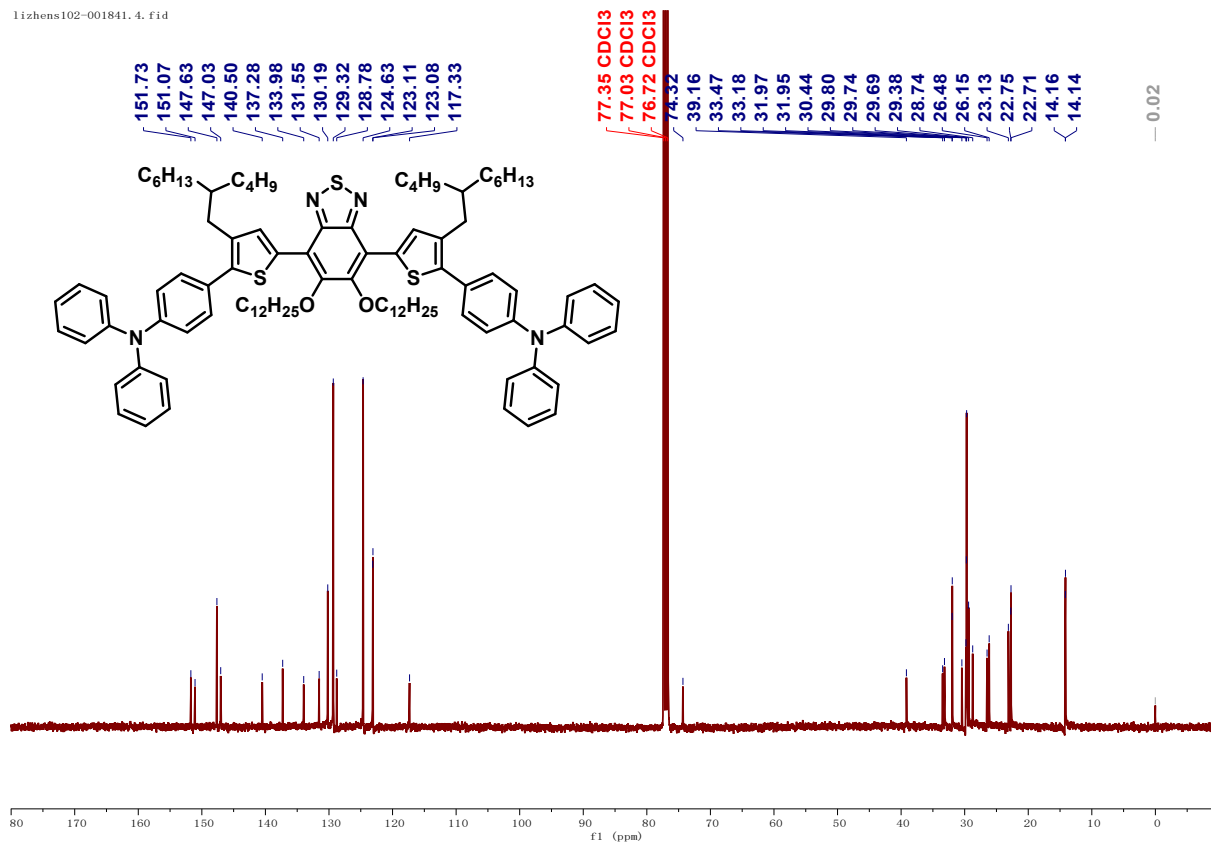


Figure S14. ^{13}C NMR spectrum of LI-162.

lizhens102-001426.1.fid

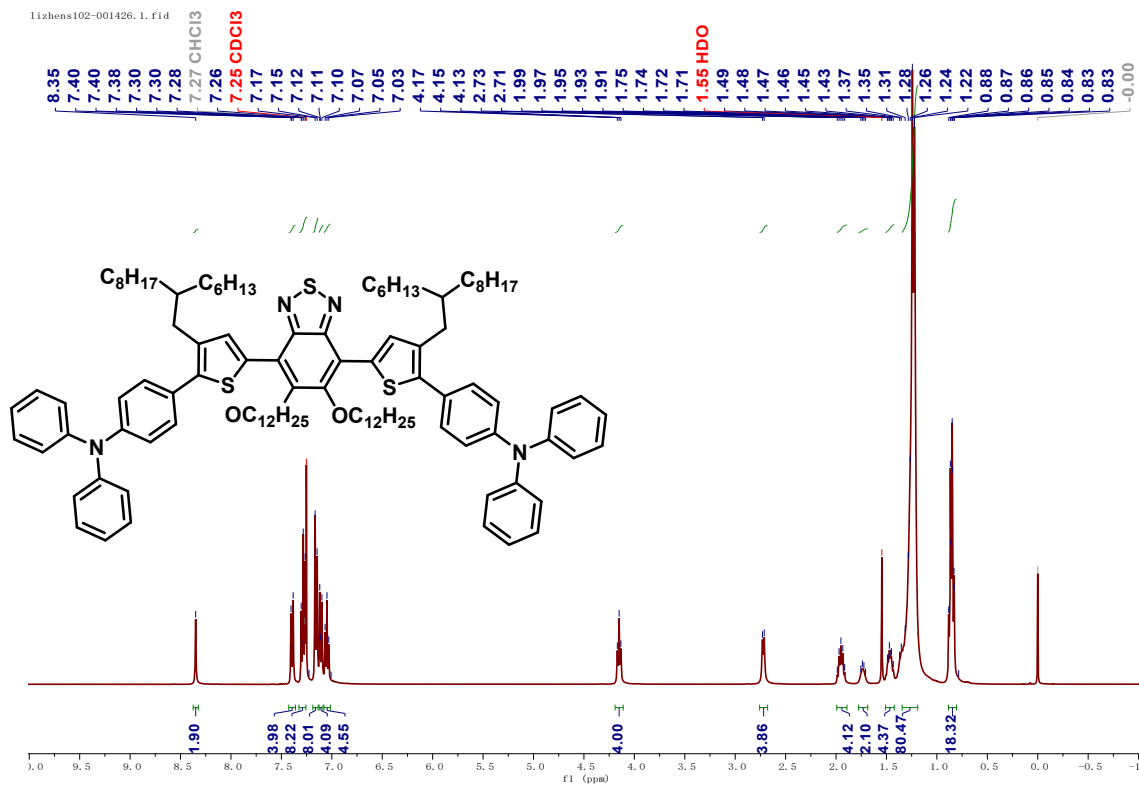


Figure S15. ^1H NMR spectrum of LI-163.

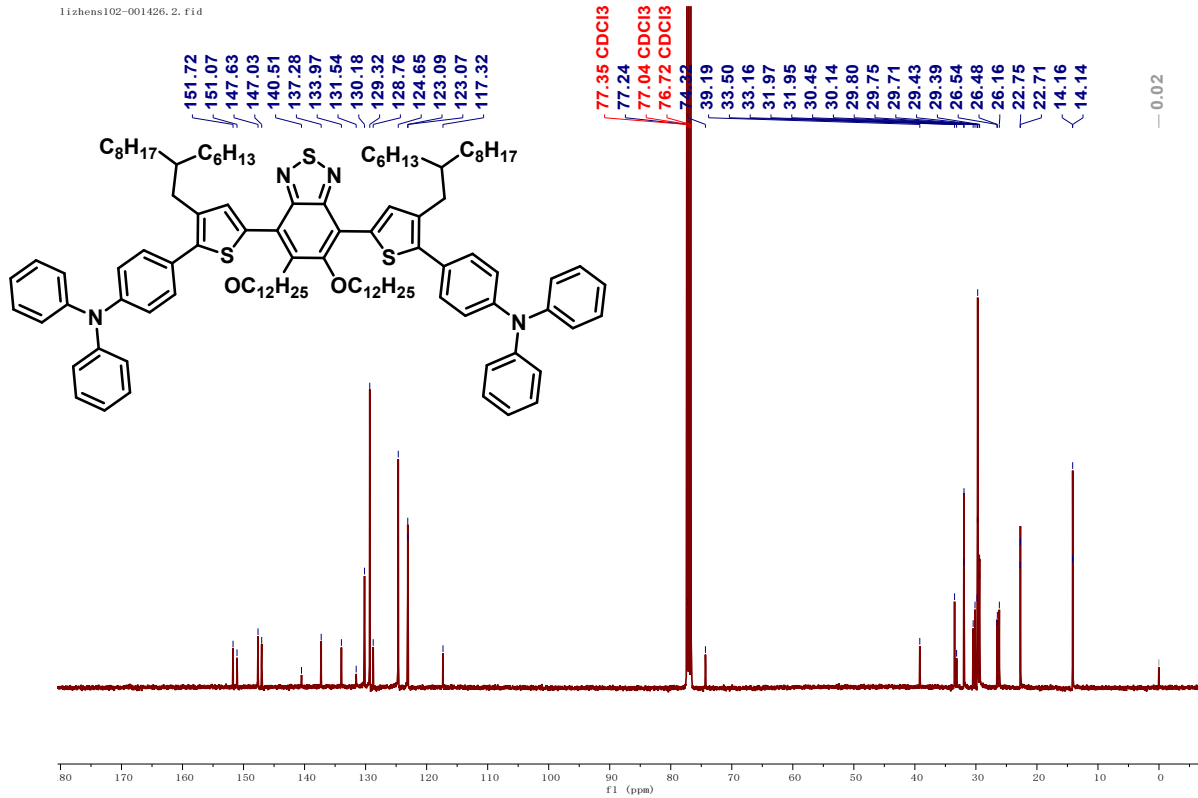


Figure S16. ¹³C NMR spectrum of LI-163.

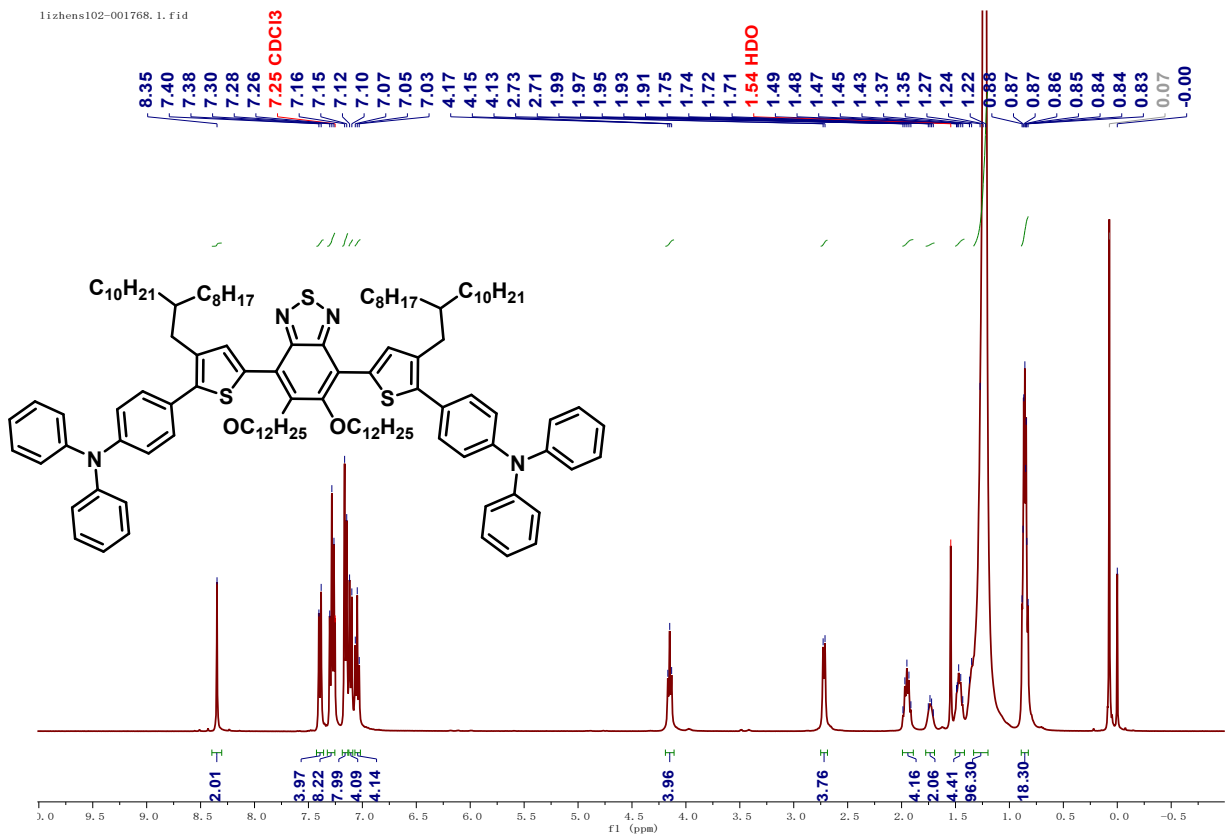


Figure S17. ¹H NMR spectrum of LI-164.

lizhens102-001768.2.fid

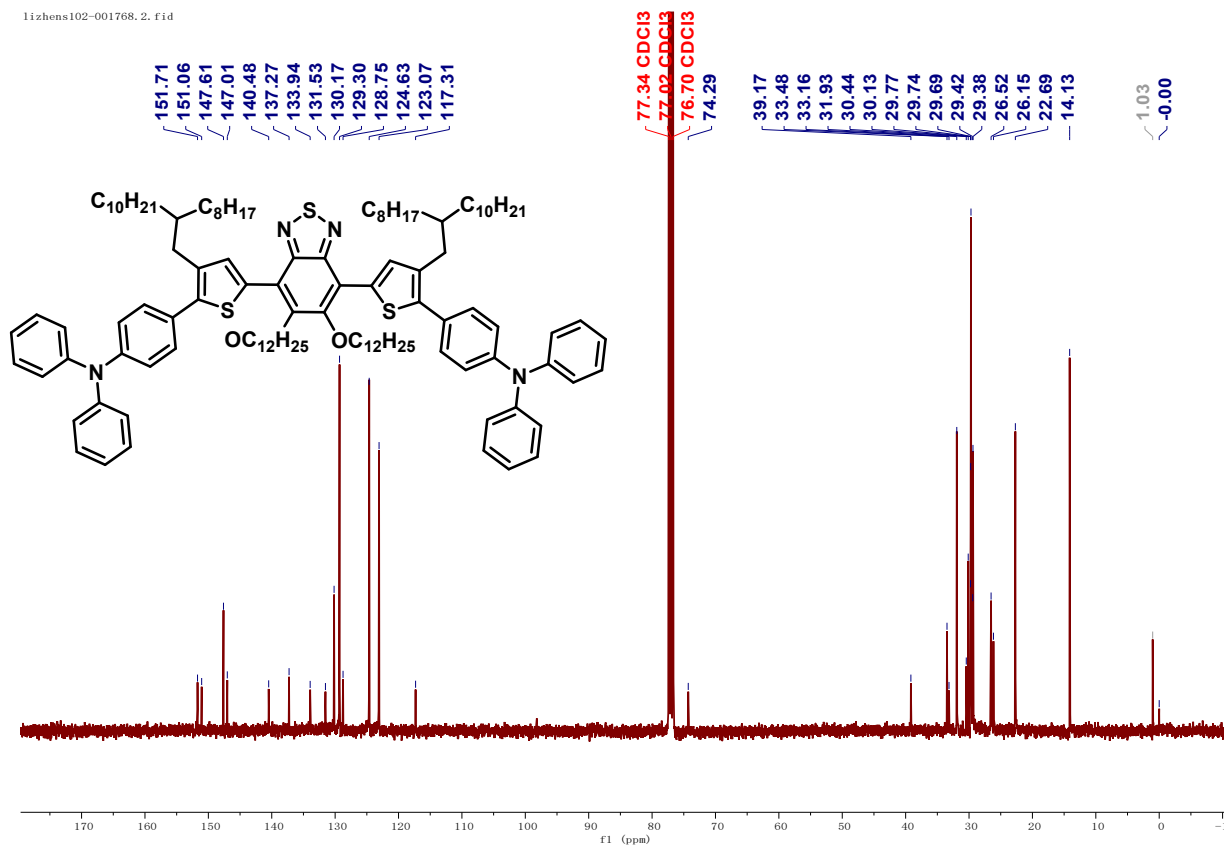


Figure S18. ^{13}C NMR spectrum of LI-164.

