# Development of bishydrazide-based fluorescent probes for the

## imaging of cellular peroxynitrite (ONOO<sup>-</sup>) during ferroptosis

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### **Table of Contents**

1. Materials and instruments	S3
2. Preparation method of ONOO- solution	S3
3. Cytotoxicity experiment	
4. Fig. S1-S18	

#### 1. Materials and instruments

All solvents and reagents were commercially available and used without further purification. Doubly distilled water was used in all the experiments. Thin-layer chromatography (TLC) analysis was performed on silica gel plates and column chromatography was conducted over silica gel (mesh 200-300), both of which were purchased from the Qingdao Ocean Chemicals. Fluorescence spectra and relative fluorescence intensity were measured with a Hitachi F-4600 spectrofluorimeter with a 10 mm quartz cuvette. UV/vis spectra were obtained with a Shimadzu UV-2700 spectrophotometer. High-resolution mass spectra (HRMS) for the characterization of structures were collected using a Bruker apex-Ultra mass spectrometer (Bruker Daltonics Corp., USA) in electrospray ionization (ESI) mode. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on an AVANCE III 400 MHz Digital NMR Spectrometer, using tetramethylsilane (TMS) as internal reference. LC-MS were collected using an Agilent 6510 Q-TOF LC/MS.

#### 2. Preparation method of ONOO- solution:

Under the condition of ice bath, HCl solution (0.6 mol/L, 10mL) was added to the mixed solution containing NaNO<sub>2</sub> (0.6 mol/L, 10mL) and H<sub>2</sub>O<sub>2</sub> (0.7 mol/L, 10mL), which was stirred vigorously, followed by NaOH solution (1.5 mol/L, 20mL). Finally, a very small amount of MnO<sub>2</sub> was added and filtered. Note that all the above solutions were prepared from deionized water, and the prepared ONOO<sup>-</sup> must be stored in the dark at -20 ° C.

ONOO<sup>-</sup> Concentration calibration: The UV spectrophotometer was used for testing. Firstly, 2mL of NaOH solution with a concentration of 0.1mol /L was used to sweep the baseline, and then X  $\mu$ L of prepared ONOO<sup>-</sup> solution was added to V mL of NaOH solution with a concentration of 0.1mol /L. The concentration of ONOO<sup>-</sup> solution can be obtained by the following equation. Where A<sub>302nm</sub> is the absorbance of ONOO<sup>-</sup> at 302nm, V is the total volume of solution, and X is the volume of added ONOO<sup>-</sup> solution.

$$c = \frac{A_{302nm} * V}{1.67X}$$

#### 3. Cytotoxicity experiments

HeLa cells were seeded in a 96-well plate at a cell density of 8000 cells/well, cultured for 24 hours, and then replaced with cultures containing different concentrations of **Rh-3** for 24 hours. After that, add 5 mg/mL of MTT solution 10  $\mu$ L to each well to each well, continue to incubate for 3 h, then add 100 mL of dimethyl sulfoxide (DMSO) to dissolve the precipitate, and measure the absorbance of the resulting solution with a microplate reader.



**Fig. S1.** Time-dependent fluorescence spectra (A) and fluorescent intensity (I<sub>561</sub>) (B) of 5  $\mu$ M **Rh-3** in the presence of 20  $\mu$ M ONOO<sup>-</sup> in PBS (pH = 7.4, 20% DMF).  $\lambda_{ex} = 505$  nm



Fig. S2. (A) Fluorescence spectra of 5  $\mu$ M Rh-3 at various pH. (B) Fluorescence spectra of 5  $\mu$ M Rh-3 treated with 30  $\mu$ M ONOO<sup>-</sup> at various pH. (C) Quantified fluorescence intensity (I<sub>561</sub>) for (A) and (B).  $\lambda_{ex} = 505$  nm.



Fig. S3. HRMS data of the reaction products of Rh-3 and ONOO<sup>-</sup>.



Fig. S4. Fluorescence spectra of 5  $\mu$ M Rh-3 in1;1 solution of acetic acid and water.  $\lambda_{ex} = 450$  nm.



Fig. S5. Survial of HeLa cells in the presence of **Rh-3** at various concentrations measured using MTT assay.









**Fig. 9.** <sup>13</sup>C NMR (MeOD-*d*<sub>4</sub>) of **Rh-1**.



Fig. 10. LC-MS data of Rh-1.



**Fig. 11.** <sup>1</sup>H NMR (CDCl<sub>3</sub>) of **Rh-2**.



**Fig. 12.** <sup>13</sup>C NMR (CDCl<sub>3</sub>) of **Rh-2**.











Fig. 15. LC-MS data of Rh-3.



**Fig. 16.** <sup>1</sup>H NMR (MeOD-*d*<sub>4</sub>) of **Rh-COOH**.



**Fig. 17.** <sup>13</sup>C NMR (MeOD-*d*<sub>4</sub>) of **Rh-COOH**.



Fig. 18. LC-MS data of Rh-COOH.