

## Asymmetrical Organic D- $\pi$ -A Conjugate with 'V'-Shape Crystal Packing: Quest to Transcend the Limits of Photophysical Properties and Applications

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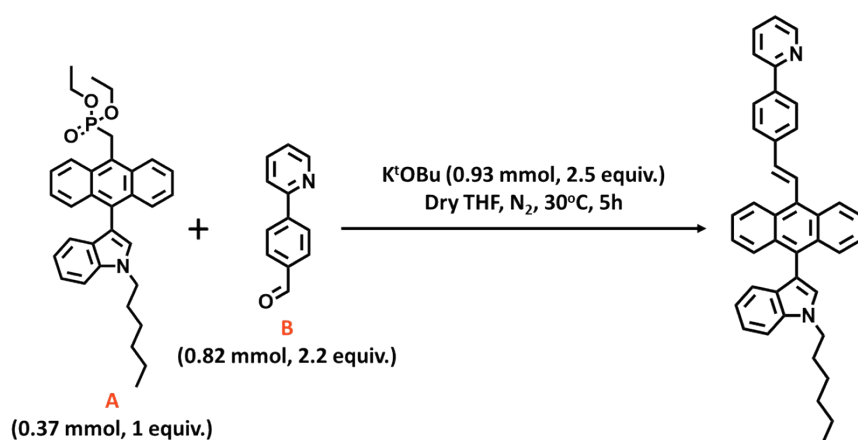
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**Scheme S1.** Synthetic pathway to prepare **(E)-1-hexyl-3-(10-(4-(pyridin-2-yl)styryl)anthracen-9-yl)-1H-indole (IAPY)<sup>1</sup>**

### Synthetic procedure and characterization of IAPY<sup>1</sup>

Diethyl ((10-(1-hexyl-1H-indol-3-yl)anthracen-9-yl)methyl)phosphonate (0.37 mmol, 1 equiv., 0.2 g), 4-(pyridin-2-yl)benzaldehyde (0.82 mmol, 2.2 equiv., 0.15 g) and K<sup>t</sup>OBu base (0.93 mmol, 2.5 equiv., 0.1 g) were taken in a two-neck round bottom flask and allowed for vacuum drying for 5 min. Later, N<sub>2</sub> was purged throughout the flask and kept sealed with a silicon septum. An N<sub>2</sub>-containing balloon was placed at one end, and at the other neck, dry THF (15 mL) was injected into the flask to stir for 5 h. The progress of the reaction was observed by eluting TLC into hexane/ EtOAc (12% EtOAc into hexane (v/v)) medium to see an intense green fluorescent product spot formed at R<sub>f</sub> = 0.29. The reaction mixture was then worked up with 30 mL of an EtOAc and brine water mixture of 1:2 EtOAc:H<sub>2</sub>O. The organic layer was separated, passed through a Na<sub>2</sub>SO<sub>4</sub> layer, finally to prepare a slurry with silica gel (60–120 mesh) for performing column chromatography (10% EtOAc into hexane (v/v)), which yielded (71%, 0.15 g) product which is a solid yellowish compound under room light but a bright yellowish-green fluorescent compound under a UV-365 nm bulb. NMR spectroscopy: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25 °C, δ): 8.74 (s, 1H), 8.46 (d, *J* = 8.76 Hz, 2H), 8.13–7.93 (m, 5H), 7.82–7.75 (m, 4H), 7.53–7.43 (m, 3H), 7.32–7.23 (m, 5H), 7.14–7.01 (m, 3H), 4.30 (t, *J* = 7.04 Hz, 2H), 1.99 (quint, *J* = 7.32 Hz, 2H), 1.46–1.23 (m, 6H), 0.90 (t, *J* = 7.04 Hz, 3H). <sup>13</sup>C NMR: (100 MHz, CDCl<sub>3</sub>, 25°C, δ): 156.9, 149.8, 138.8, 138.1, 136.9, 136.1, 132.4, 131.6, 130.2, 129.8, 129.7, 128.9, 127.9, 127.3, 127.2, 127.0, 126.2, 126.1, 125.3, 124.9, 122.2, 121.8, 120.6, 120.5, 119.5, 111.9, 109.6, 46.6, 31.5, 30.3, 26.8, 22.6, 14.0. IR (KBr, cm<sup>-1</sup>): 3445, 3048, 2944, 2849, 1580, 1467, 1431, 1345, 1322, 1233, 1147, 1017, 958. M.p.: 154–156°C. Mass spectrometry: HRMS (ESI) *m/z*: calcd for C<sub>41</sub>H<sub>36</sub>N<sub>2</sub> 556.2878, found: 557.2674 [M + H]<sup>+</sup>.

### Materials and Methods

#### Steady-state absorption and fluorescence measurements

Solid-state absorption spectra were recorded using a JASCO-500 spectrophotometer, and the solution-state absorption spectra were recorded using a UV-vis-NIR spectrophotometer (Hitachi F7000, Japan). Solution-state emission spectra were obtained using an FP-6300 spectrometer (JASCO), and by using a 10 mm path-length quartz cuvette, while the solid-state emission spectra were received using a fluorimeter (Fluorolog, HORIBA). All the emission spectra were obtained at the corresponding absorption wavelengths.

## Absolute/relative quantum yield and time-resolved lifetime decay measurement

The absolute quantum yield ( $\Phi_f$ ) values for solid samples were measured using the calibrated integrating sphere method with a fluorimeter (Fluorolog, HORIBA), and absolute errors within  $\sim \pm 2\%$  have been included while for the relative quantum yield (relative  $\Phi_f$ ) in the solution state, the error is  $\pm 5\%$ .

Time-resolved fluorescence measurements were measured using a time-correlated single-photon counting (TCSPC) unit (Horiba Deltaflex). The laser used for all the samples was of 510 nm. All the measurements were undergone at room temperature. The decay fitting was done by keeping the  $\chi^2$  value close to 1.

The average lifetime was obtained by fitting the decay profiles to a tri- or bi-exponential function eqn-1.  $\text{Fit} = A1.\exp(-t/\tau1) + A2.\exp(-t/\tau2) + A3.\exp(-t/\tau3) \dots\dots\dots(\text{eq-1})$   
 $\alpha1, \alpha2$  are the weighted components and  $\tau1, \tau2, \tau3$  are the individual lifetime components of the decay. The qualities of the fit were understood by judging the chi square ( $\chi^2$ ) values. The rate constants are calculated by using:  $k_r = [\Phi_f / \tau_{\text{avg}}] \text{ s}^{-1}$ ;  $k_{nr} = [1 - \Phi_f / \tau_{\text{avg}}] \text{ s}^{-1}$

## Powder X-ray diffraction and IR spectra

The PXRD spectra of the pristine, ground, and recovered forms of the compound **IAPY** were recorded by using a Rigaku Ultima IV X-ray diffractometer, and keeping the parameters constant for all samples, with a step width of 0.21 and a scan rate of 21 min<sup>1</sup> from 5–501 (Cu K $\alpha$  radiation,  $\lambda = 1.54 \text{ \AA}$ ). And an FT-IR spectrometer (FT/IR-4200, Jasco) was utilized to record the IR spectra of the samples. Solid samples were mixed with KBr to form pellets to record the spectra.

## Calculation of crystallinity from PXRD

Crystallinity is considered as the ratio between the area of crystalline peaks to the area of all peaks. Here the crystallinity is revealed as a percentage just by multiplying the ratio with 100. The deconvolution of the PXRD raw data was completed using oringin lab software (Origin Pro 8.5, student version) having a good increment of peaks. Then by selecting the plot the 'Analysis' menu should be clicked followed by the 'Peaks and Baseline' and 'Peak Analyzer' to reach to the 'Open Dialogue'. Here, the 'Peak Analyzer Window' will be opened to perform 'Integrate Peaks.' Followed by the 'Next' option, the 'Constant Y' should be selected to well set up the baseline. Again, 'Next' and 'Find'

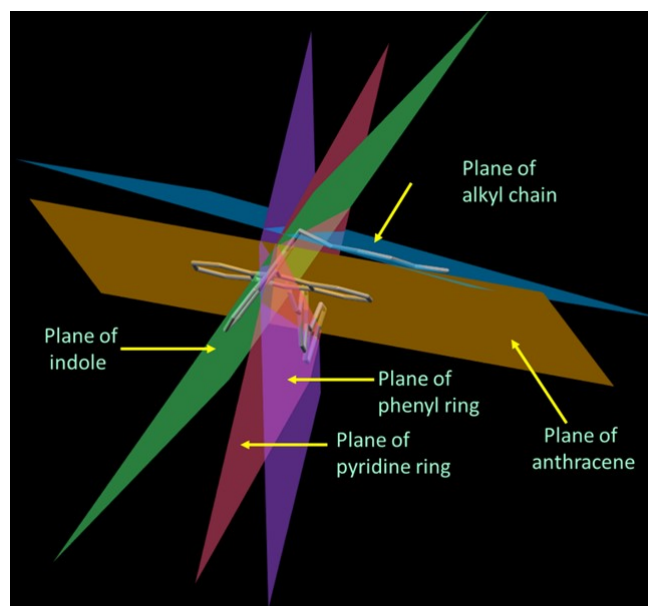
options should be clicked and 'Untick' to 'Enable Auto Find' should be done. Next 'Add' option should be clicked and all the relevant peaks should be selected accordingly. Next, the 'Done' option should be clicked. In the 'Peak Finding Setting' the 'Direction' should be selected as 'Positive' and 'Next' button should be clicked again. Here 'Adjust and Preview Graph' should be done. Next the 'Finish' button has to be clicked. Now, from the grown window, 'Select Data' then 'Select Integral Results' should be copied and pasted in a new excel sheet. It was named as 'Area of crystalline peaks'. Here sum it in excel. The area will be obtained. Therefore, the same process should be undergone again but this time not specific peaks but the 'Total area' should have to be integrated and the total area thus can be calculated using the excel sheet. Finally, the area under relevant crystalline peaks should be divided by the total area under the PXRD signal and multiplied by 100 to get the percentage of crystallinity.<sup>2</sup>

#### Differential scanning calorimetry analysis

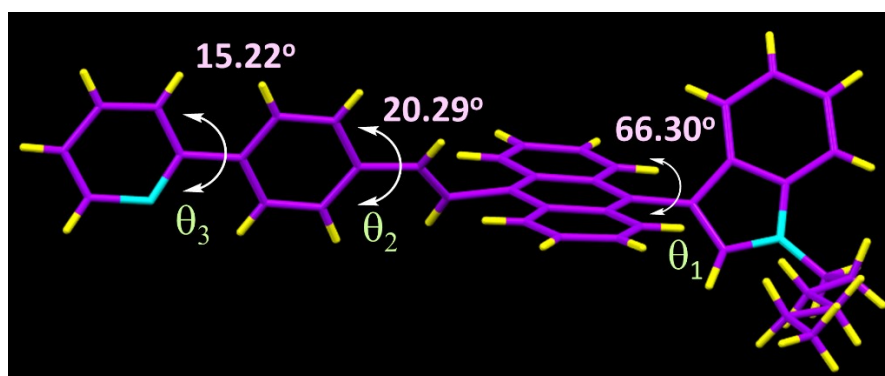
DSC thermograms were recorded by using a Themys One<sup>+</sup> (Setaram) instrument keeping the temperature in the range of 30–300 °C with a ramp rate of 2 °C min<sup>-1</sup> under a N<sub>2</sub> atmosphere.

#### Density functional theory studies

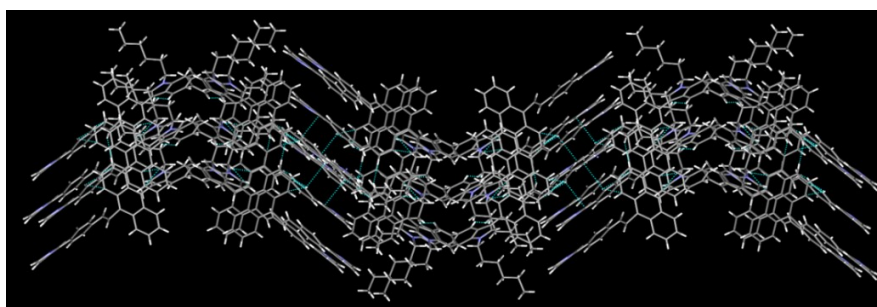
All the density functional theoretical (DFT) calculations were undergone using the ORCA Version 5.0.3 quantum chemical software package.<sup>3,4</sup> Our DFT calculations were done using different DFT functionals (viz., CAM-B3LYP, PBE0, B3LYP, WB97X, BHandHLYP and M06-2X) and 6-31G\* basis set to compare with experimental results.<sup>5</sup> The excited states (S<sub>1</sub>) geometry optimization was done using time-dependent DFT (TDDFT). Root mean square deviation (RMSD) calculations were performed using the Kabsch algorithm.<sup>6</sup> All structural and MOs were made visualized using Avogadro software.<sup>7</sup>



