

Supporting Information

Beyond nanoparticle-based oral drug delivery: transporter-mediated absorption and disease targeting

Hana Cho^{a,†}, Kang Moo Huh^{b,†}, Hyun Ji Cho^a, Bogeon Kim^a, Min Suk Shim^c, Yong-Yeon Cho^{a,d}, Joo Young Lee^{a,d}, Hye Suk Lee^{a,d}, Young Jik Kwon^e, Han Chang Kang^{a,d,*}

^a Department of Pharmacy, College of Pharmacy, The Catholic University of Korea, Bucheon, 14662, Republic of Korea

^b Department of Polymer Science and Engineering & Materials Science and Engineering, Chungnam National University, Daejeon 34134, Republic of Korea

^c Division of Bioengineering, Incheon National University, Incheon 22012, Republic of Korea

^d Regulated Cell Death (RCD) Control·Material Research Institute, The Catholic University of Korea, Bucheon, 14662, Republic of Korea

^e Department of Pharmaceutical Sciences, University of California, Irvine, CA 92697, USA

[†] HC and KMH contributed equally to this work.

* Corresponding author (HCK): hckang@catholic.ac.kr

Supplemental Table 1. A) Quantitative protein expression levels and B) relative mRNA expression levels of transporters/receptors in the human intestine^{1,2}

| A) Quantitative protein expression levels of transporters (fmol/mg membrane protein) | | | | |
|---|-----------------|----------------|--------------|--------------|
| Transporter | Duodenum | Jejunum | Ileum | Colon |
| ABCB1 | 371 | 657 | 796 | 499 |
| ABCC3 | 1115 | 577 | 711 | 2327 |
| ASBT | 35 | 196 | 1057 | 36 |
| OATP2B1 | 436 | 511 | 411 | 667 |
| OCT1 | 736 | 655 | 728 | 613 |
| OCT3 | 78 | 58 | 64 | 91 |
| PEPT1 | 2574 | 37220 | 4915 | 286 |
| B) Relative mRNA expression levels of transporters | | | | |
| Transporter | Duodenum | Ileum | Colon | |
| ASBT | 0.09 | 0.42 | < 0.01 | |
| CNT1 | 0.06 | 0.06 | < 0.01 | |
| CNT2 | 0.32 | 0.15 | < 0.01 | |
| ENT2 | < 0.01 | < 0.01 | 0.09 | |
| OATP2B1 | 0.02 | 0.06 | 0.08 | |
| OCTN1 | 0.53 | 1.27 | 1.99 | |
| OCTN2 | 0.09 | 0.10 | 0.82 | |
| PEPT1 | 3.87 | 3.79 | 0.54 | |
| SERT | 0.04 | 0.09 | < 0.01 | |

Supplemental Table 2. The solubility of substrate candidates for transporter/receptor-mediated oral drug delivery

