# **Supporting Information**

# A quick access to CF<sub>3</sub>-containing functionalized benzofuranyl, benzothiophene and indolyl heterocycles under catalyst-free conditions

Ayazoddin Aunoddin Kazi,<sup>1,2</sup> Nadimpalli Manjuladevi,<sup>1</sup> Salla Suresh Kumar,<sup>1,2</sup> Anamika Sharma,<sup>2,3\*</sup> L Ravithej Singh<sup>1,2\*</sup>

<sup>1</sup>Fluoro-Agrochemicals Division, CSIR-Indian Institute of Chemical Technology, Hyderabad – 500 007, India

<sup>2</sup>Chemical Sciences Division, Academy of Scientific and Innovative Research, Ghaziabad -201 002, India

<sup>3</sup>Department of Natural Products and Medicinal Chemistry, CSIR-Indian Institute of Chemical Technology, Hyderabad – 500 007, India

\*Corresponding authors: anamika.aug14@gmail.com (Dr. Anamika Sharma); lrsingh@iict.res.in (Dr. L. R. Singh)

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#### 1. General Information

All the chemicals were purchased from Sigma-Aldrich (Sigma-Aldrich, Germany). The solvents used were of analytical and HPLC reagent grade. Magnetic resonance spectra (<sup>1</sup>H and <sup>13</sup>C) were recorded with Bruker, and chemical shift values are reported in  $\delta$  units (ppm) using TMS as internal standard. Follow-up of the reactions and checks of the purity of the compound were done by TLC on silica-gel-protected aluminum sheets 60 F254 (Merck), and the spots were detected by exposure to UV light at  $\lambda = 254$  nm. Analytical HPLC was performed on Shimadzu system using a Phenomenax C18 column (3µm, 4.6×50 mm) by dissolving the sample in CH<sub>3</sub>CN only. Labsolution software was used for data processing. Buffer A: 0.1% TFA in H<sub>2</sub>O, buffer B: 0.1% TFA in CH<sub>3</sub>CN were used in HPLC. High resolution mass spectrometry (HRMS) was performed using a Bruker ESI-QTOF mass spectrometer in positive-ion mode.

#### 2. Preparation of benzofuran derivative

#### 2.1. General procedure for preparation of benzofuran derivative

In an oven-dried flask was added THF (10.0 vol) and DIPA (4.0 eq.) under nitrogen atmosphere. The flask was then cooled to  $-78^{\circ}$ C and then *n*-butyllithium (4.0 eq. of 2.5 M in THF) was added dropwise in inert atmosphere. The reaction mixture was allowed to stir at  $-78^{\circ}$ C for 40 min to generate LDA. After generation of LDA, **2-BTP** (1.7 eq.) was added dropwise to the flask. The reaction mixture was stirred at  $-78^{\circ}$ C for 15 min. In parallel, **1** (1 eq.) was dissolved in dry THF and was added dropwise to the stirring reaction for next 30-90 min while maintaining temperature  $-78^{\circ}$ C. The reaction was monitored by TLC for completion of reaction. After completion, reaction was quenched with aq. NH<sub>4</sub>Cl and extracted three times with EtOAc. The combined organic phase was washed twice with brine. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The crude product was purified by 100-200 silica gel column chromatography using ethyl acetate: hexane as mobile phase to afford **2a-2p** in 40-90 % yield.

#### 2.2. Physical and spectral data



Yield: 90 % Colour: Pale yellow solid Melting point: 135-137°C

<sup>1</sup> H NMR	δ 7.46 (d, <i>J</i> = 7.5 Hz, 1H), 7.37 (t, <i>J</i> = 7.8 Hz, 1H), 7.13 (t, <i>J</i> = 7.5 Hz,
(400 MHz, CDCl <sub>3</sub> ):	1H), 7.07 (d, $J = 8.2$ Hz, 1H), 5.67 (s, 1H), 5.37 (qd, ${}^{3}J_{H-F} = 7.9$ , 1.6
	Hz, 1H), 2.44 (s, 1H).
<sup>13</sup> C NMR	δ 164.9 (d, ${}^{3}J_{C-F}$ = 5.0 Hz), 157.2, 131.4, 126.0, 125.6, 124.0, 123.1 (q,
(125 MHz, CDCl <sub>3</sub> ):	${}^{1}J_{C-F} = 267.5$ Hz), 111.1, 94.6 (q, ${}^{2}J_{C-F} = 37.5$ Hz), 72.4
<sup>19</sup> F NMR	δ-57.6
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>10</sub> H <sub>7</sub> O <sub>2</sub> F <sub>3</sub> : 215.0314 [M–H] <sup>+</sup> ; found 215.0325
(APCI) m/z:	

(Z)-6-methoxy-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2b)



Yield: 80 % Colour: Pale orange solid Melting point: 110-112°C

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): HRMS (APCI) m/z:

δ 7.33 (d, J = 8.3 Hz, 1H), 6.66-6.61 (m, 1H), 6.61 (s, 1H), 5.60 (s, 1H), 5.35 (qd,  ${}^{3}J_{H-F} = 7.9$ , 0.8 Hz, 1H), 3.81 (s, 3H). δ 165.6 (q,  ${}^{3}J_{C-F} = 5.0$  Hz), 162.6, 158.6, 125.9, 117.8, 123.1 (q,  ${}^{1}J_{C-F} = 267.0$  Hz), 110.3, 97.0, 94.6 (q,  ${}^{2}J_{C-F} = 36.0$  Hz). 72.0, 55.8 δ -57.6

Calcd for  $C_{11}H_9O_3F_3$ : 229.0471 [M-OH]<sup>+</sup>; found 229.0475

(Z)-5-chloro-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2c)

Colour: White solid

Melting point: 125-127°C

Yield: 80 %



<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): HRMS (APCI) m/z: δ 7.44 (d, J = 1.8 Hz, 1H), 7.34-7.32 (m, 1H), 7.01 (d, J = 8.6 Hz, 1H), 5.69 (s, 1H), 5.39 (qd,  ${}^{3}J_{H-F} = 7.8$ , 1.8 Hz, 1H), 2.45 (bs, 1H). δ 164.4 (d,  ${}^{3}J_{C-F} = 5.0$  Hz), 155.6, 131.3, 129.1, 127.7, 125.8, 122.7 (q,  ${}^{1}J_{C-F} = 267.0$  Hz), 112.2, 95.1 (q,  ${}^{2}J_{C-F} = 36.0$  Hz), 72.1 δ -57.7 Calcd for C<sub>10</sub>H<sub>6</sub>O<sub>2</sub>ClF<sub>3</sub>: 248.9925 [M–H]<sup>-</sup>; found 248.9938

(Z)-5-bromo-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2d)



<sup>1</sup>H NMR
(400 MHz, CDCl<sub>3</sub>):
<sup>13</sup>C NMR
(100 MHz, CDCl<sub>3</sub>):
<sup>19</sup>F NMR
(376 MHz, CDCl<sub>3</sub>):
HRMS
(APCI) m/z:

Yield: 80 % Colour: Light orange solid Melting point: 119-121°C

δ 7.58 (d, J = 1.6 Hz, 1H), 7.49-7.46 (m, 1H), 6.96 (d, J = 8.6 Hz, 1H), 5.68 (s, 1H), 5.39 (qd,  ${}^{3}J_{H-F}$  = 7.8, 1.7 Hz, 1H), 2.51 (s, 1H). δ 164.3 (q,  ${}^{3}J_{C-F}$  = 5.0 Hz), 156.0, 134.2, 128.7, 128.1, 122.8 (q,  ${}^{1}J_{C-F}$  = 267.0 Hz), 116.3, 112.8, 95.2 (q,  ${}^{2}J_{C-F}$  = 36.0 Hz), 72.0. δ -57.7

Calcd for C<sub>10</sub>H<sub>6</sub>O<sub>2</sub>BrF<sub>3</sub>: 292.9420 [M–H]<sup>+</sup>; found 292.9439

(Z)-7-ethoxy-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2e)



Yield: 70 % Colour: White solid Melting point: 102-104°C

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):

<sup>13</sup>C NMR
(75 MHz, CDCl<sub>3</sub>):
<sup>19</sup>F NMR
(376 MHz, CDCl<sub>3</sub>):
HRMS
(APCI) m/z:

δ 7.06 – 7.01 (m, 2H), 6.95 – 6.91 (m, 1H), 5.67 (d, *J* = 9.7 Hz, 1H), 5.37 (qd, <sup>3</sup>*J*<sub>*H-F*</sub> = 7.9, 1.2 Hz, 1H), 4.19 (q, *J* = 7.0 Hz, 2H), 2.42 (bs, 1H), 1.43 (t, *J* = 7.0 Hz, 3H) δ 164.7 (q, <sup>3</sup>*J*<sub>*C-F*</sub> = 5.3 Hz), 145.8, 143.6, 127.5, 124.7, 123.1 (q, <sup>1</sup>*J*<sub>*C*-*F*</sub> = 267.8 Hz), 117.1, 117.1, 94.6 (q, <sup>2</sup>*J*<sub>*C-F*</sub> = 36.0 Hz), 72.7, 65.6, 15.0 δ -57.6

Calcd for  $C_{12}H_{11}O_3F_3$ : 243.0627 [M-OH]<sup>+</sup>; found 243.0629

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<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):

## <sup>19</sup>F NMR

(376 MHz, CDCl<sub>3</sub>): HRMS (APCI) m/z: Yield: 70 % Colour: Pale orange Melting point: 128-130°C

δ 6.89 (s, 1H), 6.62 (s, 1H), 5.59 (s, 1H), 5.34 (qd, ${}^{3}J_{H-F} = 7.9, 1.7$
Hz, 1H), 3.85 (s, 3H), 3.83 (s, 3H)
δ 165.9 (q, ${}^{3}J_{C-F}$ = 5.0 Hz), 151.7, 151.5, 145.9, 123.2 (q, ${}^{1}J_{C-F}$ =
267.0 Hz), 115.9, 107.9, 95.6, 94.7 (q, ${}^{2}J_{C-F}$ = 36.0 Hz), 72.9, 56.7,
56.3
δ-57.5
Calcd for $C_{12}H_{11}O_4F_3$ : 259.0577 [M-H <sub>2</sub> O] <sup>+</sup> ; found 259.0580

(Z)-3-methyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2g)



Yield:70% yield Colour: Light brown solid Melting point: 110-112°C

<sup>1</sup> H NMR	δ 7.42-7.40 (m, 1H), 7.38 – 7.32 (m, 1H), 7.13 (t, $J = 7.5$ Hz, 1H),
(400 MHz, CDCl <sub>3</sub> ):	7.08 (d, $J = 8.2$ Hz, 1H), 5.28 (q, ${}^{3}J_{H-F} = 7.9$ Hz, 1H), 2.42 (s, 1H),
	1.67 (s, 3H).
<sup>13</sup> C NMR	169.1 (q, ${}^{3}J_{C-F} = 5.3$ Hz), 155.8, 131.0, 130.1, 124.0, 123.7, 123.4 (q,
(75 MHz, CDCl <sub>3</sub> ):	${}^{1}J_{C-F} = 267.0 \text{ Hz}$ ), 111.1, 92.0 (q, ${}^{2}J_{C-F} = 36.0 \text{ Hz}$ ), 77.6, 28.5.
<sup>19</sup> F NMR	δ -57.2.
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Caled for C., H. O. F.: 213 0522 [M-OH]+: found: 213 0526
(APCI) m/z:	Calculor $C_{1119}C_{213}$ . 213.0322 [10-011], 100100. 213.0320.

