

Supporting Information

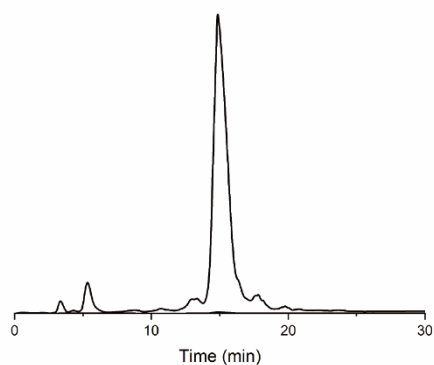
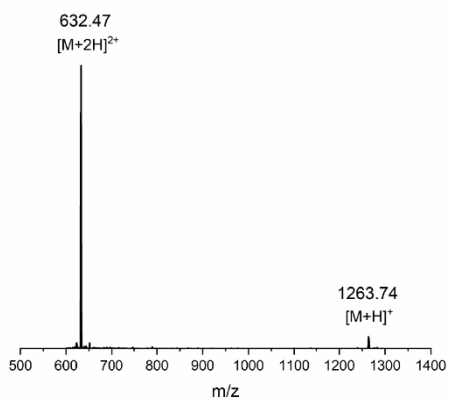
Self-assembled Branched Polypeptides as Amelogenin Mimics for Enamel Repair

Yue Li,^{‡^a} Rong Chang,^{‡^a} Yang-Jia Liu,^b Feng Chen^{*^b} and Yong-Xiang Chen^{*^a}

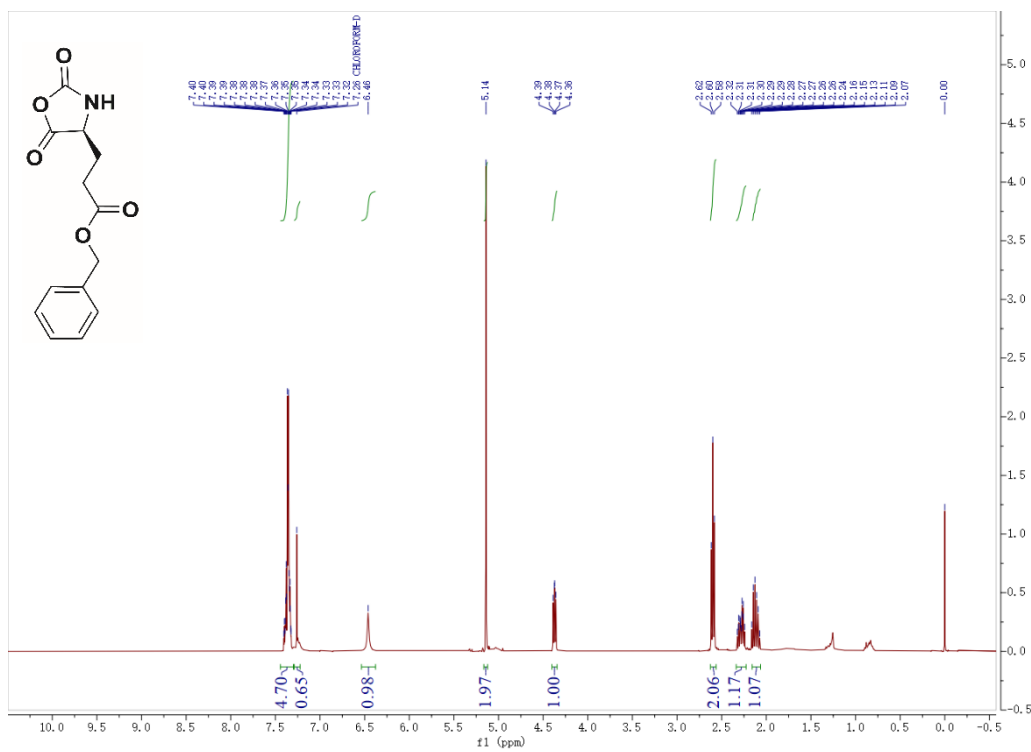
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^bCentral Laboratory Peking University Hospital of Stomatology, Beijing 100081, China. E-mail: chenfeng2011@hsc.pku.edu.cn.

a



b



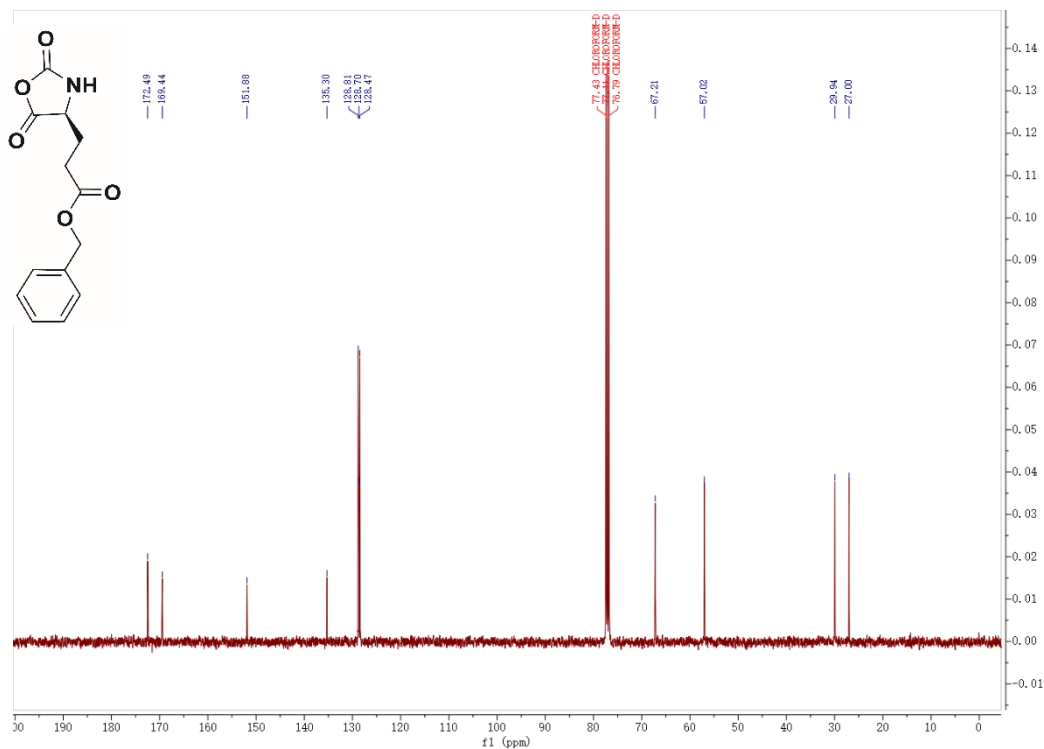
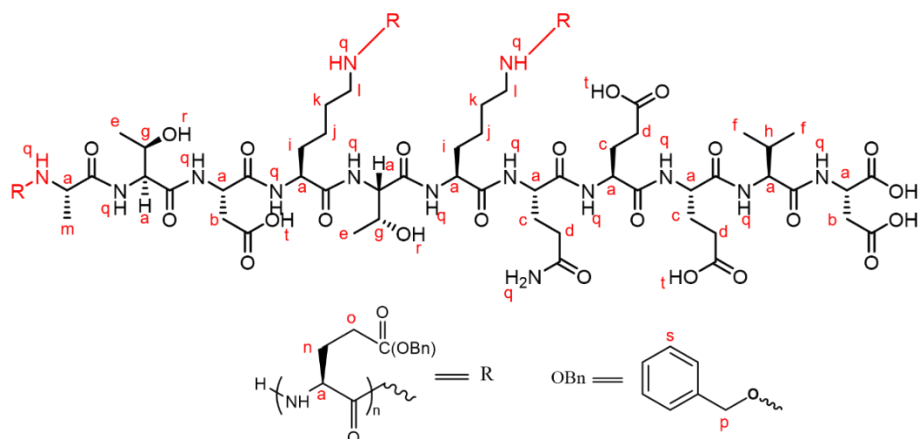
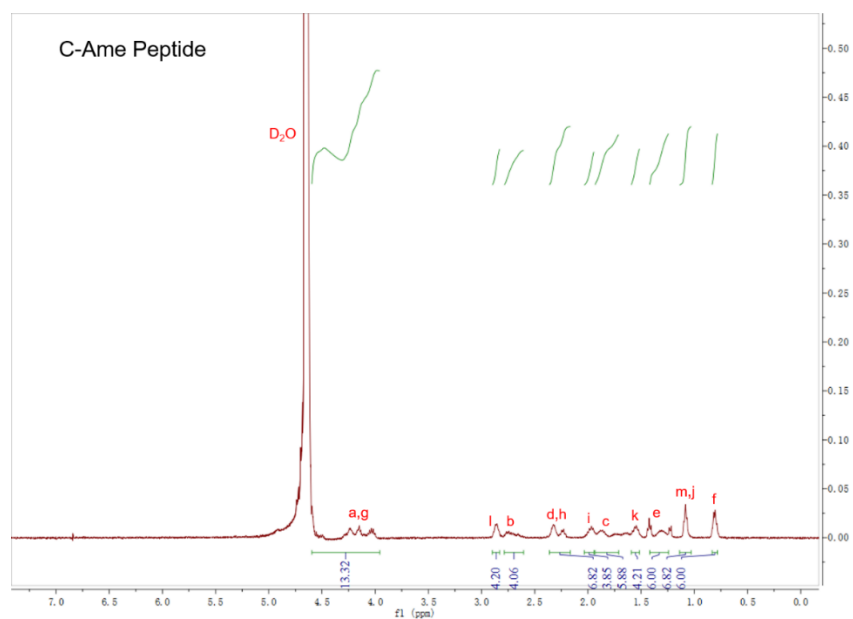


Fig. S1 a) Electrospray ionization mass spectrometry (ESI-MS) and high-performance liquid chromatography (HPLC) analysis of C-Ame Peptide. Retention time is about 14.87 min in a linear gradient of 5-45% B for 35 min (HPLC solvent A: water, 0.06% TFA; B: 80% CH₃CN/water, 0.06% TFA). b) ¹H NMR and ¹³C NMR spectra of BLG-NCA monomer in CDCl₃.

