

## Supporting Information

### **Laser-Induced Graphene/Gold Nanoparticle Hybrid Sensor for Enhanced Electrochemical Detection of Paracetamol**

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## Reagents and instruments

Paracetamol (Acetaminophen, AP), potassium ferricyanide ( $K_3[Fe(CN)_6]$ ), and potassium ferrocyanide ( $K_4[Fe(CN)_6]$ ) were purchased from Damas-beta (Shanghai, China). The polyimide film (25  $\mu\text{m}$ , DuPont, USA), silver/silver chloride ink (EN-06B8, Enson, China), silver ink (JLL-20, JuLong, China), polyethylene terephthalate (PET) film (10  $\mu\text{m}$ ), screen-printed carbon electrodes (SPCE), and screen-printed gold electrodes (SPAEC) were all bought on Taobao. The glassy carbon electrode (GCE) was acquired from ChenHua Instrument Co. Ltd. Phosphate buffered saline (10 mM KCl, pH = 7.4), chloroauric acid trihydrate ( $HAuCl_4 \cdot 3H_2O$ ), catechol (CC), resorcinol (RC), hydroquinone (HQ), ethanol, and acetone were purchased from Sigma Aldrich, China. Dopamine (DA), L-ascorbic acid (AA), and urea (UA) were sourced from Macklin, Shanghai, China. All the reagents mentioned above are of analytical grade. Ultrapure water ( $\geq 18 \text{ M}\Omega \cdot \text{cm}$ ) was produced by the Hitachi water purification system (Shanghai, China).

The scanning electron microscope (SEM) images were captured using a Zeiss Gemini SEM 300. Powder X-ray diffraction (PXRD) was performed on a SmartLab SE diffractometer (Rigaku, Tokyo, Japan) with Cu  $K\alpha$  radiation. Raman spectra were collected using a Renishaw inVia Raman microscope. X-ray photoelectron spectroscopy (XPS) was conducted on a Thermo Fisher ESCALAB 250Xi using an Al  $K\alpha$  laser (Thermo Fisher Scientific, USA). Contact angle images were obtained using a Theta Flex (BioLin, Sweden).

## Preparation of AuNPs/LIG electrode

First, a 10.6  $\mu\text{m}$   $\text{CO}_2$  laser was used to mark out the working electrode area of the three-electrode system on a 100  $\mu\text{m}$  PI substrate. Then, 100 mM  $\text{HAuCl}_4$  solution was drop-cast onto the obtained LIG working electrode at a loading volume of 10  $\mu\text{L}$ . After the chloroauric acid precursor solution dried at room temperature, a second laser treatment was conducted under the same conditions as the first to obtain an AuNPs/LIG composite nanoelectrode. The settings for both laser treatments were: laser power of 2.5 W, speed of 150  $\text{mm}\cdot\text{s}^{-1}$ , line spacing of 0.08 mm, and the laser mode was vector mode. Once the laser-marked electrode was prepared, screen printing technology was utilized to quickly print conductive silver paste and silver/silver chloride paste, which served as the reference electrode and connection wires in the three-electrode system, through pre-designed screen meshes and a scraper, followed by curing in an oven at 80°C for 5 minutes. Finally, a layer of PET film was applied over the AuNPs/LIG electrode as a passivation layer to isolate the working electrode area. The pattern of the three-electrode system was pre-designed using CAD software.

