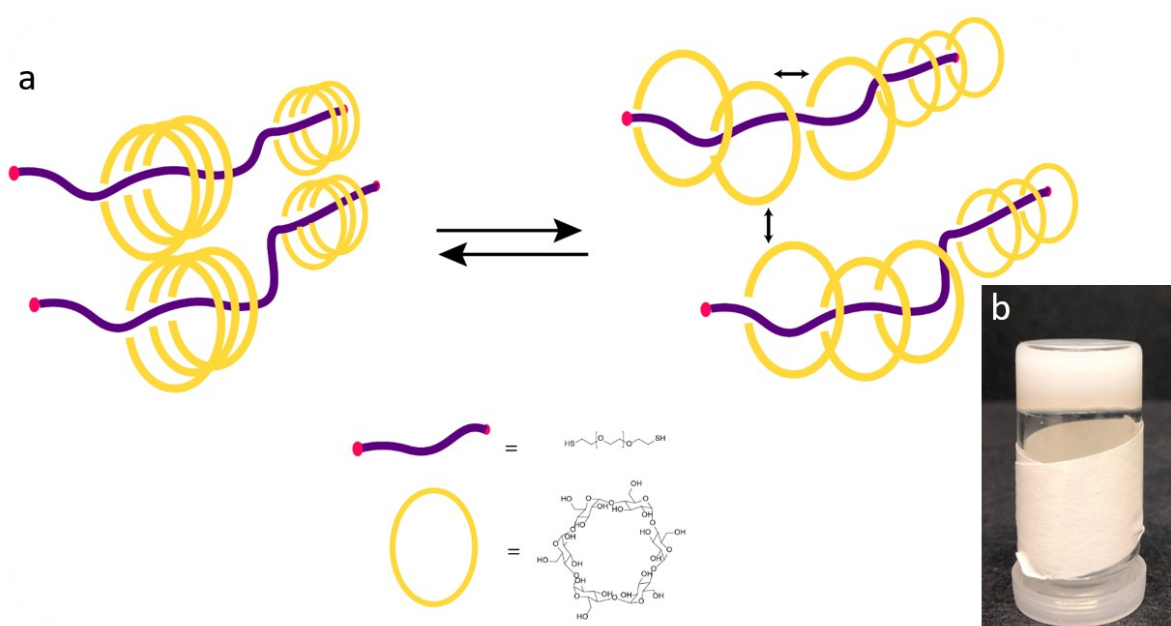


Granular Polyrotaxane Microgels As Injectable Hydrogels for Corneal Tissue Regeneration