(Z)-4,6-dimethoxy-3-methyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2h)



**Yield:** 60 % yield **Colour:** Brown solid **Melting point:** 102-104°C

<sup>1</sup> H NMR	δ 6.26 (d, $J = 1.9$ Hz, 1H), 6.16 (d, $J = 1.9$ Hz, 1H), 5.24 (q, ${}^{3}J_{H-F} =$
(500 MHz, CDCl <sub>3</sub> ):	7.9 Hz, 1H), 3.85 (s, 3H), 3.79 (s, 3H), 2.49 (bs, 1H), 1.75 (s, 3H).
<sup>13</sup> C NMR	δ 169.5 (q, ${}^{3}J_{C-F}$ = 5.0 Hz), 163.5, 157.8, 157.4, 123.5 (q, ${}^{1}J_{C-F}$ = 267.0
(100 MHz, CDCl <sub>3</sub> ):	Hz), 108.9, 94.3, 91.6 (q, ${}^{2}J_{C-F}$ = 36.0 Hz), 89.0, 77.8, 55.9, 55.7, 27.0
<sup>19</sup> F NMR	δ-57.2
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>13</sub> H <sub>13</sub> O <sub>4</sub> F <sub>3</sub> : 273.0733 [M-OH] <sup>+</sup> ; found 273.0737
(APCI) m/z:	

(Z)-4-chloro-3-methyl-7-nitro-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2i)



Yield: 70 % Colour: Yellow solid Melting point: 107-109°C

<sup>1</sup>H NMR
(400 MHz, CDCl<sub>3</sub>):
<sup>13</sup>C NMR
(100 MHz, CDCl<sub>3</sub>):
<sup>19</sup>F NMR
(376 MHz, CDCl<sub>3</sub>):

δ 8.11 (s, 1H), 7.67 (d, J = 2.2 Hz, 1H), 5.47 (q,  ${}^{3}J_{H-F} = 8.0$  Hz, 1H), 2.81 (bs, 1H), 1.74 (s, 3H).  $\delta 167.1$  (q,  ${}^{3}J_{C-F} = 5.0$  Hz), 147.9, 136.3, 133.1, 130.0, 129.5, 126.1, 122.4 (q,  ${}^{1}J_{C-F} = 269.0$  Hz), 95.4 (q,  ${}^{2}J_{C-F} = 37.0$  Hz), 76.5, 28.7  $\delta$  -57.8

#### Calcd for C<sub>11</sub>H<sub>7</sub>O<sub>4</sub>NClF<sub>3</sub>: 291.9983 [M-OH]<sup>+</sup>; found 291.9984

(APCI) m/z:

HRMS

(Z)-5-chloro-3,6-dimethyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2j)



(Z)-3,5-dimethyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2k)



**Yield:** 50 % Colour: Yellow solid Melting point: 95-97°C

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): HRMS

(APCI) m/z:

 $\delta$  7.20 (s, 1H), 7.15 – 7.12 (m, 1H), 6.96 (d, J = 8.3 Hz, 1H), 5.24 (q,  ${}^{3}J_{H-F} = 7.9$  Hz, 1H), 2.35 (s, 3H), 1.66 (s, 3H).  $\delta$  169.4 (q,  ${}^{3}J_{C-F}$  = 5.0 Hz), 153.7, 133.6, 131.3, 129.9, 123.9 123.3 (q,  ${}^{1}J_{C-F} = 268.0$  Hz), 110.5, 91.5 (q,  ${}^{2}J_{C-F} = 36.0$  Hz), 77.6, 28.4, 21.0. δ-57.2 Calcd for C<sub>12</sub>H<sub>11</sub>O<sub>2</sub>F<sub>3</sub>: 227.0678 [M-OH]<sup>+</sup>; found 227.0683

(Z)-3,5-dimethyl-7-nitro-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (21)



Colour: Brown solid Melting point: 123-125°C

**Yield: 55 %** 

$NO_2$	
<sup>1</sup> H NMR	δ 7.93 (d, $J = 2.4$ Hz, 1H), 7.51 (d, $J = 1.5$ Hz, 1H), 5.43 (q, ${}^{3}J_{H-F} =$
(500 MHz, CDCl <sub>3</sub> ):	7.9 Hz, 1H), 2.54 (s, 1H), 2.45 (s, 3H), 1.72 (s, 3H)
<sup>13</sup> C NMR	δ 167.7 (q, ${}^{3}J_{C-F}$ = 5.0 Hz), 147.4, 134.7, 134.3, 132.7, 130.2, 126.3,

(125 MHz, CDCl <sub>3</sub> ):	122.6 (q, ${}^{1}J_{C-F} = 268.8 \text{ Hz}$ ), 94.5 (q, ${}^{2}J_{C-F} = 36.3 \text{ Hz}$ ), 76.6, 28.6, 20.8.
<sup>19</sup> F NMR	δ-57.7
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	C <sub>12</sub> H <sub>10</sub> O <sub>4</sub> NF <sub>3</sub> : 272.0529 [M-OH] <sup>+</sup> ; found 272.0533
(APCI) m/z:	

(Z)-7-methoxy-3-phenyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2m)



Yield: 50 % Colour: Pale yellow solid Melting point: 108-110°C

δ 7.40 – 7.35 (m, 4H), 7.34 – 7.30 (m, 1H), 7.09 (d, <i>J</i> = 8.4 Hz, 1H),
6.72 (d, $J = 2.2$ Hz, 1H), 6.65 – 6.62 (m, 1H), 5.08 (q, ${}^{3}J_{H-F} = 7.9$ Hz,
1H), 3.83 (s, 3H), 2.69 (s, 1H).
δ 170.0 (q, ${}^{3}J_{C-F}$ = 5.1 Hz), 162.5, 157.8, 142.3, 128.6, 128.2, 125.4,
123.3 (q, ${}^{1}J_{C-F}$ = 269.0 Hz), 122.6, 110.8, 96.9, 94.4 (q, ${}^{2}J_{C-F}$ = 36.5
Hz), 81.4, 55.9
δ-57.3
Calcd for C <sub>17</sub> H <sub>13</sub> O <sub>3</sub> F <sub>3</sub> : 305.0784 [M-H <sub>2</sub> O] <sup>+</sup> ; found 305.0786

(Z)-5-bromo-3-phenyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2n)



Yield: 70 % Colour: Brown liquid Melting point: NA

δ 7.43 – 7.40 (m, 1H), 7.34 – 7.27 (m, 5H), 7.25 (d, <i>J</i> = 2.1 Hz, 1H),
7.00 (d, $J = 8.6$ Hz, 1H), 5.05 (q, ${}^{3}J_{H-F} = 8.0$ Hz, 1H), 2.76 (s, 1H).
$\delta 167.7$ (d, ${}^{3}J_{CF} = 5.0$ Hz), 154.1, 140.2, 133.0, 131.8, 127.7, 127.5,
126.9, 124.1, 115.4, 111.7, 94.0 (q, ${}^{2}J_{C-F} = 37.0$ Hz), 80.4
δ-57.4
Calcd for C <sub>16</sub> H <sub>10</sub> O <sub>2</sub> BrF <sub>3</sub> : 352.9783 [M-OH] <sup>+</sup> ; found 352.9786

(Z)-5-chloro-3-phenyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (20)



Yield: 70 % Colour: Yellow solid Melting point: 138-140°C

<sup>1</sup>H NMR

(400 MHz, CDCl<sub>3</sub>):

<sup>13</sup>C NMR

(100 MHz, CDCl<sub>3</sub>):

<sup>19</sup>F NMR

(376 MHz, CDCl<sub>3</sub>):
HRMS

(APCI) m/z:

δ 7.50 – 7.48 (m, 1H), 7.39 – 7.38 (m, 1H), 7.38 – 7.36 (m, 3H), 7.36 – 7.34 (m, 1H), 7.33 – 7.32 (m, 1H), 7.07 (d, *J* = 8.6 Hz, 1H), 5.12 (q, <sup>3</sup>*J*<sub>*H-F*</sub> = 7.8 Hz, 1H), 1.25 (s, 1H) δ 168.8, 155.2, 141.3, 134.0, 132.9, 128.8, 128.5, 128.0, 125.1, 116.5, 112.8, 95.0 (q, <sup>1</sup>*J*<sub>*C-F*</sub> = 36.0 Hz), 81.4. δ -57.5 Calcd for C<sub>16</sub>H<sub>10</sub>ClO<sub>2</sub>F<sub>3</sub>: 309.0289 [M-OH]<sup>+</sup>; found 309.0289

(Z)-3-phenyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzofuran-3-ol (2p)



Yield: 55 % Colour: Brown liquid Melting point: NA

F F	
<sup>1</sup> H NMR	δ 7.41 – 7.31 (m, 6H), 7.24 – 7.16 (m, 2H), 7.13 – 7.08 (m, 1H), 5.11
(400 MHz, CDCl <sub>3</sub> ):	$(q, {}^{3}J_{H-F} = 8.0 \text{ Hz}, 1\text{H}), 2.75 \text{ (s, 1H)}$
<sup>13</sup> C NMR	δ 169.1 (q, ${}^{3}J_{C-F}$ = 5.0 Hz), 156.3, 141.9, 131.1, 130.6, 128.6, 128.2,
(100 MHz, CDCl <sub>3</sub> ):	125.2, 124.8, 124.2, 123.2 (q, ${}^{1}J_{C-F} = 267.0$ Hz), 111.1, 94.4 (q, ${}^{2}J_{C-F}$
	= 37.0 Hz), 81.6
<sup>19</sup> F NMR	δ -57.3
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for $C_{16}H_{11}O_2F_3 = 275.0678 \text{ [M-OH]}^+$ ; found 275.0681
(APCI) m/z:	

## 3. Preparation of indolin derivatives

## 3.1. General Procedure for preparation of indolin derivative

Similar procedure as described above was followed for the synthesis of **4a-o** and the product was obtained in 40-90% yield.