| Transporter | Slightly soluble (> 1 mg/mL) | Very slightly soluble or practically insoluble (< 1 mg/mL) |
|----------------------|--|---|
| ASBT | Cholic acid sodium salt (150 mg/mL), deoxycholic acid sodium salt (330 mg/mL), glycolic acid sodium salt (200 mg/mL), taurocholic acid sodium salt (100 mg/mL) | Benzothiazepine (and derivatives)*, cholic acid (0.175 mg/mL), deoxycholic acid (0.044 mg/mL), naphthol derivates (0.74 mg/mL) |
| ATB ^{0,+/-} | Carnitine (32 mg/mL), propionyl-L-carnitine (10 mg/mL in acidic pH) | Acetylcarnitine (0.355 mg/mL) |
| CNT2 | Adenosine (7 mg/mL), cladribine (4.52 mg/mL), didanosine (27.3 mg/mL), floxuridine (49 mg/mL), formycin B (10 mg/mL), inosine (47 mg/mL), mizoribine (20 mg/mL), ribavirin (20 mg/mL), uridine (50 mg/mL), zidovudine (53 mg/mL) | Clofarabine*, fluoropyrimidine (0.467 mg/mL), guanosine* |
| ENT1 | Adenosine (7 mg/mL), capecitabine (26 mg/mL), cladribine (4.52 mg/mL), cytosine (8 mg/mL), fialuridine (2 mg/mL), fludarabine (9.2 mg/mL), gemcitabine (19 mg/mL), ribavirin (20 mg/mL), uridine (50 mg/mL), thymidine (50 mg/mL), thymine (3.82 mg/mL) | Guanine*, guanosine* |
| GLUT2 | Glucose (133 mg/mL) | - |
| GLUT5 | Fructose (36 mg/mL) | - |
| MCT1 | β-D-Hydroxybutyric acid (25 mg/mL in PBS), γ-hydroxybutyric acid (494 mg/mL), L-lactic acid (100 mg/mL), pyruvic acid (1g/mL), salicylates (2.24 mg/mL), valproic acid (1.3 mg/mL) | Nateglinide (0.0088 mg/mL) |
| OATP2B1 | Aliskiren (100 mg/mL), fexofenadine (2 mg/mL) | Amiodarone (0.72 mg/mL), atorvastatin (0.0204 mg/mL), bosentan (0.01 mg/mL), DHEAS (0.0081 mg/mL), estrone-3-sulphate*, glibenclamide*, talinolol (0.0451 mg/mL), telmisartan*, L-thyroxine (0.105 mg/mL) |
| OCTN1 | Acetylcholine (100 mg/mL), carnitine (32 mg/mL), doxorubicin hydrochloride (10 mg/mL), entecavir (2.4 mg/mL), ergothioneine (50 mg/mL), gabapentin (10 mg/mL), imatinib (200 mg/mL), ipratropium (83 mg/mL), metformin (33 mg/mL), oxaliplatin (6 mg/mL), pregabalin (36 mg/mL), pyrilamine (80 mg/mL), verapamil (25 mg/mL) | Mitoxantrone (0.734 mg/mL), quinidine (0.14 mg/mL), tiotropium (0.0176 mg/mL) |
| OCTN2 | Carnitine (32 mg/mL), cephaloridine (257.7 mg/mL), emetine (100 mg/mL), entecavir (2.4 mg/mL), imatinib (200 mg/mL), ipratropium (83 mg/mL), verapamil (25 mg/mL) | Etoposide*, spironolactone*, tiotropium (0.0176 mg/mL) |
| OSTα/β | Cholic acid sodium salt (150 mg/mL), deoxycholic acid sodium salt (330 mg/mL), glycolic acid sodium salt (200 mg/mL), PGE2 (1.05 mg/mL), taurocholic acid sodium salt (100 mg/mL) | Cholic acid (0.175 mg/mL), deoxycholic acid (0.044 mg/mL), DHEAS (0.0081 mg/mL), digoxin* |
| PAT1 | Betaine (50 mg/mL), L-tryptophan (11.4 mg/mL) | - |
| PCFT | - | Folic acid (0.01 mg/mL), 5-Methyltetrahydrofolate* |
| PEPT1 | 5-Aminolevulinic acid (50 mg/mL), carnosine (125 mg/mL), cephalexin (10 mg/mL), penicillin G (benzylpenicillin) (100 mg/mL), D-Phe-Ala (14.11 mg/mL), valacyclovir (174 mg/mL) | Cefadroxil (0.399 mg/mL), glibenclamide*, nateglinide (0.0088 mg/mL) |
| SGLT1 | Glucose (133 mg/mL), galactose (100 mg/mL) | - |
| SMVT1 | Pantothenic acid (50 mg/mL) | Biotin (0.22 mg/mL), lipoic acid* |
| SVCT1 | L-Ascorbic acid (10 mg/mL) (mg/mL) | - |

* : water insoluble

Supplemental Table 3. The change in the expression level of GI transporters/receptors in the status of disease³⁻²⁵.