a



b

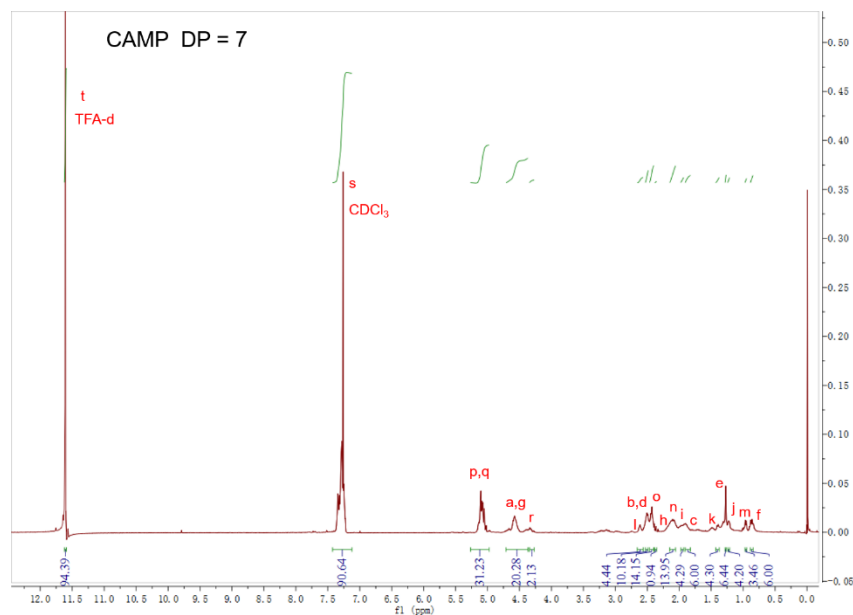


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Chemical Shift	4.25	2.75	1.87	2.32	1.46	0.85	4.17	2.25	1.99	1.09	1.58	2.82	1.11
Integration	11	4	6	6	6	6	2	1	4	4	4	4	3

Chemical Formula: $C_{51}H_{86}N_{14}O_{23}$

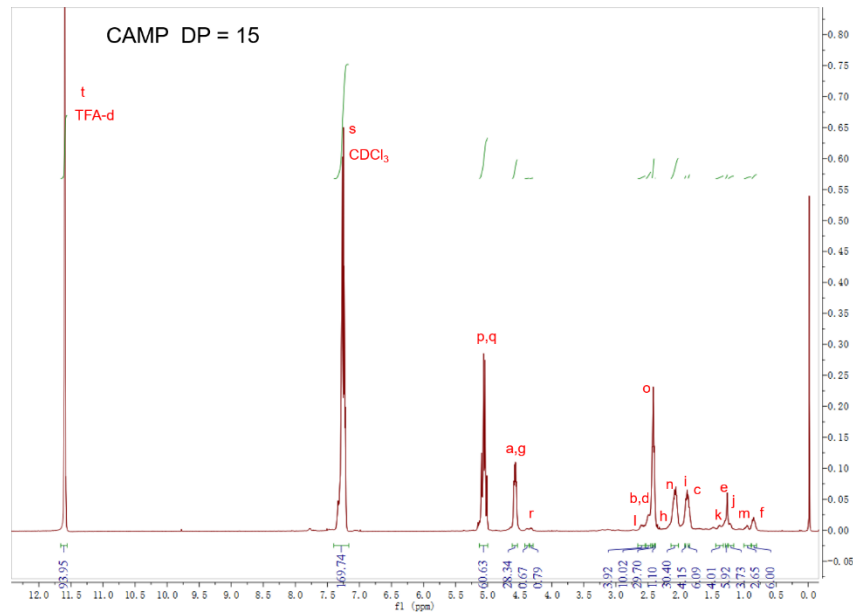
Exact Mass: 1262.60

Molecular Weight: 1263.32



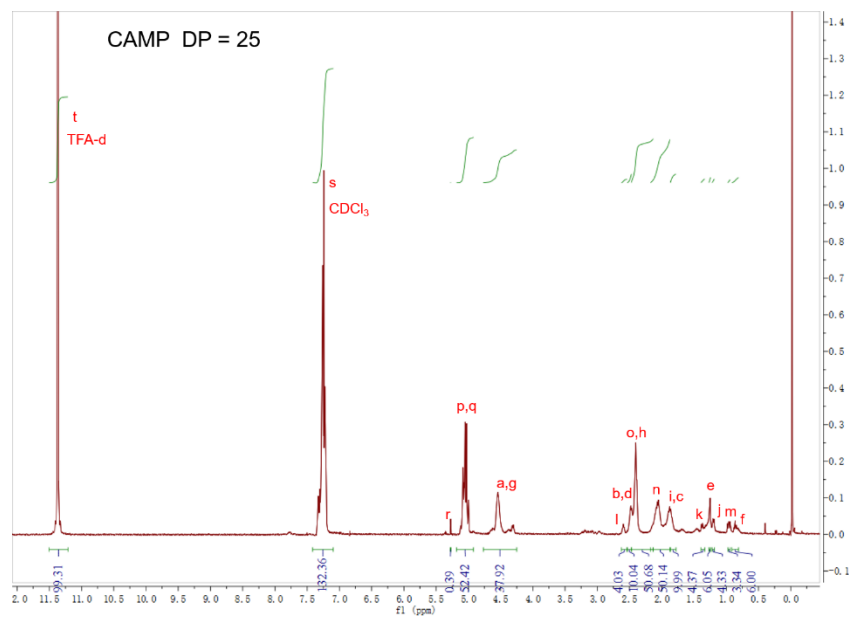
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Chemical Shift	4.62	2.54	1.85	2.54	1.26	0.85	4.62	2.39	2.01	1.25	1.47	2.63	0.95	2.18	2.48	4.28
Integration	18	4	6	6	6	6	2	1	4	4	4	4	3	14	14	2

$$\text{Molecular Weight: } 1263.32 + (18-11) \times (237.25-18.02) = 2797.93$$



	a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	r
Chemical Shift	4.62	2.54	1.85	2.54	1.26	0.85	4.62	2.39	2.01	1.25	1.47	2.63	0.95	2.18	2.48	4.28
Integration	26	4	6	6	6	6	2	1	4	4	4	4	3	30	30	2

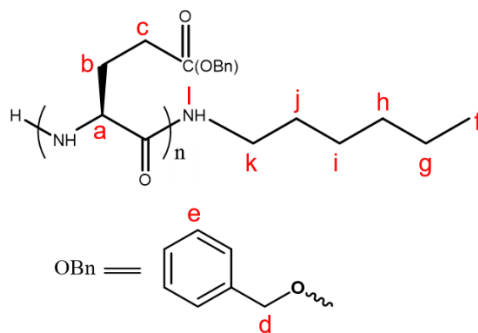
$$\text{Molecular Weight: } 1263.32 + (26-11) \times (237.25-18.02) = 4551.77$$



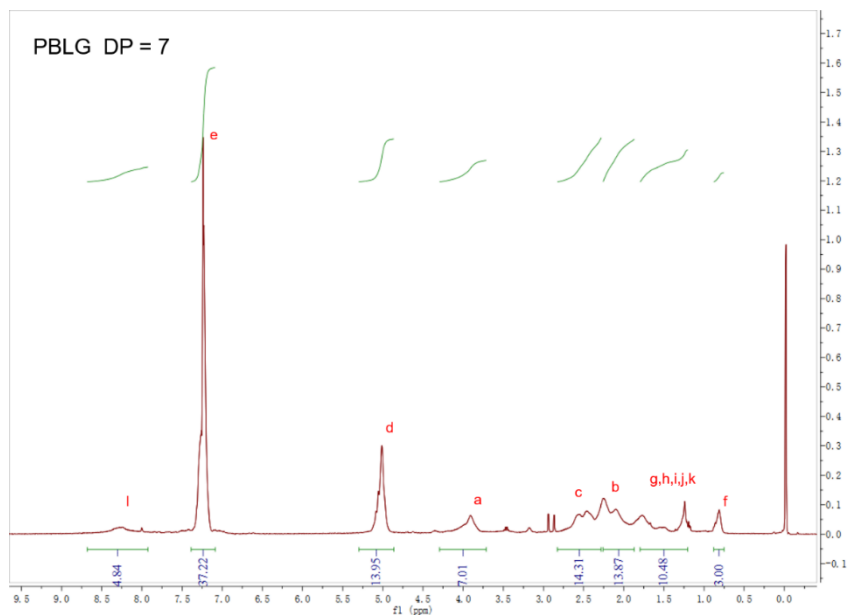
	a	b	c	d	e	f	g	h	i	j	k	l	m	n	o	r
Chemical Shift	4.62	2.54	1.85	2.54	1.26	0.85	4.62	2.39	2.01	1.25	1.47	2.63	0.95	2.18	2.48	4.28
Integration	36	4	6	6	6	6	2	1	4	4	4	4	3	50	50	2

