

Supporting Information

A simple turn-off fluorescent chemosensor based on Schiff base structure for ultrafast and highly selective trace detection of Cu²⁺ ions in aqueous solutions

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1.1 Calculation of the limit of detection (LOD) values

The LOD values were derived from fluorescence titration experiments based on a plot of fluorescence intensity and iron nitrate concentration:

$$\text{LOD} = 3\sigma/k$$

Where σ is the standard deviation of the blank sample and k is the slope of the line of best fit.¹

1.2 Cytotoxicity experiments

Living HepG-2 cells were provided by the School of Chemical and Pharmaceutical Engineering, Jilin Institute of Chemical Engineering. Inoculate cells overnight into a 96 well cell culture plate supplemented with 10% FBS (fetal bovine serum) in DMEM at 37 °C and 5% CO₂ atmosphere. Various concentrations (0, 10, 20, 30, 40, 50 μM) of the probe DHP were added to the cell culture plate after the cells were washed with phosphate-buffered saline (PBS) three times. The cells were incubated overnight at 37 °C under a 5% CO₂ atmosphere. After incubation, the original medium was exchanged with new 100 μL 10 % FBS (fetal calf serum), followed by adding 10 μL MTT (0.5 mg/mL). After 4 h, the medium was removed, and 200 μL DMSO was added to each well. The absorbance at 570 nm was measured with a Spectramax microwell plate reader, and the cell viability towards the HepG-2 cells line were measured using the equation:²

$$\text{Cell viability (\%)} = \text{Mean absorbance (Treated cell)} / \text{Mean absorbance (Control cell)}$$

1.3 Cell fluorescence imaging

During cell imaging experiments, the cells were divided into diverse groups and imaged after different treatments. HepG-2 cells were fixed in 24-well plates, washed with PBS, and then incubated in the dark for 10 min with the addition of MTT. The probes DHP and different concentrations of Cu^{2+} ions (0, 20, 40, 60, 80, 100 μM) were added and incubated for 30 min, respectively. The cells of each well were washed with 3 times PBS buffer after each step. The cells were eventually fixed on a circular slide and imaged by confocal electron fluorescence microscopy.^{3,4}

