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Electronic Supplementary Material

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3 **A CRISPR/Cas12a-based Fluorescent Method for the** 4 **Amplified Detection of Total Antioxidant Capacity**

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12 68349046, Dr. Yue He, E-mail: yuehe@cric.cn

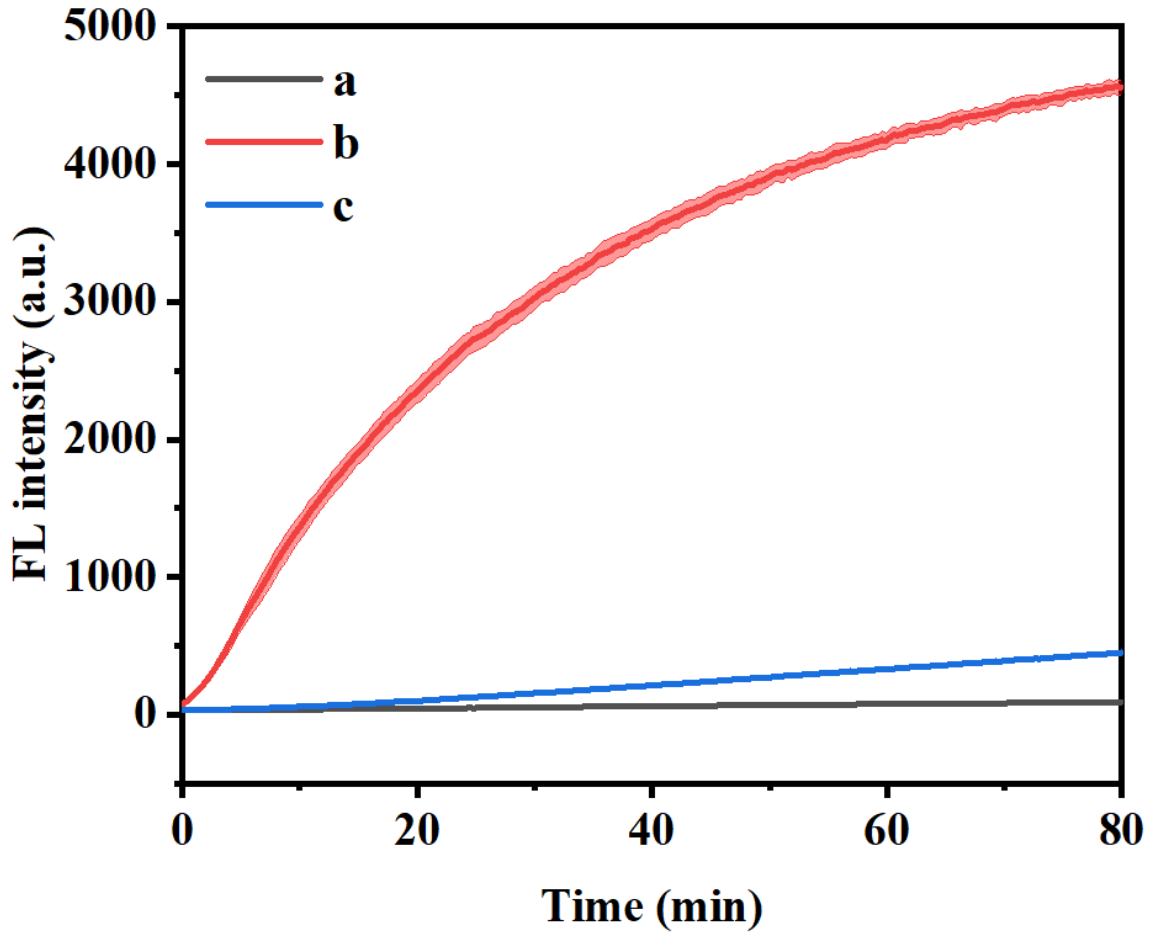
14 **This file contains:**

15 1. Supporting figures 1-6 with legends

16 2. Supporting tables 1-3

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18 1. Supporting figures 1-5 with legends



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20 **Fig. S1** Time-dependent *trans*-cleavage signals of Cas12a under different treatments: (a)

