

## Electronic Supplementary Information (ESI)

### **Improving the catalytic ability of peptide-based artificial glycosidase through tyrosine strategy**

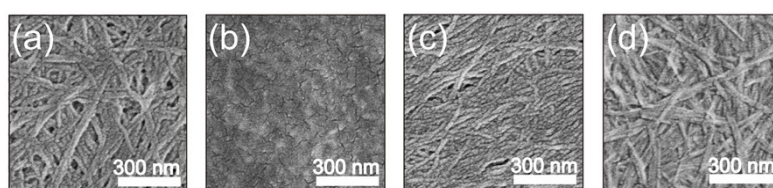
Lijun Yang <sup>a</sup>, Yi Tian <sup>a</sup>, Yutong Wang <sup>a</sup>, Wei Qi <sup>\*, a,c,d</sup> and Mengfan Wang <sup>\*,a,b,d</sup>

- a. School of Chemical Engineering and Technology, State Key Laboratory of Chemical Engineering, Tianjin University, Tianjin 300350, P. R. China.
- b. School of Life Sciences, Tianjin University, Tianjin 300072, P. R. China.
- c. The Co-Innovation Centre of Chemistry and Chemical Engineering of Tianjin, Tianjin 300072, P. R. China.
- d. Tianjin Key Laboratory of Membrane Science and Desalination Technology, Tianjin 300350, P. R. China.

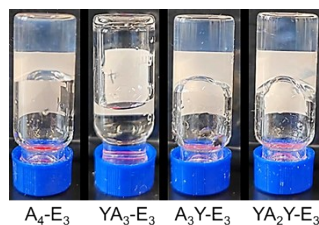
\* Corresponding author: [mwang@tju.edu.cn](mailto:mwang@tju.edu.cn), [qiwei@tju.edu.cn](mailto:qiwei@tju.edu.cn)

**Table S1** Peptides for the construction of glycosidase-like catalysts

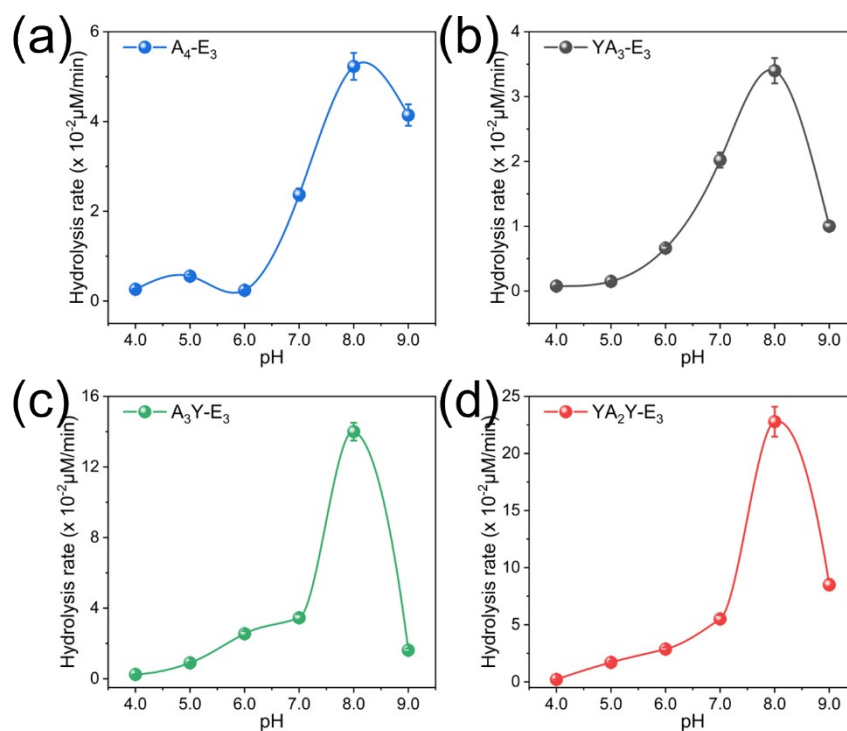
Peptide assemblies	Peptide sequence
A <sub>4</sub> -E <sub>3</sub>	Fmoc-AAAAEEE-CONH <sub>2</sub>
YA <sub>3</sub> -E <sub>3</sub>	Fmoc-YAAAE-CONH <sub>2</sub>
A <sub>3</sub> Y-E <sub>3</sub>	Fmoc-AAAYEEE-CONH <sub>2</sub>
YA <sub>2</sub> Y-E <sub>3</sub>	Fmoc-YAAYEEE-CONH <sub>2</sub>



**Fig. S1** SEM images of (a) A<sub>4</sub>-E<sub>3</sub>, (b) YA<sub>3</sub>-E<sub>3</sub>, (c) A<sub>3</sub>Y-E<sub>3</sub> and (d) YA<sub>2</sub>Y-E<sub>3</sub>.

