

Fluorescence-enhanced supra-amphiphilics based on pillar[5]arene: construction, controllable self-assembly and application in cell-imaging

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1. Materials and methods

Materials

All reagents were commercially available and used as supplied without further purification. Solvents were either employed as purchased or dried according to procedures described in the literature. The cell lines used in the study were purchased from Jiangsu A.K.X. Material Technology Co., Ltd.

Measurements

NMR spectroscopy. ^1H and ^{13}C NMR spectra were recorded on a Bruker AV400 spectrometer.

Fluorescence spectroscopy. Steady-state fluorescence spectra were recorded in a conventional quartz cell (light path 10 mm) on a Varian Cary Eclipse equipped with a Varian Cary single-cell peltier accessory to control temperature.

UV/Vis spectroscopy. UV/Vis spectra and the optical transmittance were recorded in a quartz cell (light path 10 mm) on a Shimadzu UV-3600 spectrophotometer equipped with a PTC-348WI temperature controller.

ESI-MS spectroscopy. Electrospray ionization mass spectra (ESI-MS) were measured by Agilent 6520 Q-TOF-MS.

Cytotoxicity experiments. HeLa cells were incubated in Dulbecco's modified Eagle's medium (DMEM). The medium was supplemented with 10% fetal bovine serum and 1% Penicillin-Streptomycin. Cells were seeded in 96-well plates (5×10^4 cell mL^{-1} , 0.1 mL per well) for 4 h at 37°C in 5% CO_2 . Then the cells were incubated with different groups for 4 h. The relative cellular viability was determined by the MTT assay.

Confocal laser scanning microscopy. Cells were seeded in 6-well plates (5×10^4 cell mL^{-1} , 2 mL per well) for 24 h at 37°C in 5% CO_2 . The cells were incubated with the corresponding solution for 4 h. Then the medium was removed, and the cells were

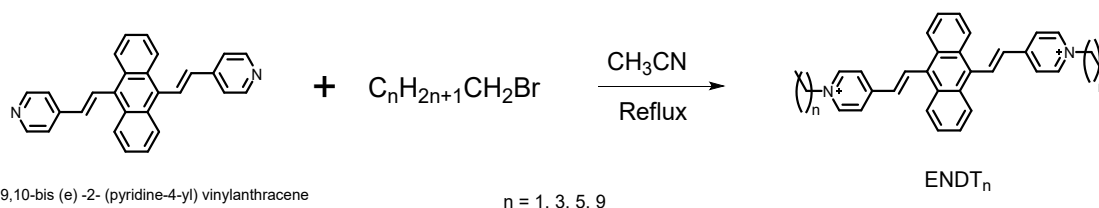
washed with phosphate buffer solution for three time. Finally, the cells were subjected to observation by a confocal laser scanning microscope.

TEM microscopy. High-resolution Transmission electron microscopy (TEM) images were acquired using a Tecnai 20 high-resolution transmission electron microscope operating at an accelerating voltage of 200 keV. The sample for high-resolution TEM measurements was prepared by dropping the solution onto a copper grid. The grid was then air-dried.

SEM microscopy. Scanning electron microscopy (SEM) investigations were carried out on a HitachiS-3400 SEM instrument.

DLS spectroscopy. Solution samples were examined on a laser light scattering spectrometer (BI-200SM) equipped with a digital correlator (TurboCorr) at 636 nm at a scattering angle of 90°. The hydrodynamic diameter (Dh) was determined by DLS experiments at 25°C.

2. Synthesis of ENDT_n



Scheme S1. Synthetic route to ENDT_n .

ENDT_n was prepared successfully by refluxing compound 9,10-bis (e) -2-(pyridine-4-yl) vinylanthracene and bromo-alkanes in MeCN (30 mL). In a typical process, compound 9,10-bis (e) -2-(pyridine-4-yl) vinylanthracene (0.49 g, 1.0 mmol) and excess bromoethane (1.08 g, 10.0 mmol) were added to MeCN (30 mL) and then reflux overnight. After the reaction was finished, the reaction mixture was cold to room temperature, ENDT_1 was filtered off and washed with MeCN for 3 times. The yield was about 50.3%. Compound ENDT_3 , ENDT_5 , ENDT_9 are repeat by this step.

ENDT_1 : Red solid, 56.7 %; m.p. > 250 °C. ^1H NMR (400 MHz, D_2O) δ (ppm): 8.68(d, 4H), 8.24(d, 2H), 8.10(m, 4H), 8.01(m, 4H), 7.46(m, 4H), 6.90(d, 2H), 4.53(t, 4H), 1.58(m, 6H). ^{13}C NMR (100 MHz, D_2O) δ (ppm): 151.8, 143.2, 136.3, 131.4, 130.5, 128.0, 126.3, 125.2, 124.1, 56.2, 15.6. LRMS spectra of ENDT_1 . Calcd. For $\text{C}_{32}\text{H}_{30}\text{N}_2$ ($[\text{M} - 2\text{Br}]^{2+}$): 221.12, found: 221.11. IR(KBr) ν : 3432, 3020, 2923, 1610, 1556, 1513, 1466, 1462, 1441, 1377, 1335, 1208, 1161, 1008, 864, 753, 710 cm^{-1} .

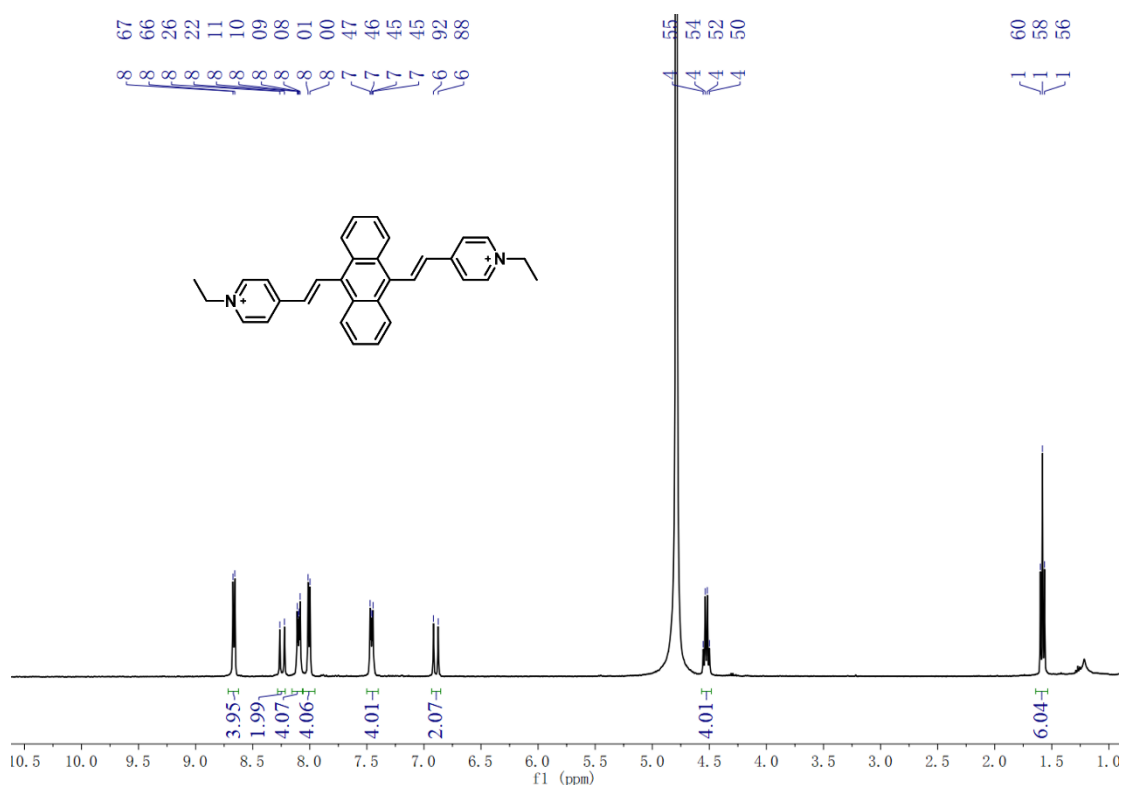


Figure S1: ^1H NMR spectra (400 MHz, 298 K, D_2O) of compound ENDT_1 .

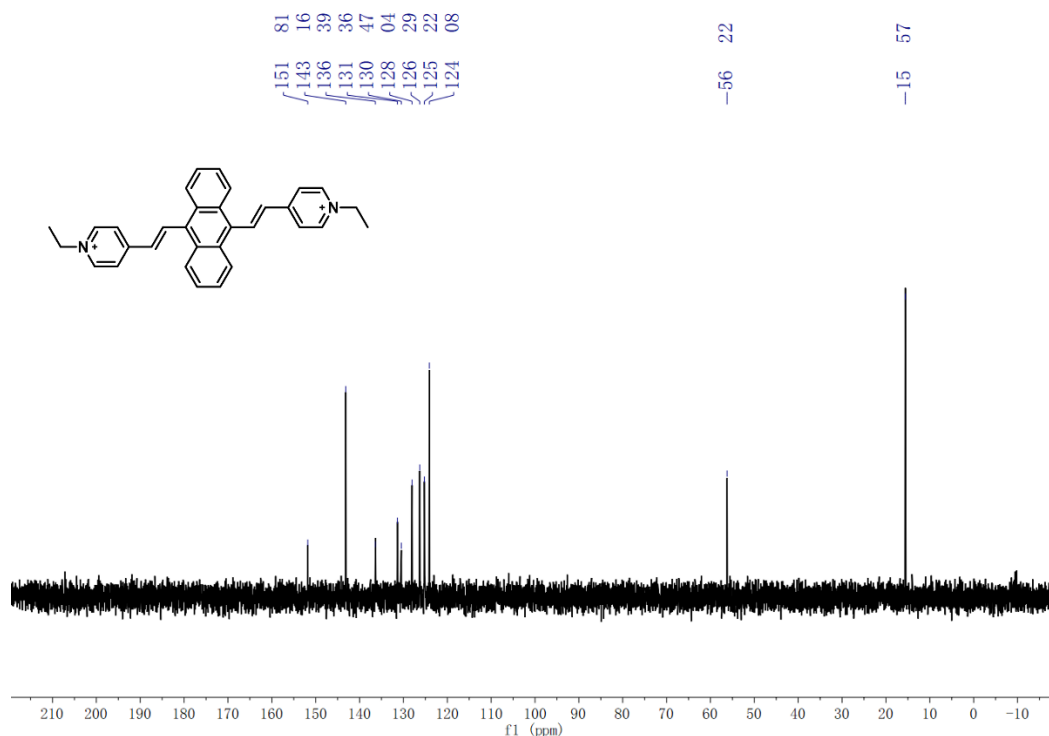


Figure S2. ^{13}C NMR spectra (100 MHz, 298 K, D_2O) of compound ENDT_1 .