## 3.2. Physical and spectral data

(Z)-3-methyl-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4a)

HO	Yield: 80 % Colour: Brown liquid Melting point: NA
<sup>1</sup> H NMK	$\delta$ 7.46 - 7.40 (m, 3H), 7.28 (s, 1H), 5.74 (q, ${}^{3}J_{H-F} = 8.5$ Hz, 1H), 3.13
(500 MHz, CDCl <sub>3</sub> ):	(s, 3H), 2.28 (s, 1H) 1.69 (s, 3H)
<sup>13</sup> C NMR	δ 153.4 (d, ${}^{3}J_{C-F}$ = 4.0 Hz), 140.7, 135.5, 130.5, 126.7, 123.4, 122.5
(100 MHz, CDCl <sub>3</sub> ):	(q, ${}^{1}J_{C-F} = 267.0$ Hz), 117.0, 104.8 (q, ${}^{2}J_{C-F} = 37.0$ Hz), 78.8, 40.4,
	27.3.
<sup>19</sup> F NMR	δ-59.5
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>12</sub> H <sub>12</sub> O <sub>3</sub> NF <sub>3</sub> S: 290.0457 [M-OH] <sup>+</sup> ; found 290.0458
(APCI) m/z:	

(Z)-3-methyl-1-tosyl-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4b)

HO N Ts F F	Yield: 70 % Colour: White solid Melting point: 167-169°C
<sup>1</sup> H NMR	$\delta$ 7.72 – 7.66 (m, 1H), 7.54 – 7.48 (m, 2H), 7.40 – 7.34 (m, 1H), 7.25
(500 MHz, CDCl <sub>3</sub> ):	(d, $J$ = 7.5 Hz, 1H), 7.21 (d, $J$ = 7.4 Hz, 1H), 7.16 (d, $J$ = 8.5 Hz, 2H), 5.79 (qd, ${}^{3}J_{H-F}$ = 8.6, 1.4 Hz, 1H), 2.34 (s, 3H), 1.77 (s, 1H), 1.05 (s, 3H)
<sup>13</sup> C NMR	δ 154.0 (d, ${}^{3}J_{C-F}$ = 6.0 Hz), 145.3, 140.3, 136.0, 134.0, 130.1, 130.0,
(100 MHz, CDCl <sub>3</sub> ):	128.2, 126.8, 123.1, 122.3 (q, ${}^{1}J_{C-F}$ = 268.0 Hz), 118.7, 107.2 (q, ${}^{2}J_{C-F}$ = 38.0 Hz), 27.1, 21.6.
<sup>19</sup> F NMR	δ-58.8
(376 MHz, CDCl <sub>3</sub> ):	
HRMS (APCI) m/z:	Calcd for C <sub>18</sub> H <sub>16</sub> O <sub>3</sub> NF <sub>3</sub> S: 366.0770 [M-H <sub>2</sub> O] <sup>+</sup> ; found 366.0771

tert-butyl (Z)-3-hydroxy-3-methyl-2-(2,2,2-trifluoroethylidene)indoline-1-carboxylate (4c)

HO	<b>Yield:</b> 60 %
	Colour: White solid
	Melting point: 153-155°C
<sup>1</sup> H NMR	δ 7.88 (d, J = 8.2 Hz, 1H), 7.50 – 7.42 (m, 1H), 7.39 – 7.29 (m, 1H),
(400 MHz, CDCl <sub>3</sub> ):	7.21 - 7.09 (m, 1H), 5.65 (s, 1H), 1.68 (s, 3H), 1.64 (s, 9H), 1.63 (s,
	1H).
<sup>13</sup> C NMR	δ 174.3, 173.3, 149.7, 143.2, 130.7, 130.3, 124.9, 123.4, 115.9, 96.3,
(100 MHz, CDCl <sub>3</sub> ):	87.8, 85.2, 31.1, 28.1

 <sup>19</sup>F NMR
 δ -57.2

 (470 MHz, CDCl<sub>3</sub>):

 HRMS
 Calcd for  $C_{16}H_{18}O_3NF_3$ : 330.1312 [M+H]<sup>+</sup>; found 330.1349

 (APCI) m/z:

(Z)-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4d)

ЮН	<b>Yield:</b> 80 %
	Colour: Brown solid
	Melting point: 130-132°C
Ňs F F	
<sup>1</sup> H NMR	$\delta$ 7.52 – 7.49 (m, 2H), 7.45 – 7.41 (m, 1H), 7.31 – 7.27 (m, 1H), 5.81
(500 MHz, CDCl <sub>3</sub> ):	(qd, ${}^{3}J_{H-F} = 8.4$ , 1.6 Hz, 1H), 5.55 (s, 1H), 2.97 (s, 3H), 2.53 (bs, 1H)
<sup>13</sup> C NMR	δ 149.3 (q, ${}^{3}J_{C-F}$ = 5.0 Hz), 142.2, 131.4, 130.9, 126.9, 125.4, 122.2
(125 MHz, CDCl <sub>3</sub> ):	(q, ${}^{1}J_{C-F} = 267.5$ Hz), 117.8, 109.2 (q, ${}^{2}J_{C-F} = 36.3$ Hz), 73.9, 39.0.
<sup>19</sup> F NMR	δ-59.7
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>11</sub> H <sub>10</sub> O <sub>3</sub> NF <sub>3</sub> S: 276.0301 [M-OH] <sup>+</sup> ; found 276.0300
(APCI) m/z:	

(Z)-5,6-dimethoxy-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4e)



(Z)-5-chloro-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4f)



Yield: 70 % Colour: Off white solid Melting point: 147-149°C

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):

δ 7.50 – 7.47 (m, 1H), 7.43 (d, J = 8.3 Hz, 1H), 7.42 – 7.38 (m, 1H), 5.81 (qd,  ${}^{3}J_{H-F}$  = 8.3, 1.7 Hz, 1H), 5.54 (d, J = 7.6 Hz, 1H), 3.00 (s, 3H), 2.49 (d, J = 7.8 Hz, 1H)

<sup>13</sup> C NMR	δ 155.2 (q ${}^{3}J_{C-F}$ = 5.0 Hz), 145.2, 140.0, 136.4, 134.7, 130.5, 127.0
(125 MHz, DMSO-	(q, ${}^{1}J_{C-F} = 267.5$ Hz), 123.5, 112.9 (q, ${}^{2}J_{C-F} = 36.3$ Hz), 77.4, 43.1.
$d_6$ +CDCl <sub>3</sub> ):	
<sup>19</sup> F NMR	δ-59.8
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>11</sub> H <sub>9</sub> O <sub>3</sub> ClNF <sub>3</sub> S: 309.9911 [M-OH] <sup>+</sup> ; found 309.9910
(APCI) m/z:	

(Z)-5-bromo-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4g)



Yield: 70 % Colour: Pale yellow solid Melting point: 140-142°C

1013 1 1	
<sup>1</sup> H NMR	$\delta$ 7.63 (d, $J = 1.6$ Hz, 1H), 7.59 – 7.50 (m, 1H), 7.37 (d, $J = 8.5$ Hz,
(400 MHz, CDCl <sub>3</sub> ):	1H), 5.81 (qd, ${}^{3}J_{H-F} = 8.3$ , 1.6 Hz, 1H), 5.54 (s, 1H), 2.99 (s, 3H),
	2.62 (bs, 1H)
<sup>13</sup> C NMR	δ 149.0 (q, ${}^{3}J_{C-F}$ = 3.8 Hz), 139.8, 134.3, 131.6, 127.4, 121.0 (q,
(100 MHz, DMSO-	${}^{1}J_{C-F} = 266.0$ Hz) 118.3, 117.9, 107.1 (q, ${}^{2}J_{C-F} = 36.0$ Hz), 71.4,
$d_6$ +CDCl <sub>3</sub> ):	37.2.
<sup>19</sup> F NMR	δ-59.7
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>11</sub> H <sub>9</sub> O <sub>3</sub> BrNF <sub>3</sub> S: 353.9406 [M-OH] <sup>+</sup> ; found 353.9404
(APCI) m/z:	

(Z)-5-fluoro-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4h)

ЮН	Yield: 65 %
F	Colour: Brown
	Melting point: 138-140°C
Ms F F	
<sup>1</sup> H NMR	δ 7.63 – 7.62 (m, 1H), 7.56 – 7.54 (m, 1H), 7.38 – 7.36 (m, 1H),
(400 MHz, CDCl <sub>3</sub> ):	5.81 (qd, ${}^{3}J_{H-F} = 8.3$ , 1.6 Hz, 1H), 5.54 (m, 1H), 2.99 (s, 3H), 2.62
	(bs, 1H).
<sup>13</sup> C NMR	δ 159.7 (d, ${}^{\text{Ar}}J_{C-F}$ = 245.4 Hz), 149.5 (q, ${}^{3}J_{C-F}$ = 4.0 Hz), 136.4, 134.2
(100 MHz, DMSO-	(d, ${}^{\text{Ar}}J_{C-F} = 8.2 \text{ Hz}$ ), 121.0 (q, ${}^{1}J_{C-F} = 268.0 \text{ Hz}$ ), 117.7 (d, ${}^{\text{Ar}}J_{C-F} =$
$d_6$ +CDCl <sub>3</sub> ):	8.4 Hz), 115.4 (d, ${}^{\text{Ar}}J_{C-F}$ = 24.0 Hz), 111.5 (d, ${}^{\text{Ar}}J_{C-F}$ = 24.6 Hz),
	107.0 (q, ${}^{2}J_{C-F}$ = 36.0 Hz), 71.4, 36.7
<sup>19</sup> F NMR	δ -59.8, -114.0
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>11</sub> H <sub>9</sub> O <sub>3</sub> NF <sub>4</sub> S: 294.0206 [M-OH] <sup>+</sup> ; found 294.0205
(APCI) m/z:	

(Z)-1-(methylsulfonyl)-5-phenyl-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4i)



Yield: 80 % Colour: Pale yellow Melting point: 140-142°C

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):

<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 7.72 (d, J = 1.7 Hz, 1H), 7.66 – 7.63 (m, 1H), 7.58 – 7.54 (m, 3H), 7.49 – 7.44 (m, 2H), 7.41 – 7.37 (m, 1H), 5.84 (qd, <sup>3</sup>J<sub>H-F</sub> = 8.3, 1.6 Hz, 1H), 5.62 (s, 1H), 3.03 (s, 3H), 2.43 (bs, 1H). δ 149.5, 141.4, 140.4, 139.8, 132.0, 129.8, 129.1, 128.0, 127.1, 127.0 (q, <sup>1</sup>J<sub>C-F</sub> = 268.8 Hz), 123.9, 118.1, 109.0 (q, <sup>2</sup>J<sub>C-F</sub> = 36.3 Hz), 74.2, 39.2 δ -59.7

(376 MHz, CDCl<sub>3</sub>): HRMS (APCI) m/z:

<sup>19</sup>F NMR

Calcd for  $C_{17}H_{14}O_3NF_3S$ : 352.0614 [M-OH]<sup>+</sup>; found 352.0612

(Z)-5-chloro-3-methyl-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4j)