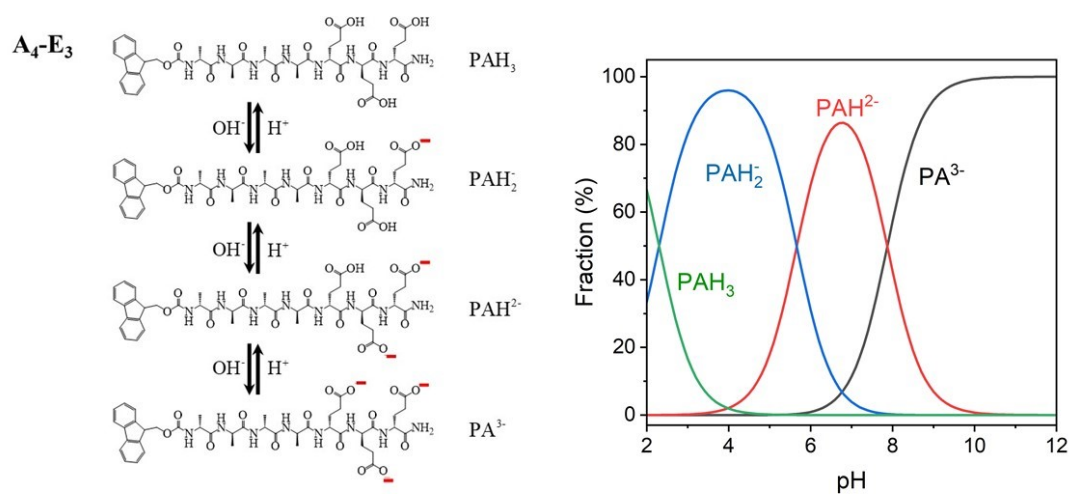


**Fig. S2** Photographic images of peptide assemblies

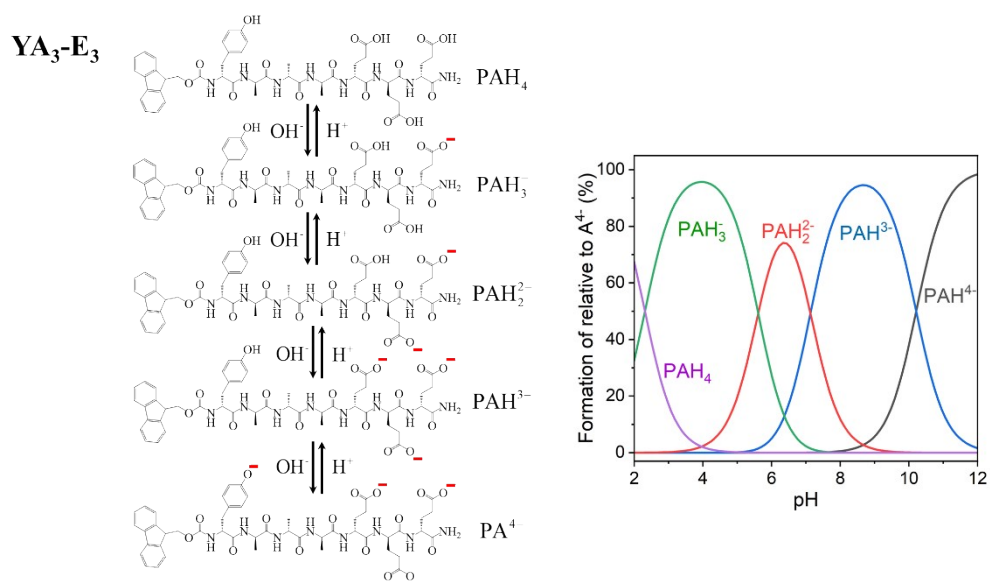


**Fig. S3** pH-*V* profiles for the hydrolysis of *p*-NPG catalyzed