

## Supplementary Information

### **I-motif sensor-based fluorometric detection of Mpox using multiplex-LAMP**

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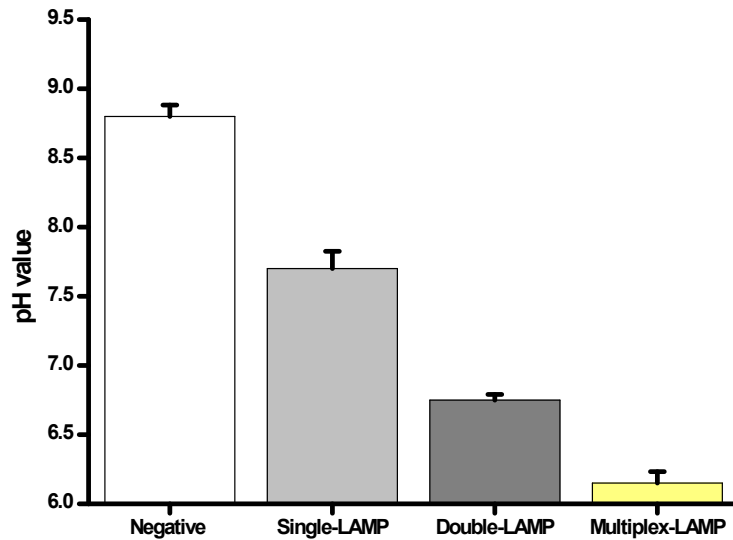
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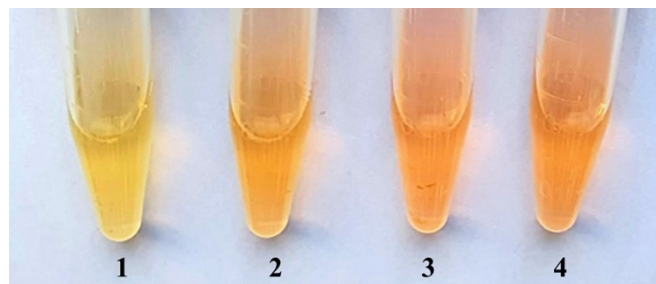
Target gene	LAMP Primers
<b>B6R (204 mers): Envelope protein gene</b> cgfttggaagaaaaaatgaaatactcttggaaatgatactgttacgtgtcctaagcggaa tgtcaatcttcaatgtagatcacggatcgtgtcaaccagttaaaggaaaatactcattggg gaacataactatcaactgtgatgttgatagaggattattggcttcgtacataactgtac agctaattcttgg	<b>FIP:</b> CGATCCGTGATCTAATTGAAGAGATCTTCTTGGAAATGATACTGTTACG <b>BIP:</b> AAGGAAAATACTCATTGGGGAACACGAAGCACCAATAACCTCAT <b>F3:</b> CGTTGTGAAGAAAAAATGGAA <b>B3:</b> CCAAGAATTAGCTGTACAAGTT
<b>N3R (231 mers): Complement binding protein gene</b> gtaagtagtcaatacagggaatagaatattattcatgtgattatacaacaaccgctctac aattaacaacattactttataacggcgaagaatactgaaattgatactgaaaaaagc cactaataaaaacagttgtaattactcaggcttagactacaaaaatggtcgtatagcga agattgtataattatctcagatctttagttagaagaatg	<b>FIP:</b> TCAATTTTCAAGTATATTCTTCGCCGTATTTCATGTGATTATACTAACAACCG <b>BIP:</b> AACAGTTGGTTAATTACTTCAGGCTAATTATACAATCTTCGCTATCGA <b>F3:</b> GTAAGTAGTCAATACGAGGAATT <b>B3:</b> CATTCTTCTAATAAAGATCTGAGA
<b>E9L (205 mers): DNA polymerase gene</b> aaacagtaatcgttaccacttaaacgatataacagtaactgtatgagattgggtaata aatacagaaggaaactcttatcgaagtgacactctatctagaataagtagatcttggg atatcgaatcaggtatttcttagcgaacagttacgtggatcgtcacaatgataacatccat tghtaatcttctg	<b>FIP:</b> ACTTCGATAAGAAGTTTCTTCTGTATCGATATAACAGTAACCTGTATGC <b>BIP:</b> AAGTACGATCTTGGGATATCGAATCTGTGACGATCCACGTAAC <b>F3:</b> AAACAGTAATCGTTTACCCTT <b>B3:</b> GACAAAAGATTAACAATGGATGTT
Target gene	RT-qPCR Primers
<b>B6R (92 mers): Envelope protein gene</b> cggatcgtgtcaaccagttaaaggaaaatactcattggggaacataataactcaactgtg atgttgatagaggattattgtgcttcgt	<b>Forward:</b> CGGATCGTGTCAACCAGTTA <b>Reverse:</b> ACGAAGCACCAATAACCTCATA
<b>N3R (116 mers): Complement binding protein gene</b> actaacaaccgctcacaattaacaacattctttataacggcgaagaatatactgaaatt gatagatcgaaaaagccactaataaaaacagttgtaattactcaggctt	<b>Forward:</b> ACTAACAACCGTCTACAATTAAC <b>Reverse:</b> AAGCTGAAGTAATTAACCAACTG
<b>E9L (141 mers): DNA polymerase gene</b> actgtatgagattgggtaataatacagaaggaaactcttatcgaagtgacactctat atctagaataagtagatcttggatcgaatcaggtatttcttagcgaacagttacgt ggatcgtcacaatg	<b>Forward:</b> ACTTGTATGCGAGATTGGGTTAATA <b>Reverse:</b> CATTGTGACGATCCACGTAAC
<b>I-motif sequences (red colored one "Rb" mainly used in our detection system)</b>	
<input type="checkbox"/> <b>H-telo:</b> TAACCCTAACCCCTAACCCCTAACCC <input checked="" type="checkbox"/> <b>Rb:</b> CCGCCCAAAACCCCC <input type="checkbox"/> <b>HIF-1<math>\alpha</math>:</b> CCCGCCCTCTCCCA <input type="checkbox"/> <b>c-MYC:</b> CCCACCTTCCCCACCCCTCCCCACCCCTCCCC	

