Electronic supporting information for

A portable fluorescence detection device based on a smartphone employing carbon nanodots for Mn²⁺ sensing

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1. Synthesis of N,S-CDs

N,S-CDs were synthesized through pyrolysis of lactose and L-cysteine by following previously reported [R1]. Briefly, 0.4000 g of lactose and 0.1000 g of L-cysteine were homogenously dissolved by 8 mL of sodium hydroxide solution with continuous stirring. Subsequently, the mixed solution was heated at 90°C on a hotplate for 5 minutes. The color of the mixture solution was changed from colorless to light yellow and brown, then left at room temperature, and finally, deionized water into the brown solution to a total volume of 16.00 mL. Parallelly, the CDs were synthesized by the same procedure as N,S-CDs but without adding L-cysteine.

2. Characterization of the N,S-CDs

The N,S-CDs underwent pyrolysis in a highly alkaline solution, utilizing lactose as a carbon precursor and L-cysteine as the source of N and S atoms. The undoped CDs, which serve as a comparison, were produced using the same process but without including L-cysteine. The yielded products performed complete characterization in previous studies [R1]. The morphology of N,S-CDs was investigated using a transmission electron microscope (TEM). The N,S-CDs exhibited a spherical morphology and a narrow size distribution. The average particle size, measured by the ImageJ program as shown in Fig. S1(a), was around 3.1±0.4 nm. The UV-visible and fluorescence spectrometer were utilized to investigate the optical characteristics of the N,S-CDs. The absorption of N,S-CDs demonstrates a distinctive absorption peak at a wavelength of 301 nm. Additionally, the peak of shoulder absorption appeared at a wavelength of 330 nm, which can be attributed to the transition of electrons in the carbon nanostructure, specifically the π - π * or $n-\pi^*$ transition [R2]. The absorption peak of the N,S-CDs exhibited a distinct distinctive peak in contrast with ordinary CDs, as illustrated in Fig. S1(b). In addition, the fluorescence emission characteristic of the N,S-CDs was examined, as represented in Fig. S1(c). The characteristic excitation and emission wavelengths of N,S-CDs were observed to be 420 and 513 nm, respectively. Furthermore, Fourier Transform Infrared (FT-IR) spectroscopy was employed to analyze the constituent functional groups inside the N,S-CDs, as depicted in Fig. S1(d). The peak at 3573.9 cm⁻¹ corresponds to the stretching vibrations of the O-H bond, while the broad absorption band at 2819.8 cm⁻¹ indicates the presence of O-H and carboxylic acid groups. Additionally, the signal observed at a wavenumber of 1570.0 cm⁻¹ can be attributed to the presence of the O-C-O functional group. The 1415.2 cm⁻¹ peak indicates the distinctive -CH₂- chain structure. Furthermore, the presence of N-H and C-N functional groups was verified by observing characteristic peaks at 1741.65 and 1217.0 cm⁻¹, respectively. The results obtained from these approaches confirm the successful preparation of N,S-CDs.



Fig. S1 (a) TEM image of the N,S-CDs, (b) UV–Vis absorption spectra of CDs and N,S-CDs (inset: photographs of CDs (A) and N,S-CDs (B)) under UV irradiation), (c) FT-IR spectrum of CDs and N,S-CDs, (d) fluorescence excitation and emission spectrum of the N,S-CDs at excitation wavelength of 420 nm.



Fig. S2 Linear calibration graph and images of solutions acquired by the smartphone camera for detecting Mn^{2+} in the 0.1 M acetic-acetate buffer (pH 5.0).



Fig. S3 The Δ RGB intensities determined from (a) the same crop and (b) the different crop on the same day and a different day of N,S-CDs in the presence of 5 μ M measured in the 0.1 M acetic-acetate buffer (pH 5.0) using a smartphone.

Table S1 The performance characteristics comparison of smartphone-based portable fluorescence

 sensor as metal ion sensing

Materials	Target analytes	Working range (µM)	Detection limit (µM)	Samples	[Ref.]
Carbon dots	Pb ²⁺	0-0.200	0.00289	Tap water, lake water	[R3]
Carbon dots	Hg ²⁺ , Pb ²⁺ , Cu ²⁺	0.020-0.320(Hg ²⁺),		River water	[R4]
		0.2-6.4 (Pb ²⁺),	0.0058 (Hg ²⁺), 0.12 (Pb ²⁺), 0.076 (Cu ²⁺)		
		0.125–2(Cu ²⁺)			
Cadmium telluride quantum dots	Ni ²⁺	0.34-1.70 (20-100 μgL ⁻¹)	0.11(6.2 µgL ⁻¹)	Tea	[R5]
Carbon dots	Cu ²⁺		0.344	-	[R6]
Rhodamine B-Gold nanoparticles	Zn ²⁺	1.5 - 91.7	1.5	TT 7	[R7]
		$(0.1 - 6 \text{ mgL}^{-1})$	(0.1 mgL ⁻¹)	Water	
Silver nanoclusters	Hg^{2+}	0.050-2.50	0.0090	Porphyra	[R8]
		$(0.010 - 0.5 \ \mu g \ mL^{-1})$	(1.8 µgL ⁻¹)		
N, S-doped carbon dots	Mn^{2+}	1-5	0.5	Cosmetics	This work

Materials	Detection Method	Working range (µM)	Detection limit (µM)	Interference s	Samples	[Ref.]
Silicon nanoparticle clusters	Fluorescence, Colorimetry	1-10	1.2	Cu ²⁺	Fingerprints	[R9]
Silicon nanoparticles	Fluorescence	2.5-250	1.1	Fe ³⁺ , Cu ²⁺	Tap water	[R10]
Copper nanoparticles	Fluorescence	2.5-250	1.6	Fe^{3+}, Pt^{2+}	-	[R11]
ZnCdS/ZnS QD/carboxymethyl chitosan/g-C ₃ N ₄ nanosheet nanocomposite film	Fluorescence	1.961-16.67	-	Hg ²⁺	Water samples	[R12]
N-C QD/CCSCD nanocomposites*	Fluorescence	0-21.1	0.0053	Fe ³⁺ , Mg ²⁺	Water samples	[R13]
N, S-doped carbon dots	Fluorescence	0-5	0.12	-	Tap water, wastewater	[R14]
Histidine functionalized carbon quantum dots	Fluorescence	0.064-0.65	0.034	-	Whole blood	
		$(3.50-35.5 \ \mu g \ L^{-1})$	$(1.85 \ \mu g \ L^{-1})$			[R15]
N, S-doped carbon dots	Fluorescence- based smartphone	1-5	0.5	-	Cosmetics	This work

Table S2 The performance characteristics comparison of the proposed sensor with other nanomaterials-based fluorescence sensors for the determination of Mn^{2+}

*N-C QD/CCSCD nanocomposites ; Carboxymethyl chitosan crosslinked with cyclodextrin containing hydrogen-bonded N-C QDs

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