

Supplementary information

Rapid in situ forming PEG hydrogels for mucosal drug delivery

Taj Yeruva^{1*}, Robert Morris², Luke Zhao¹, Peter Kofinas², Gregg A. Duncan^{1*}

¹Fischell Department of Bioengineering, University of Maryland, College Park, MD 20742, USA

²Department of Chemical & Biomolecular Engineering, University of Maryland, College Park, MD 20742, USA

*Correspondence to: Gregg Duncan, email: gduncan@umd.edu; Taj Yeruva, email: tyeruva@umd.edu

Table S1. Gelation and degradation times of rapid forming PEG gels.

4-arm PEG-SH (%w/v)	4-arm PEG-SH (%w/v)	Gelation time	
		10kD PEG	20 kD PEG
0.5	0.5	~4 min	~1.5 min
0.5	1	~1.5 min	~50 s
1	0.5	~2 days	~5 min
1	1	~30 s	~30 s
1.5	1	~30 s	~25 s
1.5	1.5	~10 s	~15 s
2	1	>2 hr.	~20s

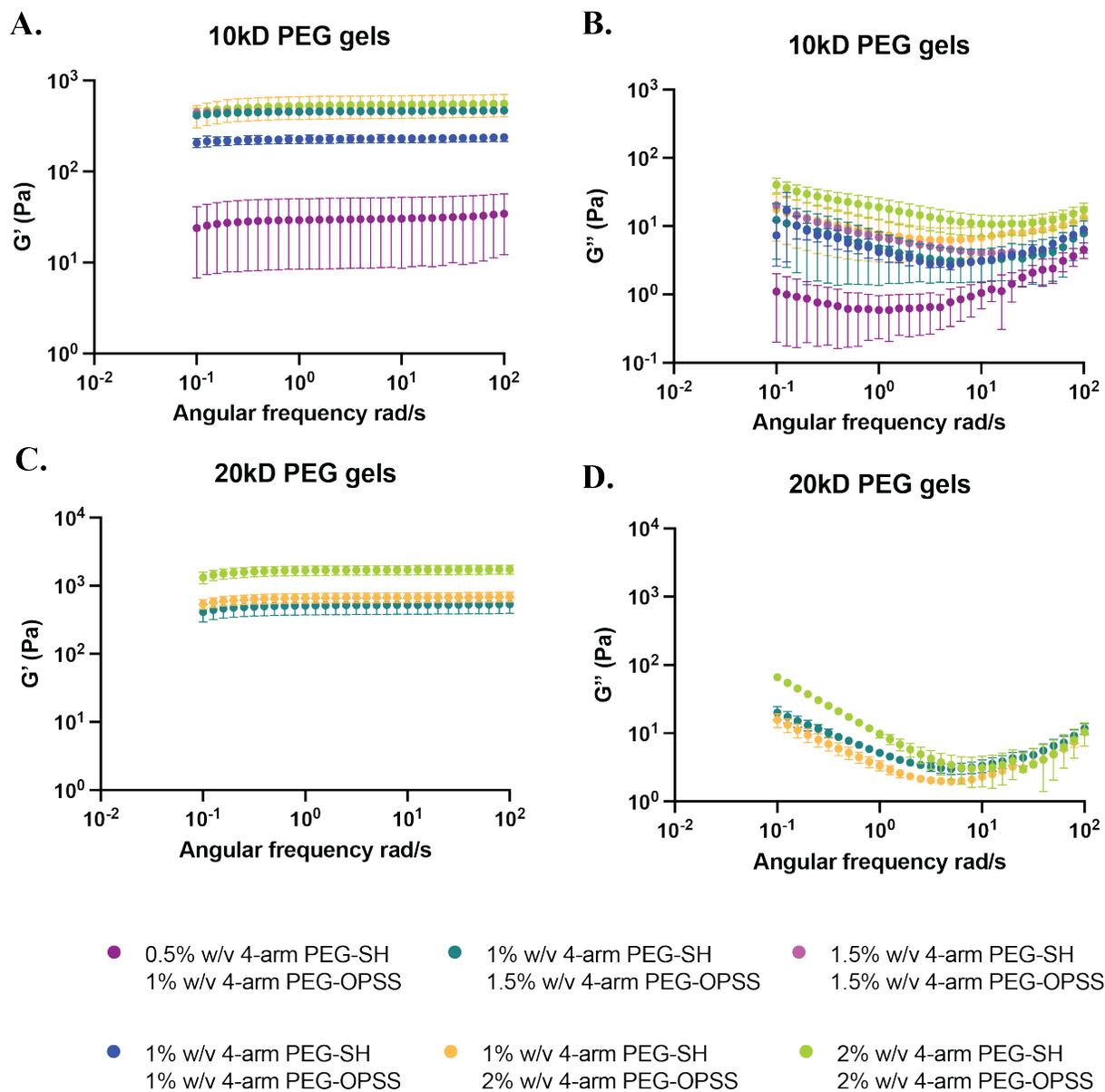
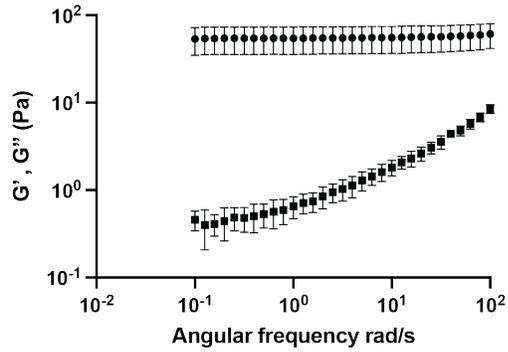
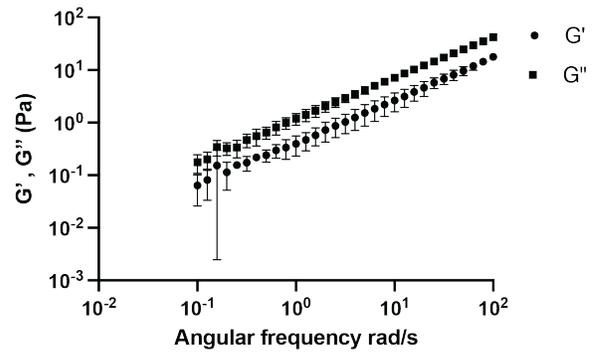