**Fig. S1** Different planes of IAPY

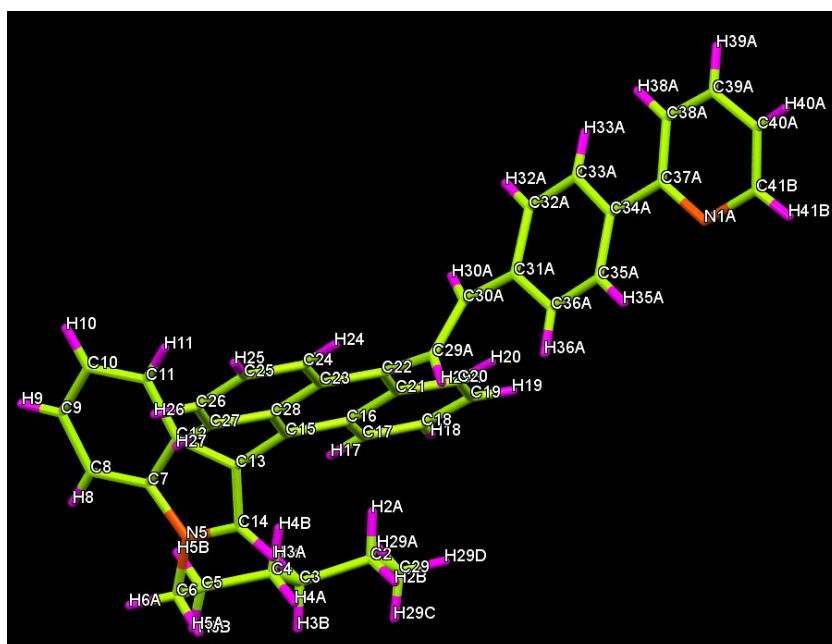


**Fig. S2** single-crystal with the dihedral angles in three different twisted sites



**Fig. S3** 'V'-shaped herringbone packing of IAPY

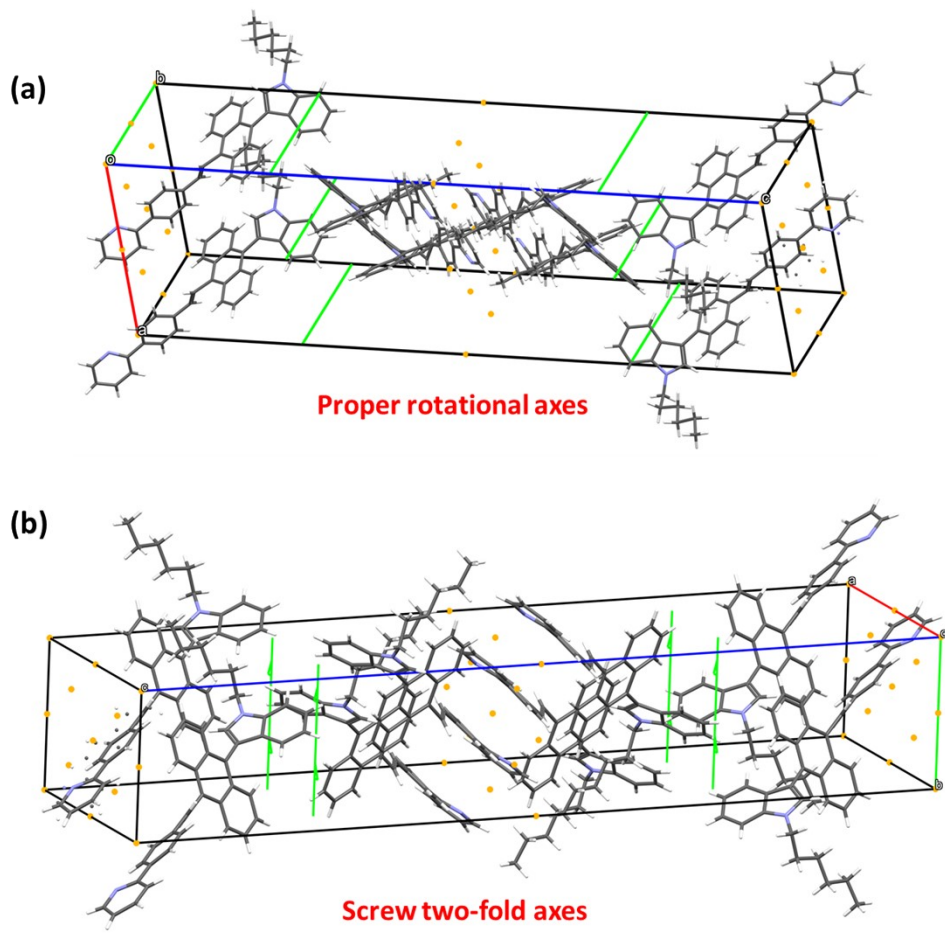




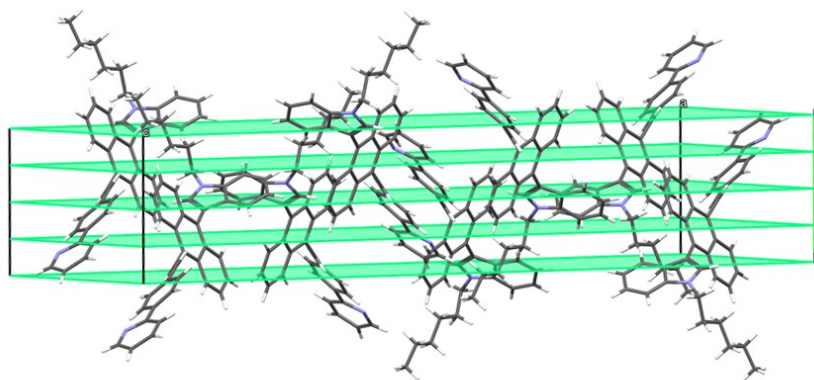
**Fig. S4** Label atom for IAPY

**Table S1** Short-contacts

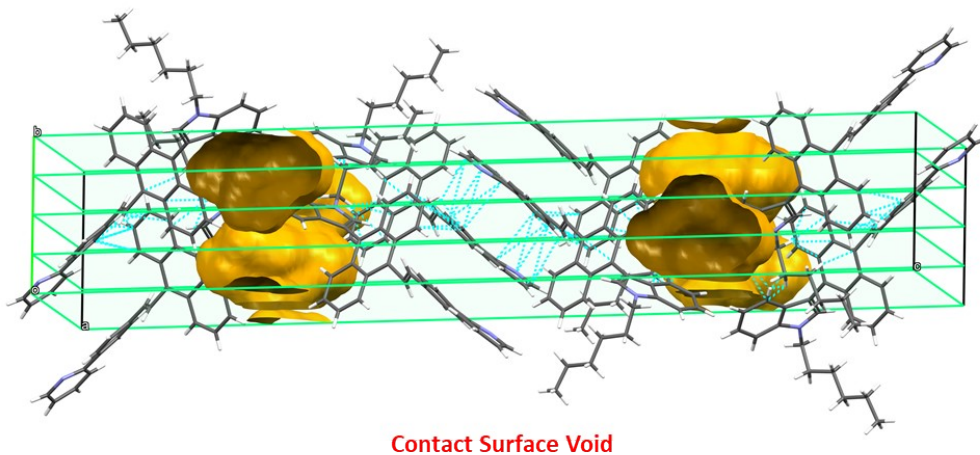
| No | Atom1 | Atom2 | Length (Å) |
|----|-------|-------|------------|
| 1  | C13   | H27   | 2.60       |
| 2  | C13   | H17   | 2.55       |
| 3  | C12   | C27   | 3.34       |
| 4  | C12   | H27   | 2.76       |
| 5  | N1A   | H35A  | 2.48       |
| 6  | C14   | C17   | 3.22       |
| 7  | C14   | H17   | 2.64       |
| 8  | C14   | C4    | 3.33       |
| 9  | C8    | H6A   | 2.89       |
| 10 | C24   | H29E  | 2.89       |
| 11 | H24   | C29A  | 2.46       |
| 12 | C20   | C30A  | 3.22       |
| 13 | H20   | C30A  | 2.63       |
| 14 | H20   | C29A  | 2.72       |
| 15 | C38A  | H33A  | 2.57       |
| 16 | H38A  | C33A  | 2.55       |
| 17 | H38A  | H33A  | 1.97       |
| 18 | H3B   | H29C  | 2.36       |
| 19 | H30A  | H32A  | 2.37       |
| 20 | C36A  | H29E  | 2.86       |
| 21 | H36A  | C29A  | 2.85       |
| 22 | H36A  | H29E  | 2.39       |



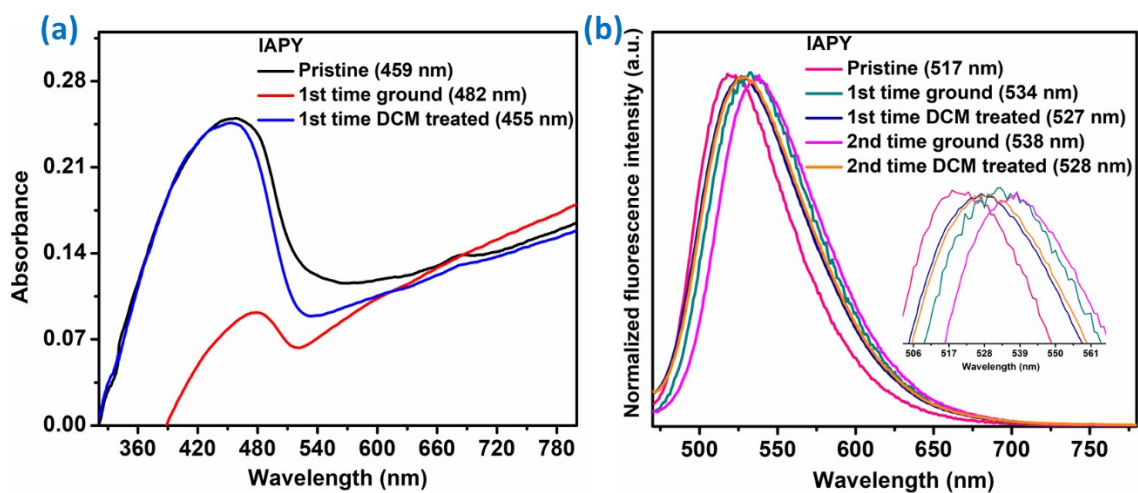
**Fig. S5** Rotational and screw-axes of IAPY



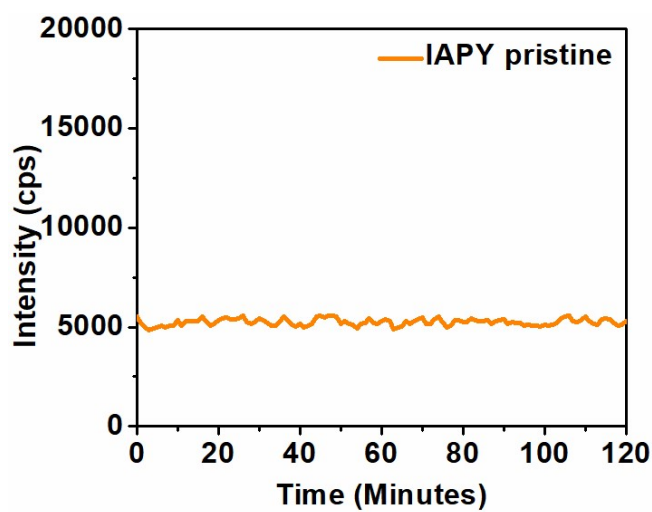
**Fig. S6** Glide plains in packing of IAPY



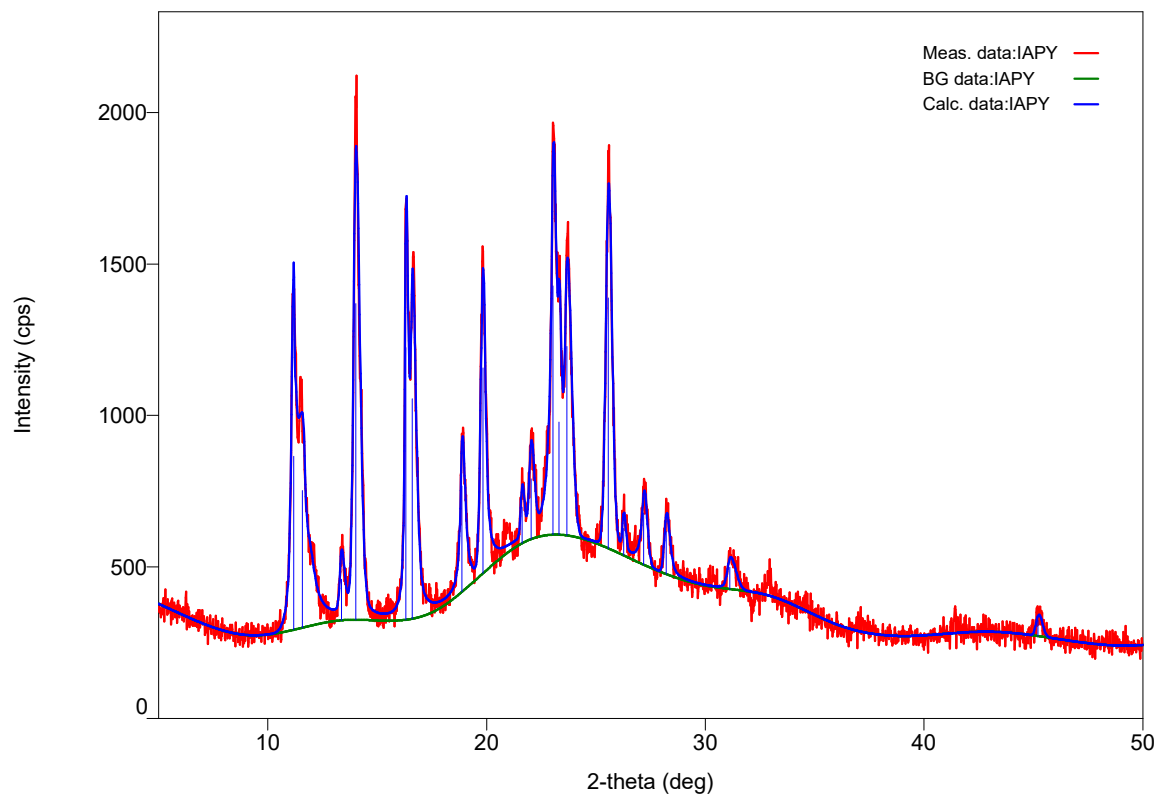
**Fig. S7** Contact surface void of IAPY



**Fig. S8a** Solid-state (a) absorbances (b) emission and MIEE properties of IAPY



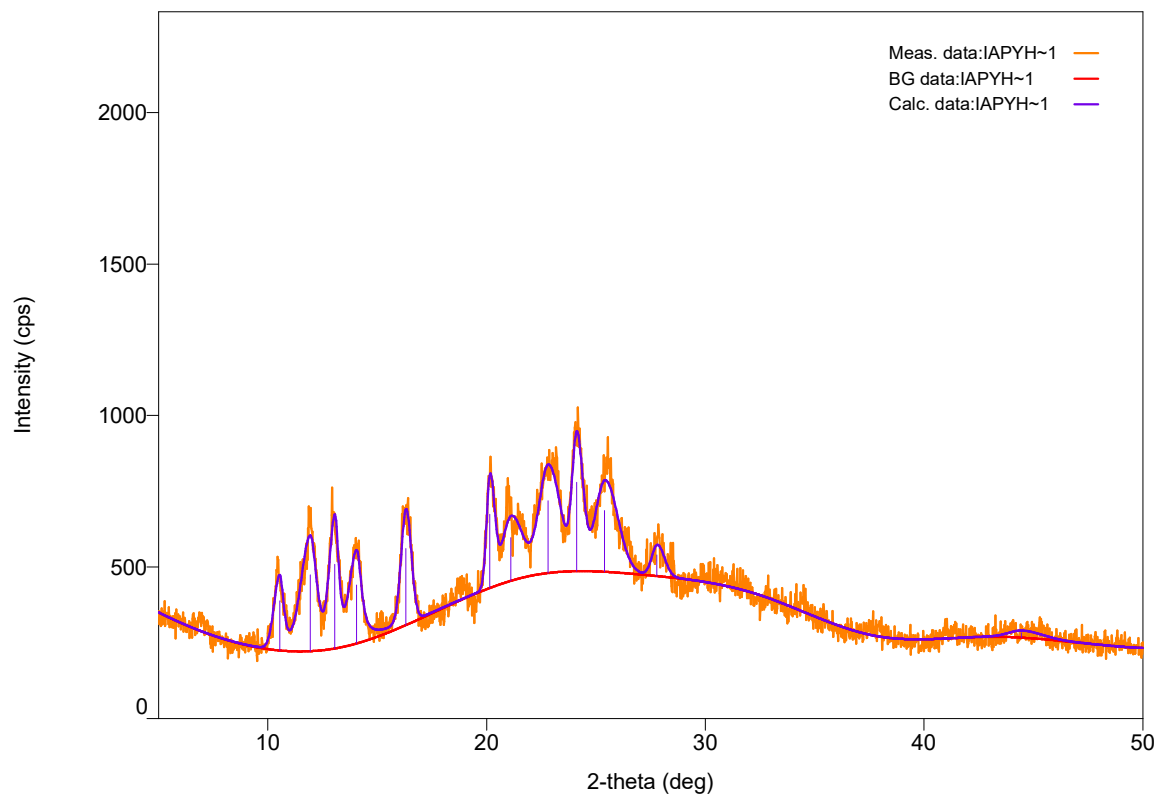
**Fig. S8b** Photostability of IAPY pristine



**Fig. S9** Software generated dendrogram for **IAPY** pristine

**Table S2** Peak list for **IAPY** pristine PXRD

| No. | 2-theta(deg) | d(ang.)    | Height(cps) | FWHM(deg) | Int. I(cps deg) | Int. W(deg) | Asym. factor |
|-----|--------------|------------|-------------|-----------|-----------------|-------------|--------------|
| 1   | 11.182(5)    | 7.907(3)   | 573(24)     | 0.166(10) | 133(11)         | 0.23(3)     | 2.2(2)       |
| 2   | 11.584(13)   | 7.633(8)   | 453(21)     | 0.64(2)   | 404(11)         | 0.89(7)     | 2.2(2)       |
| 3   | 13.366(10)   | 6.619(5)   | 135(12)     | 0.26(3)   | 41(4)           | 0.30(6)     | 0.58(6)      |
| 4   | 14.016(2)    | 6.3136(11) | 1044(32)    | 0.342(7)  | 420(8)          | 0.40(2)     | 0.58(6)      |
| 5   | 16.309(6)    | 5.430(2)   | 899(30)     | 0.177(11) | 217(13)         | 0.24(2)     | 0.53(11)     |
| 6   | 16.606(8)    | 5.334(3)   | 727(27)     | 0.291(15) | 288(14)         | 0.40(3)     | 0.53(11)     |
| 7   | 18.881(5)    | 4.6962(11) | 349(19)     | 0.254(18) | 134(5)          | 0.38(3)     | 0.6(2)       |
| 8   | 19.836(3)    | 4.4723(7)  | 678(26)     | 0.255(9)  | 221(6)          | 0.33(2)     | 1.1(2)       |
| 9   | 21.62(2)     | 4.106(4)   | 120(11)     | 0.20(5)   | 26(4)           | 0.22(6)     | 0.7(4)       |
| 10  | 22.03(3)     | 4.031(6)   | 199(14)     | 0.26(3)   | 57(7)           | 0.29(6)     | 0.7(4)       |
| 11  | 23.028(7)    | 3.8591(12) | 821(29)     | 0.26(2)   | 285(19)         | 0.35(3)     | 0.61(9)      |
| 12  | 23.309(12)   | 3.8132(19) | 372(19)     | 0.16(2)   | 79(13)          | 0.21(5)     | 0.61(9)      |
| 13  | 23.665(12)   | 3.7565(18) | 624(25)     | 0.382(17) | 323(13)         | 0.52(4)     | 0.61(9)      |
| 14  | 25.567(8)    | 3.4813(11) | 827(29)     | 0.325(7)  | 306(7)          | 0.37(2)     | 0.87(9)      |
| 15  | 26.267(14)   | 3.3901(17) | 89(9)       | 0.17(4)   | 17(3)           | 0.19(5)     | 0.87(9)      |
| 16  | 27.186(8)    | 3.2775(10) | 170(13)     | 0.28(2)   | 63(4)           | 0.37(5)     | 0.8(4)       |
| 17  | 28.22(3)     | 3.160(3)   | 140(12)     | 0.29(3)   | 51(4)           | 0.36(6)     | 0.7(3)       |
| 18  | 31.12(2)     | 2.872(2)   | 72(8)       | 0.44(6)   | 36(6)           | 0.50(14)    | 0.5(4)       |
| 19  | 45.21(9)     | 2.004(4)   | 52(7)       | 0.31(6)   | 17(4)           | 0.33(12)    | 0.7(8)       |



**Fig. S10** Software generated dendrogram for IAPY ground

**Table S3** Peak list for IAPY ground PXRD

| No. | 2-theta(deg) | d(ang.)   | Height(cps) | FWHM(deg) | Int. I(cps deg) | Int. W(deg) | Asym. factor |
|-----|--------------|-----------|-------------|-----------|-----------------|-------------|--------------|
| 1   | 10.540(14)   | 8.387(11) | 165(13)     | 0.52(3)   | 104(5)          | 0.63(8)     | 1.48(15)     |
| 2   | 11.94(2)     | 7.405(13) | 252(16)     | 0.84(3)   | 260(7)          | 1.03(9)     | 1.48(15)     |
| 3   | 13.062(12)   | 6.773(6)  | 278(17)     | 0.488(19) | 167(5)          | 0.60(5)     | 1.48(15)     |
| 4   | 14.057(17)   | 6.295(8)  | 193(14)     | 0.62(4)   | 148(7)          | 0.77(9)     | 1.48(15)     |
| 5   | 16.309(7)    | 5.431(2)  | 253(16)     | 0.546(18) | 152(6)          | 0.60(6)     | 0.96(15)     |
| 6   | 20.145(13)   | 4.404(3)  | 244(16)     | 0.43(2)   | 111(8)          | 0.46(6)     | 0.78(9)      |
| 7   | 21.12(3)     | 4.204(6)  | 142(12)     | 1.12(9)   | 170(10)         | 1.19(17)    | 0.78(9)      |
| 8   | 22.81(3)     | 3.896(5)  | 238(15)     | 1.15(5)   | 291(11)         | 1.22(12)    | 0.78(9)      |
| 9   | 24.113(13)   | 3.688(2)  | 294(17)     | 0.57(2)   | 180(8)          | 0.61(6)     | 0.78(9)      |
| 10  | 25.40(3)     | 3.504(4)  | 202(14)     | 1.21(6)   | 261(9)          | 1.29(14)    | 0.78(9)      |
| 11  | 27.79(3)     | 3.208(4)  | 69(8)       | 0.67(8)   | 50(5)           | 0.72(16)    | 0.78(9)      |
| 12  | 44.45(13)    | 2.037(6)  | 16(4)       | 1.8(4)    | 30(9)           | 1.9(10)     | 0.7(7)       |

