

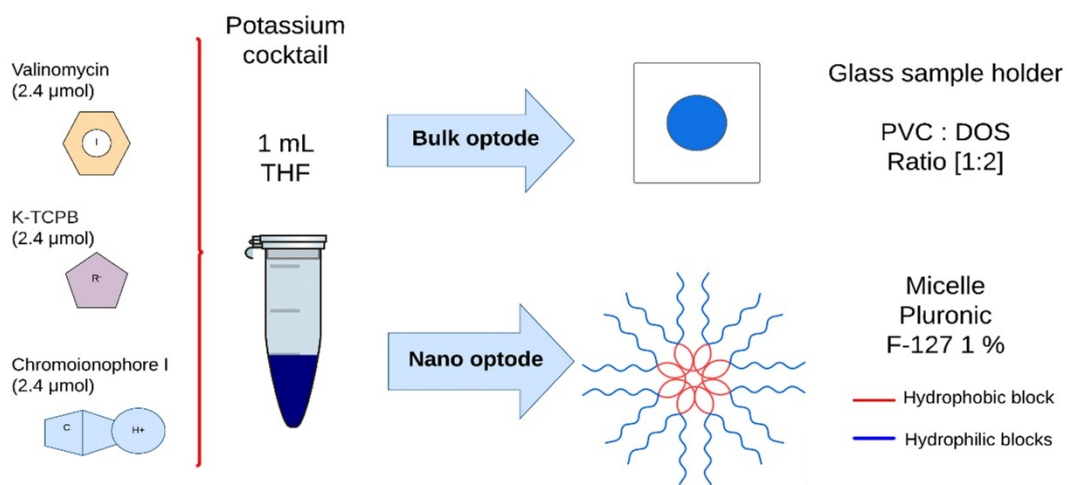
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## Supplementary information

2 **Comparative analysis of a bulk optode based on a valinomycin ionophore and a**  
3 **nano-optode in micelles with Pluronic F-127 for the quantification of potassium in**  
4 **aqueous solutions.**

