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Electronic Supplementary Information

Effective design for PEGylated polyion complex (PIC) nanoparticles to enhance cell– PIC interaction utilising block copolymer combinations of mismatching ionic chain lengths

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1. Characterization of polymers



Figure S1. A ¹H-NMR spectrum of PEG_{2k}-PAsp (10 mg mL⁻¹, D₂O) at 80 °C with TMSP as an internal standard. DP was calculated to be 56.



Figure S2. A ¹H-NMR spectrum of PEG_{2k} -PAsp (10 mg mL⁻¹, D₂O) at 80 °C with TMSP as an internal standard. DP was calculated to be 69.



Figure S3. A ¹H-NMR spectrum of PEG_{2k} -PAsp (10 mg mL⁻¹, D₂O) at 80 °C with TMSP as an internal standard. DP was calculated to be 93.



Figure S4. A ¹H-NMR spectrum of PEG_{5k}-PAsp (10 mg mL⁻¹, D₂O) at 80 °C. DP was calculated to be 70.



Figure S5. A ¹H-NMR spectrum of homo-PLL (10 mg mL^{-1} , D_2O) at 25 °C with TMSP as an internal standard. DP was calculated to be 64.



Figure S6. A ¹H-NMR spectrum of homo-PLL (10 mg mL^{-1} , D_2O) at 25 °C with TMSP as an internal standard. DP was calculated to be 100.



Figure S7. A ¹H-NMR spectrum of PEG_{2k}–PLL (10 mg mL⁻¹, D₂O) at 25 °C. DP was calculated to be 81.



Figure S8. An SEC calibration curve of PEO standards



Figure S9. A size-exclusion chromatogram of PEG_{2k}–PAsp₅₆.



Figure S10. A size-exclusion chromatogram of PEG_{2k}-PAsp₆₉.



Figure S11. A size-exclusion chromatogram of PEG_{2k}-PAsp₉₃.



Figure S12. A size-exclusion chromatogram of PEG_{5k}–PAsp.



Figure S13. A size-exclusion chromatogram of homo-PLL₆₄.



Figure S14. A size-exclusion chromatogram of homo-PLL $_{100}$.



Figure S15. A size-exclusion chromatogram of PEG_{2k}-PLL.

2. Characterization of PICs



Figure S16. TEM images of the PICs: (a) vesicles, (b) 2k-micelle, and (c) 5k-micelle.





Figure S17. TEM images of Acetylated and Guanidinylated vesicles



Figure S18. Cryo-TEM images of vesicle PIC (56,100) with 50%-CL (Scale bar: 200 nm). The thickness of the PIC membrane was estimated to be 12.0 ± 2.5 nm (N = 28).

Equation S1. The equation to determine the mixing ratio of polyanion and polycation solutions for the PIC fabrication

 $\frac{(conc. of polyanion) \times (polyanion solution volume) \times (DP of polyanion) \times (ionization degree of carboxylate)}{MW of polyanion}$

 $= \frac{(conc. of polycation) \times (polycation solution volume) \times (DP of polycation) \times (ionization degree of amine)}{MW of polycation}$

Note:

Conc. Of polyanion and polycation are set to be 1 mg/mL. Ionization rate of carboxyl at pH 7.4 is 0.92.³¹ Ionization rate of amine at pH 7.4 is 0.95.³¹

Scheme S1. Schematic drawing of TNBS assay.



Figure S19. A TNBS assay calibration curve obtained using homo-PLL



Figure S20. A TNBS assay calibration curve obtained using PEG_{2k}-PLL



Figure S21. A Cy3-fluorescence calibration curve obtained using PEG_{2k}-PAsp-Cy3



Figure S22. A Cy3-fluorescence calibration curve obtained using PEG_{5k}-PAsp-Cy3

Equation S2. The equation to determine the crosslinking degree

Remaining amine ratio (%)

 $= \frac{(DP of polycation) \times (ionization degree of amino groups) \times (apparent conc. of polycation from TNBS assay)}{(MW of polycation)}$

 $\times \frac{(MW \ of \ polyanion)}{(conc. \ of \ polyanion) \times (DP \ of \ polyanion) \times (ionization \ degree \ of \ carboxylate)} \times 100$

Crosslinking degree (%) = 100 - [amount of free amino groups (%)]

Equation S3. The equation for calculation of PEG weight fraction, f_{PEG} , of PICs

 $f_{PEG} = \frac{(PEG \ MW) \times (mole \ fraction \ of \ PEG \ ylated \ ionomer)}{total \ molecular \ weight \ of \ charge \ neutralized \ complex \ in \ mole}$



Figure S23. Autocorrelation functions of PIC vesicles



Figure S24. Autocorrelation functions of non- and modified- PIC vesicles

3. Cell experiment results



Fig. S25 The result of calcein leakage assay of PIC vesicle in HeLa cells obtained by CLSM. Blue: nuclei; Red: PIC; Green: calcein; Yellow: PIC+calcein colocalization. Briefly, HeLa was incubated with medium containing PIC (56, 100) (equal to final PAsp conc. 0.02 mg/mL) and calcein (final conc. 0.1 mg/mL) for 24 hours. Calcein leakage indicating endosomal escape, which will dye the cell with green color of whole cell body upon calcein dissolution into cytosol.



Figure S26. PIC retention on the HeLa cell surface observed after 24 h of incubation. (Blue: nucleus; Red: PICs). Aggregates are indicated by white arrows. The contrast of pictures was adjusted to yield a clear appearance of the PIC position.



2k micelle



Figure S27. Line profilings of fluorescence intensity across HeLa cells treated with (a) PIC vesicles, (b) 2kmicelle, and (c) 5k-micelle. Left: CLSM images of HeLa cells used for line profiling analysis (scan lines are shown in each image as a white arrow). Right: Line profiling results (Green line: An intensity profile of the green

fluorescence from Cellbrite® (Biotium, Fremont, CA, a cellular membrane staining dye with green fluorescence, used according to manufacturer's protocol). Blue line: An intensity profile of blue fluorescence from Hoecsht3324; Red line: An intensity profile of red fluorescence from Cy3-labelled PICs.)



Scale bar: 50 um

Figure S28. Fluorescence microscopy images of RAW 264.7 cells after 24 h of incubation with the acetylated PIC vesicle in the absence (left) and presence (right) of dextran sulphate (DexSO₄) 0.5 mg/mL. Blue: Nucleus (Hoechst 33342); red: acetylated vesicles.



(a) HeLa

(b) RAW 264.7



(c) Caco2







Figure S29. Cytotoxicity assays of PICs in different cell lines. (a) HeLa, (b) RAW 264.7, (c) Caco2 and (d) DC2.4.