lizhens102-001949.1.fid

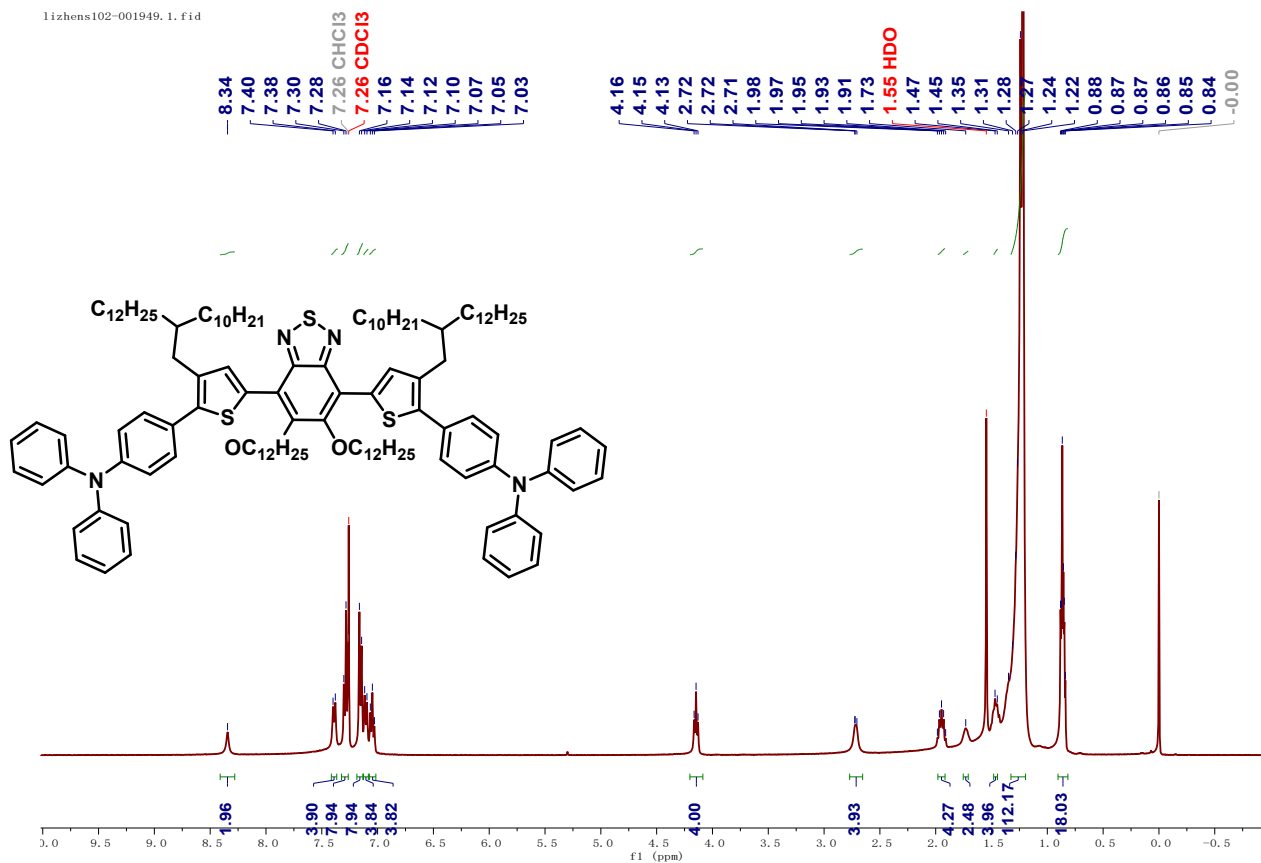


Figure S19. ^1H NMR spectrum of LI-165.

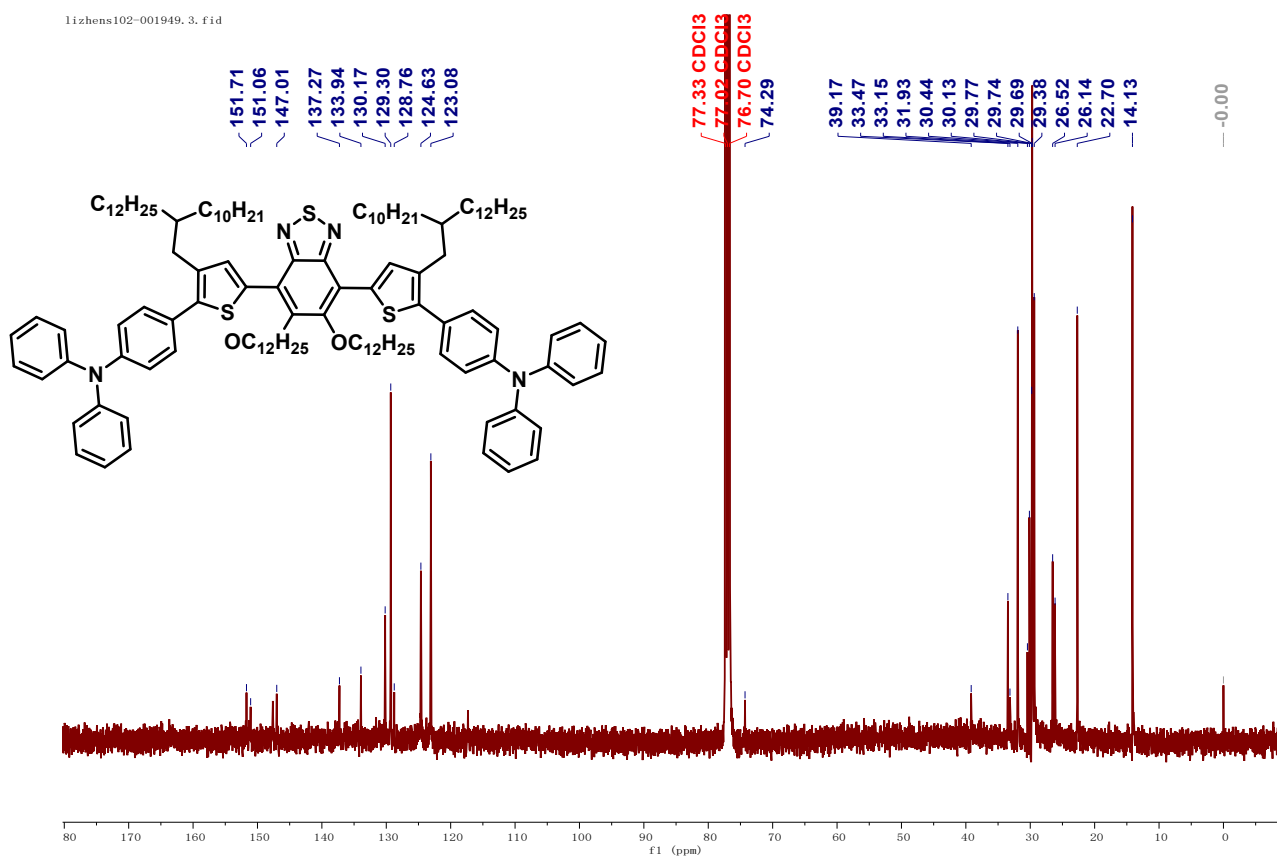
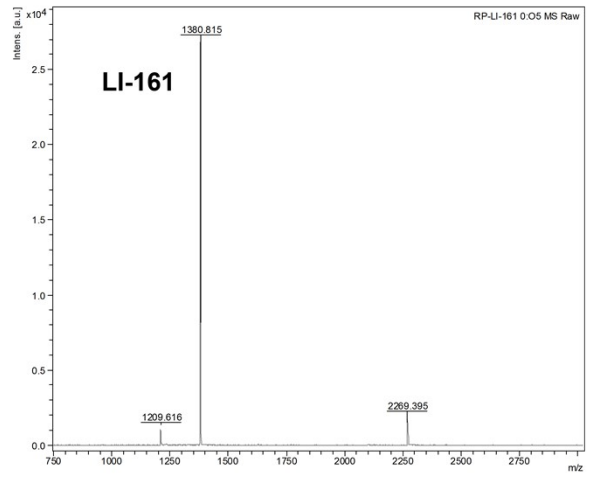
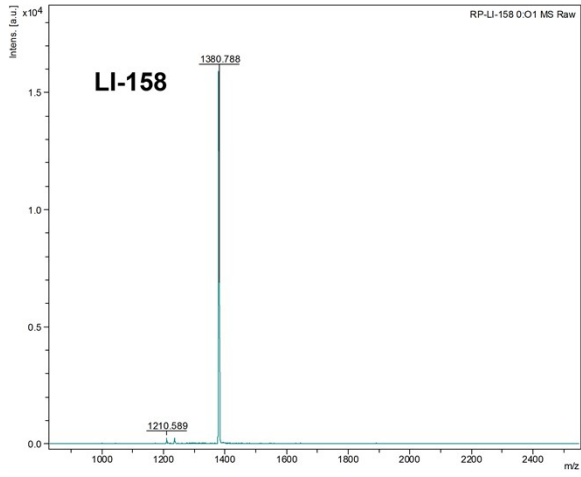
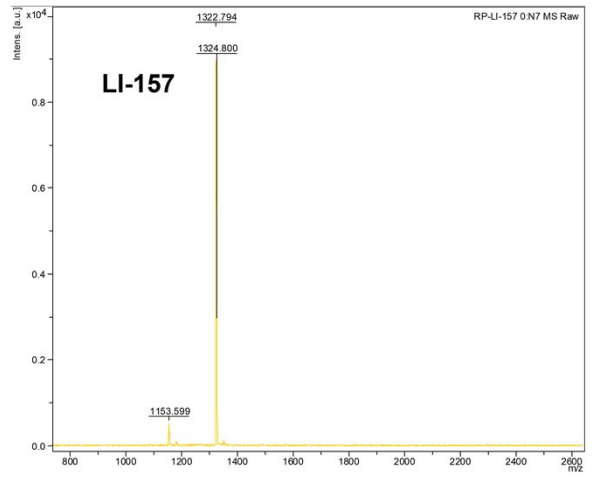
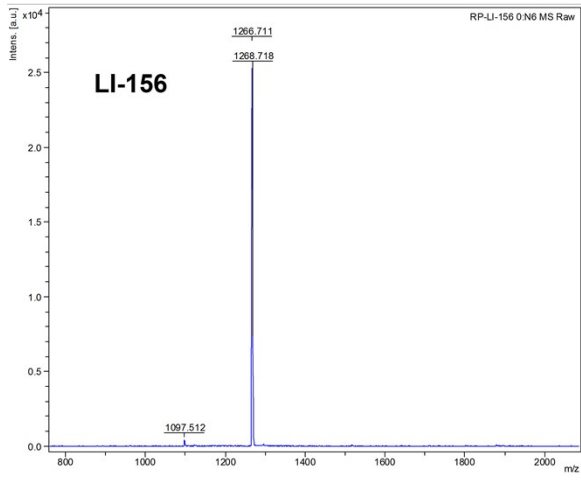
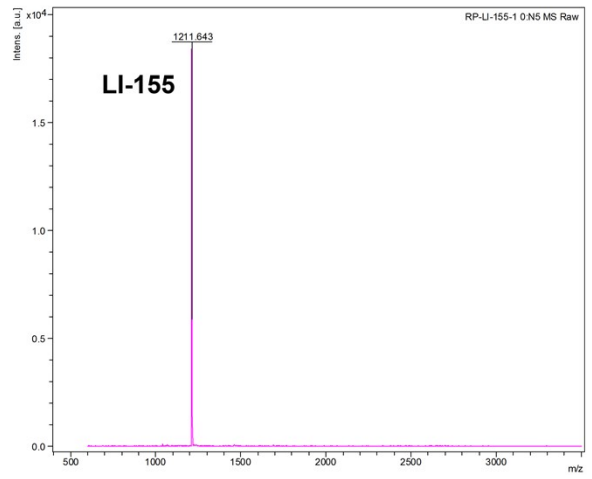
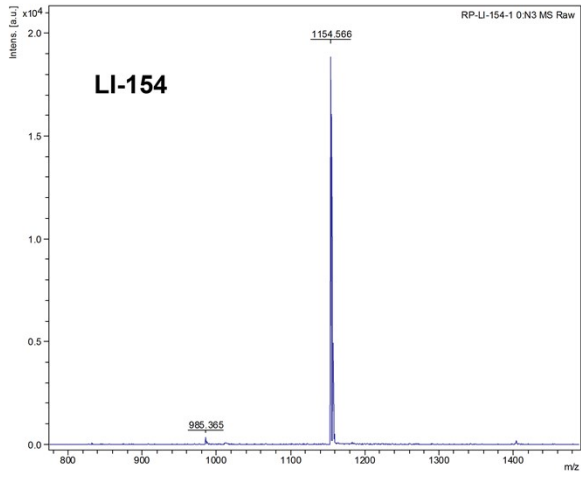


Figure S20. ¹³C NMR spectrum of LI-165.



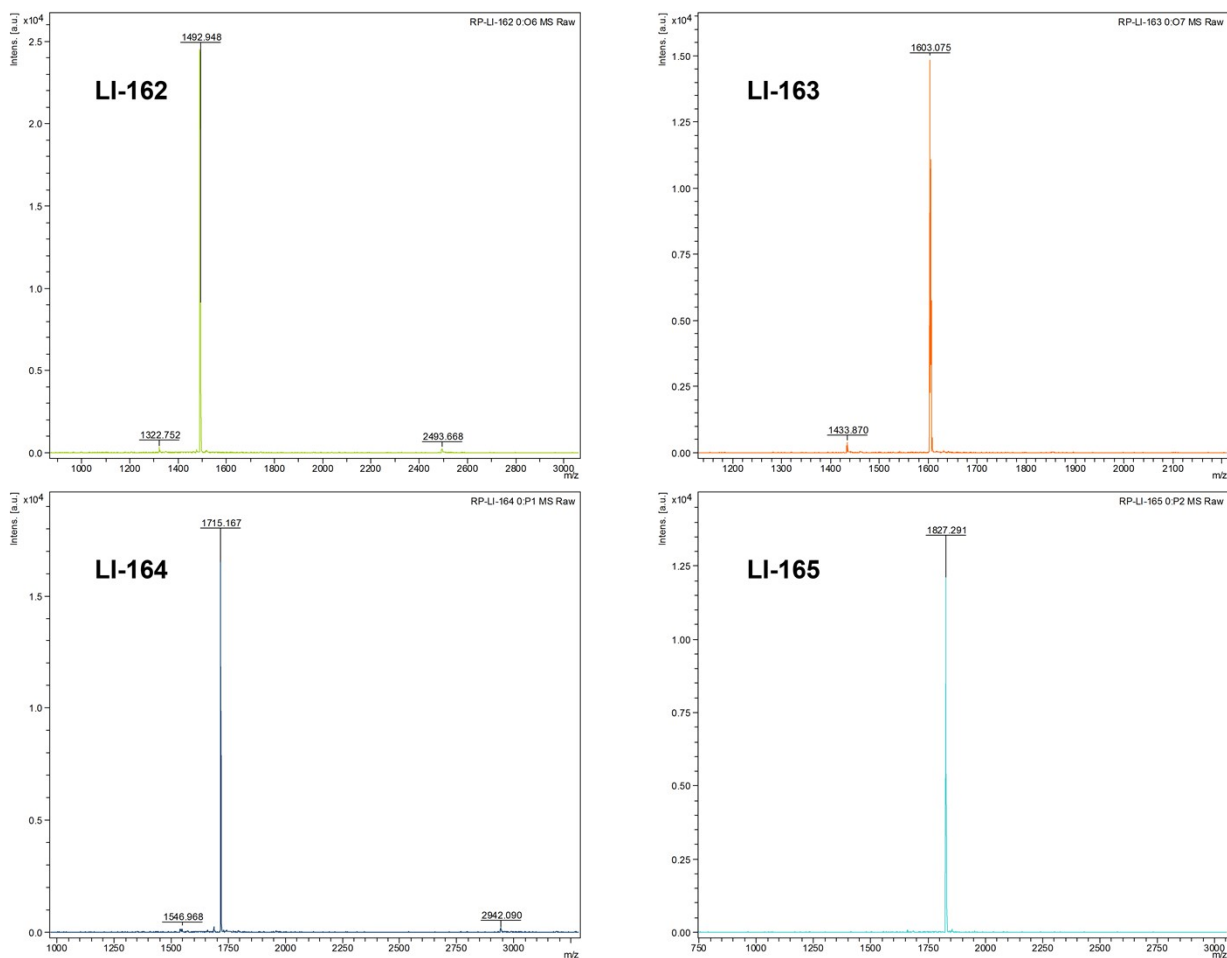


Figure S21. MALDI-TOF spectra of dyes.