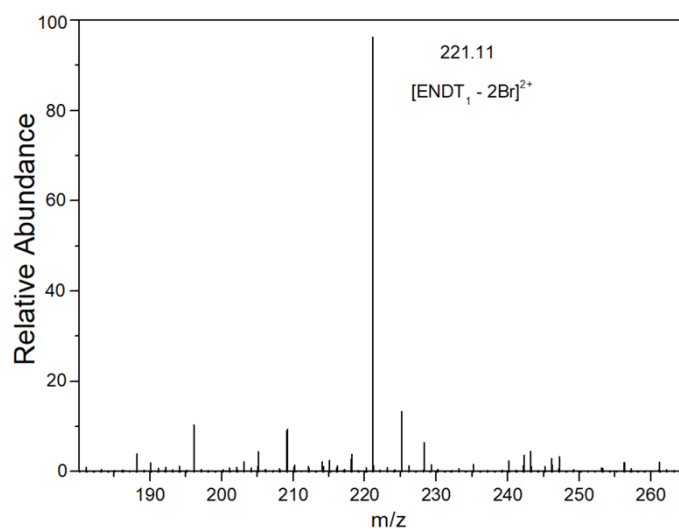


Figure S3. LRMS spectra of ENDT_1 . Calcd. For $\text{C}_{32}\text{H}_{30}\text{N}_2$ ($[\text{M} - 2\text{Br}]^{2+}$): 221.12, found: 221.11.

ENDT₃: Red solid, 55 %; m.p. > 250 °C. ^1H NMR (400 MHz, CDCl_3) δ (ppm): 8.64 (s, 4H), 8.31 (d, $J = 8.0$ Hz, 2H), 8.12 (s, 4H), 7.99 (s, 4H), 7.48 (s, 4H), 6.92 (d, $J = 8.0$ Hz, 2H), 4.47 (s, 4H), 1.93 (s, 4H), 1.31 (m, 4H), 0.90 (d, $J = 4.0$ Hz, 6H). ^{13}C NMR (100 MHz, $\text{CD}_3\text{CN}/\text{DMSO}$) δ (ppm): 152.9, 144.9, 138.5, 133.1, 132.4, 129.4, 126.9, 126.5, 125.2, 60.5, 33.2, 19.2, 13.4. IR(KBr) ν : 3464, 3392, 3041, 2963, 2882, 1634, 1604, 1510, 1455, 1379, 1311, 1167, 1094, 988, 844, 755 cm^{-1} . LRMS spectra of ENDT_3 . Calcd. For $\text{C}_{36}\text{H}_{38}\text{N}_2$ ($[\text{M} - 2\text{Br}]^{2+}$): 249.15, found: 249.13.

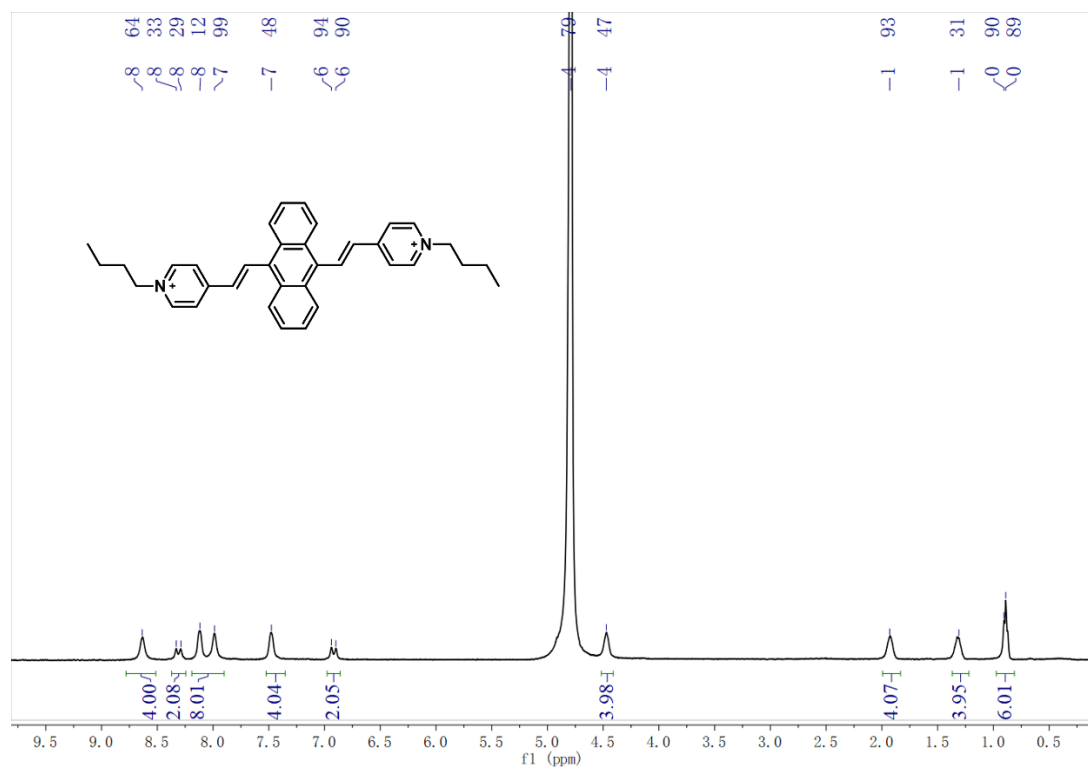


Figure S4. ¹H NMR spectra (400 MHz, 298 K, D₂O) of compound ENDT₃.

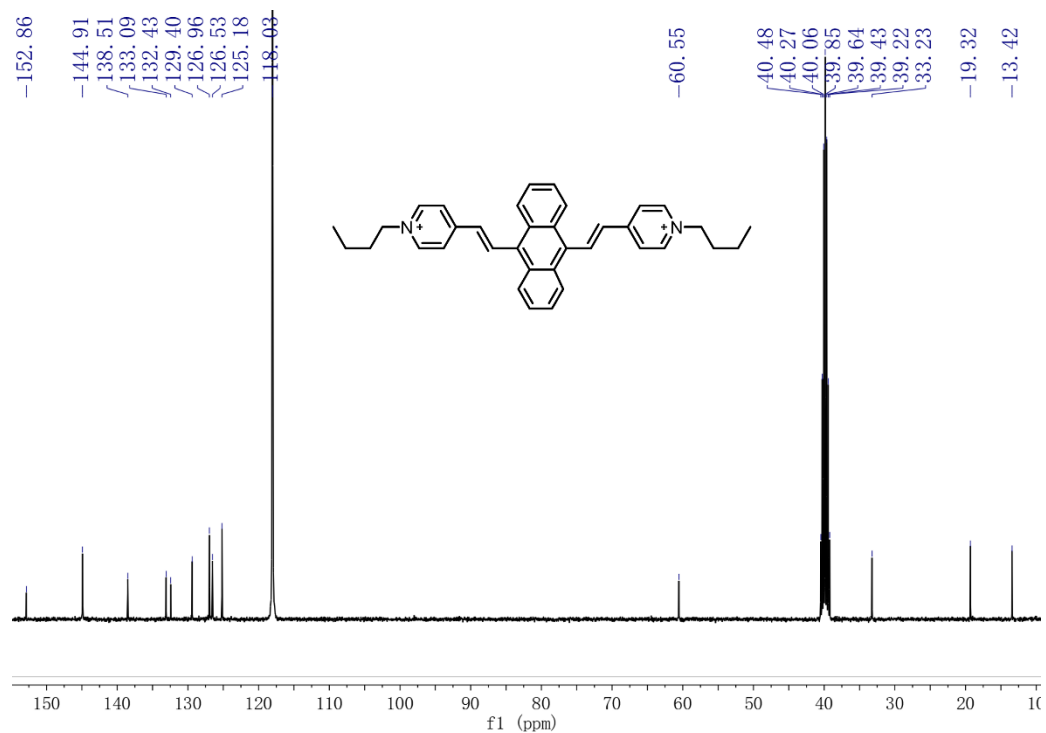


Figure S5. ¹³C NMR spectra (100 MHz, 298 K, CD₃CN/DMSO) of compound ENDT₃.

