

## Supplementary Information

### Ultraportable and Rapid Solid Sample Preparation System Utilizing Twin-Screw Mechanism for Diagnostic Applications

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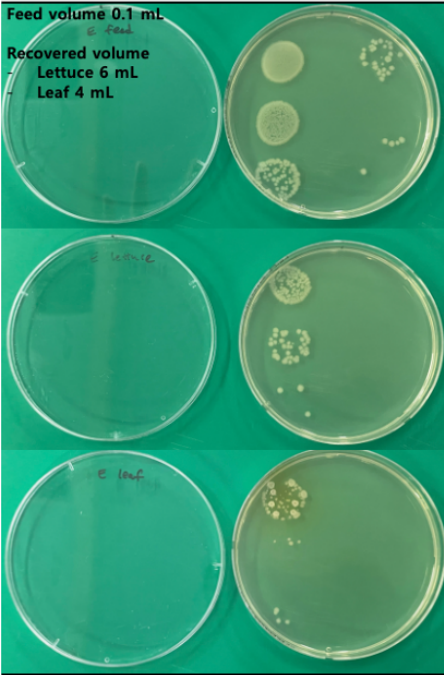
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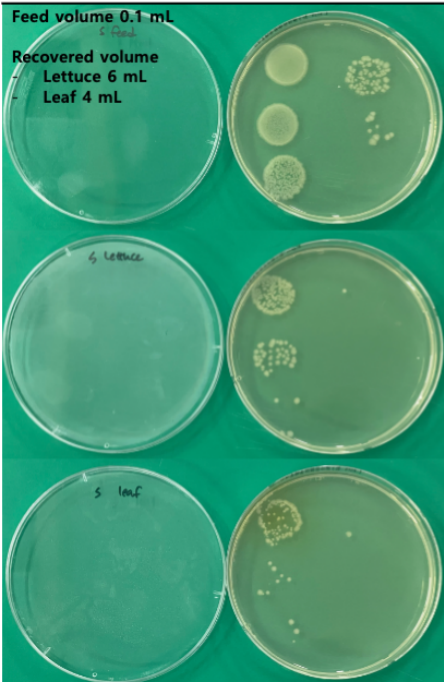
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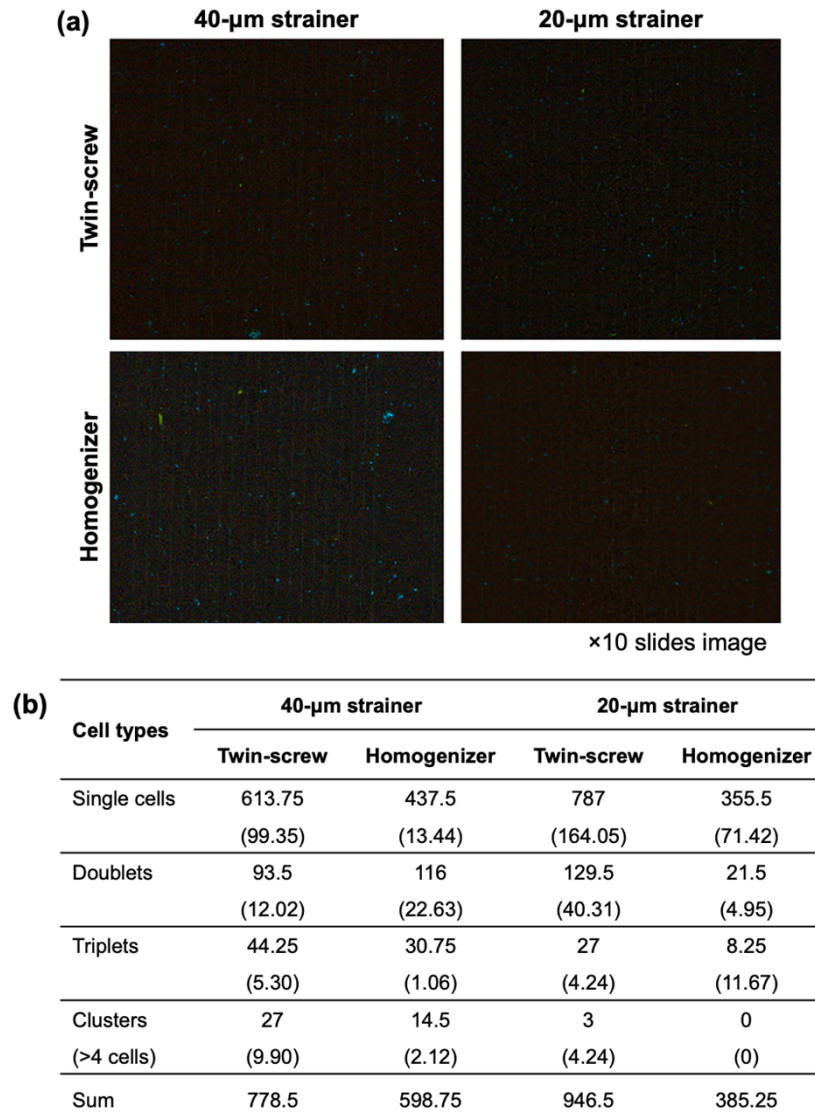
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<b>(a) <i>E. coli</i></b>		Sample type (volume)	Colony count [CFU/50 $\mu$ L]	Concentration [CFU/mL]	Total bacteria [CFU/sample]
	<b>Feed</b> (0.1 mL)	set 1	$3.5 \times 10^5$	$7.0 \times 10^6$	$7.0 \times 10^5$
		set 2	$4.0 \times 10^5$	$8.0 \times 10^6$	$8.0 \times 10^5$
	<b>Lettuce</b> (6 mL)	set 1	$3.7 \times 10^3$	$7.4 \times 10^4$	$4.44 \times 10^5$
		set 2	$3.0 \times 10^3$	$6.0 \times 10^4$	$3.6 \times 10^5$
	<b>Leaf</b> (4 mL)	set 1	$5.2 \times 10^2$	$1.04 \times 10^4$	$4.16 \times 10^4$
		set 2	$4.0 \times 10^2$	$8.0 \times 10^3$	$3.2 \times 10^4$