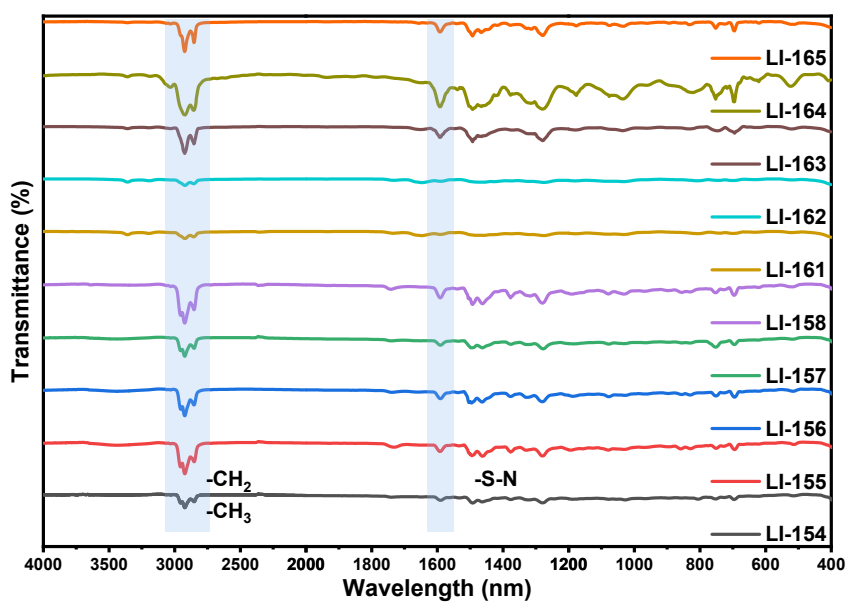


Figure S22. FTIR spectra of organic dyes in the range of 4000-400 cm^{-1} .

Photophysical property of organic dyes

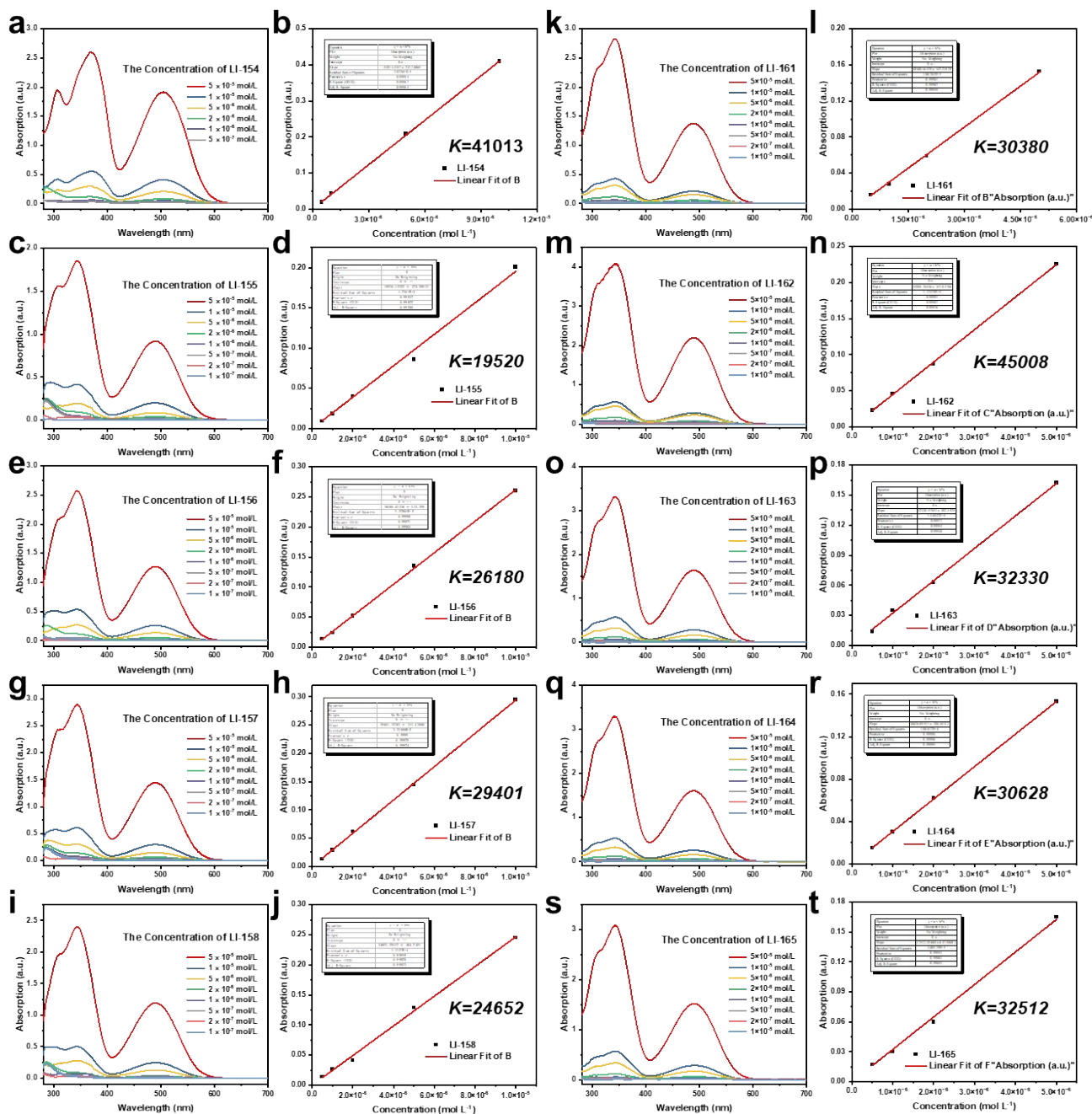


Figure S23. The UV absorption spectra of LI-154 (a), LI-155 (c), LI-156 (e), LI-157 (g), LI-158 (i), LI-161 (k), LI-162 (m), LI-163 (o), LI-164 (q) and LI-165 (s) with different concentrations in CH_2Cl_2 solution. The molar extinction coefficient (504 nm) fitting curve of LI-154 (b), the molar extinction coefficient (490 nm) fitting curve of dyes (d, f, h, j, l, n, p, r, t).

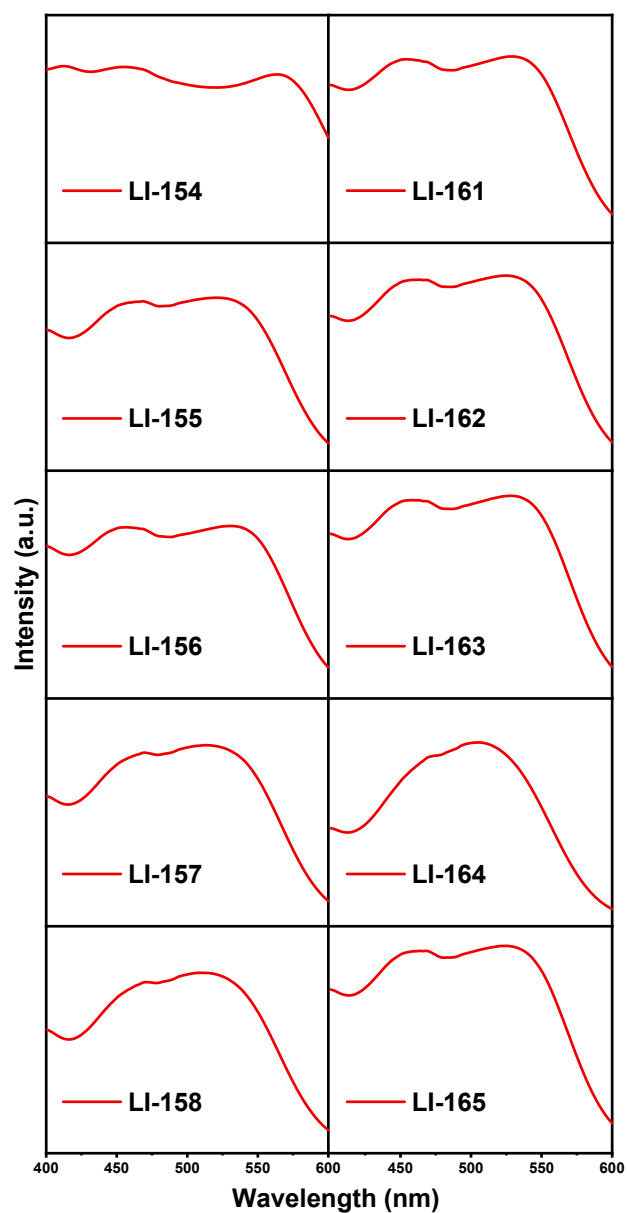


Figure S24. Excitation spectra of dyes in CH_2Cl_2 solution at the concentration of $5 \times 10^{-5} \text{ mol L}^{-1}$.

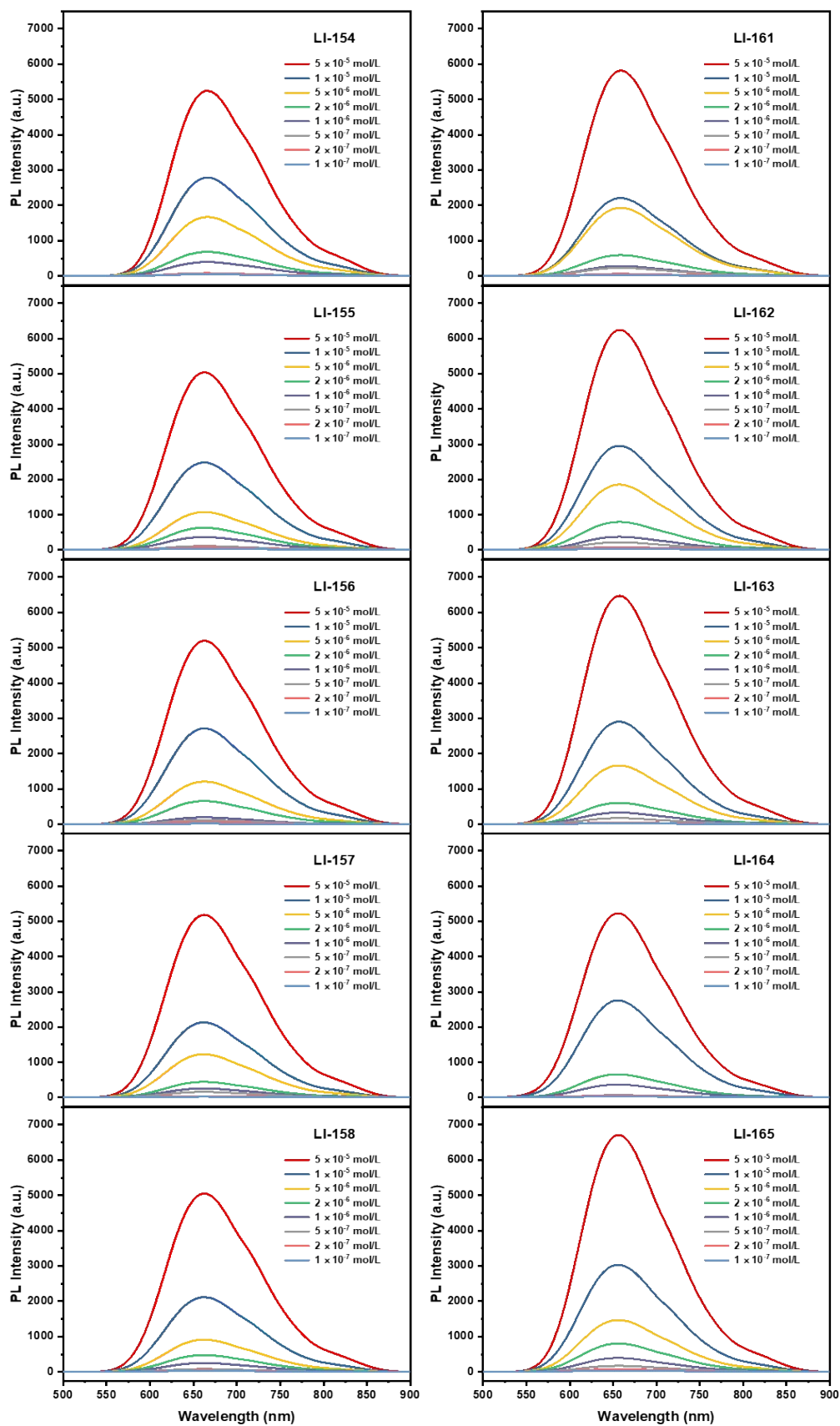


Figure S25. Photoluminescence spectra of dyes in CH_2Cl_2 solution with different concentrations ($\lambda_{\text{ex}} = 560 \text{ nm}$).

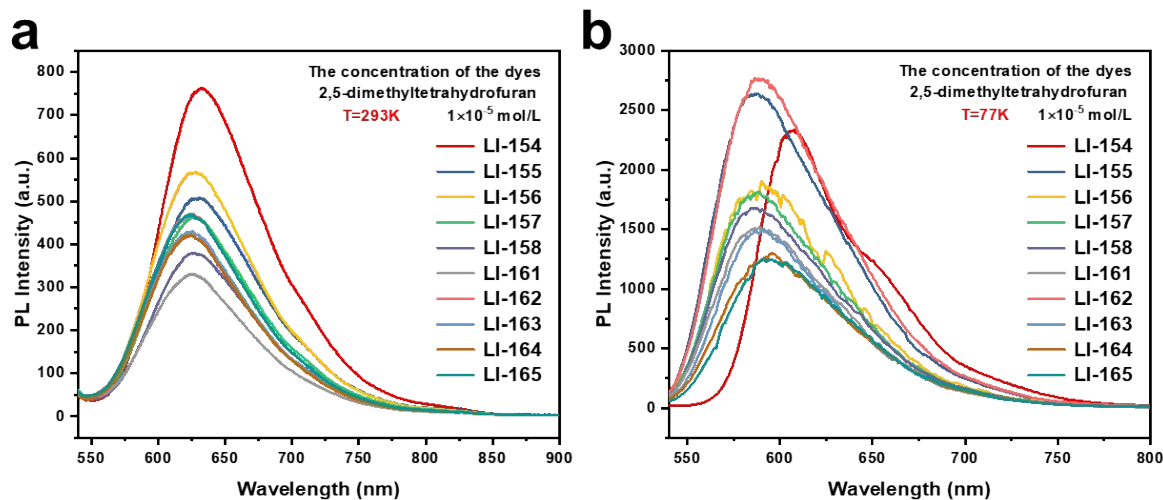


Figure S26. Photoluminescence spectra of dyes in 2,5-dimethyltetrahydrofuran solution at 293K (a) and 77 K (b) ($1 \times 10^{-5} \text{ mol L}^{-1}$, $\lambda_{\text{ex}} = 520 \text{ nm}$).