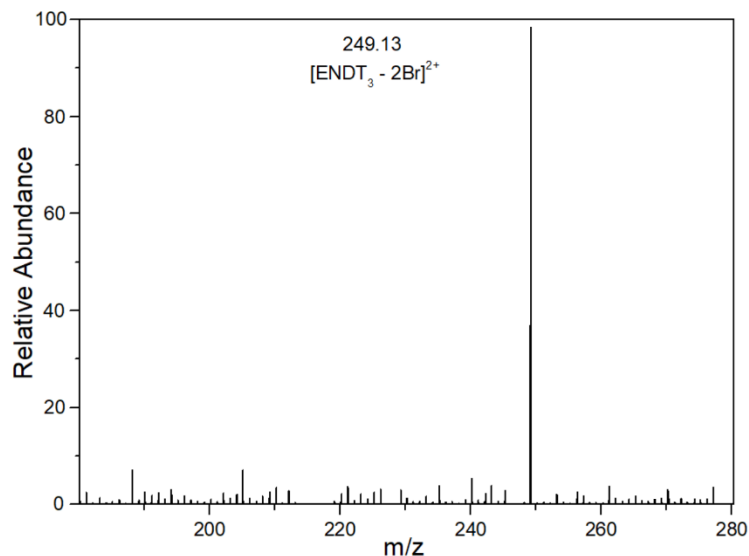


Figure S6. LRMS spectra of ENDT₃. Calcd. For C₃₆H₃₈N₂ ([M - 2Br]²⁺): 249.15, found: 249.13.

ENDT₅: Red solid, 52 %; m.p. > 250 °C. ¹H NMR (400 MHz, Acetonitrile-d₃) δ (ppm): 8.74 (d, *J* = 8.0 Hz, 2H), 8.69 (d, *J* = 4.0 Hz, 4H), 8.41-8.38 (m, 4H), 8.25 (d, *J* = 4.0 Hz, 4H) 7.64-7.61 (m, 4H), 7.23 (d, *J* = 8.0 Hz, 4H), 4.51 (t, *J* = 8.0 Hz, 4H), 2.00 (t, *J* = 8.0 Hz, 4H), 1.36 (s, 12H), 0.91 (t, *J* = 6.0 Hz, 6H). ¹³C NMR (100 MHz, Acetonitrile-d₃) δ (ppm): 153.4, 144.8, 138.8, 133.1, 132.5, 129.7, 127.2, 126.6, 125.4, 118.4, 61.6, 31.3, 25.8, 22.7, 13.8. IR(KBr) ν: 3462, 3385, 3037, 2956, 2927, 2867, 1611, 1508, 1457, 1440, 1258, 1164, 1092, 1015, 858, 799, 743 cm⁻¹. LR-MS spectra of ENDT₅. Calcd. For C₄₀H₄₆N₂ ([M - 2Br]²⁺): 277.18, found: 277.16.

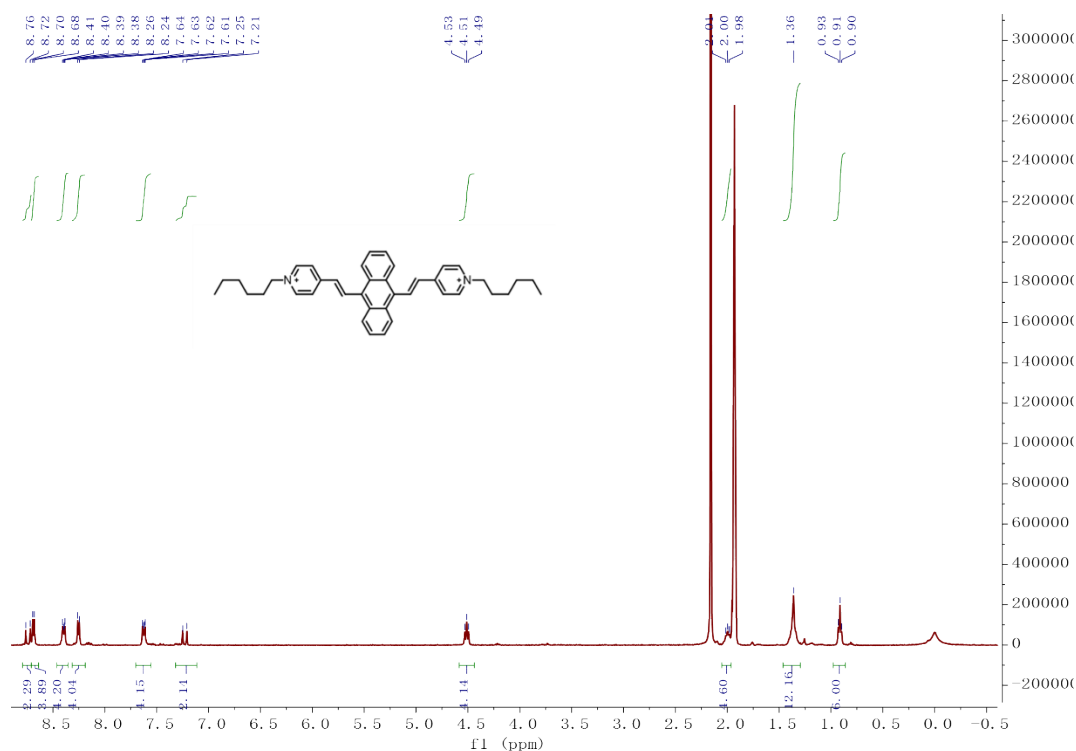


Figure S7. ^1H NMR spectra (400 MHz, 298 K, CD_3CN) of compound ENDT_5 .

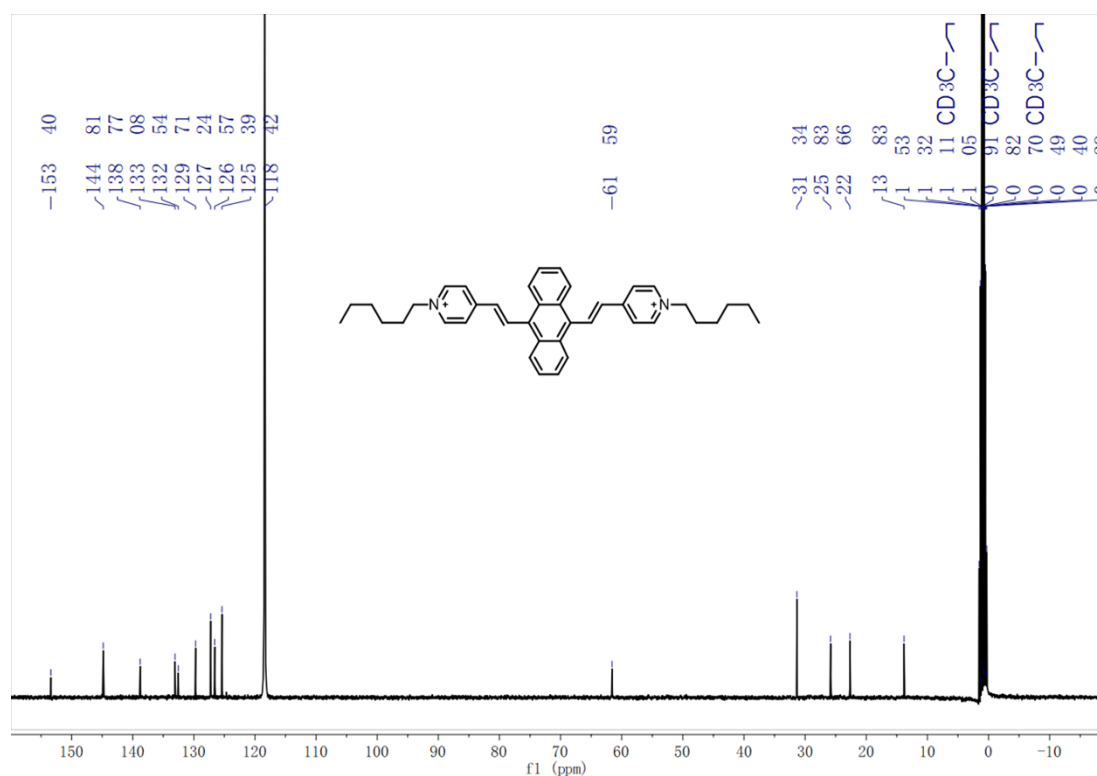


Figure S8. ^{13}C NMR spectra (100 MHz, 298 K, CD_3CN) of compound ENDT_5 .

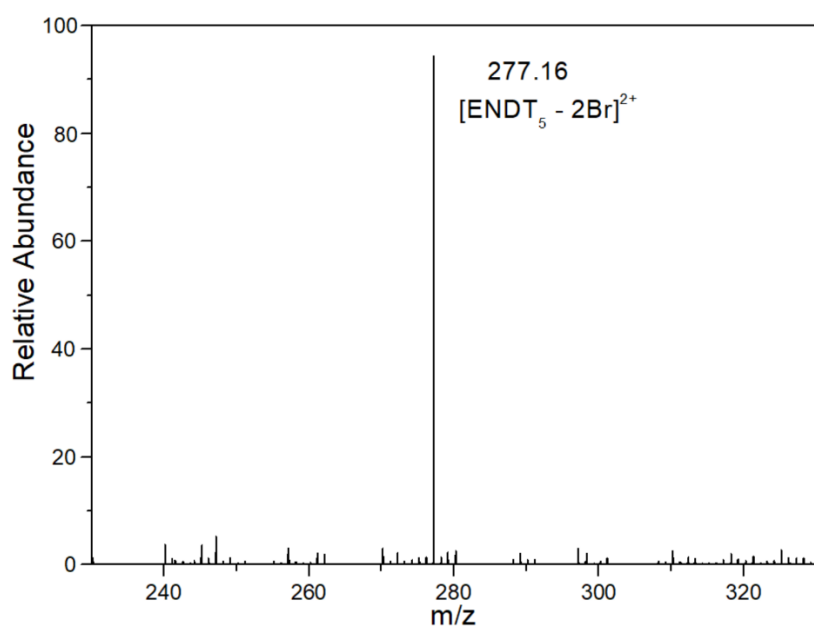


Figure S9. LRMS spectra of ENDT_5 . Calcd. For $\text{C}_{40}\text{H}_{46}\text{N}_2$ ($[\text{M} - 2\text{Br}]^{2+}$): 277.18, found: 277.16.