Figure S1. Bulk rheological properties of rapid in situ forming PEG gels. (A), (B) Storage modulus (G') and (C), (D) Loss modulus (G'') as a function of frequency at 10% strain ($n=3$).

A.



B.



C.

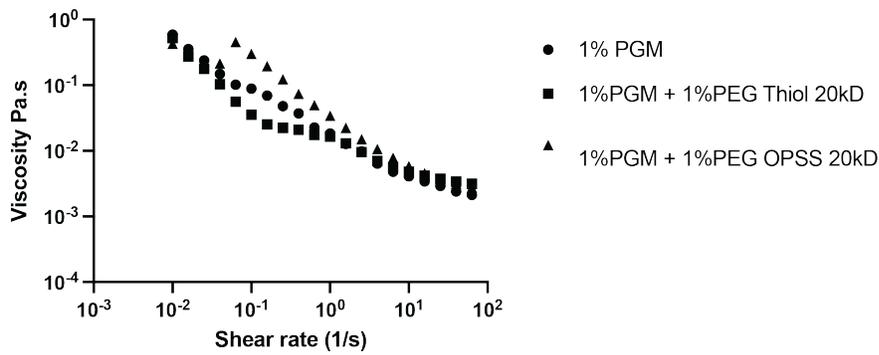


Figure S2. Bulk rheological properties of control gels used for mucoadhesion. Storage modulus (G') and Loss modulus (G'') as a function of frequency at 10% strain of (A) 2% w/v 4-arm PEG-DBCO 10kD crosslinked with 2% w/v 4-arm PEG-Azide 10kD ($n=3$), (B) 4% w/v chitosan ($n=3$). (C) Flow sweep ($n=1$) showing no instant crosslinking between mucin chains and PEG-Thiol or PEG-OPSS indicating mucoadhesion at 0 hr. is driven by polymer-mucin chain entanglement and hydrogen bonding.

