

Supporting Information

Efficient Co-Upcycling of Glycerol and CO₂ into Valuable Products Enabled by a Bifunctional Ru-Complex Catalyst

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‡ Dedicated to Prof. Michael Grätzel on the occasion of his 80th birthday.

1. Experimental procedure

FTIR and *in-situ* FTIR measurements:

FTIR spectra were recorded with a Nicolet iS50 spectrometer equipped with an MCT detector. The complex **2** was ground into a powder and placed into sample cells. A liquid film was formed by dripping an aqueous solution of glycerol and K₂CO₃ onto the catalyst powder. After purging with CO₂ for half an hour, the background spectrum was measured by heating to 90 °C under a CO₂ atmosphere. The *in-situ* FTIR measurements were monitored in real-time with a time resolution of 10 min per spectrum at 90 °C.

Calculations of TONs and TOFs:

$$TON_{formate} = \frac{n_{formate}}{n_{cat}}$$

$$TON_{lactate} = \frac{n_{lactate}}{n_{cat}}$$

$$TOF_{formate} = \frac{TON_{formate}}{t}$$

$$TOF_{lactate} = \frac{TON_{lactate}}{t}$$

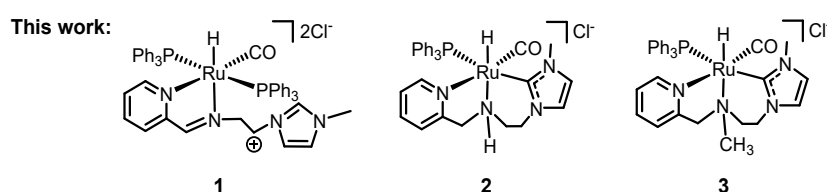
Standard procedures for the upcycling of CO₂ and glycerol and catalyst stability test.

Standard procedure: After water and glycerol (5 mL/5 mL) were mixed and degassed, the degassed solution, catalyst (0.1 μmol) and base (24 mmol) were added into the autoclave with a magnetic stirrer, and then the autoclave was closed and flushed with CO₂ for 3 times. Subsequently, after the autoclave was pressured to the designed pressure with CO₂, it was heated to the desired temperature, the stirrer was turned on and the reaction was performed for 20 hours. At the end of the reaction, the autoclave was cooled to room temperature, then 0.5 mL of D₂O and DMF as an internal standard were added to the reaction mixture for ¹H NMR analysis.

Catalyst stability test: Under standard reaction conditions, the reaction temperature is maintained at 200 °C. Stirring is paused at 5, 20, 50, and 100 hours. A small amount of reaction liquid is then extracted from the reactor using internal pressure for ¹H NMR analysis.

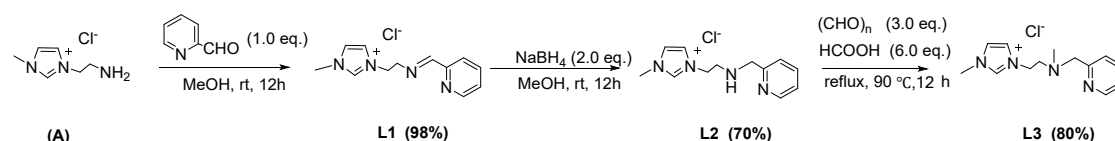
Degassing method:

After the reaction solution was added into a round-bottom flask, it was sealed and connected with the vacuum line. Subsequently, the flask was immersed in liquid nitrogen until the solution was frozen completely. The frozen mixture solution was pumped to vacuum for 5 min, after which the flask was flushed with nitrogen and the solution was allowed to thaw. The freeze-thaw cycle was performed 3 times. After the final cycle, the solution was allowed to thaw completely at room temperature. The degassed solvent was then transferred to the reaction vessel using a syringe with a long needle under a N₂ atmosphere.



Scheme S1. Ru complexes for co-upcycling of CO₂ and glycerol.

1.1 Synthesis and characterization of complex



Scheme S2. Synthesis of ligand **L1**, **L2**, **L3**.

The Ru-CNN complexes **1**, **2** and **3** were prepared according to previous reports with small modification^[1].

Synthesis of 1-methyl-3-{2-[(pyridin-2-ylmethylene)imino]-ethyl}-1H-imidazol-3-ium Chloride (ligand L1, CN^{im}N): A mixture of 2-chloroethylamine hydrochloride and 1-methylimidazole (2.0 eq.) was stirred in dried acetonitrile at 90 °C for 12 h. The precipitate was filtered, washed with EtOH, and then dried in vacuo to give the hydrochloride of **(A)** as a white solid (85%). After the white solid was dissolved in minimal water, the pH value of the aqueous solution was adjusted to 8-9 with KOH solution and stirred for 30 minutes. Subsequently, all water was removed by rotary evaporation and the residue was extracted by the mixture solution of THF/EtOH (1:4, 50mL). The extraction solution was collected and the solvent was removed under

reduced pressure to give the desired product (**A**) as a pale-yellow liquid. Product **A** (1.616g, 10 mmol) was dissolved in 30 mL anhydrous methanol, and pyridine-2-carboxaldehyde (1.071g, 10 mmol) was added to the solution followed by stirring the mixture at room temperature for 12 h. And then the solvent was removed in vacuo to yield the product as brownish yellow solid (2.5 g, 98%). ¹H NMR (400 MHz, DMSO-*d*₆, δ): 9.19 (s, 1H), 8.65 (m, 1H), 8.34 (s, 1H), 7.97 (dt, *J* = 7.9 Hz, 1.2 Hz, 1H), 7.90 (td, *J* = 7.7, 1.8 Hz, 1H), 7.79 (t, *J* = 1.8 Hz, 1H), 7.70 (t, *J* = 1.8 Hz, 1H), 7.49 (m, 1H), 4.54 (t, *J* = 6.0 Hz, 2H), 4.04 (t, *J* = 5.6 Hz, 2H), 3.85 (s, 3H). ¹³C NMR (100 MHz, DMSO-*d*₆, δ): 165.0, 154.1, 149.9, 137.5, 125.9, 123.9, 123.1, 121.2, 59.1, 50.0, 36.2. HRMS (ESI-TOF) *m/z*: [M-Cl]⁺ calcd for C₁₂H₁₅N₄⁺, 215.1297, found 215.1275.

Synthesis of 1-methyl-3-{2-[(pyridin-2-ylmethyl)-amino]-ethyl}-1H-imidazol-3-ium Chloride (ligand L2, CN^HN): NaBH₄ (0.379 g, 10 mmol) was added to a round bottom flask, and then methanol solution of ligand **L1** (2.500g, 10 mmol) was added in it cooled with an ice-water bath. This mixture solution was stirred at room temperature for 12 h. At the end of the reaction, the reaction was quenched with 10% HCl aqueous solution. The pH of the resulting aqueous solution was adjusted to 7 with saturated aqueous NaHCO₃. All solvents were removed by rotary evaporation and residue was extracted with CH₂Cl₂. The extraction solution was purified by column chromatography {200-300 mesh silica gel, dichloromethane/ethanol (10:1), R_f = 0.3} to give ligand **L2** as a pale yellow solid (1.76 g, 70%). ¹H NMR (400 MHz, DMSO-*d*₆, δ): 9.19 (s, 1H), 8.49 (m, 1H), 7.77 (t, *J* = 1.8 Hz, 1H), 7.74 (dd, *J* = 7.7, 1.8 Hz, 1H), 7.70 (t, *J* = 1.8 Hz, 1H), 7.35 (d, *J* = 7.7 Hz, 1H), 7.25 (m, 1H), 4.26 (t, *J* = 5.7 Hz, 2H), 3.87 (s, 3H), 3.80 (s, 2H), 2.91 (t, 1H, *J* = 5.7 Hz, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆, δ): 160.2, 149.2, 137.4, 137.0, 123.7, 123.1, 122.5, 122.3, 54.1, 49.0, 48.3, 36.2. HRMS (ESI-TOF) *m/z*: [M-Cl]⁺ calcd for C₁₂H₁₇N₄⁺, 217.1453, found 217.1450.

Synthesis of 1-methyl-3-{2-[methyl(pyridin-2-ylmethyl)-amino]-ethyl}-1H-imidazol-3-ium Chloride (ligand L3, CN^{Me}N): A mixture of ligand **L2** (801 mg, 3.18 mmol), paraformaldehyde (286.2 mg, 9.54 mmol) and formic acid (878.7 mg, 0.72 mL, 19.09 mmol) was stirred at 110 °C for 12 h, and then a saturated Na₂CO₃ solution was added in it until the solution became the alkaline. The resulting mixture was vigorously stirred for 2 h at 110 °C and solvent was removed by rotary

evaporation. The residue was extracted with CH_2Cl_2 , and the CH_2Cl_2 solution was evaporated under vacuum to give ligand **L3** as a yellow brown oil. (677 mg, 80%). ^1H NMR (400 MHz, $\text{DMSO-}d_6$, δ): 9.38 (s, 1H), 8.47 (m, 1H), 7.78 (t, $J = 1.8$ Hz, 1H), 7.76-7.67 (m, 2H), 7.25 (m, 1H), 7.13 (d, $J = 7.8$ Hz, 1H), 4.35 (t, $J = 5.7$ Hz, 2H), 3.91 (s, 3H), 3.65 (s, 2H), 2.79 (t, $J = 5.7$ Hz, 2H), 2.25 (s, 3H). ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$, δ): 158.9, 149.2, 137.4, 137.0, 123.5, 123.1, 123.1, 122.7, 63.0, 56.1, 47.0, 42.3, 36.2. HRMS (ESI-TOF) m/z : $[\text{M-Cl}]^+$ calcd for $\text{C}_{13}\text{H}_{19}\text{N}_4^+$, 231.1610, found 231.1602.