ENDT_9 : Dark red solid, 49 %; m.p. > 250 °C. ^1H NMR (400 MHz, Acetonitrile- d_3) δ (ppm): 8.78 – 8.74 (m, 4H), 8.37 (d, $J = 4.0$ Hz, 4H), 8.28 (d, $J = 8.0$ Hz, 2H), 7.63 (t, $J = 4.0$ Hz, 2H), 7.52 (s, 6H), 7.22 (d, $J = 8.0$ Hz, 2H), 4.55 (t, $J = 8.0$ Hz, 4H), 3.44 (t,

$J = 6.0$ Hz, 4H), 1.39-1.21 (m, 28H), 0.88 (m, 6H). ^{13}C NMR (100 MHz, Acetonitrile- d_3) δ (ppm): 153.4, 144.8, 138.8, 133.1, 132.5, 129.7, 127.2, 126.5, 125.3, 118.5, 61.6, 33.3, 32.1, 31.3, 29.6, 29.1, 26.1, 22.9, 19.5, 13.9, 13.3. IR(KBr) ν : 3440, 3024, 2914, 2846, 1606, 1513, 1457, 1376, 1321, 1164, 1101, 981, 846, 757 cm^{-1} . LRMS spectra of ENDT_9 . Calcd. For $\text{C}_{48}\text{H}_{62}\text{N}_2$ ($[\text{M} - 2\text{Br}]^{2+}$): 333.24, found: 333.28.

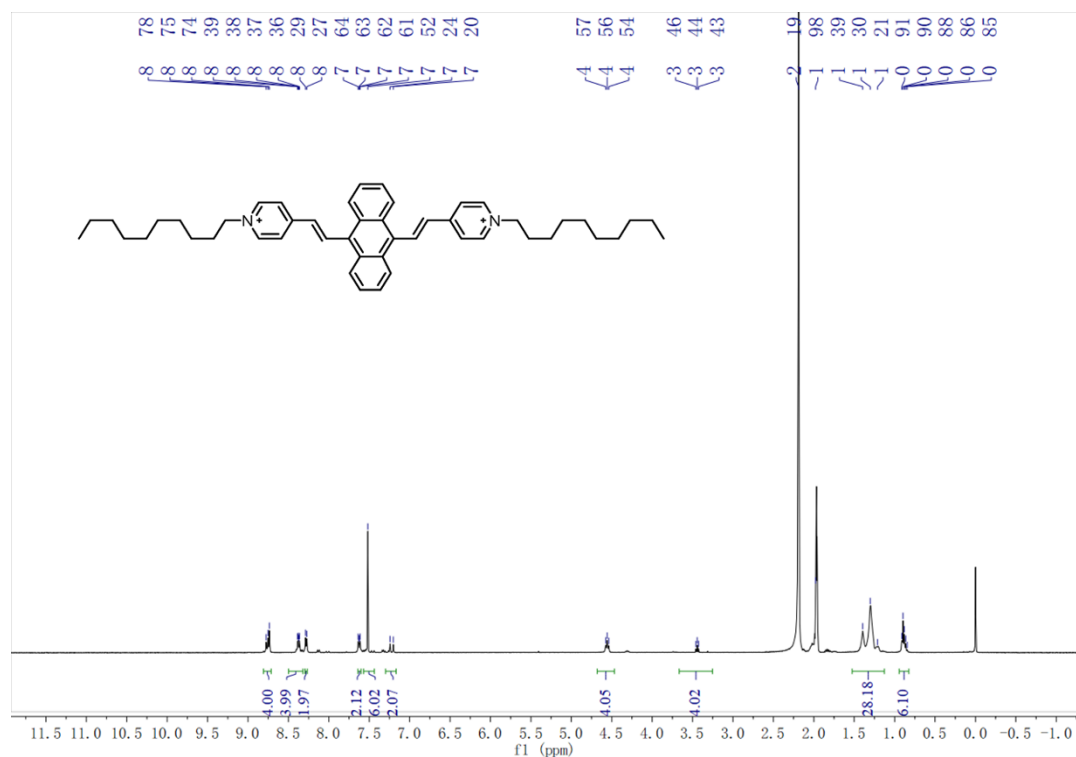


Figure S10. ^1H NMR spectra (400 MHz, 298 K, CD_3CN) of compound ENDT_9 .

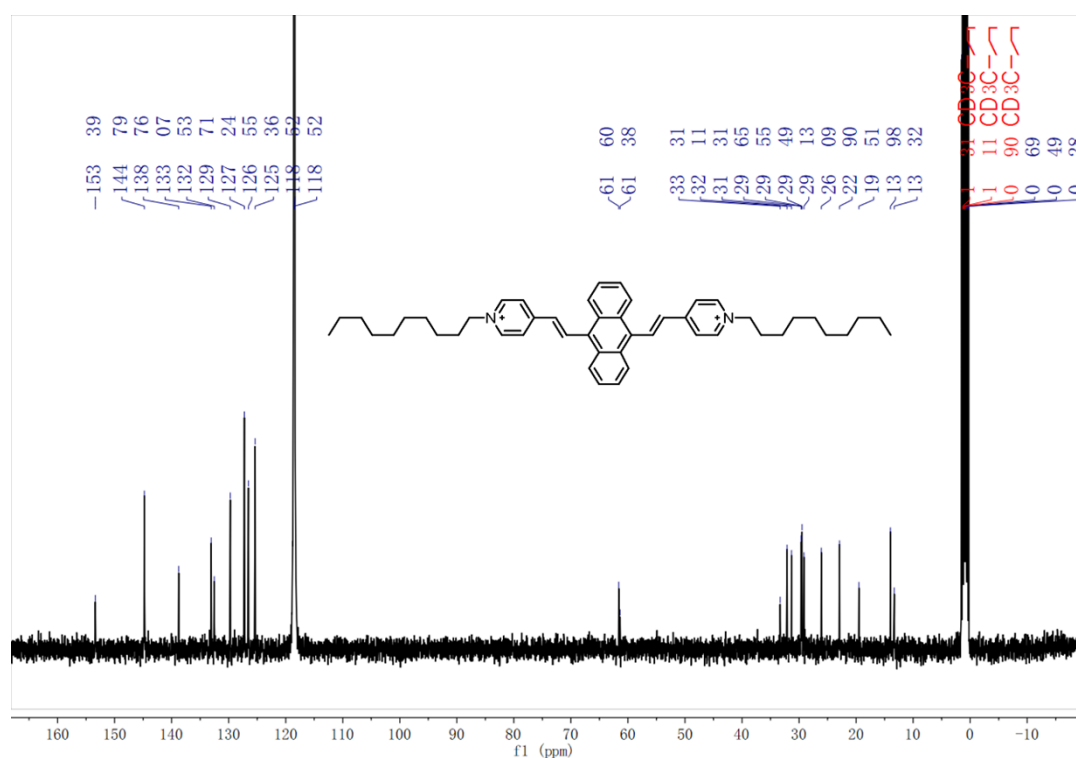


Figure S11. ^{13}C NMR spectra (100 MHz, 298 K, CD_3CN) of compound ENDT_9 .

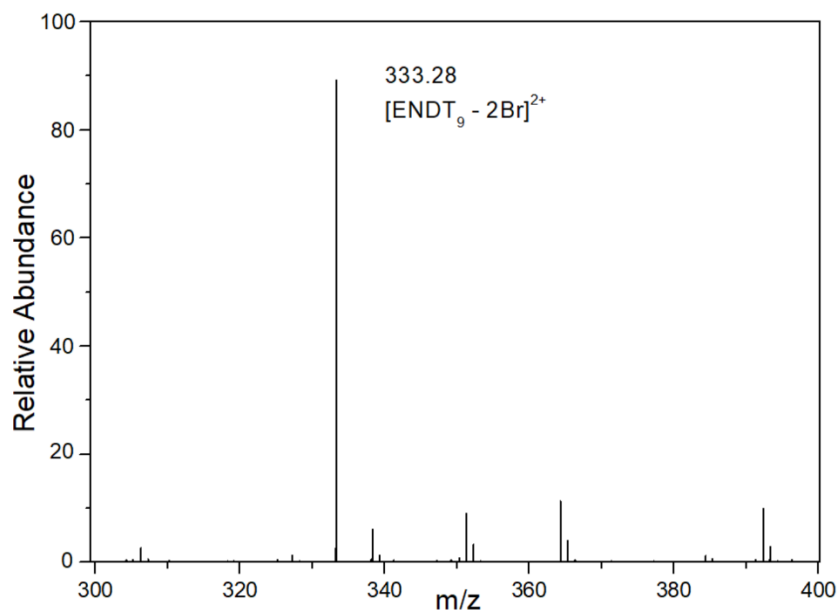
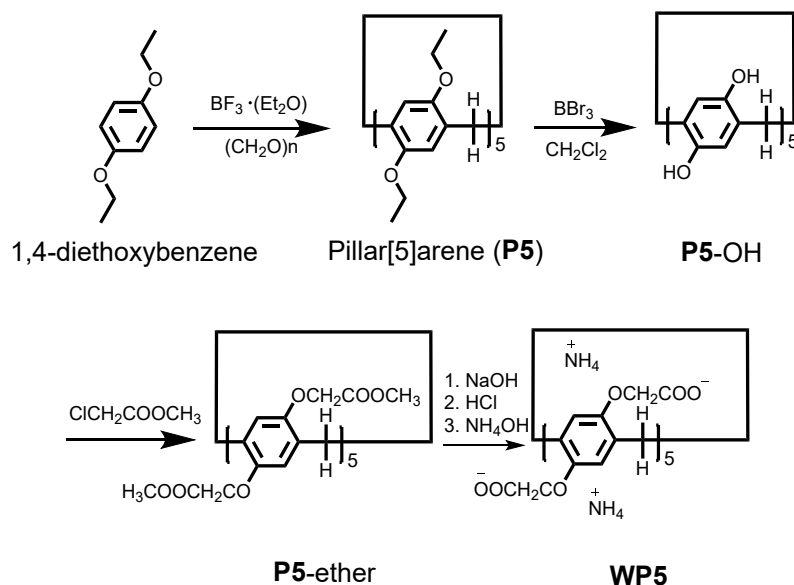


Figure S12. LRMS spectra of ENDT_9 . Calcd. For $\text{C}_{48}\text{H}_{62}\text{N}_2$ ($[\text{M} - 2\text{Br}]^{2+}$): 333.24, found: 333.28.