by peptide assemblies, (a)

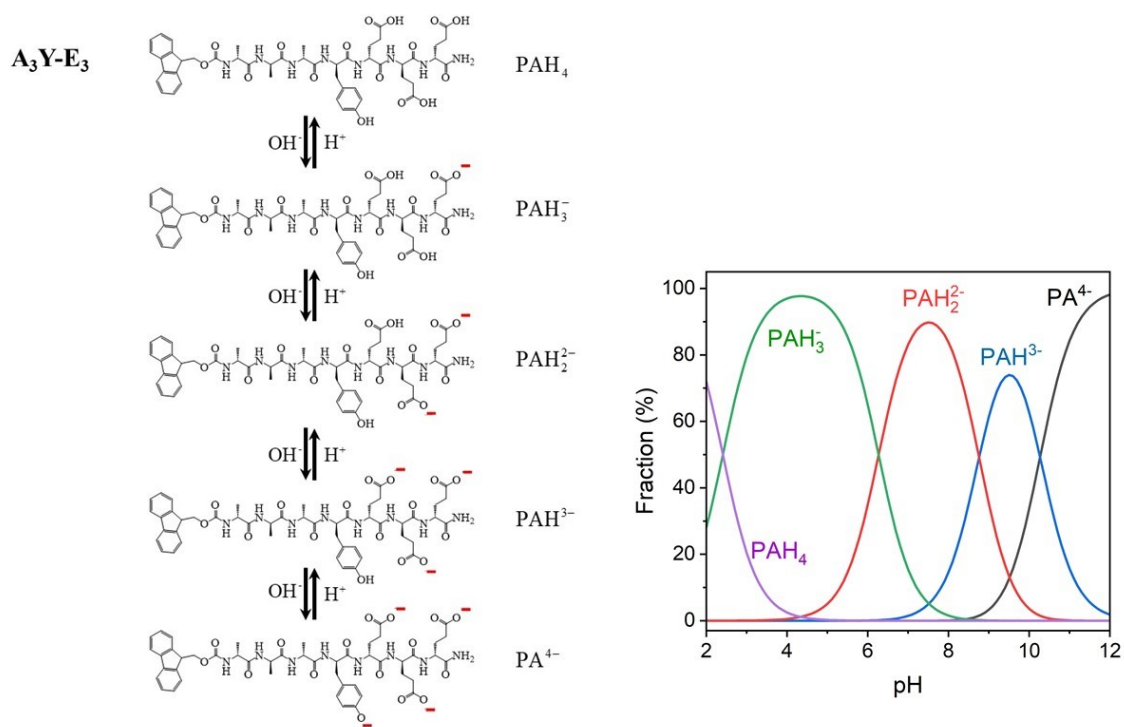
A<sub>4</sub>-E<sub>3</sub>, (b) YA<sub>3</sub>-E<sub>3</sub>, (c) A<sub>3</sub>Y-E<sub>3</sub> and (d) YA<sub>2</sub>Y-E<sub>3</sub>.



