Supporting Information for: Mass spectrometry reflects key aspects of copper-amyloid β chemistry

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Figure S1. CID spectrum of $[A\beta42+2H+Cu(II)]^{4+}$ at a collision potential of 32 V (normalised collision energy *ca.* 0.03 eV/Da), showing similar results as in (Everett et al. 2021).



Figure S2. (a) Full MS spectra of 10 μ M A β_{1-16} construct and 5 μ M CuSO₄. The metal-free (violet), Cubound (green), and double Cu-bound (red) A β_{1-16} are present as doubly, triply and quadruply charged ions. **(b)** Charge-deconvolution of the spectra in Panel (a). Deconvolution was carried out with UniDec (Marty et al. 2015; Reid et al. 2019), and peptides with zero, one, and two bound copper ions are indicated with a blue circle, green triangle, and red triangle, respectively. **(c)** Isotopic distribution of Cu-bound A β_{1-16} at charge state 3+. Simulated distributions assuming 100% Cu(II) are overlaid in orange.



Figure S3. (a) Isotope distribution of the b_{14} fragment at a collision voltage of 22 V (equivalent to a normalised collision energy of ca. 0.035 eV/Da) and the calculated percentage of Cu(II) (red dots show the best-fit simulation of the Cu(I)/Cu(II) ratio. **(b)** Energy-dependence of the relative intensity of a_{14} and b_{14} fragments after collisional activation of metal-free murine and human A β_{1-16} (dark and light green curves for a_{14} and b_{14} , respectively) and of Cu(II)-complexes of murine and human A β_{1-16} (dark and light blue curves for a_{14} and b_{14} , respectively). Note that no copper-free a_{14} or b_{14} fragments were detected in the spectra of the metal-free precursor. Error bars show the standard deviation of triplicate experiments, illustrating excellent reproducibility and consistency.



Figure S4. Energy-dependence of the relative intensity of the decarboxylated precursor after collisional activation of Cu(II)-bound murine and human A β_{1-16} .

Experimental Section

Full-length $A\beta_{1-42}$ was purchased from Bachem AG (Bubendorf, Switzerland). LC-MS grade water was obtained from Fisher Scientific (Schwerte, Germany), and other reagents, including custom peptides, from Merck (Darmstadt, Germany). MS measurements were performed with a Synapt XS instrument. Samples were loaded into glass emitters produced in-house with a Sutter P-97 Flaming/Brown micropipette puller and transferred into the gas phase with nano-electrospray ionisation in direct-infusion positive-ion mode. For full-length amyloid β and its copper complex, the 4+ charge state was isolated in the quadrupole of the instrument; for the smaller custom peptides, we selected the 3+ precursor. Collisional activation was performed in the Trap cell. In fragmentation experiments, we acquired data for 3 minutes per spectrum and averaged the fragment intensities over this time. We subsequently normalised this average intensity to the total ion current to allow comparison between experiments.

References

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