$$\text{Molecular Weight: } 1263.32 + (36-11) \times (237.25-18.02) = 6744.07$$

c

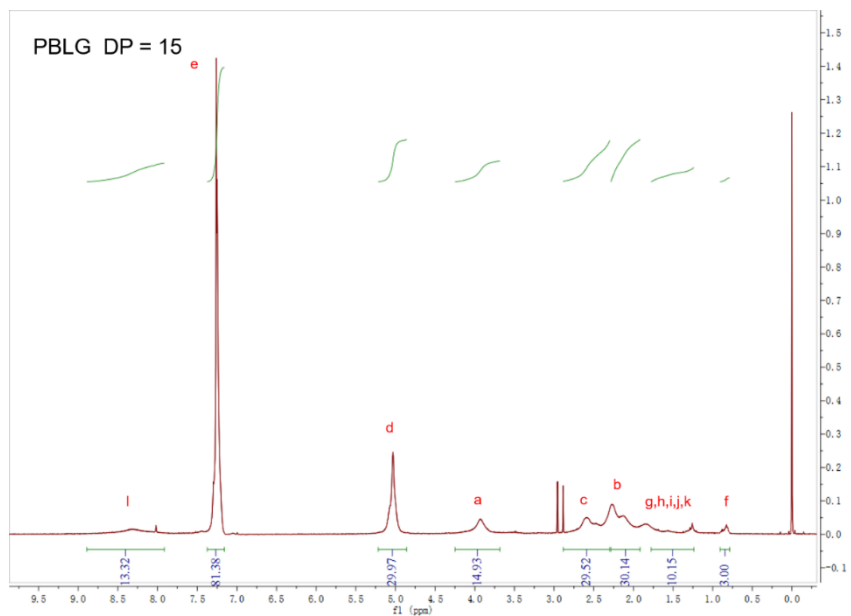


d



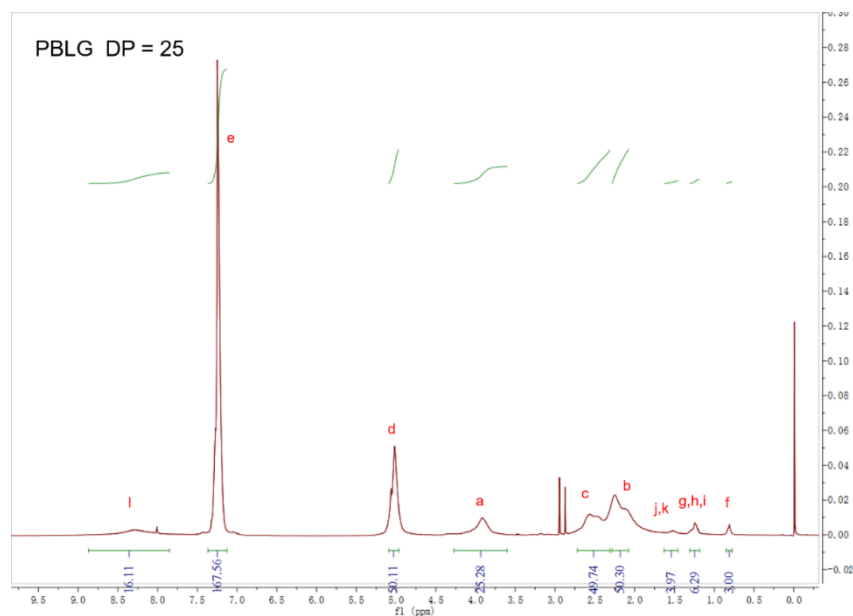
	a	b	c	d	f	g	h	i	j	k	l
Chemical Shift	3.96	2.25	2.52	5.02	0.85	1.25-1.78					8.25
Integration	7	14	14	14	3	10					5

Molecular Weight: $101.19+7 \times (237.25-18.02) = 1635.8$



	a	b	c	d	f	g	h	i	j	k	l
Chemical Shift	3.96	2.25	2.52	5.02	0.85	1.25-1.78					8.25
Integration	15	30	30	30	3	10					13

Molecular Weight: $101.19+15 \times (237.25-18.02) = 3389.64$



	a	b	c	d	f	g	h	i	j	k	l
Chemical Shift	3.96	2.25	2.52	5.02	0.85	1.25-1.78					8.25
Integration	25	50	50	50	3	10					16

Molecular Weight: $101.19 + 25 \times (237.25 - 18.02) = 5581.94$

Fig. S2 a) The structure of C-Ame Peptide and CAMPs. b) ^1H NMR spectra of C-Ame Peptide in D_2O , CAMP_7 , CAMP_{15} and CAMP_{25} in CDCl_3 (15% TFA-d). c) The structure of hexylamine and PBLGs. d) ^1H NMR spectra of PBLG_7 , PBLG_{15} , PBLG_{25} in CDCl_3 .

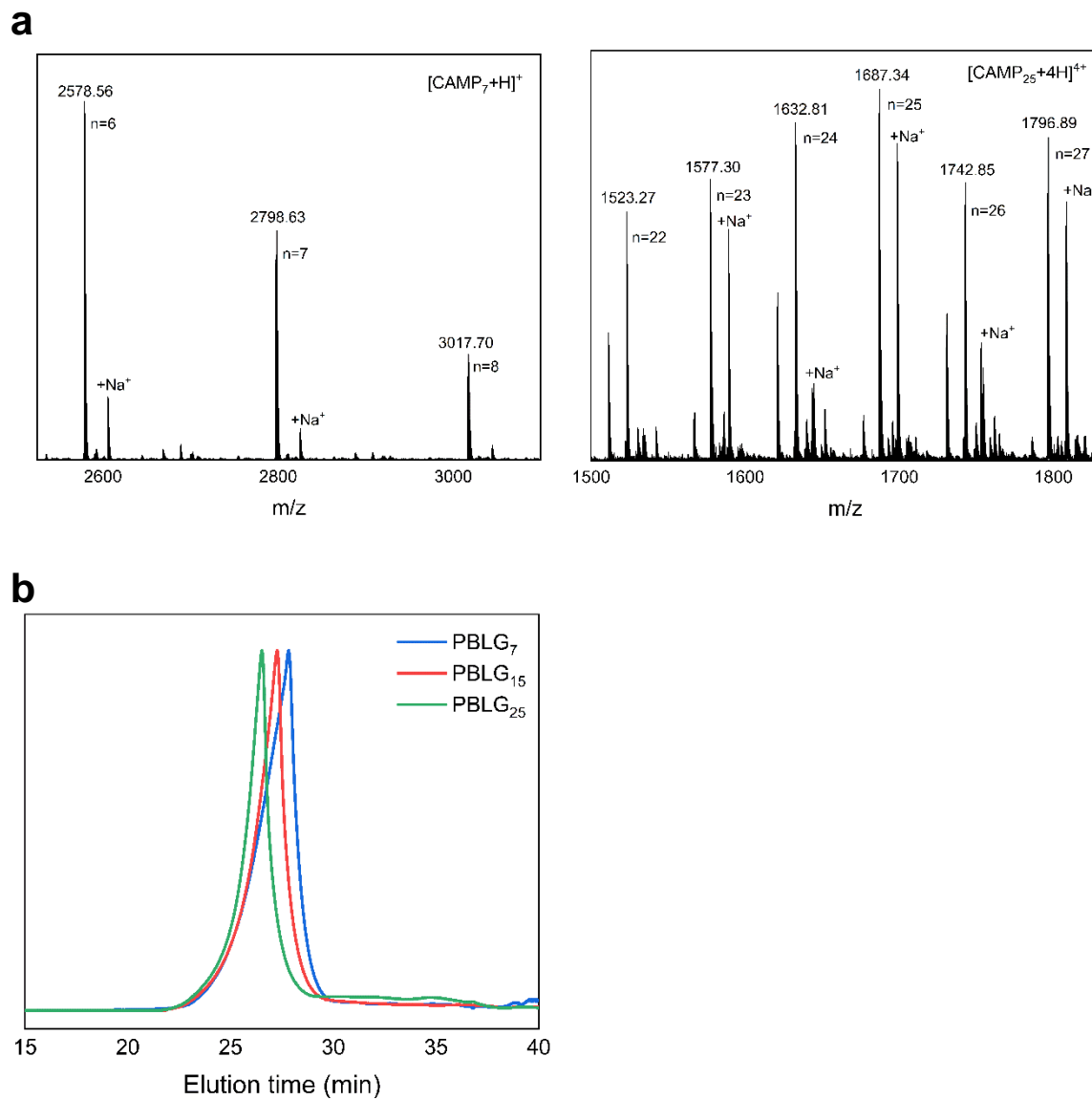


Fig. S3 a) Q-TOF-MS analysis of CAMP_7 and CAMP_{25} . b) GPC analysis of PBLG_7 , PBLG_{15} and PBLG_{25} .