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Supplementary Information



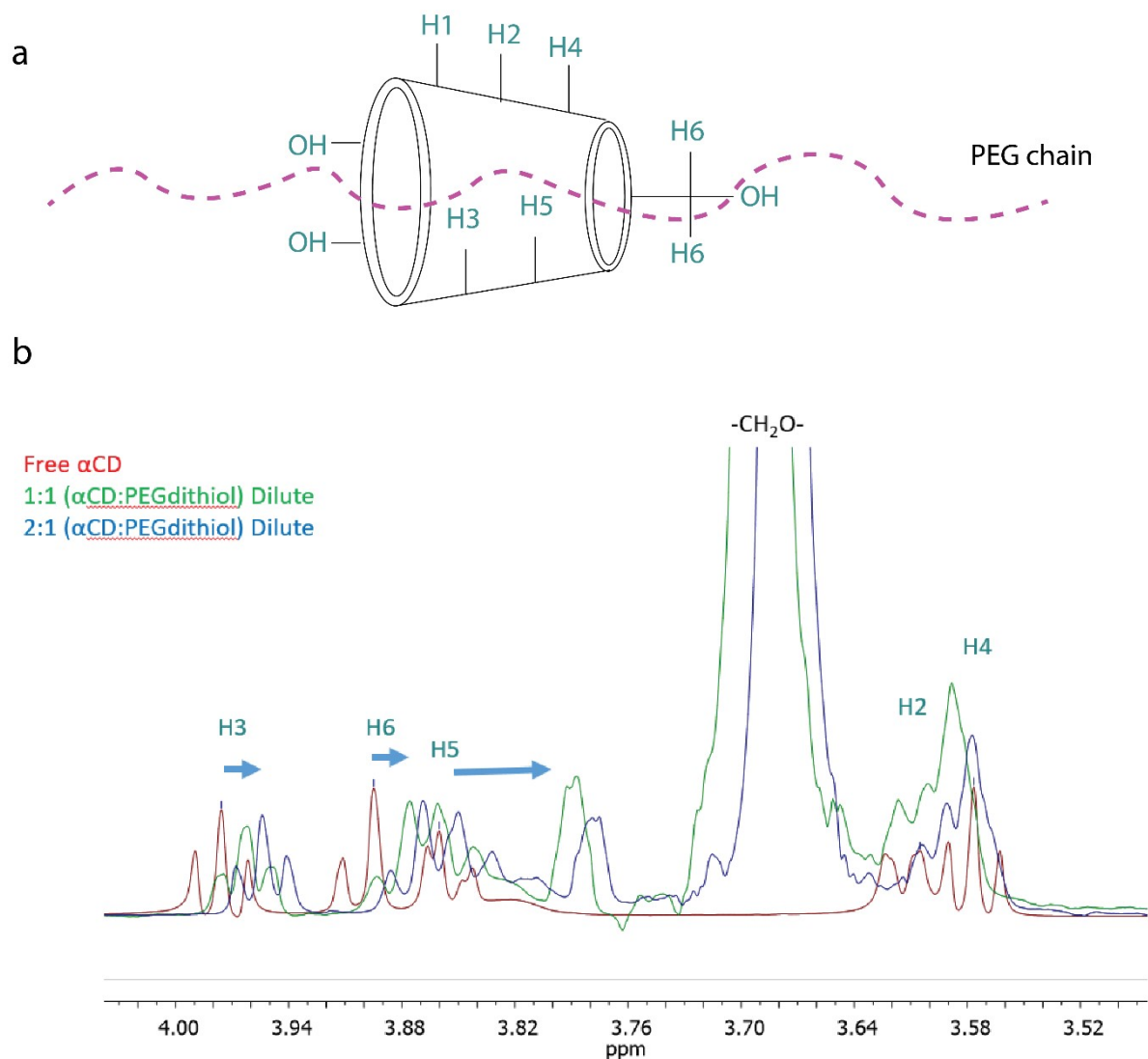


Figure S2. (a) Diagram of inclusion between CD macrocycle and PEG chain. (b) chemical shifts of α CD and PEG in 1:1 and 2:1 (α CD:PEG) ratios.

Table S1. Chemical shifts of protons indicating shifts between H3 and H5 protons indicating inclusion of PEG chain in CD.

		H1	H3	H6	H5	H2	H4
Ratio	Free αCD	5.040	3.976	3.894	3.860	3.614	3.577
1:1	Complex	5.038	3.962	3.876	3.790	3.615	3.588
	$\Delta\delta$	-0.002	-0.014	-0.018	-0.070	0.001	0.011
2:1	Complex	5.027	3.954	3.869	3.775	3.604	3.577
	$\Delta\delta$	-0.013	-0.022	-0.025	-0.085	-0.01	0

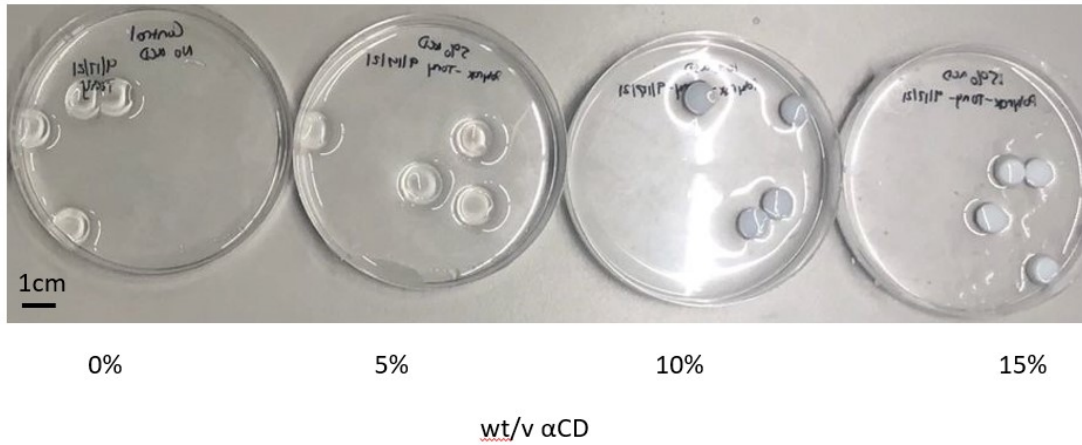


Figure S3. Synthesized bulk hydrogels after 2 days in PBS.