Yield: 50 % Colour: Light yellow liquid Melting point: NA

δ 7.42 – 7.36 (m, 2H), 7.32 (d, $J$ = 8.5 Hz, 1H), 5.74 (q, ${}^{3}J_{C-F}$ =
8.5 Hz, 1H), 3.15 (s, 3H), 2.33 (bs, 1H), 1.68 (s, 3H).
δ 153.0 (d, ${}^{3}J_{C-F}$ = 5.0 Hz), 139.2, 137.3, 132.1, 130.4, 123.7,
122.2 (q, ${}^{1}J_{C-F} = 267.5$ Hz), 118.2, 105.0 (q, ${}^{2}J_{C-F} = 36.3$ Hz),
78.7, 40.7, 27.4
δ-59.5
Calcd for C <sub>12</sub> H <sub>11</sub> O <sub>3</sub> NClF <sub>3</sub> S: 324.0067 [M-OH] <sup>+</sup> ; found
324.0069

(Z)-5-bromo-3-methyl-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4k)



Nethylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (2 Yield: 70 % Colour: Light yellow liquid Melting point: NA

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): <sup>13</sup>C NMR δ 7.56 – 7.51 (m, 2H), 7.28 (s, 1H), 5.74 (q,  ${}^{3}J_{H-F} = 8.5$  Hz, 1H), 3.15 (s, 3H), 2.38 (s, 1H), 1.68 (s, 3H). δ 152.9 (q,  ${}^{3}J_{C-F} = 5.0$  Hz), 139.7, 137.6, 133.3, 126.7, 122.2 (q,

(100 MHz, CDCl <sub>3</sub> ):	${}^{1}J_{C-F} = 267.0$ Hz), 119.6, 118.5, 104.9 (q, ${}^{2}J_{C-F} = 37.0$ Hz), 78.8,
	40.8, 27.5.
<sup>19</sup> F NMR	δ -59.5
(376 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>12</sub> H <sub>11</sub> O <sub>3</sub> NBrF <sub>3</sub> S: 367.9562 [M-OH] <sup>+</sup> ; found
(APCI) m/z:	367.9561

(Z)-1-(methylsulfonyl)-3-phenyl-2-(2,2,2-trifluoroethylidene)indolin-3-ol (41)

Yield: 60 % Colour: Pale brown Melting point: 157-159°C

Ms F F	
<sup>1</sup> H NMR	δ 7.55 (d, <i>J</i> = 8.1 Hz, 1H), 7.47 – 7.43 (m, 1H), 7.42 – 7.33 (m, 5H),
(500 MHz, CDCl <sub>3</sub> ):	$7.30 - 7.25$ (m, 2H), $5.57$ (q, ${}^{3}J_{H-F} = 8.0$ Hz, 1H), $2.94$ (s, 3H), $2.77$ (s,
	1H)
<sup>13</sup> C NMR	δ 153.4 (q, ${}^{3}J_{C-F}$ = 5.0 Hz), 141.8, 140.3, 131.0, 128.9, 128.8, 126.9,
(100 MHz, CDCl <sub>3</sub> ):	126.4, 125.1, 122.4 (q, ${}^{1}J_{C-F} = 268.0$ Hz), 117.2, 108.4 (q, ${}^{2}J_{C-F} = 37.0$
	Hz), 83.0, 39.7
<sup>19</sup> F NMR	δ-59.6
(471 MHz, CDCl <sub>3</sub> ):	
HRMS	Calcd for C <sub>17</sub> H <sub>14</sub> O <sub>3</sub> NF <sub>3</sub> S: 352.0614 [M-OH] <sup>+</sup> ; found 352.0613
(APCI) m/z:	

(Z)-5-chloro-1-(methylsulfonyl)-3-phenyl-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4m)



(Z)-5-bromo-1-(methylsulfonyl)-3-phenyl-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4n)



(Z)-5-bromo-3-(2-fluorophenyl)-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3ol (40)



#### 4. Preparation of benzothiophene derivative

Similar procedure as described above was followed for the synthesis of **6a** and the product was obtained in 80% yield.

(Z)-3-methyl-2-(2,2,2-trifluoroethylidene)-2,3-dihydrobenzo[b]thiophen-3-ol (6a)



Yield: 80 % Colour: White solid Melting point: 86-88°C

<sup>1</sup>H NMR  $\delta$  7.42 – 7.40 (m, 1H), 7.33 – 7.29 (m, 1H), 7.24 – 7.20 (m, 2H), 6.13 (400 MHz, CDCl<sub>3</sub>): (q,  ${}^{3}J_{H-F} = 8.0$  Hz, 1H), 2.48 (s, 1H), 1.63 (s, 3H) <sup>13</sup>C NMR  $\delta$  158.9 (q,  ${}^{3}J_{C-F} = 4.0$  Hz), 141.8, 134.7, 130.2, 126.2, 124.3, 123.9 (100 MHz, CDCl<sub>3</sub>): (q,  ${}^{1}J_{C-F} = 268.0$  Hz), 122.1, 109.1 (q,  ${}^{2}J_{C-F} = 35.0$  Hz), 85.3, 31.4 <sup>19</sup>F NMR  $\delta$  -59.7 (376 MHz, CDCl<sub>3</sub>): HRMS Calcd for C<sub>11</sub>H<sub>9</sub>OF<sub>3</sub>S: 229.0293 [M-OH]<sup>+</sup>; found 229.0291 (APCI) m/z:

#### 5. Further derivatization

#### 5.1. Synthesis of 2-(2,2,2-trifluoroethyl)benzofuran-3(2H)-one (2aa)

In an oven dried nitrogen purged flask, 2a (50 mg, 1.0 eq.) and triethylamine (1.5 eq.) was suspended in toluene (1.0 mL). The reaction mixture was refluxed for 2 h. After completion (as monitored by TLC), the solvent was removed under reduced pressure. The crude was then purified using silica gel column chromatography to obtain pure product (2aa).



#### 5.2. Syntnesis of (Z)-5-metnyl-5-(prop-2-yn-1-yloxy)-2-(2,2,2-trifluoroethylided dihydrobenzofuran (2ga)

In a round bottom flask, 2g (100 mg, 1.0 eq) and sodium hydride (1.5 eq.) were suspended in DMF (2.0 mL) at 0°C. The reaction was stirred for 30 min maintaining the temperature. To the stirring mixture, propargyl bromide (1.1 eq.) was added dropwise. The reaction mixture was then stirred for 2 h. After completion (as monitored by TLC), the reaction was quenched with ice cold water and extracted 3 times with ethyl acetate. Combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The crude was then purified using silica gel column chromatography to obtain pure product (**2ga**).



<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): HRMS (APCI) m/z:

(125 MHz, CDCl<sub>3</sub>):

Yield: 80 % Colour: Colourless liquid Melting point: NA

δ 7.40 – 7.37 (m, 2H), 7.18 – 7.14 (m, 1H), 7.10 – 7.08 (m, 1H), 5.30 (q,  ${}^{3}J_{H-F}$ = 8.0 Hz, 1H), 3.80 (dd, *J* = 14.6, 2.4 Hz, 1H), 3.59 (dd, *J* = 14.6, 2.5 Hz, 1H), 2.38 (t, *J* = 2.5 Hz, 1H), 1.71 (s, 3H) δ 165.1 (q,  ${}^{3}J_{C-F}$ = 5.0 Hz), 156.6, 131.4, 125.6, 124.5, 124.1, 123.2 (q,  ${}^{1}J_{C-F}$ = 267.5 Hz), 111.2, 93.1 (q,  ${}^{2}J_{C-F}$ = 36.3 Hz), 83.3, 79.2, 74.6, 53.2, 28.8 δ -57.2

Calcd for  $C_{14}H_{11}O_2F_3$ : 269.0784 [M+H]<sup>+</sup>; found 269.0784

## 5.3. Synthesis of (Z)-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)indolin-3-one (4da)

In an oven dried nitrogen purged flask, **4d** (20 mg, 1 eq.) and pyridinium chlorochromate (1.5 eq.) was suspended in DCM (1 mL). The reaction mixture was stirred at rt for 2 h. After completion (as monitored by TLC), the reaction was filtered through celite. The celite was washed several times with DCM. The combined organic filtrate was washed with brine, dried over  $Na_2SO_4$ , and concentrated under reduced pressure. The crude was then purified using silica gel column chromatography to obtain pure product (**4da**).

Yield: 95 % Colour: Brown liquid Melting point: NA
5 7.94 (d, J = 8.4 Hz, 1H), 7.87 – 7.85 (m, 1H), 7.73 – 7.69
(m, 1H), 7.34 (t, $J = 7.5$ Hz, 1H), 6.81 (q, ${}^{3}J_{H-F} = 9.0$ Hz,
1H), 3.10 (s, 3H)
8 184.4, 151.1, 137.8, 126.7, 125.7, 123.8, 118.4, 116.2,
109.0 (q, ${}^{2}J_{C-F} = 39.0$ Hz), 40.5.
S -57.2
Calcd for C <sub>11</sub> H <sub>8</sub> O <sub>3</sub> NF <sub>3</sub> S: 292.0250 [M+H] <sup>+</sup> ; found
292.0248

5.4. Synthesis of (Z)-3-methyl-1-(methylsulfonyl)-2-(2,2,2-trifluoroethylidene)-5-((trimethylsilyl)ethynyl)indolin-3-ol (4ga)

In an oven dried nitrogen purged flask, 4g (100 mg, 1.0 eq.), triethylamine (5.0 eq.), CuI (0.01 eq.) and Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub> (0.03 eq.) was suspended in dry THF (1.0 mL). The mixture was purged with nitrogen and TMS-acetylene (1.1 eq.) was added. The reaction mixture was stirred at 80°C for 8 h. After completion (as monitored by TLC), the reaction was cooled to rt and filtered through celite. The celite was washed several times with diethyl ether. The combined organic filtrate was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated under reduced pressure. The crude was then purified using silica gel column chromatography to obtain pure product (**4ga**).



## 5.5. Synthesis of (*Z*)-3-methyl-1-(methylsulfonyl)-5-(4,4,5,5-tetramethyl-1,3,2dioxaborolan-2-yl)-2-(2,2,2-trifluoroethylidene)indolin-3-ol (4gb)

In an oven dried nitrogen purged flask, 4g (20 mg, 1.0 eq.), bis(pinacolato)diboron (1.2 eq.) and KOAc (3.0 eq.) was suspended in dry 1,4-dioxane (1.0 mL). To this mixture, catalytic amount of Pd(dppf)Cl<sub>2</sub> (< 1 mg) was added. The reaction mixture was stirred at 90°C for 2 h. After completion (as monitored by TLC), the reaction was cooled to rt and filtered through celite. The celite was washed several times with diethyl ether. The combined organic filtrate was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The crude was then purified using silica gel column chromatography to obtain pure product (**4gb**).