21 CRISPR/Cas12a system; (b) CRISPR/Cas12a system + Mn^{2+} ; (c) CRISPR/Cas12a

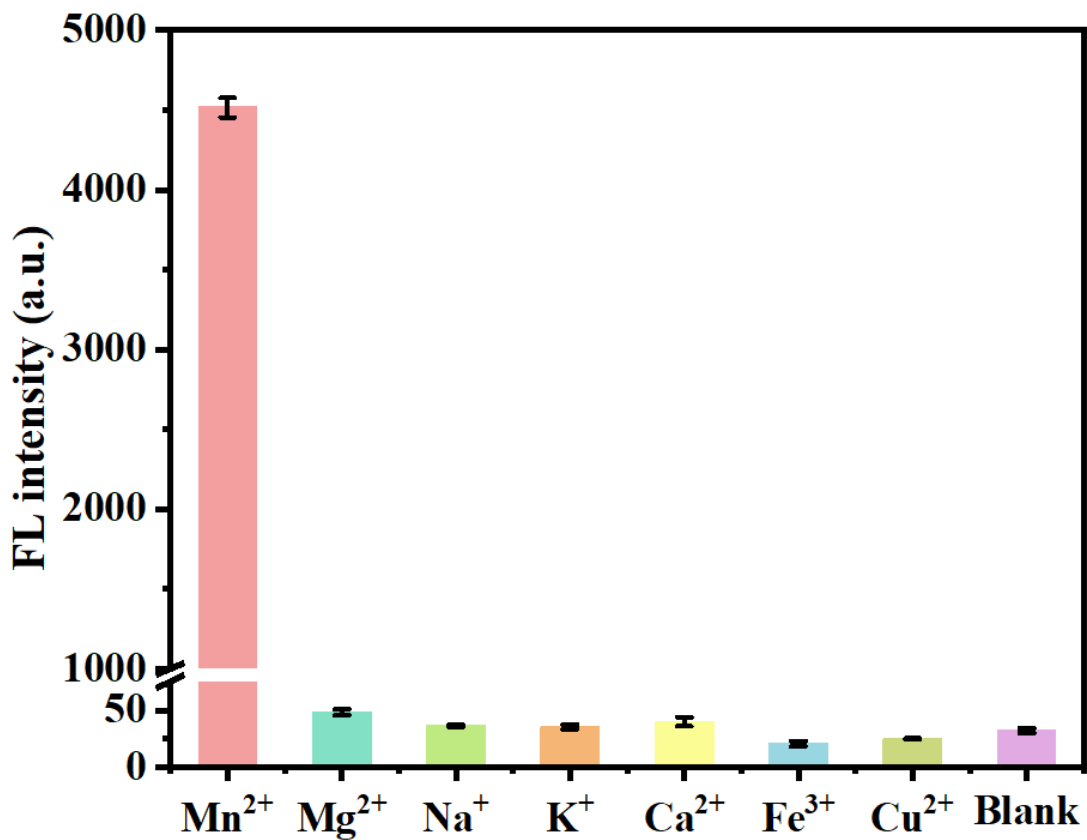
22 system + (Mn^{2+} + EDTA). The CRISPR/Cas12a system contain Cas12a-crRNA complex

23 (25 nM), substrate ssDNA (20 nM), and ssDNA-FQ (500 nM). Mn^{2+} , 30 μM ; EDTA, 30