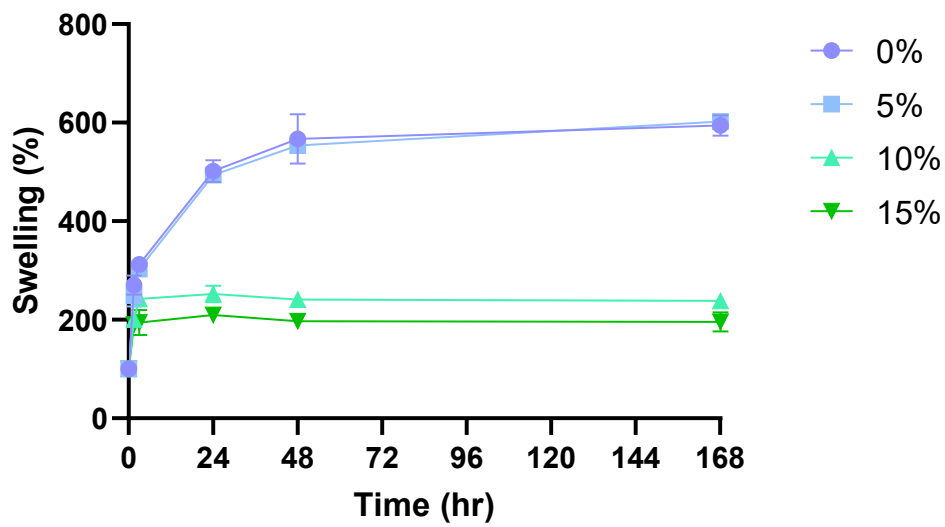


Figure S4. Swelling occurs the most with 0% α CD due to PEG chains being completely free to swell. 15% α CD has limited swelling due to secondary crosslinking between CDs.

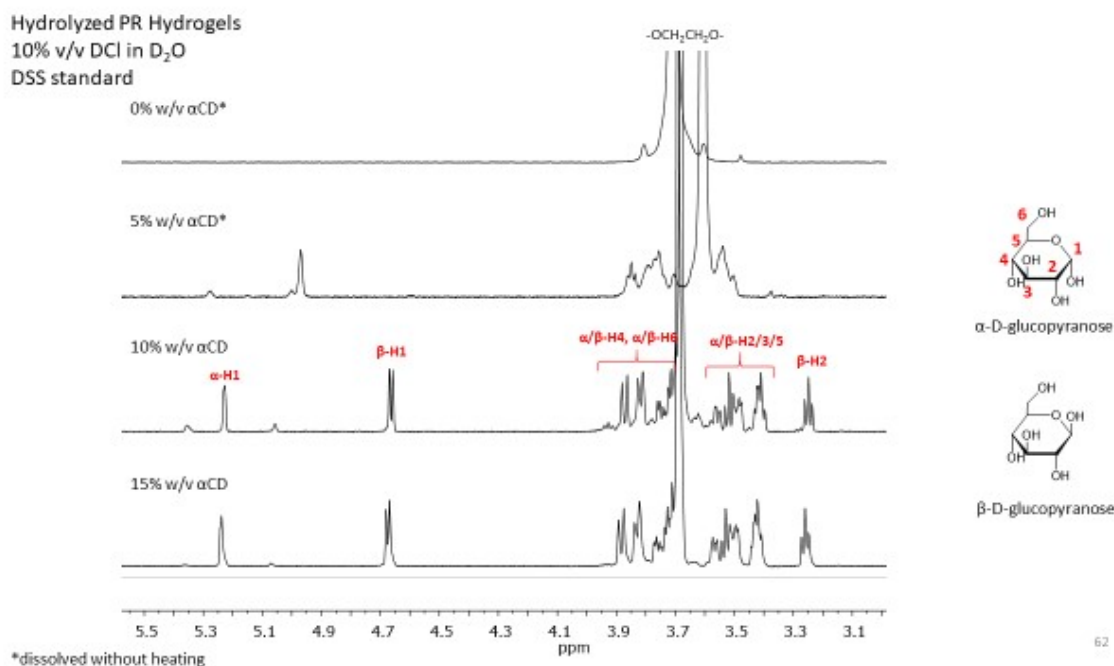


Figure S5. ¹H NMR of degraded polyrotaxane hydrogel to determine the ratio of α CD to PEG in the final hydrogel.

Table S2. Calculated α CDs per PEGdithiol.

Sample	α CD-H1 integration	PEG integration	CD/PEG
0	0	-	NA
5	1	66,7	6
10	1,67	27,9	23
15	1,65	13,2	47