**Table S1: Oligonucleotides used in this study.**

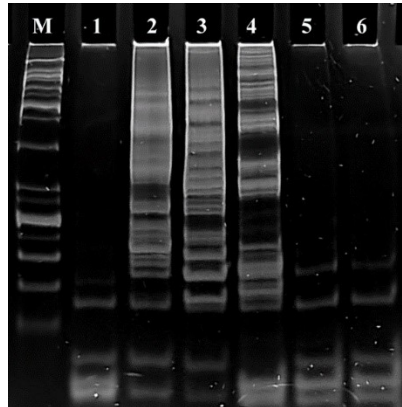
## 1. Supportive data:



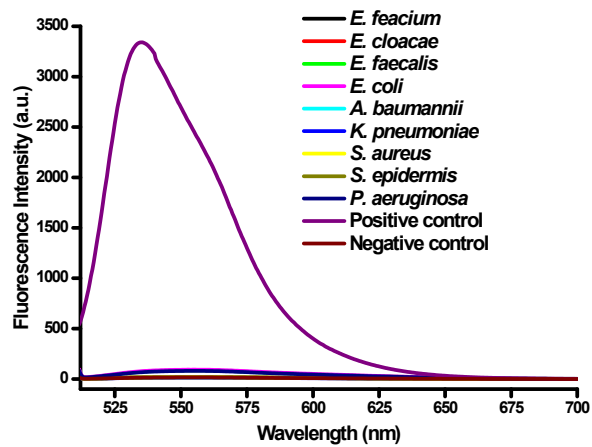
**Figure S1:** The Bar diagram represents the confirmation of pH change over different conditions of the LAMP (single, double, and multiplex) reaction. Data was measured from 5 replicates of each reaction; where error bars represent the standard deviations of five replicates.