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μM

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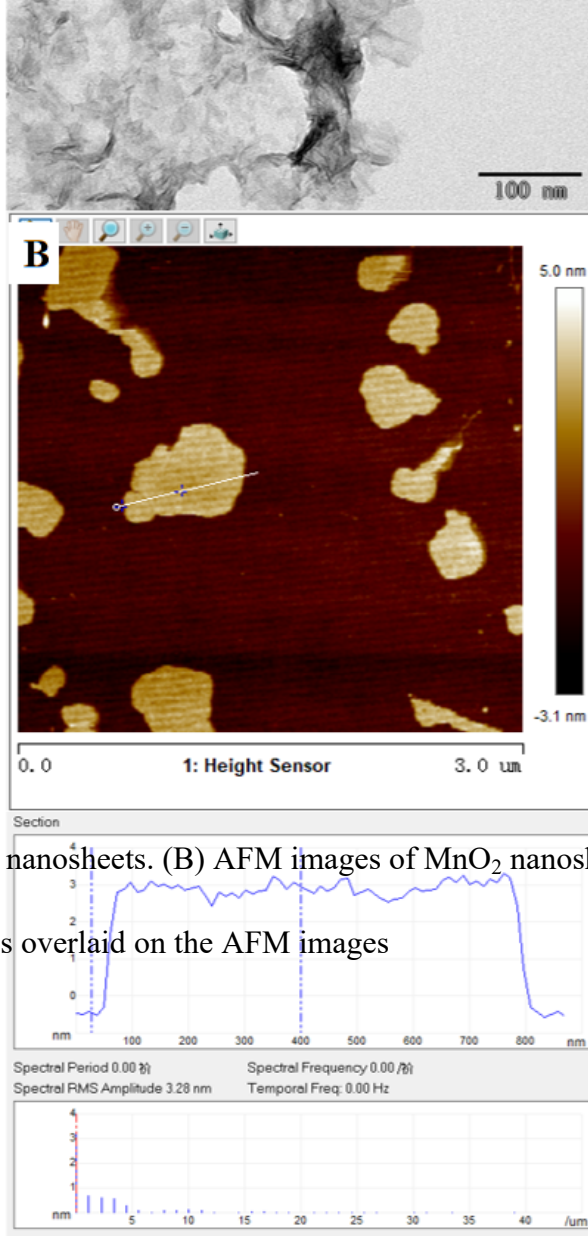
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27 **Fig. S2** Assess the effect of metal ions on the *trans*-cleavage activity of Cas12a. The
 28 concentrations of Mn²⁺ and Mg²⁺ were 30 μM. And the concentrations of other metal ions
 29 were 60 μM. Cas12a-crRNA complex, 25 nM; ssDNA-FQ, 500 nM; substrate ssDNA, 20
 30 nM. The error bars were calculated using the standard deviation (SD) of three repeated

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experiments

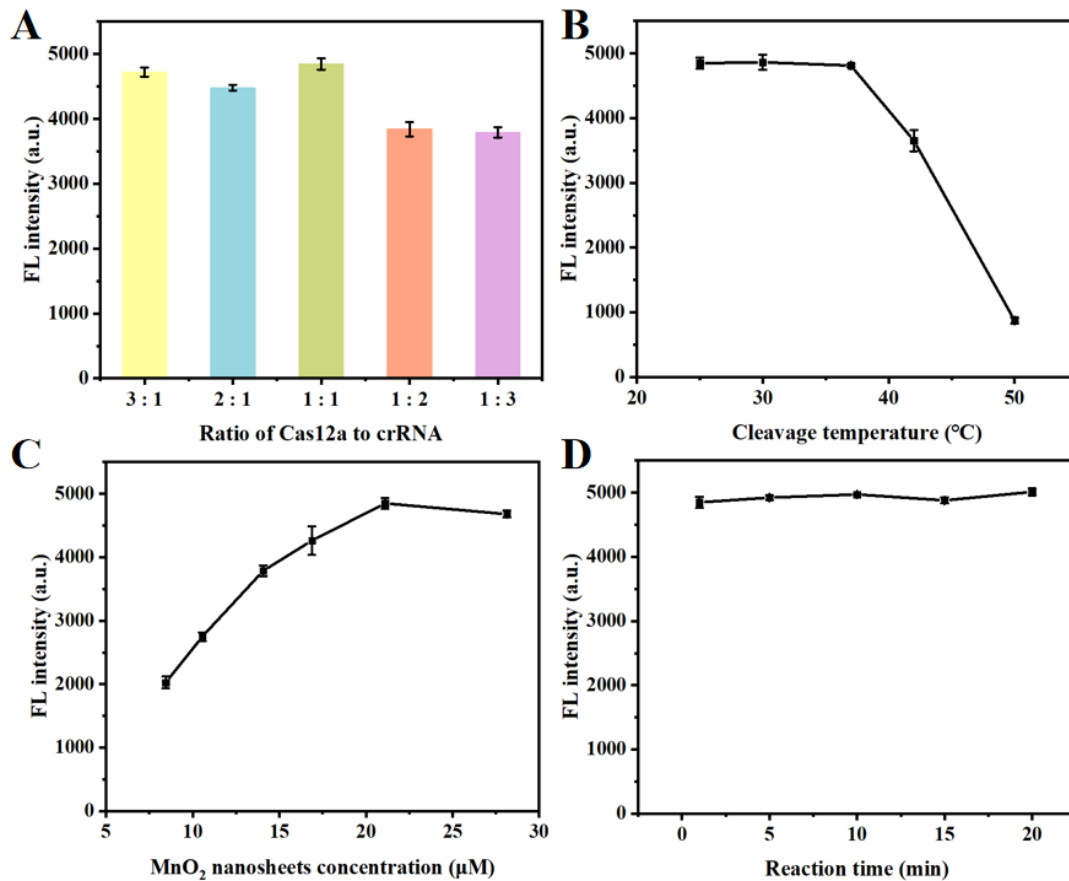
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34 **Fig. S3** (A) TEM images of MnO₂ nanosheets. (B) AFM images of MnO₂ nanosheets and