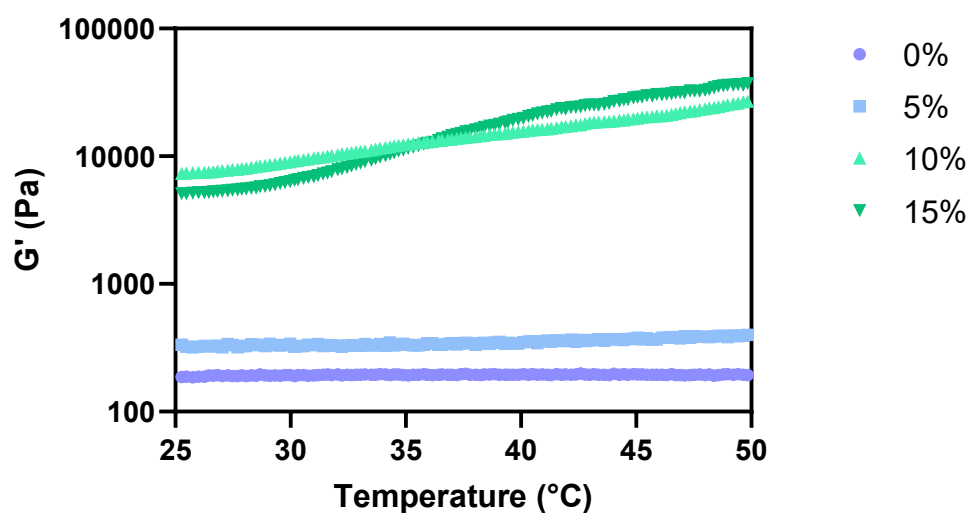


Figure S6. Thermoresponsiveness of bulk polyrotaxane gels. 10 and 15% wt/v α CD show clear increase in G' with increasing T whereas 0, 5% are negligible. Rate of temperature increase: 1°C/min.

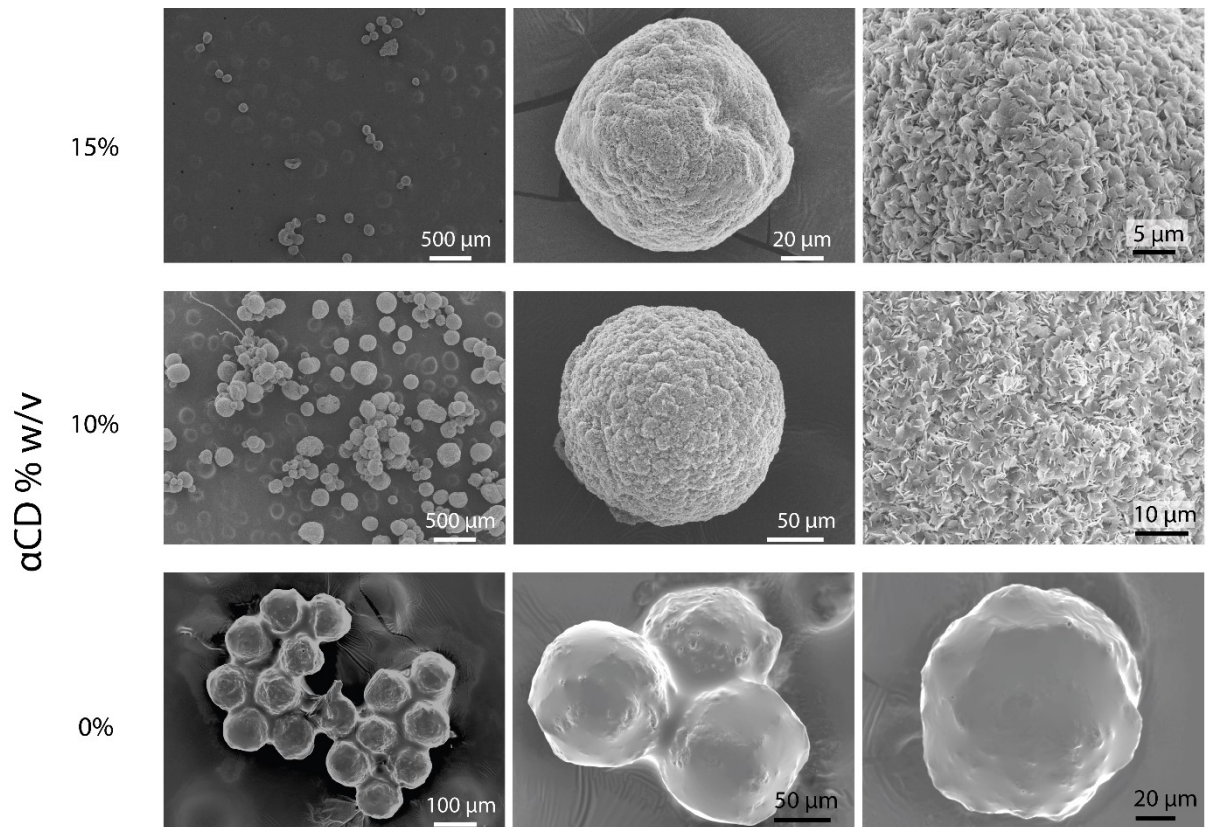


Figure S7. SEM images of 0, 10, 15% wt/v α CD GPR microgels after purification.