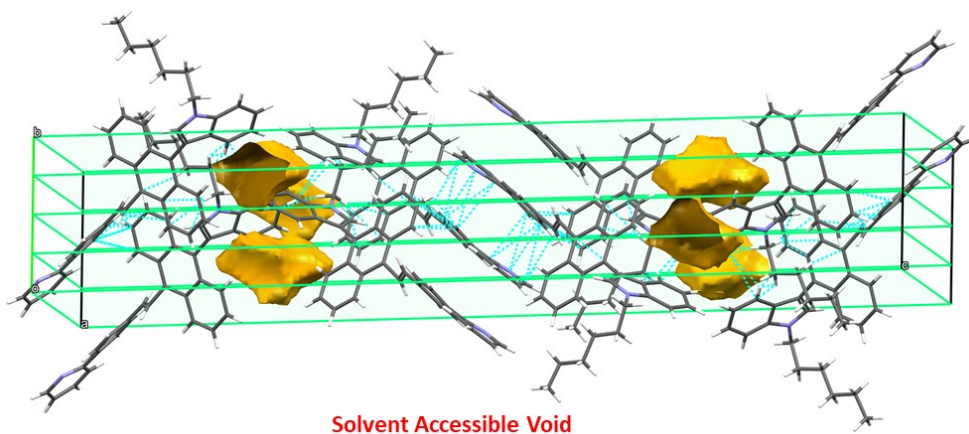


Fig. S11 Surface accessible void of IAPY

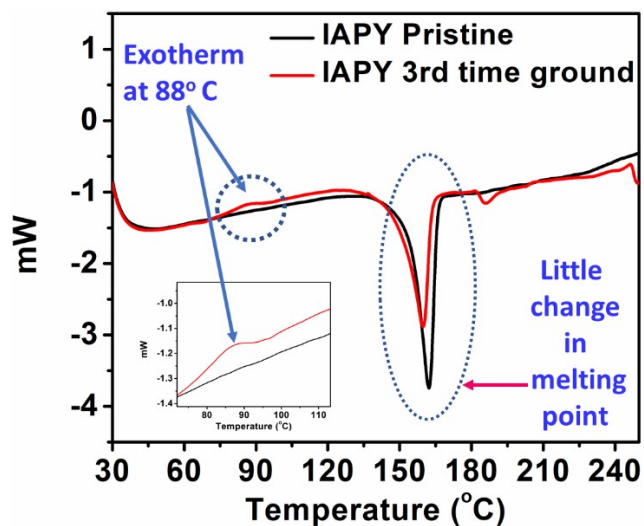


Fig. S12 DSC for the 3<sup>rd</sup> time ground form of IAPY

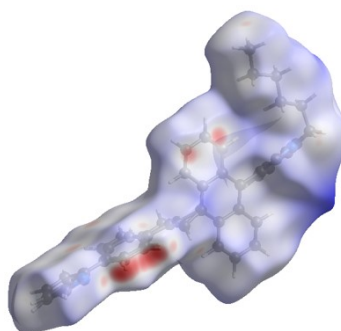
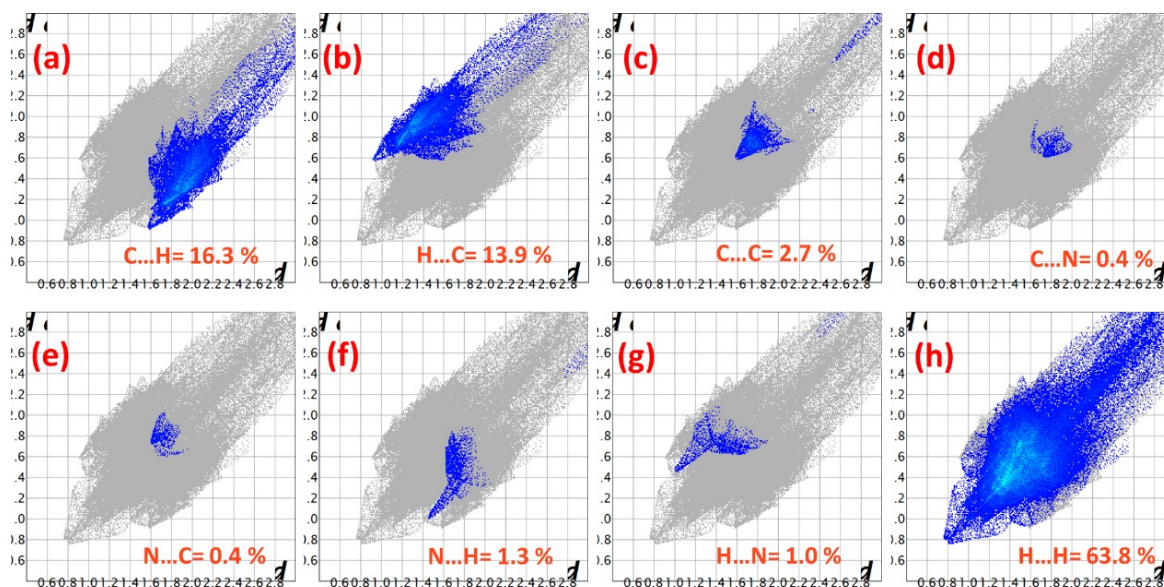


Fig. S13 Hirshfeld surface of IAPY





**Fig. S14** Hirshfeld surface and van der Waals and the non-covalent interactions to the packing for IAPY

**Table S4** Inter-molecular potentials

Calculated inter-molecular potentials:

mol1 mol2 distance energy (kJ/mol)

|    |    |         |          |
|----|----|---------|----------|
| 0  | 1  | 8.5606  | -56.7549 |
| 2  | 3  | 8.5606  | -56.7549 |
| 4  | 5  | 8.5606  | -56.7549 |
| 0  | 6  | 16.464  | -56.1803 |
| 2  | 7  | 16.464  | -56.1803 |
| 8  | 3  | 16.464  | -56.1803 |
| 0  | 2  | 8.66273 | -53.916  |
| 9  | 10 | 8.66273 | -53.916  |
| 11 | 7  | 8.66273 | -53.916  |
| 12 | 13 | 8.66273 | -53.916  |
| 7  | 6  | 8.66273 | -53.916  |
| 1  | 3  | 8.66273 | -53.916  |
| 4  | 14 | 8.66273 | -53.916  |

Hydrogen normalisation: On

Cluster Energy:

Cluster 1

PE = -122.19 kJ/mol 39 interactions  
 PE = -124.06 kJ/mol 119 interactions  
 PE = -124.16 kJ/mol 159 interactions  
 PE = -124.18 kJ/mol 179 interactions  
 PE = -124.19 kJ/mol 189 interactions  
 PE = -124.19 kJ/mol 199 interactions

Cluster 2

PE = -310.85 kJ/mol 39 interactions  
 PE = -319.49 kJ/mol 119 interactions

PE = -320.21 kJ/mol 159 interactions  
 PE = -320.42 kJ/mol 179 interactions  
 PE = -320.50 kJ/mol 189 interactions  
 PE = -320.55 kJ/mol 199 interactions

Total packing energy = -444.7 kJ/mol

$$\text{Potential} = A \cdot \exp(-Br) - Cr(-6)$$

Unified (UNI) pair-potential parameters:

| atom1 | code1 | atom2 | code2 | A        | B    | C      |
|-------|-------|-------|-------|----------|------|--------|
| C15   | 3     | C15   | 3     | 226145.2 | 3.47 | 2418.0 |
| C15   | 3     | N5    | 23    | 491494.0 | 3.86 | 2791.0 |
| C15   | 3     | H27   | 1     | 120792.1 | 4.10 | 472.8  |
| N5    | 23    | N5    | 23    | 365263.0 | 3.65 | 2891.0 |
| N5    | 23    | H27   | 1     | 228279.0 | 4.52 | 502.1  |
| H27   | 1     | H27   | 1     | 24158.0  | 4.01 | 109.2  |