Synthesis of complex $[\text{RuH}(\text{CO})(\text{PPh}_3)_2(\kappa^2\text{-CN}^{\text{im}}\text{N})](\text{Cl})_2$ (**1**): Ligand **L1** (0.25 mmol, 62.5 mg) and $\text{RuHCl}(\text{CO})(\text{PPh}_3)_3$ (0.25 mmol, 238 mg) were added to a 60 mL mixed solvent of methanol and toluene (1:3) in a two-necked flask under argon atmosphere, and the mixture was stirred at room temperature for 10 h. At the end of the reaction, the solution was concentrated to about 15 mL and 20 ml of ether was added into it to precipitate the desired complex. Subsequently, the precipitate was filtered, washed three times with ether and dried under vacuum to give complex **1** as a bright yellow solid (235 mg, 0.2 mmol, 80%). ^1H NMR ($\text{DMSO-}d_6$, 400 MHz, δ): 9.43 (s, 1H), 8.96 (s, 1H), 7.77 (m, 5H), 7.39 (m, 30H), 6.65 (t, $J = 6.6$ Hz, 1H), 4.06 (m, 4H), 3.92 (s, 3H), -11.23 (t, $J = 20.2$ Hz, 1H). $^{31}\text{P}\{^1\text{H}\}$ NMR ($\text{DMSO-}d_6$, 162 MHz, δ): 44.3 (s). HRMS (ESI-TOF) m/z : $[\text{M-2Cl}]^{2+}$ calcd for $\text{C}_{49}\text{H}_{46}\text{N}_4\text{OP}_2\text{Ru}$, 435.1095, found 435.1093. IR (KBr Disc, cm^{-1}): 1942 (CO).

Synthesis of complex $[\text{fac-RuH}(\text{CO})(\text{PPh}_3)_3-(\kappa^3\text{-CN}^{\text{H}}\text{N})]\text{Cl}$ (**2**): (Method one) Ligand **L2** (0.4 mmol, 100.8 mg), $\text{RuHCl}(\text{CO})(\text{PPh}_3)_3$ (0.4 mmol, 381 mg) and NaBH_4 (0.80 mmol, 30.26 mg) was added to a 60 mL mixed solvent of methanol and toluene in a two-necked flask under argon atmosphere. After the mixture was stirred at 90 °C for 48 h, the solvent was removed in vacuum and then extracted with dehydrated dichloromethane. The volume of dichloromethane solution was concentrated to about 5 mL, and then added with 10 ml of ether to precipitate product. The precipitate was filtered, washed three times with ether and dried under vacuum to give complex **2** as a reddish brown solid (142 mg, 0.22 mmol, 55%). The product is stable to ambient atmosphere in the solid state, and is easy to absorb water in the air. Method two: complex **2** was prepared in a manner analogous to method one except of using $\text{CH}_3\text{CH}_2\text{ONa}$ (1.2 eq.) as base and mixture of ethanol and toluene as the solvent. ^1H NMR (400 MHz, $\text{DMSO-}d_6$, δ): 7.94 (d, $J = 5.5$ Hz, 1H), 7.66

(t, $J = 7.7$ Hz, 1H), 7.43-7.26 (m, 18H), 6.86 (t, $J = 6.6$ Hz, 1H), 5.64 (brs, 1H), 4.18-3.93 (m, 5H), 3.90-3.77 (m, 1H), 3.74-3.54 (m, 2H), 2.42-2.28 (m, 1H), -12.11 (d, $J = 25.1$ Hz, 1H). $^{31}\text{P}\{^1\text{H}\}$ NMR (DMSO- d_6 , 162 MHz, δ): 48.0 (s). ^{13}C NMR (100 MHz, DMSO- d_6 , δ): 206.2 (d, $J = 14.7$ Hz, CO), 179.0 (d, $J = 87.6$ Hz, NHC carbon atom), 159.7, 154.2, 138.2, 135.2 (d, $J = 37.3$ Hz), 133.2 (d, $J = 11.1$ Hz), 130.1, 129.0 (d, $J = 9.0$ Hz), 124.9, 123.0, 122.5, 58.9, 52.6, 48.7, 38.9. HRMS (ESI-TOF) m/z : $[\text{M}-\text{Cl}]^+$ calcd for $\text{C}_{31}\text{H}_{32}\text{N}_4\text{OPRu}$, 609.1357; found, 609.1360. IR (KBr Disc, cm^{-1}): 1923 (CO).

Synthesis of complex $[\text{fac-RuH}(\text{CO})(\text{PPh}_3)-(\kappa^3\text{-CN}^{\text{Me}}\text{N})]\text{Cl}$ (3**):** Complex **3** was prepared with $\text{RuHCl}(\text{CO})(\text{PPh}_3)_3$ and Ligand **L3** by a analogous manner to complex **2** except of using $\text{CH}_3\text{CH}_2\text{ONa}$ (1.2 eq.) as base and mixture of ethanol and toluene as the solvent (pale white solid, 50%). ^1H NMR (DMSO- d_6 , 400 MHz, δ): 7.92 (d, $J = 5.4$ Hz, 1H), 7.72 (t, $J = 7.7$ Hz, 1H), 7.45-7.28 (m, 18H), 6.89 (t, $J = 6.5$ Hz, 1H), 4.15 (dd, $J = 15.4, 7.9$ Hz, 1H), 4.06 (s, 3H), 3.98 (d, $J = 15.8$ Hz, 1H), 3.78 (dd, $J = 15.3, 7.2$ Hz, 1H), 3.68 (d, $J = 15.8$ Hz, 1H), 2.93 (dd, $J = 13.4, 7.3$ Hz, 1H), 2.74 (s, 3H), 2.56 (dd, $J = 13.4, 7.9$ Hz, 1H), -13.31 (d, $J = 26.9$ Hz, 1H). $^{31}\text{P}\{^1\text{H}\}$ NMR (DMSO- d_6 , 162 MHz, δ): 44.9 (s). ^{13}C NMR (100 MHz, DMSO- d_6 , δ): 206.3 (d, $J = 15.3$ Hz, CO), 178.2 (d, $J = 86.7$ Hz, NHC carbon atom), 159.5, 154.1, 138.6, 135.7 (d, $J = 36.7$ Hz), 133.4 (d, $J = 10.5$ Hz), 130.1, 128.9 (d, $J = 8.6$ Hz), 125.0, 123.1, 123.0, 122.9, 68.6, 63.0, 53.5, 47.0, 39.2. HRMS (ESI-TOF) m/z : $[\text{M}-\text{Cl}]^+$ calcd for $\text{C}_{32}\text{H}_{34}\text{N}_4\text{OPRu}$, 623.1514, found, 623.1466. IR (KBr Disc, cm^{-1}): 1918 (CO).

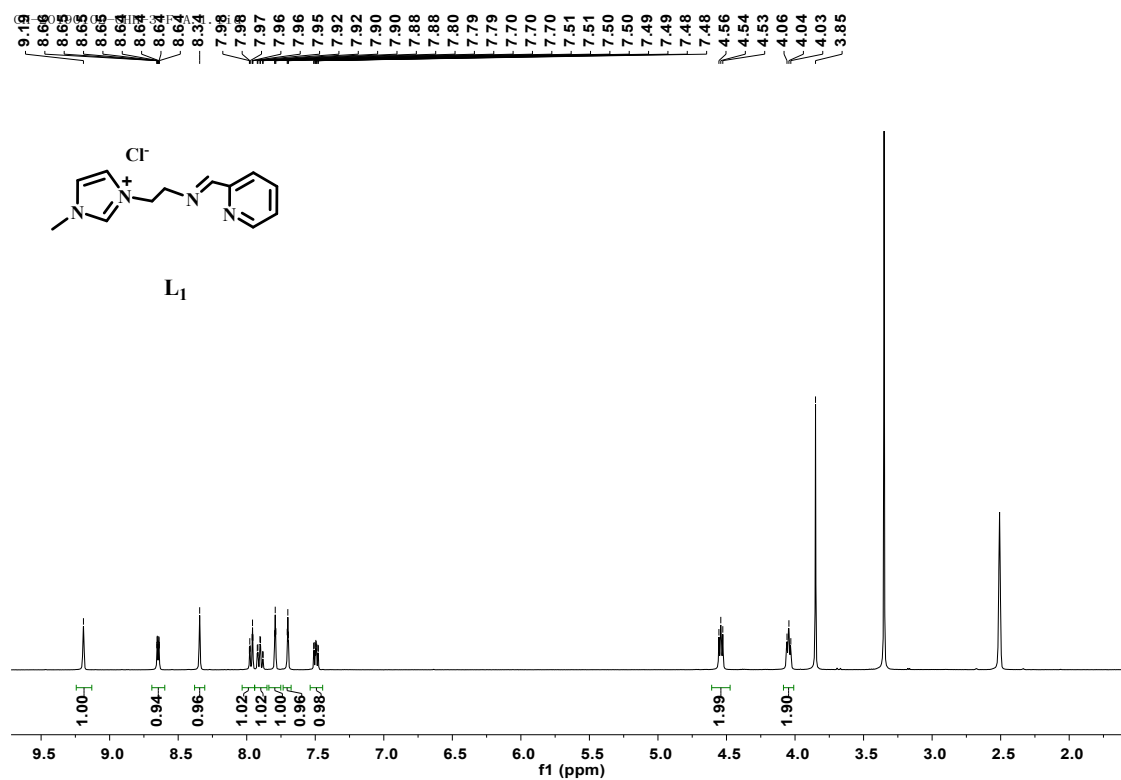


Figure S1. ¹H NMR spectrum of **L1** (400 MHz, DMSO-*d*₆).