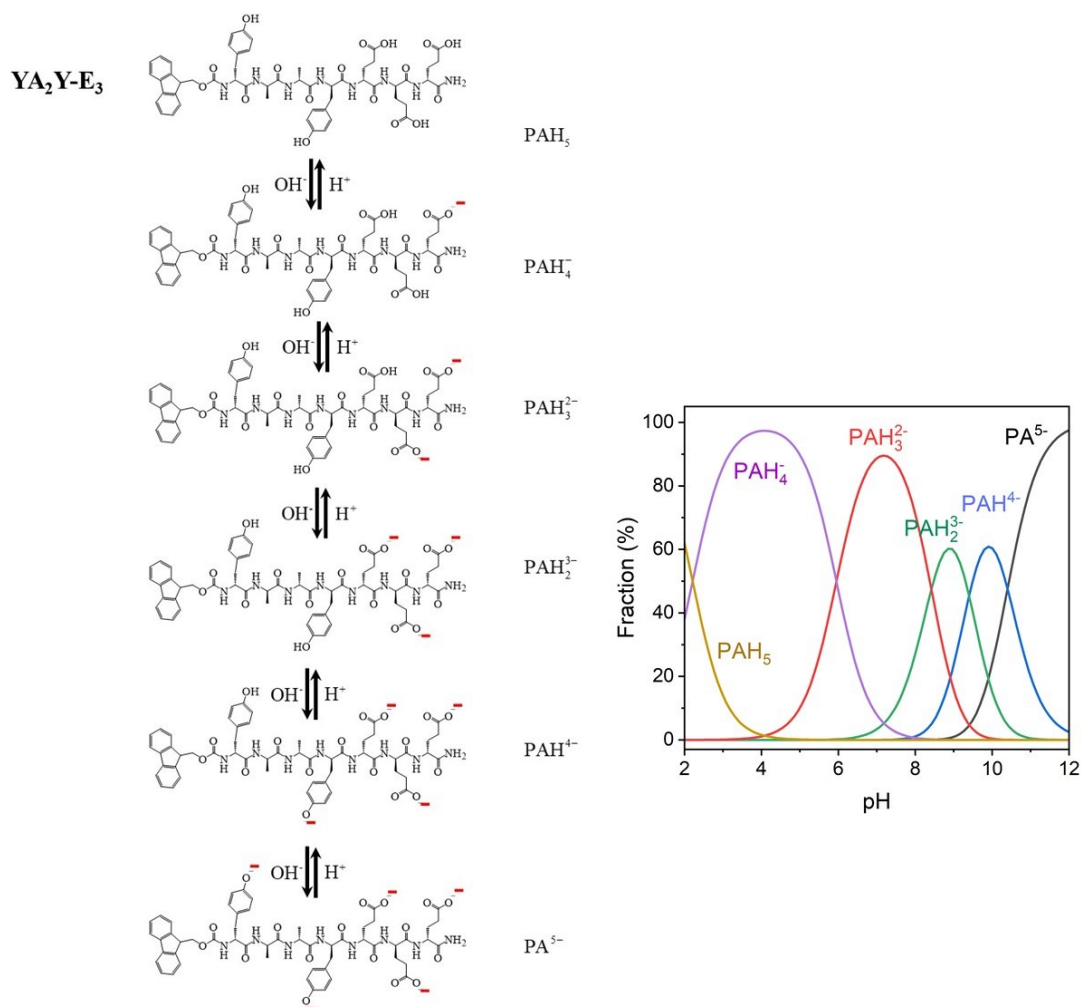
**Fig. S4** Relative abundance of different protonated and deprotonated species of A<sub>4</sub>-E<sub>3</sub>.



**Fig. S5** Relative abundance of different protonated and deprotonated species of YA<sub>3</sub>-E<sub>3</sub>.



**Fig. S6** Relative abundance of different protonated and deprotonated species of A<sub>3</sub>Y-E<sub>3</sub>.



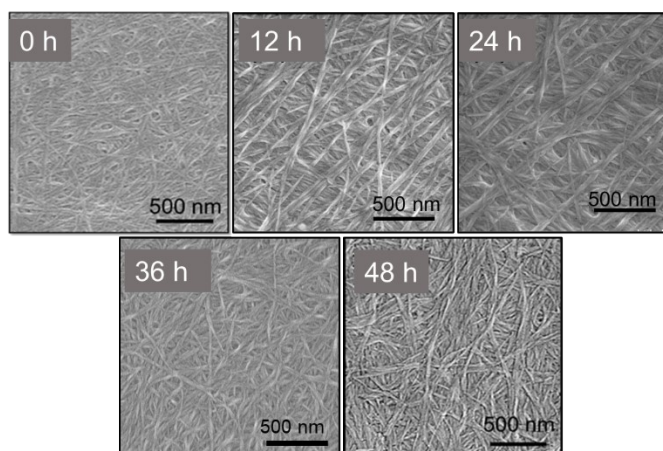
**Fig. S7** Relative abundance of different protonated and deprotonated species of YA<sub>2</sub>Y-E<sub>3</sub>.