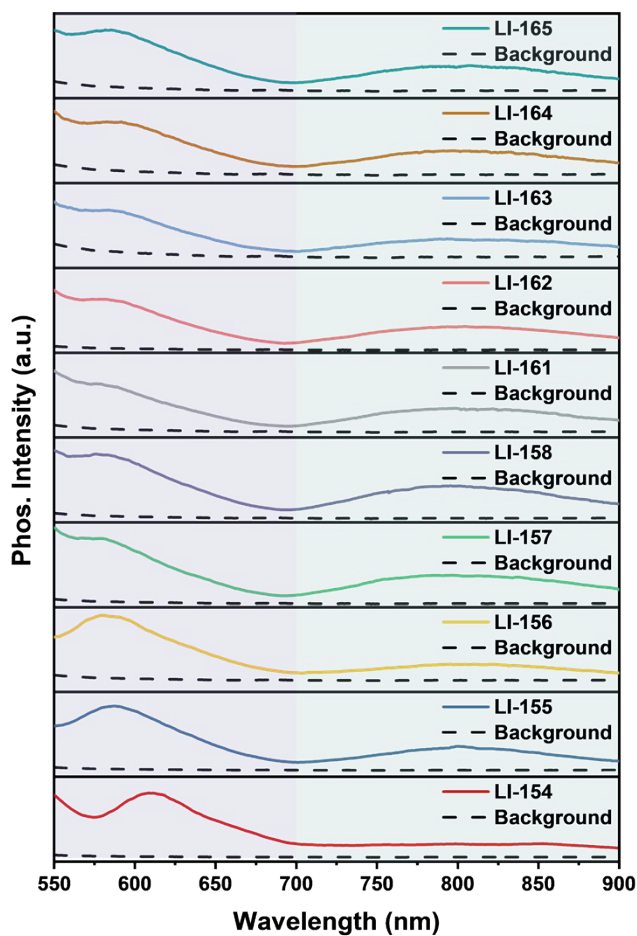


Figure S27. Phosphorescence spectra of dyes in 2,5-dimethyltetrahydrofuran solution at 77 K ($1 \times 10^{-5} \text{ mol L}^{-1}$, $\lambda_{\text{ex}} = 520 \text{ nm}$).

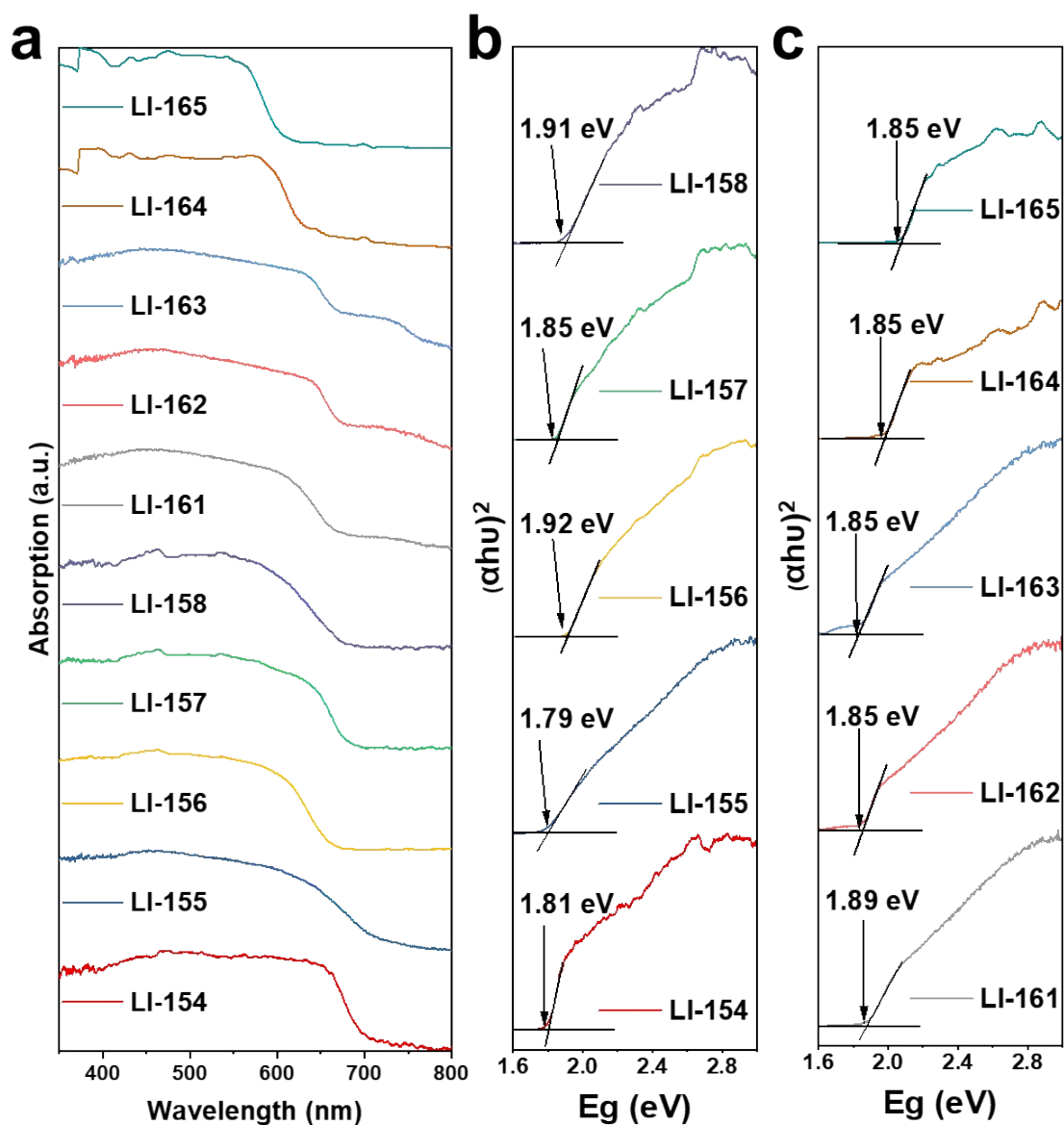
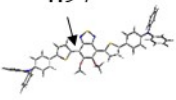
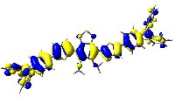
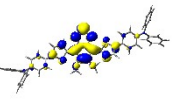
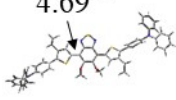
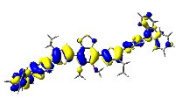
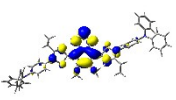
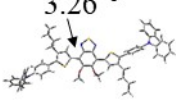
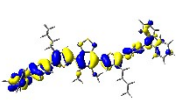
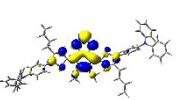
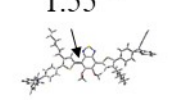
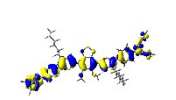
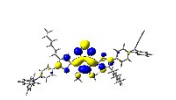
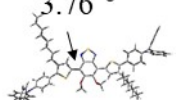
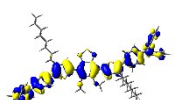
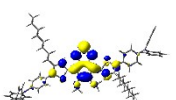
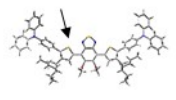
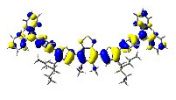
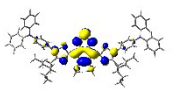
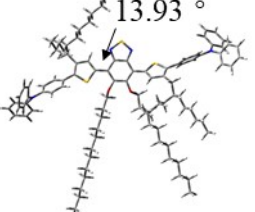
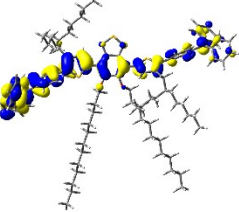
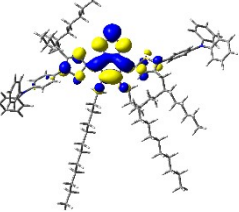


Figure S28. (a) UV-vis DRS absorption spectra of LI-154, LI-155, LI-156, LI-157, LI-158, LI-161, LI-162, LI-163 at solid state, due to LI-164 and LI-165 oily nature, a thin film UV-vis is used instead. (b, c) The bandgaps of dyes calculated by Tauc-plot.

	OPT	HOMO	LUMO
LI-154	4.97 ° 		
LI-155	4.69 ° 		
LI-156	3.26 ° 		
LI-157	1.55 ° 		
LI-158	3.76 ° 		
LI-161	13.06 ° 		
LI-162	13.93 ° 		

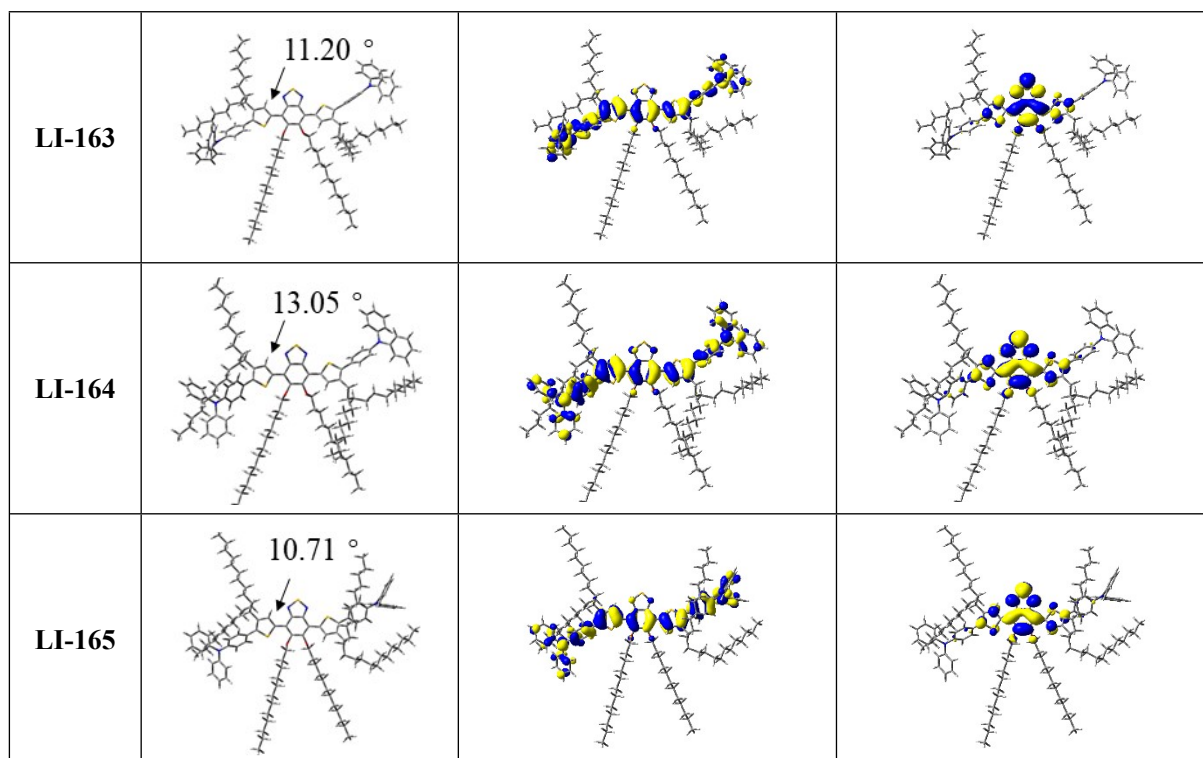


Figure S29. Optimized molecular configurations and frontier orbital distribution of organic dyes.

Table S1 Fluorescence lifetimes of organic dyes

Sample	τ_1 (ns)	A_1	τ_2 (ns)	A_2	τ_{ave} (ns)
LI-154	1.30	0.35	4.74	0.65	3.53
LI-155	1.96	0.52	4.22	0.48	3.05
LI-156	1.44	0.52	4.62	0.48	2.97
LI-157	2.78	0.34	7.65	0.66	6.01
LI-158	2.67	0.33	6.85	0.67	5.48
LI-161	1.82	0.50	4.90	0.50	3.35
LI-162	3.10	0.44	8.85	0.56	6.33
LI-163	3.32	0.47	8.92	0.53	6.31
LI-164	1.55	0.43	4.13	0.57	3.02
LI-165	1.64	0.63	3.72	0.37	2.40

For organic dyes **LI-154-LI-158** and **LI-161-LI-163**, fluorescence lifetimes were measured at solid state, for **LI-164** and **LI-165** with oily nature, a thin film is used instead. ($\lambda_{ex} = 375$ nm, $\lambda_{em} = 690$ nm).

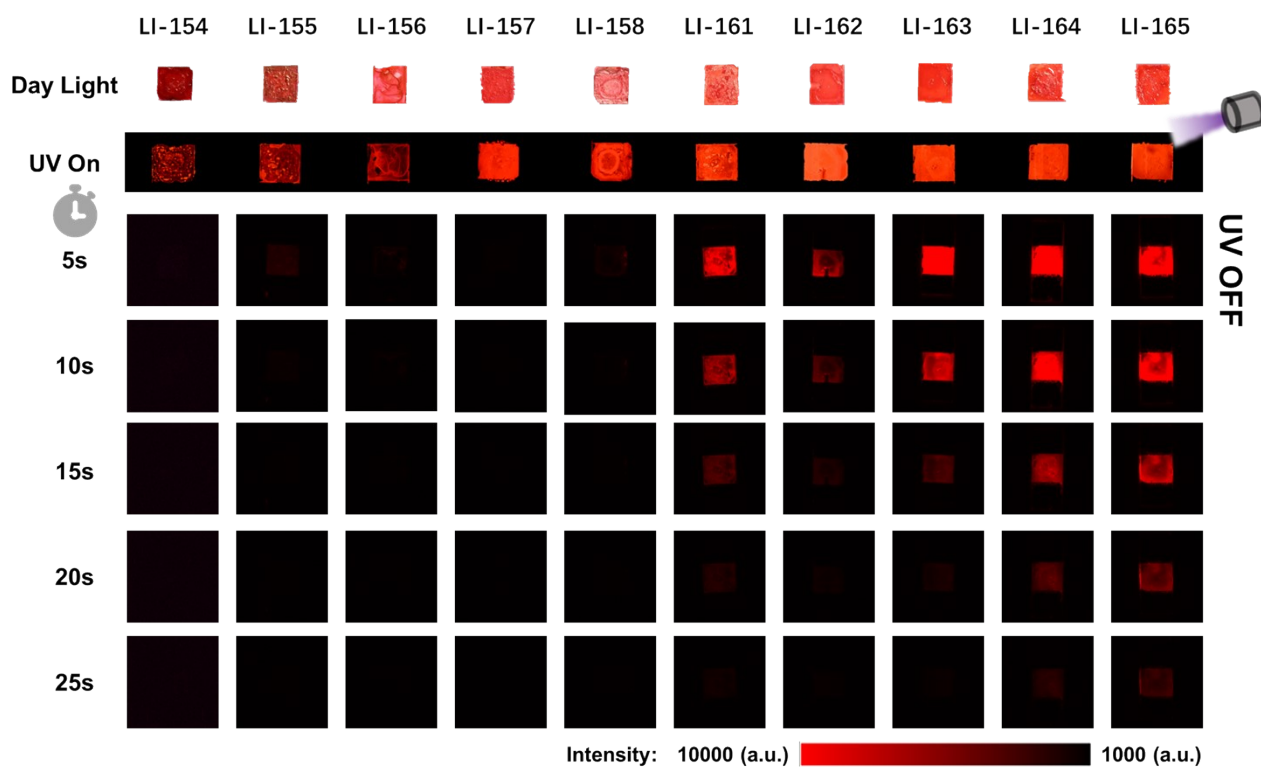


Figure S30. Photographs of organic dyes in thin film under different conditions captured by a camera, and afterglow imaging captured by a chemiluminescence imager (colors only represent phosphorescence intensity) at different time after UV off.

