Supporting Information

Light-induced β-Hydroxy Sulfone synthesis in DNA-Encoded

Libraries

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1. General Information

All reagents were obtained from commercial sources unless otherwise noted. The central dsDNA oligonucleotide with chemically modified phosphates that end in an amine terminus (HP, Figure S1) and encoding 5'-phosphorylated oligonucleotides were purchased from Genscript Biotech Corp. All other DNA oligonucleotides were obtained from General Biological System (Anhui) Co., Ltd. and assessed through the general analytical procedure for purity. DNA working solutions were prepared using DNAse/RNAse-free ultrapure water (Invitrogen), DMSO (Fisher). LC-MS running solvents were made from Optima LC-MS grade water (Fisher), Optima LC-MS grade methanol (Fisher), hexafluoroisopropanol (99+% purity, Sigma-Aldrich) and HPLC-grade triethylamine (Fisher). T4 DNA Ligase was obtained from ThermoFisher Scientific Coorporation (#EL0012). The 10X ligation buffer stock used in ligation reactions was composed as follows: 400 mM Tris-HCl, 100 mM MgCl₂, 100 mM DTT, 5 mM ATP (pH 7.8 at 25 °C). Eosin Y (90+% dye content) was purchased from Sigma Aldrich. All ionic solutions, including aq. NaCl (5.0 M) and basic borate buffer (250.0 mM sodium borate/boric acid, pH 9.5), were prepared in-house. The HP used for experiments has the following chemical structure.





Chemical building blocks and reagents were purchased from various vendor sources and used without further purification. Building blocks were generally used from aliquots dissolved in water (H₂O), dimethyl sulfoxide (DMSO), and stored in 2D barcoded tubes with septa-caps at -80 °C. Solutions were transferred by

Eppendorf pipettes. Reactions and large volume DNA precipitations were generally performed in centrifuge tubes (Axygen). All reactions were agitated in a custom-made thermo mixer at 1,000.0 to 2,000.0 RPM (thermomixer was made by

a local vendor; <u>https://www.made-in-china.com</u> /products-search/hot-chinaproducts/Thermo_Mixer.html). The reaction containers are Eppendorf tubes or 96-well plates. Eppendorf tubes can be well fitted into a thermomixer to allow the on-DNA chemistry performed in combinatorial fashion. For irradiation with green light Zhongzhan Illumination (green, λ max = 535 nm, Imax = 1000 mA, 3.0 W) was used.

Solutions were centrifuged in high-speed freezing centrifuge (Eppendorf). A Vanquish UHPLC system was integrated with LTQ XL ion trap mass spectrometer (ThermoFisher Scientific) for LC-MS analysis of oligonucleotides. DNA was visualized with Molecular Imager Gel Doc XR system (BIO-RAD) after staining in an ethidium bromide solution.

2. Analytical Conditions

On-DNA reactions conducted during validation, library synthesis, or single compound synthesis was analyzed by LC-MS (ESI-MS). Samples (ca. 100.0 pmol diluted with 40.0 μ L of water) were injected onto a reverse-phase chromatography column (DNA PacTMRP 4.0 μ m, 2.1 x 50.0 mm) and eluted (30-60% solvent B over 1.5 minutes, 0.60 mL/min flow rate; Solvent A: 0.75% V/V hexafluoroisopropanol (HFIP)/0.038% V/V triethylammonium acetate (TEAA)/5.0 μ M EDTA in deionized water; Solvent B: 0.75% V/V HFIP/0.038% TEAA/5 μ M EDTA in 90/10 methanol/ deionized water) with monitoring at UV 260.0 nm wavelength and TIC. Percent of conversion (% PDT = (purity of PDT/purity of SM) x 100) for on-DNA reactions were determined by LC-MS analysis. Chromatographic purification was likewise achieved using reverse-phase liquid chromatography (XBridge® BEH C18 3.5 μ m, 4.6 x 100.0 mm), Solvent A: 0.75% v/v hexafluoroisopropanol (HFIP)/0.18% v/v N, N-Diisopropylethylamine (DIEA) in deionized water; Solvent B: MeOH. The data analysis was performed by exporting the raw instrument data (.RAW) to an

automated biomolecule deconvolution and reporting software (ProMass) which uses a novel algorithm (ZNova) to produce artifact-free mass spectra.

3. DNA Precipitation Protocol

To a DNA reaction mixture was added 5% (v/v) 5.0 M NaCl solution and 2.5 times the reaction volume of absolute ethanol. The mixture was mixed thoroughly before stored at -80 °C overnight for DNA precipitation. The slurry was then centrifuged at 4000.0 rpm for 1 hour, followed by decanting the supernatant. The DNA pellets were dried in air after supernatant was decanted. Invitrogen UltraPure distilled water was added to reconstitute the DNA solids to the needed concentration. Generally, ethanol precipitation was performed after each chemical reaction and ligation.

4. General DNA Ligation Procedure



The **DNA oligo 1** (1.0 mM, 10.0 nmol, 1.0 equiv.) was dissolved in 10.0 μ L of distilled water, to which were added **DNA oligo 2** (11.0 μ L of a 1.0 mM solution in distilled water, 1.1 equiv.), followed by the addition of 4.0 μ L of 10 X ligation buffer and 12.0 U T4 DNA ligase, add H₂O to a final volume of 40.0 μ l. The solution was agitated at 25 °C for 1 hour. The reaction mixture was quenched thermally by incubation for 20 minutes at 65 °C completely inactivates T4 DNA Ligase and the reaction mixture was analyzed by LC-MS. The DNA product **DNA oligo 3** was precipitated with 5.0 M NaCl and ethanol. Then, it was directly used for the next step. The OD measurement was performed to quantify the reaction yield, and the product quality was assessed by LC-MS.

5. Experimental Procedures

5.1.Synthesis of DNA-NH₂



Preparation of **DNA-NHFmoc**: In a tube, was placed 1-(9H-fluoren-9-yl)-3oxo-2,7,10,13,16 -pentaoxa-4- azanonadecan-19-oic acid **S1** (200.0 mM in DMSO, 600.0 µL, 40.0 equiv.) in borate buffer (250.0 mM in H₂O, pH = 9.47, 3.0 mL, 250.0 equiv.), to which were added DMT-MM (200.0 mM in H₂O, 600.0 µL, 40.0 equiv.) and **HP** (1.0 mM in H₂O, 3.0 mL, 1.0 equiv.). The mixture was agitated at 4 °C for 2 hours. After ethanol precipitation, the reaction conversion rate (100%) was monitored by LC-MS, and the product **DNA-NHFmoc** (2,400.0 nmol) was collected for the next step use.

Preparation of **DNA-NH**₂: Into a tube was loaded with **DNA-NHFmoc** (1.0 mM in H₂O, 2,400.0 μ L), to which was added with 1,200.0 μ L of 10% (v/v) piperidine in water. The mixture was agitated at 25 °C for 5 hours. Afterwards, the product **DNA-NH**₂ was obtained by ethanol precipitation and then used for the next step without further purification. The reaction conversion rate was 96% determined by LC-MS analysis.

5.2. General Procedure for Synthesis of Vinylbenzoic Acid



Reagents and conditions: (a) Potassium vinyltrifluoroborate, Pd(dppf)Cl₂, K₂CO₃, 1,4-dioxane/H₂O (2:1), 80 °C; (b) LiOH·H₂O, CH₃CN/H₂O (1:1)

General Procedure A. To a solution of brominated aromatic hydrocarbons (1.0 equiv) and potassium vinyltrifluoroborate (1.5 equiv) in 1,4-dioxane and H₂O were added K_2CO_3 (3.0 equiv) and Pd(dppf)Cl₂ (0.05 equiv). After stirring for 4 hours at 80 °C under a nitrogen atmosphere, the resulting mixture was concentrated under reduced pressure. The residue was purified by silica gel column chromatography, eluted with hexane/ethyl acetate to afford the corresponding ester-based olefin.

To a stirred solution of ester-based olefin (1.0 equiv) in CH₃CN/H₂O (1:1) was added LiOH·H₂O (2.0 equiv) at 25 °C and stirred for 2 hours. After completion, the mixture was acidified to pH = 2.0 with 1.00 N HCl aqueous solution. The residue was purified by silica gel column chromatography to afford the corresponding carboxyl olefin.

3-Cyano-5-vinylbenzoic acid (S2m)

Prepared according to general procedure A: Using methyl 3-bromo-5cyanobenzoate (590 mg, 2.47 mmol) to afford carboxyl olefin **S2m** (325 mg, 76%) as a white solid. ¹H NMR (400 MHz, DMSO-d6) δ 13.56 (s, 1H), 8.26 (dt, J = 4.5, 1.7 Hz, 2H), 8.16 (t, J = 1.6 Hz, 1H), 6.86 (dd, J = 17.7, 11.1 Hz, 1H), 6.11 (d, J = 17.7 Hz, 1H), 5.48 (d, J = 11.1 Hz, 1H). MS (ESI-MS) m/z: 174 [M + H]⁺.