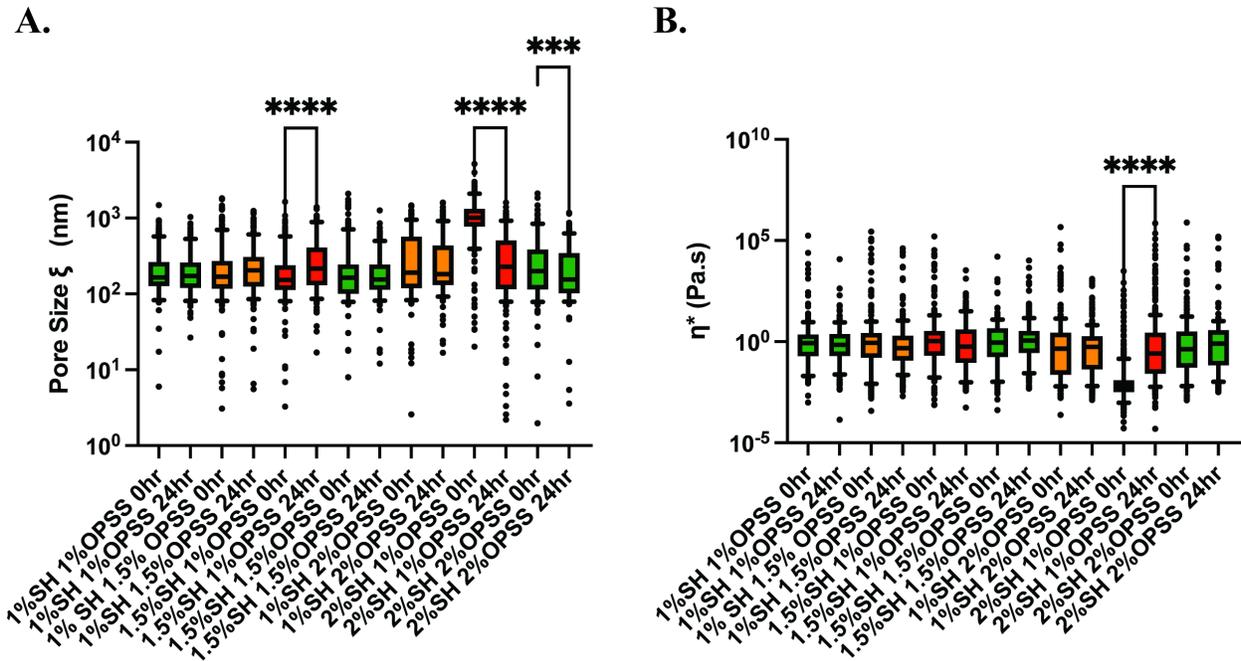


Figure S3. Micro rheological properties of 10kD PEG gels at 0 hr. and 24 hr. after mixing PEG-SH and PEG-OPSS solutions. (A) Estimated pore size based on analysis of mean square displacement (MSD) at $\tau=1s$. (B) Complex microviscosity (η^*) at a frequency $\omega = 1$ Hz calculated from measured MSD. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$ for Kruskal-Wallis test.