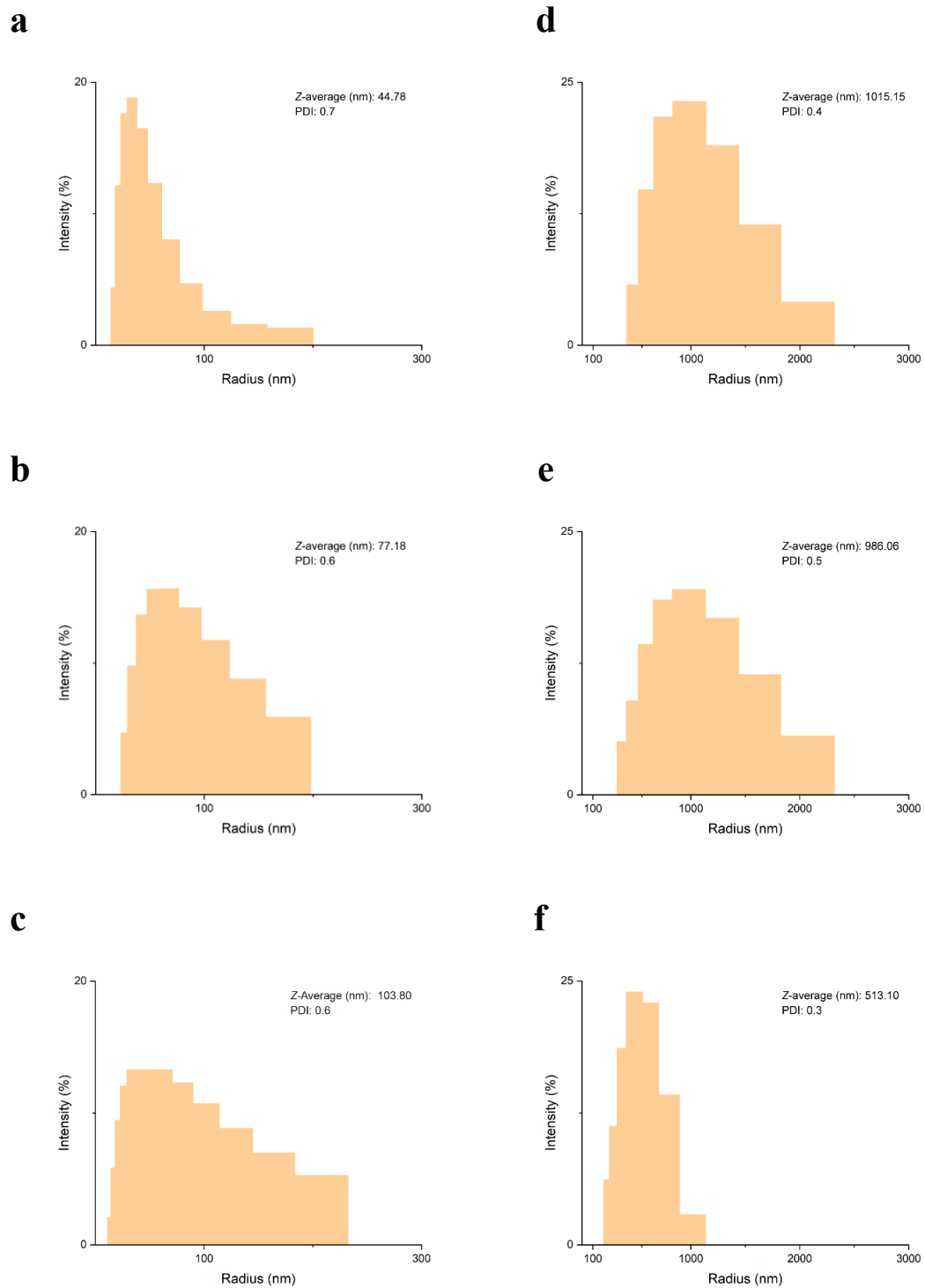
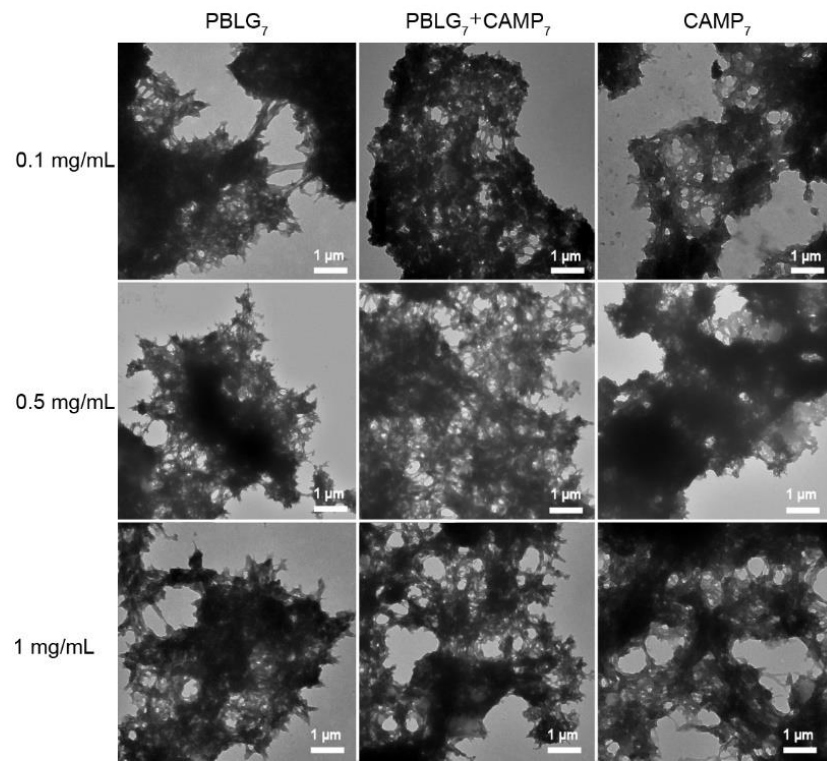
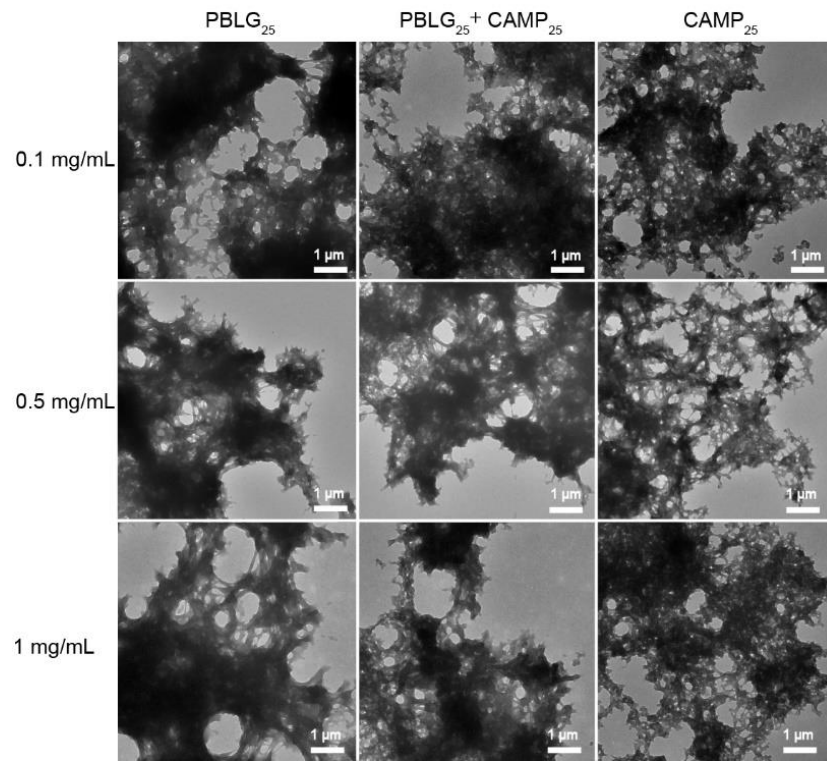


Fig. S4 Size distribution of PBLG₇ (a), PBLG₇ + CAMP₇ (1/1, n/n) (b), CAMP₇ (c), PBLG₂₅ (d), PBLG₂₅ + CAMP₂₅ (1/1, n/n) (e) and CAMP₂₅ (f) assemblies characterized by DLS.

a



b



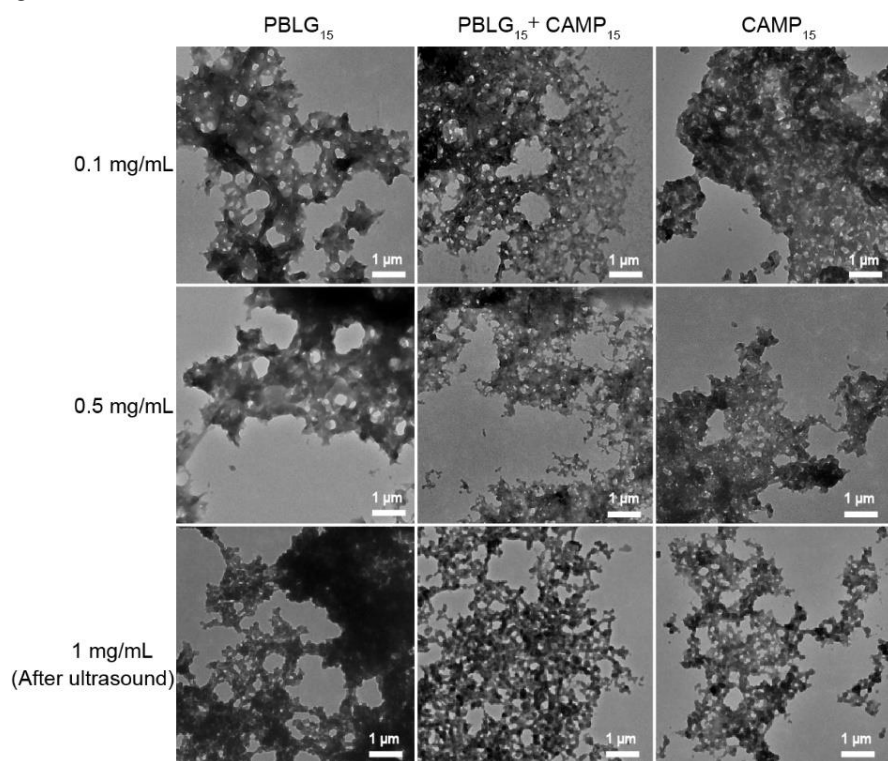
c

Fig. S5 a) TEM images of PBLG₇, PBLG₇ + CAMP₇ (1/1, n/n), CAMP₇ assemblies at different concentrations of 0.1 mg mL⁻¹, 0.5 mg mL⁻¹ and 1 mg mL⁻¹ b) TEM images of PBLG₂₅, PBLG₂₅ + CAMP₂₅ (1/1, n/n), CAMP₂₅ assemblies at different concentrations of 0.1 mg mL⁻¹, 0.5 mg mL⁻¹ and 1 mg mL⁻¹. c) TEM images of PBLG₁₅, PBLG₁₅ + CAMP₁₅ (1/1, n/n), CAMP₁₅ assemblies at different concentrations of 0.1 mg mL⁻¹, 0.5 mg mL⁻¹ and 1 mg mL⁻¹ after 20 minutes of ultrasound. Scale bar: 1 μm.