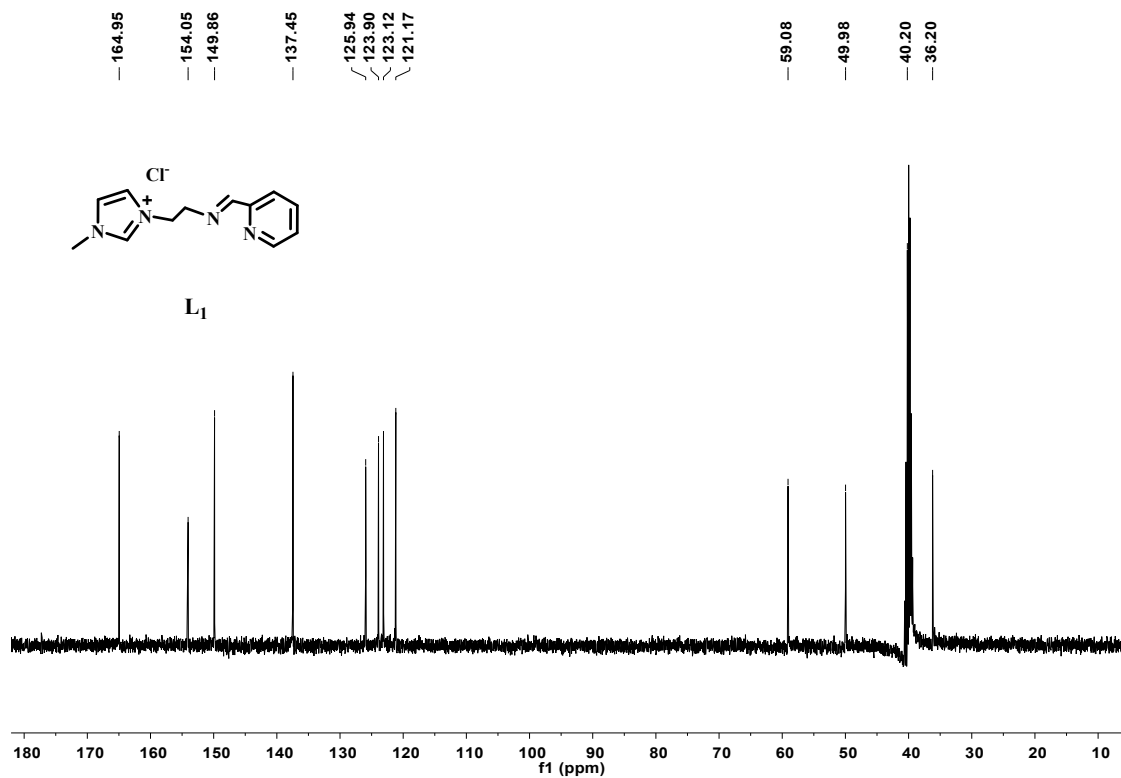


Figure S2. ^{13}C NMR spectrum of L1 (100 MHz, $\text{DMSO-}d_6$).

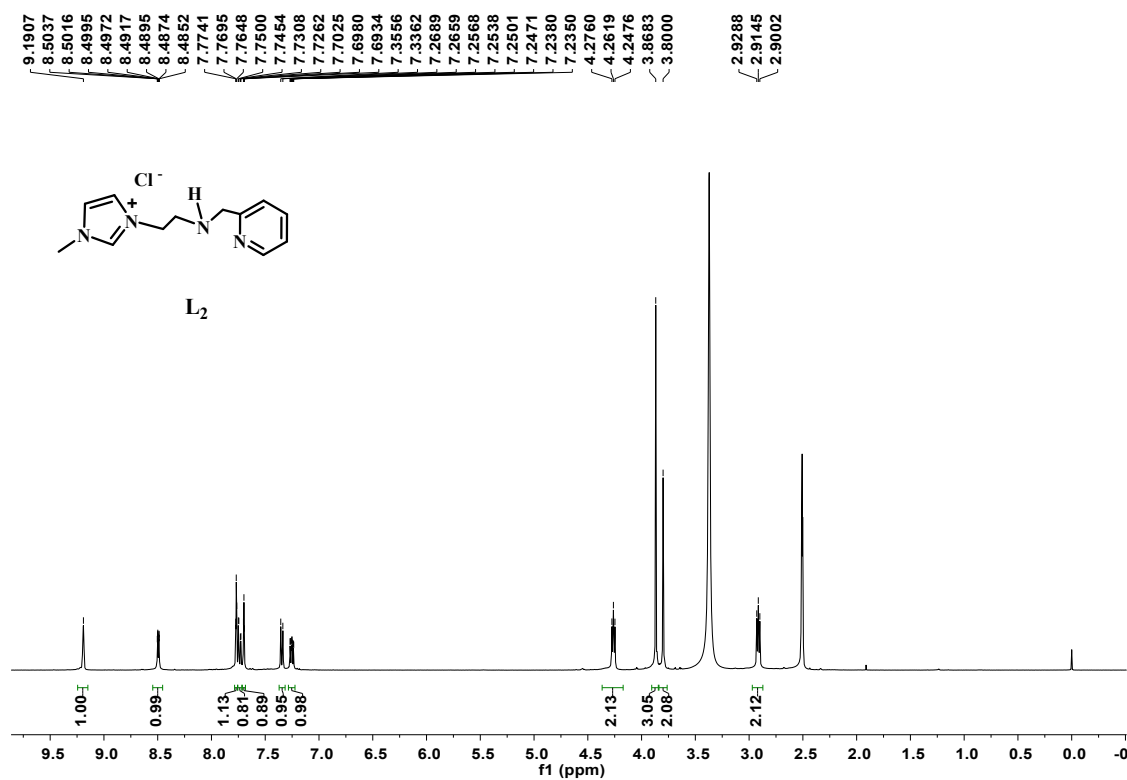


Figure S3. ^1H NMR spectrum of L2 (400 MHz, $\text{DMSO-}d_6$).

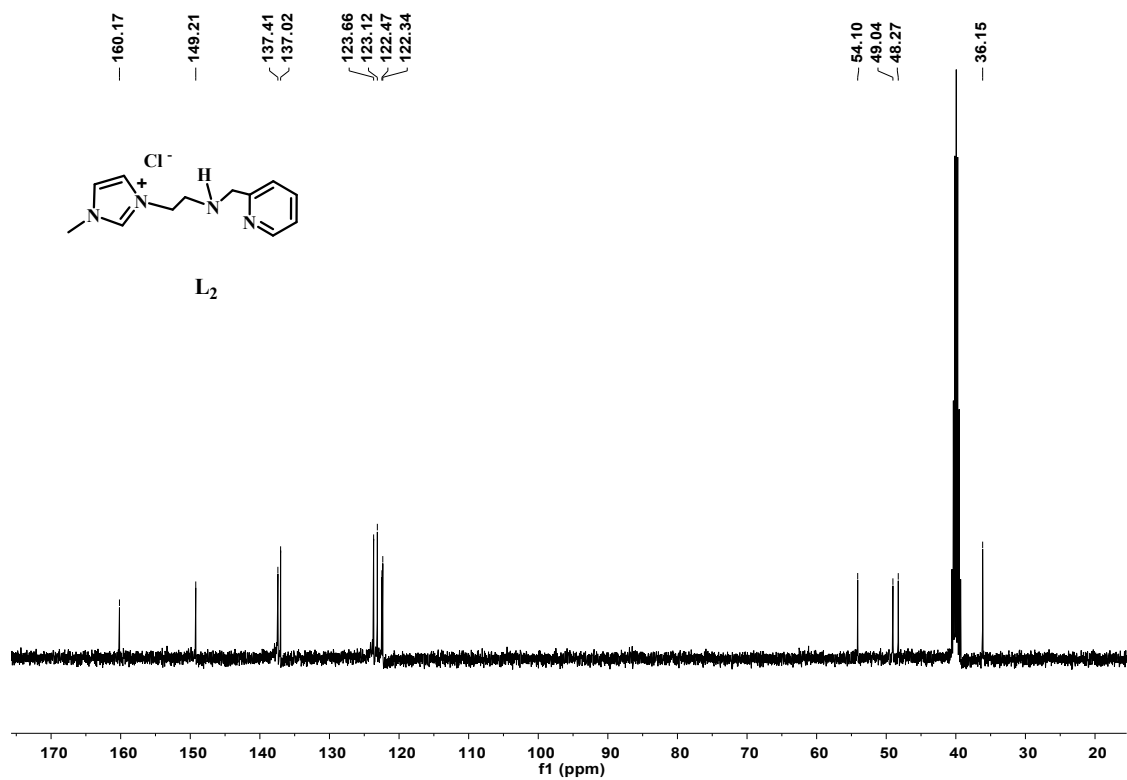


Figure S4. ^{13}C NMR spectrum of L2 (100 MHz, $\text{DMSO-}d_6$).