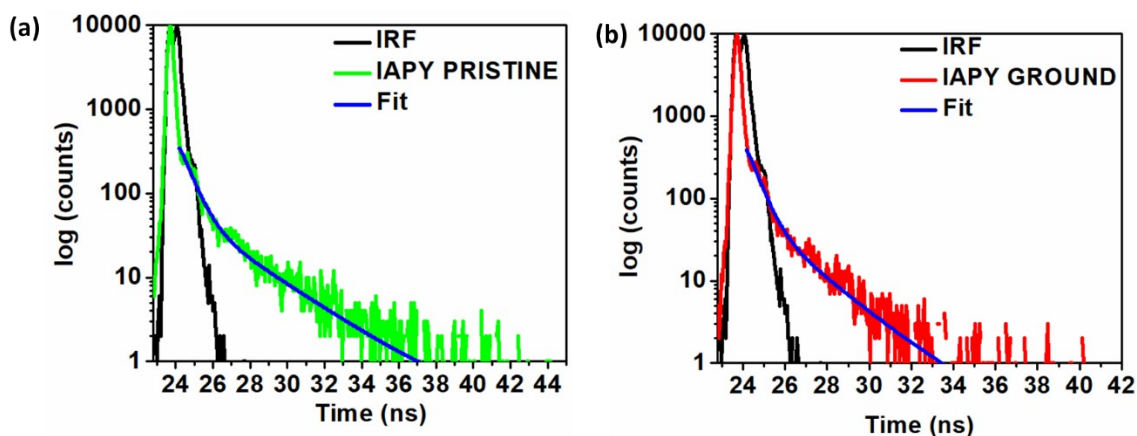


Fig. S15 Decay profile of IAPY for MIEE property

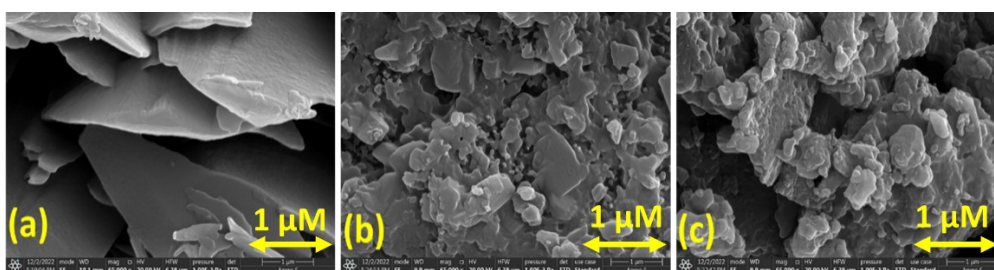


Fig. S16 SEM images for IAPY (a) pristine (b) ground (c) recovered



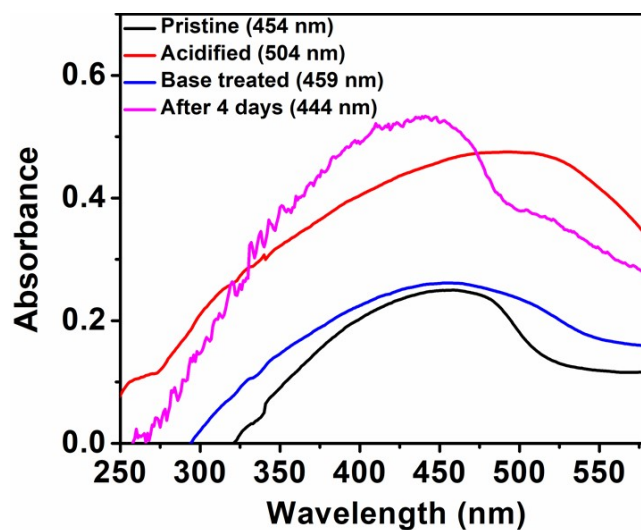


Fig. S17 Absorbances for IAPY SSAC

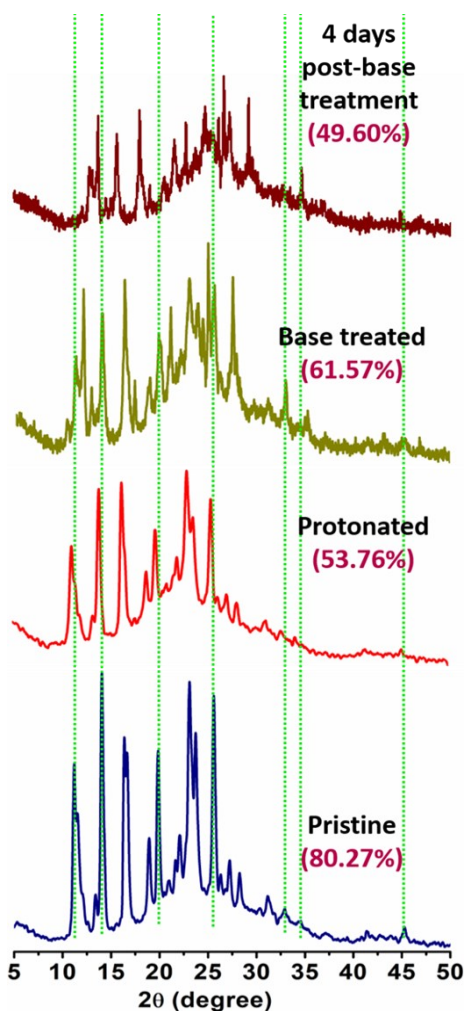


Fig. S18a Changes in PXRD pattern for IAPY due to acidofluorochromism

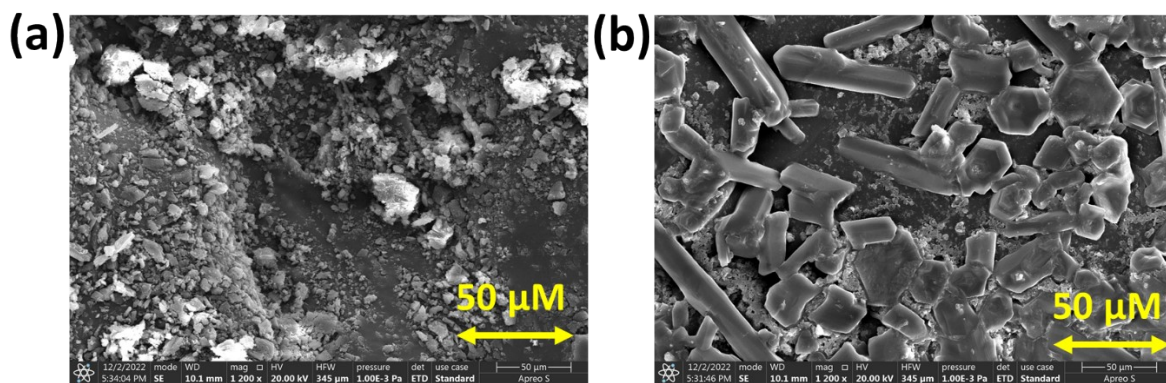


Fig. S18b SEM images for IAPY (a) acid-fumed (b) base-fumed

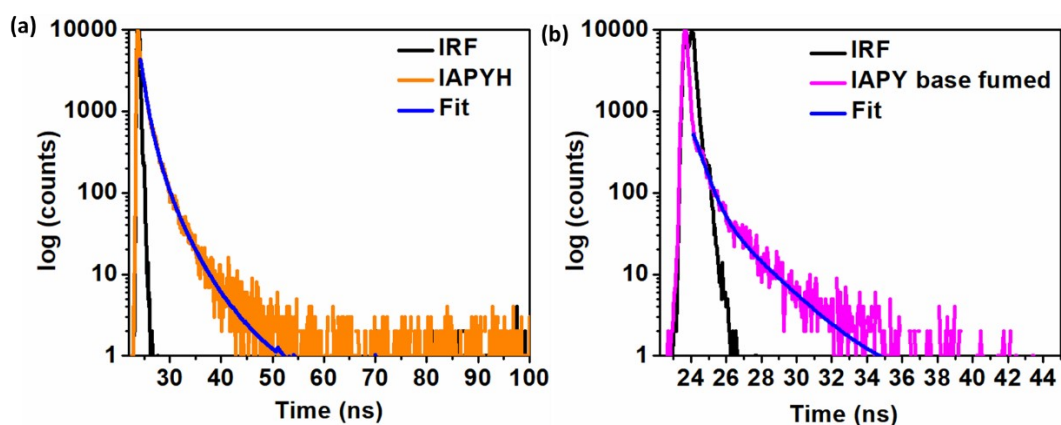


Fig. S19 Decay profile for IAPY (a) acid-fumed (IAPYH) (b) base-fumed

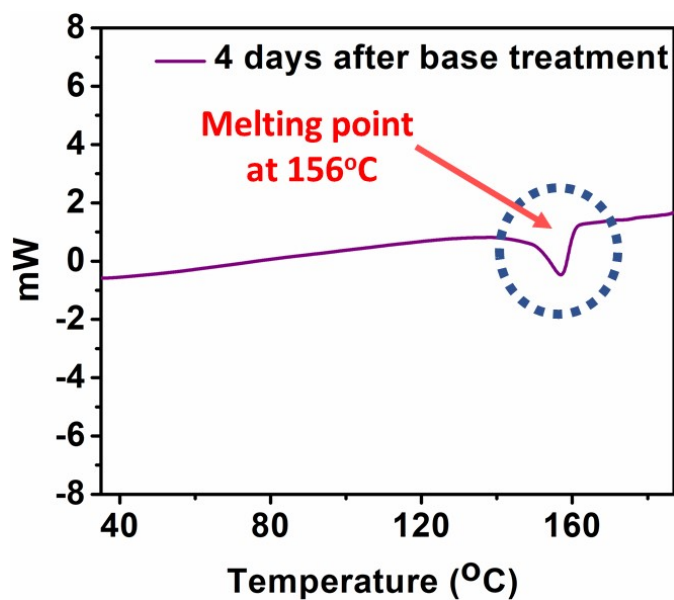


Fig. S20 DSC for the '4<sup>th</sup> day post to base treatment' form of IAPY

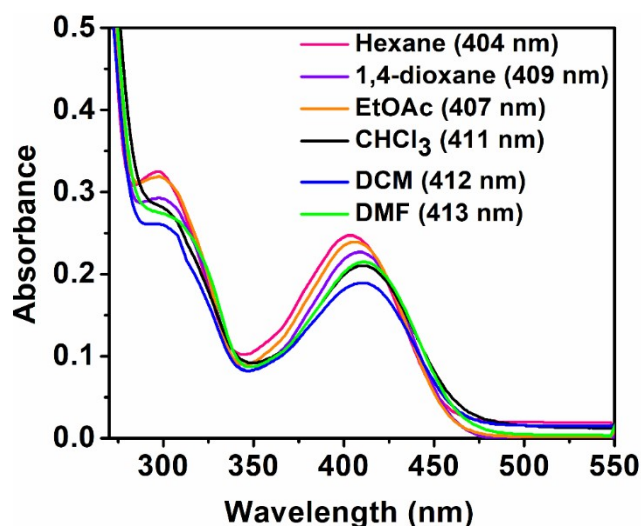


Fig. S21 Absorbances of IAPY in different solvents

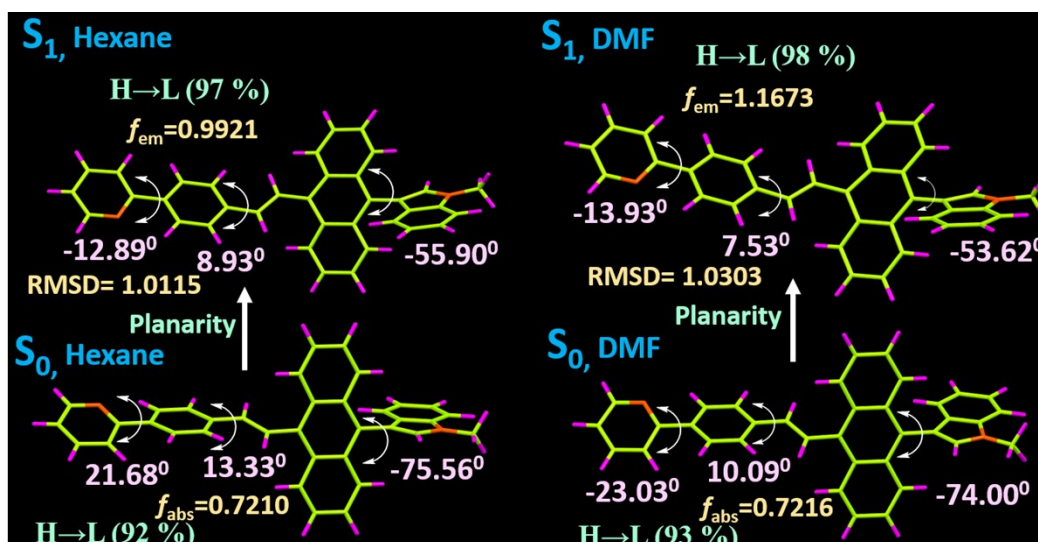


Fig. S22 TD-DFT optimized structures and calculations for IAPY in Hexane and N,N-DMF using CAM-B3LYP/6-31G\* level of theory.

Cyclic voltammetry study

Procedure and calculation: The HOMO/LUMO energy levels were calculated from the oxidation onset/reduction potentials in due course of positive/negative scans. For IAPY,  $10^{-5}$  M of DCM (dichloromethane) and  $10^{-5}$  M of N,N-DMF (dimethylformamide) solution were used with glassy carbon (GC) electrode as working electrode (WE) along with the Pt-wire and Ag/AgCl were used as a counter electrode (CE) and reference electrode (RE) respectively with a 0.1 M tetrabutylammonium perchlorate (TBAP) in the respective solvents as an electrolyte in a typical three-electrode system at normal condition.

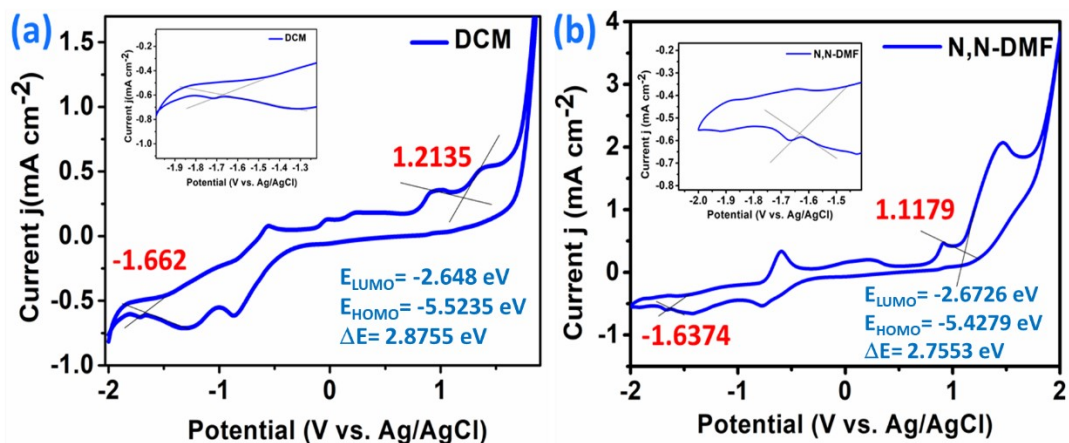


Fig. S23a HOMO-LUMO calculation from CV for IAPY in (a) DCM, and (b) N,N-DMF

$$E_{\text{HOMO}}(\text{DCM}) = -(E_{\text{OX Onset}} - 0.49 + 4.8) \text{ eV} = -(1.2135 - 0.49 + 4.8) \text{ eV} = -5.5235 \text{ eV}$$

$$E_{\text{LUMO}}(\text{DCM}) = -(E_{\text{Red Onset}} - 0.49 + 4.8) \text{ eV} = -(-1.662 - 0.49 + 4.8) \text{ eV} = -2.648 \text{ eV}$$

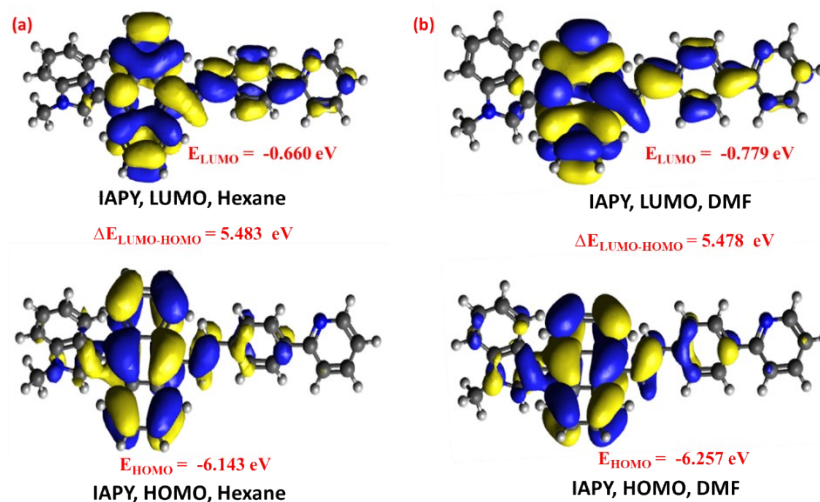
$$\text{Energy Gap (DCM)} = E_{\text{LUMO}} - E_{\text{HOMO}} = (-2.648 + 5.5235) \text{ eV} = 2.8755 \text{ eV}$$

Again,

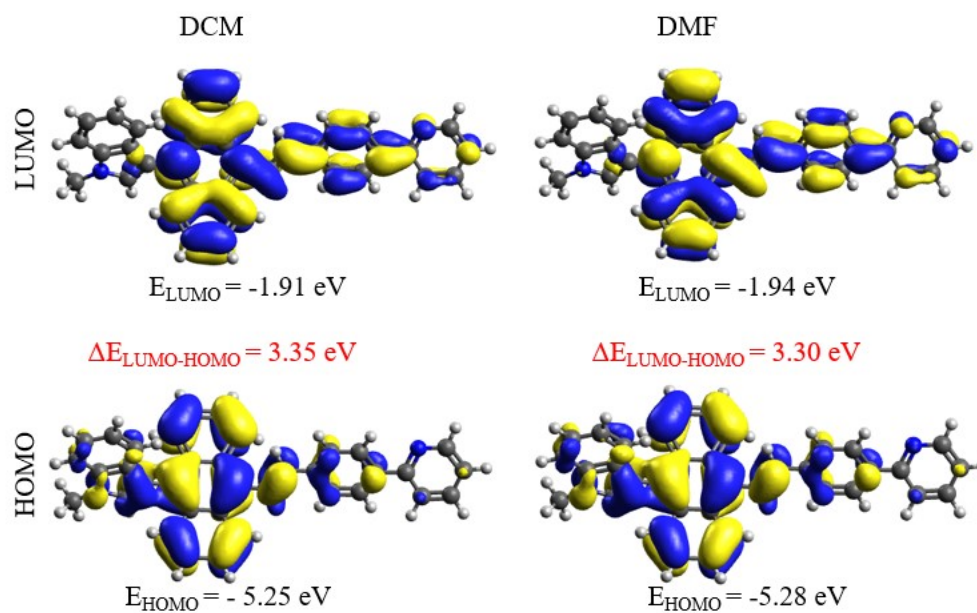
$$E_{\text{HOMO}}(\text{DMF}) = -(E_{\text{OX Onset}} - 0.49 + 4.8) \text{ eV} = -(1.1179 - 0.49 + 4.8) \text{ eV} = -5.4279 \text{ eV}$$

$$E_{\text{LUMO}}(\text{DMF}) = -(E_{\text{Red Onset}} - 0.49 + 4.8) \text{ eV} = -(-1.6374 - 0.49 + 4.8) \text{ eV} = -2.6726 \text{ eV}$$

$$\text{Energy Gap (DMF)} = E_{\text{LUMO}} - E_{\text{HOMO}} = (-2.6726 + 5.4279) \text{ eV} = 2.7553 \text{ eV}$$

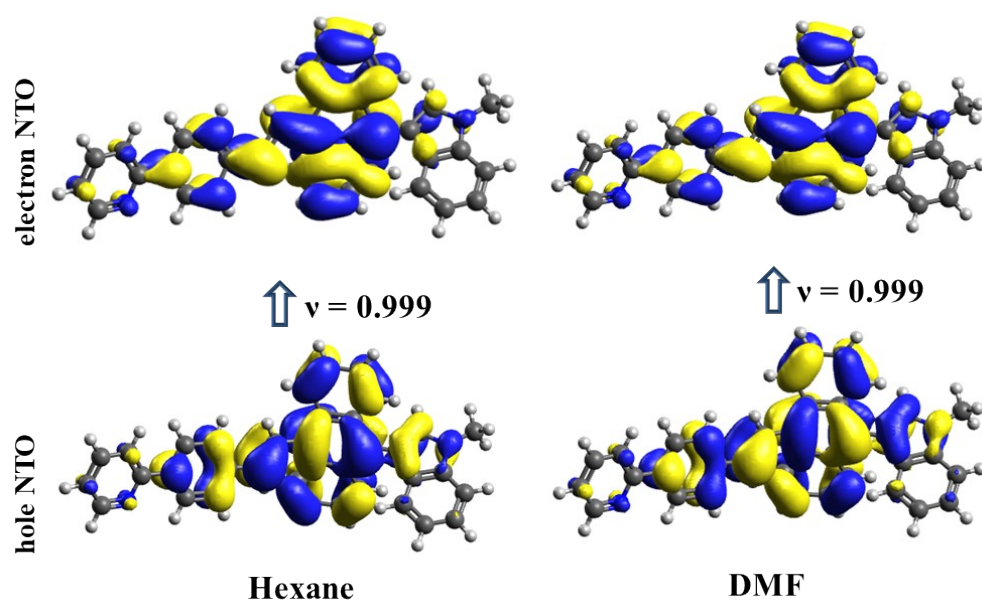


**Fig. S23b** HOMO-LUMO for IAPY in Hexane and DMF using CAM-B3LYP/6-31G\*

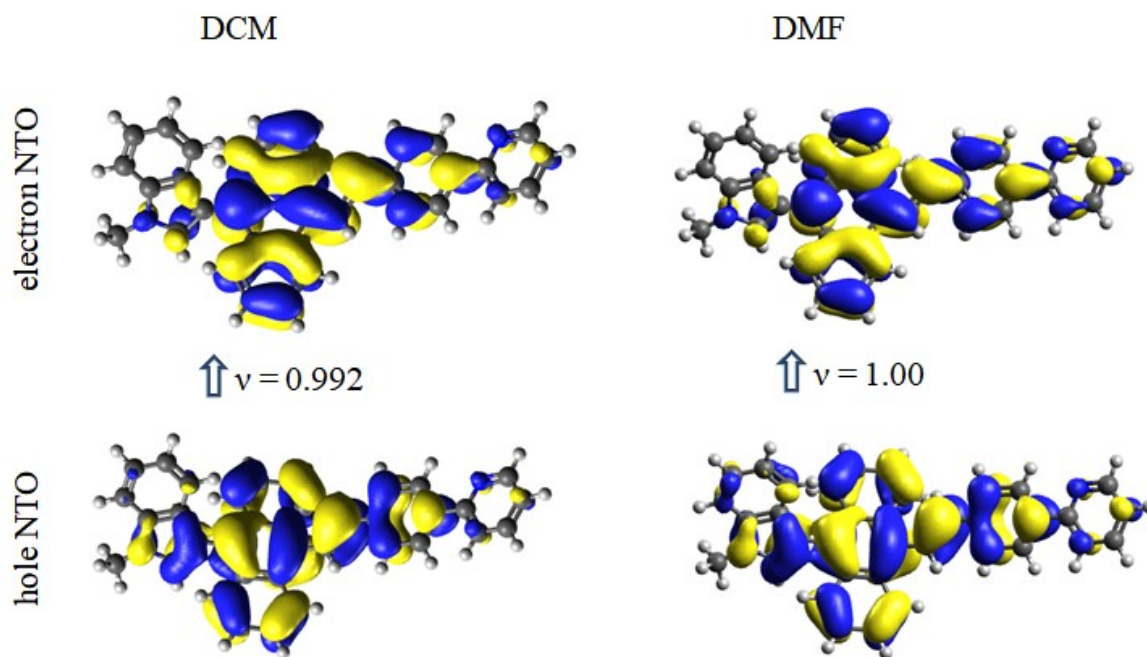


**Fig. S23c** HOMO-LUMO for IAPY in Hexane and DMF using PBE0/6-31G\* level of theory





**Fig. S23d** NTOs for **IAPY** in Hexane and DMF using CAM-B3LYP/6-31G\*



**Fig. S23e** Hole and electron NTOs in the  $S_1$  state of **IAPY** in DCM and DMF solvent with PBE0/6-31G\* level of theory

**Table S5** Calculation of Electronic Properties in DFT and DCM Solvent

| Properties/<br>Functionals      | B3LY<br>P    | CAM-<br>B3LYP | M062X        | BHnad<br>HLYP | PBE0         | WB97X        |
|---------------------------------|--------------|---------------|--------------|---------------|--------------|--------------|
| HOMO<br>(DMF)                   | -4.9482      | -6.2572       | -6.1992      | -5.8986       | -5.2752      | -7.18        |
| LUMO<br>(DMF)                   | -1.8683      | -0.779        | -1.2049      | -0.8648       | -1.9327      | 0.019        |
| <b>Energy<br/>gap<br/>(DMF)</b> | <b>3.080</b> | <b>5.478</b>  | <b>4.994</b> | <b>5.034</b>  | <b>3.343</b> | <b>7.199</b> |
| HOMO<br>(DCM)                   | -4.9274      | -6.2353       | -6.2675      | -5.8744       | -5.2505      | -7.1559      |
| LUMO<br>(DCM)                   | -1.8461      | -0.7561       | -1.1696      | -0.8399       | -1.9057      | 0.0456       |
| <b>Energy<br/>gap<br/>(DCM)</b> | <b>3.081</b> | <b>5.479</b>  | <b>5.098</b> | <b>5.035</b>  | <b>3.345</b> | <b>7.202</b> |

To bring clarity, we have calculated the  $S_0$  state HOMO-LUMO energy gap using the different functions enlisted above.

