Supplementary Information

A sandwich-type photoelectrochemical biosensor based on

$Ru(bpy)_3^{2+}$ sensitized In_2S_3 for a flatoxin B_1 detection

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Materials and Reagents

Indium nitrate hydrate, aelenium dioxide (SeO₂), thioacetamide (C₂H₅NS), 3mercaptopropionic acid (MPA), tris(2,2'-bipyridine)dichlororuthenium(II) hexahydrate (Ru(bpy)₃Cl₂·6H₂O), sodium sulfide nonahydrate (Na₂S·9H₂O), zearalenone (ZEN), aflatoxin B₁ (AFB₁), potassium chloride (KCl), potassium ferricyanide (K₃Fe(CN)₆) and L-ascorbic acid (AA) were purchased from Macklin Biochemical Co., Ltd. (Shanghai, China). Sodium dihydrogen phosphate dihydrate (NaH₂PO₄·2H₂O) was gained from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). Sodium chloride (NaCl) was purchased from Damao Chemical Reagent Factory (Tianjin, China). Potassium ferrocyanide trihydrate (K₄Fe(CN)₆·3H₂O) was obtained from Tianjin Guangfu Fine Chemical Research Institute (Tianjin, China). Ochratoxin A (OTA) was obtained from Aladdin Chemistry Co., Ltd. (Shanghai, China). Nhydroxysuccinimide (NHS), 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC), bovine serum albumin (BSA), disodium phosphate (Na₂HPO₄), T-2 toxin (T-2) and AFB₁ antibody were purchased from Shanghai Sangon Biotech Ltd. Co. (Shanghai, China). Indium tin oxide (ITO) electrodes were provided by South China Science & Technology. The AFB₁ aptamer was synthesized and purified by Shanghai Sangon Biotech Ltd. Co. (Shanghai, China) with the sequence as follows:

AFB₁ aptamer: 5'-NH₂-C₆-GTT GGG CAC GTG TTG TCT CTC TGT GTC TCG TGC CCT TCG CTA GGC CC-3'

Apparatus

PEC performance measurement and electrochemical impedance spectroscopy (EIS) were carried out on a CHI 660D electrochemical workstation (CH Instruments, China). The electrolyte used in EIS was a 0.01 M phosphate buffer saline (PBS, pH 7.4) containing 0.1 M of KCl and 2 mM of K₃[Fe(CN)₆/K₄[Fe(CN)₆] (1:1), the frequency was ranged from 100 kHz to 0.01 Hz, and the peak-to-peak amplitude of the AC potential was 5 mV. A conventional three-electrode system consisting of a modified ITO working electrode, a Ag|AgCl (saturated KCl) reference electrode, and a platinum wire counter electrode was used. A full-band xenon lamp (Nanjing Yan'an Special Lighting Factory, China) was used as the light source. Scanning electron microscopy (SEM) was carried out using S-4800 microscope (Hitachi, Japan). Transmission electron microscopy (TEM) was carried out using JEM-2100 microscope (JEOL, Japan). X-ray photoelectron spectroscopy (XPS) was performed with an ES-CALAB 250 spectroscopy (Thermo Fisher Ltd, USA). Ultraviolet-visible (UV-vis) spectroscopy was conducted using a UV-3600 spectrometer (Shimadzu Co., Ltd., Japan).

PEC measurements

The photocurrent signal was measured in 0.1 M PBS (pH 7.4) containing 0.1 M AA at the applied potential of 0 V. Tris/EDTA buffer (containing 10 mM Tris-HCl and 1 mM EDTA, pH 7.8-8.2) was utilized to wash the electrode surface after each-step modification to remove any unbound materials. Then, the wet surface of the electrode was dried with nitrogen. The working area of ITO electrode was 0.45 cm². The light source was switched on once every 20 s, and the photocurrent was generated when the electrode surface was subjected to light energy.