Electronic Supplementary Information for

Binaphthyl-based chiral covalent organic frameworks for chiral drugs separation

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1. Materials and instruments

The reagents and solvents are commercially available and used without further purification. Building block 2, 4, 6-triformylphloroglucinol (TP) wassynthesized according to previously published procedure. (*R*) and (*S*)-DCDB were synthesized according to our reported procedure. ¹

The powder diffractometer (XRD) patterns were collected by a D8 ADVANCEX-ray with Cu Kα radiation $(\lambda = 1.5405 \text{ Å})$. The total surface areas of the catalysts were measured by the BET (Brunauer–Emmer–Teller) method using N_2 adsorption at 77 K, this was done by the Micromeritics ASAP 2000 sorption/desorption analyzer. HRTEM (High resolution transmission electron microscopy) analysis was performed on a JEOL 2100 Electron Microscope at an operating voltage of 200 kV. Scanning electron microscopy (SEM) images were taken on a SUB010 scanning electron microscope with acceleration voltage of 20 kV. Elemental analyses for C, H and N were obtained on a Perkin-Elmer analyzer model 240. Infrared (IR) samples were prepared as KBr pellets, and spectra were obtained in the 400-4000 cm-1 range using a Perkin-Elmer 1600 FTIR spectrometer. ¹³C solid-state NMR spectra were recorded on a MERCURY plus 400 spectrometer operating at resonance frequencies of 400 MHz. Thermogravimetric analyses(TGA) were carried out under flowing nitrogen at a heating rate of 10 °C·min−1 on a TA Instrument Q5 analyzer. High-resolution mass spectrometry (HRMS) analysis was carried out on a Bruker maXis ultrahigh-resolution-TOF mass spectrometer. The solid-state CD spectra were recorded on a J-815 spectropolarimeter (Jasco, Japan). ¹H NMR data were collected on an AM-400 spectrometer. Chemical shiftsare reported in δ relative to TMS. Enantiomer ratios were determined by chiral HPLC analysis using a Shimadzu LC-10AT VP series and a Shimadzu LC-10A VP UV-vis.

2. Synthesis of monomers

Under nitrogen, a mixture of (*R*)-DCDB (0.57g, 1.0 mmol), 4-aminophenylboronic acid pinacol ester (0.44 g, 2.0 mmol), Pd(PPh₃)₄ (0.08 g, 0.06 mmol) and K₂CO₃ (0.82 g, 6.0 mmol) in THF (30 mL)/H₂O (10.0 mL) was refluxed for 36 h. After removed the solvent, the residue was redissolved in CH₂Cl₂ and dried over MgSO₄. The obtained crude product was purified by the column on silica gel using CH_2Cl_2/THF (10:1, v/v) as eluent to give (*R*)-BINOLDE in 83% yield. ¹H NMR (400 MHz, CDCl₃) δ=7.88 (d, J = 7.1 Hz, 1H), 7.70 (t, J = 25.5 Hz, 1H), 7.34 (d, J = 16.0 Hz, 1H), 7.26-7.12 (m, 1H), 4.11 (d, J = 6.7 Hz, 1H), 1.13 (t, J = 6.1 Hz, 1H). IR (KBr pellet cm-1): 3203(vs), 2262(w), 1494(s), 1366(m), 1175(s), 1145(m), 1040(w), 946(m), 884(w), 826(w), 726(m), 637(w), 549(m). ¹³C NMR (400 MHz, CDCl3): δ 156.13, 144.54, 136.76, 132.97, 132.05, 129.06, 128.24, 127.75, 126.28, 122.85, 120.63, 119.97, 118.94, 62.7, 15.2. ESI-MS: m/z, Anal. Calcd: 615.14, Exp: 615.14, $[M+Na]^+$.

(*R*)-BINOLDH was obtained by stirring the suspended (*R*)-BINOLDE (0.61 g, 1.0 mmol) in CH2Cl² (10.0mL) with excess BBr₃ (570 µL, 6.0 mmol) for 24 h. Quenching reaction with ice water, the organic layer was separated, and the aqueous layer was completely extracted with CH_2Cl_2 , dried with MgSO₄ and concentrated in vacuo to provide (R)-BINOLDH (86%) as a beige crystalline solid. ¹H NMR (400 MHz, DMSO): δ=7.48-7.22 (m, 4H), 7.05 (dd, J = 42.9, 27.0 Hz, 1H), 0.00 (d, J = 63.5 Hz, 2H), 0.00 (dd, J = 44.3, 13.7 Hz, 3H), 0.00 (d, J = 43.5 Hz, 1H). IR (KBr pellet cm-1): 3203(vs), 2262(w), 1586(w), 1490(s), 1377(m), 1195(s), 1149(w), 1020(w), 946(w), 884(w), 816(w), 726(m), 636(m), 549(m). ¹³C NMR (400 MHz, CDCl3): δ 155.21, 146.84, 144.34, 137.62, 132.75, 131.26, 128.07, 127.04, 126.76, 124.31, 119.86, 118.73, 115.60. ESI-MS: m/z, Anal. Calcd: 537.10, Exp: 537.11, [M+Na]⁺. (S)-BINOLDH was synthesized following the same method mentioned above except that (*S*)-DCDB was used instead of (*R*)-DCDB.