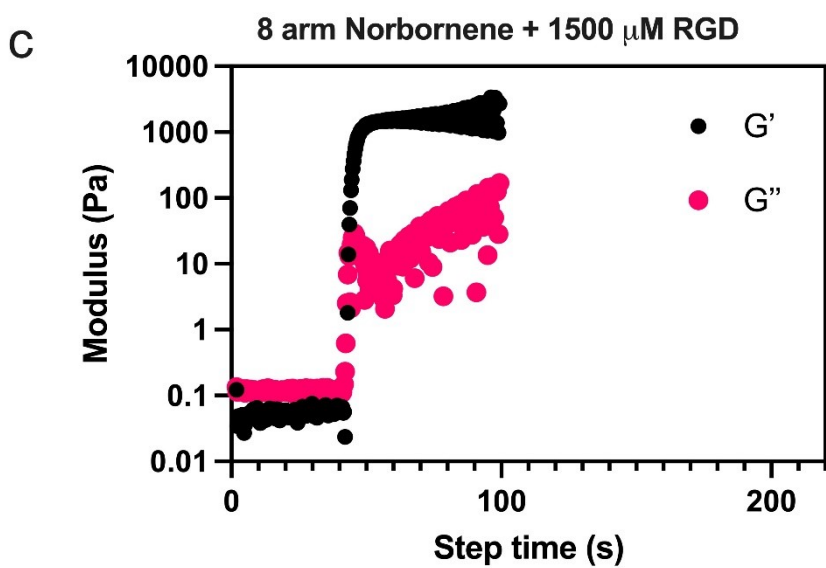
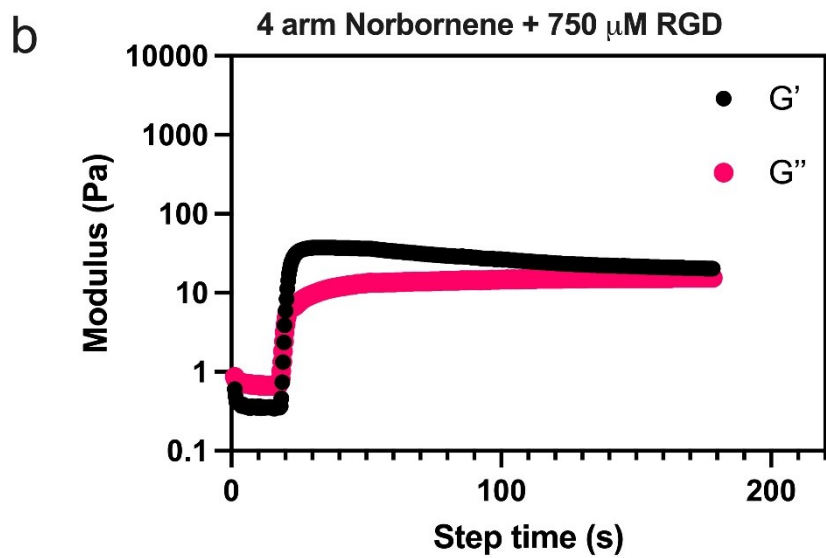
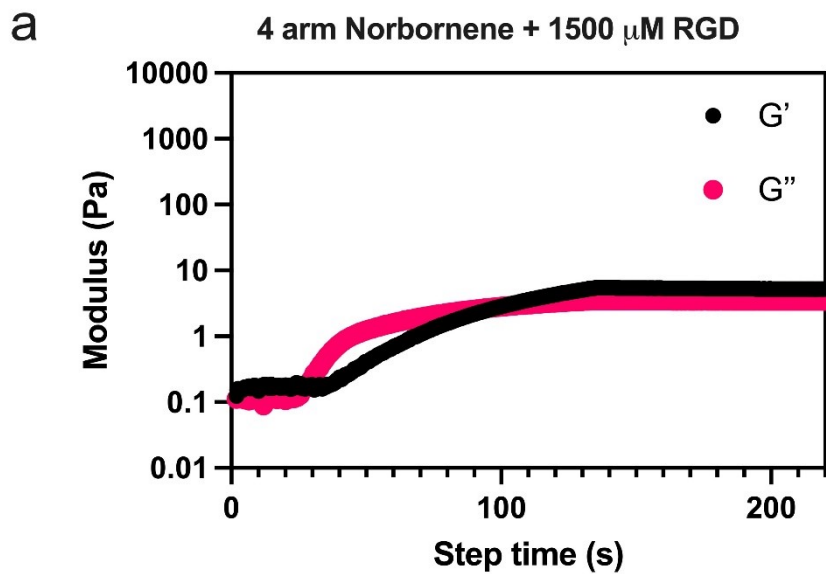


Figure S8. Hydrogel gelation kinetics for different formulations of (15% wt/v α CD) polyrotaxane bulk hydrogels with (a) 4 arm norbornene (1.64 mM) and RGD (1500 μ M) (b) 4 arm norbornene (1.5 mM) and RGD (750 μ M) and (c) 8 arm norbornene (0.273 mM) and RGD (1500 μ M) in MilliQ with 2 mM LAP. The concentration of thiol to norbornene functional groups was kept at 1:1.