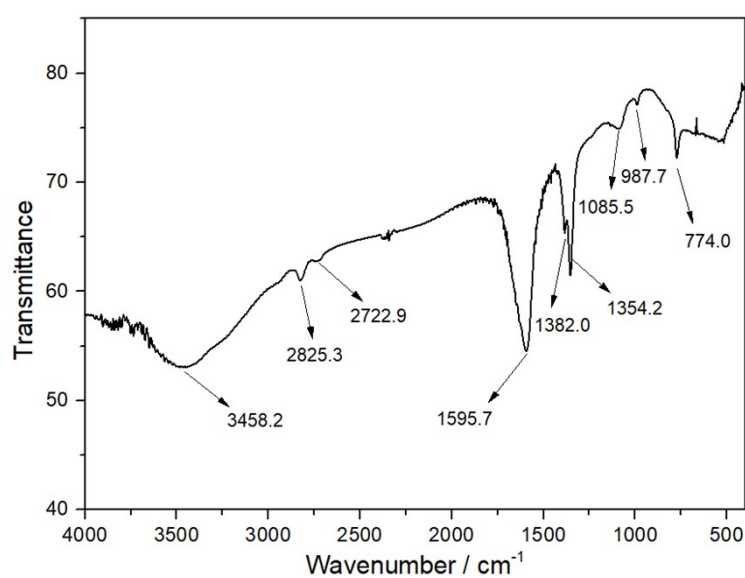


Fig. S1 IR spectrum of DHP

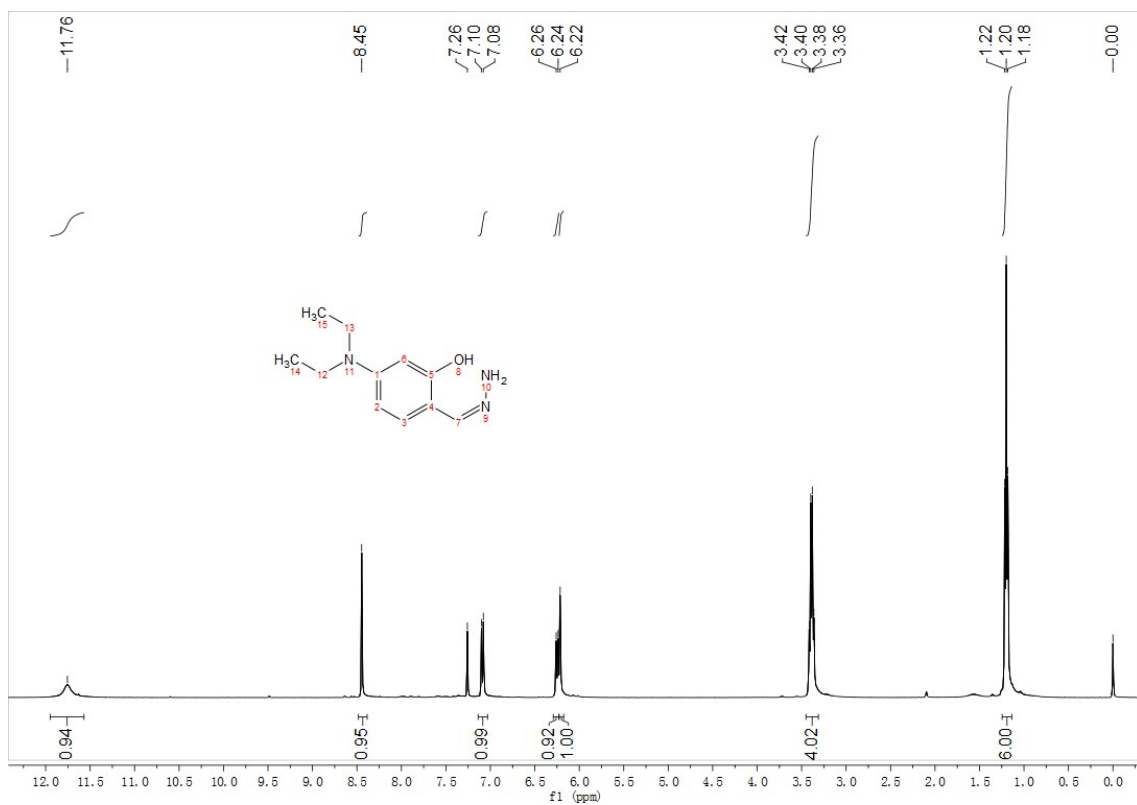


Fig. S2 ¹H NMR spectrum of DHP

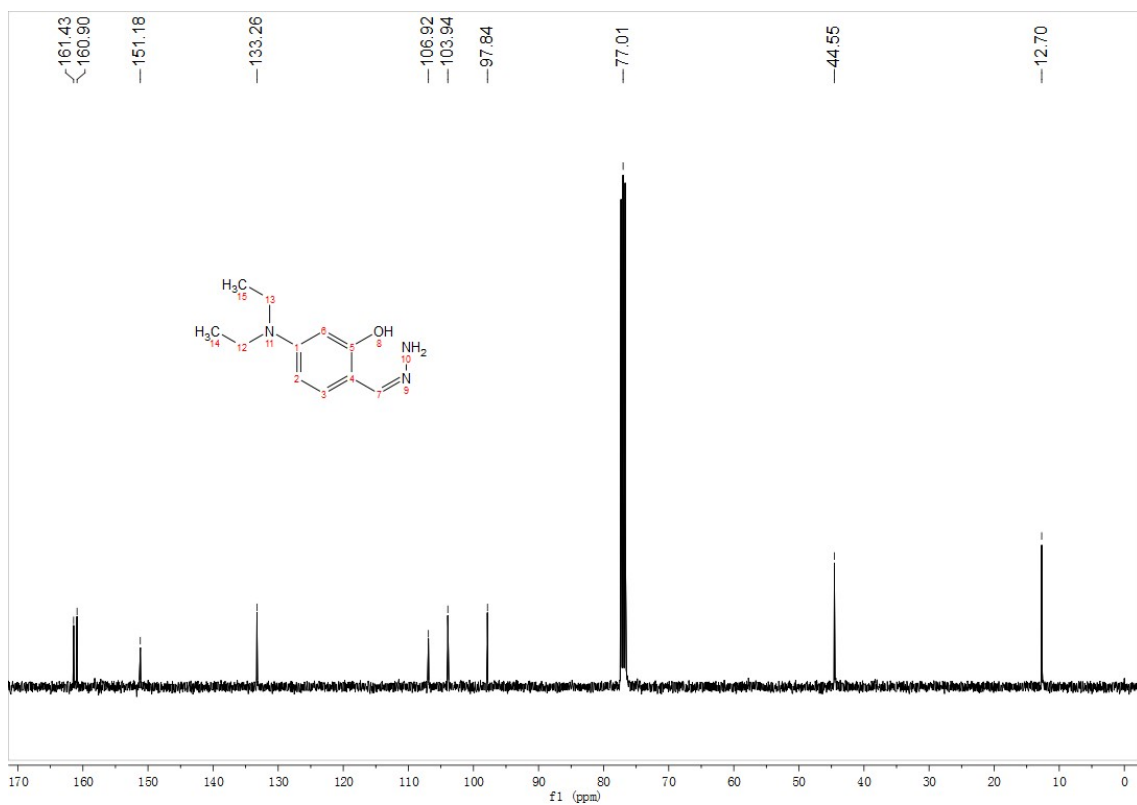


Fig. S3 ¹³C NMR spectrum of DHP