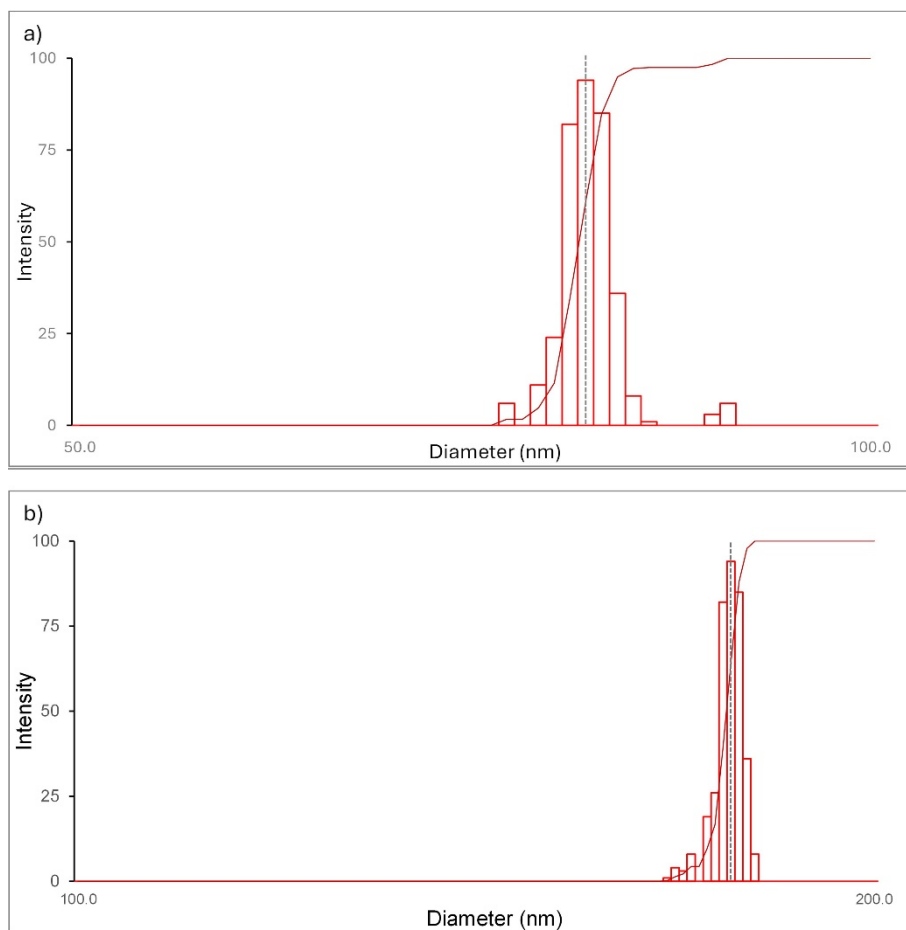
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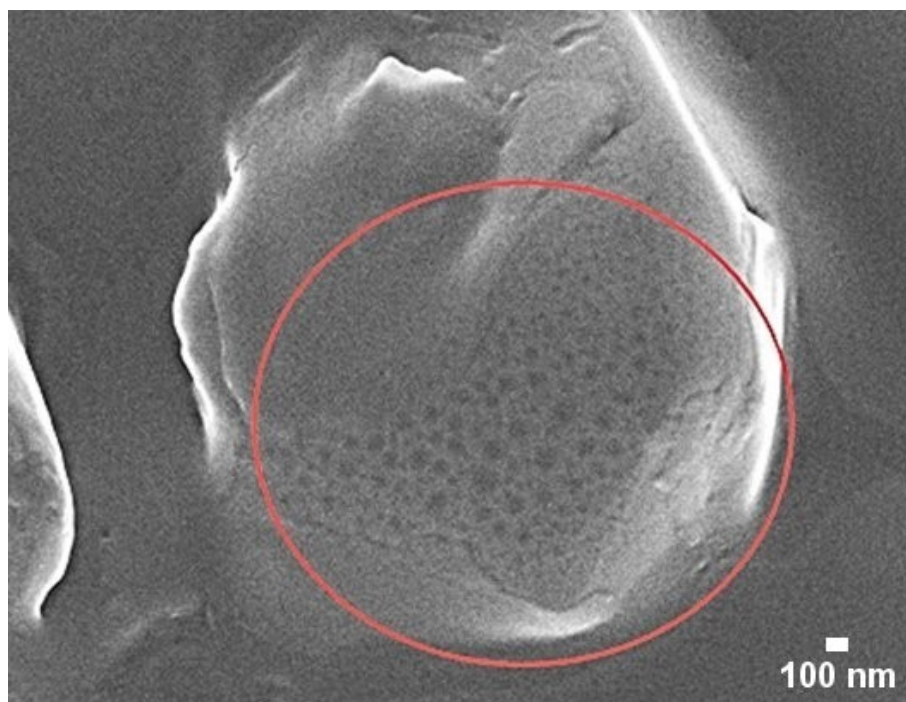
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9 **Figure S1.** Experimental scheme for the synthesis of BO and NO.



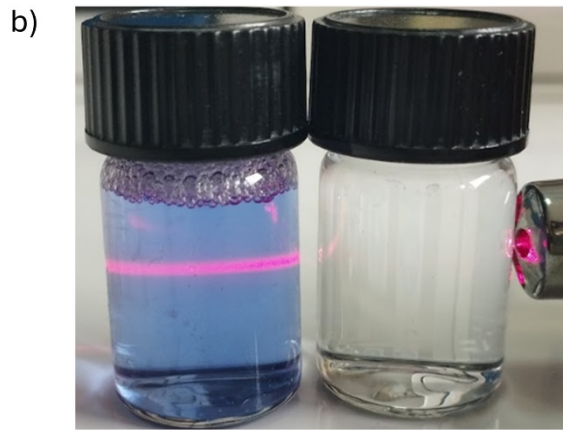
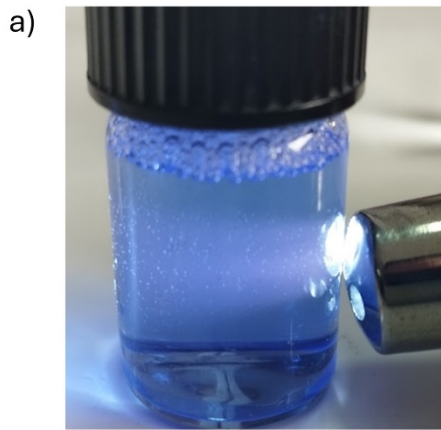
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11 **Figure S2.** DLS analysis for a) Pluronic F-127 micelles and b) NO micelles.