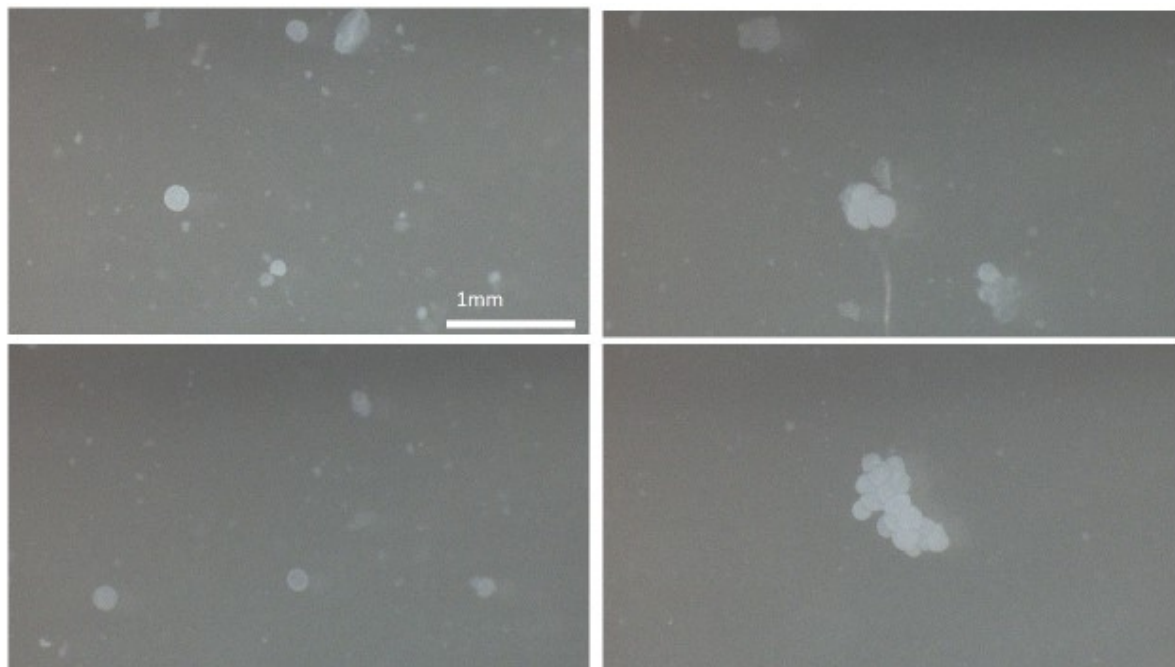


Figure S9. 8 arm RGD GPR microgels in water.

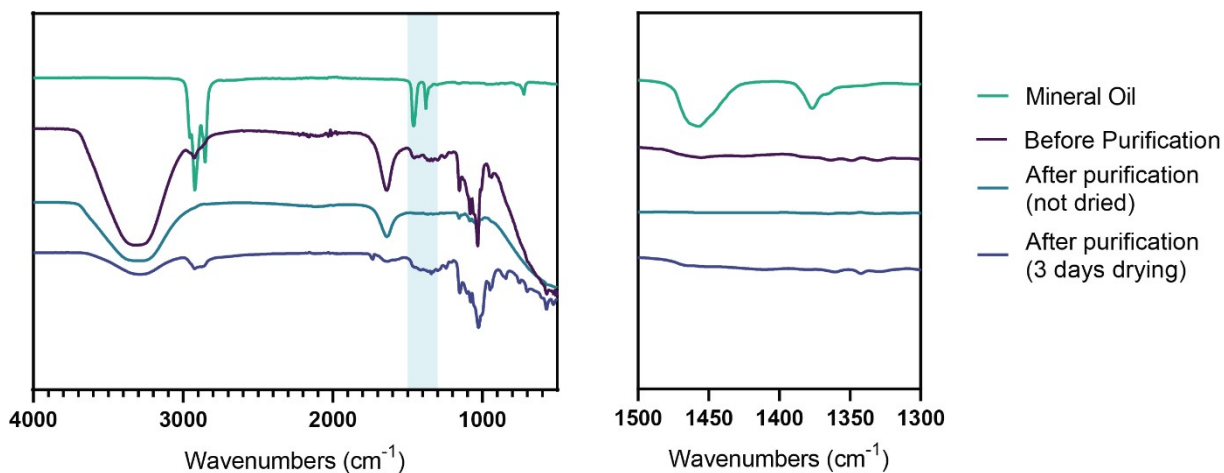


Figure S10. FTIR of purified GPR showcasing oil removal from GPR after synthesis. Residual oil removed after purification.