**Table S6** Optimized Ground State ( $S_0$ ) Geometry in DCM solvent

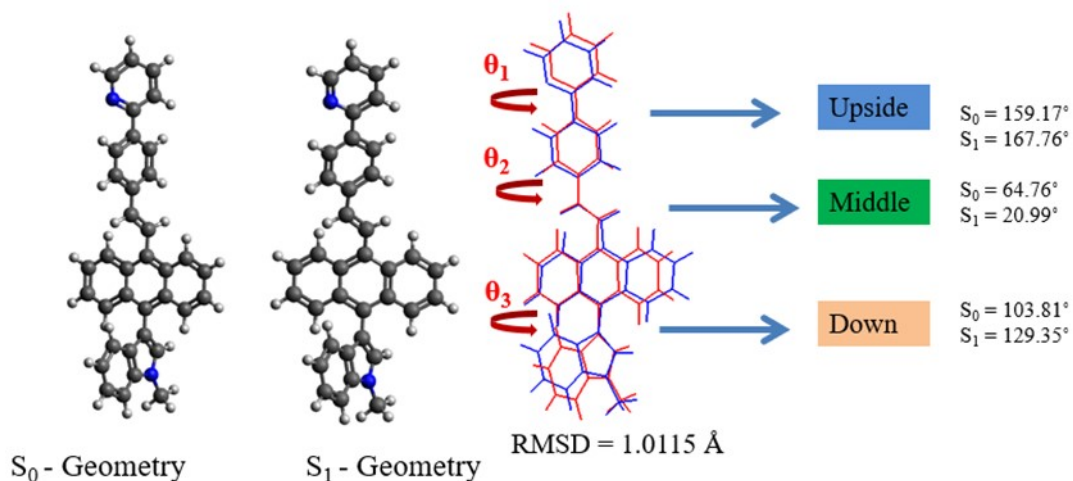
| CAM-B3LYP/6-31G*    |                    |                   | PBE0/6-31G*         |                   |                   |
|---------------------|--------------------|-------------------|---------------------|-------------------|-------------------|
| C 1.07347291682372  | 3.88433021727958   | 0.42451449178687  | C 1.09675809495630  | 3.94076758973123  | 0.29174089804622  |
| C 2.49350070384042  | 3.83698551135463   | 0.47306915401305  | C 2.51291162068535  | 3.88593439739794  | 0.34858260806975  |
| C 3.14904229923654  | 2.67540096307094   | 0.19775084156227  | C 3.16306268548559  | 2.70543882965744  | 0.11611257177854  |
| C 2.44190210324303  | 1.48111278548665   | -0.15655914231186 | C 2.45048908230953  | 1.50850662508112  | -0.20244795149290 |
| C 1.00872589136294  | 1.52924306062491   | -0.20981696836379 | C 1.01294700008879  | 1.56331464105161  | -0.26148099643988 |
| C 0.36126925420874  | 2.76829466529378   | 0.10308607963833  | C 0.37522784686478  | 2.81322945416165  | 0.00807829957909  |
| C 3.11892637761039  | 0.27478190422099   | -0.42326876480818 | C 3.12348640985442  | 0.28589168281792  | -0.41937250371791 |
| C 2.37883399020110  | -0.87660069649140  | -0.75347808449077 | C 2.37528956815091  | -0.87417791292036 | -0.71582295623856 |
| C 0.94301439881026  | -0.82903571249970  | -0.82218321425391 | C 0.93588869441062  | -0.82140794403248 | -0.79094590699058 |
| C 0.26533236431603  | 0.37371880750948   | -0.53136035353027 | C 0.25953990333077  | 0.39643448982317  | -0.53718972053350 |
| C 3.03575393596665  | -2.11432730185689  | -1.05568372677117 | C 3.02738481107403  | -2.11727033904051 | -0.98658849558119 |
| C 2.33539278005498  | -3.22490584291252  | -1.41376679096456 | C 2.32149563800939  | -3.23829108141187 | -1.32324222885219 |
| C 0.91784894908563  | -3.17084747905852  | -1.51434851167674 | C 0.90838394969629  | -3.17904057943907 | -1.43489334929189 |
| C 0.2522297136738   | -2.01695881072710  | -1.23268018736188 | C 0.24429741366029  | -2.01045383499373 | -1.18194836491111 |
| C -1.21000952835334 | 0.47171008454059   | -0.58266508400646 | C -1.20713163101175 | 0.49733405193351  | -0.57844379178990 |
| C -2.05668883067337 | -0.30400573456424  | 0.10621356585420  | C -2.06074815724340 | -0.30732569385619 | 0.08338843151809  |
| C -3.52464122775007 | -0.22876990208648  | 0.08812769452386  | C -3.52090584351502 | -0.22901920490326 | 0.07189911326763  |
| C 4.59792956498018  | 0.20626702182805   | -0.34265848739182 | C 4.59605905769347  | 0.21095042790740  | -0.32717872530006 |
| C 5.38136707453010  | -0.39941976507822  | 0.70515895519634  | C 5.37209591469378  | -0.43890017902373 | 0.69692361364202  |
| C 6.74115793990797  | -0.22801349369138  | 0.35568019354544  | C 6.73847794313934  | -0.24408693219780 | 0.36698739391262  |
| N 6.78893182080616  | 0.44667190805800   | -0.84257172160215 | N 6.79170533582546  | 0.48335995907054  | -0.79739599120849 |
| C 5.49988795674845  | 0.69978006353582   | -1.24758361963616 | C 5.50937868689622  | 0.75201905993750  | -1.20248474319621 |
| C 5.06164170436113  | -1.06375005647281  | 1.89810478086589  | C 5.04744990275500  | -1.15502871339813 | 1.85936273964629  |
| C 6.08758004091065  | -1.53427578359152  | 2.69837329751746  | C 6.07342825420314  | -1.65313682779107 | 2.64825341925485  |
| C 7.43590166049439  | -1.35388470737124  | 2.33262895652473  | C 7.42381160015771  | -1.44920366407882 | 2.30234570610782  |
| C 7.78147449171555  | -0.70123086381787  | 1.16165958299959  | C 7.77587616850363  | -0.74361766943563 | 1.16023971362773  |
| C 7.99382077677895  | 0.80578116349947   | -1.56055746906685 | C 7.99929440908851  | 0.87878119368254  | -1.48384004820099 |
| C -4.24571604198201 | 0.56849285979158   | -0.81068839449402 | C -4.24930788123358 | 0.61082774784401  | -0.78777390546940 |
| C -5.63054093329171 | 0.60924922710488   | -0.77691471357566 | C -5.63454034517169 | 0.64950920651695  | -0.74608737358999 |
| C -6.35447179714612 | -0.14784319854742  | 0.15246685717531  | C -6.35781714367768 | -0.15184690692216 | 0.15155407319908  |
| C -5.63879217698574 | -0.95651392452219  | 1.03953039938692  | C -5.63578787686443 | -1.00275941632865 | 0.99844995976041  |
| C -4.25307205272181 | -0.99401992468059  | 1.00712762174553  | C -4.24962978409465 | -1.03848995906042 | 0.95880153535980  |
| H 0.5575777715496   | 4.81087715830113   | 0.65676500171717  | H 0.58366395313587  | 4.87795479868118  | 0.49038223669266  |
| H 3.05237932146487  | 4.72816777770488   | 0.74149178042555  | H 3.07886622221516  | 4.781166919911673 | 0.59203617372346  |
| H 4.23024860396985  | 2.63482246186236   | 0.24791024987191  | H 4.24505016294826  | 2.65738329478869  | 0.17885216921260  |
| H -0.72154710021132 | 2.81018573083889   | 0.09737207270248  | H -0.70865005523285 | 2.86484110902271  | -0.00394959033906 |
| H 4.11702774519867  | -2.14758943568299  | -1.00052547666944 | H 4.11034631564260  | -2.15163238688717 | -0.93110228086562 |
| H 2.85675363994718  | -4.14974926617911  | -1.64059576884985 | H 2.84216756650519  | -4.16977142610403 | -1.52885923772714 |
| H 0.36709636237017  | -0.405182614877441 | -1.82937348768184 | H 0.35315861597625  | -4.06226272675012 | -1.73945754486917 |
| H -0.82434116780475 | -1.98135239052495  | -1.33402156883678 | H -0.83222811064481 | -1.97121758176665 | -1.30071191003321 |
| H -1.61043306743673 | 1.26791180304675   | -1.20712953660168 | H -1.61096327510409 | 1.32242919535697  | -1.16448604802990 |
| H -1.64643317594969 | -1.06355880770309  | 0.76934192808829  | H -1.64887112717733 | -1.08926931735189 | 0.72121062637664  |
| H 5.32050061190783  | 1.22347262113492   | -2.17722306532673 | H 5.33725706543588  | 1.31541831140455  | -2.11105921112200 |
| H 4.02368090191730  | -1.20578449215382  | 2.18493998298803  | H 4.00748886737632  | -1.31659146796556 | 2.13219207366437  |
| H 5.85373480682168  | -2.05122754827525  | 3.62411633186790  | H 5.83497019265312  | -2.21046669544770 | 3.55040549600823  |
| H 8.21878959213242  | -1.73434987880844  | 2.98156361640066  | H 8.20498887338734  | -1.85174892748332 | 2.94171975360326  |
| H 8.82080530786655  | -0.56248636689111  | 0.88128421424003  | H 8.81780299367327  | -0.58564973737287 | 0.89528720982183  |
| H 8.62572440923082  | 1.46128901322413   | -0.95419108735056 | H 8.62254601130210  | 1.50695421462112  | -0.83872257212265 |
| H 7.71520990950545  | 1.32279491947581   | -2.47359959244135 | H 7.72835691957044  | 1.44800762534838  | -2.37549898074541 |
| H -6.18349768812007 | -1.55637140368816  | 1.75906703838410  | H -6.17861513463664 | -1.63629549488360 | 1.69250808130007  |



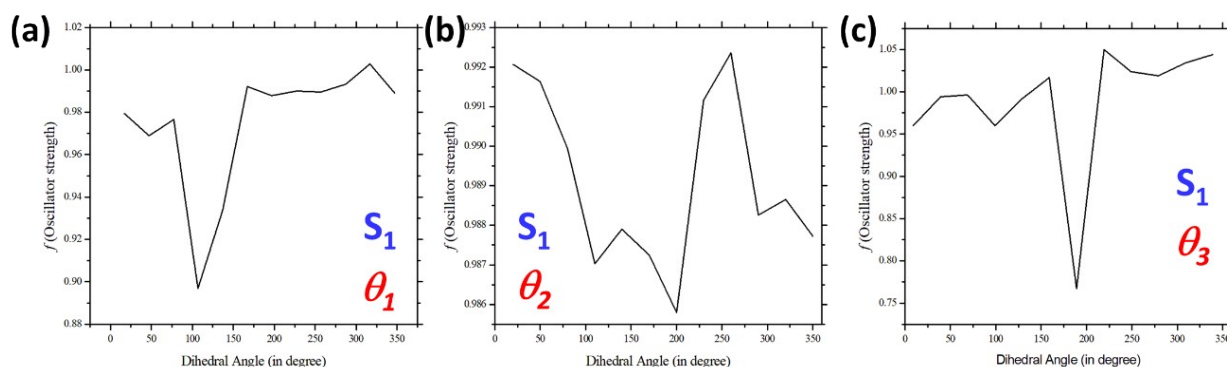


**Table S8** DSE properties of **IAPY**

| Compound   | Solvent           | ET <sub>30</sub> | $\lambda_{\text{abs.max}}$ (nm) | $\lambda_{\text{em.max}}$ (nm) | Stokes shift (cm <sup>-1</sup> ) | $\Phi_f$ (%) |
|--|-------------------|------------------|---------------------------------|--------------------------------|----------------------------------|--------------|
| <b>IAPY</b><br>with<br>$\lambda_{\text{em.max}}$ (Solid)<br>= 517 nm | Hexane            | 31.0             | 404                             | 515                            | 90090.09                         | 20.22        |
|  | 1,4-dioxane       | 36.0             | 409                             | 518                            | 91743.119                        | 17.73        |
|  | EtOAc             | 38.1             | 407                             | 522                            | 86956.521                        | 14.54        |
|  | CHCl <sub>3</sub> | 39.1             | 411                             | 521                            | 90909.09                         | 13.33        |
|  | DCM               | 40.7             | 412                             | 514                            | 98039.216                        | 12.79        |
|  | N,N-DMF           | 43.2             | 413                             | 526                            | 88495.575                        | 9.79         |



**Fig. S24** Geometry comparison between Ground state ( $S_0$ ) geometry and  $S_1$ -state optimized geometry IAPY in DMF using CAM-B3LYP/6-31G\* level of theory.