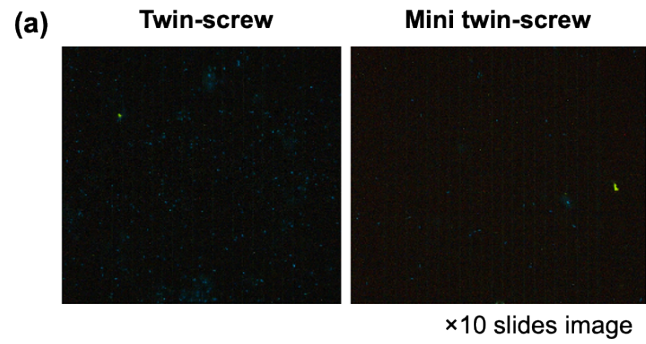
  

<b>(b) <i>S. epidermidis</i></b>		Sample type (volume)	Colony count [CFU/50 $\mu$ L]	Concentration [CFU/mL]	Total bacteria [CFU/sample]
	<b>Feed</b> (0.1 mL)	set 1	$9.0 \times 10^5$	$1.8 \times 10^7$	$1.8 \times 10^6$
		set 2	$4.3 \times 10^3$	$8.6 \times 10^4$	$5.16 \times 10^5$
	<b>Lettuce</b> (6 mL)	set 1	$4.3 \times 10^3$	$8.6 \times 10^4$	$5.16 \times 10^5$
		set 2	$4.0 \times 10^3$	$8.0 \times 10^4$	$4.8 \times 10^5$
	<b>Leaf</b> (4 mL)	set 1	$8.7 \times 10^2$	$1.74 \times 10^4$	$6.96 \times 10^4$
		set 2	$8.0 \times 10^2$	$1.6 \times 10^4$	$5.4 \times 10^4$

**Figure S1.** Agar plates showing colony-forming units (CFU) of (a) *E. coli* and (b) *S. epidermidis* recovered from lettuce and leaf samples after processing, along with the detailed colony counting results (raw data corresponding to Figure 2e in the article).



**Figure S2.** Cell counting process for evaluating the tissue dissociation efficiency of the twin-screw device and homogenizer (raw data corresponding to Figure 3d and 3e in the article): (a) Fluorescence microscopy images of dissociated tissue suspensions loaded onto a hemocytometer for counting DAPI-stained nuclei. (b) Quantitative analysis of cell types based on mean counts and standard deviations. Data are presented as Mean (Standard deviations).



(b) Cell types	Twin-screw	Mini twin-screw
Single cells	622	176
Doublets	198	16
Triplets	141	9
Clusters(>4 cells)	199	11

**Figure S3.** Cell counting process for evaluating the single cell ratio obtained by the twin-screw and miniaturized twin-screw modules (raw data corresponding to Figure 4c in the article): (a) Fluorescence microscopy images of dissociated tissue suspension loaded onto a hemocytometer for counting DAPI-stained nuclei. (b) Quantitative analysis of cell types based on total counts.