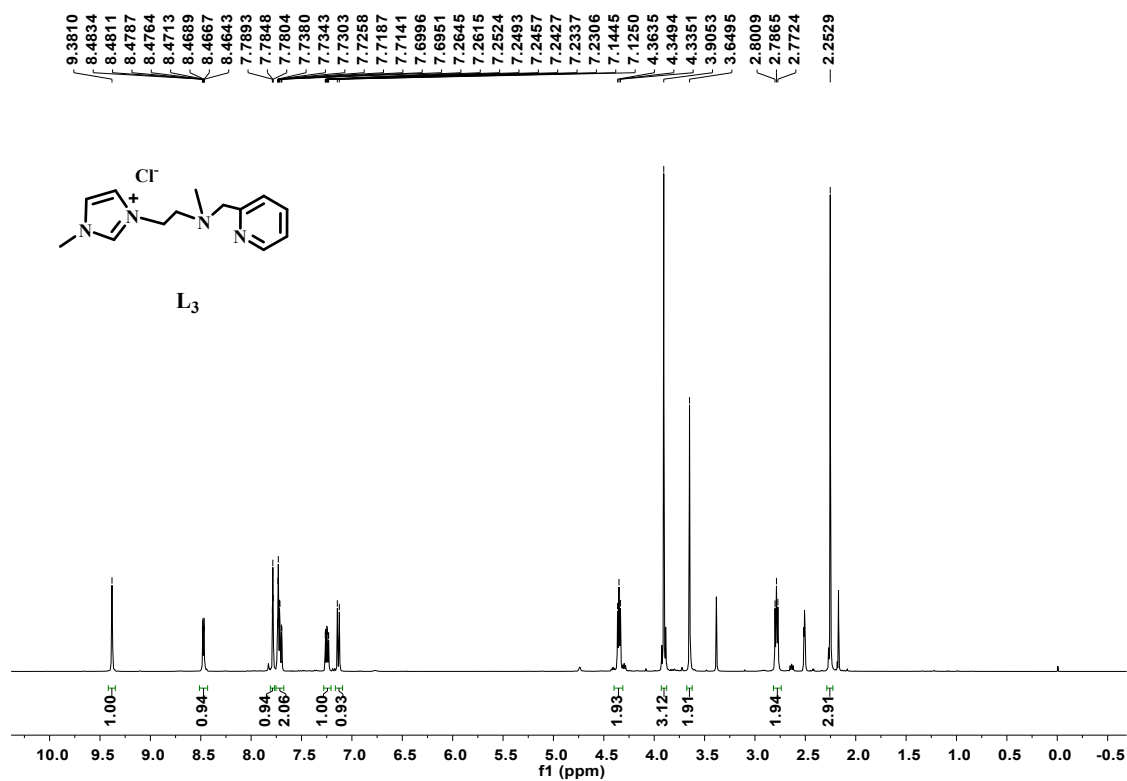


Figure S5. ^1H NMR spectrum of L3 (400 MHz, $\text{DMSO-}d_6$).

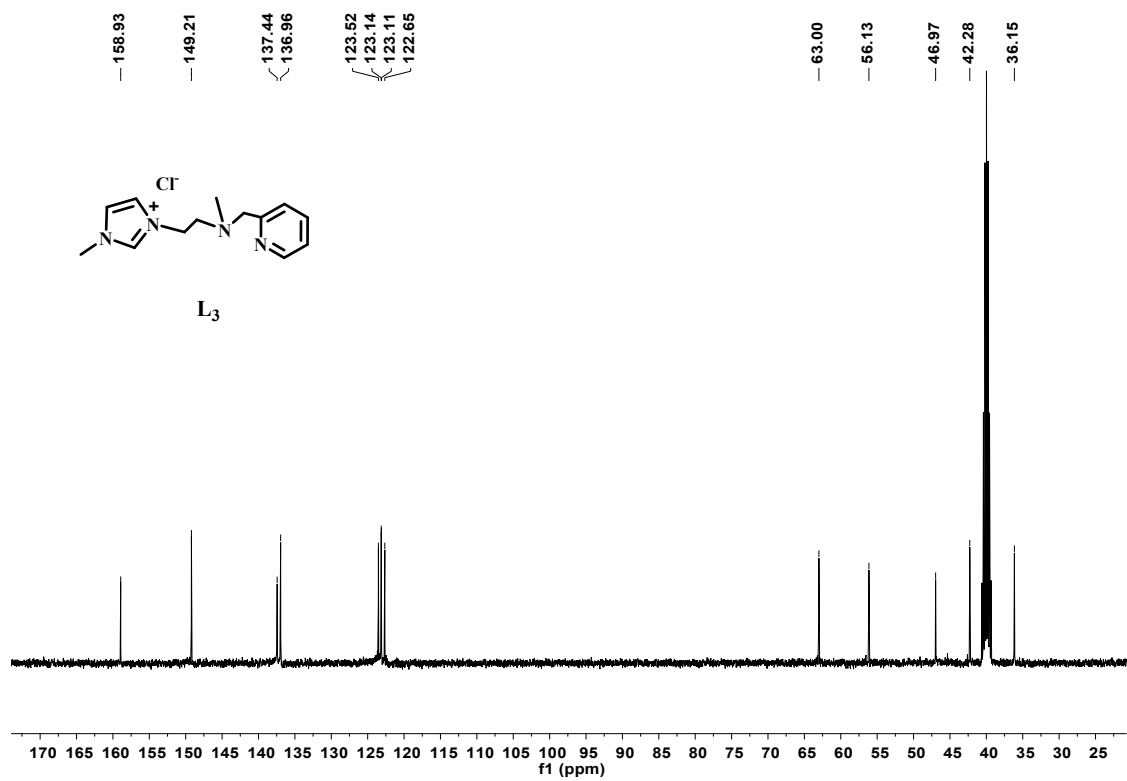


Figure S6. ^{13}C NMR spectrum of **L3** (100 MHz, $\text{DMSO-}d_6$).

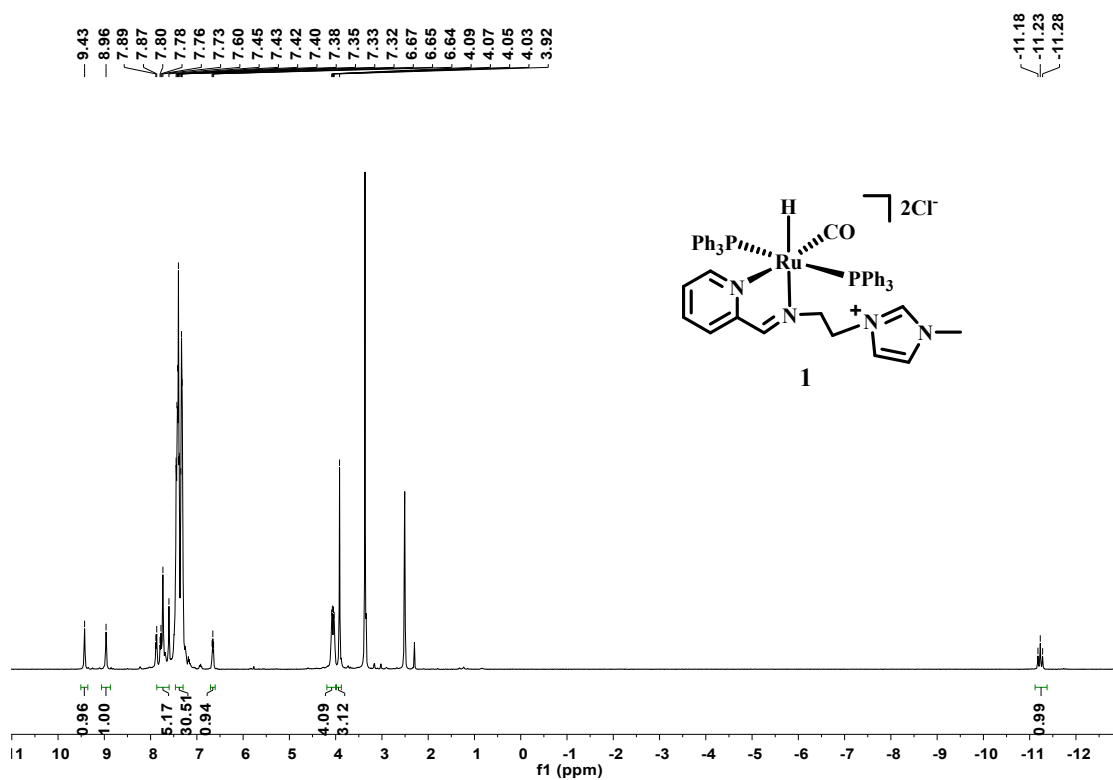


Figure S7. ^1H NMR spectrum of complex **1** (400 MHz, $\text{DMSO-}d_6$).