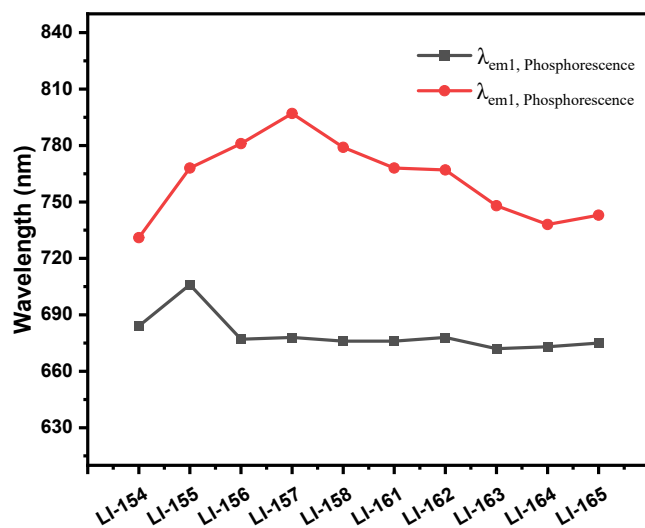


Figure S31. The phosphorescence emission wavelengths of organic dyes.

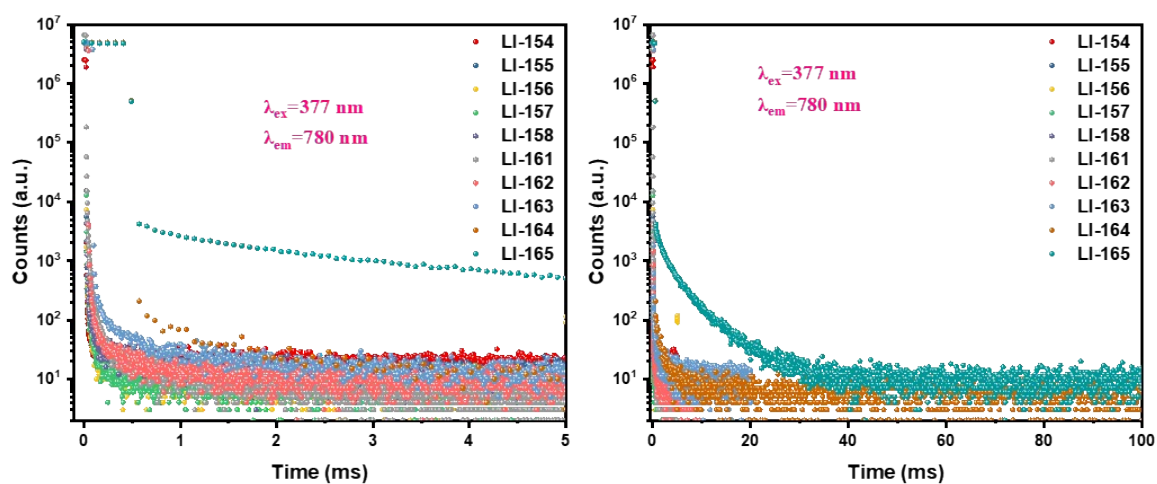


Figure S32. Phosphorescence decay of organic dyes in thin film. ($\lambda_{\text{ex}} = 377 \text{ nm}$, $\lambda_{\text{em}} = 780 \text{ nm}$).

Table S2 Phosphorescence lifetimes of organic dyes in thin film ($\lambda_{\text{ex}} = 377 \text{ nm}$, $\lambda_{\text{em}} = 780 \text{ nm}$)

Sample	τ_1 (ms)	A_1	τ_2 (ms)	A_2	τ_3 (ms)	A_3	τ_{ave} (ms)
LI-154	0.004	0.470	0.037	0.051	0.730	0.015	0.027
LI-155	0.006	0.590	0.046	0.139	0.460	0.017	0.024
LI-156	0.006	0.380	0.039	0.108	0.630	0.008	0.023
LI-157	0.004	2.754	0.050	0.152			0.006
LI-158	0.005	1.870	0.045	0.196	1.017	0.021	0.019
LI-161	0.007	2.32	0.044	0.227	0.730	0.027	0.018
LI-162	0.017	0.670	0.350	0.041			0.036
LI-163	0.057	0.380	1.190	0.046			0.179
LI-164	0.660	0.081	4.600	0.014			1.254
LI-165	1.254	2.339	5.256	0.949			2.409

Characterization of PCN, PCN/Pt and dye/PCN/Pt

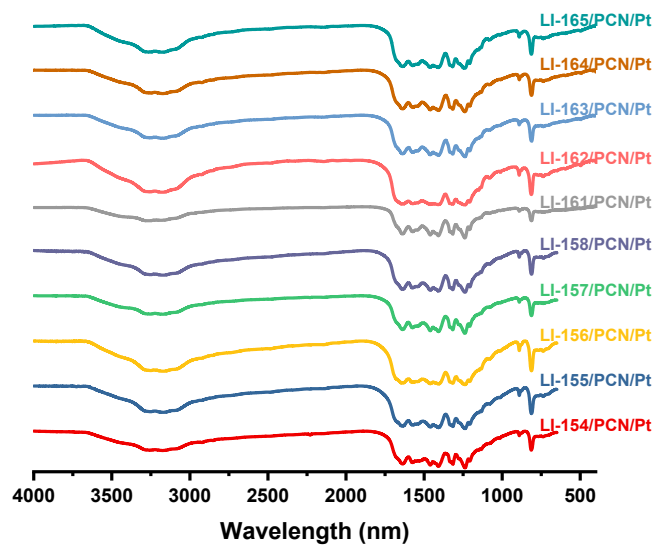


Figure S33. FTIR spectra of PCN, PCN/Pt and dye/PCN/Pt.

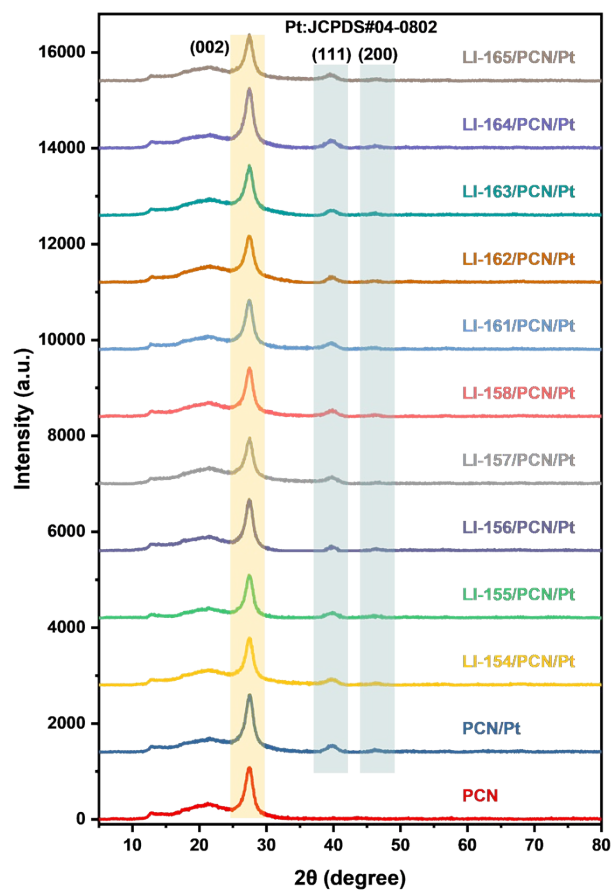


Figure S34. XRD patterns of PCN, PCN/Pt and dye/PCN/Pt.

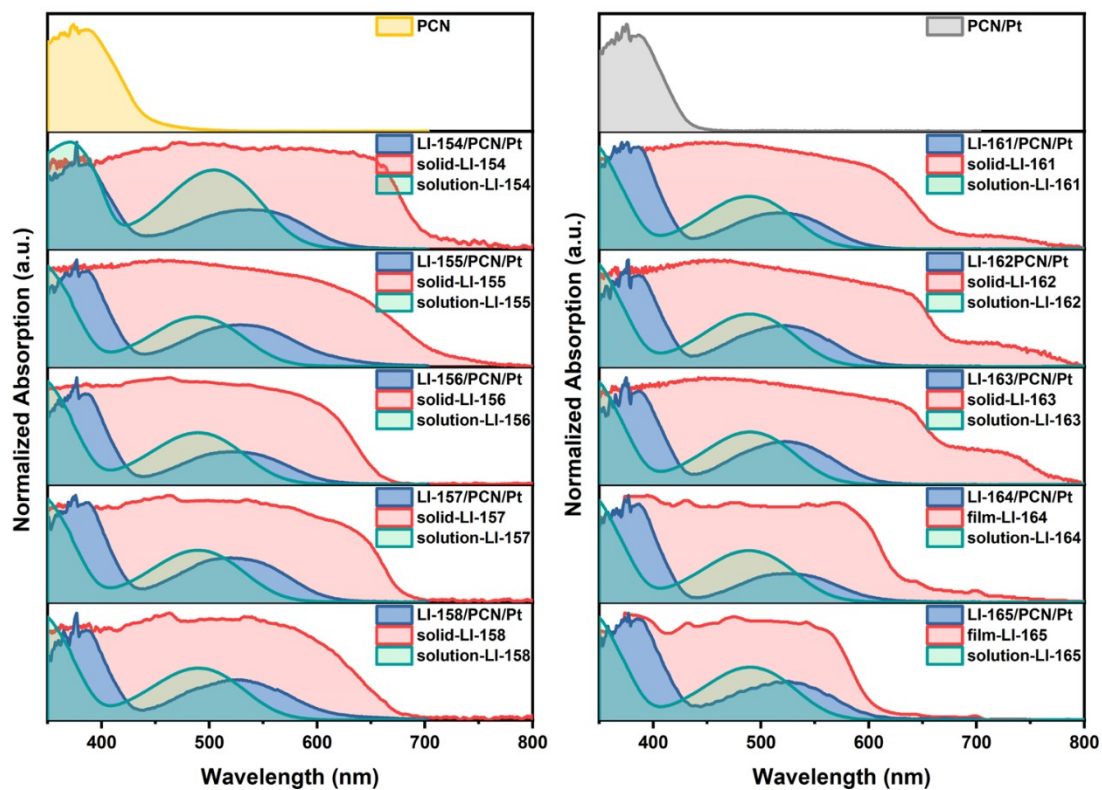


Figure S35. UV-vis DRS absorption spectra of dye/PCN/Pt (blue line), dye at solid or film state (red line), and UV-vis absorption spectra of dye in CH_2Cl_2 solution ($1 \times 10^{-5} \text{ mol L}^{-1}$, green line).

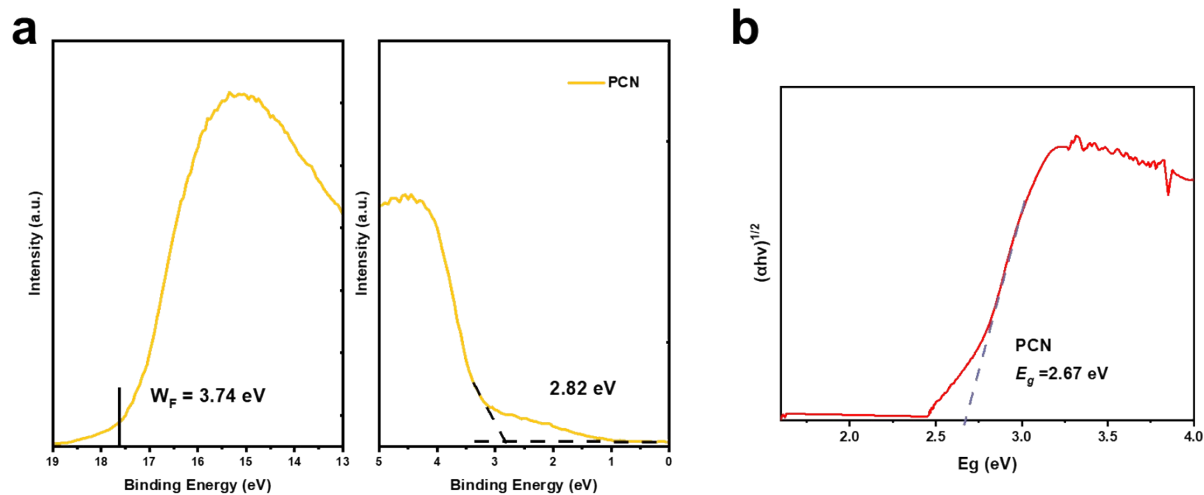


Figure S36. (a) UPS spectra of PCN, HOMO energy level is calculated according to the following equation: E_{HOMO} (vs. vacuum conditions) = $-(E_{\text{photon}} - \text{width of UPS spectra})$, where E_{photon} denotes the excitation energy of He I (21.22 eV). (b) The bandgaps of PCN calculated by Tauc-plot.