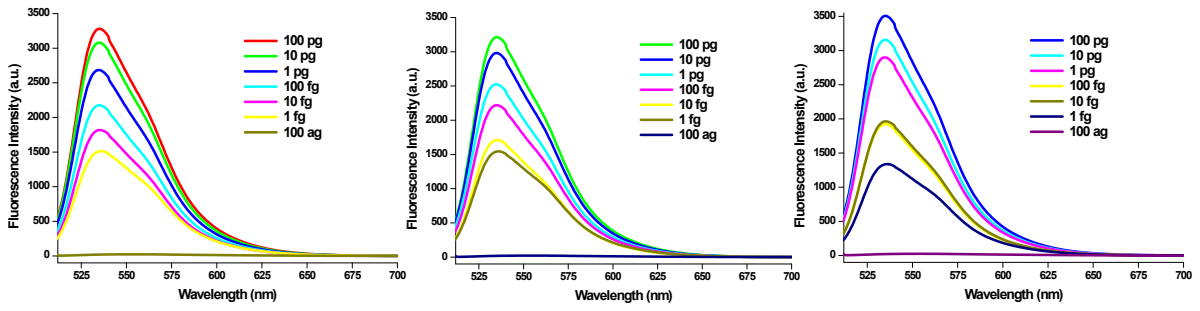
**Figure S2:** Confirming i-motif formation by the TO color change, whereas, after post-LAMP amplification, TO was added. Here, Lane 1: Multiplex LAMP without any target (negative control), Lane 2: Single-LAMP, Lane 3: Double-LAMP, Lane 4: Multiplex-LAMP (positive control). In Figure S2, we can clearly see the color difference between the negative and positive controls of multiplex-LAMP (lane 1 & lane 4). Additionally, from left to right (conversion from light yellow to dense yellow), while the color intensity difference is not significantly recognizable, still upon careful observation; even to the naked eye, the slight variations are more apparent.



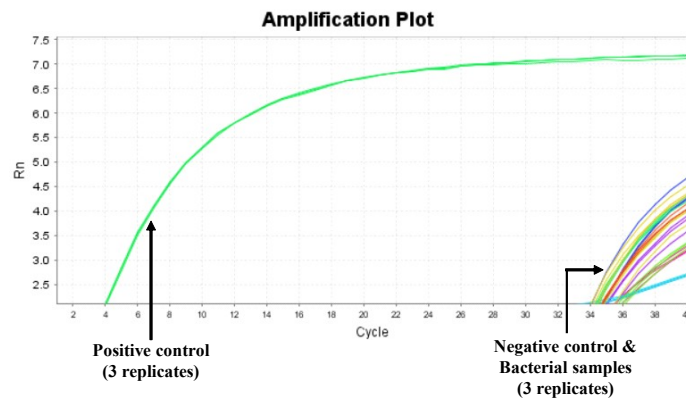
**Figure S3:** Temperature-dependent multiplex-LAMP gradient study to show LAMP primers suitability by 20% non-denaturing PAGE. Lane M: 25/100 bp DNA marker, Lane 1: at 65°C negative control, Lane 2: at 65°C positive control, Lane 3: at 60°C positive control, Lane 4: at 55°C positive control, Lane 5: at 60°C negative control, Lane 6: at 55°C negative control.



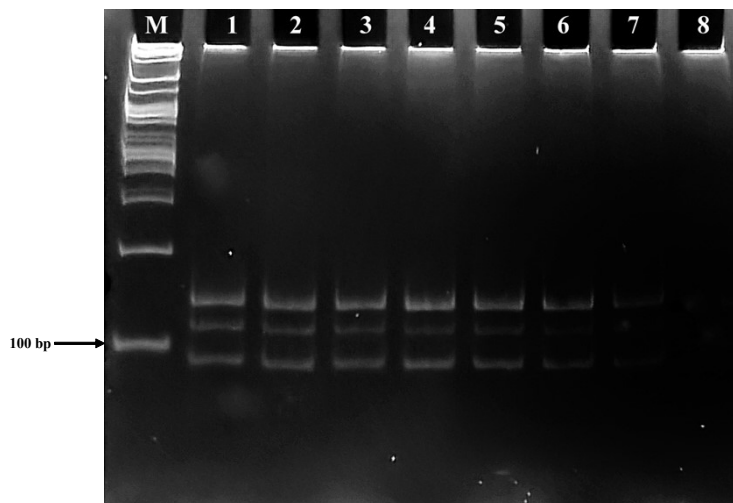
**Figure S4:** Fluorescence measurement of multiplex-LAMP mediated i-motif-TO fluorometric assay selectivity study by using nine different bacterial genomes; including negative and positive control of our system. All fluorescence emission peaks were measured at 535 nm (Thiazole Orange; TO) wavelength.



