Supporting Information for:

Laccase-catalyzed functionalization of phenol-modified carbon nanotubes: from grafting of metallopolyphenols to enzyme self-immobilization

Umberto Contaldo,^a Solène Gentil, ^{a,b} Elise Courvoisier-Dezord,^c Pierre Rousselot-Pailley,^c Fabrice Thomas^a, Thierry Tron,^{c*} and Alan Le Goff^{*a}

1. Materials and Methods

Chemicals and general procedures.

Multiwalled carbon nanotubes (MWCNT; 10 nm diameter, purity > 99%) were obtained from Sigma-Aldrich and were used as received without any purification. All the reagents were purchased from Sigma–Aldrich and were used without further purification. All solvents were of analytical grade. 4-hydroxybenzenediazonium tetrafluoroborate¹ and 4-ferrocenylphenol³⁸ were prepared as previously described. Distilled water was passed through a Milli-Q water purification system. The electrochemical experiments were carried out in a three-electrode electrochemical cell using a Biologic VMP3 Multi Potentiostat. The MWCNT electrodes were used as working electrodes. Pt wire was used ad counter electrode and the reference electrode was based on the Ag/AgClO₄ 10⁻² M reference electrode in MeCN and Ag/AgCl in water. All potentials are given versus Fc/Fc⁺ in MeCN and the standard hydrogen electrode in water (E°_{Ag/AgCl(KCl sat)} = +0.197 V *vs* NHE at 25°C). All current densities are normalized towards the geometrical surface of the glassy carbon electrode (0.071 cm²). Experiments were repeated on at least three electrodes.

The X-band EPR spectra were recorded on an EMX plus spectrometer equipped with a high sensitivity cavity and a Bruker nitrogen flow cryostat. Spectra were simulated using the Easyspin software package² operating in Matlab. First, MWCNTs were chemically-modified with 4-hydroxybenzenediazonium tetrafluoroborate according to a previously-described procedure.^{3,4} 200 μ L of 1 mg mL⁻¹ dispersion ofphenol-modified MWCNTs were prepared in 100 mM Acetate buffer (pH 5.5), 100 mM DMPO and 10 % DMSO. Laccase (2 μ L of a 150 μ M stock solution) was injected in the EPR tube and subsequently frozen at 100 K.

XPS analysis was performed using a Thermoelectron ESCALAB 250 device (ICGM, France). The X-ray excitation was provided by a monochromatic Al-K α (h ν =1486.6 eV) source. The analyzed area was ~0.15 mm². The background signal was removed using the Shirley⁵ method. The surface atomic concentrations were determined from photoelectron peaks areas using the atomic sensitivity factors reported by Scofield⁶. Binding energies (BE) of all core levels were referred to the C=C of C1s carbon at 284.4 eV. *Caution: Perchlorate salts present a potential explosion risk; use with caution!*

UV-visible spectroscopy of reactivity of laccase with 4-ferrocenylphenol (FcPhOH). For all spectroscopical analysis was used spectrophotometer Lambda 650 (Perkin Elmer). The quartz glass cuvette (Hellma® Micro Cuvette, QS Quartz) was prepared with: 20 μ L of 1 mM FcPhOH in 100% DMSO and 960 μ L of 0.1 M Acetate buffer, pH 5.5. An FcPhOH spectrum was first recorded, after baseline correction (0.1 M Acetate buffer, pH 5.5, 2 % DMSO). Then 20 μ L of Laccase 150 μ M in 50 mM Acetate buffer, pH 5 was injected into the cuvette, mixed and then a spectrum was recorded after a total time of 2 minutes and then each 5 minutes thereafter.

A single wave time evolution, $\lambda = 350$ nm, of the reactivity of the laccase with FcPhOH was recorded by injection of 20 µL of Laccase 150 µM in 50 mM Acetate buffer, pH 5 into the cuvette filled with 980 µL of 0.1M Acetate buffer, pH 5.5, 20 µM FcPhOH.

MALDI-TOF MS experiments

The MALDI-TOF experiments were performed with a Bruker Daltonics Autoflex Speed mass spectrometer in reflectron positive mode. Dithranol was used as matrix. The matrix solution was prepared by dissolving dithranol in tetrahydrofuran (THF) (2mg ml⁻¹). Samples were diluted in THF. 2 μ l of the sample solution were mixed with 4 μ l of matrix solution then 1 μ l of the mixture solution was deposited onto the MALDI target plate and dried at room temperature. Calibration was performed externally by using a peptide calibration mixture (Peptide Calibration Standard II -Bruker Daltonics).

Preparation of MWCNT electrode modified with 4-hydroxybenzenediazonium tetrafluoroborate and laccase

MWCNT film were prepared by drop coating 20 μ L of a 5 mg mL⁻¹ dispersion of MWCNTs in 1-Methyl-2-pyrrolidinone (NMP) onto the glassy carbon (GC) electrode surface. The deposit was then allowed to dry under vacuum.

Electrografting of 4-hydroxybenzenediazonium on the surface of the MWCNT electrode was performed by cyclic voltammetry in MeCN with 0.1 M tetrabutylammonium hexafluorophosphate (TBAP) as the supporting electrolyte.

The modified MWCNT GC electrode were then incubated in 20 μ L of Lac3 solution (70 μ M in Acetate buffer 50 mM pH 5) for 6 h at 4°C.

Figure S1. XPS survey spectra the phenol-modified MWCNT electrodes modified with FcPhOH in the (a) presence and (b) absence of laccase.



References

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