Yield: 80 % Colour: Viscous colourless liquid Melting point: NA

<sup>1</sup>H NMR

(400 MHz, CDCl<sub>3</sub>):

<sup>13</sup>C NMR

(175 MHz, DMSO-d<sub>6</sub>+CDCl<sub>3</sub>):
<sup>19</sup>F NMR
(376 MHz, CDCl<sub>3</sub>):
HRMS

(APCI) m/z:

δ 7.88 (s, 1H), 7.87-7.84 (m, 1H), 7.41 (d, *J* = 8.0 Hz, 1H), 5.74 (q, <sup>3</sup>*J*<sub>*H-F*</sub> = 8.0 Hz, 1H), 3.12 (s, 3H), 2.11 (s, 1H), 1.70 (s, 3H), 1.35 (d, *J* = 1.9 Hz, 12H) δ 153.4, 143.1, 142.3, 136.7, 135.6, 129.9, 115.9, 104.7 (q, <sup>2</sup>*J*<sub>*C-F*</sub> = 36.8 Hz), 84.0, 39.4, 29.6, 24.9, 24.7

δ-59.5

Calcd for  $C_{18}H_{23}NBF_3O_3S$ : 416.1309 [M-OH]<sup>+</sup>; found 416.1307

## 5.6. Synthesis of (Z)-3-methyl-1-(methylsulfonyl)-5-phenyl-2-(2,2,2trifluoroethylidene)indolin-3-ol (4gc)

In an oven dried nitrogen purged flask, 4g (100 mg, 1 eq.), phenyl boronic acid (1.2 eq.), K<sub>2</sub>CO<sub>3</sub> (3.0 eq.) were suspended in 1,4-dioxane:H<sub>2</sub>O (1:1; 2.0 mL). The mixture was purged with nitrogen and Pd(PPh<sub>3</sub>)<sub>4</sub> (0.05 eq.) was added. The reaction mixture was stirred at 100°C for 8 h. After completion (as monitored by TLC), the reaction was cooled to rt and filtered through celite. The celite was washed several times with diethyl ether. The combined organic filtrate was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated under reduced pressure. The crude was then purified using silica gel column chromatography to obtain pure product (**4gc**).



Yield: 90 % Colour: Brown liquid Melting point: NA

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):

<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):

<sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): HRMS (APCI) m/z: δ 7.65 – 7.56 (m, 4H), 7.48 – 7.44 (m, 3H), 7.40 – 7.36 (m, 1H), 5.77 (q,  ${}^{3}J_{H-F} = 8.0$  Hz, 1H), 3.18 (s, 3H), 2.26 (s, 1H), 1.73 (s, 3H) δ 149.5 (q,  ${}^{3}J_{C-F} = 5.0$  Hz), 141.4, 140.4, 139.8, 132.0, 129.8, 129.1, 128.0, 127.1, 123.9, 121.9 (q,  ${}^{1}J_{C-F} = 267.5$  Hz), 118.0, 109.0 (q,  ${}^{2}J_{C-F} = 36.3$  Hz), 74.1, 39.1 δ -59.5 Calcd for C<sub>18</sub>H<sub>16</sub>O<sub>3</sub>NF<sub>3</sub>S: 366.0770 [M-OH]<sup>+</sup>; found 366.0769

#### 6. X-Ray Crystal Data

X-ray data for the compound was collected at room temperature on a Bruker D8 QUEST instrument with an I $\mu$ S Mo microsource ( $\lambda = 0.7107$  A) and a PHOTON-III detector. The raw data frames were reduced and corrected for absorption effects using the Bruker Apex 3 software suite programs [1]. The structure was solved using intrinsic phasing method and further refined with the SHELXL program and expanded using Fourier techniques [2]. Anisotropic displacement parameters were included for all non-hydrogen atoms. All C bound H atoms were positioned geometrically and treated as riding on their parent C atoms [C-H = 0.93-0.97 Å, and Uiso(H) = 1.5Ueq(C) for methyl H or 1.2Ueq(C) for other H atoms].

#### Crystal structure determination of 2a



**Crystal Data** for  $C_{10}H_7O_2F_3$  (*M*=216.16 g/mol): monoclinic, space group P2<sub>1</sub> (no. 4), *a* = 8.510(8) Å, *b* = 4.956(4) Å, *c* = 11.481(9) Å,  $\beta$  = 106.91(3)°, *V* = 463.3(7) Å<sup>3</sup>, *Z* = 2, *T* = 294.15 K,  $\mu$ (MoK $\alpha$ ) = 0.146 mm<sup>-1</sup>, *Dcalc* = 1.549 g/cm<sup>3</sup>, 9200 reflections measured (5.002° ≤ 2 $\Theta$  ≤ 60.688°), 2488 unique (*R*<sub>int</sub> = 0.0696, R<sub>sigma</sub> = 0.0724) which were used in all calculations. The

final  $R_1$  was 0.0629 (I > 2 $\sigma$ (I)) and  $wR_2$  was 0.1962 (all data). CCDC 2352135 deposition number contains the supplementary crystallographic data for this paper which can be obtained free of charge at <u>https://www.ccdc.cam.ac.uk/structures/</u>

#### Crystal structure determination of 4g



**Crystal Data** for C<sub>11</sub>H<sub>9</sub>BrF<sub>3</sub>NO<sub>3</sub>S (M =372.16 g/mol): orthorhombic, space group Pna2<sub>1</sub> (no. 33), a = 5.4629(7) Å, b = 21.481(3) Å, c = 11.2799(13) Å, V = 1323.7(3) Å<sup>3</sup>, Z = 4, T = 294.15 K,  $\mu$ (MoK $\alpha$ ) = 3.305 mm<sup>-1</sup>, *Dcalc* = 1.868 g/cm<sup>3</sup>, 9707 reflections measured (6.74° ≤ 2 $\Theta$  ≤ 56.542°), 2982 unique ( $R_{int}$  = 0.0297,

 $R_{sigma} = 0.0459$ ) which were used in all calculations. The final  $R_1$  was 0.0293 (I > 2 $\sigma$ (I)) and  $wR_2$  was 0.0628 (all data). **CCDC 2366081** deposition numbers contains the supplementary crystallographic data for this paper which can be obtained free of charge at https://www.ccdc.cam.ac.uk/structures/

#### 7. Computational Data

For better understanding of the electronic effect of the intermediate formed in the plausible reaction mechanism, density functional theory (DFT) was performed in the gas phase using Gaussian09 software. To perform the DFT calculations, B3LYP (Becke three-parameter Lee–Yang–Parr exchange correlation functional) and the 6-311G++(d,p) as basis set was used [3]. Geometries were optimized, and the frequency calculations showed no negative eigenvalue. The optimized geometries were used to perform natural bond orbital (NBO) calculations to calculate the atomic charges present in the molecule [4]. Later this was used to evaluate Fukui indices for calculating nucleophilic centre whether at position 1 or 2 (**Table S1**) using below equation.

## $f \mathrel{+}= Qk(N+1) - Qk(N)$

Where, Qk is charge on atom k; N is neutral system (or system which participates in the reaction mechanism); N+1 is one electron added to the N system. The data are tabulated in **Table 2**. Fukui indices provide an insight into the most probable nucleophilic (and electrophilic) attack site. Based on the reaction mechanism there arise two possibilities, the attack can probably take place *via* site 1 or site 2. Upon comparison, site 1 is favoured for nucleophilic attack compared to position 2. The Fukui indices are positive at site 1 compared to the negative values at site 2. The formation of 5-exo-dig cyclization is justified as negative Fukui indices are insignificant [5, 6].

**Table S1**. Fukui indices for understanding the nucleophilic probability at the respective atom.



Intermediat e	Charge on atom			f+	
	Χ	1	2	For 1	For 2
2g'	-0.673	0.089	-0.214	0.108	-0.105
<b>4a'</b>	-0.855	0.139	-0.275	0.014	-0.006
4b'	-0.845	0.086	-0.196	0.065	-0.100
4c'	-0.745	0.146	-0.216	0.136	-0.057

6a'	0.121	0.145	-0.277	0.054	-0.012
-NH <sub>2</sub>	-0.832	0.157	-0.277	0.056	-0.014

## References

- [1] B. APEX, Madison, WI, USA, (2016).
- [2] G.M. Sheldrick, *Acta Crystallographica Section A: Foundations and Advances*, **71** (2015) 3-8.
- [3] A. Frisch, *Wallingford*, USA, 25p, 470 (2009).
- [4] A.E. Reed, L.A. Curtiss, F. Weinhold, *Chem. Rev.*, **88** (1988) 899-926.
- [5] T. Otsuka, N. Okimoto, H. Saito, M. Taiji, J. Phys. Conf. Ser., 1290 (2019) 012021.
- [6] P. Bultinck, D. Clarisse, P.W. Ayers, R. Carbo-Dorca, Phys. Chem. Chem. Phys., 13 (2011) 6110-6115.

#### 8. Spectra of the synthesized compounds

#### <sup>1</sup>H NMR for 2a at 400 MHz



## <sup>13</sup>C NMR for 2a at 125 MHz



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

#### <sup>19</sup>F NMR for 2a at 376 MHz

--57.58 OH Parameter Value C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/3a-w/ 3g/02-9023-LRS-19F.jdx 1 Data File Name 2 Title 3 Comment 4 Origin 5 Owner 6 Site Bruker BioSpin GmbH nmr 7 Spectro 8 Author spect 9 Solvent 10 Temperature 11 Pulse Sequence 12 Number of Scans CDCI3 298.2 zgfhigqn.2 16 12 Number of Scarts 13 Receiver Gain 14 Relaxation Delay 15 Pulse Width 16 Acquisition Time 17 Acquisition Date 18 Modification Date 210 3.0000 18.0000 0.7340 2022-06-02T13:14:26 2022-06-02T00:00:00 376.46 19 Spectrometer Frequency 20 Spectral Width 89285.0 21 Lowest Frequency 22 Nucleus 23 Acquired Size -82292.7 19F 65536 24 Spectral Size 131072