3. Synthesis of (*R***)- and (***S***)-BHTP-COF**

An *o*-dichlorobenzene (*o*-DCB)/*n*-BuOH (0.5/0.5 mL) mixture of (*R*)- or (*S*)-BINOLDH (0.12 mmol, 64.5 mg), TP (0.08 mmol, 16.8 mg) and acetic acid (30 μL) in a Pyrex tube (35 mL) were degassed via three freezepump-thaw cycles. The tube was sealed and heated under $N₂$ at 120 °C in an oil bath for 3 days. The solids were separated via centrifugation after cooling to room temperature. The powder was washed several times with dichloromethane and ethanol via centrifugation and dried under vacuum to produce (*R*)- or (*S*)-

BHTP-COF in 92 % yield. Elemental Analysis (%) calcd for C₁₄H₁₁N₂O: C, 75.32; N, 12.55; H, 4.97. Found (%):

C, 76.15; N, 12.13; H, 5.31.

4. General procedure for column packing

The stationary material materials including (*R*)- or (*S*)-**BHTP-COF** materials were packed with the same method. (*R*)- or (*S*)**-BHTP-COF** was dispersed in a mixture of *n*-hexane/isopropanol (9:1, v/v) for 10 min. The suspension was then packed into an empty stainless-steel column (20 cm long x 4.2 mm id) under 40 MPa using *n*-hexane/isopropanol as the displacement liquid. The prepared columns were conditioned with *n*-hexane/isopropanol (9:1, v/v) at a flow rate of 0.5 mL/min for 2 h before chromatographic experiments.

Separation factor (*α*) and resolution factor (*Rs*) were obtained from the following equations:

 $\alpha = (t_{R2} - t_0)/(t_{R1} - t_0)$ *Rs* = 2(t_{R2} - t_{R1})/(w₁ + w₂)

Where t_{R1} and t_{R2} represent the retention times of right-handed or left-handed enantiomers ($t_{R2} > t_{R1}$), and w_1 and w_2 are the widths of the bases formed by triangulation of the peaks, respectively. The column void time is t_0 .

5. Figures S1-S5

Fig. S1 Characterization of (*R*)-**BHTP-COF**. (a) Measured and simulated PXRD patterns for (*R*)-**BHTP-COF**. Compared to the pattern generated from the *P3* space group (pink line), (*R*)-**BHTP-COF** unequivocally crystallizes in the *R3* space group (blue line). (b) TGA trace of (*R*)-**BHTP-COF**. (c) SEM image of (*R*)-**BHTP-COF**. (d) HR-TEM image of (*R*)-**BHTP-COF**. (e) N² adsorption and desorption isotherms of (*R*)-**BHTP-COF** at 77 K. Its pore width distribution is inserted.

Fig. S2 Characterization of (*S*)-**BHTP-COF**. (a) FT-IR spectra of (*S*)-**BHTP-COF** and its monomers. (b) Solidstate ¹³C CP-MAS NMR spectrum of (*S*)-**BHTP-COF**. (c) SEM image of (*S*)-**BHTP-COF**. (d) TGA trace of (*S*)- **BHTP-COF**. (e) SEM image of (*S*)-**BHTP-COF**. (f) Measured and simulated PXRD patterns for (*S*)-**BHTP-COF**. Compared to the pattern generated from the *P3* space group (pink line), (*S*)-**BHTP-COF** unequivocally

crystallizes in the R3 space group (blue line). (g) Crystal structure of (S)-BHTP-COF. (h) N₂ adsorption and desorption isotherms of (*S*)-**BHTP-COF** at 77 K. Its pore width distribution is inserted.

Fig. S3 (a) HPLC romatograms of ibuprofen enantiomers on the (*R*)-**BHTP-COF** packed column using the same eluent conditions obtained for 20 inject runs. (b) PXRD pattern of the (*R*)-**BHTP-COF** after 20 catalytic cycles.

Fig. S4 The molecular models of the ibuprofen *R*-enantiomer (a) and *S*-enantiomer (b) interacting with (*R*)-

BHTP-COF (The bond distances given in angstroms). All density functional theory (DFT) calculations were performed with the Gaussian16 package. ² The geometry optimizations were carried out in the gas phase at the B3LYP level of theory ^{3,4} with additional Grimme's D3 dispersion correction (Becke-Johnson damping), ⁵ with the def2-SVP basis set. ^{6, 7} The 3D structures shown were illustrated using CYLview. ⁸

Fig. S5 HPLC chromatograms of the six racemic drugs including ibuprofen(a), mandelic acid (b), ketoprofen(c), naproxen(d), propranolol (e), cetirizine (f) with the (*S*)-**BHTP-COF** packed column (mobile phase n-hexane/isopropanol (9:1, v/v), 0.3 mL/min flow rate, detection wavelength 254 nm, temperature $25 °C$).

6. Tables S1-S4

Table S1. The structure model of (*R*)-**BHTP-COF** with *R3*mode.

Table S2. The structure model of (*S*)-**BHTP-COF** with *R3*mode.

Table S3. Comparison of separation performance of three racemic drugs by HPLC columns packed with (*R*)- **BHTP-COF**, and several commercial CSPs.

C.S.: Cannot be seperated.

Table S4. Enantioseparation data on the (*S*)-**BHTP-COF** packed HPLC column.

7. References

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