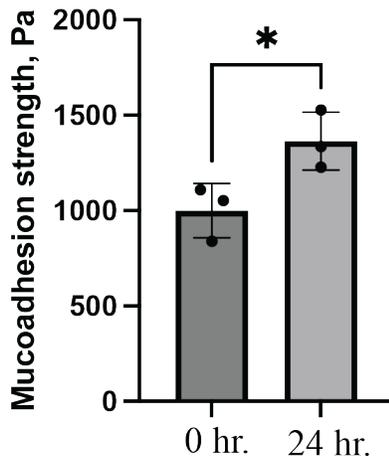


Figure S4. Mucoadhesive properties of 0.5% w/v 4-arm PEG-SH 10kD and 1% w/v 4-arm PEG-OPSS 10kD gels at 0 hr. and 24hr. after application to porcine intestinal tissue ($n=3$). * $p < 0.05$ for Welch's t test.

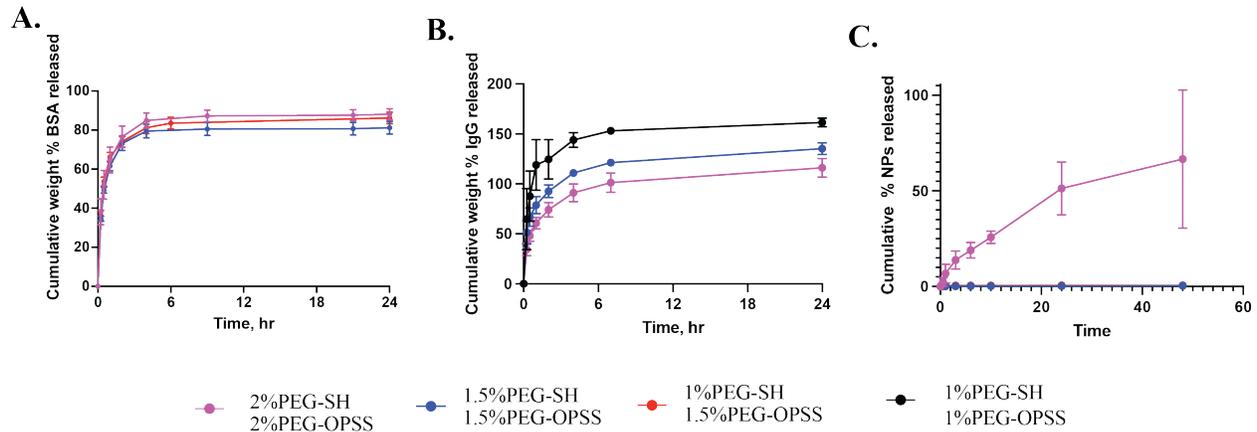


Figure S5. Release kinetics of model therapeutic cargo from different formulations of rapid forming PEG hydrogels. Cumulative release profile of (A) BSA from 10kD PEG gels (n=3), (B) IgG from 10kD PEG gels (n=3), and (C) 20nm nanoparticles (NP) from 20kD PEG gels (n=3) over 24 hours.

FITC labeled IgG was used for these release experiments. More than 100% release was observed likely due to fluorescence dequenching following IgG hydrolysis over time.[1]