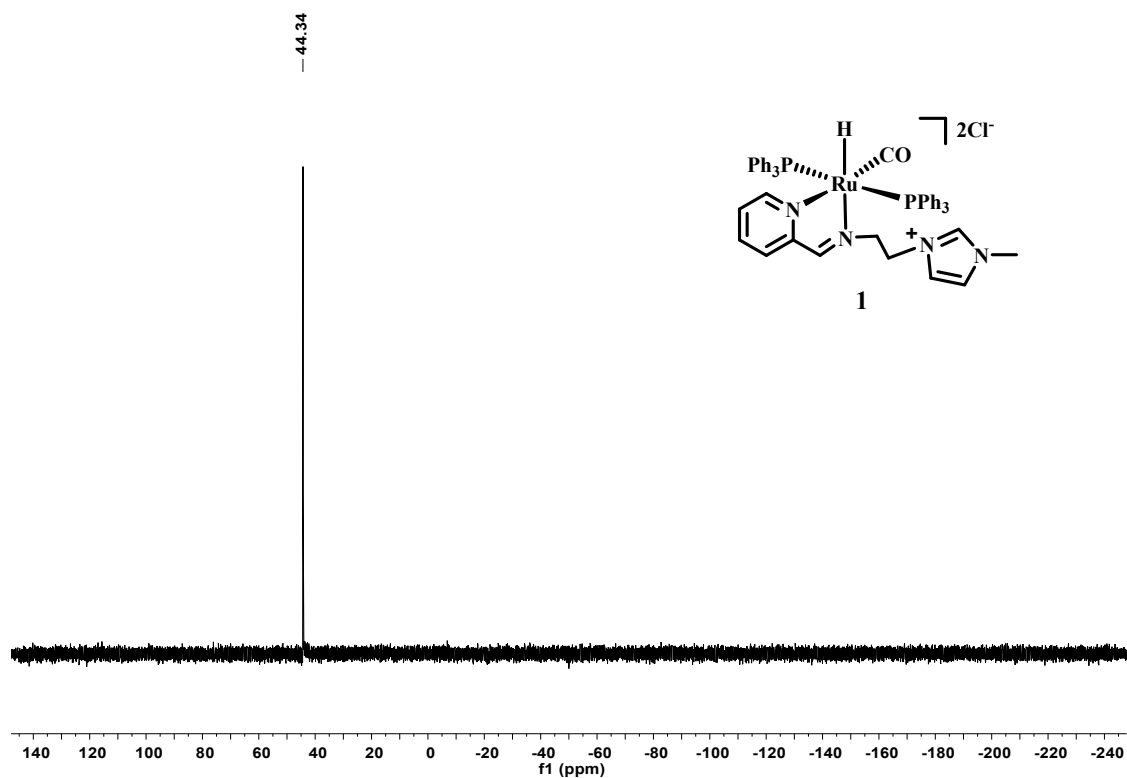


Figure S8. $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum of complex **1** (162 MHz, $\text{DMSO-}d_6$).

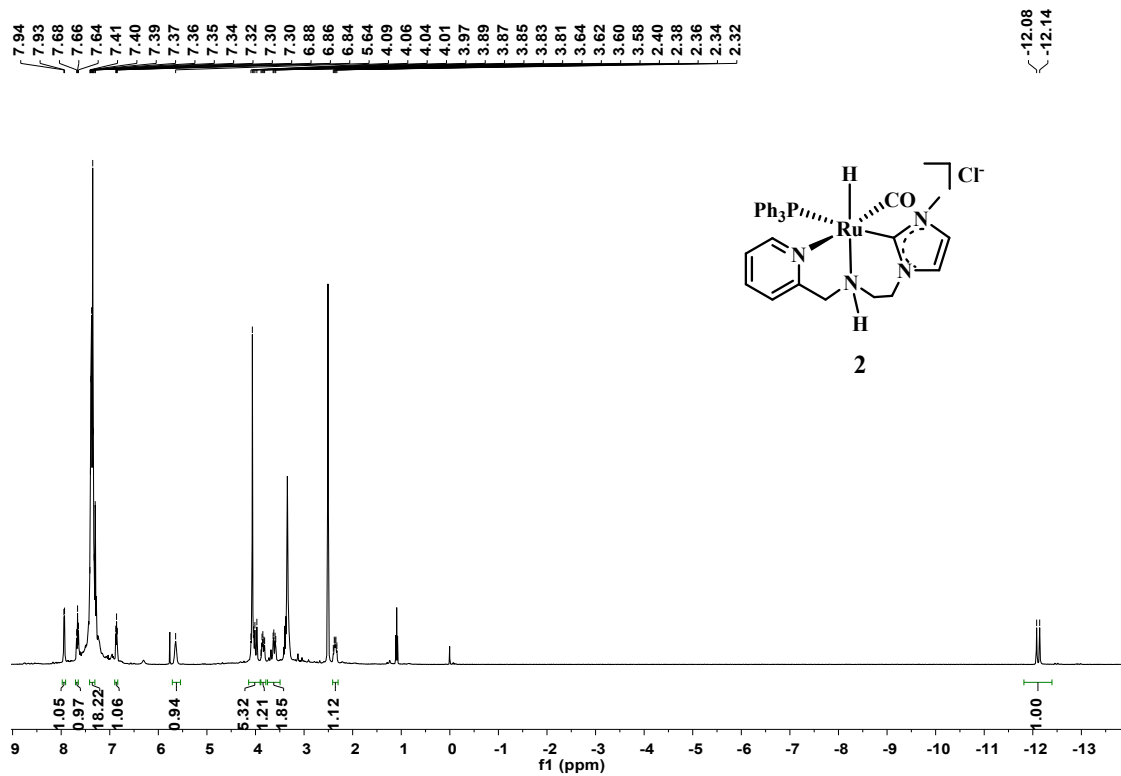


Figure S9. ^1H NMR spectrum of complex **2** (400 MHz, $\text{DMSO-}d_6$).

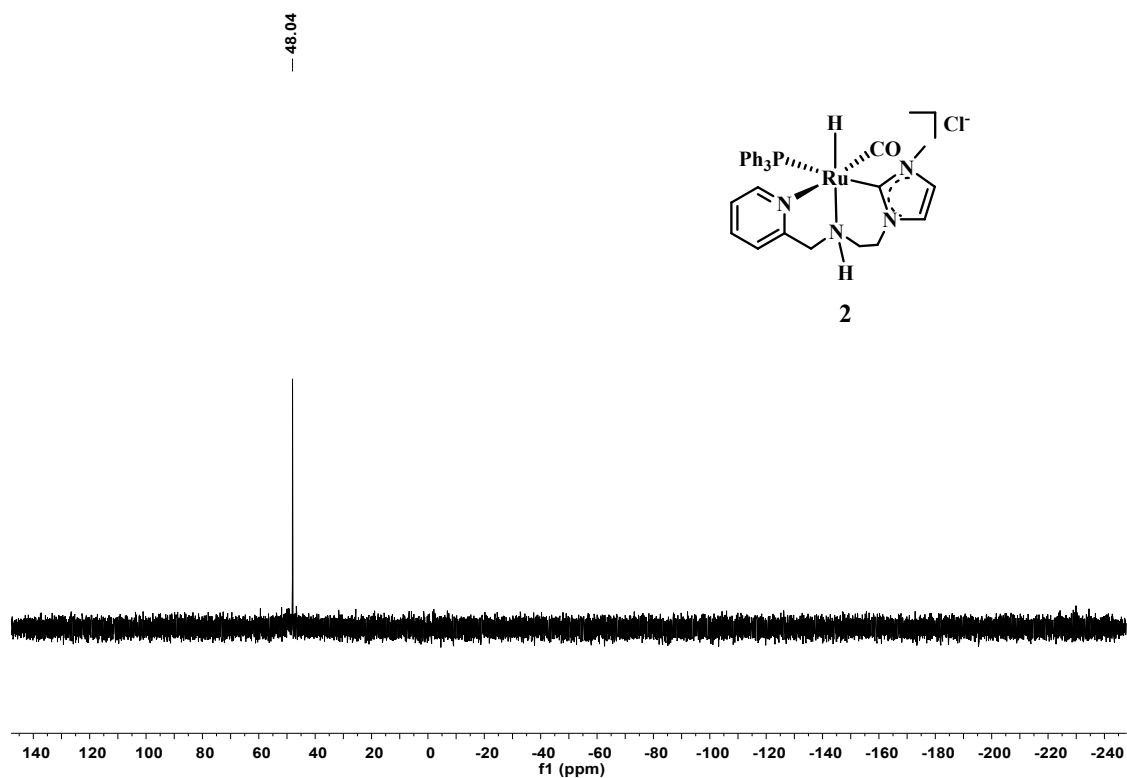


Figure S10. $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum of complex **2** (162 MHz, $\text{DMSO-}d_6$).