| Transporter | Disease | Species/cell line | mRNA | Protein |
|----------------------|-------------------------|-------------------|---------------------------------------|---|
| ASBT | Diabetes | Rat | Ileum ↑ | - |
| | IBD (UC) | Human | Colon (2.8-fold ↓) | - |
| | IBD (CD) | Human | - | 31 % ↓ |
| | Obesity | Rat | Ileum ↑ | Ileum (3.0-fold ↓) |
| | Obstructive cholestasis | Human | Duodenum (4.0-fold ↓) | - |
| FcRn | Infection | IPEC-J2 | | ↑ |
| FRα | Colorectal cancer | Human | | Colorectal Cancer ↑ |
| FRβ | IBD | Human | | Macrophage cc |
| GLUT2 | Diabetes | Human | Duodenum (3-fold ↑) | - |
| GLUT5 | Diabetes | Human (T2DM) | Duodenum (3-fold ↑) | 2.5-fold ↑ |
| | IBD (CD) | Mouse | Small intestine ↓ | |
| Mannose receptor | IBD | Human | | Macrophage ↑ Dendritic cell ↑ |
| MCT1 | Alzheimer disease | APP/PS1 Mouse | - | Small intestine (1.9-fold ↓) |
| | IBD (CD) | Human | Colon (2.0-fold ↓) | Colon (3.9-fold ↓) |
| | IBD (UC) | Human | Colon (3.0-fold ↓) | - |
| OATP2B1 | Cushing syndrome | Human (T2DM) | Duodenum (1.3-fold ↑) | - |
| | IBD (CD) | Human | Ileum (7-fold ↑) | - |
| | IBD (UC) | Human | Ileum (4-fold ↑) Colon (13-fold ↑) | - |
| OCTN2 | IBD (CD) | Human | Ileum (50.0-fold ↓) | - |
| | IBD (UC) | Human | | Colon ↓ |
| OSTα | IBD (UC) | Human | Colon (5.0-fold ↓) | - |
| | Cholestasis | BDL-C57BL/6 | - | Ileum (1.7-fold ↓) |
| OSTβ | Cholestasis | BDL-C57BL/6 | - | Ileum (5.0-fold ↓) |
| | IBD (UC) | Human | Colon (3.4-fold ↓) | - |
| PCFT | Diabetes | Rat | Jejunum (12.3-fold ↑) | - |
| PEPT1 | Cushing syndrome | Caco-2 | - | Membrane (1.8-fold ↑) |
| | Hyperthyroidism | Caco-2 | 4.0-fold ↓ | 3.5-fold ↓ |
| | Hyperthyroidism | Rat | 1.4-fold ↓ | 1.4-fold ↓ |
| | IBD | Human | - | Colon ↑ |
| | Obesity | Caco-2 | - | Membrane (2.2-fold ↑) Intracellular (2.0-fold ↓) |
| SGLT1 | Diabetes | Human | Duodenum (3-fold ↑) | - |
| SMVT | IBD (UC) | Mouse | - | Colon (4.0-fold ↓) |
| SVCT1 | IBD (UC) | Human | 2.94-fold ↓ | - |
| SVCT2 | IBD (CD) | Human | Ileum (7-fold ↑) Colon (8-fold ↑) | - |
| Transferrin receptor | IBD (CD) | Rat | - | Colon (1.4-fold ↑) |

* IBD, inflammatory disease; CD, Crohn's disease; UC, Ulcerative colitis; T2DM, Type 2 diabetes mellitus; BDL, bile duct ligation