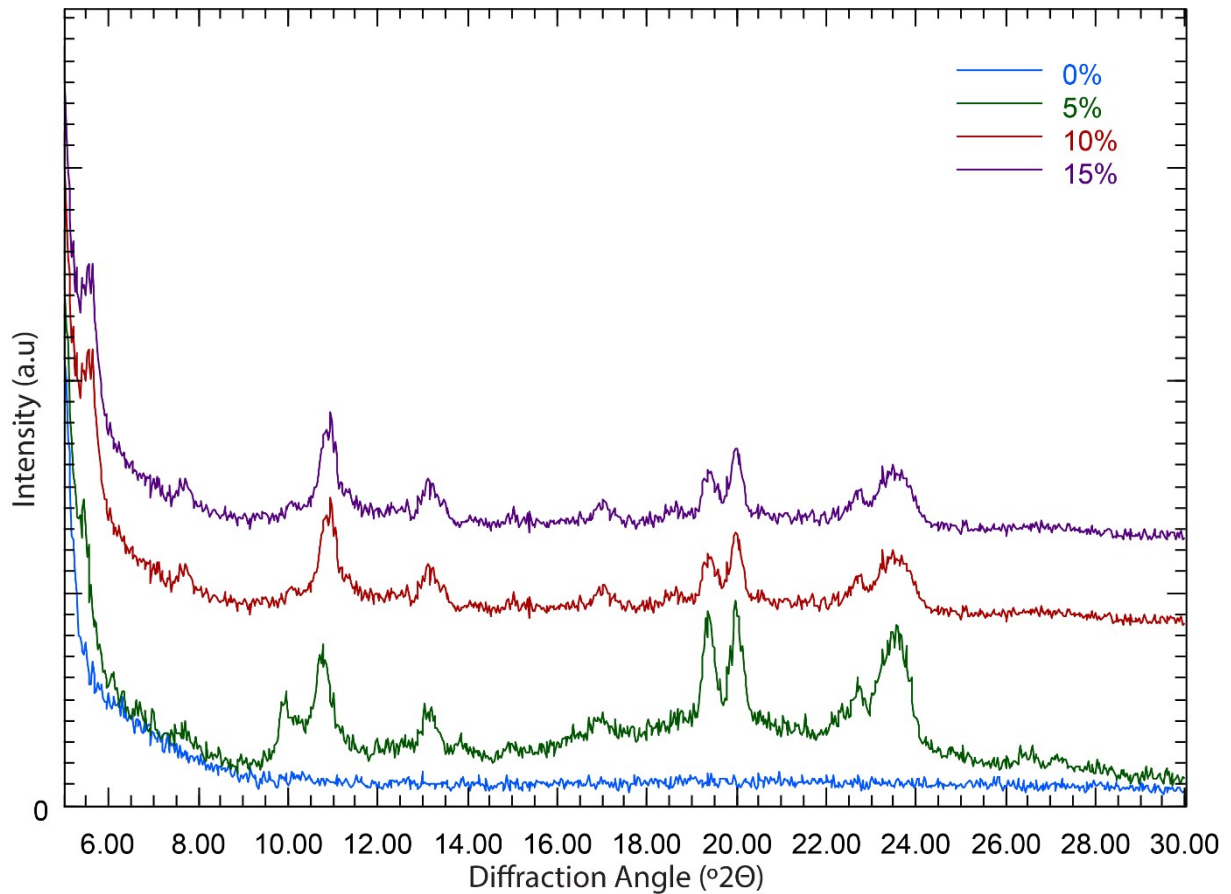


Figure S11. XRD of GPR dried microgels. No diffraction patterns observed for PEG microgels (0% α CD). Diffraction patterns appear with addition of 5 – 15% α CD.

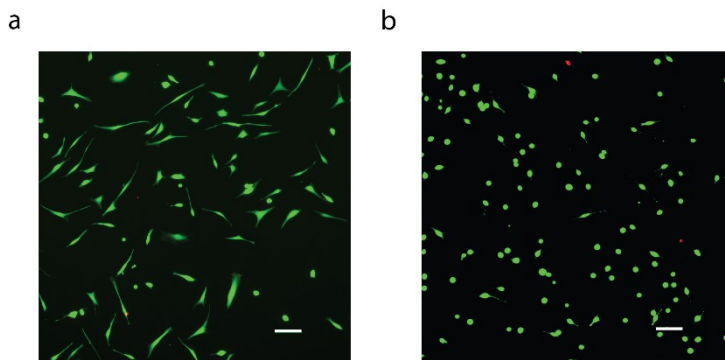


Figure S12. (a) Live/dead images of corneal keratocytes in differentiation media. (b) Morphological changes in human corneal keratocytes after 24 hours exposed to GPR supernatant. Scale bar = 100 μ m.