3-Methyl-4-vinylbenzoic acid (S2p)

Prepared according to general procedure A: Using methyl 4-bromo-3methylbenzoate (2.00 g, 8.77 mmol) to afford carboxyl olefin **S2p** (803 mg, 57%) as a white solid. ¹H NMR (400 MHz, DMSO-d6) δ 12.84 (s, 1H), 7.79 – 7.71 (m, 2H), 7.62 (d, J = 7.9 Hz, 1H), 6.99 (dd, J = 17.5, 11.0 Hz, 1H), 5.83 (dd, J = 17.4, 1.3 Hz, 1H), 5.44 (dd, J = 11.1, 1.3 Hz, 1H), 2.36 (s, 3H). MS (ESI-MS) m/z: 163 [M + H]⁺.



3-Chloro-4-vinylbenzoic acid (S2r)

Prepared according to general procedure A: Using methyl 4-bromo-3chlorobenzoate (3.00 g, 12.10 mmol) to afford carboxyl olefin **S2r** (1.49 g, 68%) as a white solid. ¹H NMR (400 MHz, DMSO-d6) δ 13.30 (s, 1H), 7.97 – 7.81 (m, 2H), 7.05 (dd, J = 17.5, 11.1 Hz, 1H), 6.03 (dd, J = 17.5, 0.9 Hz, 1H), 5.59 (dd, J = 11.1, 0.9 Hz, 1H). MS (ESI-MS) m/z: 183 [M + H]⁺.



S2w

4-Vinylfuran-2-carboxylic acid (S2w)

Prepared according to general procedure A: Using methyl 4-bromofuran-2carboxylate (2.50 g, 12.30 mmol) to afford carboxyl olefin **S2w** (1.39 g, 82%) as a white solid. ¹H NMR (400 MHz, DMSO-d6) δ 13.22 (s, 1H), 8.20 – 7.93 (m, 1H), 7.54 – 7.37 (m, 1H), 6.57 (dd, J = 17.6, 10.9 Hz, 1H), 5.70 (dd, J = 17.6, 1.4 Hz, 1H), 5.22 (dd, J = 10.9, 1.4 Hz, 1H). MS (ESI-MS) m/z: 139 [M + H]⁺.



5-Vinylthiophene-3-carboxylic acid (S2x)

Prepared according to general procedure A: Using methyl 5-bromothiophene-3-carboxylate (2.00 g, 9.09 mmol) to afford carboxyl olefin **S2x** (930 mg g, 66%) as a white solid. ¹H NMR (400 MHz, DMSO-d6) δ 12.74 (s, 1H), 8.20 – 8.05 (m, 1H), 7.40 (d, J = 1.4 Hz, 1H), 6.89 (dd, J = 17.5, 10.9 Hz, 1H), 5.60 (d, J = 17.5 Hz, 1H), 5.23 (d, J = 10.9 Hz, 1H). MS (ESI-MS) m/z: 155 [M + H]⁺.



4-Vinylthiophene-2-carboxylic acid (S2y)

Prepared according to general procedure A: Using methyl 4-bromothiophene-2-carboxylate (2.00 g, 9.09 mmol) to afford carboxyl olefin **S2y** (1.04 g g, 74%) as a white solid. ¹H NMR (400 MHz, DMSO-d6) δ 13.14 (s, 1H), 7.93 (d, J = 1.5 Hz, 1H), 7.80 (d, J = 1.4 Hz, 1H), 6.70 (dd, J = 17.7, 11.0 Hz, 1H), 5.76 (dd, J = 17.7, 1.2 Hz, 1H), 5.24 (dd, J = 10.9, 1.2 Hz, 1H). MS (ESI-MS) m/z: 155 [M + H]⁺.



S2s

3-Bromo-4-vinylbenzoic acid (S2s)

A solution of methyltriphenylphosphanium bromide (3.56 g, 9.97 mmol, 1.2 equiv) in 1, 4-dioxane (40 mL) was treated with K_2CO_3 (1.72 g, 12.47 mmol, 1.5 equiv) for 30 minutes under nitrogen atmosphere followed by the addition of methyl 3-bromo-4-formylbenzoate (2.00 g, 8.31 mmol, 1.0 equiv) at room temperature for overnight. After completion, the resulting mixture was concentrated under reduced pressure. The residue was purified by reversed-phase flash chromatography to afford methyl 3-bromo-4-vinylbenzoate (1.53 g, 76%) as a white solid.

To a stirred solution of methyl 3-bromo-4-vinylbenzoate (1.50 g, 6.25 mmol, 1.0 equiv) in CH₃CN/H₂O (1:1) was added LiOH·H₂O (525 mg, 12.5 mmol, 2.0 equiv) at 25 °C and stirred for 2 hours. After completion, the mixture was acidified to pH=2 with 1 N HCI. The residue was purified by silica gel column chromatography to afford 3-bromo-4-vinylbenzoic acid (**S2s**, 1.24 g, 88%) as a white solid. ¹H NMR (400 MHz, DMSO-d6) δ 13.28 (s, 1H), 8.09 (d, J = 1.6 Hz, 1H), 7.93 – 7.87 (m, 1H), 7.87 – 7.80 (m, 1H), 7.01 (dd, J = 17.4, 11.0 Hz, 1H), 6.00 (dd, J = 17.4, 0.9 Hz, 1H), 5.58 (dd, J = 11.0, 0.9 Hz, 1H). MS (ESI-MS) m/z: 227 [M + H]⁺.

5.3. General Acylation Procedure of DNA-Conjugated Olefins



The modified carboxyl olefin (200.0 mM in DMSO, 10.0 μ L, 40.0 equiv.) and DMT-MM (200.0 mM in H₂O, 10.0 μ L, 40.0 equiv.) were mixed, which followed by

the addition of **DNA-NH**² (1 mM in H₂O, 50.0 μ L, 50.0 nmol, 1.0 equiv.) and borate buffer (250 mM, pH = 9.5, 50.0 μ L). The reaction was quenched in 1 hour and precipitated with cold ethanol. The reaction conversion rates were in the range of 92% to >99%, determined by LC-MS analysis.

5.4. General Procedure for Synthesis of Sodium Sulfinates

$$\begin{array}{c} O \\ R - \overset{O}{\overset{}_{\text{H}}} - \text{Cl} \\ O \\ H_2 O, 50 \ ^{\circ}\text{C}, 4 \ \text{h} \end{array} \xrightarrow{O} R \xrightarrow{O} Na^{+}$$

General Procedure B. A solution of Na₂SO₃ (1.5 equiv) in H₂O was treated with Na₂CO₃ (2.0 equiv) for 20 minutes at 50 °C under nitrogen atmosphere followed by the addition of the corresponding sulfonyl chloride (1.0 equiv) dropwise and the resulting mixture was stirred for 2 hours at 50 °C. After completion of the reaction, the solvents were removed in vacuo, and the white residue was redissolve in hot EtOH. The resulting mixture was filtered, the filtrate was concentrated under reduced pressure to provide the corresponding sulfonate.

M. W. = 170.2

Sodium 2-cyclobutylethane-1-sulfinate (4c)

Prepared according to general procedure B: Using 2-cyclobutylethane-1sulfonyl chloride (1.00 g, 5.93 mmol) to afford sodium 2-cyclobutylethane-1sulfinate (460 mg) as a crude product without further purification. Crude ¹H NMR (400 MHz, Deuterium Oxide) δ 2.39 – 2.25 (m, 3H), 2.11 – 1.98 (m, 2H), 1.93 – 1.75 (m, 2H), 1.70 – 1.57 (m, 4H).

$$M. W. = 156.2$$

Sodium cyclopentanecarboxylate (4h)

Prepared according to general procedure B: Using cyclopentanesulfonyl chloride (1.00 g, 5.49 mmol) to afford sodium cyclopentanecarboxylate (576 mg) as a crude product without further purification. Crude ¹H NMR (400 MHz, Deuterium Oxide) δ 3.45 – 2.55 (m, 1H), 2.08 – 1.68 (m, 5H), 1.66 – 1.55 (m, 3H).

5.5. General Reaction Conditions for the Photocatalytic Sulfone Synthesis



General reaction conditions: A 0.6-mL centrifuge tube was charged with the DNA starting materials **1** (10.0 nmol, 1.0 equiv., 2.0 mM in H₂O, 5.0 μ L, 0.8 mM final concentration), **2** or **4** (600.0 mM in H₂O, 5.0 μ L, 240.0 mM final concentration), nitrobenzene (Ph-NO₂, 2,000.0 mM in DMSO, 0.5 μ L, 80.0 mM final concentration) and photocatalyst (Eosin Y/HCI, 1:2, premixed, 250.0 mM in DMSO, 2.0 μ L, Eosin Y with 40.0 mM final concentration). The reaction mixture was agitated and irradiated using green LEDs (535 nm) for 1 h at 40 °C. Afterwards, the sample was analyzed by LC-MS after EtOH precipitation. The reaction conversion rates were in the range of 36% to >99%, determined by LC-MS analysis.