3. Synthesis of **WP5**

WP5 was prepared according to previous report,^{S1,S2} and the ^1H NMR was showed in Figure S13.



Scheme S2. Synthesis of water soluble pillar[5]arene (**WP5**).

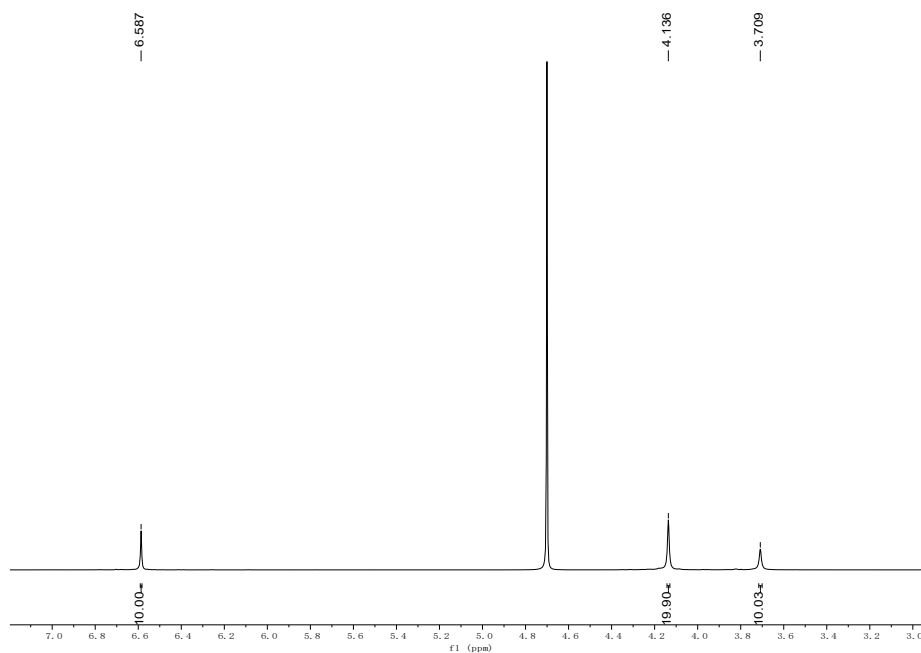


Figure S13. ^1H NMR (400 MHz, 298K, D_2O) spectrum of **WP5**.

4. Host-guest interaction between **WP5** and **ENDT_n**

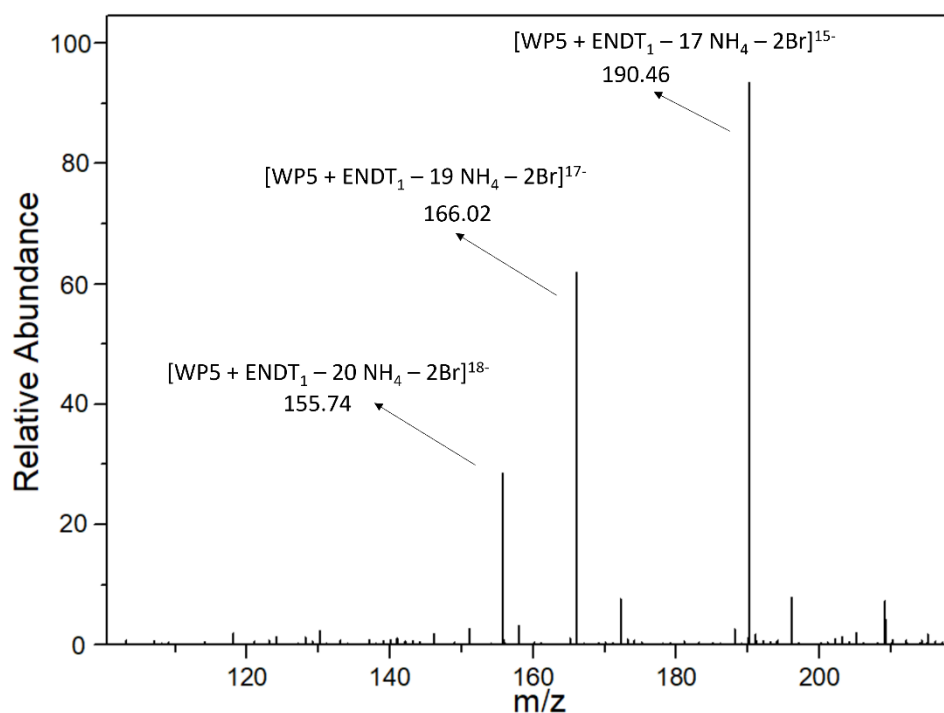


Figure S14. LR-ESI-MS spectra of $(\text{WP5})_2 \supset \text{ENDT}_1$. Calcd. For $\text{C}_{142}\text{H}_{110}\text{N}_2\text{O}_6\text{O}$ ($[\text{2WP5} + \text{ENDT}_1 - 20\text{NH}_4 - 2\text{Br}]^{18-}$): 155.75, found: 155.74; $\text{C}_{142}\text{H}_{114}\text{N}_3\text{O}_6\text{O}$ ($[\text{2WP5} + \text{ENDT}_1 - 19\text{NH}_4 - 2\text{Br}]^{17-}$): 165.98, found: 166.02; $\text{C}_{142}\text{H}_{122}\text{N}_5\text{O}_6\text{O}$ ($[\text{2WP5} + \text{ENDT}_1 - 17\text{NH}_4 - 2\text{Br}]^{15-}$): 190.51, found: 190.46.

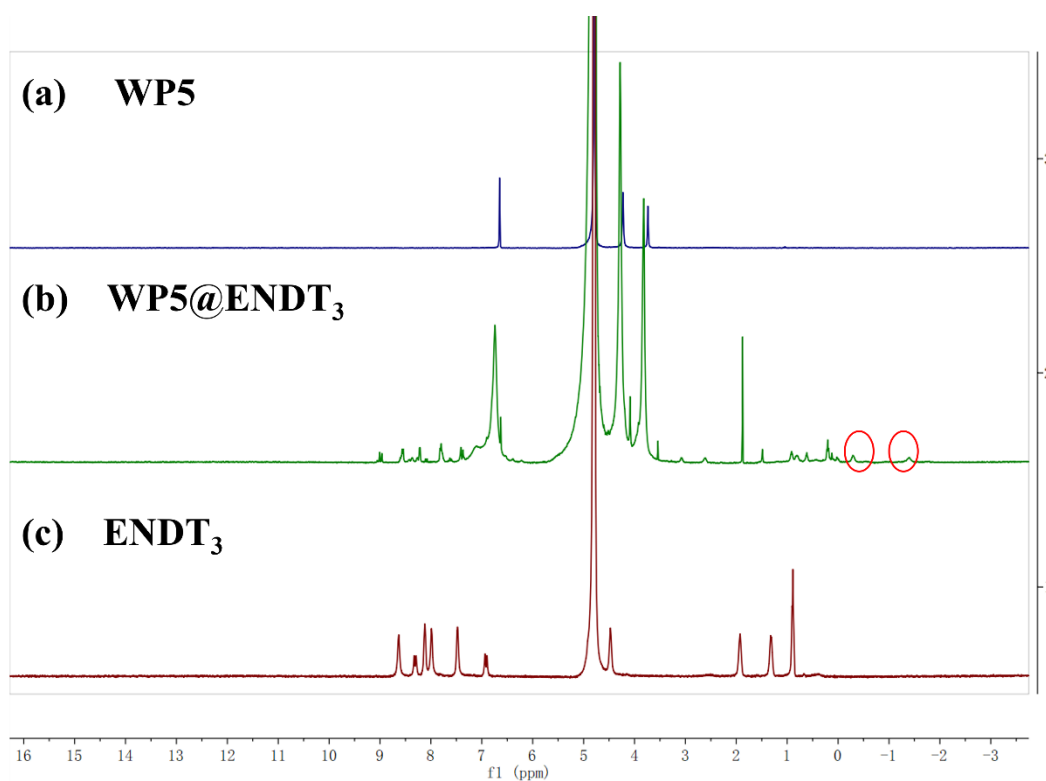


Figure S15. ^1H NMR spectra (400 MHz, D_2O , 298 K) of (a) ENDT_3 (5.0 mM), (b) $\text{ENDT}_3 + \text{WP5}$ ($[\text{ENDT}_3] = 5.0$ mM, $[\text{WP5}] = 10.0$ mM), and (c) ENDT_3 (10.0 mM).