**Fig. S25** Dihedral scanning for emission oscillator strength at different twisted sites of **IAPY**

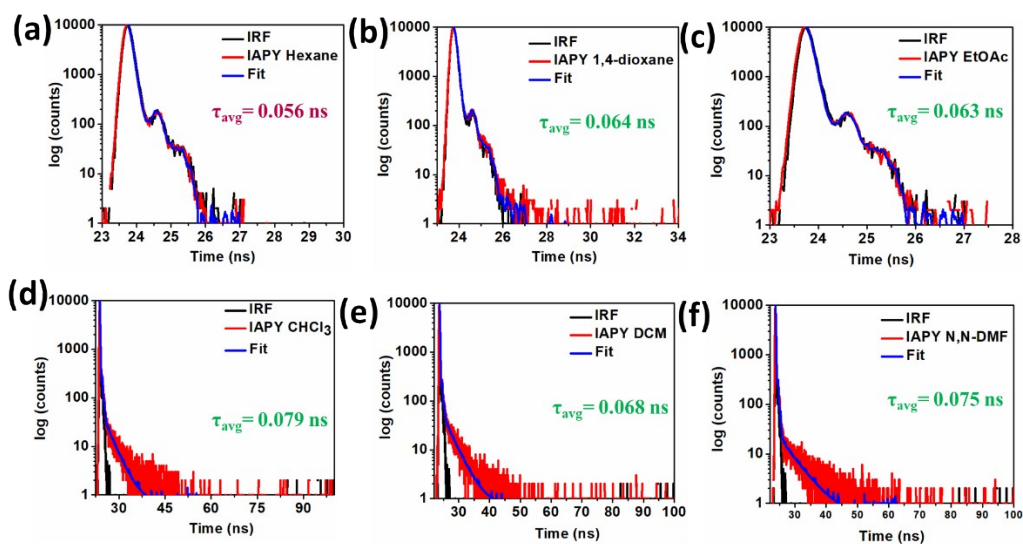


Fig. S26 Decay profiles of IAPY in different solvents

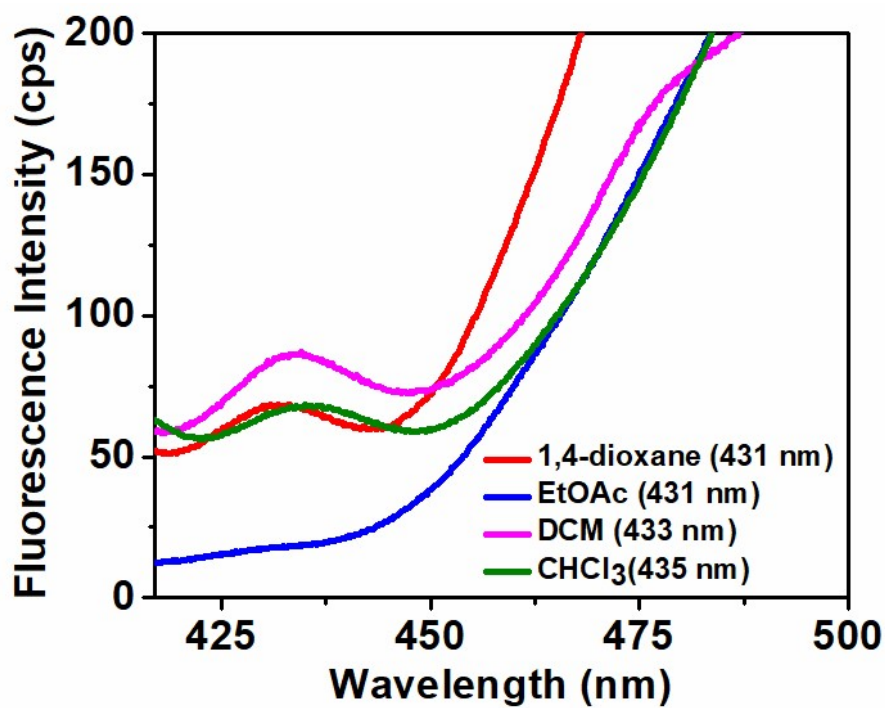


Fig. S27 LE state emissions of IAPY in different solvents

## CIE chromaticiy diagram 1931

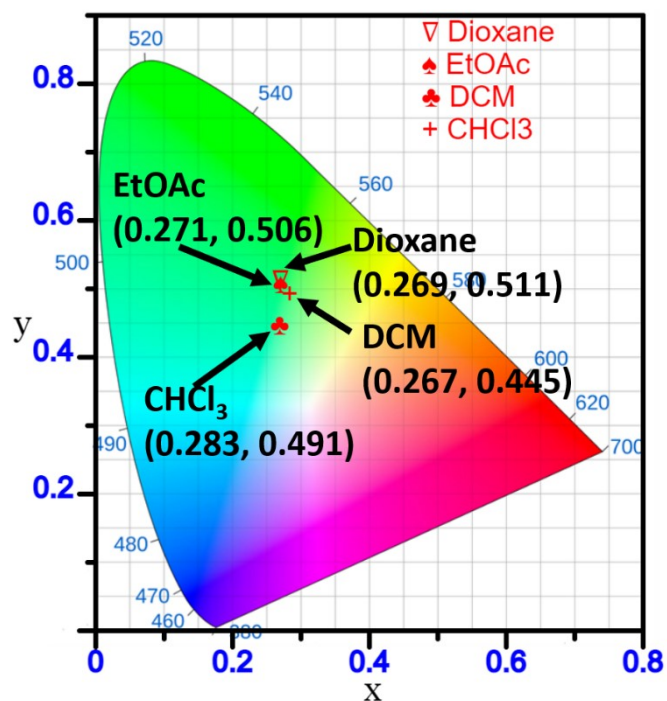


Fig. S28 CIE-coordinates of whitish emission of IAPY

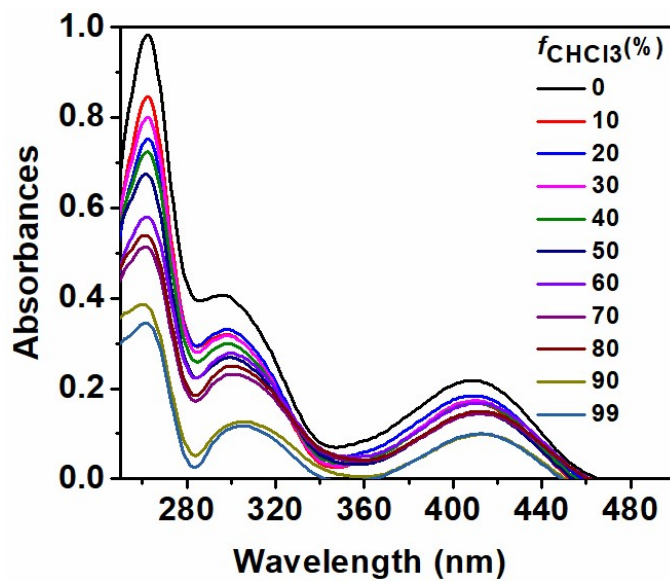
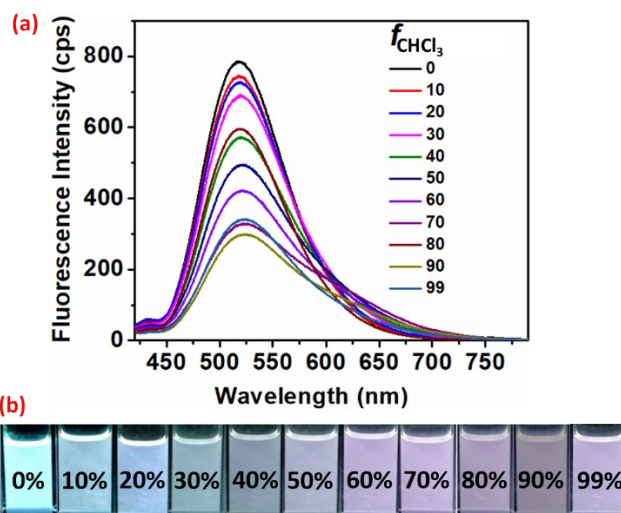
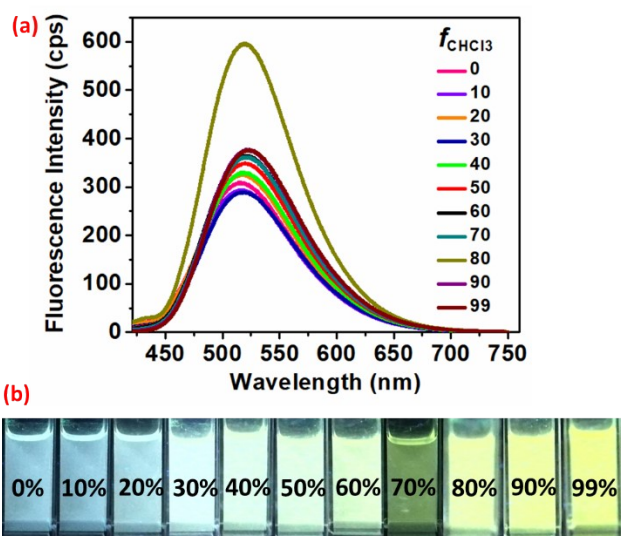


Fig. S29a Absorbances for whitish emission for IAPY in the binary mixture of 1,4-dioxane and CHCl<sub>3</sub>.



**Fig. S29b** Whitish emission for IAPY in the binary mixture of 1,4-dioxane and  $\text{CHCl}_3$ . (The percentage of  $\text{CHCl}_3$  has been increased gradually, and the excitation wavelength was in the range of 409-416 nm). The CIE coordinates for 0% to 99% are as: **0% (0.27, 0.52), 10% (0.27, 0.52), 20% (0.28, 0.58), 30% (0.29, 0.52), 40% (0.30, 0.52), 50% (0.32, 0.51), 60% (0.32, 0.50), 70% (0.34, 0.50), 80% (0.39, 0.46), 90% (0.34, 0.51), 99% (0.31, 0.52).**



**Fig. S30** Whitish emission for IAPY in the binary mixture of EtOAc and  $\text{CHCl}_3$ . (The percentage of  $\text{CHCl}_3$  has been increased gradually, and the excitation wavelength was in the range of 411-418 nm).

**Table S9** Results of times-resolved studies, altogether

| Compound/Solvent          | $\chi^2$  | $\lambda_{em}$<br>max<br>(nm) | $\tau_1$   | $\tau_2$   | $\tau_3$  | $\tau_4$ | $\alpha_1$ | $\alpha_2$ | $\alpha_3$ | $\alpha_4$ | $\tau_{avg}$<br>(ns) | $\Phi_f$<br>(%) | $k_f(s^{-1})$<br>$\times 10^{-6}$ | $k_{nr}(s^{-1}) \times 10^{-6}$ |
|---------------------------|-----------|-------------------------------|------------|------------|-----------|----------|------------|------------|------------|------------|----------------------|-----------------|-----------------------------------|---------------------------------|
| IAPY<br>Pristine          | 1.01<br>2 | 517                           | 0.606<br>8 | 2.960<br>7 | -         | -        | 0.903<br>2 | 0.096<br>7 | -          | -          | 0.834<br>4           | 26.6<br>4       | 319.27<br>1                       | 879.194                         |
| IAPY<br>Ground            | 1.06<br>7 | 534                           | 0.498<br>3 | 2.110<br>0 |           |          | 0.907<br>9 | 0.092<br>0 |            |            | 0.646<br>7           | 53.3<br>2       | 824.49<br>3                       | 721.818                         |
| IAPY H                    | 1.00<br>0 | 530                           | 0.600<br>4 | 1.410<br>0 | 3.44<br>9 |          | 0.626<br>9 | 0.318<br>9 | 0.053<br>1 |            | 1.023<br>7           | 1.16            | 11.331                            | 965.517                         |
| IAPY<br>base<br>fumed     | 1.05<br>8 | 539                           | 0.518<br>6 | 2.101<br>6 |           |          | 0.909<br>8 | 0.090<br>8 |            |            | 0.580<br>6           | 5.89            | 101.44<br>7                       | 1620.909                        |
| IAPY<br>Hexane            | 1.03<br>7 | 515                           | 0.055<br>9 | 0.596<br>2 |           |          | 0.490<br>7 | 0.509<br>3 |            |            | 0.055<br>6           | 20.2<br>2       | 3636.6<br>9                       | 14348.92                        |
| IAPY 1,4-<br>dioxane      | 1.06<br>9 | 518                           | 0.064<br>5 | 0.063<br>4 |           |          | 0.484<br>2 | 0.515<br>7 |            |            | 0.064                | 17.7<br>3       | 2770.3<br>12                      | 12854.68<br>8                   |
| IAPY<br>EtOAc             | 1.02<br>3 | 522                           | 0.062<br>9 | 0.062<br>5 |           |          | 0.494<br>9 | 0.505<br>1 |            |            | 0.063                | 14.5<br>4       | 2307.9<br>36                      | 13565.07<br>9                   |
| IAPY<br>DCM               | 1.07<br>1 | 514                           | 3.177      | 0.064      |           |          | 0.001<br>2 | 0.998<br>8 |            |            | 0.068                | 13.3<br>3       | 1960.2<br>94                      | 12745.58<br>8                   |
| IAPY<br>CHCl <sub>3</sub> | 1.00<br>6 | 521                           | 2.943<br>3 | 0.075<br>9 |           |          | 0.001<br>1 | 0.998<br>8 |            |            | 0.079                | 12.7<br>9       | 1618.9<br>87                      | 11039.24<br>1                   |
| IAPY<br>N,N-DMF           | 1.01<br>9 | 526                           | 0.071<br>9 | 5.098<br>8 |           |          | 0.999<br>5 | 0.000<br>5 |            |            | 0.074                | 9.79            | 1322.9<br>72                      | 12190.54<br>05                  |

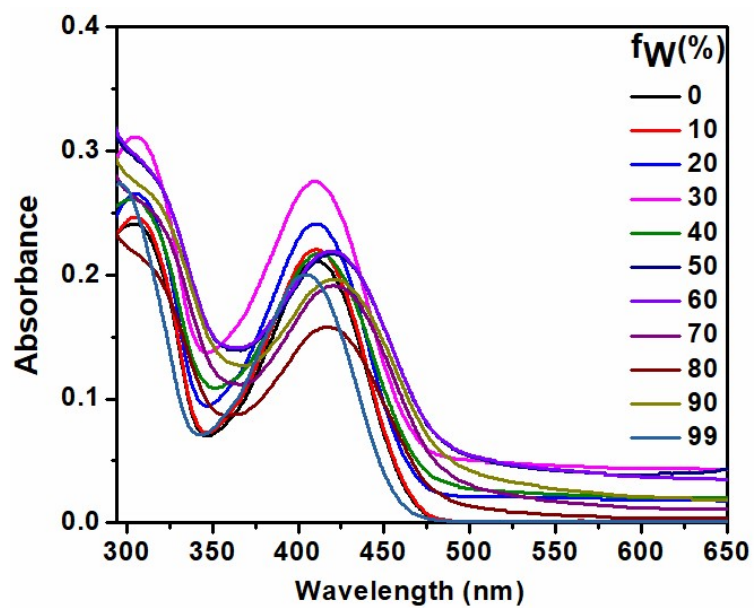


Fig. S31 Absorbances for AIE-study of IAPY

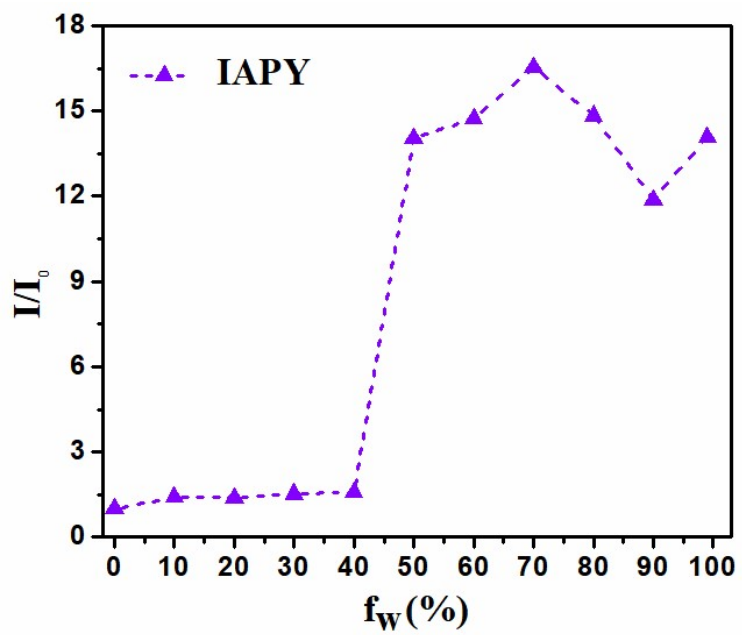


Fig. S32 Intensity of emission increment for AIE at  $10^{-5}$ M concentration of IAPY

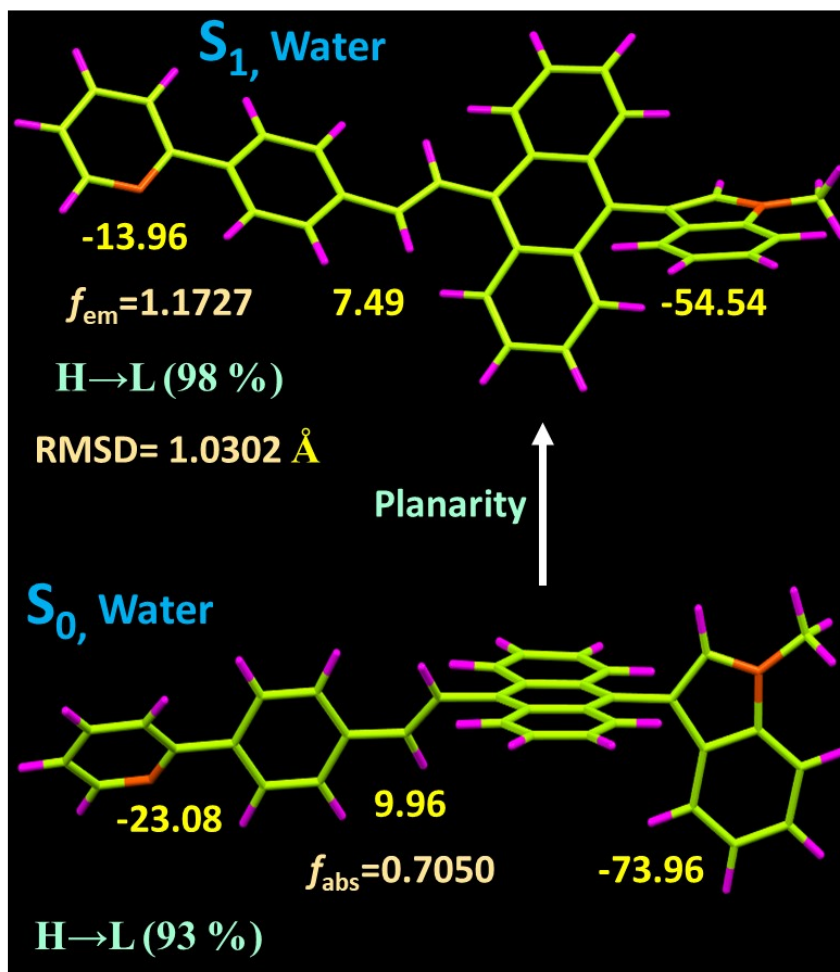
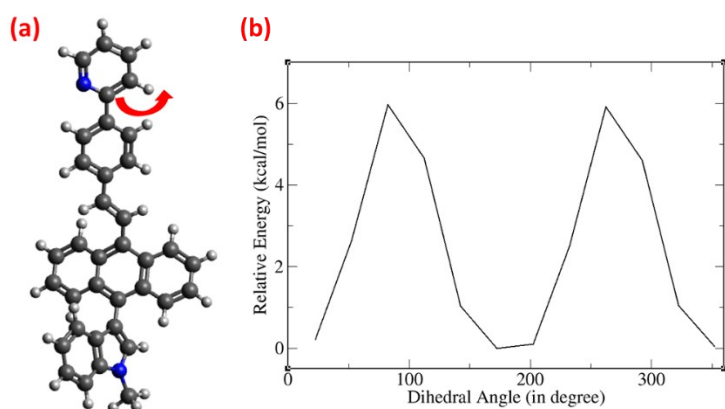


Fig. S33 TD-DFT optimized structure of IAPY in water



The rotation barrier is very small ~6 kcal/mol. That's free rotation takes place and excited state energy can decay non radiatively.

Fig. S34a Rotation barrier of the Pyridine group in S<sub>1</sub> state



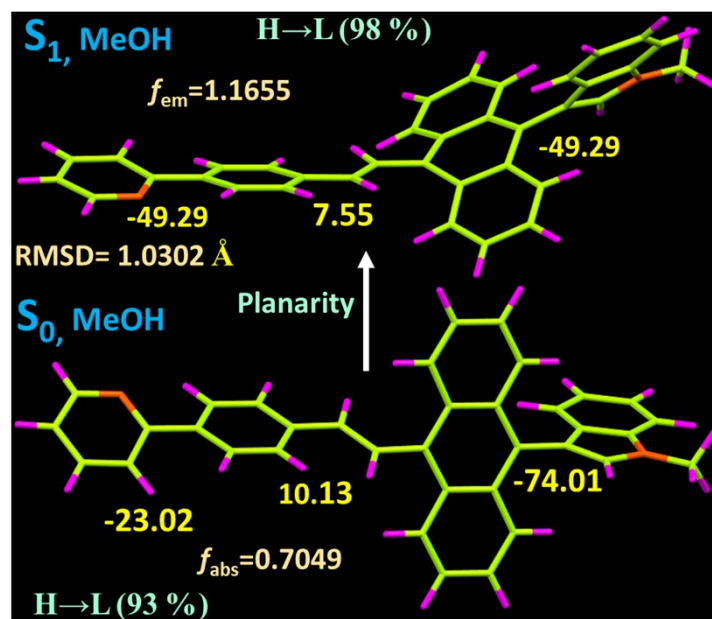


Fig. S34b TD-DFT optimized structure of IAPY in MeOH

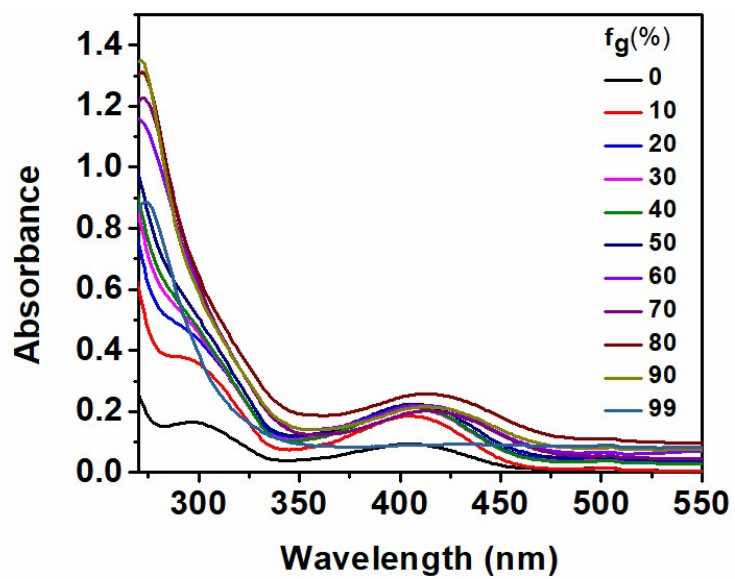
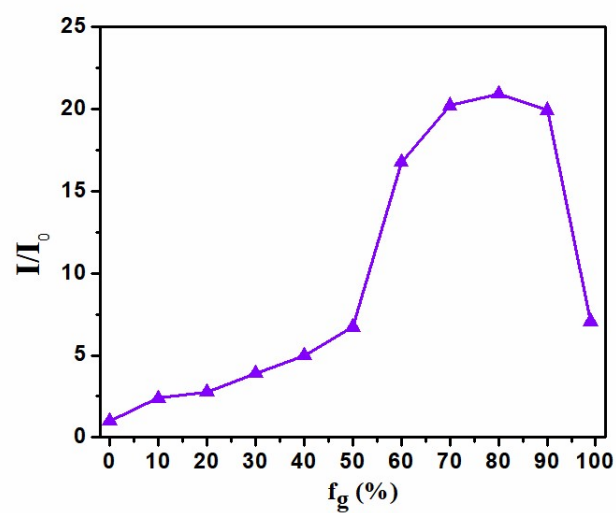
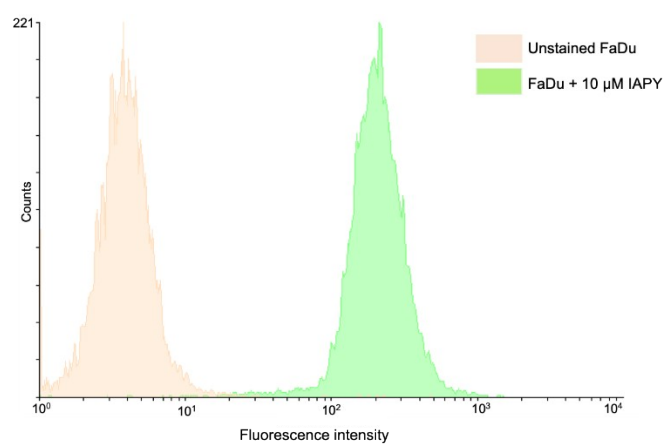


