Supplementary Information

An Integrated Microfluidic Chip for Rapid and Multiple Antimicrobial Susceptibility Testing

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Fig. S1. AST and MICs assays off-chip using traditional broth dilution method.

Fig. S2. Absorption spectra of CIP before and after freeze-drying. C0 and C1 represent the concentrations of 128 μ g mL⁻¹ and 64 μ g mL⁻¹ of CIP after a 10-fold dilution, respectively.

Table S1. Comparison of the integrated microfluidic chip in this work with the previously reported microfluidic platforms for AST and MIC determination.

Video S1. The process of self-priming and segmentation captured using high-speed camera.

Video S2. The process of self-priming and segmentation simulated using commercial finite element method (FEM) software.

References

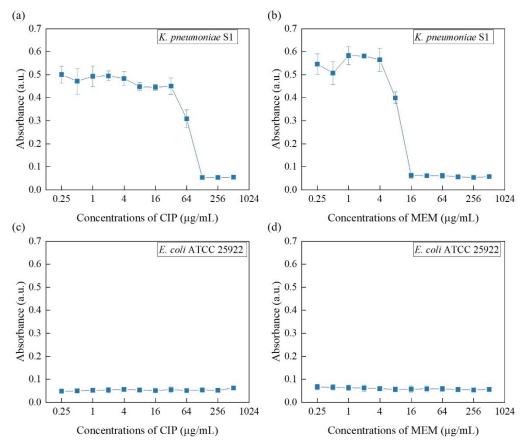


Fig. S1. AST and MICs assays off-chip using traditional broth dilution method. AST for K. pneumoniae S1 inoculated with CIP (a) and MEM (b), and E. coli ATCC 25922 inoculated with CIP (c) and MEM (d) using broth dilution method, respectively. The error bars represent the standard deviation of three replicates.

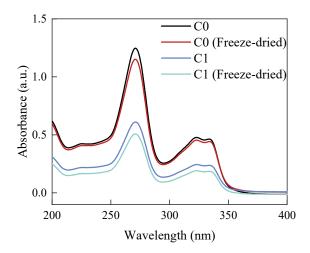


Fig. S2. Absorption spectra of CIP before and after freeze-drying. C0 and C1 represent the concentrations of 128 μ g mL⁻¹ and 64 μ g mL⁻¹ of CIP after a 10-fold dilution, respectively.

| Sample loading | Label | Multiplex detection | MIC determination | Incubation | Reference |
|----------------------------------|------------|---------------------|--------------------------|------------|-----------|
| methods | probe | | methods | time | S |
| Air pressure-driven | Label-free | Two rows of cell | Bacterial growth | Less than | 1 |
| with electropneumatic controller | | traps | rates | 30 min | |
| Pump-driven | Resazurin | Seven linear | Relative increment of | 8–9 h | 2 |
| | | channels | fluorescence intensity | | |
| Centrifugal force- | Label-free | Multiplexed drug | Morphological | 3 h | 3 |
| driven | | testing | changes and number | | |
| | | | of bacteria | | |
| Self-partitioning | Label-free | 192 nanoliter-sized | Morphological | Within 3 | 4 |
| SlipChip | | compartments | changes and number | hours | |
| | | | of bacteria | | |
| Pre-degassing and | Resazurin | - | Digital resazurin | About 3 h | 5 |
| vacuum-driven | | | assay | | |
| Self-priming and | Label-free | Eight detection | Number of bacteria | 2 h | This work |
| vacuum-driven | | areas | | | |

Table S1. Comparison of the integrated microfluidic chip in this work with the previously reported microfluidic platforms for AST and MIC determination.

References

- Ö. Baltekin, A. Boucharin, E. Tano, D. I. Andersson and J. Elf, Proc. Natl. Acad.
 Sci. U.S.A., 2017, **114**, 9170–9175.
- 2 M. Osaid, Y.-S. Chen, C.-H. Wang, A. Sinha, W.-B. Lee, P. Gopinathan, H.-B. Wu and G.-B. Lee, Lab Chip, 2021, **21**, 2223–2231.
- 3 S. Hwang and J. Choi, Lab Chip, 2023, 23, 229–238.
- 4 X. Li, X. Liu, Z. Yu, Y. Luo, Q. Hu, Z. Xu, J. Dai, N. Wu and F. Shen, Lab Chip, 2022, **22**, 3952–3960.
- 5 W. Wu, G. Cai, Y. Liu, Y. Suo, B. Zhang, W. Jin, Y. Yu and Y. Mu, Lab Chip, 2023, **23**, 2399–2410.