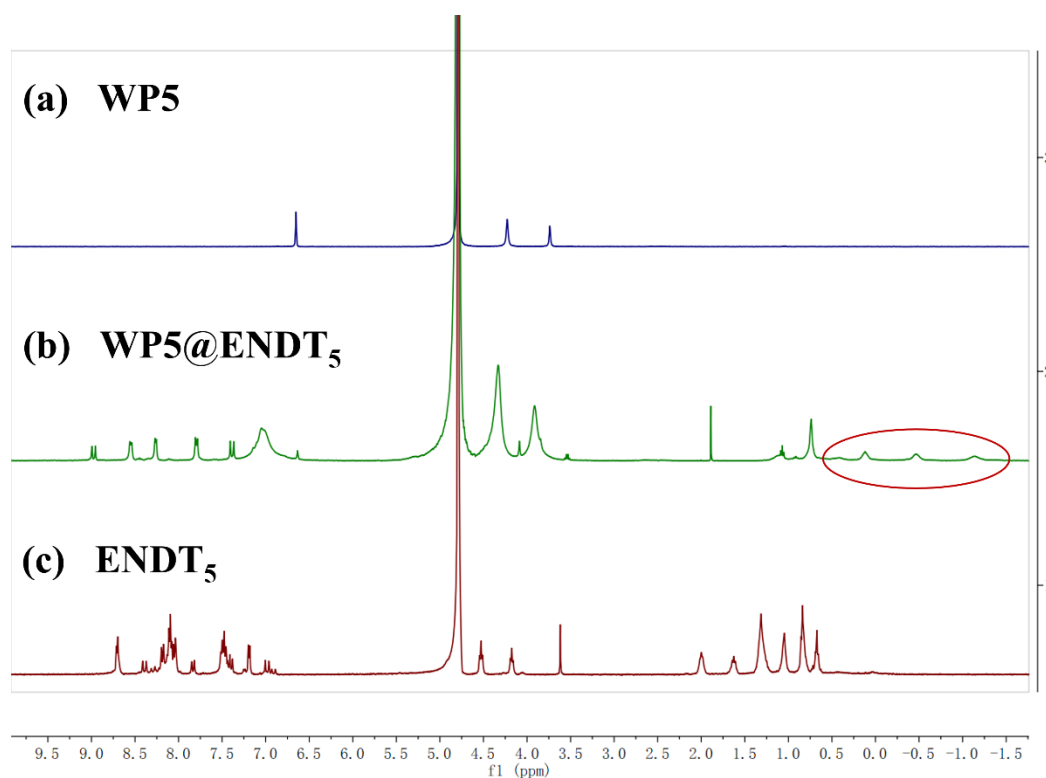


Figure S16. ^1H NMR spectra (400 MHz, D_2O , 298 K) of (a) ENDT_5 (5.0 mM), (b) $\text{ENDT}_5 + \text{WP5}$ ($[\text{ENDT}_5] = 5.0$ mM, $[\text{WP5}] = 10.0$ mM), and (c) ENDT_5 (10.0 mM).

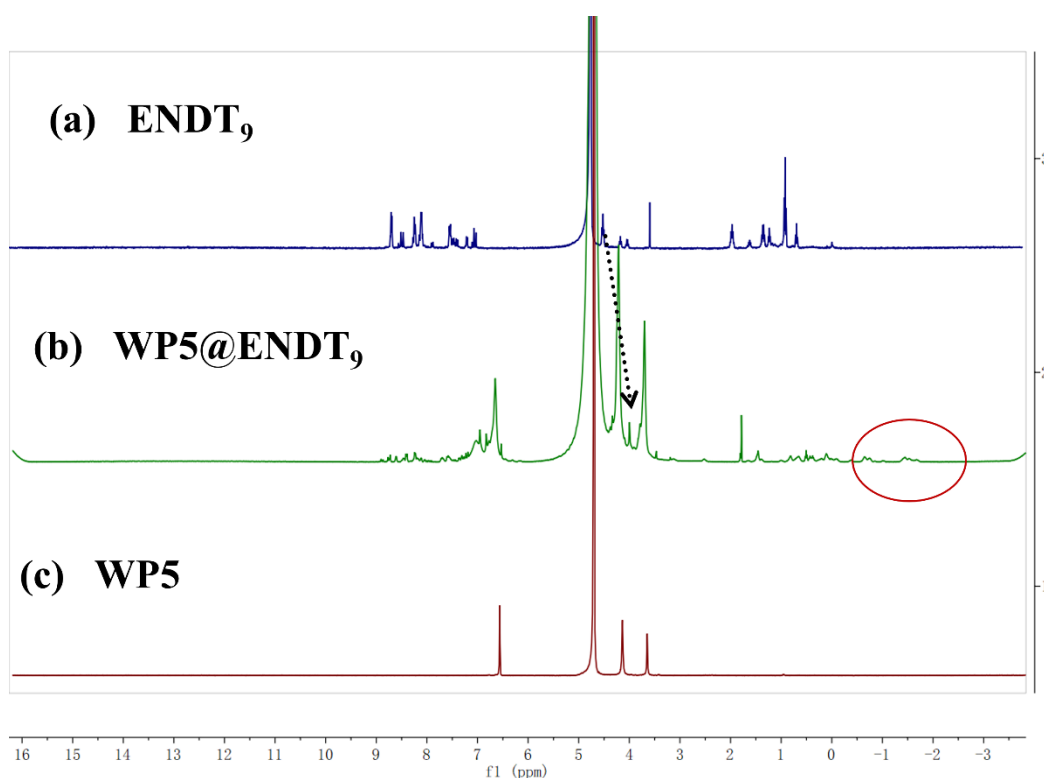


Figure S17. ^1H NMR spectra (400 MHz, D_2O , 298 K) of (a) ENDT_9 (5.0 mM), (b) ENDT_9 + WP5 ($[\text{ENDT}_9] = 5.0$ mM, $[\text{WP5}] = 10.0$ mM), and (c) ENDT_9 (10.0 mM).

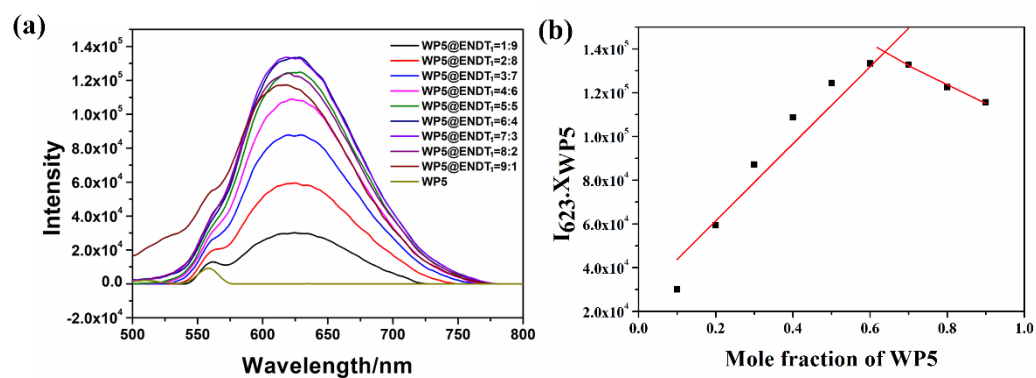


Figure S18. (a) Fluorescence titration of ENDT_1 (10 μM) with different concentration of WP5. (b) Job's plot for ENDT_1 upon addition of WP5 in aqueous medium ($[\text{ENDT}_1] + [\text{WP5}] = 10$ μM).

5. Self-assembly properties

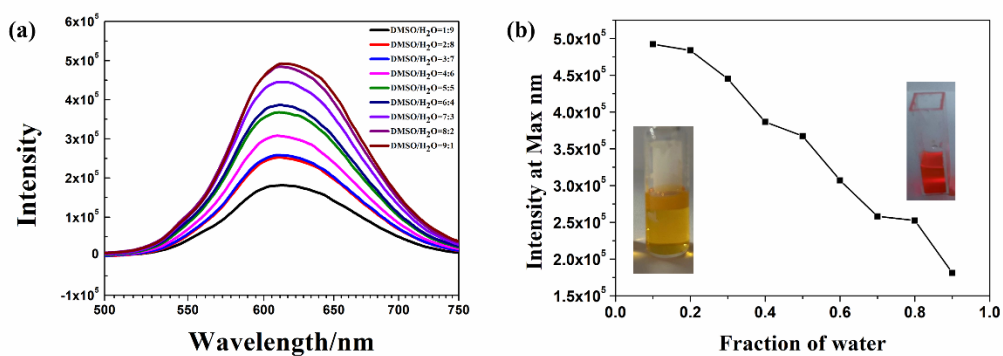


Figure S19. (a) Fluorescence emission of ENDT_1 (10 $\mu\text{mol/L}$) in different ratio of $\text{H}_2\text{O}/\text{DMSO}$ mixture. (b) Fluorescence emission of ENDT_1 (10 $\mu\text{mol/L}$) in Max ratio of $\text{H}_2\text{O}/\text{DMSO}$ mixture.