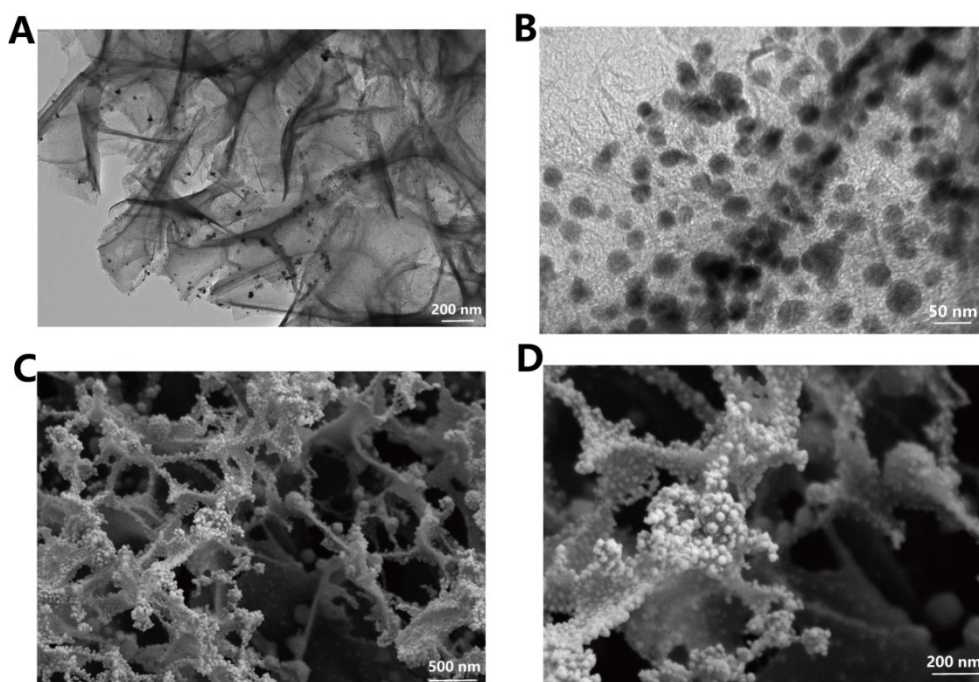
### **Electrochemical detection using AuNPs/LIG electrodes**

To investigate the preparation conditions of AuNPs/LIG and the effect of external strain on electrochemical behavior, cyclic voltammetry (CV) measurements were conducted within a voltage range of -0.2 V to +0.6 V in a 0.1 M KCl solution containing 5.0 mM  $\text{K}_3[\text{Fe}(\text{CN})_6]^{3-/4-}$ . In a 0.1 M PBS solution with a pH of 7, differential pulse voltammetry (DPV) was employed at a scan rate of 0.05 V/s to detect the electroactivity of acetaminophen within the range of -0.2V to +0.6V.