-50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

#### HRMS for 2a in APCI (-) MODE.

-10 -20 -30

-40

10 0



#### <sup>1</sup>H NMR for 2b at 500 MHz



#### 

#### <sup>13</sup>C NMR for 2b at 100 MHz



#### <sup>19</sup>F NMR for 2b at 376 MHz



## HRMS for 2b in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2c at 500 MHz



## <sup>13</sup>C NMR for 2c at 100 MHz



#### <sup>19</sup>F NMR for 2c at 376 MHz



## HRMS for 2c in APCI (-) MODE.



#### <sup>1</sup>H NMR for 2d at 400 MHz



#### <sup>13</sup>C NMR for 2d at 100 MHz



## <sup>19</sup>F NMR for 2d at 376 MHz



-80

-90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

## HRMS for 2d in APCI (-) MODE.

-10 -20 -30

-40 -50 -60 -70

10 0



#### <sup>1</sup>H NMR for 2e at 500 MHz



#### <sup>13</sup>C NMR for 2e at 75 MHz



## <sup>19</sup>F NMR for 2e at 376 MHz



HRMS for 2e in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2f at 400 MHz



## <sup>13</sup>C NMR for 2f at 100 MHz



#### <sup>19</sup>F NMR for 2f at 376 MHz





#### <sup>1</sup>H NMR for 2g at 400 MHz



# <sup>13</sup>C NMR for 2g at 75 MHz



#### <sup>19</sup>F NMR for 2g at 376 MHz



## HRMS for 2g in APCI (+) MODE.


#### <sup>1</sup>H NMR for 2h at 500 MHz



## <sup>13</sup>C NMR for 2h at 100 MHz



130 120 110 100 90 f1 (ppm) 210 200 170 160 -10

#### <sup>19</sup>F NMR for 2h at 376 MHz



#### HRMS for 2h in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2i at 400 MHz



## <sup>13</sup>C NMR for 2i at 100 MHz



## <sup>19</sup>F NMR for 2i at 376 MHz



## HRMS for 2i in APCI (+) MODE.



## <sup>1</sup>H NMR for 2j at 400 MHz



## <sup>13</sup>C NMR for 2j at 100 MHz



#### <sup>19</sup>F NMR for 2j at 376 MHz



# HRMS for 2j in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2k at 400 MHz



## <sup>13</sup>C NMR for 2k at 100 MHz



## <sup>19</sup>F NMR for 2k at 376 MHz



## HRMS for 2k in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2l at 500 MHz



## <sup>13</sup>C NMR for 2l at 125 MHz



## <sup>19</sup>F NMR for 2l at 376 MHz



# HRMS for 2l in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2m at 400 MHz



## <sup>13</sup>C NMR for 2m at 100 MHz



## <sup>19</sup>F NMR for 2m at 376 MHz

Parameter	Value
1 Data File Name	C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/3a-w/3q/ 02-4790-LRS-19F.jdx
2 Title	
3 Comment	
4 Origin	Bruker BioSpin GmbH
5 Owner	nmr
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	CDCl3
10 Temperature	298.2
11 Pulse Sequence	zgfhigqn.2
12 Number of Scans	16
13 Receiver Gain	210
14 Relaxation Delay	3.0000
15 Pulse Width	18.0000
16 Acquisition Time	0.7340
17 Acquisition Date	2023-02-02T16:28:55
18 Modification Date	2023-02-02T16:29:00
19 Spectrometer Frequency	376.46
20 Spectral Width	89285.0
21 Lowest Frequency	-82292.7
22 Nucleus	19F
23 Acquired Size	65536
24 Spectral Size	131072



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1(ppm)

# HRMS for 2m in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2n at 400 MHz



## <sup>13</sup>C NMR for 2n at 100 MHz



#### <sup>19</sup>F NMR for 2n at 376 MHz

---57.44 HO Ph Br Parameter Value C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/3a-w/3n/ 07-2485-LRS-19F.jdx 1 Data File Name 2 Title 3 Comment 4 Origin 5 Owner 6 Site Bruker BioSpin GmbH nmr 7 Spectrometer 8 Author spect 9 Solvent 10 Temperature 11 Pulse Sequence CDCI3 295.5 zgfhigqn.2 16 210 12 Number of Scans 13 Receiver Gain 14 Relaxation Delay 3.0000 15 Pulse Width 16 Acquisition Time 17 Acquisition Date 18.0000 0.7999 2022-09-07T14:03:19 19 Spectrometer Frequency 20 Spectral Width 2022-09-07T14:03:00 376.46 89285.0 21 Lowest Frequency 22 Nucleus 23 Acquired Size -82292.7 19F 71424 24 Spectral Size 131072 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm) 10 ò -10 -20 -30 -40 -50 -60 -70 -80

## HRMS for 2n in APCI (+) MODE.



#### <sup>1</sup>H NMR for 20 at 500 MHz



#### <sup>13</sup>C NMR for 20 at 100 MHz



## <sup>19</sup>F NMR for 20 at 376 MHz

---57.46 HO,\_Ph Parameter Value CL C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/3a-w/3o/ 28-9656-LRS-19F.jdx 1 Data File Name 2 Title 3 Comment 4 Origin 5 Owner 6 Site Bruker BioSpin GmbH nmr 7 Spectrometer 8 Author spect 8 Author 9 Solvent 10 Temperature 11 Pulse Sequence 12 Number of Scans 13 Receiver Gain 14 Relaxation Delay 15 Pulse Width 16 Acquisition Time 17 Acquisition Time 18 Modification Date 18 Spectrometer CDCl3 298.2 zgfhigqn.2 16 210 3.0000 18.0000 0.7340 2022-10-28T12:03:21 2022-10-28T12:03:00 18 Modification Date 19 Spectrometer Frequency 20 Spectral Width 21 Lowest Frequency 22 Nucleus 23 Acquired Size 24 Spectral Size 376.46 89285.0 -82292.7 19F 65536 131072 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

HRMS for 20 in APCI (+) MODE.



### <sup>1</sup>H NMR for 2p at 400 MHz



## <sup>13</sup>C NMR for 2p at 100 MHz



## <sup>19</sup>F NMR for 2p at 376 MHz

Parameter	Value
1 Data File Name	C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/3a-w/3p/ 02-4788-LRS-19F.jdx
Title	
3 Comment	
4 Origin	Bruker BioSpin GmbH
5 Owner	omr
5 Site	
7 Spectrometer	spect
3 Author	
Solvent	CDCI3
10 Temperature	298.2
11 Pulse Sequence	zafhiaan. 2
12 Number of Scans	16
13 Receiver Gain	210
14 Pelavation Delay	3 0000
15 Pulse Width	18.0000
16 Acquisition Time	0.7340
17 Acquisition Date	2022 02 02716-21-24
19 Modification Date	2022 02 02 10 21 22
10 Produced up to Produce	2023-02-02110:21:00
Frequency	370.40
20 Spectral Width	89285.0
21 Lowest Frequency	-82292.7
22 Nucleus	19F
23 Acquired Size	65536
24 Spectral Size	131072

32

-40 -50 -60 -70 -80

-90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

# HRMS for 2p in APCI (+) MODE.

10 0 -10 -20 -30



#### <sup>1</sup>H NMR for 4a at 400 MHz



## <sup>13</sup>C NMR for 4a at 100 MHz



#### <sup>19</sup>F NMR for 4a at 376 MHz

74. 62----Parameter Value C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/5a-q/5a/ 17-1043-LRS-19F.jdx но 1 Data File Name 2 Title 3 Comment Ν ·F 0=\$=0**F** 4 Origin 5 Owner 6 Site Bruker BioSpin GmbH F nmr 7 Spectrometer 8 Author spect 9 Solvent CDCI3 10 Temperature 11 Pulse Sequence 298.2 zgfhigqn.2 12 Number of Scans 13 Receiver Gain 16 210 14 Relaxation Delay 3.0000 15 Pulse Width 16 Acquisition Time 18.0000 0.7340 17 Acquisition Date 18 Modification Date 2022-06-17T12:20:08 2022-06-17T00:00:00 19 Spectrometer Frequ ncy 376.46 20 Spectral Width 89285.0 21 Lowest Frequency -82292.7 22 Nucleus 23 Acquired Size 19F 65536 24 Spectral Size 131072 10 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm) 0

## HRMS for 4a in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4b at 500 MHz



## <sup>13</sup>C NMR for 4b at 100 MHz



## <sup>19</sup>F NMR for 4b at 376 MHz

----58.79

Parameter	Value
1 Data File Name	C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/5a-q/5b/ 20-8790-LRS-19F.jdx
2 Title	
3 Comment	
4 Origin	Bruker BioSpin GmbH
5 Owner	nmr
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	CDCl3
10 Temperature	298.2
11 Pulse Sequence	zgfhigqn.2
12 Number of Scans	16
13 Receiver Gain	210
14 Relaxation Delay	3.0000
15 Pulse Width	18.0000
16 Acquisition Time	0.7340
17 Acquisition Date	2022-10-20T14:33:54
18 Modification Date	2022-10-20T14:34:00
19 Spectrometer Frequency	376.46
20 Spectral Width	89285.0
21 Lowest Frequency	-82292.7
22 Nucleus	19F
23 Acquired Size	65536
24 Spectral Size	131072



## 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

## HRMS for 4b in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4c at 400 MHz



### <sup>13</sup>C NMR for 4c at 100 MHz



## <sup>19</sup>F NMR for 4c at 470 MHz



# HRMS for 4c in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4d at 400 MHz



## <sup>13</sup>C NMR for 4d at 125 MHz



## <sup>19</sup>F NMR for 4d at 376 MHz

Parameter	Value
1 Data File Name	C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/5a-q/5g/ 31-4218-LRS-19F.jdx
2 Title	
3 Comment	
4 Origin	Bruker BioSpin GmbH
5 Owner	nmr
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	CDCl3
10 Temperature	298.2
11 Pulse Sequence	zafhigan.2
12 Number of Scans	16
13 Receiver Gain	210
14 Relaxation Delay	3.0000
15 Pulse Width	18.0000
16 Acquisition Time	0.7340
17 Acquisition Date	2023-01-31T11:00:33
18 Modification Date	2023-01-31T11:00:00
19 Spectrometer Frequency	376.46
20 Spectral Width	89285.0
21 Lowest Frequency	-82292.7
22 Nucleus	19F
23 Acquired Size	65536
24 Spectral Size	131072

-60 -70 -80

-90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

HRMS for 4d in APCI (+) MODE.