5.6. Scale-up Procedure of On-DNA Sulfone



A 0.6-mL centrifuge tube was charged with the DNA starting materials **1o** (100.0 nmol, 1.0 equiv., 2.0 mM in H₂O, 50.0 μ L, 0.8 mM final concentration), **2** (600.0 mM in H₂O, 50.0 μ L, 240.0 mM final concentration), nitrobenzene (Ph-NO₂, 2,000.0 mM in DMSO, 5.0 μ L, 80.0 mM final concentration) and photocatalyst (Eosin Y/HCl, 1:2, premixed, 250.0 mM in DMSO, 20.0 μ L, Eosin Y with 40.0 mM final concentration). The reaction mixture was agitated and irradiated using green LEDs (535 nm) for 1 h at 40 °C. Afterwards, the sample was analyzed by LC-MS

after EtOH precipitation. The reaction conversion rate was 84% determined by LC-MS analysis.

6. Structure, Purity, and Conversion



6.1. The Structure of Carboxyl Olefins

6.2. The Structure and Purity of DNA-Conjugated Compounds 1a-1z



6.3. The Structure of Sodium Sulfinates



6.4. The Structure and Conversion Rate of DNA-Conjugated 3a-3z



6.5. The Structure and Conversion Rate of DNA-Conjugated 5a-5v



^aWhen the sodium sulfinate (**4h**) used was a crude product synthesized in the

laboratory, the conversion rate of **5h** was 94%. When the sodium sulfinate (**4h**) used was obtained from commercial source (99% purity), the conversion rate of **5h** was 95%.

7. Synthesis of A Prototype DEL

Primer & Tags. The DNA Tags all 5 -ends were phosphorylated.

| | Primer-F 13nt | Tag A-F 17nt | Tag B-F 10nt | |
|----|------------------|--------------------------------|-----------------|----|
| 5' | TACGYYYYYYGGA | TGCGXXXXXXXXXTG | ΤΥΥΥΥΥΥΥΥΥ | 3' |
| 3' | GG ATGCXXXXXXCCT | ACGC YYYYYYYYYA <mark>(</mark> | CAXXXXXXXXXXAGG | 5' |
| | Primer-R 19nt | Tag A-R 10nt | Tag B-R 16nt | |

Primer Ligation: The **DNA-NH**² (M.W. = 5184, 1.0 mM, 100.0 nmol, 1.0 equiv.) was dissolved in 100.0 µL water. Primer duplexes were added (M.W. = 9949, 110.0 µL of a 1.0 mM solution in water, 110.0 nmol, 1.1 equiv.), followed by 40.0 µL 10X ligation buffer and 120.0 U T4 DNA ligase, and H₂O to a final volume of 400.0 µl. The reaction mixture was incubated at 25 °C for 1 hour. The DNA product was precipitated with NaCl and ethanol and taken on to the first cycle of library synthesis without further purification. The product was 96 nmol determined by O.D. measurement.

DEL Cycle 1: A 1.0 mM solution of the primer-elongated AOP headpiece (**P**-**DNA-NH**₂, 96.0 nmol, 96.0 µL) was split into 6 wells (16.0 nmol/well). To each well was then added 6.4 µL of 10X ligation buffer, 19.2 U T4 DNA ligase, 1 of 6 Tag A solutions (M.W. = 8404, 17.6 µL of 1.0 mM stock solutions in water), and H₂O to a final volume of 64.0 µl. The ligation was agitated at 25 °C for 1 hour. The DNA was precipitated to afford the pellet (**A-P-DNA-NH**₂), which was then dissolved in 16.0 µL of distilled H₂O.

To a solution of 1 of 6 carboxyl olefins (200.0 mM in DMSO, 3.2 μ L, 40.0 eq) and DMT-MM (200.0 mM in H₂O, 3.2 μ L, 40.0 equiv.), which followed by the addition of **A-P-DNA-NH₂** (1.0 mM in H₂O, 16.0 μ L, 16.0 nmol/well, 1.0 equiv.) and borate buffer (pH = 9.5, 16.0 μ L). The acylation mixture was agitated at 25 °C for 1 hour. All reactions were monitored by LC-MS. After completion, the reactions

were pooled and then precipitated with ethanol to yield 72.0 nmol of crude cycle 1 products, the cycle 1 products were dissolved in 72.0 μ L of distilled H₂O. The OD measurement was used to quantify the reaction yields.

DEL Cycle 2: The cycle 1 products (72.0 nmol, 72.0 μ L) were split into 6 wells (12.0 nmol/well, 12.0 μ L/well). To each well was then added 4.8 μ L of 10X ligation buffer, 14.4 U T4 DNA ligase, 1 of 6 Tag B solutions (M.W. = 8103.0, 13.2 μ L of 1.0 mM stocks in water), and H₂O to a final volume of 48.0 μ l. The reaction mixture was agitated at 25 °C for 1 hour. The DNA was precipitated to afford the pellet.

Then, to each well (**B-A-P-DNA-NH₂-BB1**, 12.0 nmol, 1.0 equiv., 2.0 mM in H₂O, 6.0 μ L, 0.8 mM final concentration) was added 1 of 6 sodium sulfinates (600.0 mM in H₂O, 6.0 μ L, 240.0 mM final concentration), nitrobenzene (Ph-NO₂, 2,000.0 mM in DMSO, 0.6 μ L, 80.0 mM final concentration) and photocatalyst (Eosin Y/HCl, 1:2, premixed, 250.0 mM in DMSO, 2.4 μ L, Eosin Y with 40.0 mM final concentration). The reaction mixture was agitated and irradiated using green LEDs (535 nm) for 1 h at 40 °C. All reactions were monitored by LC-MS. After completion, the reactions were pooled, precipitated with ethanol, and purified by reverse-phase HPLC, yielded 45.0 nmol of products.

8. Control Experiment

8.1. General Procedure for the Synthesis of S4



Into a tube was loaded with **S3** (20.0 μ L, 200.0 mM in DMSO, 80.0 equiv.), HATU (12.5 μ L, 200.0 mM in DMSO, 50.0 equiv.), DIEA (125.0 μ L, 200.0 mM in DMSO, 500.0 equiv.) and **DNA-NH**₂ (50.0 μ L, 1.0 mM in H₂O, 1.0 equiv.). The reaction mixture was agitated at 25.0 °C for 2 hours and monitored by LC-MS. After ethanol precipitation, the obtained solid was dissolved in distilled water and then directly used for the next step without further purification.

8.2. General Procedure for Photoreaction of S4



A 0.6-mL centrifuge tube was charged with the DNA starting materials **S4** (10.0 nmol, 1.0 equiv., 2.0 mM in H₂O, 5.0 μ L, 0.8 mM final concentration), **2** (600.0 mM in H₂O, 5.0 μ L, 240.0 mM final concentration), nitrobenzene (Ph-NO₂, 2,000.0 mM in DMSO, 0.5 μ L, 80.0 mM final concentration) and photocatalyst (Eosin Y/HCl, 1:2, premixed, 250.0 mM in DMSO, 2.0 μ L, Eosin Y with 40.0 mM final concentration). The reaction mixture was agitated and irradiated using green LEDs (535 nm) for 1 h at 40 °C. Afterwards, the sample was analyzed by LC-MS after EtOH precipitation. The reaction was determined by LC-MS analysis.

9. Characterization Data

9.1. Mass Spectrum of Starting Material (HP)

HP Conversion rate: 100% Expected mass: 4937.2 Observed mass: 4937.5

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|----------------------|--------------------|--------------------------|
| 1.2 | 4937. | 3.76E+00 | ok | 2.80E+00 | 100.0 |
| 9 | 5 | 5 | | 5 | 0 |



Figure S2. LC-MS Spectrum of compound HP.

9.2. Mass Spectrum of DNA-NHFmoc



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/U V Area Percent |
|-------------|---------------------------|-----------|----------------------|--------------------|---------------------------|
| 1.33 | 5407. | 1.08E+00 | ok | 1.69E+00 | 60.04 |
| 4 | 8 | 5 | ОК | 5 | 09.04 |
| 1.50 | 5406. | 2.02E+00 | ok | 7.57E+00 | 20.06 |
| 0 | 7 | 3 | ŬK | 4 | 30.90 |

Deconvoluted mass spectrum of product:



Figure S3. LC-MS Spectrum of **DNA-NHFmoc**.

9.3. Mass Spectrum of DNA-NH₂

NIN T NH₂ $DNA-NH_2$ Conversion rate: 96% Expected mass: 5184.5 Observed mass: 5185.2

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.31 | 5185.2 | 5.43E+005 | ok | 1.04E+006 | 97.61 |
| 1.36 | 5185.2 | 2.82E+005 | ok | 1.63E+004 | 1.53 |
| 1.44 | 5184.9 | 4.26E+004 | ok | 9.16E+003 | 0.86 |



Figure S4. LC-MS Spectrum of DNA-NH₂.