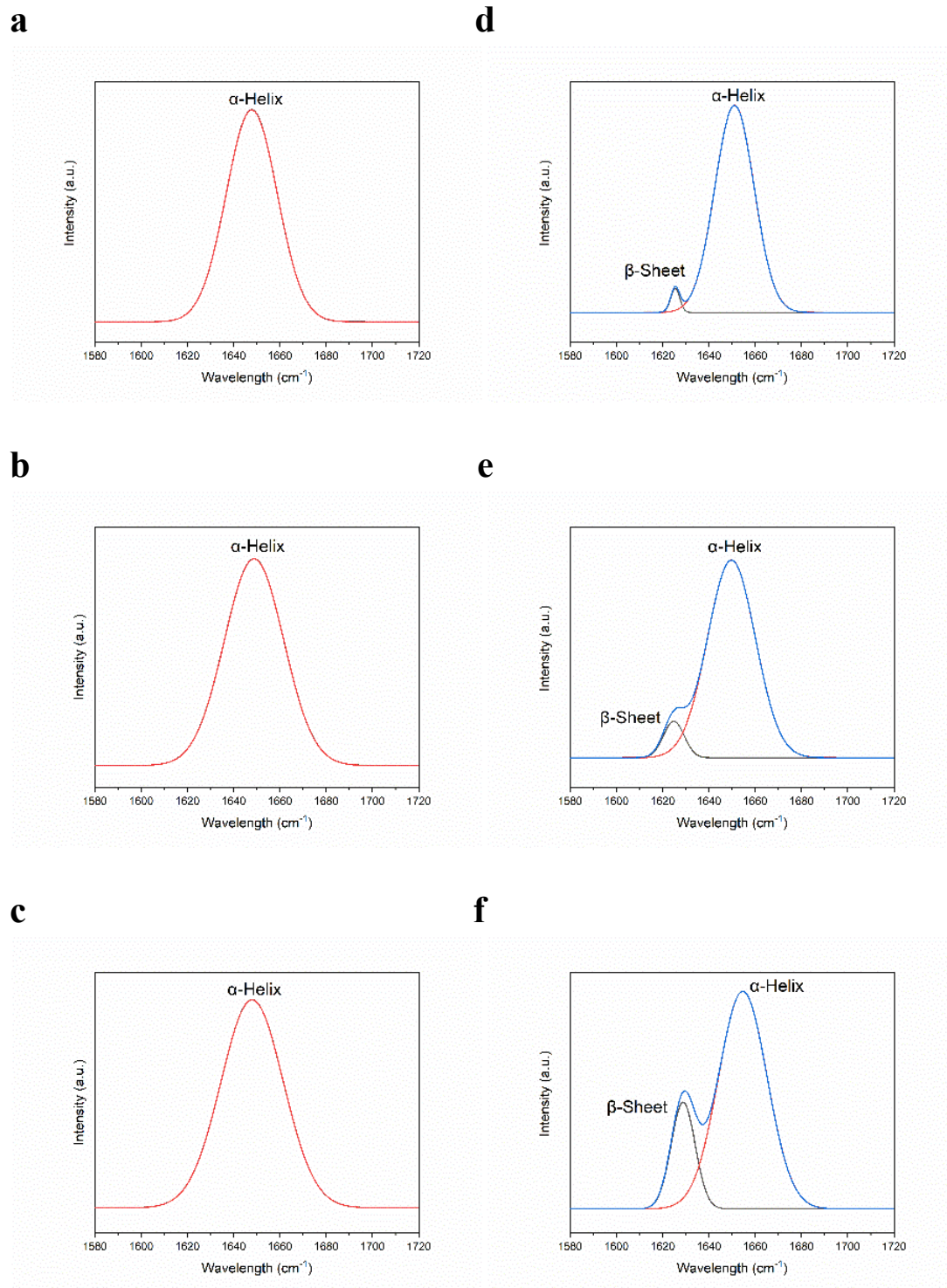


Fig. S6 FTIR spectroscopy and deconvolution of PBLG₇ (a), PBLG₇ + CAMP₇ (1/1, n/n) (b), CAMP₇ (c), PBLG₂₅ (d), PBLG₂₅ + CAMP₂₅ (1/1, n/n) (e) and CAMP₂₅ (f) assemblies at amide I spectral region.

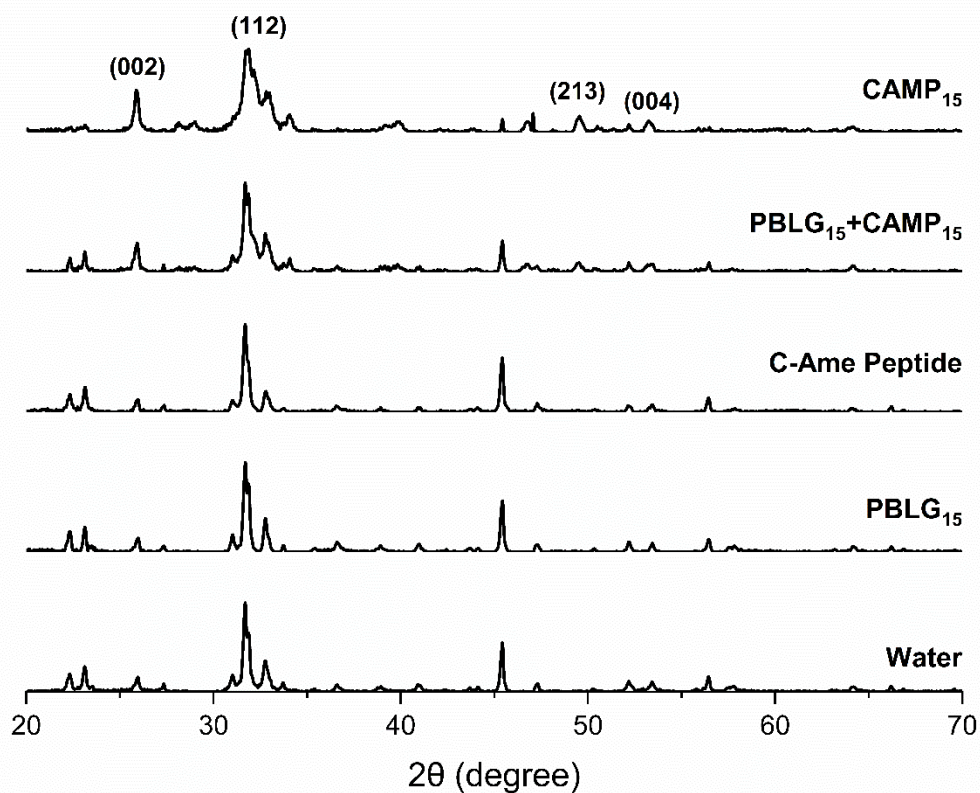
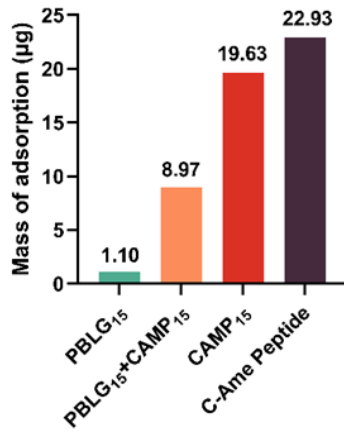
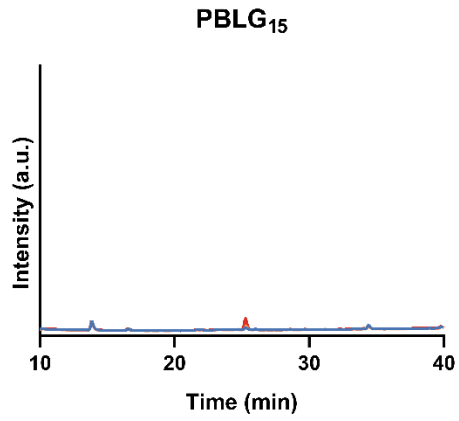
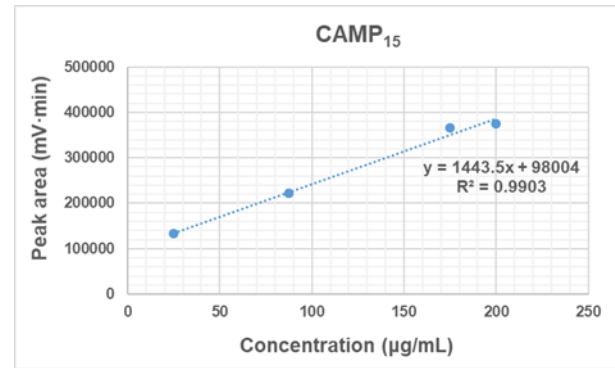
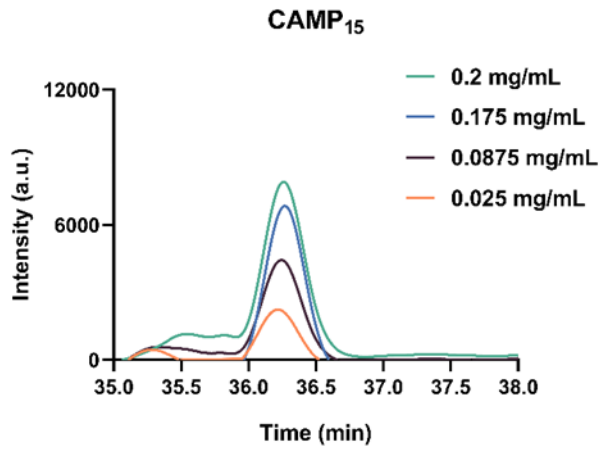
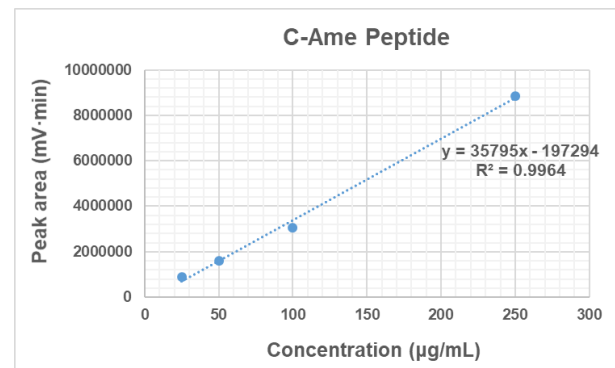
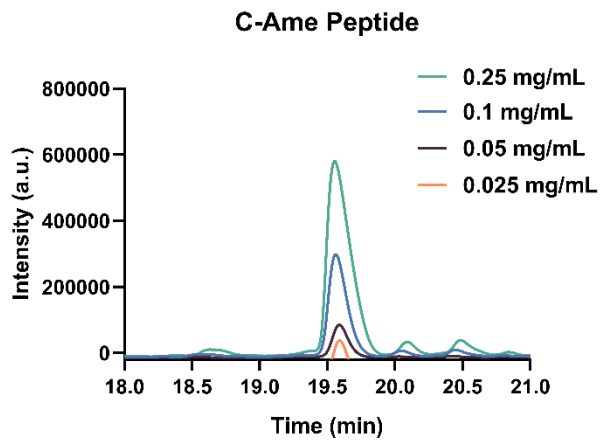
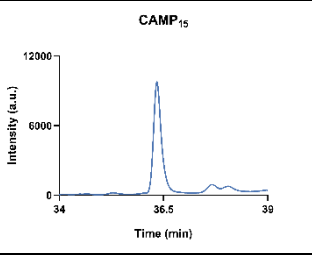
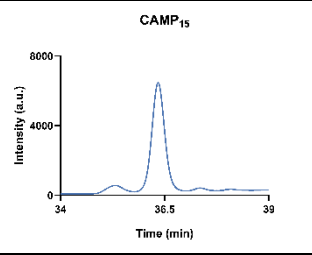
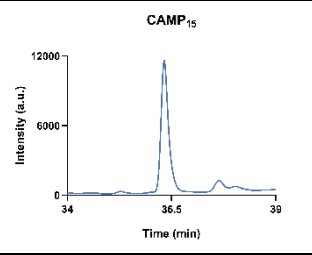