10 0

-10 -20 -30 -40 -50



#### <sup>1</sup>H NMR for 4e at 400 MHz



#### <sup>13</sup>C NMR for 4e at 100 MHz



## <sup>19</sup>F NMR for 4e at 376 MHz



-90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -21 f1 (ppm)

-70 -80

# HRMS for 4e in APCI (+) MODE.

10 0 -10 -20 -30 -40 -50 -60



## <sup>1</sup>H NMR for 4f at 400 MHz



## <sup>13</sup>C NMR for 4f at 125 MHz in CDCl<sub>3</sub> and DMSO-d<sub>6</sub> mixture.



<sup>19</sup>F NMR for 4f at 376 MHz



---59.76

## HRMS for 4f in APCI (+) MODE.

10 0 -10 -20 -30 -40

-50 -60 -70 -80



#### <sup>1</sup>H NMR for 4g at 400 MHz



# <sup>13</sup>C NMR for 4g at 125 MHz in CDCl<sub>3</sub> and DMSO-d<sub>6</sub> mixture.



# <sup>19</sup>F NMR for 4g at 376 MHz

Parameter	Value
1 Data File Name	C:/Users/ayazm/OneDrive/ Desktop/2btp/nmr/5a-q/5j/ 01-4413-LRS-19F.jdx
2 Title	
3 Comment	
4 Origin	Bruker BioSpin GmbH
5 Owner	nmr
6 Site	
7 Spectrometer	spect
8 Author	
9 Solvent	CDCl3
10 Temperature	298.2
11 Pulse Sequence	zgfhiggn.2
12 Number of Scans	16
13 Receiver Gain	210
14 Relaxation Delay	3.0000
15 Pulse Width	18.0000
16 Acquisition Time	0.7340
17 Acquisition Date	2023-02-01T21:47:14
18 Modification Date	2023-02-01T21:47:00
19 Spectrometer Frequency	376.46
20 Spectral Width	89285.0
21 Lowest Frequency	-82292.7
22 Nucleus	19F
23 Acquired Size	65536
24 Spectral Size	131072



-90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)

# HRMS for 4g in APCI (+) MODE.

10 0 -10 -20 -30 -40 -50 -60 -70 -80



## <sup>1</sup>H NMR for 4h at 400 MHz



## <sup>13</sup>C NMR for 4h at 125 MHz in CDCl<sub>3</sub> and DMSO-d<sub>6</sub> mixture.



## <sup>19</sup>F NMR for 4h at 376 MHz



### HRMS for 4h in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4i at 500 MHz



## <sup>13</sup>C NMR for 4i at 100 MHz



## <sup>19</sup>F NMR for 4i at 376 MHz



# HRMS for 4i in APCI (+) MODE.


#### <sup>1</sup>H NMR for 4j at 400 MHz



# <sup>13</sup>C NMR for 4j at 100 MHz



# <sup>19</sup>F NMR for 4j at 376 MHz



# HRMS for 4j in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4k at 400 MHz



### <sup>13</sup>C NMR for 4k at 100 MHz



### <sup>19</sup>F NMR for 4k at 376 MHz



HRMS for 4k in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4l at 400 MHz



## <sup>13</sup>C NMR for 4l at 100 MHz



### <sup>19</sup>F NMR for 4l at 376 MHz



# HRMS for 4l in APCI (+) MODE.



## <sup>1</sup>H NMR for 4m at 300 MHz



### <sup>13</sup>C NMR for 4m at 75 MHz

Data File Name					
	C:/Users/ayazm/Desktop/nmr/ 10june/12-4976-LRS-13C.jdx				
Title					но
Comment		0.8	8 0 5 8	99 99 99	CI CI
Origin	Bridden BieSpin Carble	123	124.0 126.1	108.108	
Origin	Bruker biospin Gribh	Y	1111	512	
Site					MIS F F
unu					
Spectrometer	Avance Neo Nanobay				
Author			1.1		
Solvent	CDCl3				
0 Temperature	298.2			dh	
1 Pulse Sequence	zgpg30	mahar	بالماليا سيالها	munillimm	
2 Number of Scans	3072				
3 Receiver Gain	45	152	128	112	
4 Relaxation Delay	2.0000	f1 (ppm)	f1 (ppm)	f1 (ppm)	
5 Pulse Width	10.0000				
Acquisition Time	0.5000				
7 Acquisition Date	2024-06-12T19:24:51	1 1.			
8 Modification Date	2024-06-12T20:09:53	11			
Spectrometer Frequence	y 75.48				
Spectral Width	17856.9				
Lowest Frequency	-1371.1				
Nucleus	13C				
Acquired Size	8928		1		1
Contraction of Contract	CEE2C				

### <sup>19</sup>F NMR for 4m at 376 MHz



# HRMS for 4m in APCI (-) MODE.



#### <sup>1</sup>H NMR for 4n at 400 MHz



#### <sup>13</sup>C NMR for 4n at 100 MHz



### <sup>19</sup>F NMR for 4n at 470 MHz



## HRMS for 4n in APCI (+) MODE.



#### <sup>1</sup>H NMR for 40 at 400 MHz



### <sup>13</sup>C NMR for 40 at 100 MHz



### <sup>19</sup>F NMR for 40 at 376 MHz



# HRMS for 40 in APCI (+) MODE.



#### <sup>1</sup>H NMR for 6a at 400 MHz



## <sup>13</sup>C NMR for 6a at 100 MHz



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

### <sup>19</sup>F NMR for 6a at 376 MHz



HRMS for 6a in APCI (+) MODE.



#### <sup>1</sup>H NMR for 2aa at 400 MHz



## <sup>13</sup>C NMR for 2aa at 100 MHz



### <sup>19</sup>F NMR for 2aa at 376 MHz



## HRMS for 2aa in APCI (-) MODE.



### <sup>1</sup>H NMR for 2ga at 400 MHz



# <sup>13</sup>C NMR for 2ga at 100 MHz



# <sup>19</sup>F NMR for 2ga at 376 MHz



HRMS for 2ga in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4da at 400 MHz



#### <sup>13</sup>C NMR for 4da at 100 MHz



### <sup>19</sup>F NMR for 4da at 470 MHz



HRMS for 4da in APCI (+) MODE.



### <sup>1</sup>H NMR for 4ga at 400 MHz



## <sup>13</sup>C NMR for 4ga at 125 MHz



# <sup>19</sup>F NMR for 4ga at 376 MHz



## HRMS for 4ga in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4gb at 400 MHz



# <sup>13</sup>C NMR for 4gb at 100 MHz



#### <sup>19</sup>F NMR for 4gb at 376 MHz



HRMS for 4gb in APCI (+) MODE.



#### <sup>1</sup>H NMR for 4gc at 400 MHz



### <sup>13</sup>C NMR for 4gc at 100 MHz



# <sup>19</sup>F NMR for 4gc at 376 MHz



HRMS for 4gc in APCI (+) MODE.



HPLC chromatogram for peak purity

















































# HPLC for 2n























HPLC for 4e











# HPLC for 4h











# HPLC for 4l









S107
















#### 9. Deuterium labelling of compound 2g (HRMS of 2g')



### 10. Cartesian Coordinates of the optimized geometries of molecules

Table 1.	Cartesian Coordinates of the optimized geometry of intermediate of 2g	<b>'</b> .
	Free Energy = -874.17189674 Hartree	

-1	1																					
C	-							3.5	523	133	500	2	.0	39	71	10	0	-0	.0	39	25	600
С								2.1	L7:	140	000	2	.3	69	20	70	0	-0	.0	46	91	000
С								1.2	214	485	100	1	.3	50	88	80	0	-0	.0	40	02	300
С								1.5	543	308	400	-0	.0	08	59	50	0	-0	.0	23	36	000
С							-	2.9	92(	022	600	-0	.3	14	61	20	0	-0	• 0	22	42	200
С								3.8	381	620	100	0	.6	93	91	30	0	-0	.0	32	50	700
H							4	4.2	280	622	400	2	• 8	09	63	80	0	-0	.0	41	98	800
H U								1. 1	301	664 535	/UU 500	3 1	.4	0/	80	40 30	0	-0	.0	3⊥ 31	03 00	200
п Н								4 (	20. 238	859	200 200	1	.0	15	22	00	0	-0	0.0	31 41	03 13	500
C							(	0.4	15!	559	200	-1	.1	26	51	60	0	0	.1	64	- 3 74	100
С							(	0.5	549	998	300	-2	.1	73	39	10	0	-1	.0	04	27	400
Н								1.4	19:	176	100	-2	.7	10	95	10	0	-0	. 9	08	60	000
Η							- (	).2	273	397	500	-2	.8	81	93	40	0	-0	. 8	86	78	400
Η							(	).4	18	673	400	-1	.7	07	88	20	0	-1	. 9	97	09	800
0								3.3	324	456	300	-1	.6	30	21	90	0	-0	.0	25	95	200
H							•	4.2	25	979	000	-1	.6	59	63	30	0	0	.1	96	17	200
0 C							_ (	J.: n (	ן∠כ 1 ג	UZ3 000	300 900	- 1 - 0	. 6	29	80	30	0	1 0_	.3	82 16	34 11	900 100
C							_	2.0	) ) 7 '	794	200 200	-0	• = 1	39	22	60	0	-0	0.0	38 28	11 52	200
C								3.4	14: 14:	122	200 500	0	.2	85	21	10	0	-0	.0	17	80	100
F								4.2	21	912	500	-0	.3	49	28	10	0	-0	.9	49	56	700
F							- 3	3.5	59(	076	500	1	.6	24	50	00	0	-0	.2	72	37	500
F								4.0	)69	913	300	0	.0	75	79	50	0	1	.1	78	59	000
1	2 1	5	6	1 1	57	1	0															
2	3 1	.5	8	1.0	), )	1	• •															
3	4 1	.5	9	1.0	)																	
4	5 1	.5	11	L 1	.0																	
5	6 1	.5	16	5 1	.0																	
6	10	1.0	)																			
/ Q																						
9																						
1(	C																					
11	1 12	1.	. 0	18	1.	5	19	1.	. 0													
12	2 13	1.	. 0	14	1.	0	15	1.	. 0													
1.	3																					
⊥' 1'	± 5																					
1	5 17	1.	. 0																			
1	7																					
18	3																					
19	9 20	3.	.0																			
20	J 21	⊥. ₁	.5	2	1	$\cap$	2 1	1	0													
と. つ^	1 22 )	Τ.	. U	23	⊥•	0	Ζ4	⊥.	. 0													
23	- 3																					
24	1																					

-1	1		
С	-3.83578100	0.62845500	1.10602800
С	-3.39404500	1.94113900	1.25609100
С	-2.19171400	2.32945800	0.67077500
С	-1.40026800	1.44003200	-0.05561200
С	-1.84756200	0.10918800	-0.17628600
С	-3.07101200	-0.28368900	0.38434700
Н	-4.77765200	0.30740100	1.53970700
H	-3.98303500	2.65740800	1.82064300
Н	-1.79010100	3.33517600	0.75038800
H	-3.4124/400	-1.30132200	0.24941100
C	-0.11101900	2.09241700	-0.71194400
U U	-0.27931000 -1.12602200	2.02572600	-2.27260600
п U	-1.12002300	2.00932000	-2.32202000
н	-0.46089100	1 02804800	-2 69171100
0	0 11803300	3 33172200	-0 29472200
C	1.09039900	1.25085000	-0.33428700
C	2.13597000	0.73379200	-0.00275200
С	3.40038900	0.22024800	0.43335100
F	4.15209500	-0.28828300	-0.58284200
F	3.26906100	-0.80097000	1.34231600
F	4.18486900	1.15032600	1.04649700
Ν	-1.07603900	-0.85336900	-0.91209700
Η	-0.21840400	-0.48230000	-1.30685800
0	0.26849500	-2.93554800	-1.25453800
0	-2.01268200	-3.12501600	-0.16932400
С	-0.01618800	-2.20969200	1.29162100
Η	0.93015400	-1.68377800	1.18595100
Н	-0.71483900	-1.65052300	1.91252800
H	0.13157600	-3.21535900	1.68459100
S	-0.74951300	-2.40/66800	-0.34690100
1	2 1.5 6 1.5 7 1.0		
2	3 1.5 8 1.0 4 1.5 9 1.0		
4	5 1.5 11 1.0		
5	6 1.5 23 1.0		
6	10 1.0		
7			
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10	)		
11	12 1.0 16 1.5 17 1.0		
12	2 13 1.0 14 1.0 15 1.0		
13	3		
14	<u>1</u> -		
15			
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10	/ LØ 3.U 2 10 1 5		
⊥ č 1 (	$\begin{array}{cccccccccccccccccccccccccccccccccccc$		
т :	$\prime$ $\sim$ $\circ$ $\sim$		