9.4.¹H NMR Study of Off-DNA Carboxyl Olefins

¹H NMR spectrum for 3-cyano-5-vinylbenzoic acid (**S2m**), DMSO-d6, 400 MHz:



Figure S5. ¹H NMR spectrum of **S2m**.

1H NMR spectrum for 3-methyl-4-vinylbenzoic acid (S2p), DMSO-d6, 400 MHz:



Figure S6. ¹H NMR spectrum of **S2p**.

¹H NMR spectrum for 3-chloro-4-vinylbenzoic acid (**S2r**), DMSO-d6, 400 MHz:



Figure S7. ¹H NMR spectrum of **S2r**.

¹H NMR spectrum for 4-vinylfuran-2-carboxylic acid (**S2w**), DMSO-d6, 400 MHz:



Figure S8. ¹H NMR spectrum of **S2w**.

¹H NMR spectrum for 5-vinylthiophene-3-carboxylic acid (**S2x**), DMSO-d6, 400 MHz:



Figure S9. ¹H NMR spectrum of **S2x**.

¹H NMR spectrum for 4-vinylthiophene-2-carboxylic acid (**S2y**), DMSO-d6, 400 MHz:





¹H NMR spectrum for 3-bromo-4-vinylbenzoic acid (**S2s**), DMSO-d6, 400 MHz:



Figure S11. ¹H NMR spectrum of **S2s**.

9.5. Mass Spectrum of DNA-Conjugated Olefins



LC Spectrum:



S26

| RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|----------------------|--------------------|--------------------------|
| 1.78 | 5238. | 9.94E+00 | ok | 2.00E+00 | 100.0 |
| 0 | 8 | 3 | | 5 | 0 |



Figure S12. LC-MS Spectrum of Compound 1a.

1b

Purity: >99% Expected mass: 5266.6 Observed mass: 5267.0

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|----------------------|--------------------|--------------------------|
| 1.41 | 5267. | 1.00E+00 | ok | 1.43E+00 | 100.0 |
| 4 | 0 | 5 | | 5 | 0 |



Figure S13. LC-MS Spectrum of Compound 1b.



LC Spectrum:



| | RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/UV Area Percent |
|---|-------------|---------------------------|-----------|----------------------|--------------------|--------------------------|
| | 1.3 | 5292. | 5.98E+00 | ok | 1.10E+00 | 100.0 |
| 5 | | 5 | 5 | | 5 | 0 |

Deconvoluted mass spectrum of product:



Figure S14. LC-MS Spectrum of Compound 1c.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|----------------------|--------------------|--------------------------|
| 1.3 | 5306. | 7.01E+00 | ok | 1.21E+00 | 100.0 |
| 6 | 4 | 5 | | 5 | 0 |



Figure S15. LC-MS Spectrum of Compound 1d.

1e

Purity: 93% Expected mass: 5278.6 Observed mass: 5278.2

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.35 | 5278.2 | 2.80E+005 | ok | 1.30E+005 | 92.97 |
| 1.38 | 5277.7 | 6.80E+004 | ok | 9.87E+003 | 7.03 |



Figure S16. LC-MS Spectrum of Compound 1e.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|----------------------|--------------------|--------------------------|
| 1.4 | 5306. | 4.84E+00 | ok | 1.18E+00 | 100.0 |
| 0 | 4 | 5 | | 5 | 0 |

Deconvoluted mass spectrum of product:



Figure S17. LC-MS Spectrum of Compound 1f.







| RT (min) | Base Peak Mass (Da) | Intensity | Spectra I Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|----------------------|--------------------|--------------------------|
| 1.3 | 5278. | 7.46E+00 | ok | 1.25E+00 | 100.0 |
| 3 | 5 | 5 | | 5 | 0 |



Figure S18. LC-MS Spectrum of Compound 1g.

1h Purity: >99% Expected mass: 5278.6 Observed mass: 5278.1

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.34 | 5278.1 | 2.39E+005 | ok | 1.25E+005 | 100.00 |



Figure S19. LC-MS Spectrum of Compound 1h.


LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.489 | 5315.1 | 1.26E+005 | ok | 1.88E+005 | 100.00 |



Figure S20. LC-MS Spectrum of Compound 1i.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.470 | 5315.0 | 9.75E+004 | ok | 1.62E+005 | 100.00 |



Figure S21. LC-MS Spectrum of Compound 1j.

1k Purity: >99% Expected mass: 5328.7 Observed mass: 5328.2



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.42 | 5328.2 | 3.30E+005 | ok | 9.92E+004 | 100.00 |



Figure S22. LC-MS Spectrum of Compound 1k.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.38 | 5332.3 | 7.13E+005 | ok | 1.42E+005 | 100.00 |



Figure S23. LC-MS Spectrum of Compound 1I.

₹ N H CN 1m Purity: 94% Expected mass: 5339.7 Observed mass: 5340.0



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.37 | 5340.0 | 5.95E+005 | ok | 1.21E+005 | 93.72 |
| 1.41 | 5339.8 | 6.37E+004 | ok | 8.12E+003 | 6.28 |



Figure S24. LC-MS Spectrum of Compound 1m.

1n Purity: >99% Expected mass: 5332.7 Observed mass: 5332.1



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.40 | 5332.1 | 2.61E+005 | ok | 1.31E+005 | 100.00 |







10 Purity: >99% Expected mass: 5314.7 Observed mass: 5315.2

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.484 | 5315.2 | 1.04E+005 | ok | 7.62E+005 | 100.00 |



Figure S26. LC-MS Spectrum of Compound 10.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.41 | 5327.9 | 2.24E+005 | ok | 1.49E+005 | 100.00 |



Figure S27. LC-MS Spectrum of Compound **1p**.

1q Purity: >99% Expected mass: 5332.7 Observed mass: 5332.4



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.350 | 5332.4 | 5.89E+005 | ok | 1.49E+005 | 100.00 |







LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.40 | 5349.0 | 5.23E+005 | ok | 1.50E+005 | 100.00 |



Figure S29. LC-MS Spectrum of Compound 1r.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.41 | 5394.2 | 4.09E+005 | ok | 1.29E+005 | 100.00 |



Figure S30. LC-MS Spectrum of Compound 1s.

1t

Purity: >99% Expected mass: 5342.7 Observed mass: 5341.5



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.429 | 5341.5 | 3.63E+004 | ok | 5.93E+004 | 100.00 |



Figure S31. LC-MS Spectrum of Compound 1t.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.267 | 5225.7 | 4.48E+004 | ok | 7.34E+003 | 5.32 |
| 1.315 | 5315.2 | 6.27E+005 | ok | 1.31E+005 | 94.68 |



Figure S32. LC-MS Spectrum of Compound 1u.







| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.31 | 5335.3 | 8.49E+003 | ok | 7.05E+003 | 5.01 |
| 1.38 | 5304.4 | 5.05E+005 | ok | 1.34E+005 | 94.99 |



Figure S33. LC-MS Spectrum of Compound 1v.

1w Purity: >99%

Expected mass: 5304.6 Observed mass: 5304.4



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.332 | 5304.4 | 5.50E+005 | ok | 1.25E+005 | 100.00 |







LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.37 | 5321.1 | 5.05E+005 | ok | 1.25E+005 | 92.07 |
| 1.42 | 5320.6 | 8.92E+004 | ok | 1.07E+004 | 7.93 |



Figure S35. LC-MS Spectrum of Compound 1x.

1y Purity: >99% Expected mass: 5320.7 Observed mass: 5320.6



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.334 | 5320.6 | 6.70E+005 | ok | 1.57E+005 | 100.00 |



Figure S36. LC-MS Spectrum of Compound 1y.

1z

Purity: >99% Expected mass: 5315.6 Observed mass: 5315.0



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.34 | 5315.0 | 3.22E+005 | ok | 1.37E+005 | 100.00 |



Figure S37. LC-MS Spectrum of Compound 1z.

9.6.¹H NMR Study of Sodium Sulfinates

¹H NMR spectrum for sodium 2-cyclobutylethane-1-sulfinate (**4c**), Deuterium Oxide, 400 MHz:



Figure S38. ¹H NMR spectrum of **4c**.

¹H NMR spectrum for sodium cyclopentanecarboxylate (**4h**), Deuterium Oxide, 400 MHz:



Figure S39. ¹H NMR spectrum of **4h**.

9.7. Mass Spectrum of 3a-3z

www CI 3a

Purity: 74% Conversion rate: 74% Expected mass: 5431.2 Observed mass: 5430.0

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.038 | 5184.4 | 3.68E+004 | ok | 1.07E+005 | 26.29 |
| 1.109 | 5430.0 | 1.26E+005 | ok | 2.99E+005 | 73.71 |



Figure S40. LC-MS Spectrum of DEL Compound 3a.