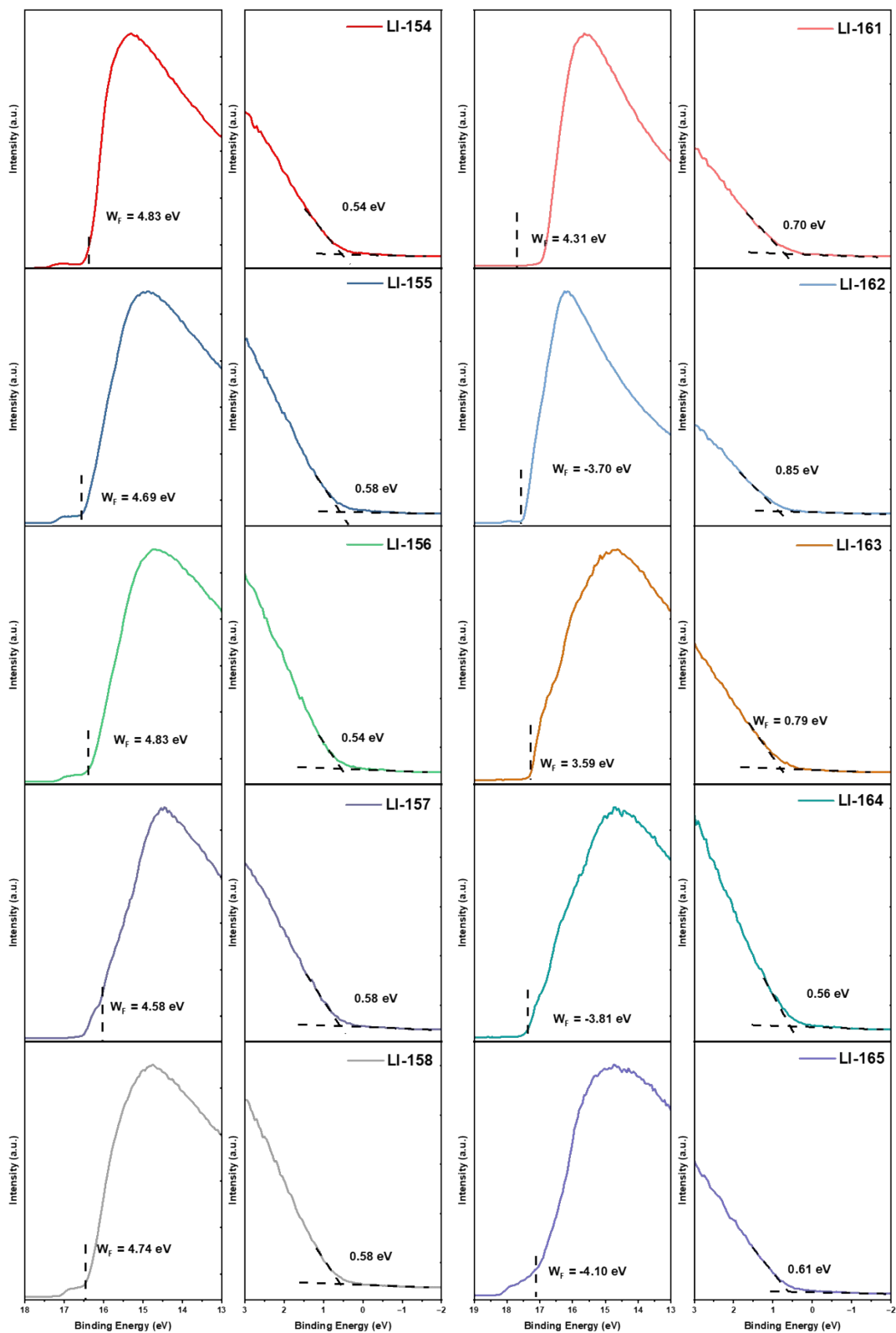


Figure S37. UPS spectra of dyes, HOMO energy levels are calculated according to the following equation: E_{HOMO} (vs. vacuum conditions) = $-(E_{\text{photon}} - \text{width of UPS spectra})$, where E_{photon} denotes the excitation energy of He I (21.22 eV).

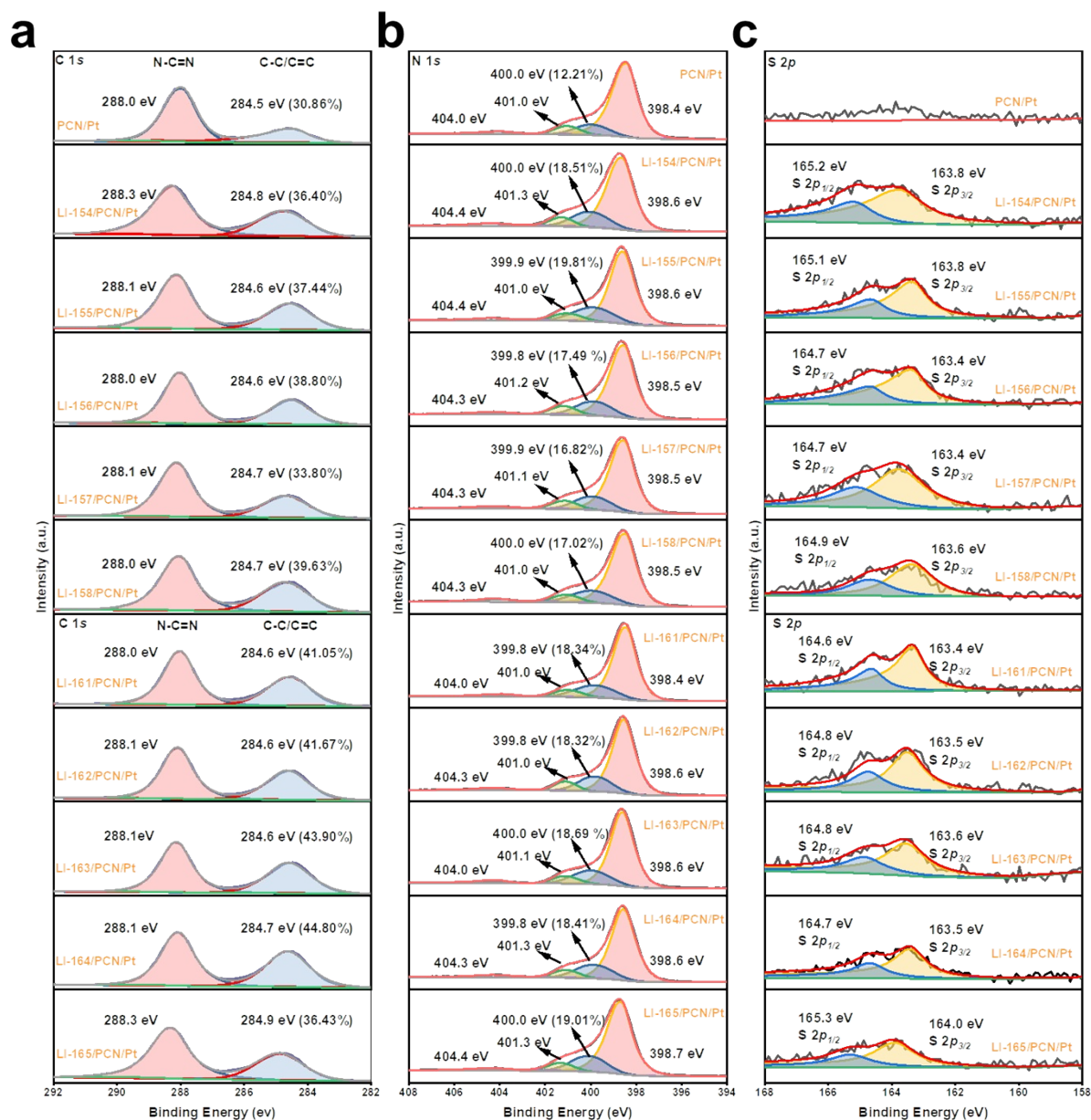


Figure S38. High-resolution XPS spectra of PCN/Pt and dye/PCN/Pt: (a) C 1 s, (b) N 1 s, and (c) S 2p.

Table S3. Relative ratios of two carbon species determined by C 1s spectra

Sample	N-C=N			C=C/C-C		
	Binding	Area	%	Binding	Area	%
	Energy (eV)			Energy (eV)		
PCN/Pt	288.0	106668	69.1%	284.5	47726	30.9%
LI-154/PCN/Pt	288.3	137729	63.6%	284.8	79048	36.4%
LI-155/PCN/Pt	288.1	118849	62.6%	284.6	71286	37.4%
LI-156/PCN/Pt	288.0	121050	61.2%	284.6	76909	38.8%
LI-157/PCN/Pt	288.1	118045	66.2%	284.7	60352	33.8%
LI-158/PCN/Pt	288.0	110030	60.4%	284.7	72394	39.6%
LI-161/PCN/Pt	288.0	143876	59.0%	284.6	100421	41.0%
LI-162/PCN/Pt	288.1	114146	58.3%	284.6	81732	41.7%
LI-163/PCN/Pt	288.1	124327	56.1%	284.6	97583	43.9%
LI-164/PCN/Pt	288.1	95069	55.2%	284.7	77438	44.8%
LI-165/PCN/Pt	288.3	115427	63.6%	284.9	66298	36.4%

Table S4. Relative ratios of four nitrogen species determined by N 1s spectra

Sample	π - π excitations			Quaternary N			Tertiary N			sp ² N		
	Bindin			Bindin			Bindin			Bindin		
	g	Area	%	g	Area	%	g	Area	%	g	Area	%
	Energy (eV)			Energy (eV)			energy (eV)			Energy (eV)		
PCN/Pt	404.0	1522	5.8	401.0	2030	7.7	400.0	3221	12.2	398.4	19629	74.3
		7	%		5	%		6	%		3	%
LI- 154/PCN/Pt	404.4	1759	5.8	401.3	2211	7.2	400.0	5682	18.5	398.6	21053	68.5
		1	%		0	%		9	%		7	%
LI- 155/PCN/Pt	404.4	1450	5.4	401.0	2075	7.7	399.9	5329	19.8	398.6	18060	67.1
		1	%		9	%		9	%		6	%
LI- 156/PCN/Pt	404.3	1447	5.3	401.2	2123	7.8	399.8	4753	17.5	398.5	18866	69.4
		0	%		2	%		3	%		2	%
LI- 157/PCN/Pt	404.3	1634	6.1	401.1	2131	7.9	399.9	4532	16.8	398.5	18662	69.2
		4	%		2	%		1	%		1	%
LI- 158/PCN/Pt	404.3	1387	5.9	401.0	1687	7.1	400.0	4024	17.0	398.5	16558	70.0
		0	%		0	%		2	%		0	%
LI- 161/PCN/Pt	404.0	1707	5.5	401.0	2274	7.3	399.8	5753	18.3	398.4	21945	69.0
		8	%		1	%		5	%		9	%
LI- 162/PCN/Pt	404.3	1362	5.4	401.0	1942	7.7	399.8	4619	18.3	398.6	17301	68.6
		7	%		6	%		7	%		9	%
LI- 163/PCN/Pt	404.0	1398	5.2	401.1	1930	7.2	400.0	5044	18.7	398.6	18630	69.0
		8	%		6	%		6	%		8	%
LI- 164/PCN/Pt	404.3	1233	6.0	401.3	1543	7.4	399.8	3825	18.4	398.6	14187	68.2
		6	%		2	%		5	%		9	%
LI- 165/PCN/Pt	404.4	1334	5.2	401.3	1830	7.0	400.0	4934	19.0	398.7	17875	68.8
		4	%		3	%		7	%		1	%

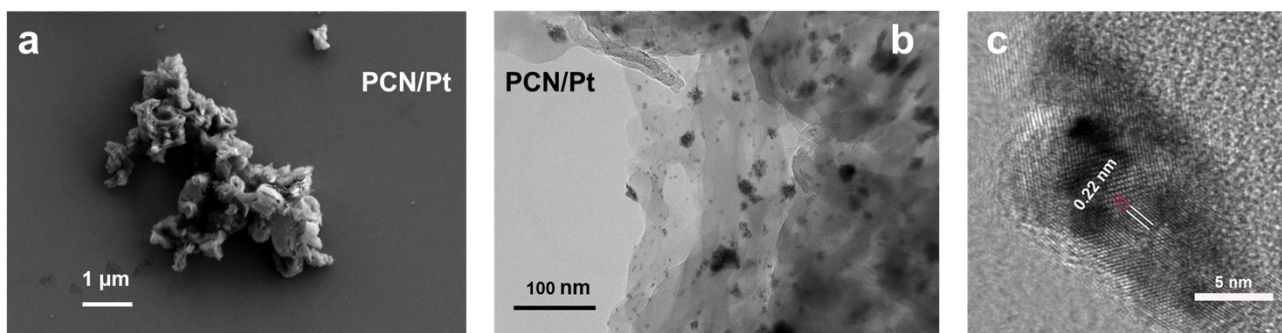


Figure S39. SEM and TEM images of PCN (a) and PCN/Pt (b). The Pt morphology on the surface of PCN (c).

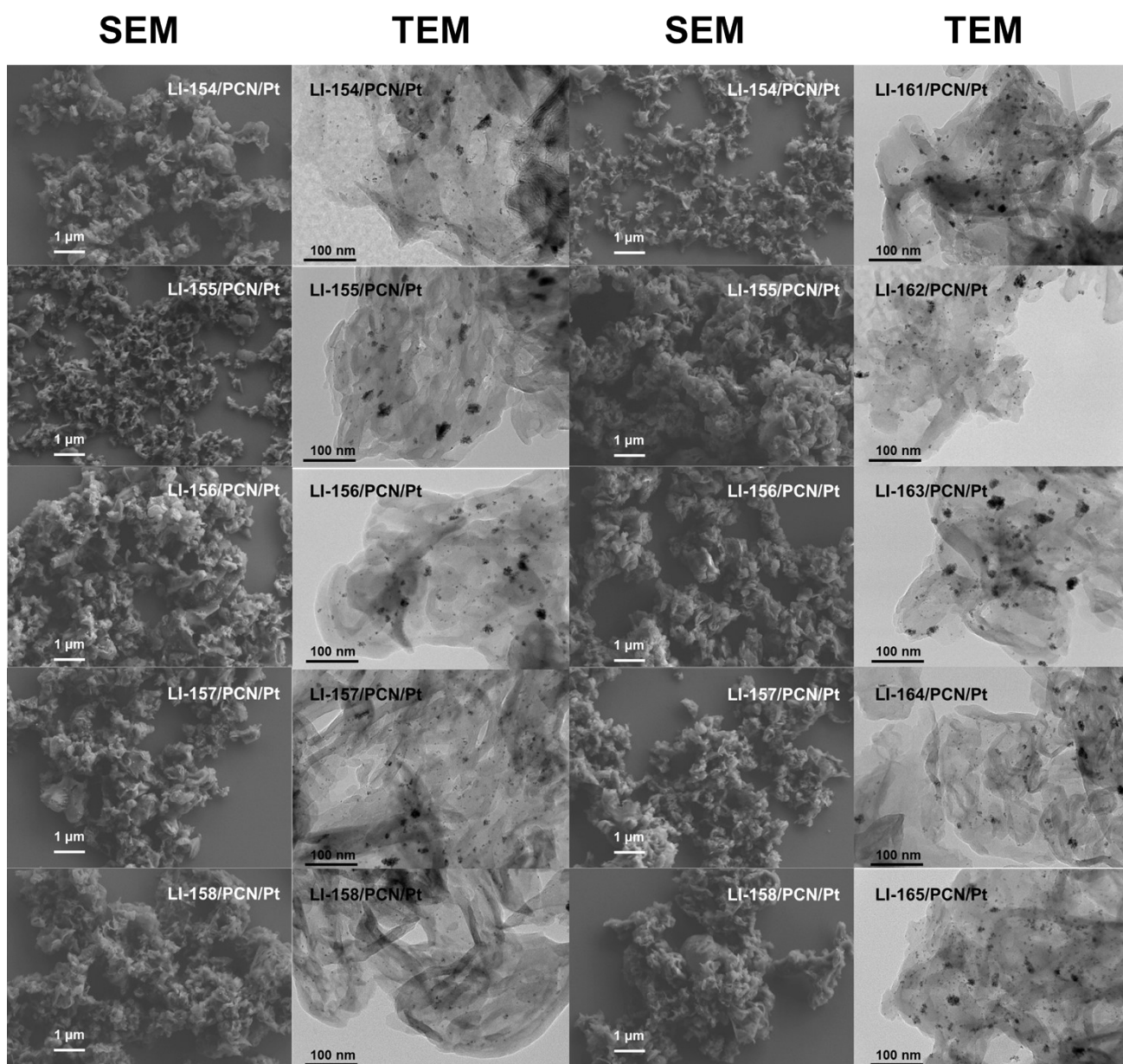


Figure S40. SEM and TEM images of dye/PCN/Pt.