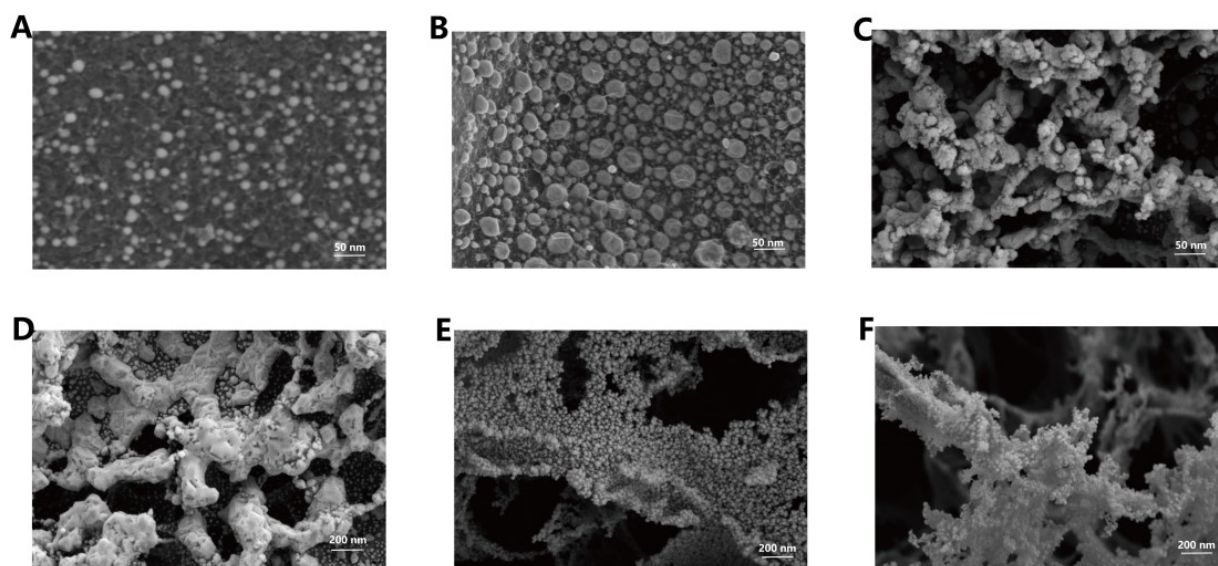
## Actual sample analysis

Tylenol brand phenacetin-containing tablets were purchased from a pharmacy, and one tablet was ground into powder. According to the acetaminophen content indicated in the instruction manual, it was prepared into a standard concentration for testing. Tap water was directly sampled from the laboratory's water pipe. After filtering out physical impurities with a 0.45  $\mu\text{m}$  filter, the tap water samples were diluted and adjusted to pH using a 0.1 M PBS solution (pH=7.0) at a ratio of 1:1 (v:v). The standard addition method was used to analyze acetaminophen in the tap water samples. The formula for determining the recovery rate is:

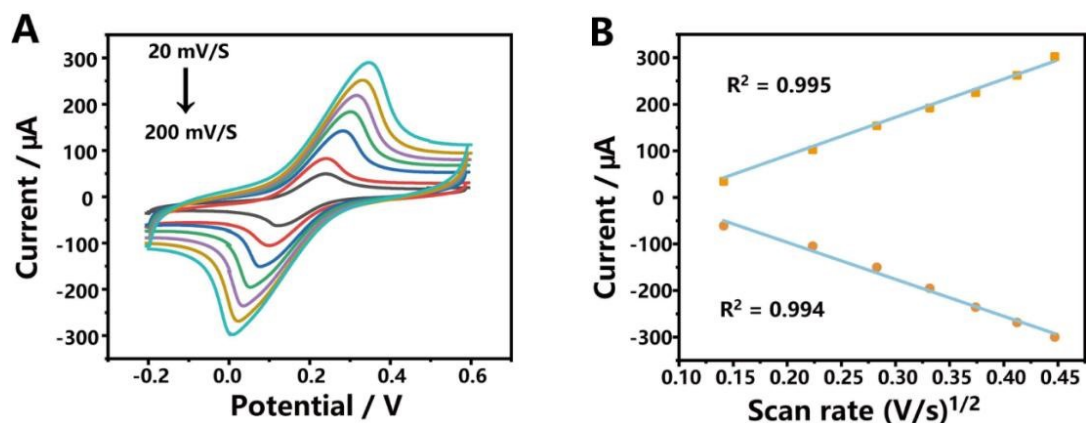
$$\text{Recovery Rate (\%)} = \text{Test Concentration} / \text{Added Concentration} \times 100\%.$$