N H óнó CI

3b Purity: 80% Conversion rate: 80% Expected mass: 5459.2 Observed mass: 5458.0



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.29 | 5281.7 | 1.91E+004 | ok | 6.51E+003 | 4.93 |
| 1.32 | 5265.7 | 6.93E+004 | ok | 1.93E+004 | 14.60 |
| 1.36 | 5458.0 | 9.90E+004 | ok | 1.06E+005 | 80.48 |



Figure S41. LC-MS Spectrum of DEL Compound **3b**.

CI 3c Purity: 76%

Conversion rate: 76% Expected mass: 5485.3 Observed mass: 5486.1



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.034 | 5390.3 | 4.28E+004 | ok | 4.28E+004 | 9.64 |
| 1.074 | 5308.9 | 2.62E+005 | ok | 6.16E+004 | 13.88 |
| 1.111 | 5486.1 | 2.77E+005 | ok | 3.39E+005 | 76.47 |



Figure S42. LC-MS Spectrum of DEL Compound 3c.



Conversion rate: 76% Expected mass: 5499.3 Observed mass: 5500.7 and 5499.2

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 0.988 | 5184.3 | 5.45E+004 | ok | 2.19E+004 | 5.23 |
| 1.062 | 5404.4 | 5.11E+004 | ok | 2.31E+004 | 5.52 |
| 1.096 | 5322.9 | 3.08E+005 | ok | 5.56E+004 | 13.28 |
| 1.123 | 5500.7 | 5.90E+005 | ok | 1.96E+005 | 46.90 |
| 1.139 | 5499.2 | 1.86E+005 | ok | 1.22E+005 | 29.07 |





OH 3e Purity: 95% Conversion rate: >99%

Conversion rate: >99% Expected mass: 5471.2 Observed mass: 5471.0



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.232 | 5375.0 | 5.47E+003 | ok | 2.59E+003 | 1.85 |
| 1.250 | 5279.8 | 1.52E+004 | ok | 4.46E+003 | 3.20 |
| 1.310 | 5471.0 | 2.81E+005 | ok | 1.32E+005 | 94.95 |

Deconvoluted mass spectrum of SM:



Figure S44. LC-MS Spectrum of DEL Compound 3e.

MANN NH 0 || 3f

Purity: >99% Conversion rate: >99% Expected mass: 5499.3 Observed mass: 5498.7

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.325 | 5498.7 | 1.17E+005 | ok | 3.62E+004 | 100.00 |









| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.219 | 5184.3 | 1.94E+005 | ok | 1.00E+005 | 45.33 |
| 1.296 | 5278.2 | 7.71E+004 | ok | 3.04E+004 | 13.76 |
| 1.325 | 5471.1 | 4.32E+005 | ok | 7.86E+004 | 35.59 |
| 1.363 | 5470.3 | 3.82E+004 | ok | 1.17E+004 | 5.32 |



Figure S46. LC-MS Spectrum of DEL Compound **3g**.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.047 | 5311.3 | 1.09E+005 | ok | 1.15E+005 | 21.03 |
| 1.117 | 5471.7 | 3.97E+005 | ok | 4.30E+005 | 78.97 |



Figure S47. LC-MS Spectrum of DEL Compound **3h**.


Purity: 83% Conversion rate: 83% Expected mass: 5507.3 Observed mass: 5507.6

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.357 | 5184.5 | 2.25E+004 | ok | 4.50E+004 | 11.22 |
| 1.447 | 5315.9 | 4.86E+003 | ok | 1.50E+004 | 3.74 |
| 1.498 | 5507.6 | 9.37E+004 | ok | 3.33E+005 | 83.04 |
| 1.728 | 5506.3 | 2.93E+002 | ok | 8.00E+003 | 1.99 |



Figure S48. LC-MS Spectrum of DEL Compound 3i.



3j Purity: 96% Conversion rate: 96% Expected mass: 5507.3 Observed mass: 5507.9

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.347 | 4979.2 | 2.99E+003 | ok | 6.42E+003 | 0.90 |
| 1.428 | 5411.2 | 2.47E+003 | ok | 1.72E+004 | 2.41 |
| 1.486 | 5507.9 | 1.61E+005 | ok | 6.84E+005 | 95.84 |
| 1.738 | 11012.7 | 9.23E+002 | ok | 6.06E+003 | 0.85 |



Figure S49. LC-MS Spectrum of DEL Compound 3j.

3k

Purity: 90% Conversion rate: 90% Expected mass: 5521.3 Observed mass: 5521.3 and 5520.5



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.23 | 4978.1 | 2.66E+004 | ok | 1.01E+004 | 1.89 |
| 1.27 | 5225.8 | 1.12E+005 | ok | 1.72E+004 | 3.21 |
| 1.29 | 5424.9 | 3.41E+004 | ok | 9.82E+003 | 1.83 |
| 1.34 | 5329.9 | 1.78E+004 | ok | 8.99E+003 | 1.68 |
| 1.40 | 5521.3 | 1.66E+006 | ok | 4.18E+005 | 78.06 |
| 1.42 | 5520.5 | 5.90E+005 | ok | 6.50E+004 | 12.13 |
| 1.48 | 5519.8 | 3.53E+004 | ok | 6.42E+003 | 1.20 |



Figure S50. LC-MS Spectrum of DEL Compound 3k.

31 Purity: 96% Conversion rate: 96% Expected mass: 5525.3 Observed mass: 5525.2



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.237 | 4978.8 | 2.55E+004 | ok | 3.76E+003 | 0.81 |
| 1.289 | 5075.6 | 1.28E+004 | ok | 2.30E+003 | 0.49 |
| 1.321 | 5334.5 | 4.16E+004 | ok | 1.08E+004 | 2.32 |
| 1.357 | 5525.2 | 1.47E+006 | ok | 4.48E+005 | 96.38 |



Figure S51. LC-MS Spectrum of DEL Compound 3I.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.215 | 4978.8 | 1.44E+004 | ok | 1.22E+003 | 0.60 |
| 1.262 | 5435.9 | 7.10E+003 | ok | 1.77E+003 | 0.88 |
| 1.278 | 5341.1 | 1.01E+004 | ok | 2.69E+003 | 1.33 |
| 1.331 | 5532.2 | 4.36E+005 | ok | 1.97E+005 | 97.20 |



Figure S52. LC-MS Spectrum of DEL Compound **3m**.

3n Purity: 90% Conversion rate: 90% Expected mass: 5525.3 Observed mass: 5525.0



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.30 | 5429.2 | 4.36E+004 | ok | 9.32E+003 | 1.92 |
| 1.41 | 5525.0 | 7.78E+005 | ok | 4.39E+005 | 90.37 |
| 1.51 | 5824.1 | 1.27E+004 | ok | 3.02E+004 | 6.22 |
| 1.58 | 6074.8 | 1.59E+004 | ok | 7.25E+003 | 1.49 |



Figure S53. LC-MS Spectrum of DEL Compound **3n**.

30

Purity: 93% Conversion rate: 93% Expected mass: 5507.3 Observed mass: 5508.1



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.439 | 4979.0 | 2.95E+003 | ok | 6.39E+003 | 0.60 |
| 1.522 | 4979.0 | 4.15E+003 | ok | 4.53E+004 | 4.29 |
| 1.575 | 5508.1 | 1.58E+005 | ok | 9.88E+005 | 93.46 |
| 1.845 | 5506.4 | 1.98E+003 | ok | 1.74E+004 | 1.65 |



Figure S54. LC-MS Spectrum of DEL Compound **30**.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.35 | 5322.7 | 4.21E+004 | ok | 1.29E+004 | 2.05 |
| 1.41 | 5521.2 | 1.02E+006 | ok | 5.73E+005 | 91.14 |
| 1.52 | 5856.8 | 4.38E+004 | ok | 4.28E+004 | 6.82 |



Figure S55. LC-MS Spectrum of DEL Compound **3p**.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.250 | 5225.7 | 2.36E+004 | ok | 3.88E+003 | 2.81 |
| 1.333 | 5524.8 | 6.47E+005 | ok | 1.28E+005 | 92.87 |
| 1.381 | 5523.9 | 5.79E+004 | ok | 5.97E+003 | 4.32 |



Figure S56. LC-MS Spectrum of DEL Compound **3q**.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.40 | 5541.6 | 7.50E+005 | ok | 3.38E+005 | 94.03 |
| 1.50 | 5540.3 | 1.43E+004 | ok | 2.15E+004 | 5.97 |



Figure S57. LC-MS Spectrum of DEL Compound 3r.

Br ОН CI 3s

Purity: 95% Conversion rate: 95% Expected mass: 5586.2 Observed mass: 5586.6

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.310 | 5394.6 | 1.47E+004 | ok | 2.91E+003 | 2.67 |
| 1.358 | 5586.6 | 3.97E+005 | ok | 1.03E+005 | 94.59 |
| 1.430 | 5468.7 | 8.13E+003 | ok | 2.99E+003 | 2.74 |



Figure S58. LC-MS Spectrum of DEL Compound 3s.