References

1. M. Drozdzik and S. Oswald, *Current Medicinal Chemistry*, 2016, **23**, 4468-4489.
2. Y. Meier, J. J. Eloranta, J. Darimont, M. G. Ismair, C. Hiller, M. Fried, G. A. Kullak-Ublick and S. R. Vavricka, *Drug metabolism and Disposition*, 2007, **35**, 590-594.
3. M. Drozdzik, I. Czekawy, S. Oswald and A. Drozdzik, *Pharmacological Reports*, 2020, **72**, 1173-1194.
4. D. Jung, A. Fantin, U. Scheurer, M. Fried and G. Kullak-Ublick, *Gut*, 2004, **53**, 78-84.
5. P. Hruz, C. Zimmermann, H. Gutmann, L. Degen, U. Beuers, L. Terracciano, J. Drewe and C. Beglinger, *Gut*, 2006, **55**, 395-402.
6. F. Annaba, K. Ma, P. Kumar, A. K. Dudeja, R. D. Kineman, B. L. Shneider, S. Saksena, R. K. Gill and W. A. Alrefai, *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 2010, **299**, G898-G906.
7. S. Sundaram, B. Palaniappan, N. Nepal, S. Chaffins, U. Sundaram and S. Arthur, *Cells*, 2019, **8**, 1197.
8. N. Patel, L. Ghali, I. Roitt, L. P. Munoz and R. Bayford, *Nanoscale Advances*, 2021, **3**, 5373-5386.
9. S. H. Lee, J. G. Song and H.-K. Han, *Acta Pharmaceutica Sinica B*, 2022, **12**, 4249-4261.
10. J. Skupsky, S. Sabui, M. Hwang, M. Nakasaki, M. D. Cahalan and H. M. Said, *Cellular and molecular gastroenterology and hepatology*, 2020, **9**, 557-567.
11. S. Pérez-Torras, I. Iglesias, M. Llopis, J. J. Lozano, M. Antolín, F. Guarner and M. Pastor-Anglada, *Journal of Crohn's and Colitis*, 2016, **10**, 850-859.
12. V. S. Subramanian, S. Sabui, G. A. Subramenium, J. S. Marchant and H. M. Said, *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 2018, **315**, G241-G248.
13. Y. Fujita, H. Kojima, H. Hidaka, M. Fujimiya, A. Kashiwagi and R. Kikkawa, *Diabetologia*, 1998, **41**, 1459-1466.
14. J. Dyer, I. Wood, A. Palejwala, A. Ellis and S. Shirazi-Beechey, *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 2002, **282**, G241-G248.
15. P. Liu, C. Gao, H. Chen, C. T. Vong, X. Wu, X. Tang, S. Wang and Y. Wang, *Acta Pharmaceutica Sinica B*, 2021, **11**, 2798-2818.
16. M. Fujiya, Y. Inaba, M. W. Musch, S. Hu, Y. Kohgo and E. B. Chang, *Inflammatory bowel diseases*, 2011, **17**, 907-916.
17. K. A. Wojtal, J. J. Eloranta, P. Hruz, H. Gutmann, J. Drewe, A. Staumann, C. Beglinger, M. Fried, G. A. Kullak-Ublick and S. R. Vavricka, *Drug Metabolism and Disposition*, 2009, **37**, 1871-1877.
18. M. Pyzik, L. K. Kozicky, A. K. Gandhi and R. S. Blumberg, *Nature Reviews Immunology*, 2023, 1-18.
19. T. M. Bui, H. L. Wiesolek and R. Sumagin, *Journal of Leucocyte Biology*, 2020, **108**, 787-799.
20. R. Thibault, P. De Coppet, K. Daly, A. Bourreille, M. Cuff, C. Bonnet, J. F. Mosnier, J. P. Galmiche, S. Shirazi-Beechey and J. P. Segain, *Gastroenterology*, 2007, **133**, 1916-1927.
21. C.-Y. Wang, S. Liu, X.-N. Xie and Z.-R. Tan, *Drug design, development and therapy*, 2017, 3511-3517.
22. S. A. Ingersoll, S. Ayyadurai, M. A. Charania, H. Laroui, Y. Yan and D. Merlin, *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 2012, **302**, G484-G492.
23. G. D'Argenio, M. Calvani, A. Casamassimi, O. Petillo, S. Margarucci, M. Rienzo, I. Peluso, R. Calvani, A. Ciccodicola and N. Caporaso, *The FASEB journal*, 2006, **20**, 2544-2546.
24. S. Basu, C. Liu, X. K. Zhou, R. Nishiguchi, T. Ha, J. Chen, M. Johncilla, R. K. Yantiss, D. C. Montrose and A. J. Dannenberg, *Am J Physiol Gastrointest Liver Physiol*, 2021, **321**, G232-g242.
25. P. Li, Y. Wang, J. Luo, Q. Zeng, M. Wang, M. Bai, H. Zhou, J. Wang and H. Jiang, *Biochem Pharmacol*, 2020, **178**, 114115.