Fig. S7 XRD spectra of the mineralized products of Water, PBLG₁₅, C-Ame Peptide, PBLG₁₅ + CAMP₁₅ (1/1, n/n) and CAMP₁₅. NaCl was removed from the mineralization solution and the mineralized products were immediately frozen with liquid nitrogen and lyophilized for getting the powder sample to be tested.

a**b****c****d**

e

			
Peak area (mV·min)	129577	138558	146056
Concentration (µg/mL)	21.87	28.09	33.29
Mass of adsorption (µg)	10.94	14.05	16.64
Average mass of adsorption (µg)	13.88 ± 2.84		

f

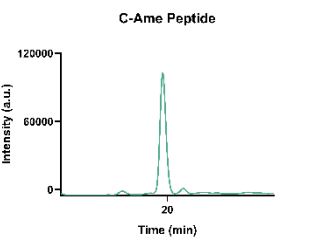
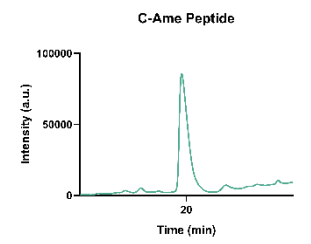
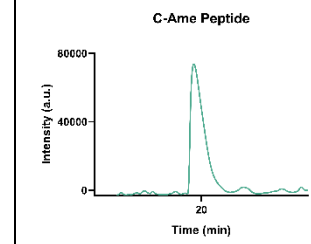
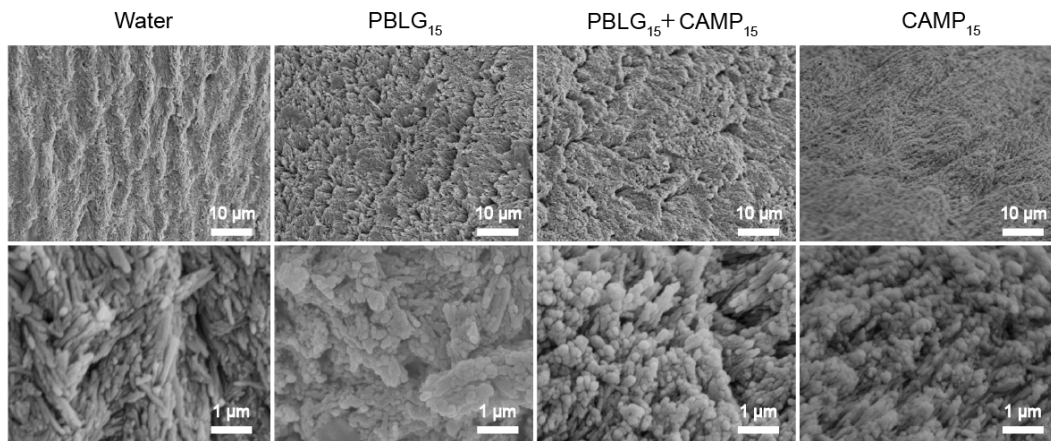
			
Peak area (mV·min)	859125	973297	1056289
Concentration (µg/mL)	29.51	32.70	35.02
Mass of adsorption (µg)	14.76	16.35	17.51
Average mass of adsorption (µg)	16.21 ± 1.38		

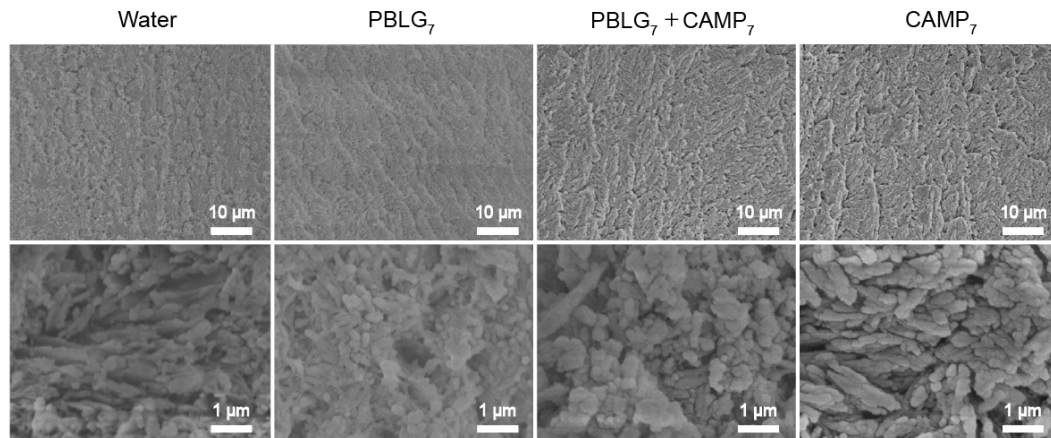
Fig. S8 Characterization of long-term adsorption capacity of polypeptides on enamel in artificial saliva. a) Average mass of PBLG₁₅, PBLG₁₅ + CAMP₁₅ (1/1, n/n), CAMP₁₅ and C-Ame Peptide coated on enamel slices using the BCA Protein Assay Kit. These data are the means (n=3). b) HPLC analysis of PBLG₁₅ after adding 500 µL 37% phosphoric acid and ultrasonic treatment. The analysis was conducted for three parallel determinations. Retention time of PBLG₁₅ is about 30.49 min in a linear gradient of 50-90% B for 40 min. c-d) Peak area-concentration standard curve of CAMP₁₅ (c) and C-Ame Peptide (d). e-f) HPLC analysis and average mass calculation of CAMP₁₅ (e) and C-Ame Peptide (f) after adding 500 µL 37% phosphoric acid and ultrasonic treatment. The

analysis was conducted for three parallel determinations. Retention time of CAMP₁₅ is about 36.41 min in a linear gradient of 30-80% B for 50 min. Retention time of C-Ame Peptide is about 19.96 min in a linear gradient of 5-45% B for 45 min. (HPLC solvent A: water, 0.06% TFA; B: 80% CH₃CN/water, 0.06% TFA)