Fig. S35 Absorbances for viscofluorochromism of IAPY

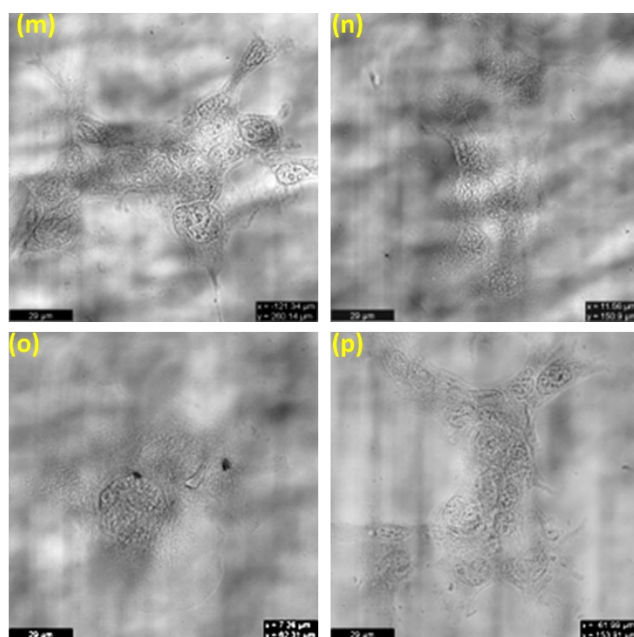




**Fig. S36** Intensity of emission increment for VIE at  $10^{-5}$ M concentration of **IAPY**



**Fig. S37** Cellular uptake of the probe **IAPY** through Fluorescence Activated Cell Sorting (FACS)



**Fig. S38** Bright field images for wash-free bioimaging of FaDu with **IAPY** (10  $\mu$ M)

### Cell Culture and confocal microscopy

The cell line was bought from ATCC and preserved in DMEM supplemented with 10% FBS with 1% (v/v) penicillin-streptomycin antibiotics. Cells were incubated at 37°C in a humidified atmosphere comprising 5% CO<sub>2</sub>. All the culture supplies were procured from Thermo Fisher Scientific.

Cell line was seeded on a lysine-coated sterile coverslip placed on a 12-well cell culture plate (1 × 10<sup>5</sup> cells per well) in DMEM containing 10% FBS with 1% antibiotics. Cells were allowed to adhere completely. After 24 h, cells were treated with formulation using 10 $\mu$ M probe concentration for 4 h. Later, cells were not washed with sterile PBS and mounted on a glass slide. Cell imaging was done using a confocal microscope (Leica LAS X).

### FACS

FaDu cells were seeded in a 12-well plate at a density of 0.2 million cells per well. The cells were treated with IAPY at 10  $\mu$ M concentration while the unstained control cells were treated with DMSO. After 24 hours of treatment, the medium was removed and cells were washed with PBS and trypsinized to obtain a single cell suspension. The cells in trypsin were centrifuged at 2000 rpm for 10 minutes at 4 degrees and resuspended in PBS for uptake analysis using BD FACS Aria II flow cytometer.

### Device fabrication, formulation with dye for screen printing and the procedure for screen printing

A market-available screen-printing liquid (SPL) white non-fluorescent dye of 4000 cP was purchased. Later 30 mg of IAPY dye was dissolved in 10 ml of DMSO and sonicated unless the solution becomes clear. Now this solution was added with 30 ml of the screen-printing liquid white non-fluorescent dye and mixed properly to prepare an **IAPY-SPL** media. Again, by using another dye **SB6** the **SB6-SPL** media was prepared in the same fashion. The QR code was drawn with CAD tool solid works, and on the PVC sheet the QR code was engraved with the universal laser system CO<sub>2</sub> laser with 10-micron wavelength with 10% into 30 watts. The **SB6-SPL** was first painted on a glass surface and dried subsequently. The surface emitted green under 365-UV lamp. Later on, the laser-cut QR code pattern was screen printed

with **IAPY-SPL** on the surface of **SB6-SPL**. With naked eyes the QR-code is not clearly visible under ambient light. Even under 365-UV lamp also the entire surface remains green. Once the acid fumigation is done the QR-code will be visible under 365-UV lamp as it glows yellow on a green surface.

### Procedure and method of data storage and anticounterfeiting applications with stimuli-responsive hidden stamp:

Three dyes- **IAPY**, **ATH4P**, and **SB6** were initially chosen as all of them are green emitters ( $\lambda_{em. max} = 501 \text{ nm} - 517 \text{ nm}$ ). An ethanolic solution of  $10^{-7} \text{ M}$  concentration was prepared of each dye and soaked with three different sponges. Now a wooden stamp was pressed on the **IAPY** solution-soaked sponge and placed on a TLC plate to get an **IAPY** led stamp. Later on, the wooden stamp was washed with acetone properly, and consecutively, two more **ATH4P** and **SB6** led stamps were prepared on the TLC plates. Now, none of the stamps are visible to the naked eye but under the UV-365 nm lamp, all the stamps glow green. Now how to identify which stamp is real and which ones are counterfeited? Let us say that the **IAPY** is the real dye; hence, the **IAPY** led stamp will be the real stamp. **IAPY** emits orange-red on acid-fumigation but immediately does not revert back to green on base-fumigation as it turns yellow. But this information is secret. The counterfeiters might mimic a dye (**SB6**) which is a green emitter but not acidofluorochromic or can mimic a dye (**ATH4P**) which can emit red on acid fuming and again revert back to green on base fuming. Because reversible acidofluorochromism is usually used in most of the cases. So, if an unknown stamp that looks alike the real one, is acid-fumed and still emits green; it means the dye is counterfeited and the stamp is unreal. Next, if an unknown stamp that looks alike again as the real one, is acid-fumed and turns red emitting then that stamp may or may not be real. But again, with base-fuming if it turns green emitting instantly, then the stamp (made of dye **ATH4P**) is fake with no doubt. But on further base-fuming if it turns yellowish then the stamp is the real stamp.

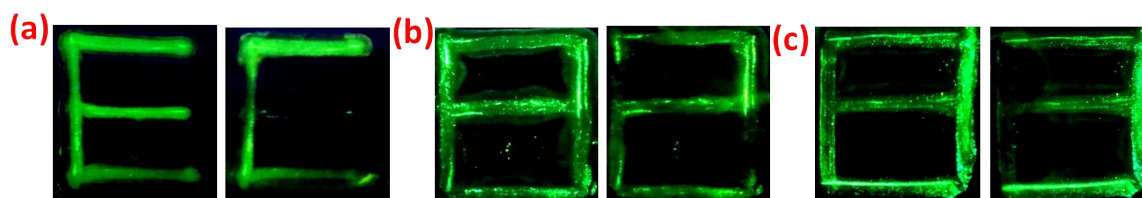


Fig. S39 Data encryption with IAPY

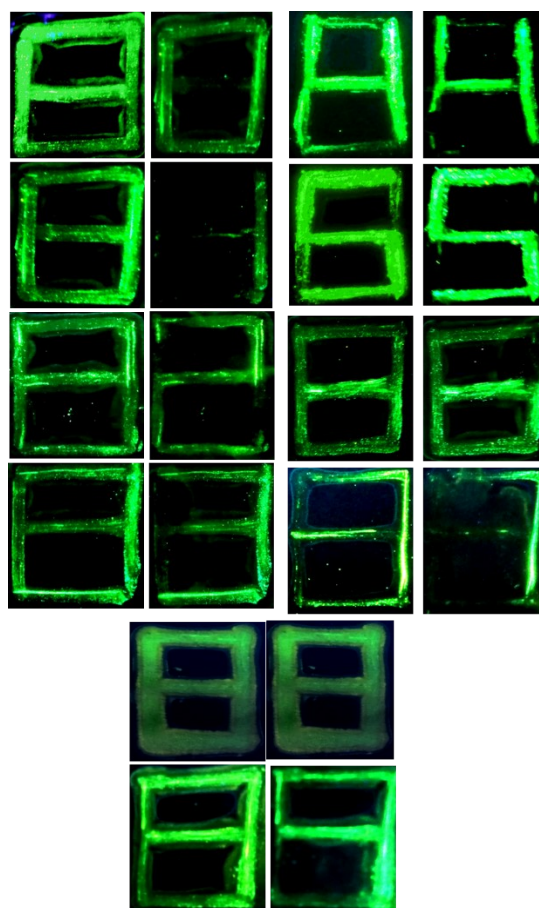


Fig. S40 Data encryption with IAPY to make different digits from '8'

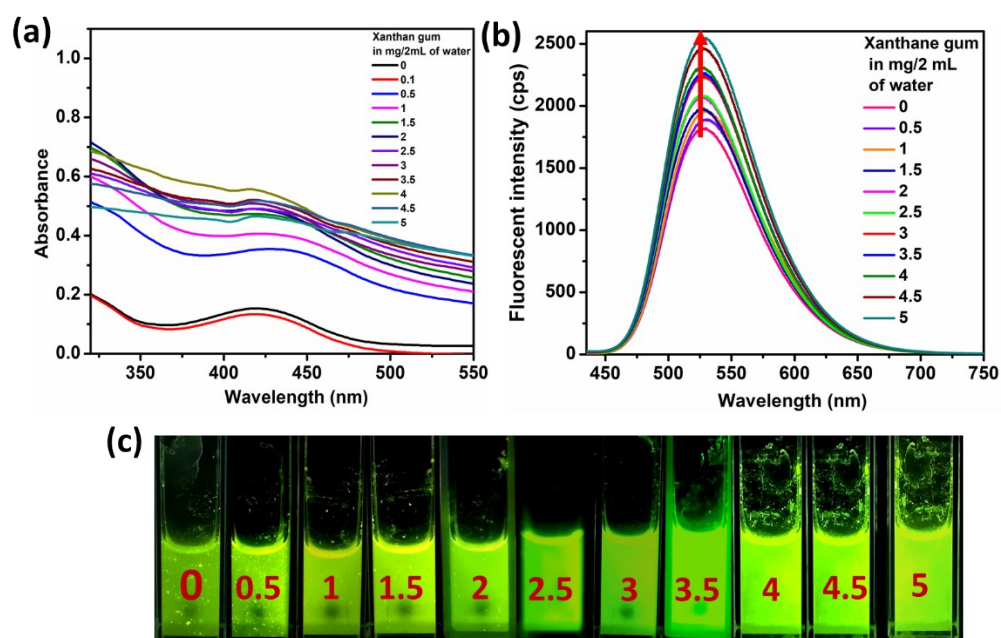
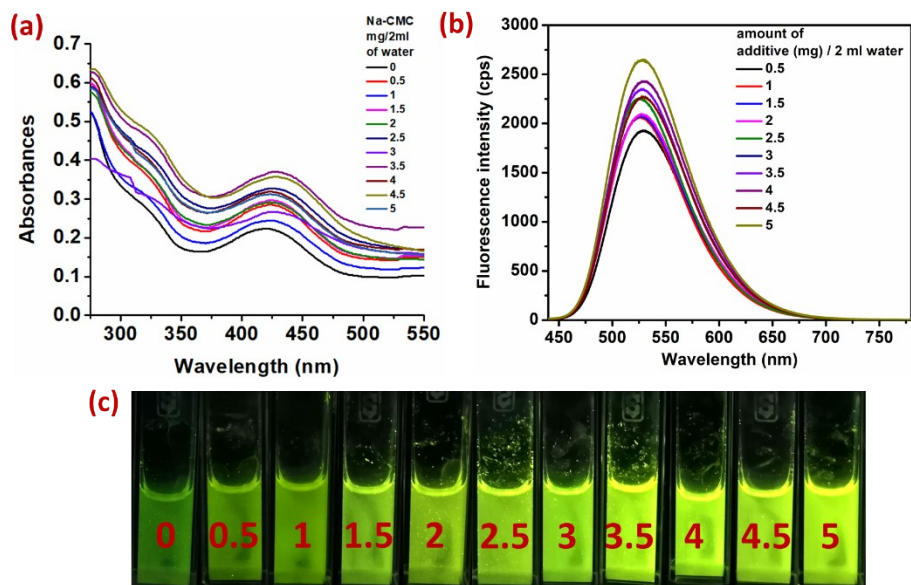
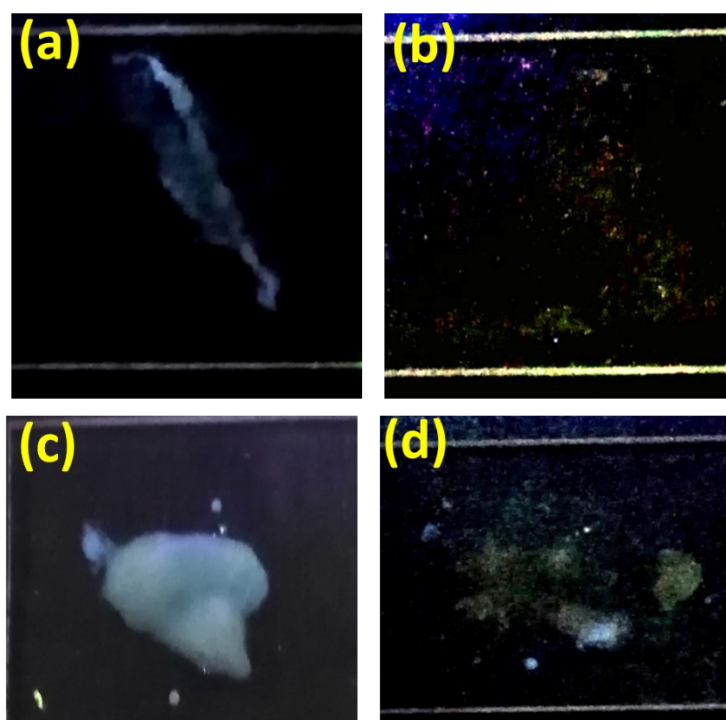


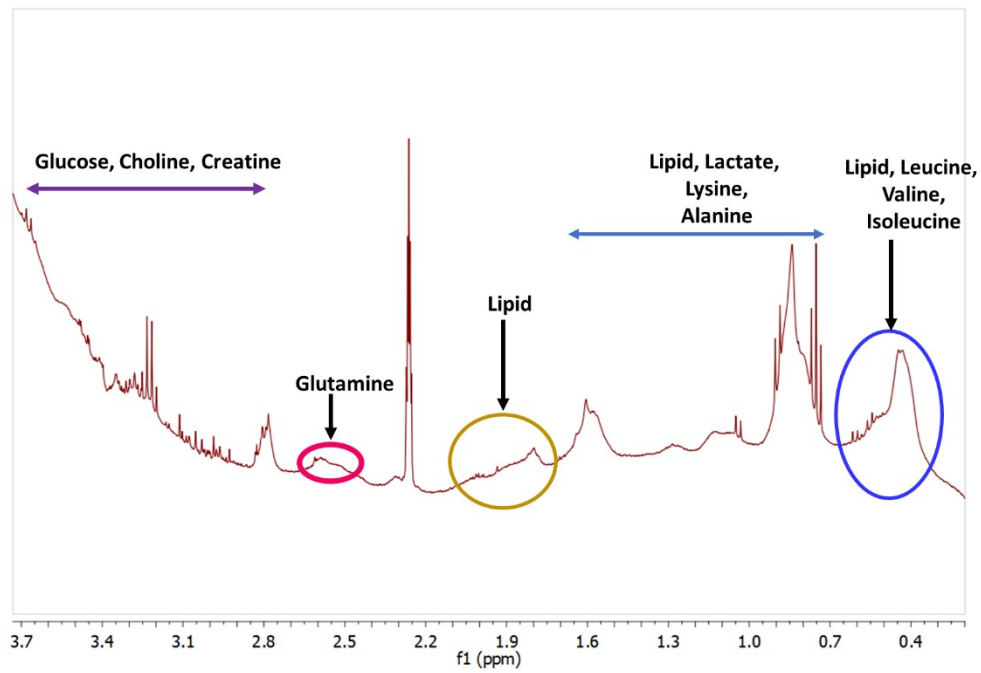
Fig. S41 Xanthan gum sensing (a) absorbances (b) emission (c) picture taken under UV-365 nm lamp (the mg of Xanthan gum taken per 2ml of water is written on each cuvette picture)



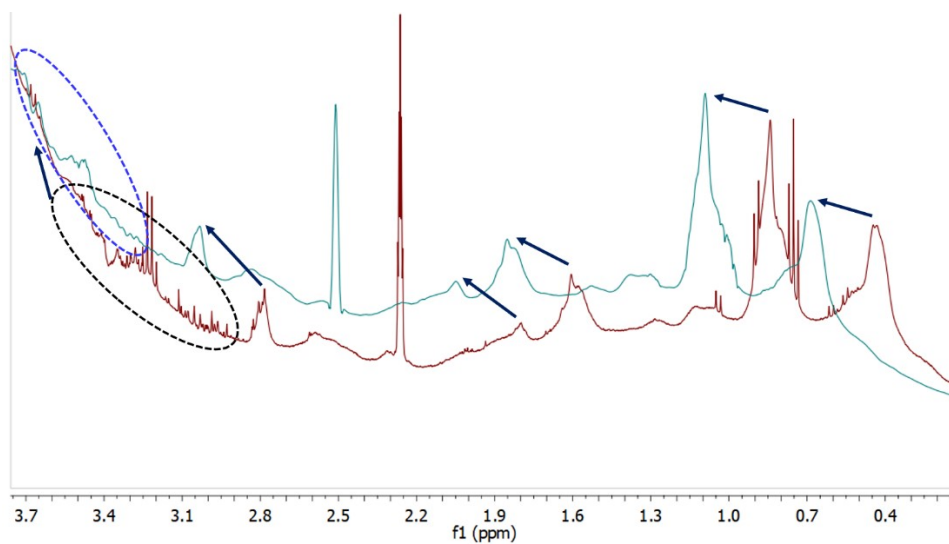
**Fig. S42** Na-CMC sensing (a) absorbances (b) emission (c) picture taken under UV-365 nm lamp (the mg of Na-CMC taken per 2ml of water is written on each cuvette picture)



**Fig. S43** False-positive taste under UV-365 nm lamp; (a) dried patches of cold drinks (b) almost no fluorescence after staining with IAPY (c) tomato sauce (d) feeble green fluorescence after staining with IAPY.