35 height profiles along the white lines overlaid on the AFM images



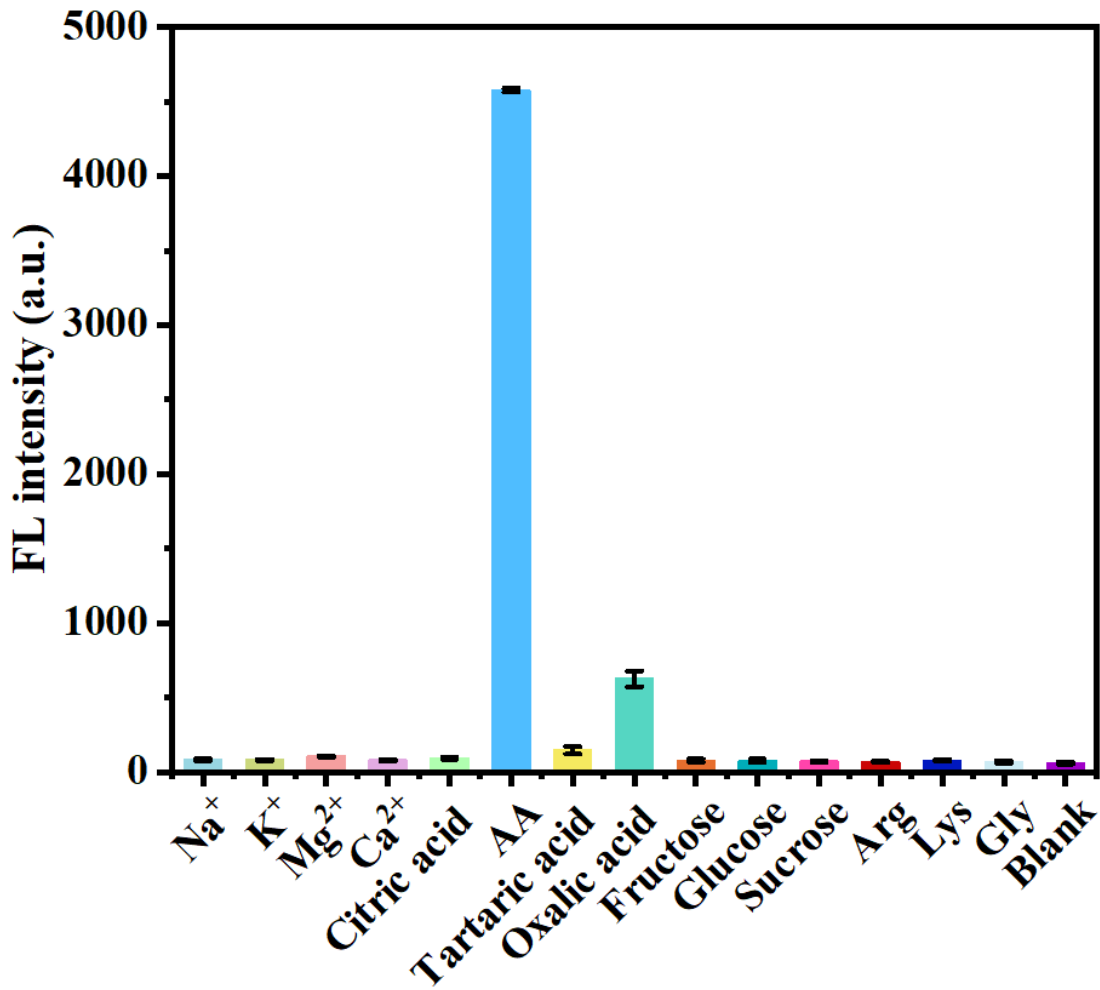
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37 **Fig. S4** Conditions optimization of the proposed FL method. (A) The molar ratio of
 38 Cas12a to crRNA. (B) The cleavage temperatures for Cas12a. (C) The concentration of
 39 MnO₂ nanosheets. (D) The reaction time between AA and MnO₂ nanosheets. Cas12a-
 40 crRNA complex, 25 nM; ssDNA-FQ, 500 nM; substrate ssDNA, 20 nM; AA, 25 μM;
 41 MnO₂ nanosheets, 21.1 μM. The error bars were calculated using the SD of three