# **Table 2.** Cartesian Coordinates of the optimized geometry of intermediate of **4a'**.Free Energy = -1442.28876931 Hartree

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23 24 1.0 31 1.0

24

25 31 2.0

26 31 2.0

27 28 1.0 29 1.0 30 1.0 31 1.0

28

29

30

31
```

## **Table 3.** Cartesian Coordinates of the optimized geometry of intermediate of 4b'.Free Energy = -1673.37611586 Hartree

-1 1			
С	0.61878700	0.49500900	-3.36441600
С	-0.59671500	1.05364700	-2.97887200
С	-1.21974900	0.61290700	-1.81709600
С	-0.69437000	-0.39636300	-0.99192400
С	0.54788900	-0.94028800	-1.38943200
С	1.18039600	-0.48490800	-2.55731000
Н	1.12691900	0.81982500	-4.26602200
Н	-1.06326800	1.82816300	-3.57938100
Н	-2.16331300	1.06106900	-1.53180100
Н	2.13666900	-0.92258900	-2.82883700
С	-1.41084000	-0.70107400	0.36791200
С	-1.53933000	-2.23239700	0.65716700
Н	-0.55866200	-2.64994900	0.86314000
Н	-2.15743700	-2.34900000	1.55043300
Н	-2.00694500	-2.77357400	-0.17508000
0	-0.80035800	-0.00527300	1.34265100
С	-2.85357200	-0.24167900	0.26873200
С	-4.00316900	0.12554300	0.36063700
С	-5.35192100	0.59101700	0.47785200
F	-6.27516500	-0.41497000	0.40587300
F	-5.69647600	1.47527700	-0.50798000
F	-5.60539600	1.23517600	1.65314200
N	1.20420400	-2.03795400	-0.71807900
Н	1.70798100	-2.59890100	-1.39733300
0	3.50351700	-2.68154000	0.07004300
0	1.75543600	-2.20686300	1.82655200
S	2.36983200	-1.86828700	0.55207900
С	2.88932100	-0.14457000	0.59213200
С	4.17548400	0.16913500	0.16005900
С	2.01625600	0.82443900	1.08476800
С	4.60312100	1.49291400	0.23098000
Н	4.82503200	-0.61274600	-0.21334100
С	2.46798100	2.14133800	1.13293900
Н	0.98688700	0.55592200	1.36580300
С	3.75732000	2.49558600	0.71858600
Н	5.60580200	1.74747900	-0.09912100
Н	1.79120700	2.90857800	1.49664700

С 4.22872600 3.92732600 0.81385800 Η 4.53171000 4.17356200 1.83795000 Η 5.08719100 4.11025100 0.16272800 Η 3.43438300 4.62474000 0.53479400 1 2 1.5 6 1.5 7 1.0 2 3 1.5 8 1.0 3 4 1.5 9 1.0 4 5 1.5 11 1.0 5 6 1.5 23 1.0 6 10 1.0 7 8 9 10 11 12 1.0 16 1.0 17 1.0 12 13 1.0 14 1.0 15 1.0 13 14 15 16 17 18 3.0 18 19 1.5 19 20 1.0 21 1.0 22 1.0 20 21 22 23 24 1.0 27 1.0 24 25 27 2.0 26 27 2.0 27 28 1.0 28 29 1.5 30 1.5 29 31 1.5 32 1.0 30 33 1.5 34 1.0 31 35 1.5 36 1.0 32 33 35 1.5 37 1.0 34 35 38 1.0 36 37 38 39 1.0 40 1.0 41 1.0 39 40

**Table 4.** Cartesian Coordinates of the optimized geometry of intermediate of 4c'.Free Energy = -1200.27486663 Hartree

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-1 1			
С	-0.12936600	3.71250400	-0.21005800
С	1.22794300	3.45187000	-0.03487300
С	1.63442900	2.13086500	0.16135700

С	0.72306200	1.07871100	0.18586800
С	-0.67308100	1.33498800	0.00505900
С	-1.06350800	2.68283900	-0.19228800
н	-0 47078400	4 73249200	-0 36569200
П П	1 05056600	1.25270000	-0 05267900
н 	1.93930000	4.23270000	-0.03207900
H	2.69003800	1.9199/500	0.29563600
Н	-2.11524000	2.88855300	-0.32425400
С	1.18792000	-0.36627700	0.48584500
С	0.97668200	-0.67330600	1.99489900
Н	-0.08604900	-0.56143600	2.21370300
н	1 27997000	-1 70249500	2,20258400
и Ц	1 55072900	0 01288900	2 62338800
0	0 51250100	-1 22220500	_0 20520000
0	0.51250100	-1.55250500	-0.30320000
	2.62263600	-0.54/43800	0.18529200
С	3.79166400	-0.74642400	-0.02109100
С	5.18729600	-0.98550500	-0.28284400
F	5.46835100	-2.30396300	-0.45674000
F	5.98727600	-0.55910600	0.73470200
F	5.63148500	-0.35177100	-1.40052100
Ν	-1.51058300	0.23569300	0.02455900
н	-0 44171900	-1 00715400	-0 27376800
C	-2 020/1000	0.22660600	-0 05427200
C	-2.03941000	1 20150400	-0.03437200
C	-4./9344/00	-1.20158400	-0.15368000
С	-4.89025700	-2.73294200	-0.14195600
Н	-4.45514400	-3.13269700	0.77738400
H	-5.93520800	-3.05265200	-0.20599800
Н	-4.34027900	-3.15316600	-0.98754300
С	-5.36553400	-0.65499100	-1.47119600
Н	-5.30080200	0.43154800	-1.49672600
н	-4 80257400	-1 06043700	-2 31699000
и Ц	-6 /129//00	-0.95760600	-1 57925900
C .	-5 52170700	-0.62425600	1 06429500
	-3.33178700	-0.02425000	1.00420300
H	-6.58583600	-0.92242000	1.03984800
H	-5.08/62900	-1.01121200	1.98618300
Н	-5.46553900	0.46248900	1.07346700
0	-3.36967100	-0.96983200	-0.06333800
0	-3.59064600	1.30594700	-0.10640900
1 2 1.5 6 1.5 7 1	.0		
2 3 1 5 8 1 0			
3 4 1 5 9 1 0			
4 5 1.5 11 1.0			
5 6 1.5 23 1.0			
6 10 1.0			
7			
8			
9			
10			
11 12 1.0 16 1.0	17 1.0		
$12 \ 13 \ 1 \ 0 \ 14 \ 1 \ 0$	15 1 0		
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т.Э 1 Л			
1 G			
16 24 1.0			
17 18 3.0			
18 19 1.5			

## **Table 5.** Cartesian Coordinates of the optimized geometry of intermediate of **6a'**.Free Energy = -1197.15275796 Hartree

-1 1			
С	-3.13075800	2.34931500	-0.20075500
С	-1.87854300	2.63859800	0.33817600
С	-0.98344000	1.59463700	0.56258800
С	-1.29978900	0.26695800	0.26291300
С	-2.56231900	-0.01976200	-0.28347600
С	-3.46318600	1.03339000	-0.50638100
Н	-3.84691900	3.14261900	-0.39233200
Н	-1.59946200	3.66070000	0.57383900
Н	-0.00279700	1.81235200	0.97408400
H	-4.43700800	0.81541500	-0.93359000
С	-0.31453100	-0.91114600	0.54377800
С	-0.40453700	-1.24307100	2.07752000
Н	-1.42116700	-1.59048200	2.27721200
Н	0.29124900	-2.05799400	2.29218100
Н	-0.17812100	-0.38605400	2.72325500
0	-0.56309300	-1.97268400	-0.23262900
С	1.08949500	-0.42567100	0.28844700
С	2.24294200	-0.15719900	0.03874200
С	3.60254500	0.15214800	-0.28180400
F	4.47964000	-0.21067300	0.70352600
F	3.81387700	1.48979200	-0.48248100
F	4.05196800	-0.46076500	-1.41501800
S	-3.02892800	-1.70739200	-0.69049500
H	-4.33482300	-1.33117500	-0.89843900
	- 1 0		
121.561.5 2315810	/ 1.0		

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3 4 1.5 9 1.0
4 5 1.5 11 1.0
5 6 1.5 23 1.0
6 10 1.0
7
8
9
10
11 12 1.0 16 1.5 17 1.0
12 13 1.0 14 1.0 15 1.0
13
14
15
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17 18 3.0
18 19 1.5
19 20 1.0 21 1.0 22 1.0
20
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23 24 1.0
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```

**Table 6.** Cartesian Coordinates of the optimized geometry of intermediate of **amine** analoguewith no protecting group.Free Energy = -854.32325388 Hartree

-1 1			
С	-3.51119600	2.00566200	-0.08603200
С	-2.24085200	2.33346700	0.38264100
С	-1.27536100	1.32578600	0.47700100
С	-1.53696800	0.00308500	0.12733000
С	-2.83748200	-0.32686800	-0.34048000
С	-3.80192300	0.69281800	-0.44369300
Н	-4.27771600	2.76991600	-0.17800900
Н	-1.99747400	3.35332900	0.66196700
Н	-0.27902500	1.58377900	0.82233600
Н	-4.79390000	0.43506700	-0.80755100
С	-0.46393200	-1.13312200	0.24513700
С	-0.57450800	-1.72493100	1.69566800
Н	-1.57028700	-2.16273500	1.80060500
Н	0.17009200	-2.51820000	1.80219800
Н	-0.42660100	-0.97136400	2.47819200
0	-0.56673500	-2.09273200	-0.70805400
С	0.90372200	-0.52467400	0.13902000
С	2.04422100	-0.14255100	0.00825300
С	3.39106500	0.30334100	-0.17341500
F	4.27561900	-0.30503700	0.67406100
F	3.54504200	1.64681000	0.04379500
F	3.87681300	0.08190500	-1.42986800
Ν	-3.14421400	-1.64591600	-0.64624900
Н	-3.88617100	-1.75377400	-1.32433400
Н	-2.25352600	-2.15792700	-0.84771000

```
1 2 1.5 6 1.5 7 1.0
2 3 1.5 8 1.0
3 4 1.5 9 1.0
4 5 1.5 11 1.0
5 6 1.5 23 1.0
6 10 1.0
7
8
9
10
11 12 1.0 16 1.0 17 1.0
12 13 1.0 14 1.0 15 1.0
13
14
15
16
17 18 3.0
18 19 1.5
19 20 1.0 21 1.0 22 1.0
20
21
22
23 24 1.0 25 1.0
24
25
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