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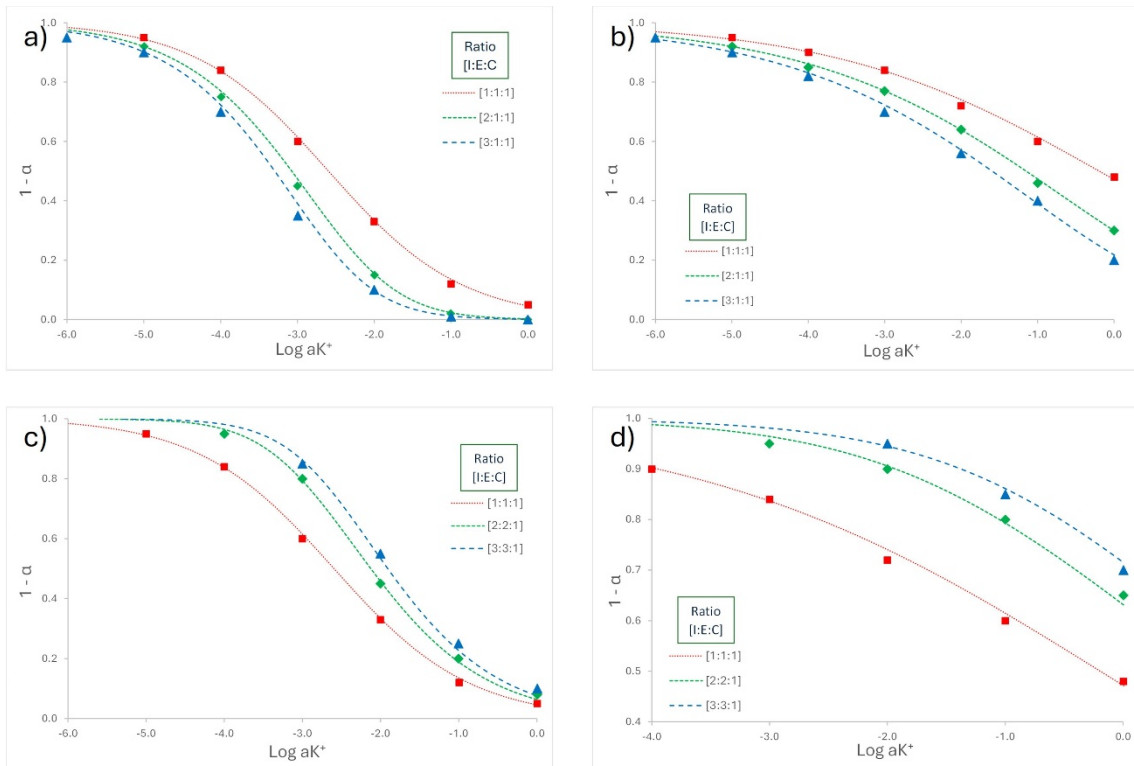
13 **Figure S3.** SEM image of NO micelles.



