# Application of Singlet Oxygen-Activatable Nanocarriers to Boost X-ray-Induced Photodynamic Therapy and Cascaded Ferroptosis for Breast Cancer

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## **Experimental methods**

## Materials

RPMI 1640 medium, fetal bovine serum (FBS) and trypsin were purchased from Gibco BRL (Gaithersberg, MD; USA). Cell counting kit-8 (CCK-8) and 4',6-diamidino-2-phenylindole (DAPI) were purchased from Beyotime Biotechnology Co., Ltd. (Nantong; China). Liperfluo was obtained from Beijing Dojindo Biotechnology Co., Ltd (Beijing, China). Annexin V-FITC/PI was purchased from Invitrogen Corporation (IVGN; USA). All other solvents were of analytical grade.

## Characterization

The proton nuclear magnetic resonance (<sup>1</sup>H NMR) spectra were recorded in CD<sub>3</sub>SOCD<sub>3</sub> on a 400-MHz spectrometer (Avance III, Bruker, Germany). The mean particle size, polydispersity, and zeta potential were measured by dynamic light scattering (DLS) with a Malvern Zetasizer (Nano-ZS, Malvern Instruments, UK). The morphology of nanoparticles was observed by transmission electron microscopy (TEM, Hitachi HT7700). The UV/Vis absorptions of nanoparticles and drug loading efficiency were detected on a UV/Vis spectrophotometer (UV-3600 Shimadzu, Japan). The fluorescence of SOSG was measured using a Hitachi F7000 fluorescence spectrophotometer. *In vivo* imaging of small animals was captured by Xenogen IVIS Lumina system (AMI-934M, USA).

### Cellular internalization of nanocarriers in vitro

For confocal laser scanning microscope (CLSM) observations, 4T1 cells were

seeded onto 14-mm coverslips at a density of  $2 \times 10^4$  cells per well in 1.0 mL of 1640 medium, and cultured at 37 °C with 5% CO<sub>2</sub> for 12 h. The cells were then incubated with NP<sub>VR</sub> or D-NP<sub>VR</sub> for 2, 4 or 6 h, washed with PBS, fixed with 4% paraformaldehyde, and then stained with phalloidin and 4',6-diamidino-2-phenylindole (DAPI) sequentially according to the manufacturer's protocol. The cellular internalization was then visualized on a Zeiss LSM 810 confocal microscope.

### **Biosafety Evaluation**

Twenty healthy BALB/c mice were randomly divided into four groups and received *i.v.* injection daily for three times with PBS, free VP + RLS3, NP<sub>VR</sub>, or D-NP<sub>VR</sub>, respectively. The orbital plexus blood sampling was collected on day 7 for routine blood count, creatinine, urea nitrogen, alanine aminotransferase, and aspartate aminotransferase detection.

#### **Statistical Analysis**

Statistical significance was analyzed using a t-test. One-way analysis of variance (ANOVA) was used for comparisons between more than two groups. Unless specially noted, data are shown as mean  $\pm$  SD (p<0.05).



Figure S1. Synthetic route of 1,2-bis(2-hydroxyethylthio)ethylene bridged D-HPE.



**Figure S2.** <sup>1</sup>H NMR spectrum of D-HPE in CDCl<sub>3</sub> recorded on an AVANCE III 400 MHz spectrometer at 25 °C.



Figure S3. <sup>1</sup>H NMR spectrum of HPE in CDCl<sub>3</sub> recorded on an AVANCE III 400 MHz spectrometer at 25 °C.



Figure S4. The diameter (A) and morphology (B) change of  $NP_{VR}$  and  $D-NP_{VR}$  with X-ray irradiation. The scale bar is 200 nm.



Figure S5. Bis(2-hydroxyethylthio)ethylene linker degradation of  $D-NP_{VR}$  with or without 4 Gy of X-ray radiation.



**Figure S6.** Fluorescent imaging of lipid peroxides in 4T1 cells. The cells were treated by various formulations including VP&RSL3, NP<sub>VR</sub>, and D-NP<sub>VR</sub>.



Figure S7. Relative viabilities after incubation with  $NP_{VR}$  or D- $NP_{VR}$  on 4T1, MCF-7 and NIH-3T3 cells.



**Figure S8.** Following *i.v.* injections, the doxorubicin fluorescence observed by CLSM at 24 h with (+) or without (-) X-ray radiation.



**Figure S9.** Body weight change following multiple *i.v.* injections of PBS, free VP&RSL3, NP<sub>VR</sub> or D-NP<sub>VR</sub>.



Figure S10. Complete blood count in BALB/c mice after *i.v.* injections (n = 5).

 Table S1. Drug loading contents (DLC) and encapsulation efficiencies (EE) of VP

 and RSL3 for NP<sub>VR</sub> and D-NP<sub>VR</sub>.

	DLC (%)		EE (%)	
	VP	RSL3	VP	RSL3
NPvr	3.39	3.22	35.9	34.1
D-NP <sub>VR</sub>	3.53	3.07	34.6	30.1