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repeated experiments

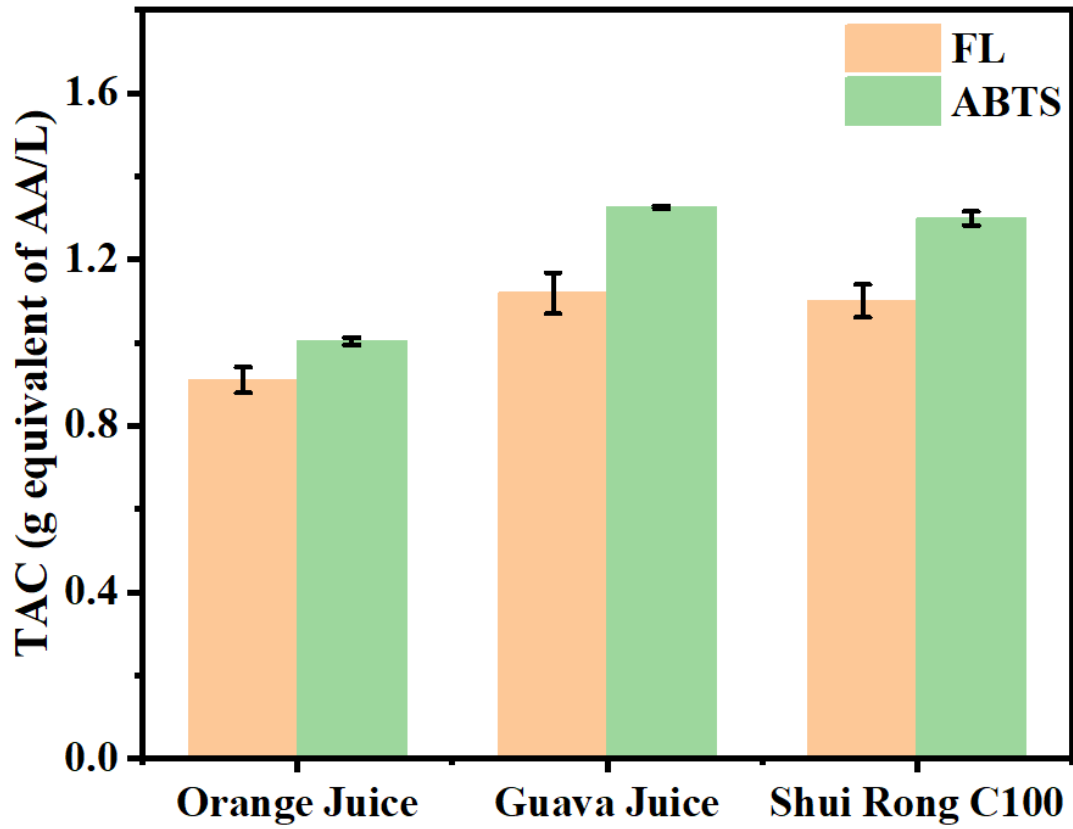
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45 **Fig. S5** The FL response intensities of the proposed FL method to Na⁺, K⁺, Mg²⁺, Ca²⁺,
 46 citric acid, tartaric acid, oxalic acid, fructose, glucose, sucrose, Arg, Lys, Gly and AA at a
 47 concentration of 20 mg·L⁻¹. The error bars were calculated using the SD of three repeated
 48 experiments

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51 **Fig. S6** Determined TAC values in the three beverage samples and comparison with

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standard ABTS assay

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55 **2. Supporting tables 1-3**