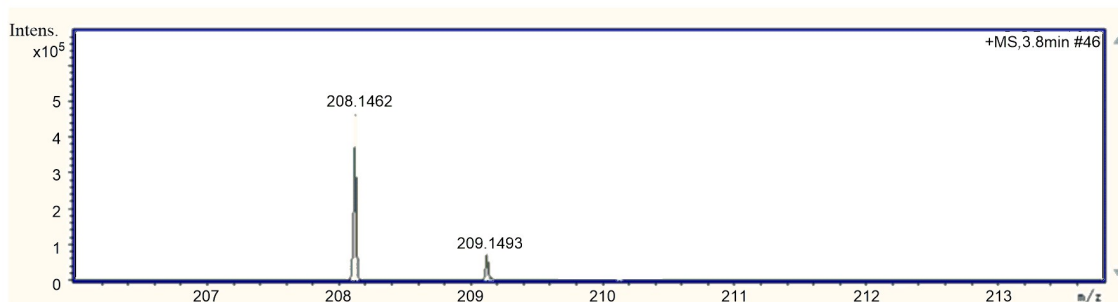


Fig. S4 HR-MS spectrum of DHP

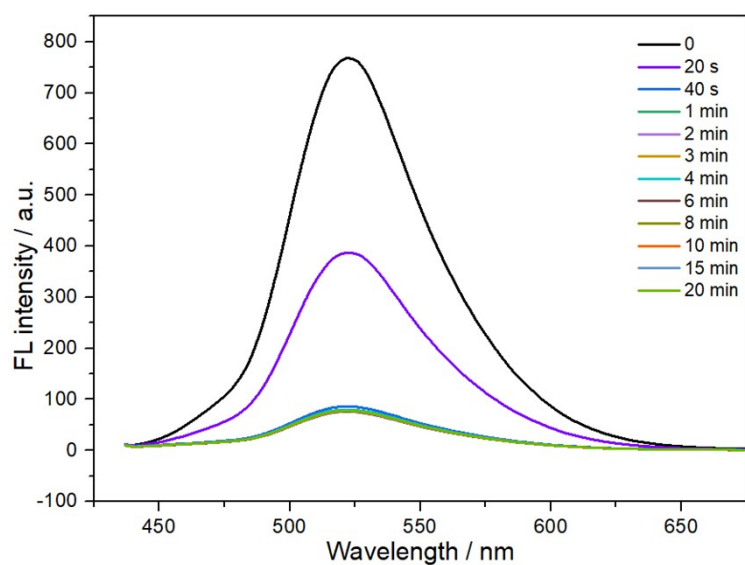


Fig. S5 Response time and stability of probe DHP (10 μ M). Fluorescence intensity of DHP-Cu²⁺ (10 μ M), EtOH/H₂O (1:1, v/v)