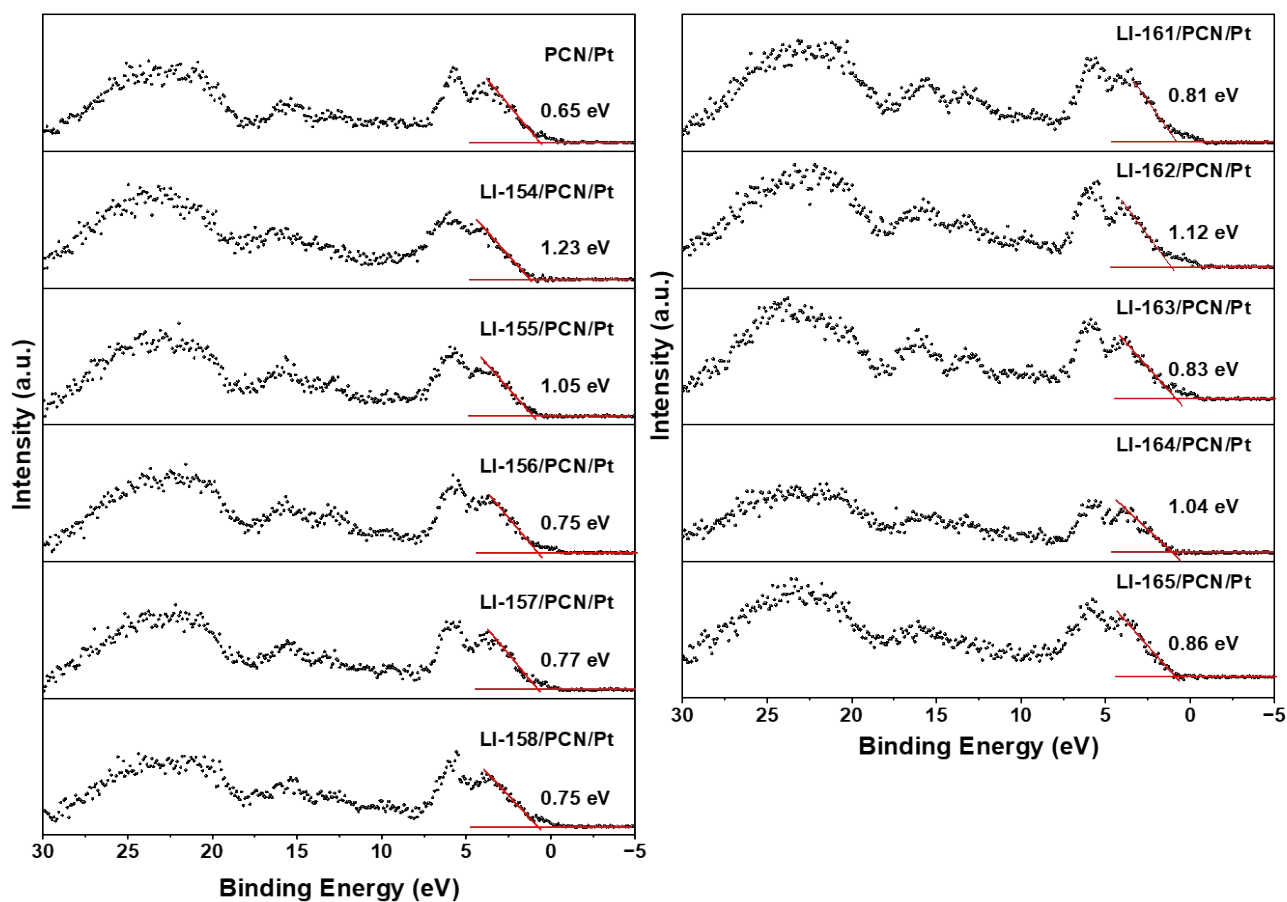


Figure S41. VB XPS spectra of PCN/Pt and dye/PCN/Pt.

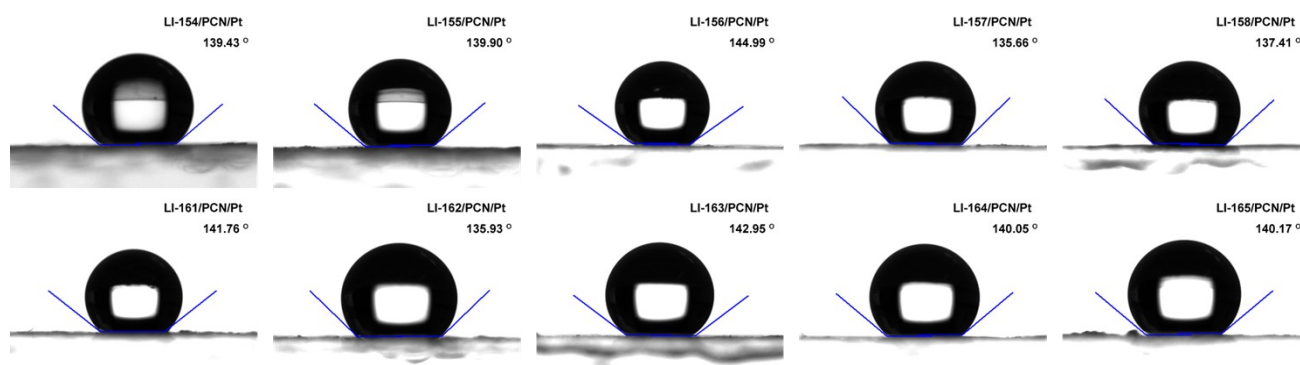


Figure S42. The water contact angles of dye/PCN/Pt electrodes.

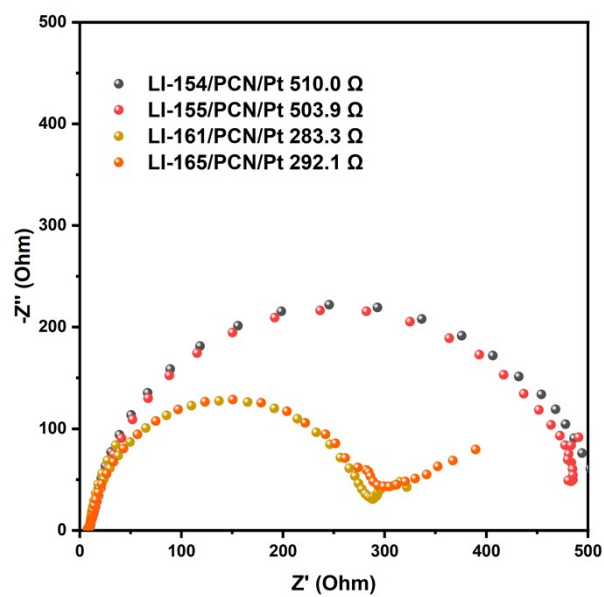


Figure S43. Electrochemical impedance spectra (EIS) Nyquist plots of dye/PCN/Pt (-1.2 V vs. RHE).

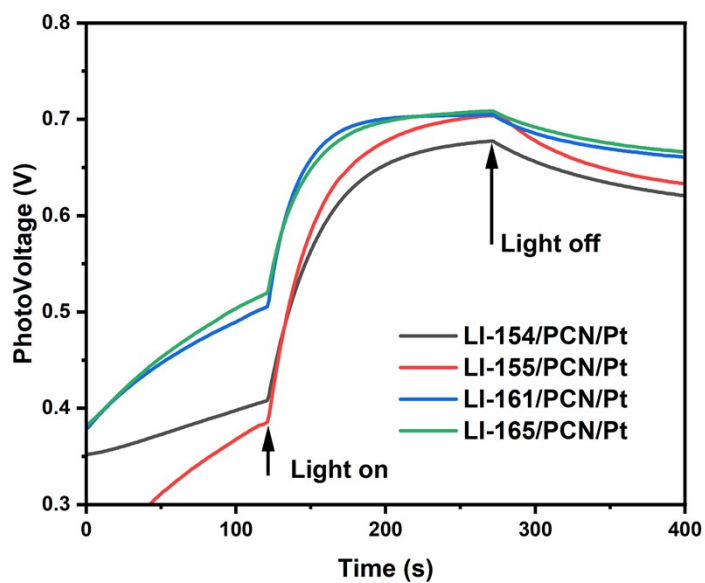


Figure S44. The transient open-circuit voltage decay (OCVD) of dye/PCN/Pt.

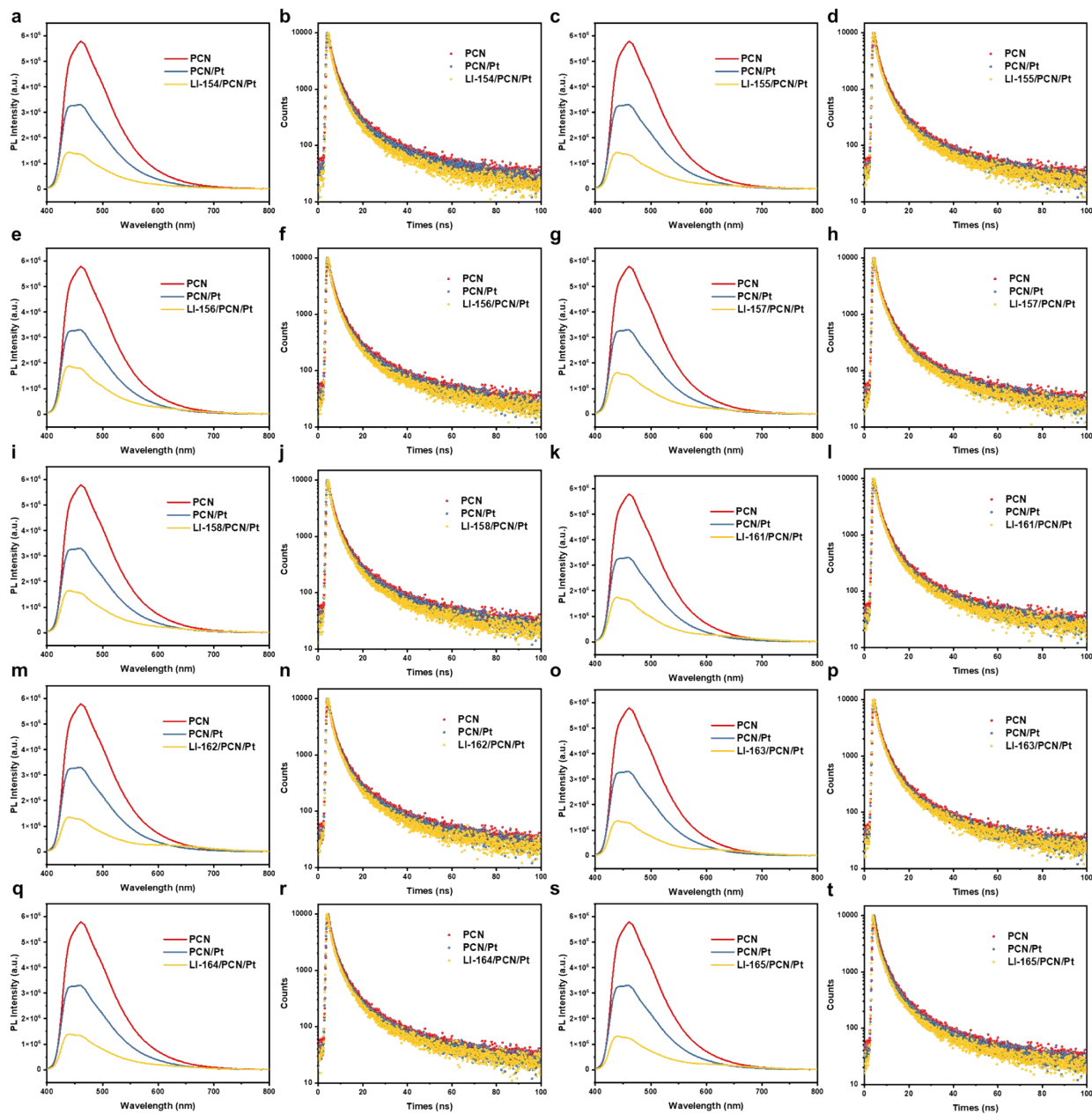
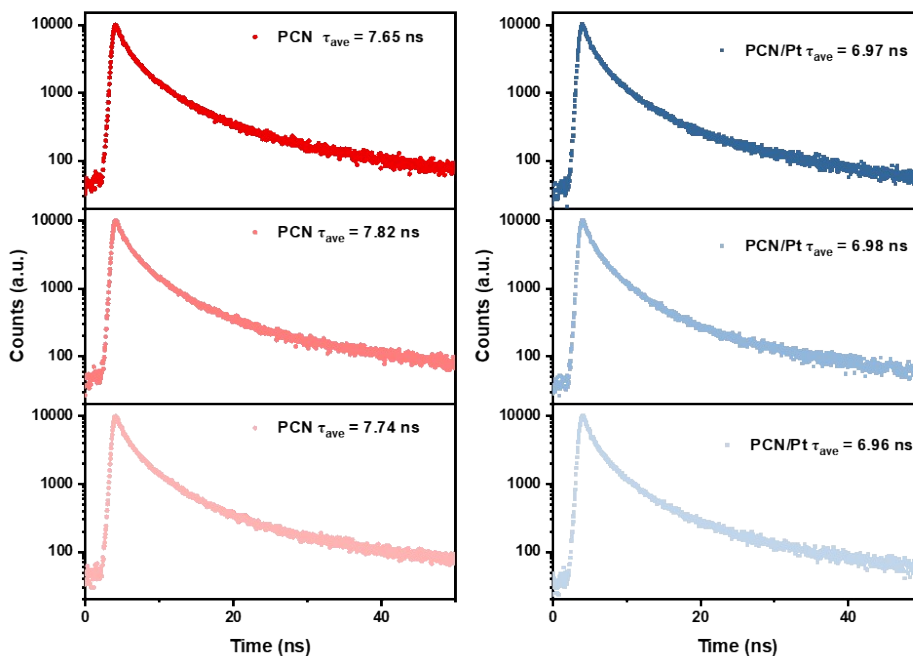


Figure S45. PL spectra of PCN, PCN/Pt and dye/PCN/Pt ($\lambda_{\text{ex}} = 350$ nm) (a, c, e, g, i, k, m, o, q, s). TRPL spectra of PCN, PCN/Pt, dye/PCN and dye/PCN/Pt ($\lambda_{\text{ex}} = 375$ nm, $\lambda_{\text{em}} = 450$ nm) (b, d, f, h, j, l, n, p, r, t)

Table S5. Fluorescence lifetimes of PCN, PCN/Pt and dye/PCN/Pt ($\lambda_{\text{ex}} = 375 \text{ nm}$, $\lambda_{\text{em}} = 450 \text{ nm}$)

Sample	τ_1 (ns)	A_1	τ_2 (ns)	A_2	τ_3 (ns)	A_3	τ_{ave} (ns)
PCN	1.17	0.27	4.21	0.50	18.89	0.23	7.74
PCN/Pt	1.17	0.25	4.11	0.50	17.49	0.25	6.97
LI-154/PCN/Pt	1.01	0.28	3.79	0.51	17.01	0.21	5.85
LI-155/PCN/Pt	1.10	0.27	4.02	0.52	18.30	0.21	6.22
LI-156/PCN/Pt	1.08	0.22	3.98	0.52	17.75	0.22	6.14
LI-157/PCN/Pt	1.12	0.28	3.91	0.50	17.23	0.22	6.06
LI-158/PCN/Pt	1.07	0.28	3.90	0.51	17.20	0.21	5.95
LI-161/PCN/Pt	1.08	0.27	3.71	0.50	16.07	0.23	5.87
LI-162/PCN/Pt	1.14	0.30	3.94	0.49	17.16	0.21	5.90
LI-163/PCN/Pt	1.18	0.30	3.99	0.48	17.49	0.22	6.11
LI-164/PCN/Pt	1.28	0.34	4.42	0.47	19.62	0.19	6.28
LI-165/PCN/Pt	1.13	0.30	3.92	0.49	17.35	0.21	5.95