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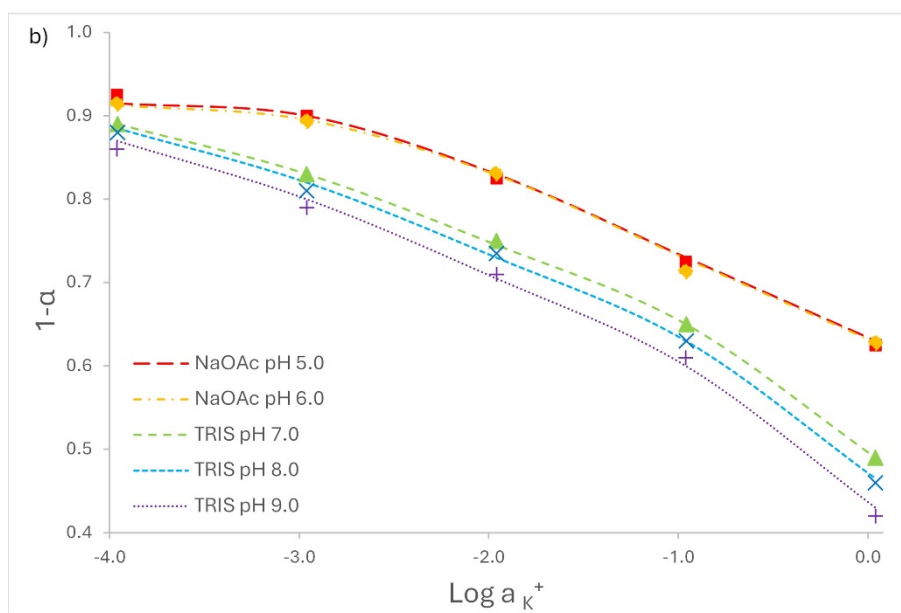
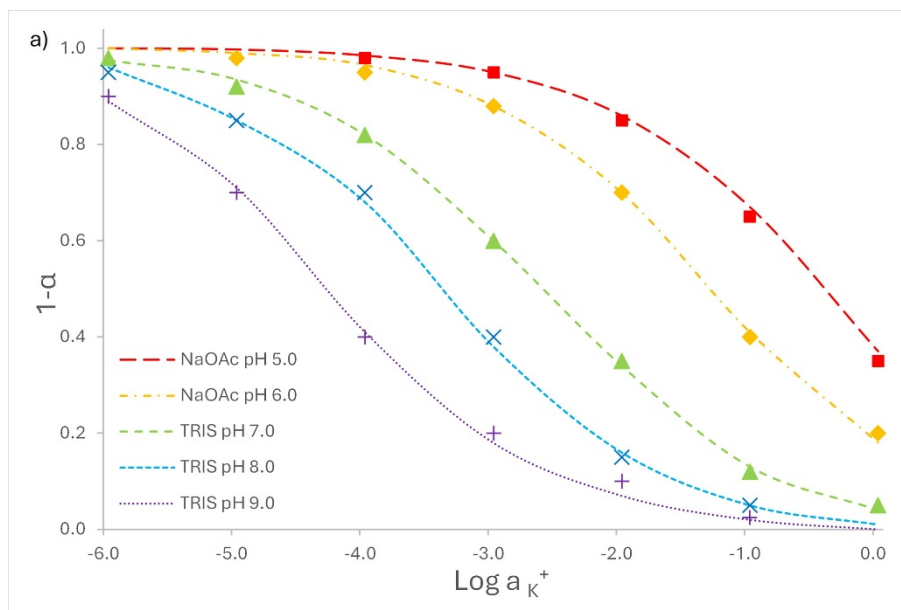
15 **Figure S4.** Tyndall effect in NO solutions. a) With a light source present. b) With a laser  
 16 beam present.

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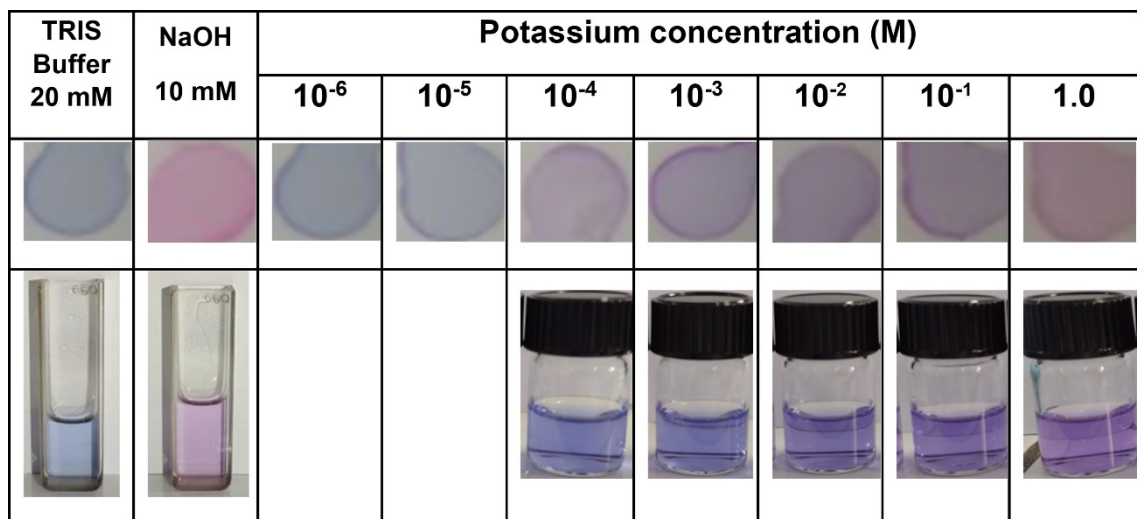
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19 **Figure S5.** Effect of increasing the ionophore valinomycin on the sigmoid model of (a)  
 20 BO and (b) NO. Effect of increasing the ion exchanger K-TCPB on the sigmoid model of  
 21 (c) BO and (d) NO. Measurements were performed in 20 mM TRIS-HCl buffer pH 7.0.