Conversion rate: 82% Expected mass: 5535.3 Observed mass: 5535.2



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.225 | 5184.2 | 1.70E+005 | ok | 3.23E+004 | 7.74 |
| 1.249 | 5226.1 | 1.26E+005 | ok | 3.60E+004 | 8.63 |
| 1.295 | 5344.0 | 9.90E+003 | ok | 4.93E+003 | 1.18 |
| 1.334 | 5535.2 | 7.91E+005 | ok | 3.44E+005 | 82.45 |



Figure S59. LC-MS Spectrum of DEL Compound 3t.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.212 | 4978.7 | 2.19E+004 | ok | 2.95E+003 | 0.82 |
| 1.245 | 5226.1 | 1.18E+005 | ok | 2.50E+004 | 6.93 |
| 1.310 | 5508.1 | 8.29E+005 | ok | 3.32E+005 | 92.25 |



Figure S60. LC-MS Spectrum of DEL Compound **3u**.







| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.23 | 4978.7 | 1.56E+004 | ok | 1.06E+004 | 1.09 |
| 1.27 | 5337.0 | 4.78E+004 | ok | 1.05E+005 | 10.76 |
| 1.35 | 5496.9 | 1.02E+006 | ok | 6.10E+005 | 62.77 |
| 1.37 | 5496.7 | 2.06E+006 | ok | 1.62E+005 | 16.71 |
| 1.43 | 5496.0 | 2.37E+005 | ok | 4.59E+004 | 4.73 |
| 1.46 | 10957.5 | 2.46E+004 | ok | 3.84E+004 | 3.95 |









| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.224 | 5183.6 | 3.36E+004 | ok | 7.29E+003 | 2.59 |
| 1.245 | 5225.8 | 2.61E+004 | ok | 1.28E+004 | 4.57 |
| 1.273 | 5075.8 | 1.44E+004 | ok | 3.70E+003 | 1.32 |
| 1.310 | 5496.7 | 6.99E+005 | ok | 2.35E+005 | 83.66 |
| 1.375 | 5496.0 | 3.86E+004 | ok | 2.21E+004 | 7.86 |



Figure S62. LC-MS Spectrum of DEL Compound **3w**.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.279 | 5353.9 | 2.05E+004 | ok | 3.03E+003 | 2.06 |
| 1.303 | 5321.9 | 2.75E+004 | ok | 3.62E+003 | 2.46 |
| 1.348 | 5513.0 | 5.94E+005 | ok | 1.27E+005 | 86.07 |
| 1.392 | 5840.4 | 4.56E+004 | ok | 1.39E+004 | 9.42 |



Figure S63. LC-MS Spectrum of DEL Compound **3x**.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.218 | 4978.8 | 1.89E+004 | ok | 3.34E+003 | 0.80 |
| 1.251 | 5225.7 | 6.54E+004 | ok | 2.28E+004 | 5.43 |
| 1.322 | 5512.9 | 7.88E+005 | ok | 3.94E+005 | 93.77 |



Figure S64. LC-MS Spectrum of DEL Compound **3y**.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.35 | 10629.3 | 1.55E+004 | ok | 6.99E+003 | 1.79 |
| 1.40 | 5492.1 | 7.09E+005 | ok | 3.75E+005 | 96.17 |
| 1.49 | 5798.7 | 2.84E+004 | ok | 7.93E+003 | 2.03 |



Figure S65. LC-MS Spectrum of DEL Compound 3z.

9.8. Mass Spectrum of 3o at A Scale of 100 nmol

₹ N H 30 Purity: 84% Conversion rate: 84%

Expected mass: 5507.3 Observed mass: 5506.5



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.252 | 5348.8 | 3.73E+003 | ok | 1.08E+004 | 6.13 |
| 1.282 | 5315.7 | 1.63E+004 | ok | 1.68E+004 | 9.50 |
| 1.323 | 5506.5 | 1.12E+005 | ok | 1.49E+005 | 84.37 |



Figure S66. LC-MS Spectrum of **3o** at a scale of 100 nmol.

9.9. Mass Spectrum of 5a-5v

ξ N Η ÓH 5a Purity: 81%

Conversion rate: 81% Expected mass: 5424.8 Observed mass: 5426.0

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.294 | 5426.0 | 4.37E+004 | ok | 1.94E+005 | 80.54 |
| 1.340 | 5314.9 | 7.30E+003 | ok | 4.68E+004 | 19.46 |



Figure S67. LC-MS Spectrum of DEL Compound 5a.

όн 5b Purity: 82% Conversion rate: 82% Expected mass: 5438.8 Observed mass: 5437.8



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.068 | 5348.6 | 8.24E+003 | ok | 1.15E+004 | 2.93 |
| 1.093 | 5437.8 | 2.69E+005 | ok | 3.23E+005 | 82.25 |
| 1.155 | 23219.2 | 2.68E+004 | ok | 5.81E+004 | 14.81 |



Figure S68. LC-MS Spectrum of DEL Compound **5b**.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.043 | 5361.0 | 7.93E+004 | ok | 2.90E+004 | 7.15 |
| 1.071 | 5317.1 | 1.06E+005 | ok | 1.43E+004 | 3.54 |
| 1.091 | 5323.7 | 7.17E+004 | ok | 7.06E+003 | 1.74 |
| 1.132 | 5478.8 | 1.21E+006 | ok | 3.10E+005 | 76.52 |
| 1.177 | 5477.6 | 4.62E+004 | ok | 4.47E+004 | 11.04 |



Figure S69. LC-MS Spectrum of DEL Compound 5c.

Ž H óн 5d Purity: 85% Conversion rate: 85% Expected mass: 5500.9

Observed mass: 5500.5 and 5499.6



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.314 | 5315.6 | 2.65E+004 | ok | 8.59E+003 | 6.63 |
| 1.362 | 5500.5 | 1.40E+005 | ok | 3.83E+004 | 29.57 |
| 1.374 | 5499.6 | 3.66E+005 | ok | 7.15E+004 | 55.15 |
| 1.422 | 26572.2 | 5.66E+003 | ok | 1.12E+004 | 8.65 |



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Figure S70. LC-MS Spectrum of DEL Compound 5d.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.303 | 5451.2 | 1.53E+004 | ok | 3.21E+004 | 30.57 |
| 1.313 | 5450.3 | 5.31E+004 | ok | 5.37E+004 | 51.24 |
| 1.336 | 5315.1 | 6.11E+003 | ok | 1.91E+004 | 18.18 |





5f Purity: 86% Conversion rate: 86% Expected mass: 5436.8 Observed mass: 5437.7

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.293 | 5437.7 | 2.75E+004 | ok | 2.49E+005 | 85.99 |
| 1.334 | 5314.9 | 4.64E+003 | ok | 4.06E+004 | 14.01 |



Figure S72. LC-MS Spectrum of DEL Compound 5f.


Conversion rate: 75% Expected mass: 5450.9 Observed mass: 5451.5 and 5451.3

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.303 | 5451.5 | 1.48E+004 | ok | 6.26E+004 | 32.09 |
| 1.316 | 5451.3 | 1.15E+005 | ok | 8.45E+004 | 43.32 |
| 1.339 | 5449.7 | 4.49E+004 | ok | 3.06E+004 | 15.68 |
| 1.371 | 5315.3 | 1.83E+003 | ok | 1.74E+004 | 8.92 |









| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.220 | 4978.6 | 2.67E+004 | ok | 1.02E+004 | 1.14 |
| 1.257 | 5411.6 | 1.45E+005 | ok | 4.68E+004 | 5.25 |
| 1.311 | 5464.2 | 4.63E+005 | ok | 8.34E+005 | 93.60 |





used was a crude product synthesized in the laboratory.

NT NH 5h

Purity: 95% Conversion rate: 95% Expected mass: 5464.9 Observed mass: 5465.4

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.050 | 4979.5 | 5.98E+004 | ok | 2.38E+004 | 1.66 |
| 1.077 | 5412.6 | 1.94E+005 | ok | 4.92E+004 | 3.44 |
| 1.116 | 5465.4 | 1.22E+006 | ok | 1.36E+006 | 94.90 |



Figure S75. LC-MS Spectrum of DEL Compound **5h**. The sodium sulfinate (**4h**) used was obtained from commercial sources.







| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.29 | 5411.7 | 3.39E+004 | ok | 3.79E+004 | 4.38 |
| 1.33 | 5315.7 | 3.19E+004 | ok | 3.38E+004 | 3.90 |
| 1.39 | 5478.5 | 2.40E+005 | ok | 3.11E+005 | 38.80 |
| 1.41 | 5477.2 | 5.02E+005 | ok | 2.06E+005 | 25.83 |
| 1.44 | 5476.6 | 1.21E+005 | ok | 3.16E+004 | 3.65 |
| 1.46 | 5476.2 | 3.19E+004 | ok | 3.23E+004 | 3.73 |
| 1.50 | 10922.8 | 9.33E+003 | ok | 7.27E+004 | 8.40 |
| 1.53 | 11029.3 | 2.19E+004 | ok | 9.81E+004 | 11.33 |





Figure S76. LC-MS Spectrum of DEL Compound 5i.