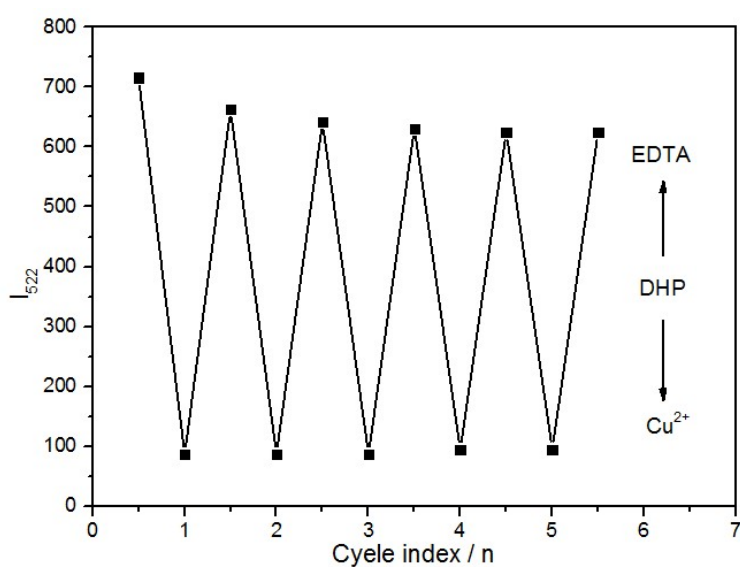


Fig. S6. The reversibility studies of DHP-Cu²⁺ with EDTA.