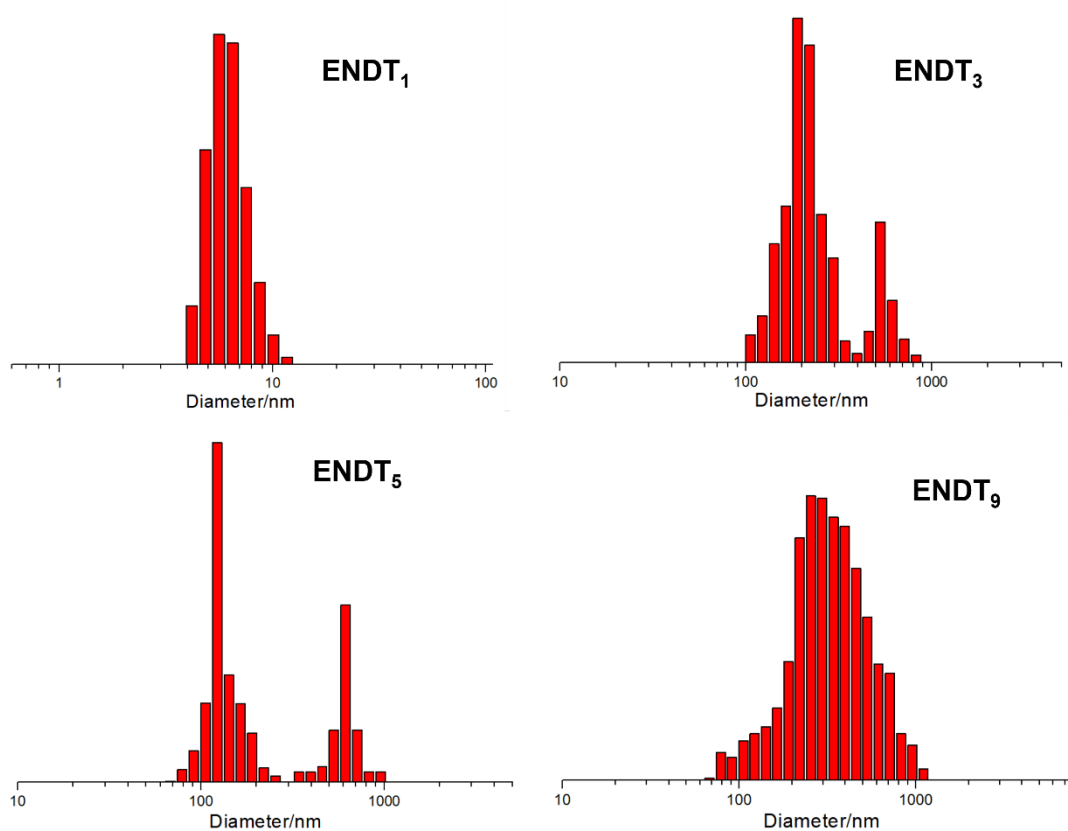


Figure S20. DLS data of ENDT_n (10 $\mu\text{mol/L}$) in water.

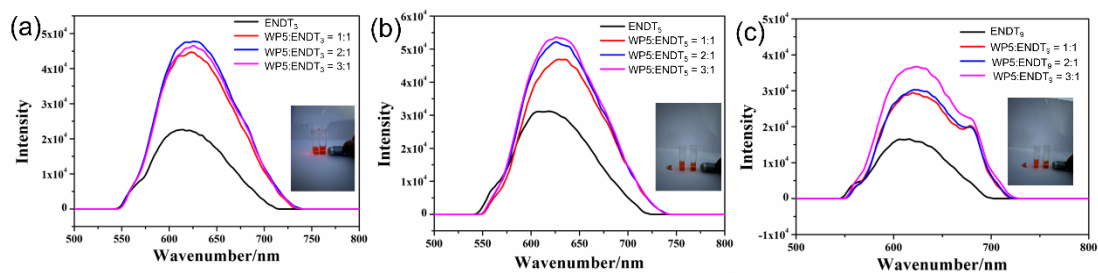


Figure S21. (a-c) Fluorescence titration of ENDT₃, ENDT₅, ENDT₉ (10 μM) with different concentration of WP5. $\lambda_{\text{ex}} = 470 \text{ nm}$. Insert: Trndall of ENDT₃, ENDT₅, ENDT₉ and WP5@ENDT₃, WP5@ENDT₅, WP5@ENDT₉.

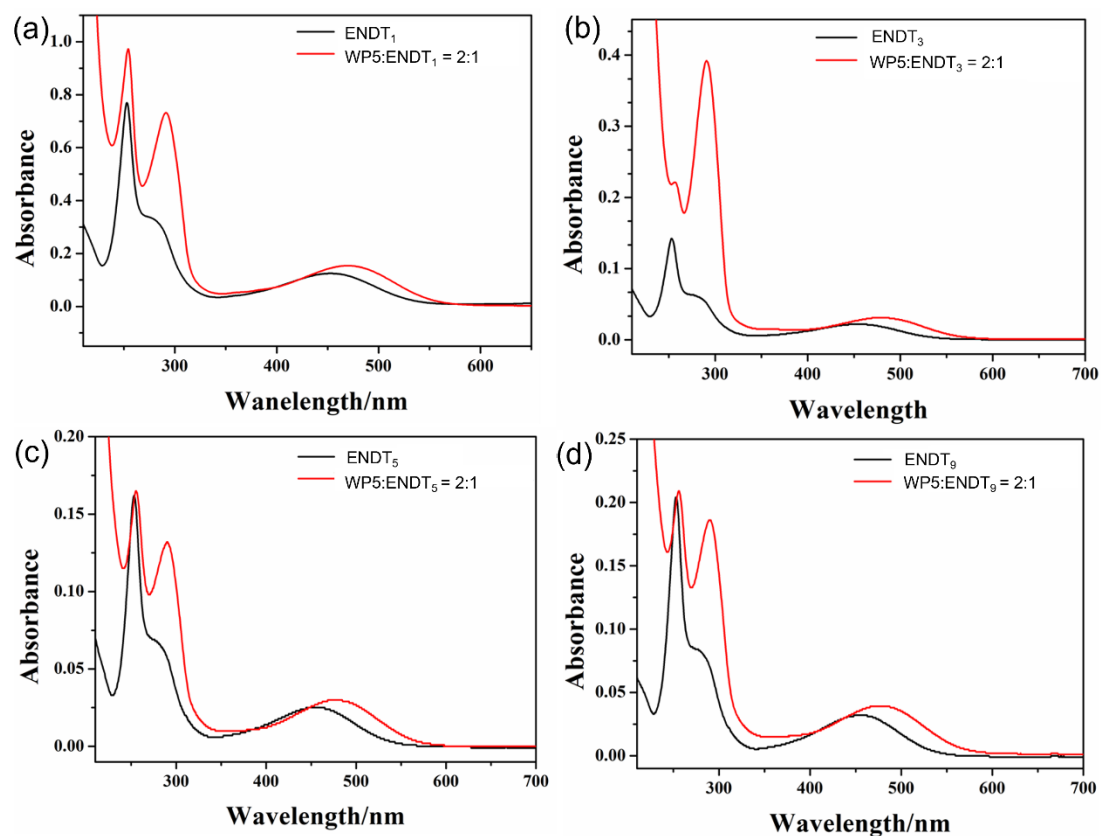


Figure S22. (a-d) UV-vis absorption spectra of ENDT-(1,3,5,9) and (WP5)₂@ENDT-(1,3,5,9). [ENDT-(1,3,5,9) = 10 μmol/L, WP5 = 20 μmol/L].

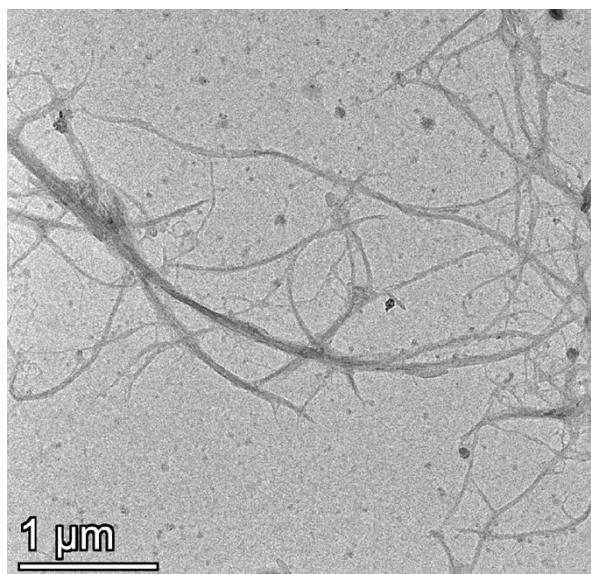


Figure S23. TEM image of ENDT₉ based assemblies in water.

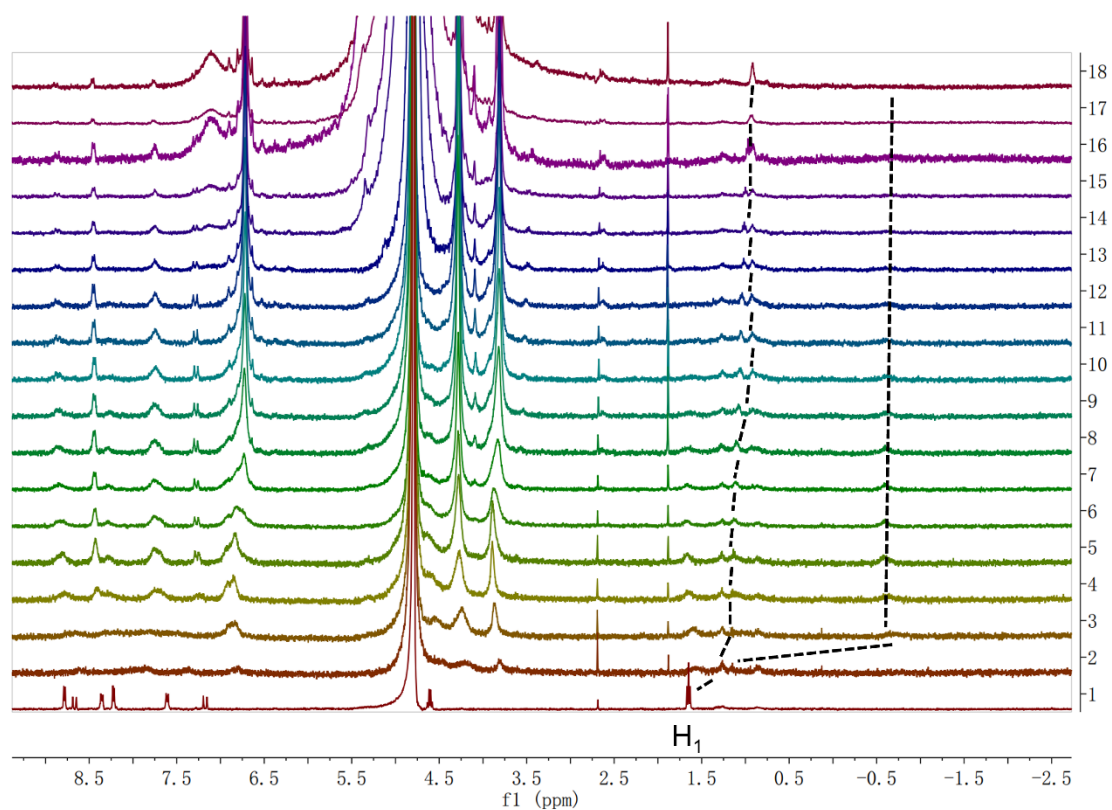


Fig. S24. ¹H NMR spectra (400 MHz, D₂O, 298 K) of ENDT₁ at a concentration of 1.00 mM upon addition of various concentrations of the molecular WP5: (1) 0 mM, (2) 0.196 mM, (3) 0.385 mM, (4) 0.566 mM, (5) 0.741 mM, (6) 0.909 mM, (7) 1.228 mM, (8) 1.525 mM, (9) 1.803 mM, (10) 2.063mM, (11) 2.308 mM, (12) 2.537 mM, (13) 2.754 mM, (14) 2.966 mM, (15) 3.151 mM, (16) 3.590 mM, (17) 3.976 mM and (18) 4.318 mM.