Purity: 95% Conversion rate: 95% Expected mass: 5472.9 Observed mass: 5473.9 and 5472.1





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.273 | 5316.2 | 1.68E+003 | ok | 6.93E+003 | 2.68 |
| 1.299 | 5473.9 | 7.74E+004 | ok | 7.34E+004 | 28.42 |
| 1.310 | 5472.1 | 5.66E+004 | ok | 1.71E+005 | 66.18 |
| 1.387 | 6037.3 | 1.10E+002 | ok | 7.03E+003 | 2.72 |



Figure S77. LC-MS Spectrum of DEL Compound 5j.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.080 | 5485.7 | 7.58E+004 | ok | 3.41E+005 | 83.88 |
| 1.148 | 10937.4 | 5.68E+003 | ok | 6.56E+004 | 16.12 |



Figure S78. LC-MS Spectrum of DEL Compound 5k.



Observed mass: 5486.0



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.077 | 5486.0 | 3.72E+005 | ok | 1.01E+006 | 78.21 |
| 1.122 | 10938.1 | 4.75E+004 | ok | 1.07E+005 | 8.31 |
| 1.143 | 5765.8 | 2.31E+004 | ok | 1.74E+005 | 13.48 |



Figure S79. LC-MS Spectrum of DEL Compound 5I.

ÓН 5m Purity: 96% Conversion rate: 96% Expected mass: 5486.9 Observed mass: 5487.7



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.346 | 4979.1 | 5.44E+003 | ok | 8.51E+003 | 0.69 |
| 1.427 | 5316.5 | 3.57E+003 | ok | 3.81E+004 | 3.07 |
| 1.475 | 5487.7 | 1.30E+005 | ok | 1.19E+006 | 96.24 |



Figure S80. LC-MS Spectrum of DEL Compound **5m**.



Purity: 83% Conversion rate: 83% Expected mass: 5490.8 Observed mass: 5491.6

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.208 | 4979.2 | 1.09E+003 | ok | 6.35E+003 | 1.62 |
| 1.246 | 5332.2 | 1.60E+003 | ok | 8.65E+003 | 2.20 |
| 1.304 | 5491.6 | 5.02E+004 | ok | 3.24E+005 | 82.52 |
| 1.382 | 11054.9 | 8.43E+002 | ok | 5.36E+004 | 13.66 |



Figure S81. LC-MS Spectrum of DEL Compound **5n**.



Conversion rate: 95% Expected mass: 5490.8 Observed mass: 5491.6



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.352 | 4979.1 | 6.14E+003 | ok | 1.02E+004 | 0.64 |
| 1.462 | 5491.6 | 1.32E+005 | ok | 1.51E+006 | 95.26 |
| 1.728 | 5490.1 | 1.36E+003 | ok | 6.50E+004 | 4.10 |



Figure S82. LC-MS Spectrum of DEL Compound **50**.

5р Purity: 78% Conversion rate: 78% Expected mass: 5490.8 Observed mass: 5490.8



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.331 | 5198.7 | 1.81E+003 | ok | 1.53E+004 | 2.13 |
| 1.380 | 5226.3 | 1.74E+003 | ok | 8.13E+003 | 1.13 |
| 1.458 | 5490.8 | 9.32E+004 | ok | 5.65E+005 | 78.29 |
| 1.543 | 5489.7 | 2.87E+003 | ok | 1.23E+005 | 17.07 |
| 1.738 | 5489.8 | 2.07E+002 | ok | 1.00E+004 | 1.39 |



Figure S83. LC-MS Spectrum of DEL Compound **5p**.

N H ÓН 5q Purity: 89%

Conversion rate: 89% Expected mass: 5551.8 Observed mass: 5551.5

LC Spectrum:



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| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.331 | 5199.1 | 8.96E+002 | ok | 9.60E+003 | 2.33 |
| 1.385 | 5225.9 | 9.66E+002 | ok | 8.62E+003 | 2.09 |
| 1.446 | 5323.9 | 3.38E+003 | ok | 2.89E+004 | 7.00 |
| 1.489 | 5551.5 | 3.69E+004 | ok | 3.66E+005 | 88.58 |



Figure S84. LC-MS Spectrum of DEL Compound **5q**.

5r

Purity: 93% Conversion rate: 93% Expected mass: 5473.8 Observed mass: 5473.1



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.039 | 5410.2 | 9.42E+004 | ok | 3.00E+004 | 6.56 |
| 1.074 | 5473.1 | 7.09E+005 | ok | 4.28E+005 | 93.44 |



Figure S85. LC-MS Spectrum of DEL Compound 5r.



Conversion rate: >99% Expected mass: 5473.8 Observed mass: 5474.4

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.268 | 5474.4 | 1.94E+005 | ok | 3.88E+005 | 100.00 |



Figure S86. LC-MS Spectrum of DEL Compound **5s**.



Conversion rate: >99% Expected mass: 5473.8 Observed mass: 5473.1



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.452 | 5473.1 | 3.65E+004 | ok | 3.16E+005 | 100.00 |



Figure S87. LC-MS Spectrum of DEL Compound 5t.





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.424 | 4979.2 | 5.52E+003 | ok | 7.94E+003 | 0.47 |
| 1.534 | 5525.1 | 1.28E+005 | ok | 6.43E+005 | 37.76 |
| 1.572 | 5524.3 | 1.23E+005 | ok | 1.02E+006 | 59.92 |
| 1.837 | 5523.4 | 1.07E+003 | ok | 3.16E+004 | 1.85 |





Figure S88. LC-MS Spectrum of DEL Compound **5u**.



Purity: 94% Conversion rate: 94% Expected mass: 5478.9 Observed mass: 5477.8 and 5477.1





| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.34 | 5477.8 | 3.43E+004 | ok | 6.24E+004 | 55.10 |
| 1.35 | 5477.1 | 5.56E+004 | ok | 4.45E+004 | 39.29 |
| 1.41 | 5476.5 | 2.82E+003 | ok | 6.36E+003 | 5.62 |



Figure S89. LC-MS Spectrum of DEL Compound **5v**.

9.10. Mass Spectra of DEL

Cycle 1 Analysis, well A1

100000 \sim Expected mass: 23546.7 Observed mass: 23548.8

LC/UV Spectrum:



Deconvoluted mass spectrum of Product:



Figure S90. LC/UV Spectrum and deconvoluted mass spectrum of cycle 1 well A1.

Cycle 1 Analysis, well A2



LC/UV Spectrum:



Deconvoluted mass spectrum of Product:



Figure S91. LC/UV Spectrum and deconvoluted mass spectrum of cycle 1 well **A2**. Cycle 1 Analysis, well **A3**







Figure S92. LC/UV Spectrum and deconvoluted mass spectrum of cycle 1 well **A3**. Cycle 1 Analysis, well **A4**





Deconvoluted mass spectrum of Product:



Figure S93. LC/UV Spectrum and deconvoluted mass spectrum of cycle 1 well **A4**. Cycle 1 Analysis, well **A5**





Deconvoluted mass spectrum of Product:



Figure S94. LC/UV Spectrum and deconvoluted mass spectrum of cycle 1 well **A5**. Cycle 1 Analysis, well **A6**



Expected mass: 23600.8 Observed mass: 23602.8

LC/UV Spectrum:



Deconvoluted mass spectrum of Product:



Figure S95. LC/UV Spectrum and deconvoluted mass spectrum of cycle 1 well **A6**. Crude cycle 1 products, pooled and precipitated.



Deconvoluted mass spectrum of Products:



Figure S96. LC/UV Spectrum and deconvoluted mass spectrum of crude pooled cycle 1 products.

Cycle 2 Analysis, well A1



Deconvoluted mass spectrum of Products:



Figure S97. LC/UV Spectrum and deconvoluted mass spectrum of well **A1**. Cycle 2 Analysis, well **A2**



Expected mass: 31748.3-31814.4



Deconvoluted mass spectrum of Products:



Figure S98. LC/UV Spectrum and deconvoluted mass spectrum of well **A2**. Cycle 2 Analysis, well **A3**





Figure S99. LC/UV Spectrum and deconvoluted mass spectrum of well **A3**. Cycle 2 Analysis, well **A4**





Figure S100. LC/UV Spectrum and deconvoluted mass spectrum of well **A4**. Cycle 2 Analysis, well **A5**




Deconvoluted mass spectrum of Products:



Figure S101. LC/UV Spectrum and deconvoluted mass spectrum of well **A5**. Cycle 2 Analysis, well **A6**



Deconvoluted mass spectrum of Products:



Figure S102. LC/UV Spectrum and deconvoluted mass spectrum of well **A6**. Crude cycle 2 products, pooled and precipitated.



Deconvoluted mass spectrum of Products:



Figure S103. LC/UV Spectrum and deconvoluted mass spectrum of crude pooled cycle 2 products.

Purified cycle 2 products, pooled and precipitated.





Deconvoluted mass spectrum of Products:



011294.0

Figure S104. LC/UV Spectrum and deconvoluted mass spectrum of purified pooled cycle 2 products.