56 **Table S1** Oligonucleotide sequences used in this work

Oligonucleotide	Sequence (5'→3')
crRNA	UAAUUUCUACUAAGUGUAGAUUCCUAGUGGGGGCGAACCC
substrate ssDNA	GGGTTCGCCCACTAGGA
ssDNA-FQ	FAM-TTATT-BHQ1

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58 **Table S2** The FL intensity of three repeated experiments and the corresponding relative
 59 standard deviation (RSD) values of the sensing solution for different concentrations of AA

Concentration (mg·L⁻¹)	Data			Average value	RSD/%
20	4595	4466	4646	4569.00	2.03
15	4358	4181	4501	4346.67	3.69
12.5	4146	4156	4051	4117.67	1.41
10	3149	3206	3123	3159.33	1.34
7.5	2416	2382	2579	2459.00	4.28
5	1365	1107	1120	1197.33	12.14
2.5	375.2	483.5	398.5	419.07	13.60
1	175.4	157.4	136.8	156.53	12.34
0.5	72.97	97.02	78.41	82.80	15.23
0.1	54.17	63.25	54.58	57.33	8.94
0	52.02	60.54	58.57	57.04	7.82
Average value					7.53

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61 **Table S3** Comparison the analytical performance of different fluorescent methods for AA

62 detection

Materials	Linear Range (μM)	Detection Limit (μM)	Reference
BSA-AuNCs	3-50	0.4	1
CrO_4^{2-} @Cd-MOFs	46.3-591	41.28	2
GQDs-hypochlorite	8-60	1.4	3
RhB@MOF nanocomposite	1-25	0.31	4
CuInZnS QDs@FeOOH	5-60	1.5	5
DBHM + Cu^{2+} sensor	0-500	2.37	6
Cu NPs/N-Ti ₃ C ₂ T _x	5-150	0.437	7
NiNCs/ Fe^{3+} composites	20-200	7.45	8
CA-CDs	5-100	0.15	9
Fe_3O_4 @SiO ₂ @UiO-PBA	5-60	2.5	10
CRISPR/Cas12a-MnO ₂	2.84-70.97	0.23	This work

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64 **References:**

- 65 1. P. Ni, S. Liu, B. Wang, C. Chen, Y. Jiang, C. Zhang, J. Chen and Y. Lu, *Journal of Hazardous*
66 *Materials*, 2021, **411**.
- 67 2. J. N. Xiao, J. J. Liu, M. Y. Liu, G. F. Ji and Z. L. Liu, *Inorganic Chemistry*, 2019, **58**, 6167-6174.
- 68 3. M. Wang, J. Chen, C. Liu, J. Qiu, X. Wang, P. Chen and C. Xu, *Small*, 2017, **13**.
- 69 4. L. Guo, Y. Liu, R. Kong, G. Chen, Z. Liu, F. Qu, L. Xia and W. Tan, *Analytical Chemistry*, 2019, **91**,
70 12453-12460.
- 71 5. Y. B. Liu, G. Y. Sun, P. Y. Ma and D. Q. Song, *Talanta*, 2024, **271**.
- 72 6. Q. Meng, J. X. Yao, M. Y. Chen, Y. J. Dong, X. Y. Liu, S. Y. Zhao, R. Qiao, C. B. Bai, C. Q. Qu and
73 H. Miao, *Analytica Chimica Acta*, 2023, **1276**.
- 74 7. J. Huang, C. Shen, H. Gu, G. Wang, P. Zhou, X. Liu, K. Yu, Y. Qin, K. Zhou, J. Zhang and Z. Chen,
75 *ACS Sustainable Chemistry & Engineering*, 2023, **11**, 17472-17481.
- 76 8. J. Li, N. Yao, X. Zhang and Y. Liu, *Dyes and Pigments*, 2024, **221**.
- 77 9. Z. Lin, Q. Zeng, W. Yao, W. Chen, C. Cai, J. Yang, X. Lin and W. Chen, *Food Chemistry*, 2024, **437**.
- 78 10. T. Wan, Z. Zhang, H. Wang, Y. Yang, H. Wang, J. Zhang, Y. Zeng and L. Li, *Spectrochimica Acta*
79 *Part A: Molecular and Biomolecular Spectroscopy*, 2024, **305**.
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