Supplementary Material

Probe-mediated fluorescent biosensor for MC-LR detection using exonuclease III as a signal amplifier

Yuyan Wang^a, Ying Zeng^a, Xiaoya Ren^a, Jun Qiu^b, Jiafeng Pan^{a*}, Fei Yang^{a,*}

^a Hunan Province Key Laboratory of Typical Environmental Pollution and Health Hazards, School of Public Health, Hengyang Medical School, University of South China, Hengyang 421001, China.

^b Hunan Children's Hospital, Hunan 410000, China.

*Corresponding author.

E-mail addresses: jiafengpan0928@163.com (J. Pan), yangfeilong@126.com (F. Yang).

Name	Sequence (from 5' to 3')
Aptamer	GGC GCC AAA CAG GAC CAC CAT GAC AAT TAC CCA TAC
	CAC CTC ATT ATG CCC CAT CTC CGC TTT TTT
Blocker	GCG GAG ATG GGG CAT TTT TTT (21nt)
	GCG GAG ATG GGG CAT AAT TTT TT (23nt)
	GCG GAG ATG GGG CAT AAT GTT TTT T (25nt)
	GCG GAG ATG GGG CAT AAT GAG TTT TTT (27nt)
Hairpin	FAM-AGA CTA GAC CGG AAC GAC GGT CTA GTC <u>T</u> (BHQ1)-ATG
probe	CCC CAT CTC CGC

 Table S1 Sequences of Oligonucleotides Used in the Experiments.

PAGE procedure

Natural polyacrylamide gel electrophoresis was performed by electrophoresis (15% PAGE). 3.8 mL of ultrapure water, 5 mL of 30% Acr-bis (29:1), 1 mL of 10 × TBE buffer, 100 μ L of APS (10%), and 4 μ L of TEMED were mixed and polymerized at 37 °C for 1 h. In addition, the loading sample was prepared by mixing 10 μ L the resulting solution, 2 μ L 100 × SYBR Green solution, and 2 μ L 6 × loading buffer. Then, the electrophoresis experiment was carried out at 45 V for 30 min and turned to 90 V for 90 min in 1× TBE buffer (pH=8.0). Finally, the PAGE gel was scanned by the gel image analysis system (Bio-Rad, USA).



Fig. S1 Effect of A: B ratio on fluorescence of sensor. Fix the concentration of B at 100 nM. Error bars = \pm SD, n = 3.



Fig. S2 Fluorescence methods. The mixture was composed of MC-LR, MC-RR, MC-YR, MC-LA, Cu^{2+} , Cd^{2+} , Pb^{2+} , and Mn^{2+} . Ns: no significance. Error bars = \pm SD, n = 3.

Method	Dynamic range	LOD (ng/L)	Reference	
	(µg/L)			
Surface-enhanced Raman spectroscopy	1.56 - 50	290	1	
(SERS) -based sensor				
Electrochemical aptamer-based sensor	3×10 ⁻² - 1	9.2	2	
Conner nanoclusters based fluorescent	1×10-2 1×103	18	3	
nuclease and the second s	1~10 - 1~10	-1. 0	5	
A silane carbon dots based fluorescent	110 3 2 20	0.6	4	
	1×10 ⁻³ - 3.20	0.6	4	
enzyme-linked immunoassay				
Dual-modal split-type immunosensor	5×10 ⁻⁵ - 5	0.03	5	
SEDS based antosensor	1×10-2 2×102	2	6	
SERS-based aplasensor	1410 - 2410	2	0	
Signal-off ECL sensing model	1×10 ⁻³ - 2×10 ²	0.2	7	
	5 10 5 1 102	0.01	0	
Au/CeO ₂ /g-C ₃ N ₄ -based	$5 \times 10^{-5} - 1 \times 10^{2}$	0.01	8	
photoelectrochemical sensors				
Raman spectroscopic dual-modal	0.1 - 50	0.5	9	
aptasensor				
Exonuclease III-assisted amplification	1×10 ⁻³ - 10	0.37	This work	
fluorescent aptasensor				

Table S2 Comparison of detection sensitivity between our MC-LR detection aptasensor

 and some previously reported methods.

References

- 1 B. Huo, L. Xia, Z. Gao, G. Li, Y. Hu, Anal. Chem, 2022, 94, 11889-11897.
- V. Vogiazi, A. A. de la Cruz, E. A. Varughese, W. R. Heineman, R. J. White, D. D. Dionysiou, ACS EST Engg, 2021, 1, 1597-1605.
- Y. Zhang, Y. Lai, X. Teng, S. Pu, Z. Yang, P. Pang, H. Wang, C. Yang, W. Yang,
 C. J. Barrow, Anal. Methods, 2020, 12, 1752-1758.
- Z. L. Xu, S. L. Ye, L. Luo, X. Hua, J. X. Lai, X. P. Cai, Q. W. Liang, H. T. Lei, Y. M. Sun, Y. Chen, X. Shen, Sci. Total Environ, 2020, 708, 134614.
- 5 J. Wei, W. Chang, A. Qileng, W. Liu, Y. Zhang, S. Rong, H. Lei, Y. Liu, Anal. Chem, 2018, 90, 9606-9613.
- 6 D. He, Z. Wu, B. Cui, Z. Jin, Food Chem, 2019, **278**, 197-202.
- 7 M. Li, H. Lin, S. K. Paidi, N. Mesyngier, S. Preheim, I. Barman, ACS sens, 2020,
 5, 1419-1426.
- 8 X. Ouyang, L. Tang, C. Feng, B. Peng, Y. Liu, X. Ren, X. Zhu, J. Tan, X. Hu, Biosens. Bioelectron, 2020, 164, 112328.
- 9 G. Zhao, Y. Du, N. Zhang, Y. Li, G. Bai, H. Ma, D. Wu, W. Cao, Q. Wei, Anal. Chem, 2023, 95, 8487-8495.