**Table S2** The protonation state of peptide catalysts under pH 8<sup>a</sup>

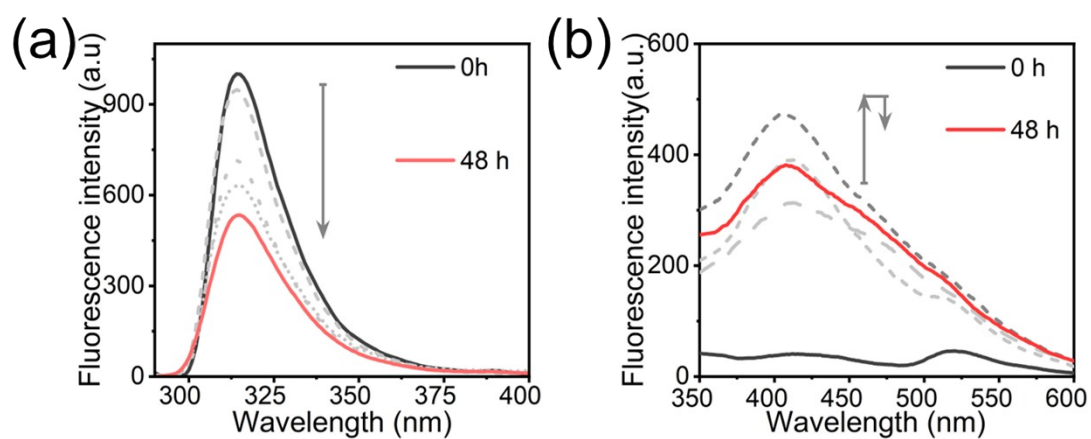
<b>A<sub>4</sub>-E<sub>3</sub></b>	PAH <sub>3</sub>	PAH <sub>2</sub> <sup>-</sup>	PAH <sup>2-</sup>	PA <sup>3-</sup>		
Fraction (%)	0	0.19	42.49	57.32		
<b>YA<sub>3</sub>-E<sub>3</sub></b>	PAH <sub>4</sub>	PAH <sub>3</sub> <sup>-</sup>	PAH <sub>2</sub> <sup>2-</sup>	PAH <sup>3-</sup>	PA <sup>4-</sup>	
Fraction (%)	0	0.046	11.58	87.82	0.55	
<b>A<sub>3</sub>Y-E<sub>3</sub></b>	PAH <sub>4</sub>	PAH <sub>3</sub> <sup>-</sup>	PAH <sub>2</sub> <sup>2-</sup>	PAH <sup>3-</sup>	PA <sup>4-</sup>	
Fraction (%)	0	1.56	83.8	14.56	0.08	
<b>YA<sub>2</sub>Y-E<sub>3</sub></b>	PAH <sub>5</sub>	PAH <sub>4</sub> <sup>-</sup>	PAH <sub>3</sub> <sup>2-</sup>	PAH <sub>2</sub> <sup>3-</sup>	PAH <sup>4-</sup>	PA <sup>5-</sup>

Fraction (%)	0	0.62	70.74	27.52	1.12	0

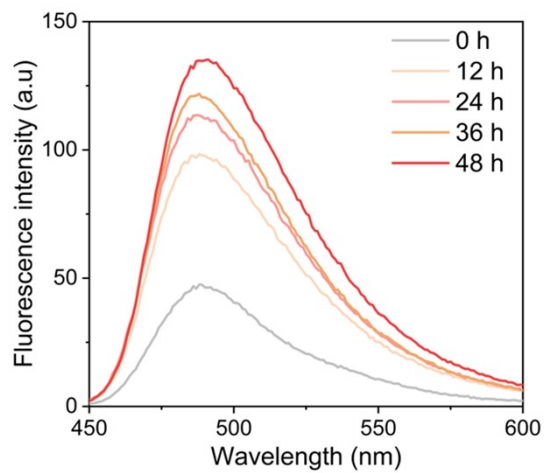
<sup>a</sup> The gray shadow indicates catalytically active states



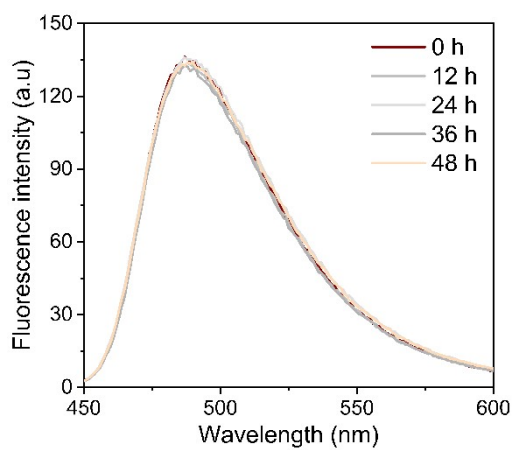
**Fig. S8** SEM images of CL-YA<sub>2</sub>Y-E<sub>3</sub> at different cross-linking time.