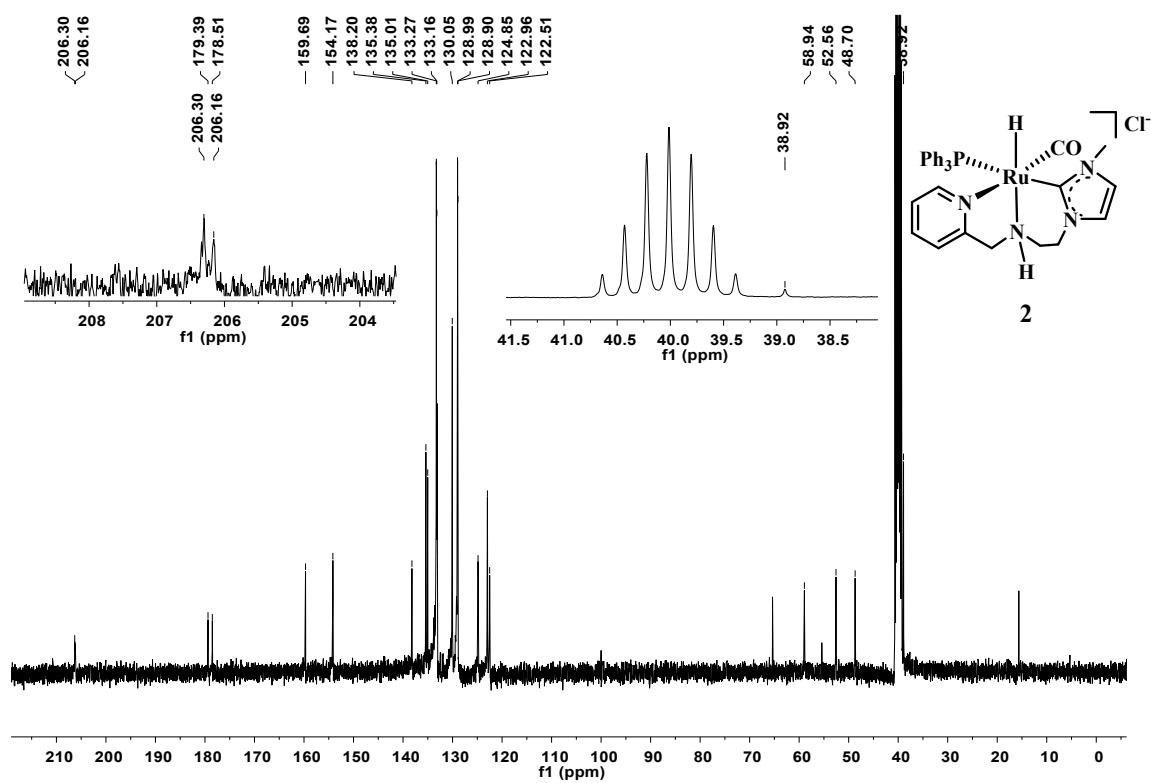


Figure S11. ^{13}C NMR spectrum of complex **2** (100 MHz, $\text{DMSO-}d_6$). Note: the two single peaks at 15.6 and 65.4 ppm are attributed to ether. Single peak at 55.4 ppm is attributed to dichloromethane.

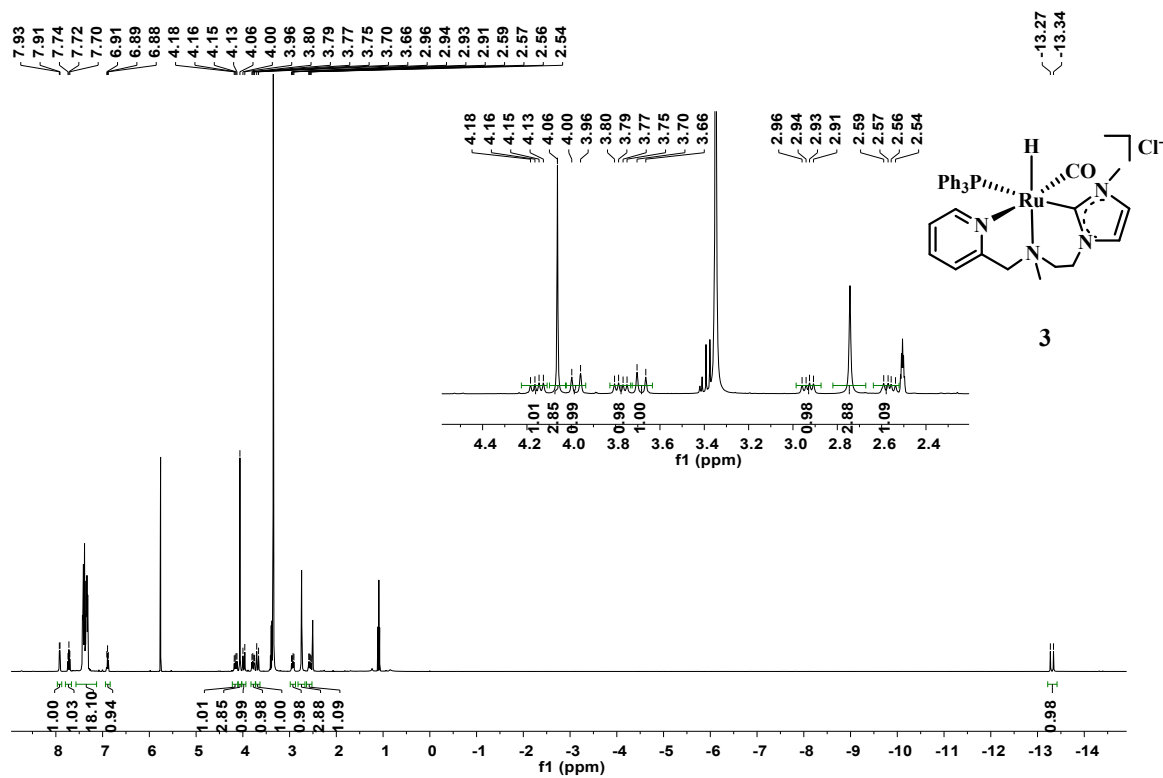


Figure S12. ^1H NMR spectrum of complex 3 (400 MHz, $\text{DMSO-}d_6$).

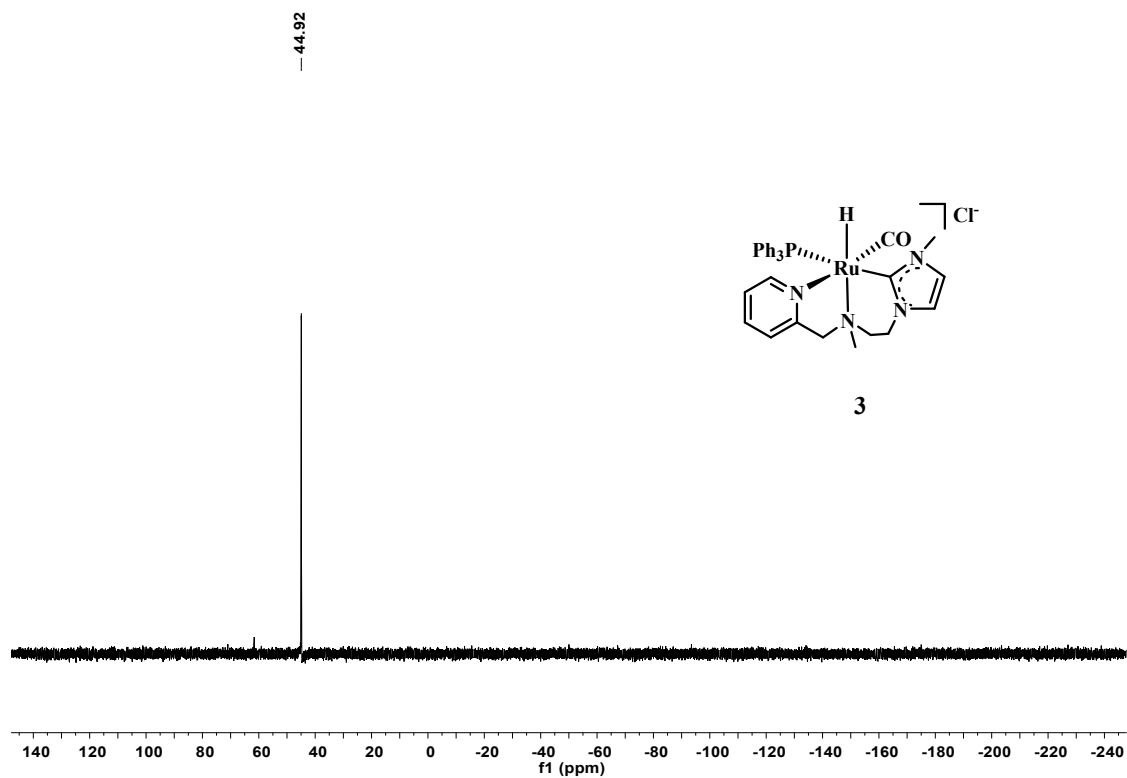


Figure S13. $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum of complex 3 (162 MHz, $\text{DMSO-}d_6$).