The non-linear curve-fitting was based on a previous report.^{S3}

$$\Delta\delta = (\Delta\delta_\infty / [G]_0) (0.5[H]_0 + 0.5([G]_0 + 1/K_a) - (0.5 ([H]_0^2 + (2[H]_0 (1/K_a - [G]_0)) + (1/K_a + [G]_0)^2)^{0.5})) \dots\dots\dots(\text{Eq. S1})$$

where $\Delta\delta$ is the chemical shift change of H₁ on ENDT₁, $\Delta\delta_\infty$ is the chemical shift change of H₁ when ENDT₁ is completely complexed, [H]₀ is the initial concentration of the WP5, and [G]₀ is the fixed initial concentration of ENDT₁.

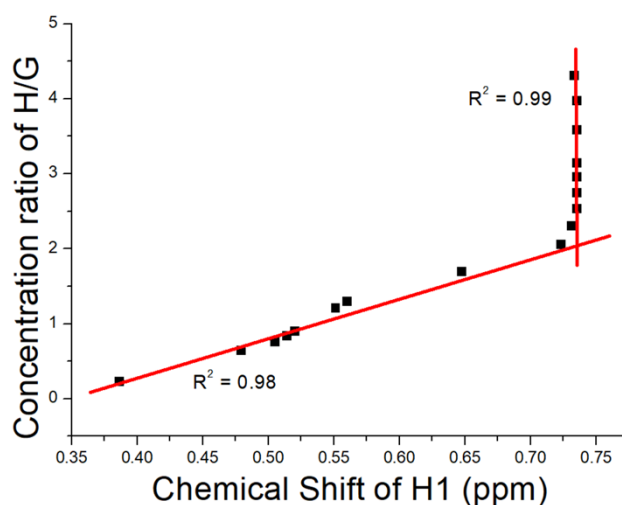


Fig. S25 Mole ratio plot for **WP5** and ENDT₁, indicating a 2 : 1 stoichiometry.

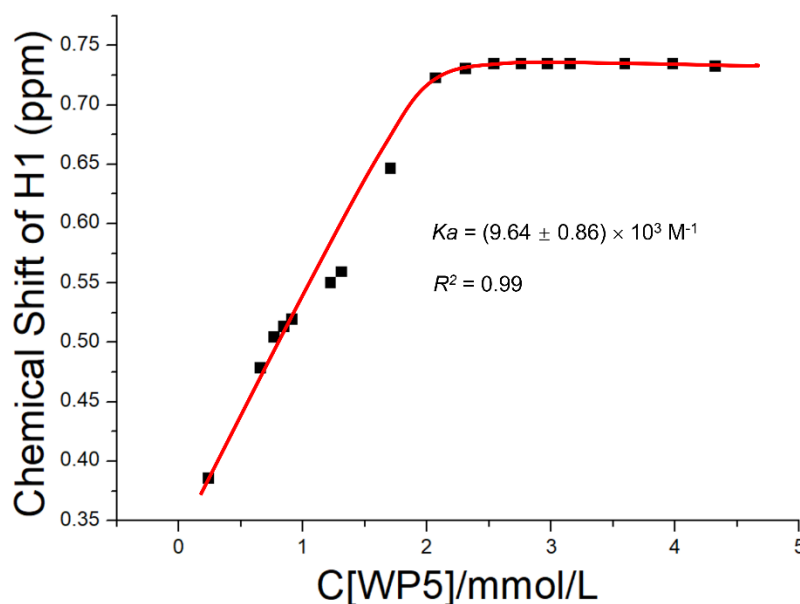


Fig. S26. The chemical shift changes of H₁ on ENDT₁ upon addition of **WP5**. The red solid line was obtained from the non-linear curve-fitting.

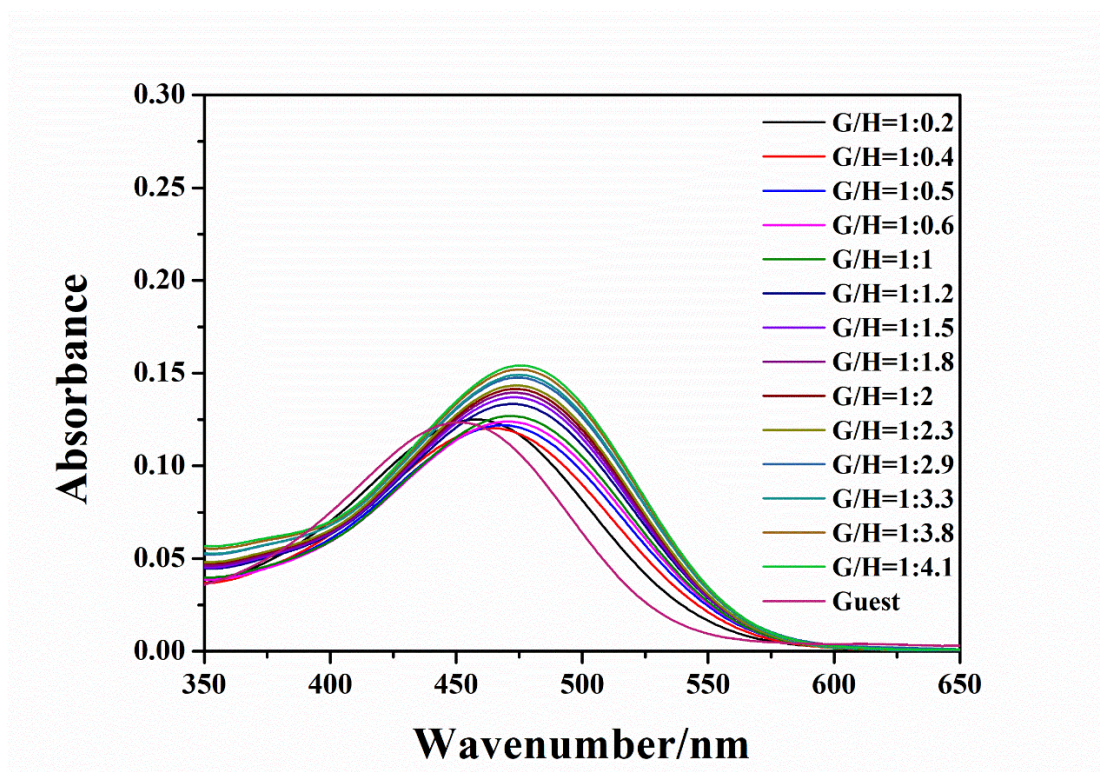


Fig. S27 UV-Vis titration of ENDT (10 μm) upon addition of WP5.

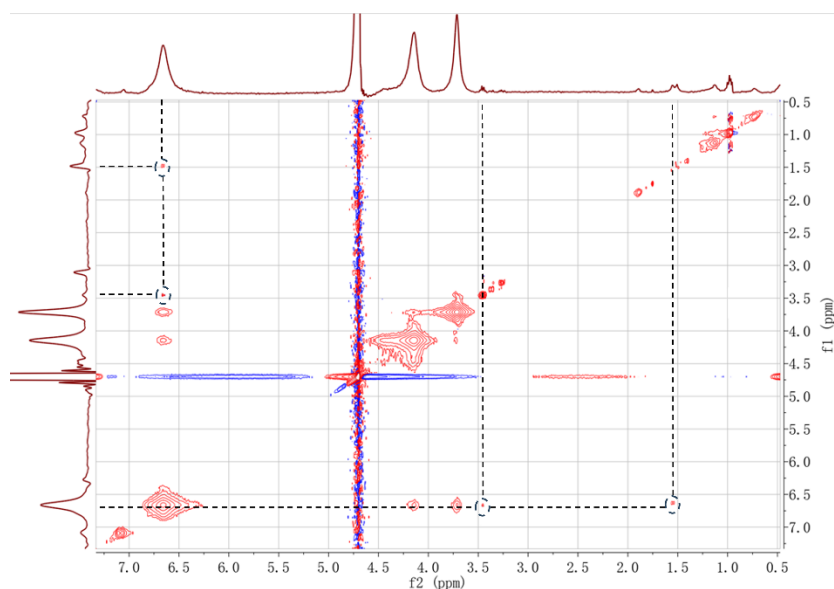


Fig. S28. 2D NOE NMR spectrum (400 MHz, D_2O , 298 K) of $(\text{WP5})_2 \gg \text{ENDT}_1$ at 1.0 mM.

6. References

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Nakamoto, *Chem. Commun.*, **2010**, *46*, 3708-3710.

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