**Figure S46.** PL decay of PCN and PCN/Pt with three parallel experiments.

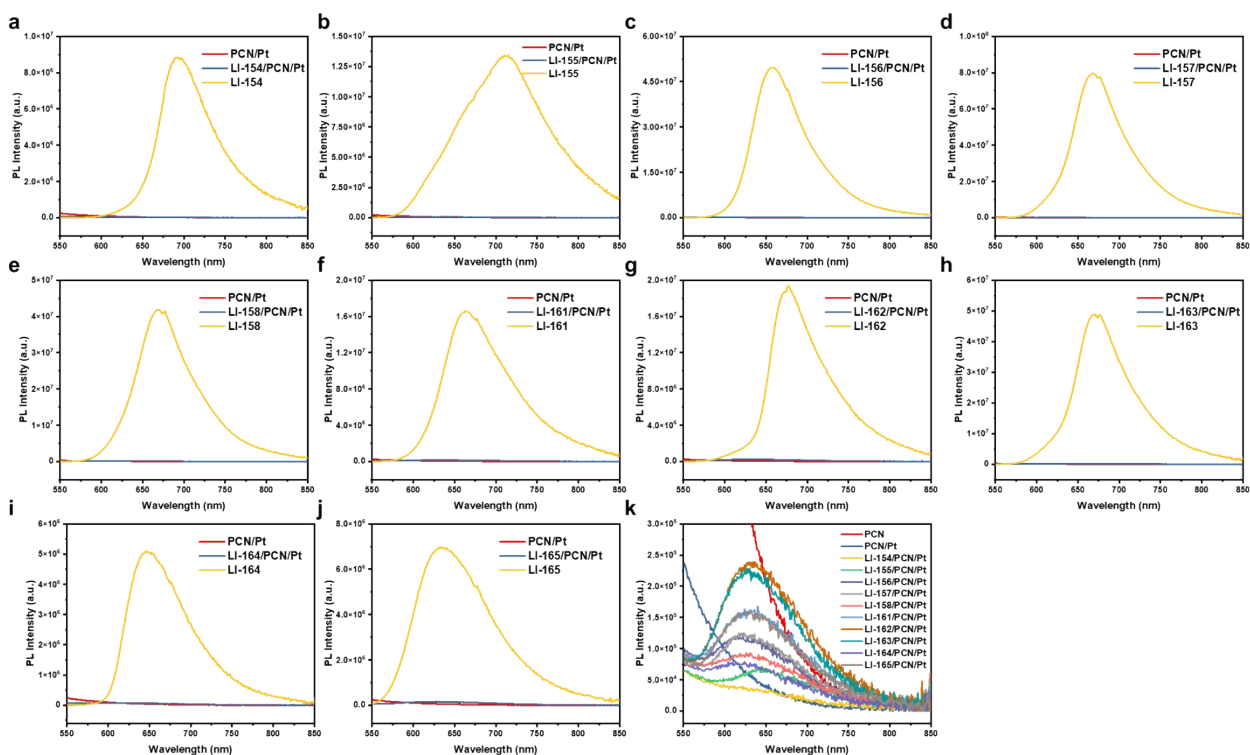


Figure S47. PL spectra of dye, PCN/Pt and dye/PCN/Pt ($\lambda_{\text{ex}} = 430 \text{ nm}$).

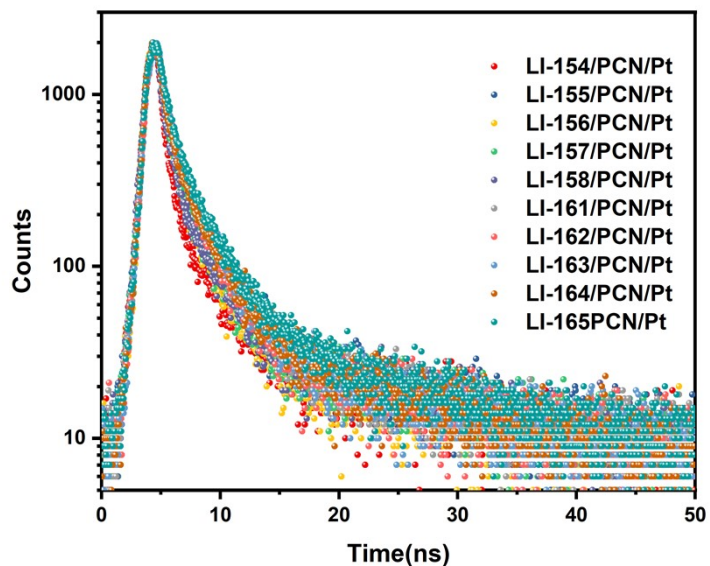


Figure S48. PL decay of dye/PCN/Pt ($\lambda_{\text{ex}} = 450 \text{ nm}$, $\lambda_{\text{em}} = 660 \text{ nm}$)

Table S6. Fluorescence lifetimes of dye/PCN/Pt ($\lambda_{\text{ex}} = 450 \text{ nm}$, $\lambda_{\text{em}} = 660 \text{ nm}$)

Sample	τ_1 (ns)	A_1	τ_2 (ns)	A_2	τ_{ave} (ns)
LI-154/PCN/Pt	0.72	0.66	5.24	0.34	2.24
LI-155/PCN/Pt	0.98	0.68	6.04	0.32	2.58
LI-156/PCN/Pt	0.98	0.72	5.22	0.28	2.15
LI-157/PCN/Pt	0.97	0.69	5.39	0.31	2.36
LI-158/PCN/Pt	0.88	0.63	5.31	0.37	2.50
LI-161/PCN/Pt	1.23	0.66	5.24	0.34	2.60
LI-162/PCN/Pt	1.16	0.66	5.10	0.34	2.51
LI-163/PCN/Pt	1.11	0.62	4.74	0.38	2.49
LI-164/PCN/Pt	1.03	0.60	5.03	0.40	2.62
LI-165/PCN/Pt	1.27	0.64	5.77	0.36	2.89

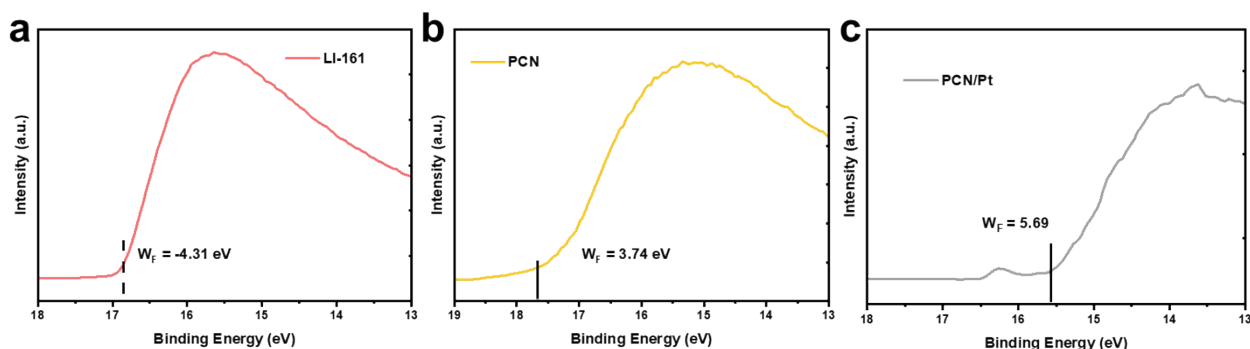


Figure S49. The calculated work function of LI-161 (a), PCN (b) and PCN/Pt (c) by UPS.

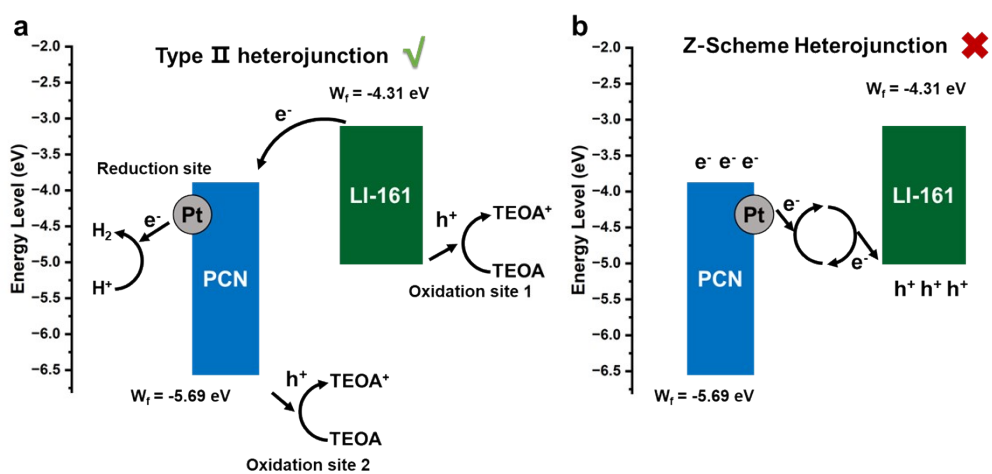


Figure S50. Proposed mechanism of LI-161/PCN/Pt system for photocatalytic hydrogen evolution.

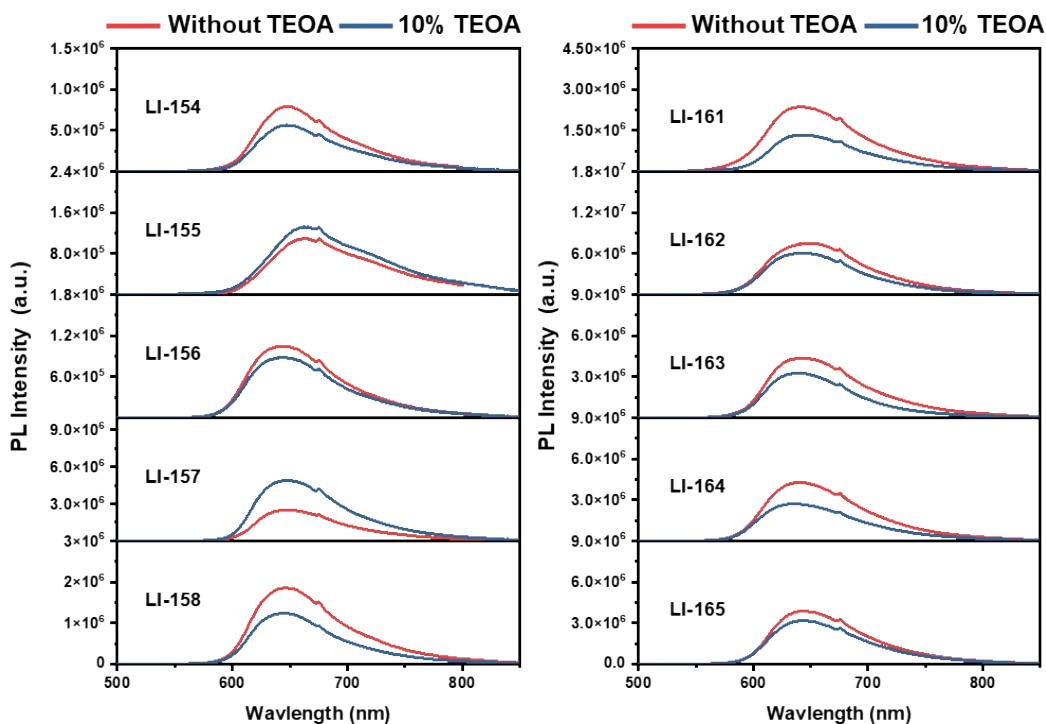


Figure S51. PL spectra of organic dyes (thin film) in water and 10% TEOA aqueous solution.

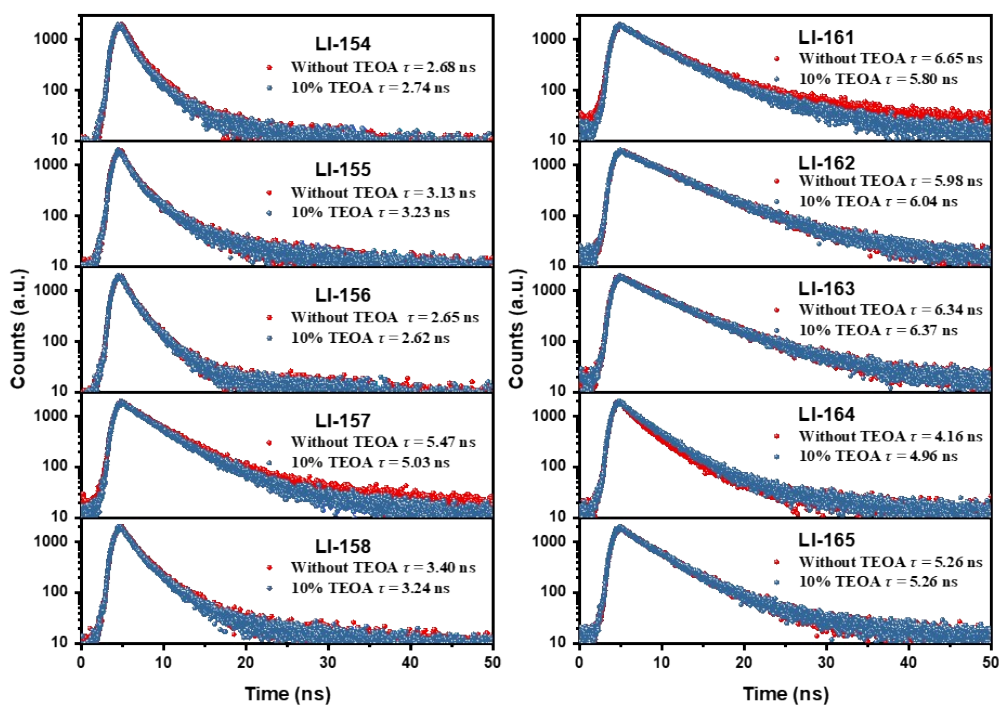


Figure S52. TRPL spectra of organic dyes (thin film) in water and 10% TEOA aqueous solution.

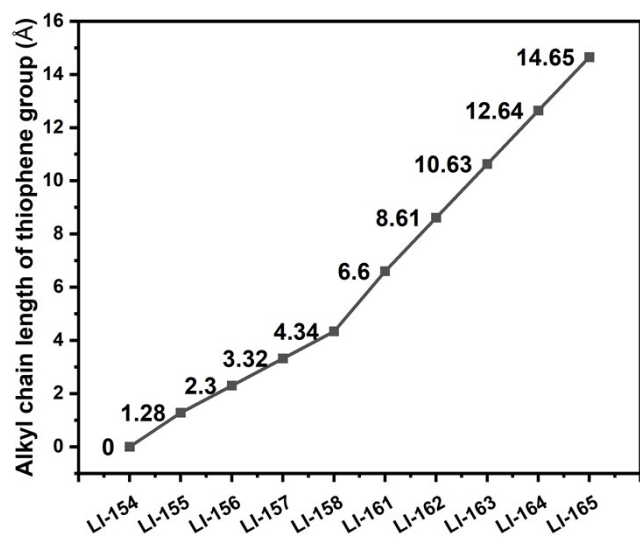


Figure S53. The effective lengths of alkyl chains substituted to thiophene moiety for isolation effect.