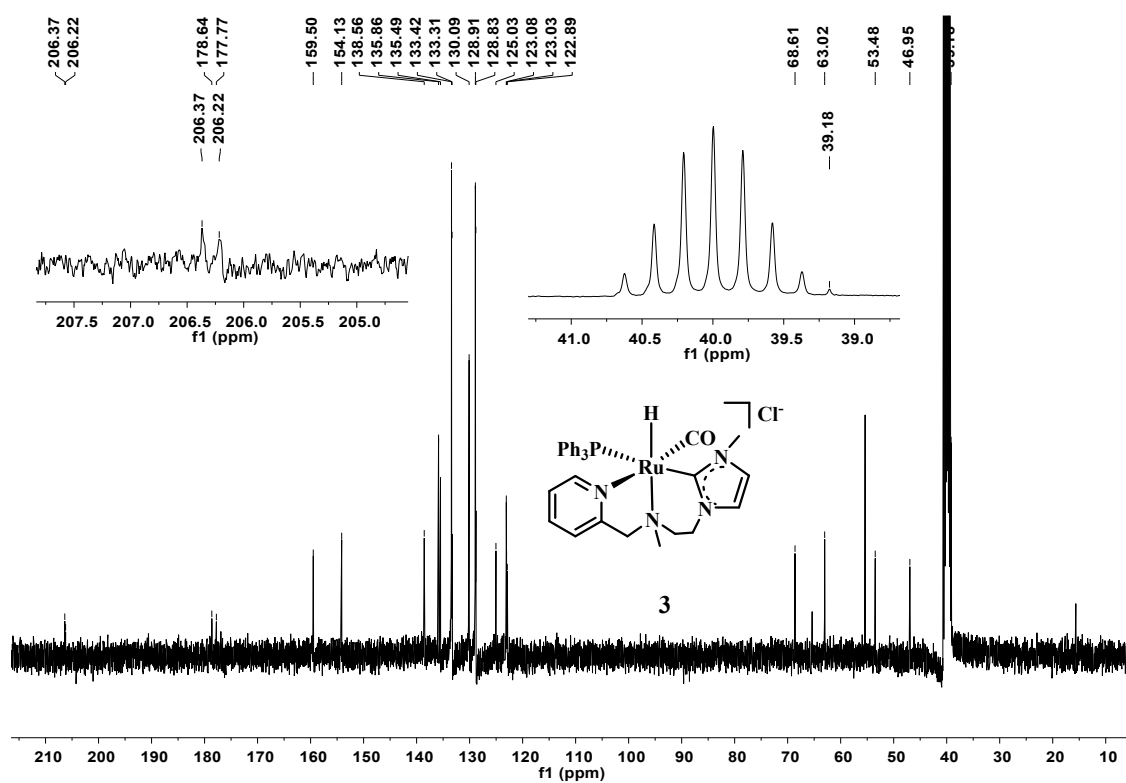


Figure S14. ¹³C NMR spectrum of complex **3** (100 MHz, DMSO-*d*₆). Note: the two single peaks at 15.6 and 65.4 ppm are attributed to ether. Single peak at 55.4 ppm is attributed to dichloromethane.

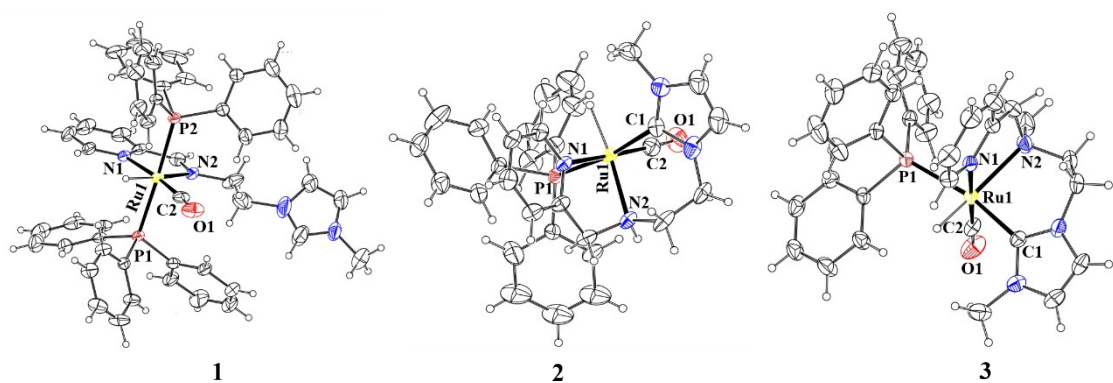


Figure S15. X-ray crystal structures of complexes **1** (BF₄⁻), **2** and **3** (BF₄⁻), the counterion is omitted (These structures are obtained from Reference¹, CCDC 2125582, 2125759 and 2125625 contain the supplementary crystallographic data for complexes **1-3**).

1.2 General procedure of transfer hydrogenation of CO₂.

Complex **1** was slightly soluble in water at room temperature, while complexes **2** and **3** were soluble in water at room temperature. A fresh stock solution of complex **1** or **2** or **3** was prepared prior to the experiment. For example, 40 μmol complex was dissolved in 100 mL degassed H₂O, and diluted to 0.02 μmol/mL with H₂O. The reactions were carried out in a Hastelloy Autoclave Reactor system equipped with a 50 mL cylinder. Under a nitrogen atmosphere, 5.0 mL of a stock solution of complex (0.02 μmol/mL, 0.1 μmol), K₂CO₃ (24 mmol), glycerol or isopropanol (5.0 ml) were added to an autoclave. The autoclave was then sealed, pressurized with CO₂ and stirred at room temperature until the pressure of CO₂ no longer dropped, and the pressurized CO₂ was adjusted to the required pressure (10-50 bar). The mixture was vigorously stirred and heated at 110-200 °C for the desired time (5-100 h). Then, the reaction mixture was cooled down to ambient temperature, and diluted with H₂O. 100-500 μl of DMF (dimethylformamide) was added as internal standard, then, the amount of formate, acetate and lactate were quantified by ¹H NMR spectroscopy in D₂O. The residual hydrogen in gas phase after reaction is analyzed by gas chromatography. The reactions were run three times, and the average numbers of mmol were used.

Representative ¹H NMR of the reaction mixtures

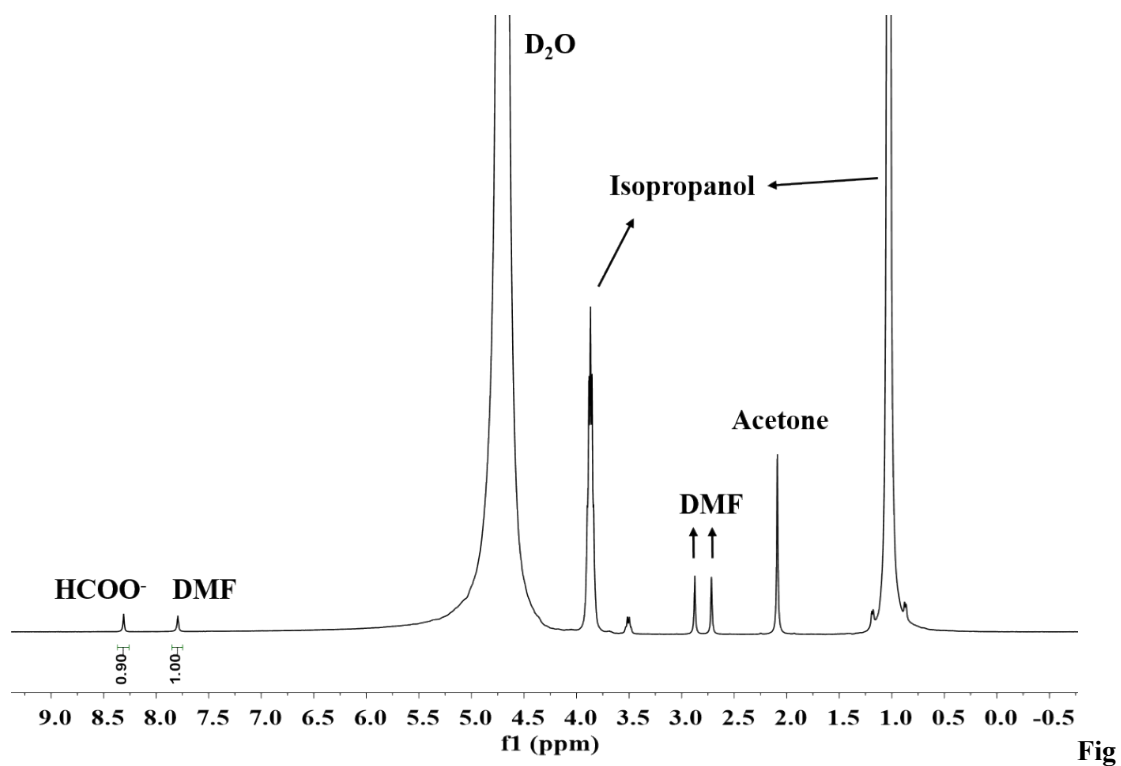


Figure S16. ^1H NMR spectrum of the reaction mixture performed with isopropanol using DMF as internal standard.

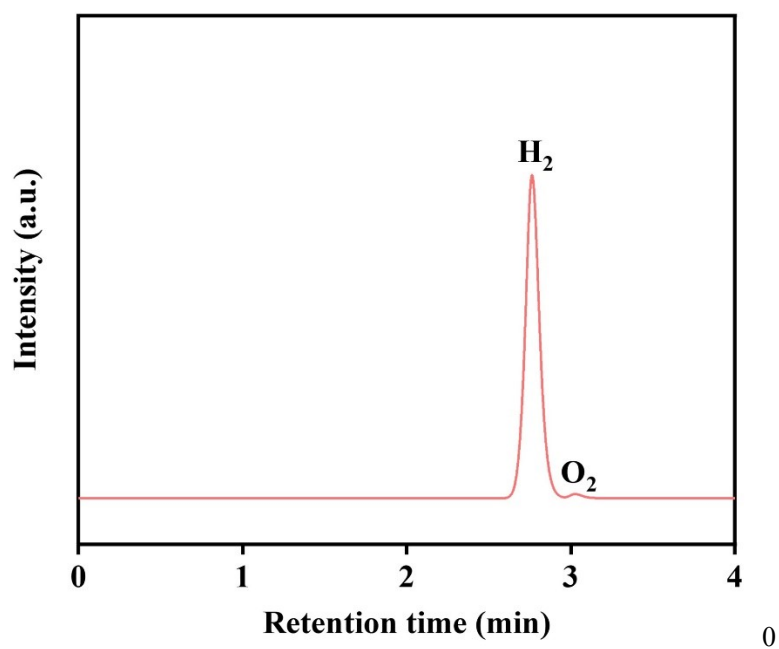


Figure S17. After reaction, hydrogen in gas phase was detected by gas chromatography.