9.11. Mass Spectra of S4 from Control Experiment

LC/UV Spectrum:



Deconvoluted mass spectrum of Product:



Figure S105. LC/UV Spectrum and deconvoluted mass spectrum of **S4**. LC/UV Spectrum:



Deconvoluted mass spectrum of Product:



Figure S106. LC/UV Spectrum and deconvoluted mass spectrum of photoreaction of **S4**.

10. DNA Damage Evaluation

10.1. Ligation Procedure for the Product 6



Tag 1: The sequences of 5 '-phosphorylated Tag 1-F chain are 5'-ACTXXXXXXAG -3', and the sequences of their complementary 5'phosphorylated Tag 1-R are 5'- GCACCTXXXXXXAGTGG -3'.

To DNA sample **HP** (5.0 nmol, 5.0 μ L, 1.0 equiv.) was added DNA **Tag 1** (5.5 nmol, 1.1 equiv.), **Tag 2** (6.1 nmol, 1.2 equiv.), followed by the addition of 10X ligation buffer (4.0 μ L) and T4 DNA ligase (12.0 U), and H₂O to a final volume of 40.0 μ l. The reaction mixture was incubated at 25 °C for 2 hours. The ligation was assessed for completion by LC-MS analysis. The reaction mixture was quenched thermally by incubation for 20 minutes at 65 °C. The crude material **6** was purified by ethanol precipitation. Product **6** (149 nt, 4.0 nmol, 80% recovery rate) was obtained as white solids.

10.2. Synthesis of Product 8



Tag 1: The sequences of 5 '-phosphorylated Tag 1-F chain are 5'-ACTXXXXXXAG -3', and the sequences of their complementary 5'- phosphorylated Tag 1-R are 5'- GCACCTXXXXXXAGTGG -3'.

Preparation of **7**: To DNA sample **3o** (5.0 nmol, 5.0 μ L, 1.0 equiv.) was added DNA **Tag 1** (5.5 nmol, 1.1 equiv.), followed by the addition of 10X ligation buffer (2.0 μ L) and T4 DNA ligase (6.0 U), and H₂O to a final volume of 20.0 μ l. The reaction mixture was incubated at 25 °C for 1 hour. The reaction mixture was quenched thermally by incubation for 20 minutes at 65 °C. The crude material **7** was purified by ethanol precipitation. Product **7** (4.3 nmol, 86% recovery rate) was obtained as white solids.

Preparation of **8**: To **7** (4.3 nmol, 4.3 μ L, 1.0 equiv.) was added DNA **Tag 2** (4.7 nmol, 1.1 equiv.), followed by the addition of 10X ligation buffer (1.72 μ L) and T4 DNA ligase (5.16 U), and H₂O to a final volume of 17.2 μ l. The reaction mixture was incubated at 25 °C for 1 hour. The ligation was assessed for completion by LC-MS analysis. The reaction mixture was quenched thermally by incubation for 20 minutes at 65 °C. The crude material **8** was purified by ethanol precipitation. Product **8** (3.44 nmol, 80% recovery rate) was obtained as white solids.

10.3. qPCR Test



The photoreaction for the synthesis of DNA-conjugated β-hydroxy sulfone was performed with a DNA conjugated compound containing a double stranded DNA coding region to mimic the library component. The product DNA-conjugated **30** was ligated with a full-length of oligonucleotide to furnish **8**. Ligation of **HP** to form DNA **6** was used as a control experiment as described in previous experimental sections. Then the amplification efficiency was analyzed by qPCR (Quant Standio 3, Thermo Fisher).

The product **6** and **8** were subjected to a stepwise dilution with 10 folds each time until it reached a final 10⁸ folds dilution. The last seven 10-folds serial dilutions were used as templates for qPCR tests using a SYBR Green Master Mix kit (Thermo) on a Real-Time PCR System (Quant studio 3). All samples were run in triplicates and subjected to PCR cycles as follows: 50 °C heat activation for 2 mins, then 95 °C heat for 10 mins followed by 30 cycles of 95 °C denaturation for 15 seconds, 60 °C annealing for 60 seconds. Melt curve stage: 95 °C denaturation for 15 seconds, 60 °C annealing for 60 seconds, and 95 °C dissociation for 15 seconds.

To further assess the amplification efficiency, the quantity of the full-length DNA templates was first normalized based on the Bioanalyzer results and qPCR with serial dilution was performed. Linear fitting was then calculated respectively based on the CT values. According to the slope, the amplification efficacy was calculated. Compared between the experimental groups, the amplification efficiency of DNA 8 (102%) was close to control experiment DNA 6 (102%) indicated no obvious impact on PCR efficiency by the reaction. Thus, in summary, the DNA remained in good integrity after the photoreaction.

10.4. Mass Spectra of the Product 6-8

NH₂

Expected mass: 46797.1 Observed mass: 46825.7

LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.297 | 7405.6 | 4.37E+003 | ok | 2.08E+004 | 7.04 |
| 1.333 | 15728.1 | 5.94E+003 | ok | 7.63E+004 | 25.84 |
| 1.383 | 44530.1 | 1.54E+003 | ok | 8.71E+003 | 2.95 |
| 1.438 | 46825.7 | 3.65E+004 | ok | 1.89E+005 | 64.18 |

Deconvoluted mass spectrum of product:



S154

Figure S107. LC-MS Spectrum of 6.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.22 | 4063.1 | 4.20E+004 | ok | 2.77E+004 | 14.44 |
| 1.29 | 5884.6 | 1.22E+004 | ok | 2.17E+004 | 11.32 |
| 1.40 | 15243.8 | 1.12E+004 | ok | 1.05E+004 | 5.50 |
| 1.46 | 15419.3 | 3.12E+005 | ok | 1.32E+005 | 68.74 |

Deconvoluted mass spectrum of product:



Figure S108. LC-MS Spectrum of 7.



LC Spectrum:



| RT (min) | Base Peak Mass (Da) | Intensity | Spectral Quality | LC/UV Peak Area | LC/UV Area Percent |
|-------------|---------------------------|-----------|---------------------|--------------------|--------------------------|
| 1.27 | 7405.6 | 6.36E+003 | ok | 2.68E+004 | 10.91 |
| 1.33 | 15728.9 | 8.81E+003 | ok | 7.70E+004 | 31.34 |
| 1.35 | 25931.8 | 4.18E+003 | ok | 2.40E+004 | 9.76 |
| 1.39 | 47126.7 | 1.68E+004 | ok | 1.18E+005 | 48.00 |

Deconvoluted mass spectrum of product:





10.5. Gel Electrophoresis

Gel electrophoresis was performed with a 12-well 3% TBE agarose gel (Invitrogen) in 1X TBE buffer which was prepared in-house. The ligation mixture was diluted to the concentration of 50.0 ng/ μ L. The DNA loading sample was prepared by adding 10.0 μ L of the diluted DNA sample and 2.0 μ L of 6X DNA loading dye. The first lane of the gel was loaded with a DNA molecular weight ladder, and 5.0 μ L of DNA-dye mixed samples was loaded into each lane. Gels were run at 120 V for 35 minutes and stained with 3X GelStain ethidium bromide

in 0.1M NaCl for 40 min. DNA fragments were visualized under a UV light device and assessed for completed ligation. For smaller scale of ligation experiment, master-mix can be prepared by pre-mixing water, 10X ligation buffer and DNA ligase before adding into the designated DNA starting material. The crude oligo was cleaned up by ethanol precipitation.



Figure S110. Gel electrophoresis image of oligo products: the DNA headpiece (**HP**), DNA-conjugated olefin (**1o**), product **3o**, the subsequent ligation crude products **7**, **6** and **8**.





Figure S111. Standard curve of the qPCR products (a) DNA 6; (b) DNA 8.

| Standard — | Target | : (DNA 8) | Relative Amount of Target (%) | Average Relative |
|------------|--------|-------------|-------------------------------------|-------------------------|
| (nM) | CT1 | Sample (nM) | | Amount of Target (%) |
| 0.045 | 14.38 | 0.035 | 77.4 | 78 |
| 0.0045 | 17.65 | 0.0035 | 78.0 | |
| 0.00045 | 20.92 | 0.00035 | 78.7 | |

Table S1. Calculation of DNA Damage.

In conclusions: The amplifiable DNA in the photoreaction product (DNA 8) is 78.0%. Thus, the DNA damage is 22% after 2 steps of chemical conversion including acylation and photoreaction, in addition to AOP installation.

11. Optimization of the photocatalyst



^aFinal concentration. ^bConversions were determined by LC-MS. ^cGeneral reaction conditions: **1o** (10.0 nmol, 1.0 equiv., 0.8 mM final concentration), **2** (240.0 mM final concentration), Ph-NO₂ (80.0 mM final concentration) and photocatalyst/HCI (1:2, premixed, photocatalyst with 40.0 mM final concentration) in DMSO/H₂O (1:4 by volume) was irradiated with green light for 1 h at 40 °C.