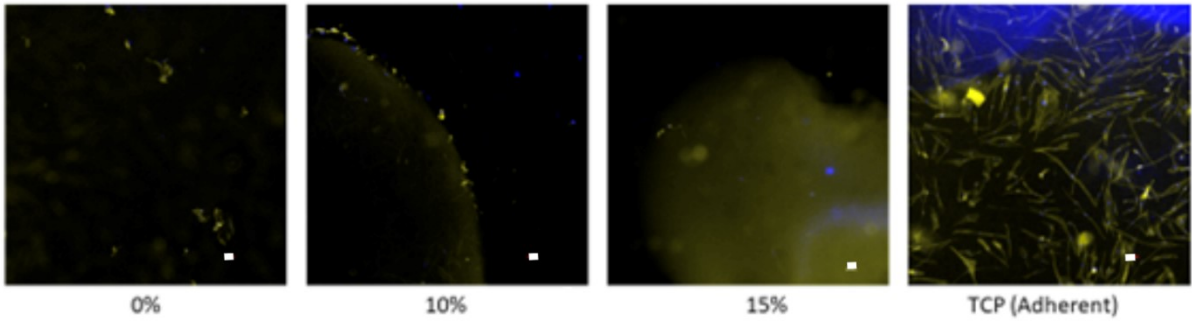


Figure S13. Bulk PR hydrogels after 2 weeks in culture with immortalized human corneal keratocytes indicating little to no adherence of cells to the bulk gel. Scale bar = 100 μ m.

Degradation by Hydrolysis

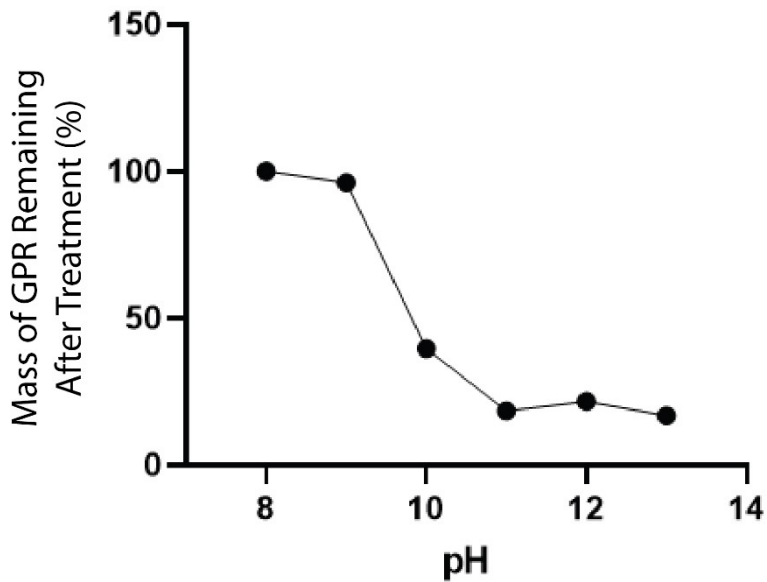


Figure S14. Degradability of GPR through basic hydrolysis monitored by mass loss.

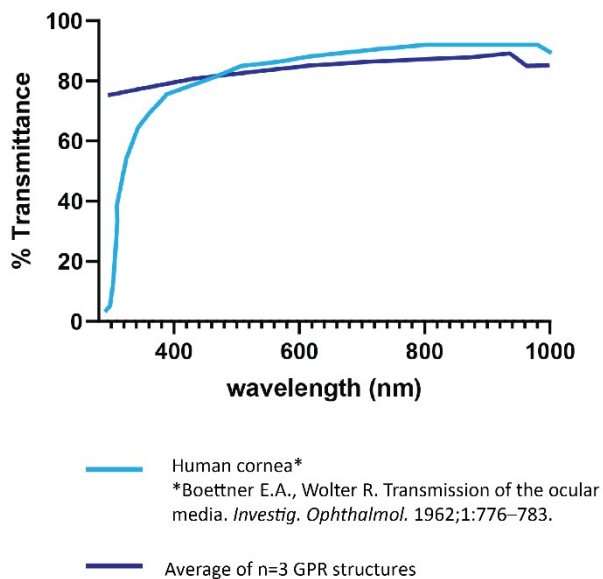


Figure S15. % Transmittance obtained by UV-Vis spectroscopy of the GPR structures after 2 weeks in culture compared to the human cornea.