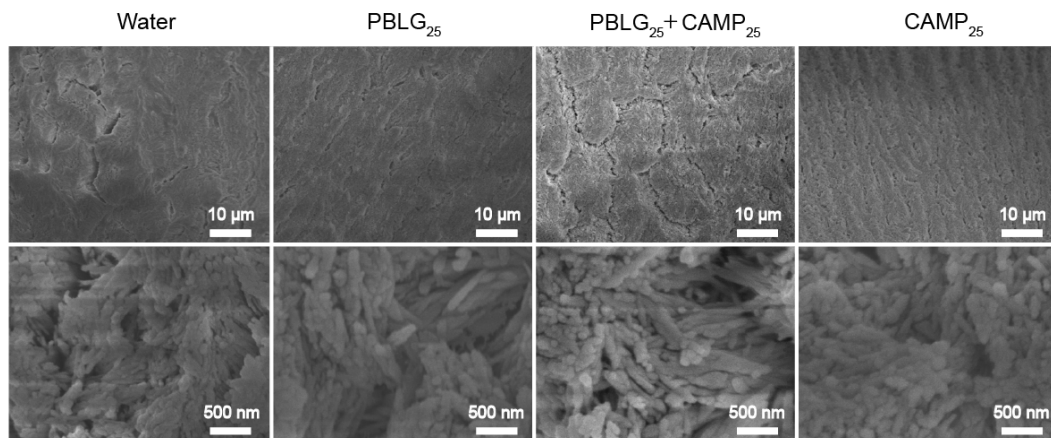
a



b



c



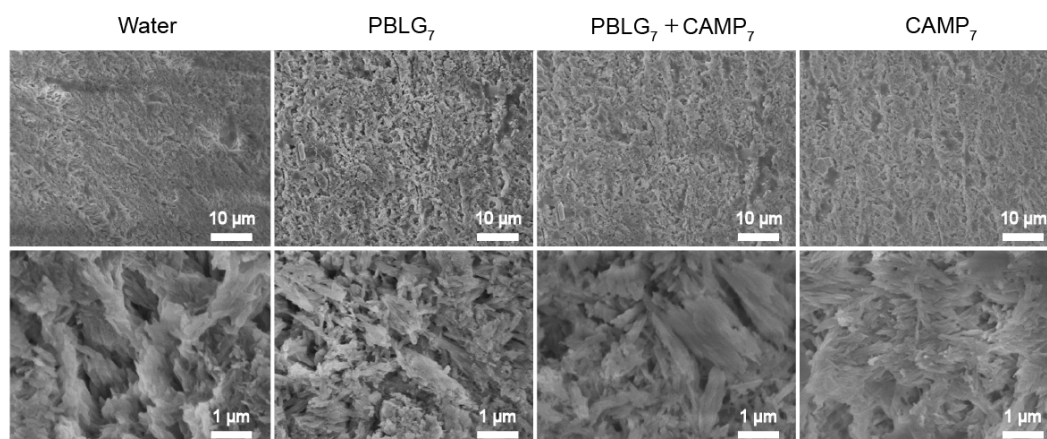
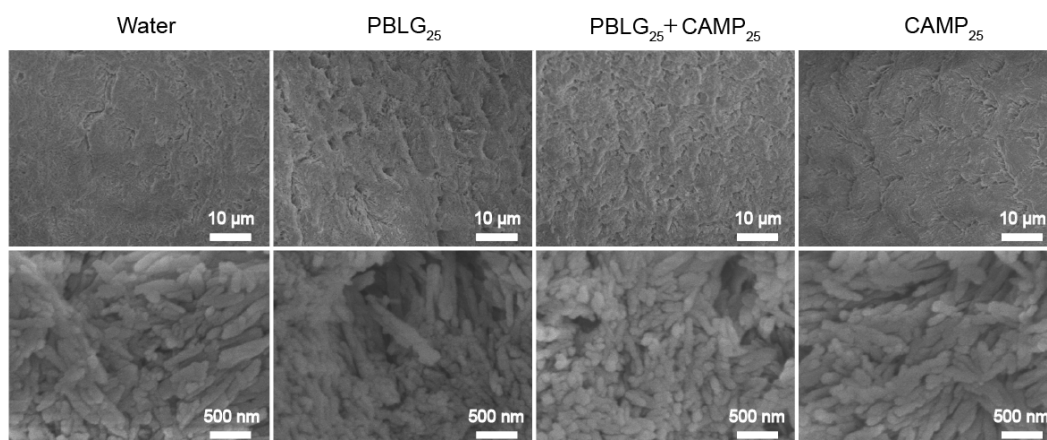
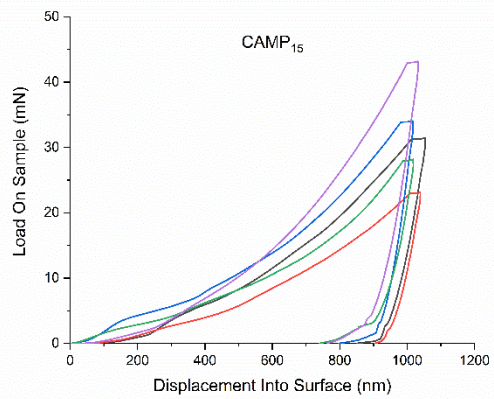
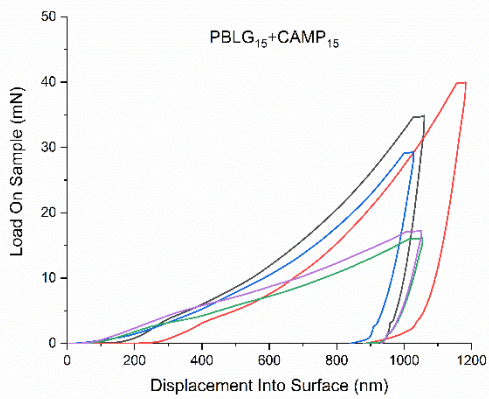
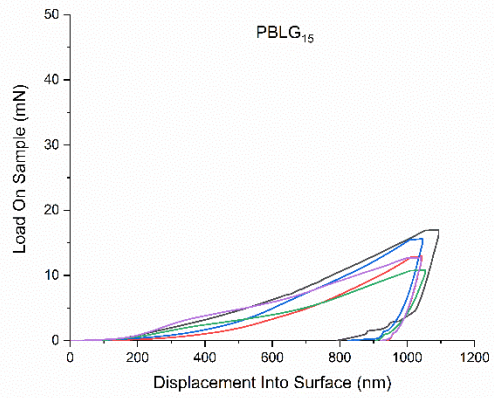
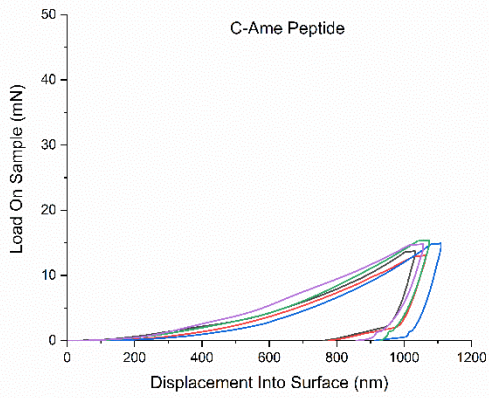
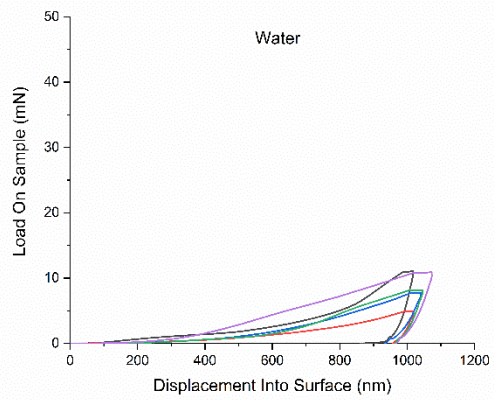
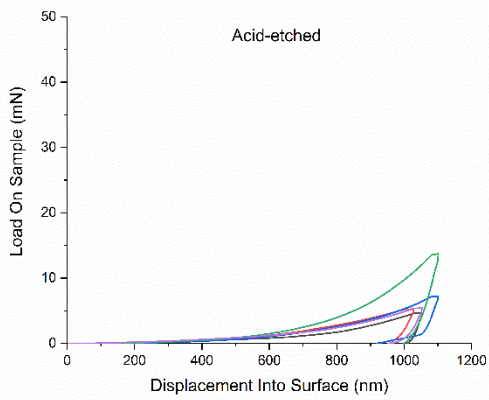
d**e**

Fig. S9 a) SEM low-magnification images and high-magnification images of Water-coated enamel, PBLG₁₅-coated enamel, PBLG₁₅ + CAMP₁₅ (1/1, n/n)-coated enamel, and CAMP₁₅-coated enamel after 3 days of incubation in artificial saliva. b) SEM low-magnification images and high-magnification images of Water-coated enamel, PBLG₇-coated enamel, PBLG₇ + CAMP₇ (1/1, n/n)-coated enamel, and CAMP₇-coated enamel after 3 days of incubation in artificial saliva. c) SEM low-magnification images and high-magnification images of Water-coated enamel, PBLG₂₅-coated enamel, PBLG₂₅ + CAMP₂₅ (1/1, n/n)-coated enamel, and CAMP₂₅-coated enamel after 3 days of incubation in artificial saliva. d) SEM low-magnification images and high-magnification images of Water-coated enamel, PBLG₇-coated enamel, PBLG₇ + CAMP₇ (1/1, n/n)-coated enamel, and CAMP₇-coated enamel after 6 days of incubation in artificial saliva. e) SEM low-magnification images and high-magnification images of Water-coated enamel, PBLG₂₅-coated enamel, PBLG₂₅ + CAMP₂₅ (1/1, n/n)-coated enamel, and CAMP₂₅-coated enamel after 6 days of

incubation in artificial saliva. Scale bars: 10 μm (a-e (top)), 1 μm (a, b, d (bottom)), and 500 nm (c, e (bottom)).



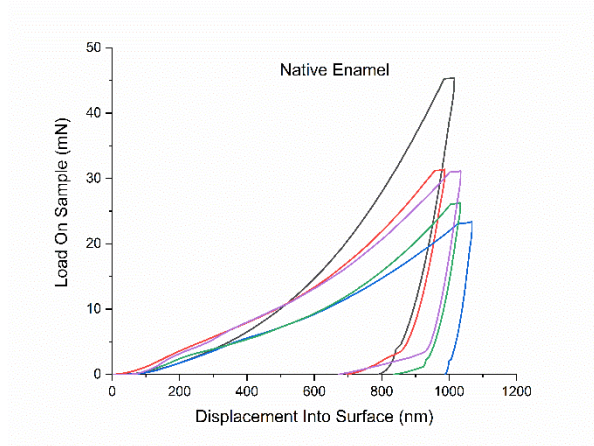
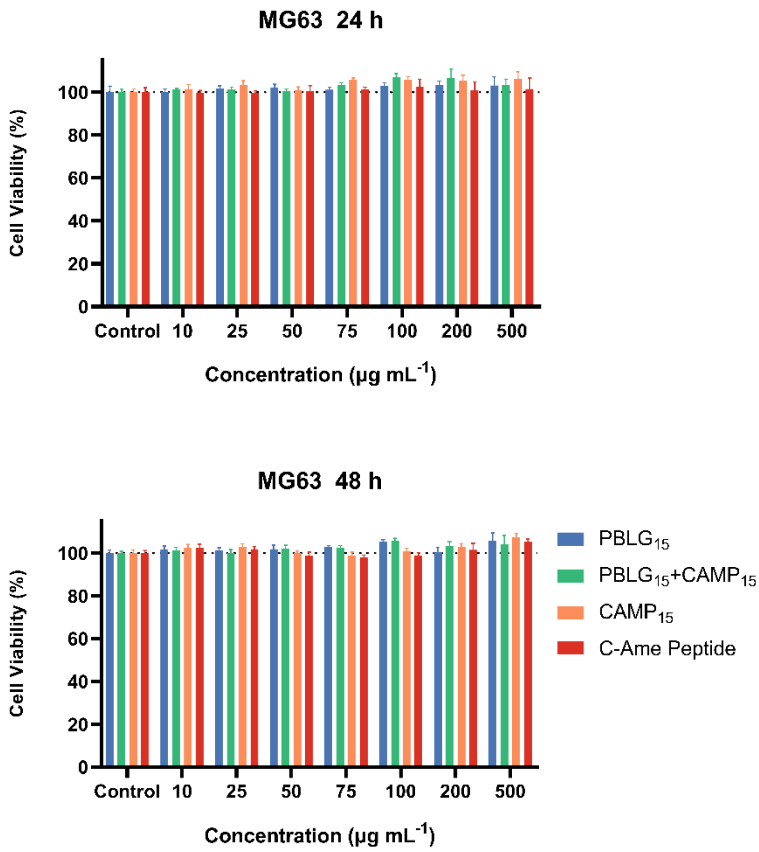
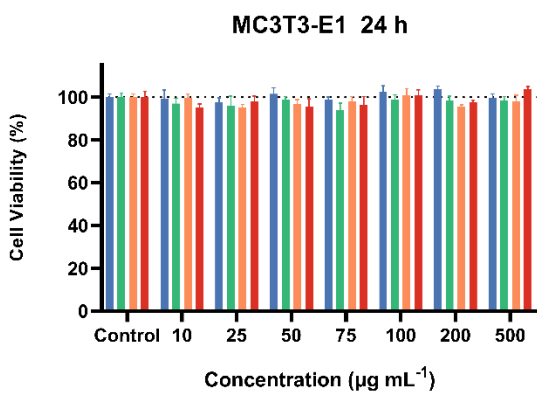