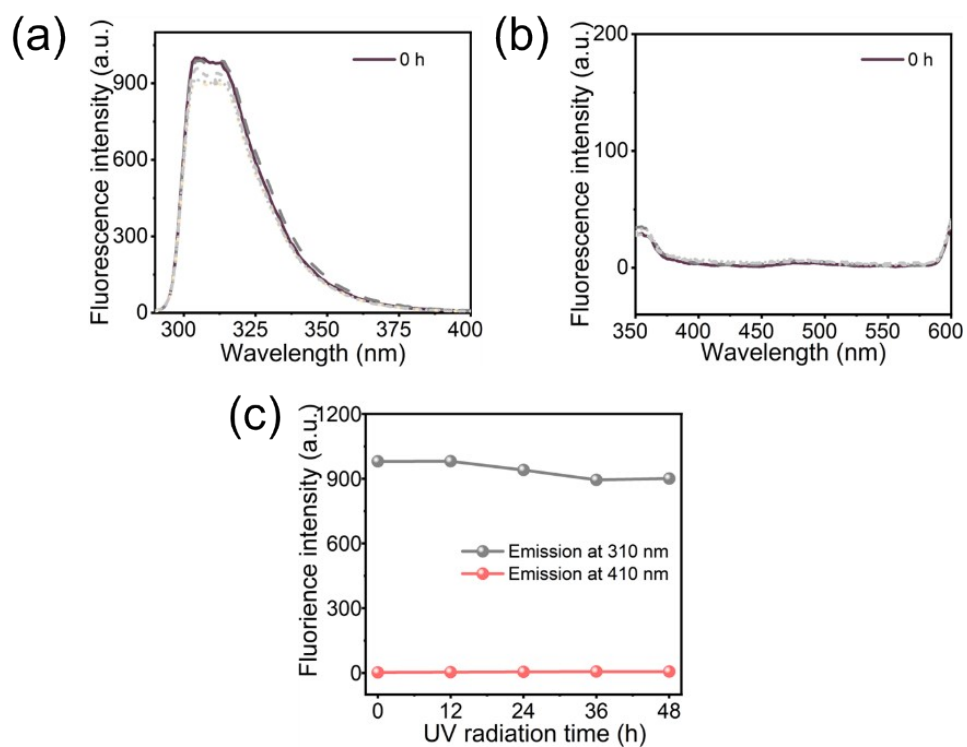
**Fig. S9** (a) Fluorescence emission spectrum of the YA<sub>2</sub>Y-E<sub>3</sub> solution (1 mM/mL) under 274 nm excitation. (b) Fluorescence emission spectrum of the YA<sub>2</sub>Y-E<sub>3</sub> solution (1 mM/mL) under 320 nm excitation.



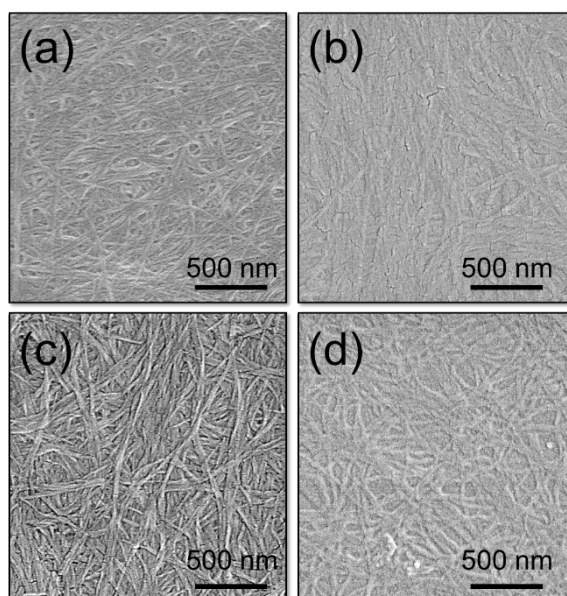
**Fig. S10** Th-T fluorescence spectrum of the CL-YA<sub>2</sub>Y-E<sub>3</sub> at different cross-linking time.