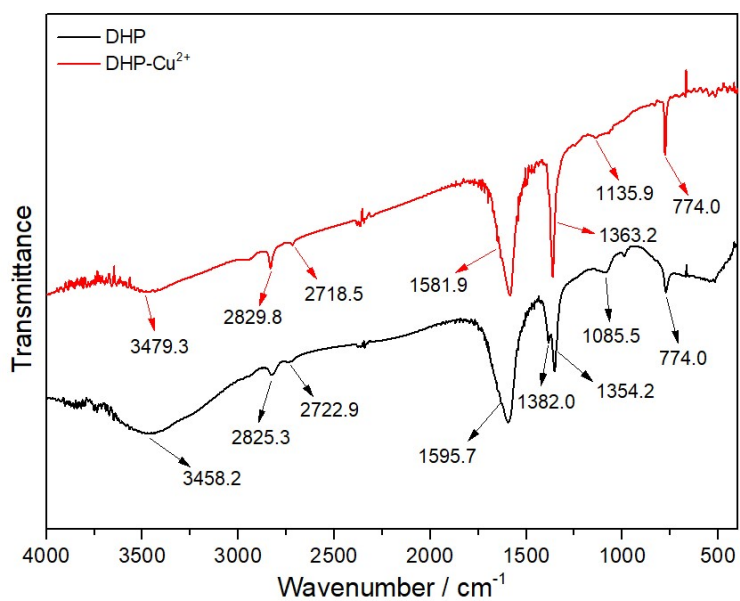


Fig. S7 IR spectrum of DHP and DHP- Cu^{2+}

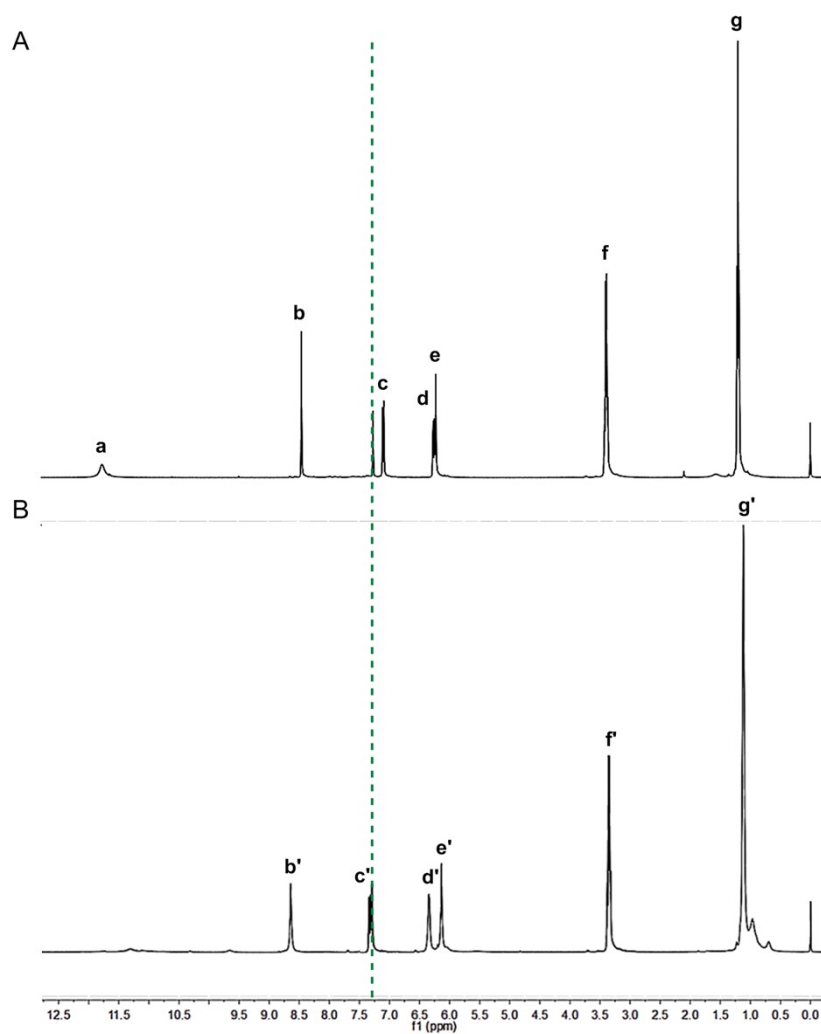


Fig. S8 ^1H NMR of DHP and DHP- Cu^{2+} in CDCl_3 (A) only DHP, (B) DHP- Cu^{2+}

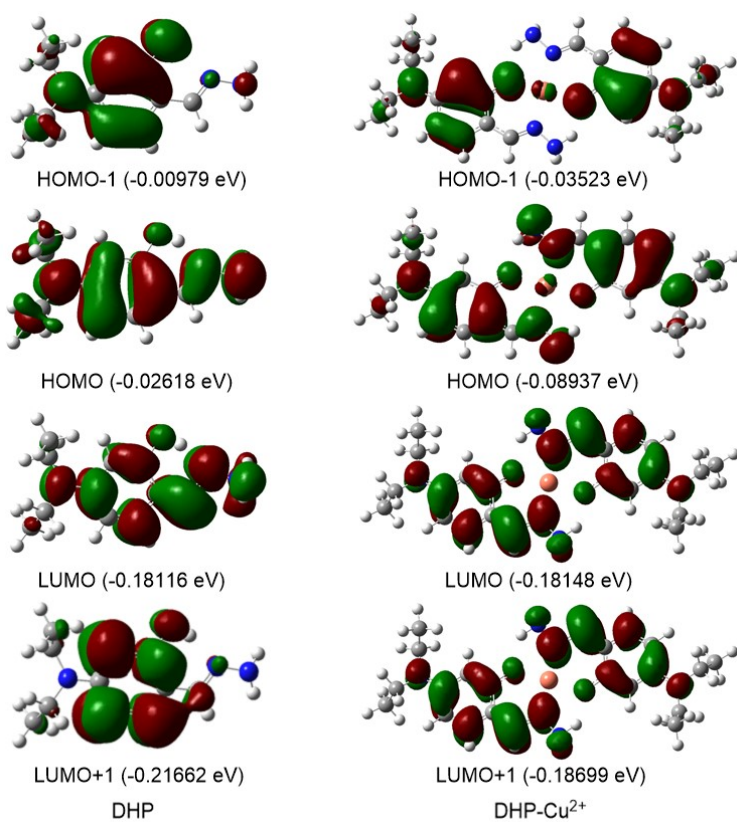


Fig. S9 Optimized molecular configuration and frontier orbitals of DHP and DHP-Cu²⁺.