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23 **Figure S6.** Sigmoid curve behavior at pH 5.0-9.0 for (a) BO and (b) NO. Measurements  
 24 were performed in 0 20 mM TRIS-HCl buffer and 20 mM acetate-acetic buffer.



25

26 **Figure S7.** Photographs of BO (top) and NO (bottom) for different concentrations of K<sup>+</sup>  
 27 in 20 mM TRIS buffer at pH 7.0.

28 **Table S1.** The logarithm of the selectivity coefficients of the optodes for potassium ions  
 29 based on the valinomycin ionophore against interfering ions.

<i>Log K<sup>+</sup> - j<sup>+</sup></i>				
j = interferer	Na <sup>+</sup>	Li <sup>+</sup>	Ca <sup>2+</sup>	Mg <sup>2+</sup>
Bulk optode	1.79	3.45	2.09	2.20
Nano optode	3.04	3.21	4.01	4.20

30 **Table S2.** Publications of potassium optical sensors

Methodology	Signal	Selective agent K <sup>+</sup>	Ion exchanger	Optical agent	Support	Type of sample	Limit of detection
ISO (this work)	Colorimetric	Valinomycin	K-TCPB	CHI	BO: PVC - DOS NO: F-127	-	BO: 1.0 μM NO: 0.1 mM
CQD <sup>71</sup>	Fluorescence quenching	CQD	-	CQD	Gelatin powder	Human blood serum	0.01 mM
POD <sup>70</sup>	Colorimetric	Valinomycin	K-TCPB	CHI	Paper substrate	-	0.1 mM
ISO <sup>72</sup>	Colorimetric	Valinomycin	DOP	CHI	PVC	Blood plasma	2.3 mM

Polyoctylthiophene nanoptodes <sup>66</sup>	Fluorescence	Valinomycin	Na-TFPB	CHI	POT - PVA - DOS	Bovine serum album	1.0 · 10 <sup>-6</sup> M
ISO <sup>67</sup>	Fluorescence	Valinomycin	Na-TFPB	SDI - SDII	Nylon membrane	Aqueous sample solution	1.0 μM
POD <sup>68</sup>	Colorimetric	Valinomycin	BARF	CHI	F-127 - DOS	Biological fluids	2.0 mM
ISO <sup>69</sup>	Colorimetric	Valinomycin	Na-TFPB	CHI	F-127 - Agarose	-	0 - 2.0 mM
ISO <sup>41</sup>	Fluorescence	Valinomycin	Na-TFPB	SD	Organosilica nanospheres - nylon filter paper	Aqueous sample solution	1.0 μM

### 31 Method of least-squares for calculated the Ke

32 The equilibrium constant of the valinomycin-potassium ion complex ( $K_e^{KI^+}$ ) could be  
33 expressed as Equation S1.

$$34 \quad \text{Log} \left( \frac{a_{K^+}}{a_{H^+}} \right) + \text{Log} K_e^{KI^+} = \text{Log} \left( \frac{\alpha}{(1-\alpha)} \frac{[C_E] - (1-\alpha)[C_C]}{[C_I] - [C_E] + (1-\alpha)[C_C]} \right)$$

35 Equation S1

36 If we defined  $\text{Log} \left( \frac{a_{K^+}}{a_{H^+}} \right) = X_i$ ;  $\text{Log} K_e^{KI^+} = C$  and

37  $\frac{\alpha}{(1-\alpha)} \frac{[C_E] - (1-\alpha)[C_C]}{[C_I] - [C_E] + (1-\alpha)[C_C]} = Y_i$ . A simple way to find C is to average the

38 distances between Xi and Yi as an Equation S2.

$$39 \quad C = \frac{\sum (Y_i - X_i)}{n} \quad \text{Equation S2}$$

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