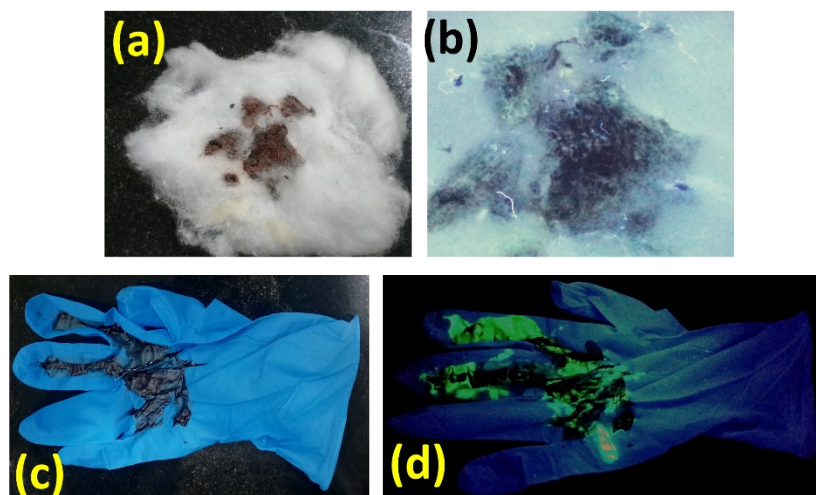


**Fig. S44a** Blood serum NMR

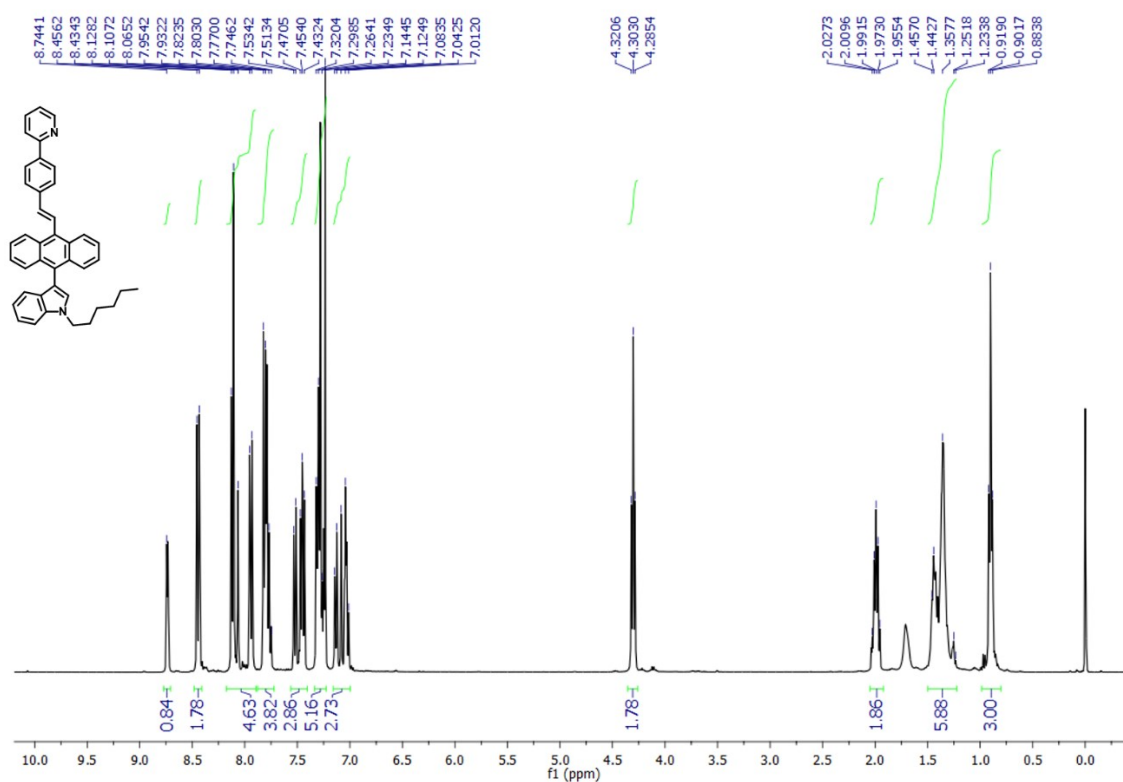


**Fig. S44b** Blood serum-NMR comparison (the maroon color line for **normal serum** and the sea-green color line for **IAPY+serum**). The peaks for **(IAPY+serum)** are de-shielded from that of **normal serum**.





**Fig. S45** Hospital-disposed of biohazards detection by fluorescence; under normal room light (a) blood-smeard cotton (c) blood containing glove; under UV-365 nm lamp (b) blood-smeard cotton (d) blood containing glove



**Fig. S46**  $^1\text{H}$  NMR of IAPY $^1$

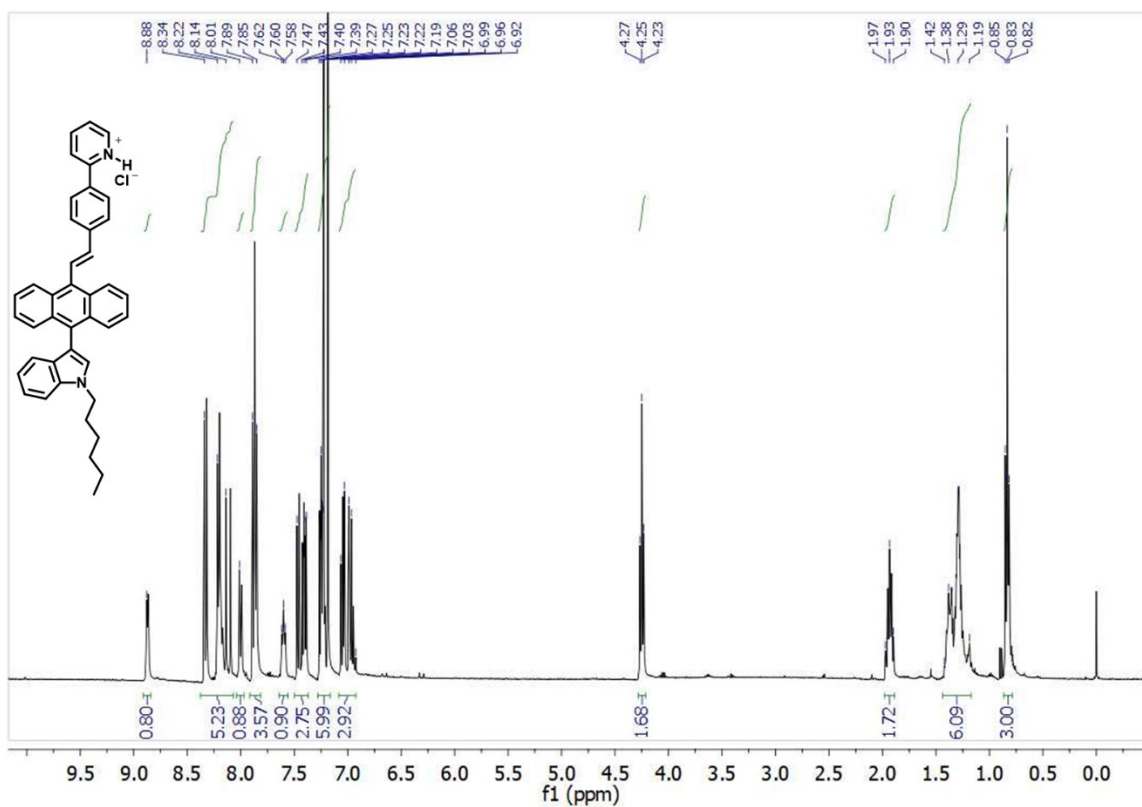


Fig. S47 <sup>1</sup>H NMR of IAPYH

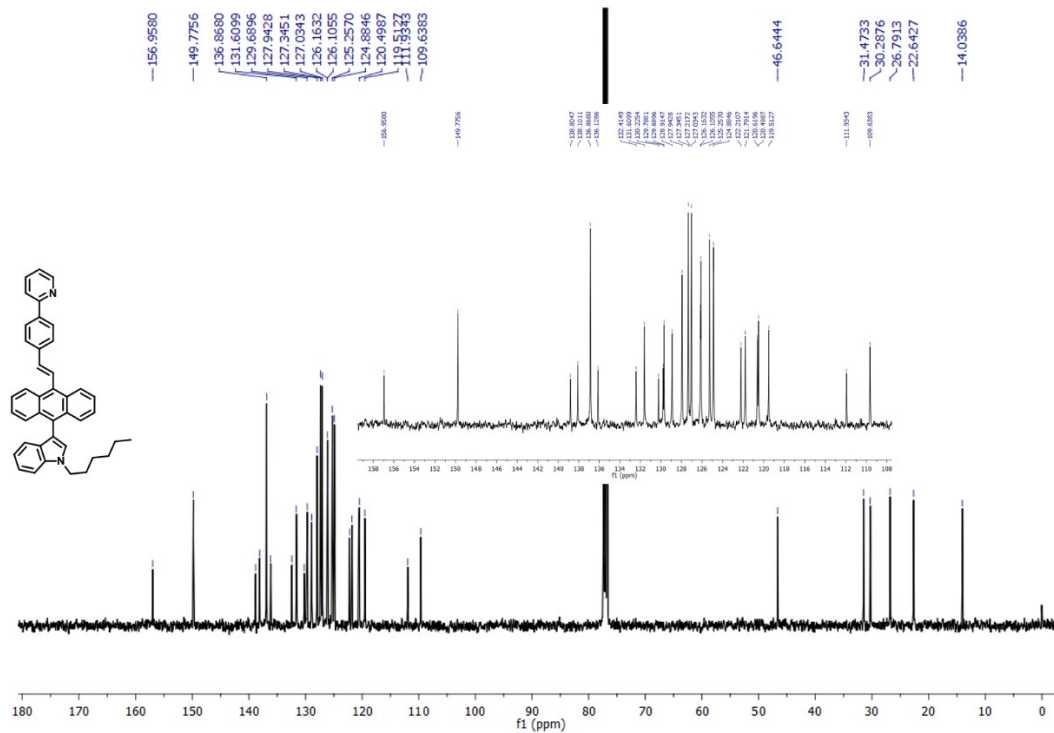


Fig. S48 <sup>13</sup>C NMR of IAPYH<sup>1</sup>



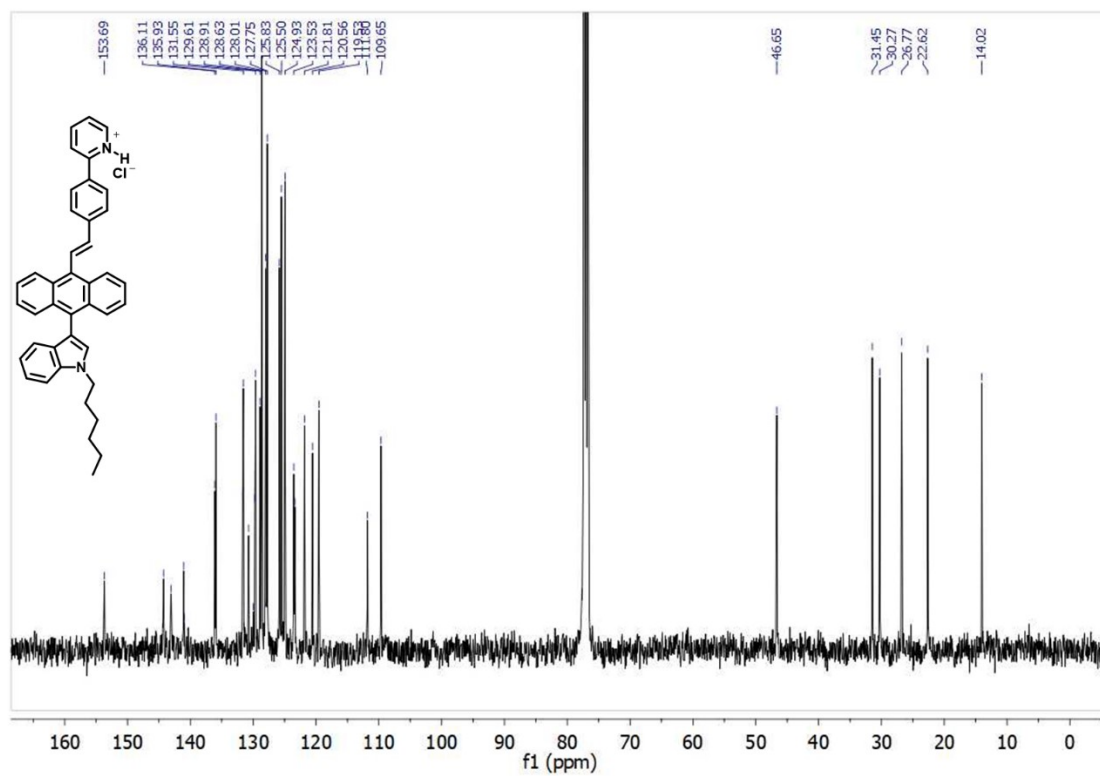


Fig. S49a <sup>13</sup>C NMR of IAPYH

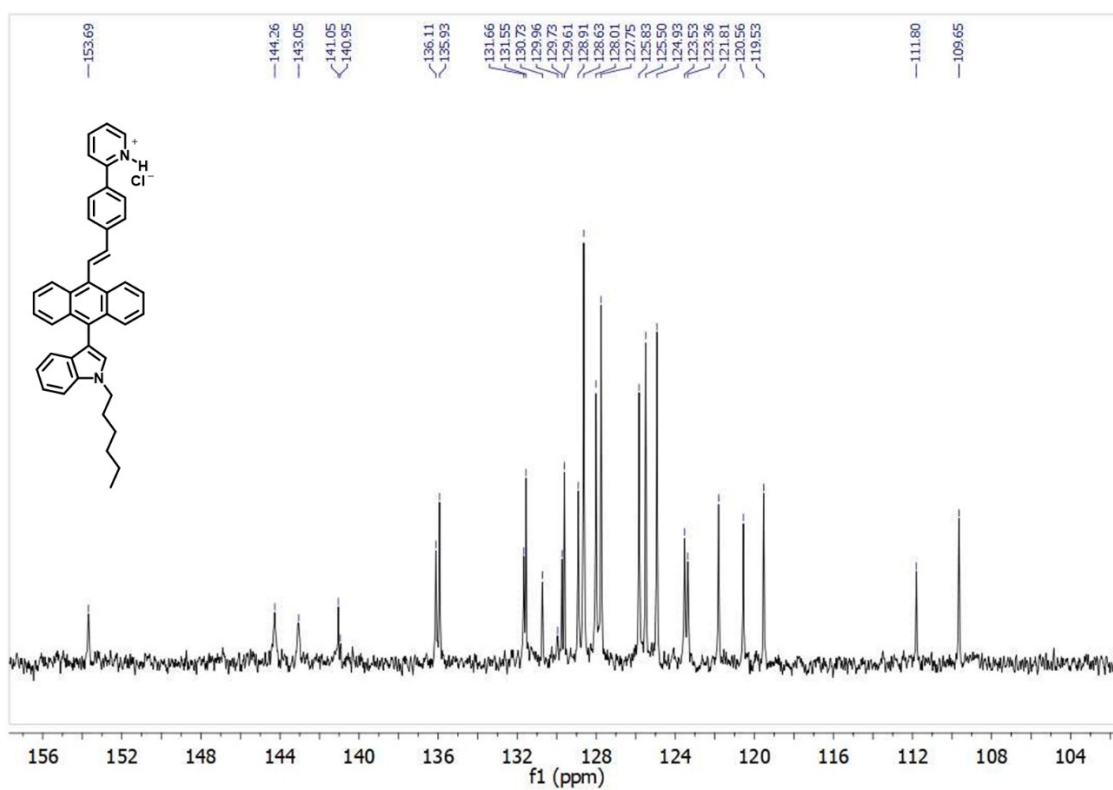


Fig. S49b <sup>13</sup>C NMR (aromatic part zoomed) of IAPYH

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END