Fig. S10 Load-displacement curves in nano-indentation tests including Acid-etched enamel, Water-coated enamel, C-Ame Peptide-coated enamel, PBLG₁₅-coated enamel, PBLG₁₅ + CAMP₁₅ (1/1, n/n)-coated enamel, CAMP₁₅-coated enamel and Native enamel.

a



b



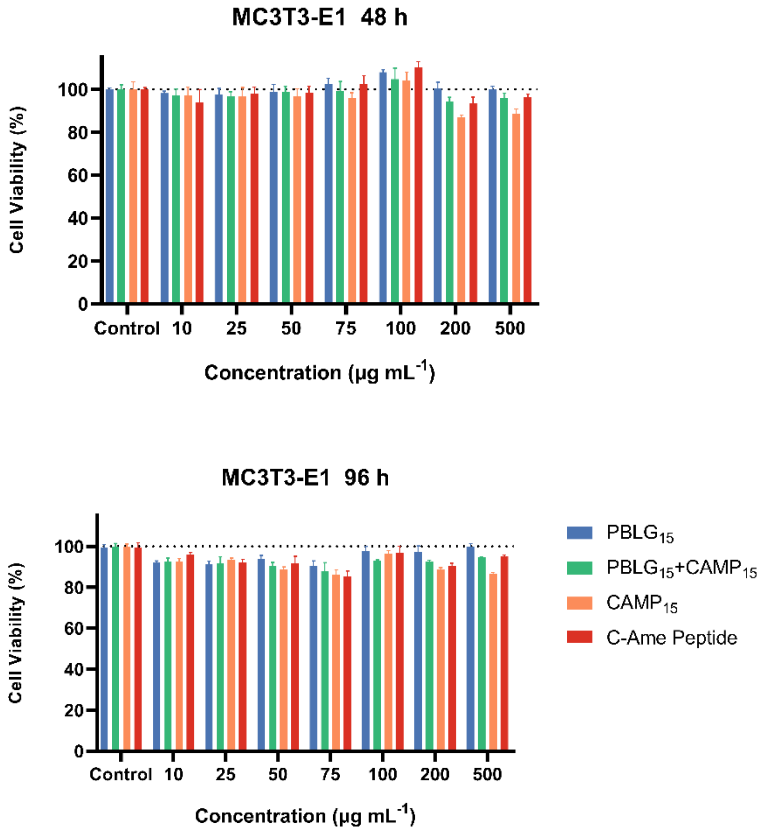
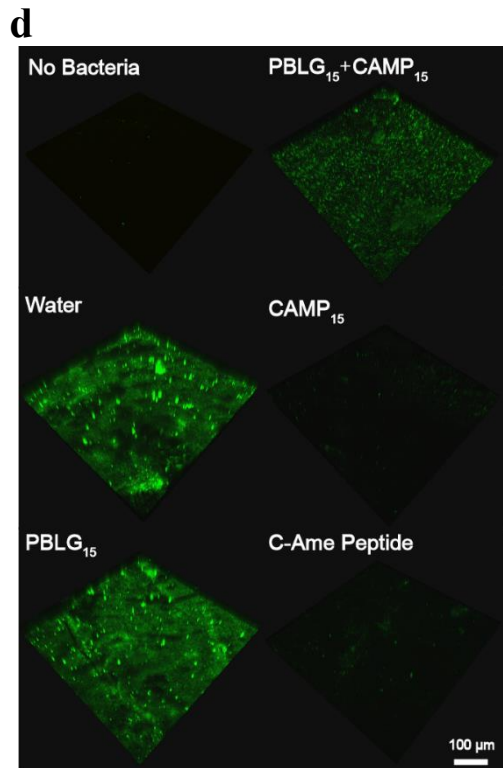
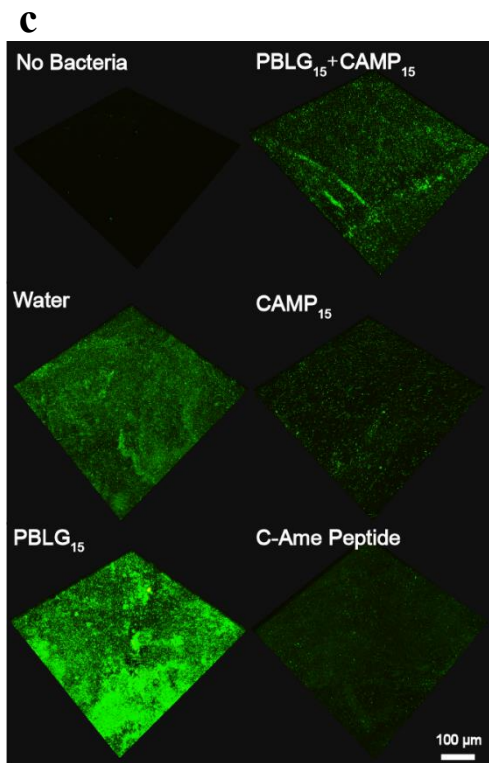
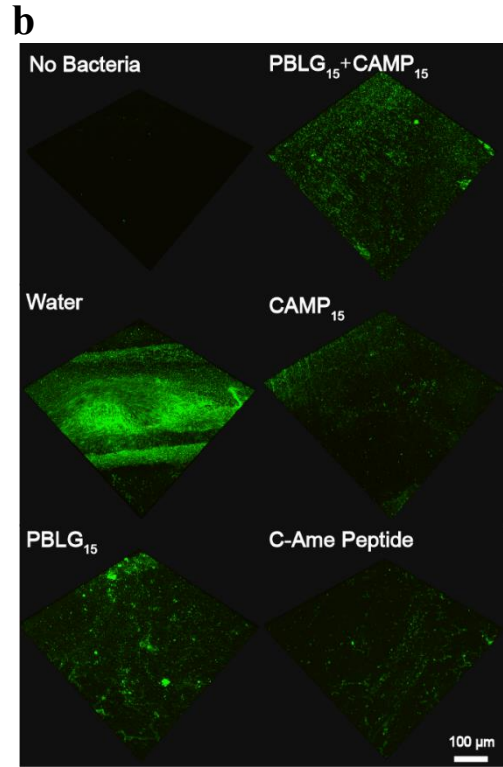


Fig. S11 Cell viability of MG63 cell for 24 h and 48 h (a), MC3T3-E1 cell for 24 h, 48 h and 96 h (b) at different concentrations (0, 10, 25, 50, 75, 100, 200, 500 $\mu\text{g mL}^{-1}$) of PBLG₁₅, PBLG₁₅ + CAMP₁₅ (1/1, n/n), CAMP₁₅ and C-Ame Peptide. These data are the means \pm SD (n=5).



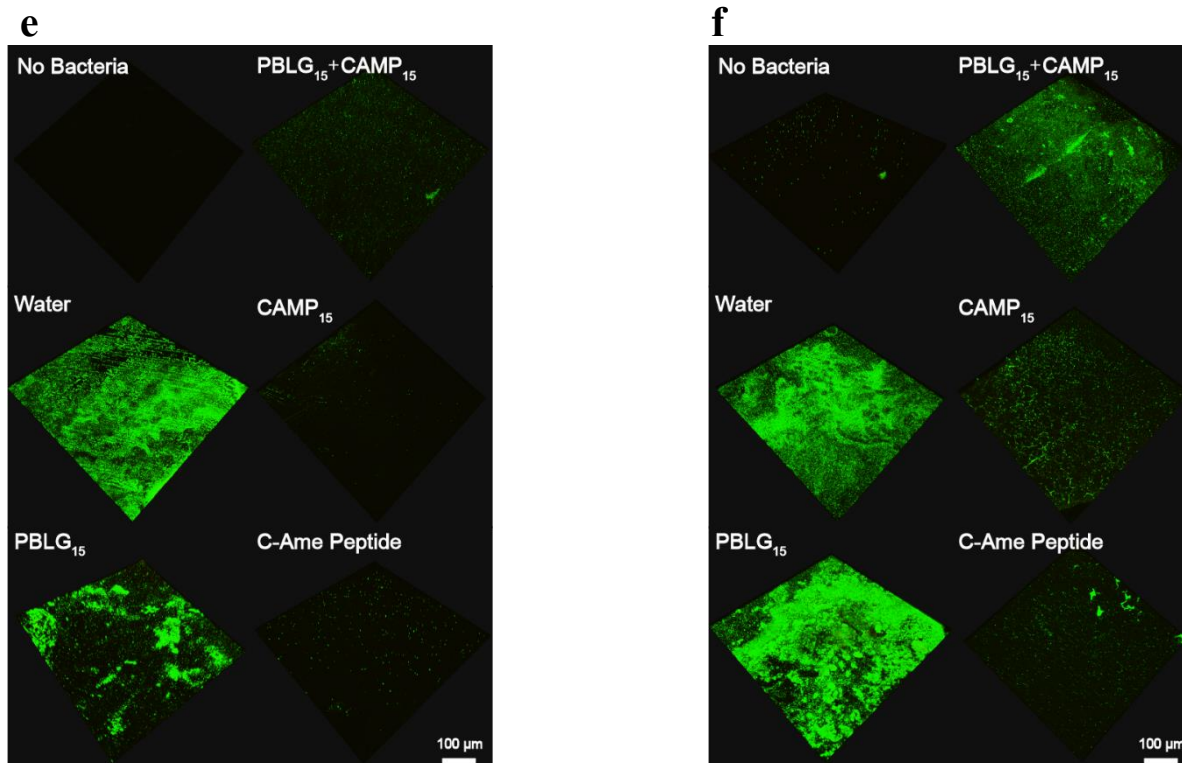


Fig. S12 CLSM images of dead *Streptococcus mutans* biofilm distribution after 24 hours (a), CLSM 3D images of *Streptococcus mutans* biofilm distribution after 48 hours (b) and 72 hours (c) on the different enamels coated with No Bacteria, Water, PBLG₁₅, PBLG₁₅ + CAMP₁₅ (1/1, n/n), CAMP₁₅ and C-Ame Peptide. CLSM 3D images of *Staphylococcus aureus* biofilm distribution after 24 hours (d), 48 hours (e) and 72 hours (f) on the different enamels coated with No Bacteria, Water, PBLG₁₅, PBLG₁₅ + CAMP₁₅ (1/1, n/n), CAMP₁₅ and C-Ame Peptide. Scale bar: 100 μm.