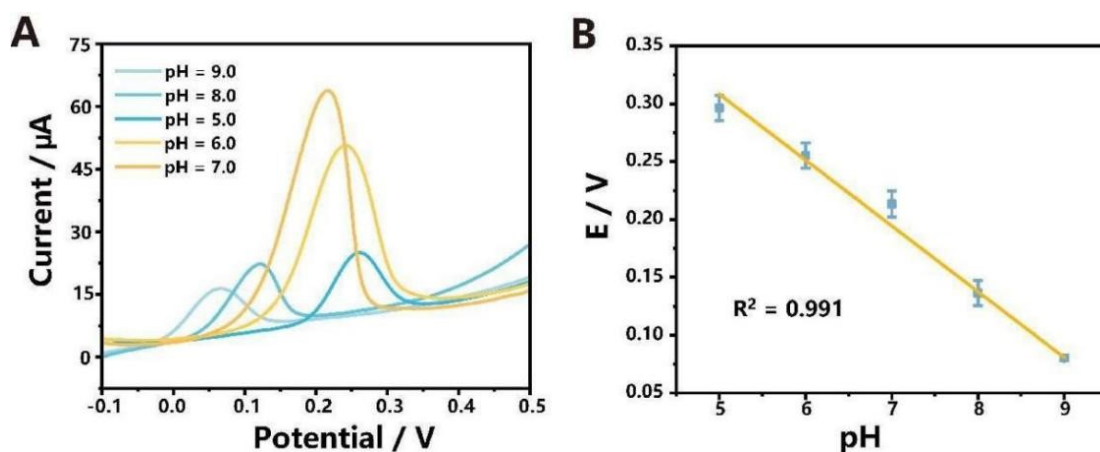
**Fig. S1** Characterization of AuNPs/LIG electrodes: (A, B) TEM image; (C, D) SEM images.



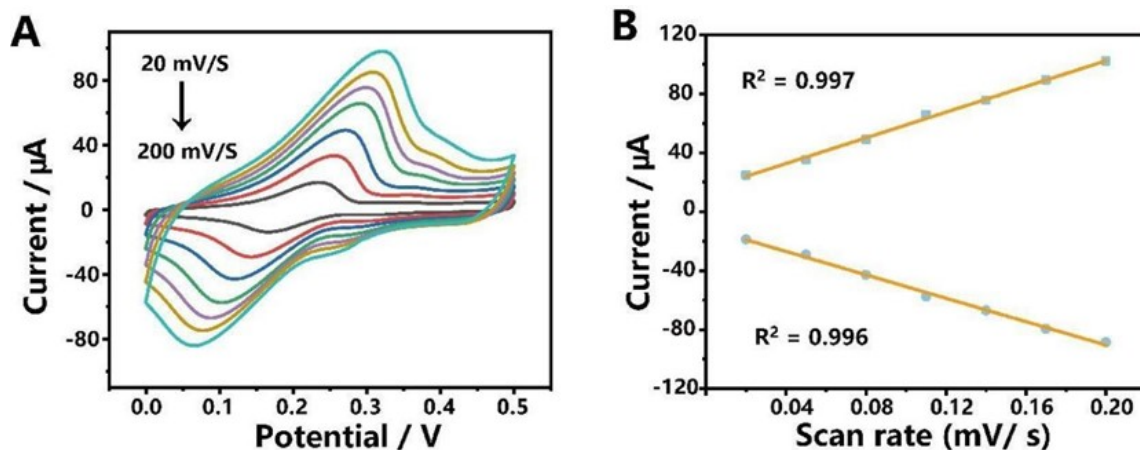
**Fig. S2** SEM images of AuNPs/LIG fabricated with different laser parameters: (A) 2.0 W; (B) 2.5 W; (C) 3.0 W. SEM images of secondary laser engraving at different speeds: (D) 100 mm·s<sup>-1</sup>; (E) 150 mm·s<sup>-1</sup>; (F) 200 mm·s<sup>-1</sup>.



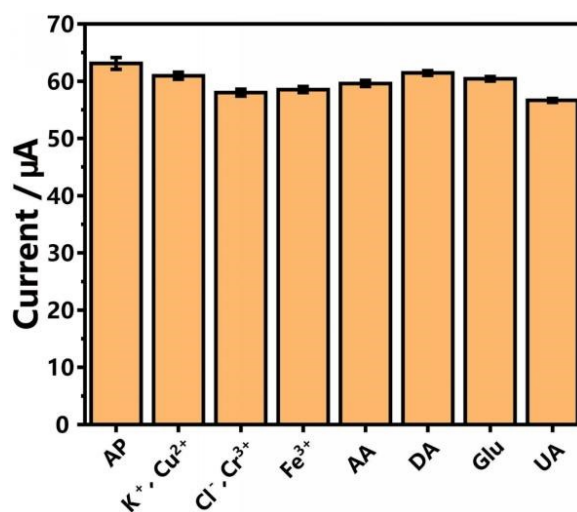
**Fig. S3** (A) Cyclic voltammograms (CVs) of the AuNPs/LIG electrode in a solution containing 0.1 mmol/L KCl and 5 mM  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  at different scan rates: 20, 50, 80, 110, 140, 170, 200 mV/s; (B) Plot of peak current versus the square root of scan rate.