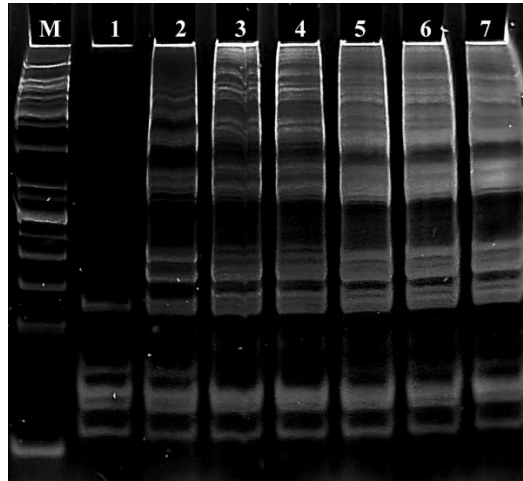
**Figure S5:** Fluorescence measurement of multiplex-LAMP mediated i-motif-TO fluorometric assay sensitivity study. A, B, and C represent the three-replication study of sensitivity experiments. All fluorescence emission peaks were measured at 535 nm (Thiazole Orange; TO) wavelength.



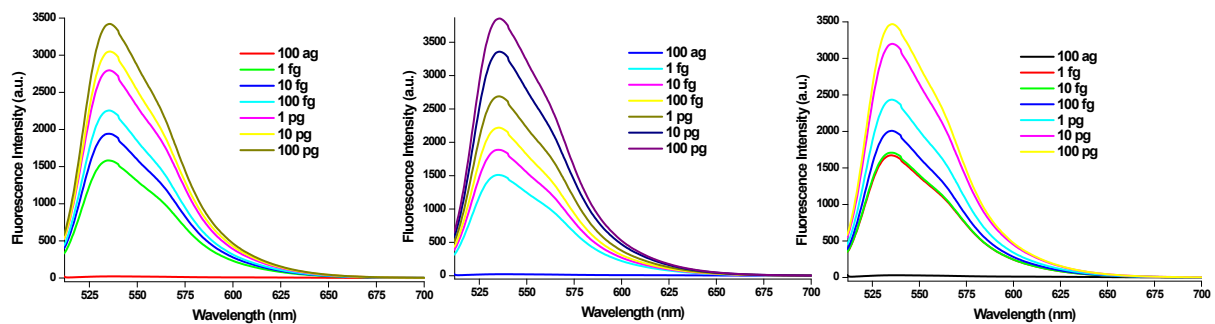
**Figure S6:** Analysis of amplification plot based on selectivity experiment of Mpox detection by Real-time PCR by using nine different bacterial genomes.



**Figure S7:** Analysis of sensitivity experiment of Mpx detection by Real-time PCR through 20% non-denaturing PAGE; where B6R gene (92 mers), N3R (116 mers), and E9L (141 mers). Here, Lane M: 100 bp DNA marker, Lane 1: 100 pg, Lane 2: 10 pg, Lane 3: 1 pg, Lane 4: 100 fg, Lane 5: 10 fg, Lane 6: 1 fg, Lane 7: 100 ag, Lane 8: negative control (without any target).



**Figure S8:** Analysis of sensitivity experiment of multiplex-LAMP mediated i-motif-TO fluorometric detection system with human serum samples to imitate real sample conditions by 20% non-denaturing PAGE. Here, Lane M: 25/100 bp DNA marker, Lane 1: 100 ag, Lane 2: 1 fg, Lane 3: 10 fg, Lane 4: 100 fg, Lane 5: 1 pg, Lane 6: 10 pg, Lane 7: 100 pg.



**Figure S9:** Fluorescence measurement of multiplex-LAMP mediated i-motif-TO fluorometric assay sensitivity study with human serum samples to imitate real sample conditions. A, B, and C represent the three-replication study of sensitivity experiments. All fluorescence emission peaks were measured at 535 nm (Thiazole Orange; TO) wavelength.