**Fig. S11** Th-T fluorescence spectrum of the CL-A<sub>4</sub>-E<sub>3</sub> at different cross-linking time.



**Fig. S12** (a) Fluorescence emission spectrum of the  $A_4-E_3$  solution (1 mM/mL) under 274 nm excitation. (b) Fluorescence emission spectrum of the  $A_4-E_3$  solution (1 mM/mL) under 320 nm excitation. (c) Fluorescence intensity of emission at 274 nm and 320 nm varies with time.



**Fig. S13** SEM images of  $YA_2Y-E_3$  before (a) and after (b) the p-NPG hydrolysis reactions. SEM images of  $CL-YA_2Y-E_3$  before (c) and after (d) the p-NPG hydrolysis reactions.



**Table S3** Comparison of YA<sub>2</sub>Y-E<sub>3</sub>, CL-YA<sub>2</sub>Y-E<sub>3</sub> with some reported glycosidase mimics in *p*-NPG (**1**) and 4-nitrophenyl  $\alpha$ -D-glucoside hydrolysis (**2**).

Catalyst	Temperature	Substrate	$K_m$ (mM)	$V_{max}$ ( $\mu$ M/min)	$k_{cat}$ ( $\times 10^{-3} s^{-1}$ )	$k_{cat}/K_m$ ( $M^{-1} min^{-1}$ )	Substrate	Reaction time	Specific activity ( $\mu$ mol/h/mg)	Reference
YA <sub>2</sub> Y-E <sub>3</sub>	30 °C	<b>1</b>	1.64	0.29	0.0032	0.117	<b>2</b>	96 h	66.2	This work
CL-YA <sub>2</sub> Y-E <sub>3</sub>			2.59	0.657	0.0073	0.168			123.4	
6 <sup>A</sup> -( <i>R</i> )-cyanohydrin CD ( <b>19</b> )	59 °C	<b>1</b>	5.36		0.0012	0.013	<b>2</b>	48 h	32.28	1
6 <sup>A</sup> ,6 <sup>D</sup> -dicarboxylic acid $\beta$ -CD ( <b>29</b> )	59 °C		n.d.		0.0170				45.0	
Catalyst 1	59 °C	<b>1</b>	13.40		0.000334	0.0015	<b>2</b>	48 h	8.28	2
Binuclear Copper(II) Complexes 12	30 °C	<b>1</b>	213		0.07	0.00204	<b>2</b>	48 h	96.70	3
Cyclodextrins 13	30 °C	<b>1</b>	80.30		0.0142	0.0106	<b>2</b>	48 h	38.62	4
Dicyanohydrin- $\beta$ - cyclodextrin (1)	59 °C	<b>1</b>	4.14		0.0103	0.149				5
Dicyanohydrin- $\alpha$ - cyclodextrin (2)			6.34	0.33	0.0093	0.0880				
Cyclodextrin Derivatives (11)	59 °C	<b>1</b>	7.60		0.0014	0.0111				6
Compound 2	59 °C	<b>1</b>	2.90		0.0012	0.0123	<b>2</b>	48 h	3.83	7

## Reference

1. J. Bjerre and M. Bols, *European Journal of Organic Chemistry*, 2010, **2010**, 3487-3500.
2. C. Rousseau, N. Nielsen and M. Bols, *Tetrahedron Letters*, 2004, **45**, 8709-8711.
3. S. Striegler, N. A. Dunaway, M. G. Gichinga, J. D. Barnett and A.-G. D. Nelson, *Inorganic Chemistry*, 2010, **49**, 2639-2648.
4. Y. Zhou, C. M. Pedersen and M. Bols, *Tetrahedron Letters*, 2013, **54**, 2458-2461.
5. F. Ortega-Caballero, J. Bjerre, L. S. Laustsen and M. Bols, *Journal of Organic Chemistry*, 2005, **70**, 7217-7226.
6. F. Ortega-Caballero, C. Rousseau, B. Christensen, T. E. Petersen and M. Bols, *Journal of the American Chemical Society*, 2005, **127**, 3238-3239.
7. J. Bjerre, T. H. Fenger, L. G. Marinescu and M. Bols, *European Journal of Organic Chemistry*, 2007, **2007**, 704-710.