**Fig. S4** (A) Cyclic voltammograms (CVs) of the AuNPs/LIG electrode in a PBS buffer solution (pH=7.0) containing 0.1 mmol/L paracetamol at different scan rates: 20, 50, 80, 110, 140, 170, 200 mV/s; (B) Plot of pH versus potential.



**Fig. S5** (A) Cyclic voltammograms (CVs) of the AuNPs/LIG electrochemical sensor in a PBS buffer solution (0.01 mol/L, pH=7.0) containing 1 mmol/L paracetamol at different scan rates: 20, 50, 80, 110, 140, 170, 200 mV/s; (B) Plot of peak current versus scan rate.



**Fig. S6** Effect of 10 mM organic or inorganic ions on the current value of AuNPs/LIG response to paracetamol. AP- paracetamol; AA- Ascorbic acid; DA- Dopamine; UA- Uric acid.

**Table S1** Comparison between reported methods and our method for detecting paracetamol.

Electrode	Detection object	Linear range	Detection limit	Application scenario	Reference
S-CTFs@NiCo <sub>2</sub> O <sub>4</sub>	PA	2 ~ 360 $\mu$ M	0.18 $\mu$ M	Actual water sample	[12]
	4-PA		0.35 $\mu$ M		
NH <sub>2</sub> -UiO-66/WC	PA	1 $\mu$ M ~ 150 $\mu$ M	0.17 $\mu$ M	Actual water sample	[20]
	PAP		0.35 $\mu$ M		
GMA	PA	0.1 $\mu$ M ~ 150 $\mu$ M	0.057 $\mu$ M	tablet	[41]
N-GSEC	PA	0.1 ~ 250 $\mu$ M	0.01 $\mu$ M	-	[15]
	4-PA		0.025 $\mu$ M		
AuNPs/LIG	PA	0.1 $\mu$ M ~ 100 $\mu$ M	0.065 $\mu$ M	Tab water and tablet	This study

**Table S2** Comparison with other laser synthesis methods.

Electrode	Detection object	Linear range	Detection limit	Application scenario	Reference
LIG	Paraquat	0.5 - 35 $\mu$ M	0.54 $\mu$ M	Actual water sample	[42]
LIG/PtNPs	glucose	300 nM - 2.1 mM	4.622 $\mu$ A/mM	Sweat	[43]
LIG/Ag NPs	H <sub>2</sub> O <sub>2</sub>	0.1 - 10 mM	7.9 $\mu$ M	Whole and skimmed milk	[44]
Co@3D NPC	clozapinealben	0.15 - 20 $\mu$ M	2.8 nM	Soothing tea	[45]



	dazole	0.1 - 7 $\mu$ M	2.5 nM	and pure milk	
AuNPs/LIG	PA	0.1 $\mu$ M - 100 $\mu$ M	0.065 $\mu$ M	Tab water and tablet	This study

**Table S3.** Detection of paracetamol in actual samples

Sample	Spiked ( $\mu$ M)	Found ( $\mu$ M)	HPLC ( $\mu$ M)	Recovery rate (%)
Tablet	10	10.68	12.78	106.8%
	20	20.12	22.89	100.6%
	50	52.33	52.27	104.66%
	80	82.16	81.58	102.7%
Tap water	10	9.78	10.77	97.8%
	20	19.25	22.17	96.25%
	50	48.76	49.59	97.52%
	80	78.63	78.32	98.29%