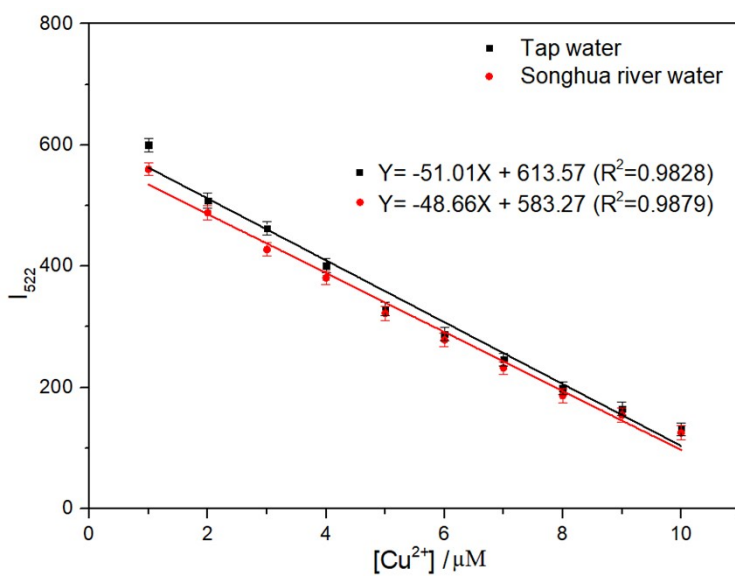


Fig. S10 Linear curves of fluorescence response of probe DHP to Cu²⁺ in tap water and river water.

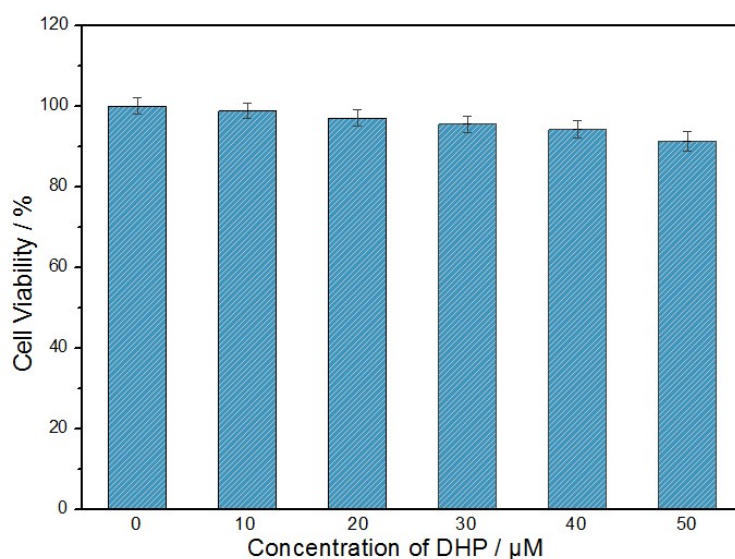


Fig. S11 Cell viability graph of probe DHP using HepG-2 cells by MTT assay after 24 h.

Table S1 Orbital energy differential of DHP and DHP-Cu²⁺.

Compound	$\Delta E_{H \rightarrow L}$ (A.U.)	$\Delta E_{H \rightarrow L+1}$ (A.U.)	$\Delta E_{H-1 \rightarrow L}$ (A.U.)
DHP	0.15498	0.19044	0.17137
DHP-Cu ²⁺	0.09211	0.09762	0.14625

Reference

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