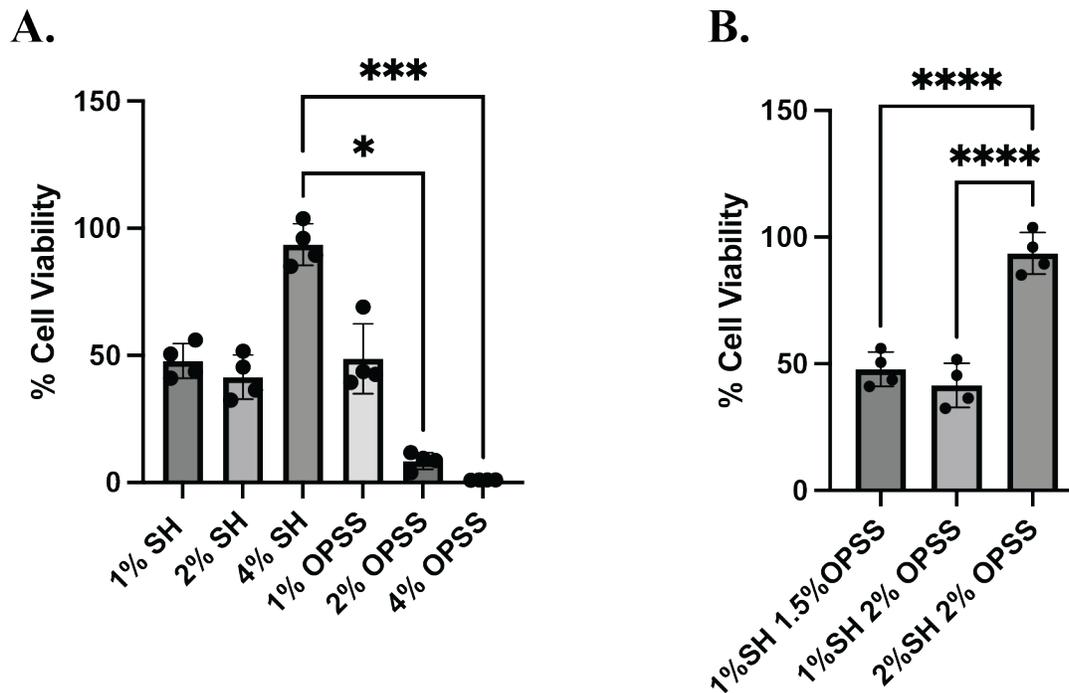


Figure S6. Biocompatibility of rapid forming 5kD PEG gels. Viability of HEK-293T cells following treatment with (A) 4-arm PEG solutions (n=4) * $p < 0.05$ *** $p < 0.001$ for Kruskal-Wallis test. (B) PEG hydrogels (n=4) **** $p < 0.0001$ for one-way ANOVA with Tukey's multiple comparison test.

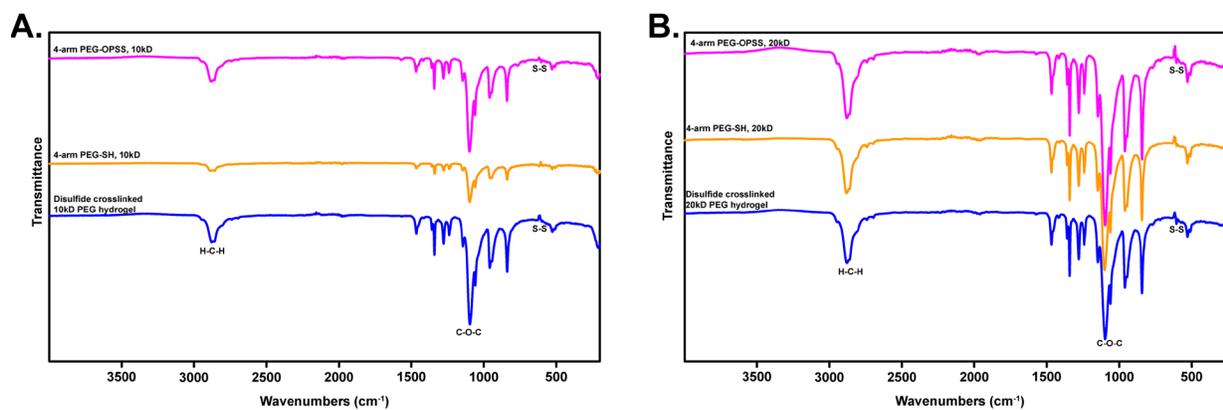


Figure S7. FTIR Spectrum of rapid forming PEG gels. (A) 10kD and (B) 20kD

References:

- [1] C. Wischke, H.-H. Borchert, Fluorescein isothiocyanate labelled bovine serum albumin (FITC-BSA) as a model protein drug: opportunities and drawbacks, *Pharmazie* 61 (2006) 770–774.