2. Additional catalytic data

Table S1. Transfer hydrogenation of CO₂ to formate with ⁱPrOH.

entry ^a	catalyst	base	HCOO ⁻ (mmol)	TON (HCOO ⁻)	Acetone (mmol)	TON (acetone)
1	1	KOH	0.802	80	1.154	115
2	2	KOH	1.823	182	2.526	253
3	3	KOH	0.388	39	0.37	37
4 ^b	2	KOH	0	0	-	-
5 ^c	2	KOH	0	0	4.252	425
6	2	/	0	0	0.431	43
7	2	NaOH	1.874	187	1.250	125
8	2	K ₂ CO ₃	2.069	207	2.004	200
9	2	KHCO ₃	1.771	177	2.922	292
10	2	Na ₂ CO ₃	0.749	75	0.746	75
11	2	TEA	1.254	125	0.825	83
12	2	TEOA	0.504	50	1.121	112
13	2	DBU	0.685	69	1.312	131
14 ^d	2	K ₂ CO ₃	2.586	259	4.329	433
15 ^e	2	K ₂ CO ₃	3.090	309	4.911	491
16 ^f	2	K ₂ CO ₃	3.103	310	3.710	371
17 ^g	2	K ₂ CO ₃	2.444	244	3.903	390
18 ^h	2	K ₂ CO ₃	1.786	179	2.673	267

^a Reaction conditions: 10 μmol complex, base (24 mmol), H₂O / ⁱPrOH (5 mL/5 mL), 50 bar CO₂, T = 150 °C, t = 20 h. TEA = triethylamine; TEOA = triethanolamine; DBU = 1,8-diazabicyclo [5.4.0] undec-7-ene. ^b without ⁱPrOH. ^c without CO₂. ^d 40 bar CO₂. ^e 20 bar CO₂. ^f 10 bar CO₂. ^g 1 bar CO₂. ^h base 12 mmol. Formate and acetone were quantified by ¹H NMR analysis in D₂O and using DMF as internal standard.

The effect of CO₂ pressure on the hydrogen-transfer reduction of CO₂ was evaluated (Table 2, entries 1-4). Interestingly, lower CO₂ pressure is favorable for CO₂ transfer hydrogenation, and the TON values of formate gradually decrease along with the raising of CO₂ pressure. We proposed two possible reasons for this phenomenon. It was reported that the transfer hydrogenation of CO₂ to formate favors basic media.^{2, 3, 4} More CO₂ will dissolve in the solution due to the higher CO₂ pressure, which reduces the pH of the solution and is unfavorable for the transfer hydrogenation of CO₂.^{3, 4} Moreover, the higher pressure is not beneficial for the formation of hydrogen

thermodynamically, and the suppressed dehydrogenation of *i*PrOH might also lead to the poor CO₂ transfer hydrogenation performance.

KOH, K₂CO₃, and KHCO₃ were all converted into HCO₃⁻ in an aqueous solution under high CO₂ pressure. Since K₂CO₃ yields 2 equivalents of HCO₃⁻, it exhibits the strongest alkalinity when compared with an equal molar amount of KOH and KHCO₃ in the presence of excess CO₂, resulting in higher activity for K₂CO₃ (Table S1, entries 2, 8-9). When the same molar concentration of K⁺ (12 mmol of K₂CO₃) was used, the TON_{formate} in the K₂CO₃ aqueous solution was found to be similar to that in KOH or KHCO₃ (Table S1, entry 18).

Table S2. Effect of temperature on the transfer hydrogenation reaction

entry	T/°C	alcohol	HCOO ⁻		Acetone	
			(mmol)	TON	mmol	TON
1	110	<i>i</i> PrOH	0.634	63	0.352	35
2	130	<i>i</i> PrOH	1.694	169	0.799	80
3	150	<i>i</i> PrOH	3.090	309	4.911	491
4	170	<i>i</i> PrOH	4.060	406	6.25	625
5	200	<i>i</i> PrOH	4.111	411	7.22	722

^aReaction conditions: 10 μmol complex **2**, K₂CO₃ (24 mmol), H₂O / *i*PrOH (5 mL/5 mL), 20 bar CO₂, t = 20 h.

Table S3. Influence of K₂CO₃ concentration on the activity of CO₂ transfer hydrogenation

entry	alcohol	K ₂ CO ₃ (mmol)	HCOOK (mmol)	TON (formate)	lactate	TON
					TON	
1	<i>i</i> PrOH	6	0.607	607	-	
2	<i>i</i> PrOH	12	1.073	1070	-	
3	<i>i</i> PrOH	24	4.034	4030	-	
4	Glycerol	6	1.177	1180	2663	185
5	Glycerol	12	1.616	1620	2728	193
6	Glycerol	24	3.391	3390	6460	228

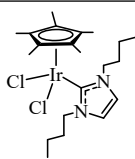
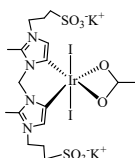
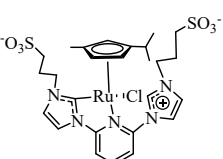
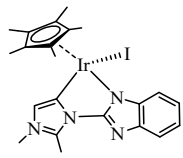
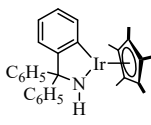
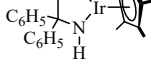
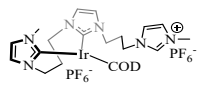
Reaction conditions: 1 μmol complex **2**, K₂CO₃ (6-24 mmol), H₂O / *i*PrOH or Glycerol (5 mL/5 mL), 20 bar CO₂, T = 170 °C, t = 20 h.

Table S4. Effect of catalyst amounts on catalytic CO₂ transfer hydrogenation with ⁱPrOH

entry	Cat./ μmol	T/°C	Time (h)	HCOOK (mmol)	TON	ⁱ PrOH conv. %	Acetone	
							mmol	TON
1	10	170	20	4.060	406	22.5	6.25	625
2	5	170	20	3.930	786	21.7	4.71	943
3	1	170	20	4.034	4030	36.3	2.20	2200
4	0.5	170	20	3.775	7550	48.2	1.51	3017
5	0.1	170	20	1.162	11600	13.3	0.41	4100
6	0.1	170	50	1.526	15300	39.0	0.59	5900
7	0.1	170	100	2.590	25900	54.5	0.73	7327

Reaction conditions: complex **2**, K₂CO₃ (24 mmol), H₂O / ⁱPrOH (5 mL/5 mL), 20 bar CO₂.

Table S5. Previously reported catalysts for the transfer hydrogenation of CO₂ and inorganic carbonates

Cat.	Cl Source	hydrogen donor	T (°C)	Time (h)	TON (formate)	TON (lactate)	Ref.
	CO ₂ (50 bar)	ⁱ PrOH	110	72	150	-	[5]
	CO ₂ (50 bar)	ⁱ PrOH	200	75	2700	-	[6]
	CO ₂ (26 bar)	glycerol	180	24	1065	1685	[7]
	CO ₂ (1 bar)	glycerol	150	12	1080	-	[8]
	CsHCO ₃	ⁱ PrOH	80	5	3200	-	[9]
	CO ₂ (1 bar)	ⁱ PrOH	80	5	655	-	
	CO ₂ (5 bar)	glycerol	200	20	200 000	875 000	[10]

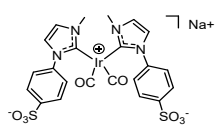
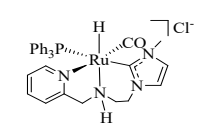
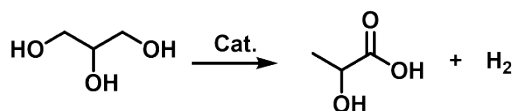
	K ₂ CO ₃	glycerol	150	6	13350	27850	[11]
	CO ₂ (20 bar)	glycerol	200	50	300000	387000	this work

Table S6. Independent glycerol dehydrogenation or CO₂ hydrogenation reactions.

entry	Cat. /μmol	T/°C	Time (h)	TON (FA)	TON (LA)	TON (AA)	P(H ₂) /MPa
1 ^a	0.1	200	20	15400	164000	11250	
2 ^b	0.1	150	5	1720			1.0
3 ^b	0.1	150	5	2840			2.0
4 ^b	0.1	150	5	4030			3.0
5 ^b	0.1	150	5	trace			0.1
6 ^b	0.1	200	20	13100			1.0
7 ^c	0.1	200	20	118000	176000	8620	

^a Independent glycerol dehydrogenation without CO₂. ^b Independent CO₂ hydrogenation without glycerol. ^c Co-upcycling of glycerol and CO₂. FA: formate; LA: lactate; AA: acetate.



Eq. 1

$$pV = nRT$$

Eq. 2

The Ru complex **2** can realize the dehydrogenation of glycerol into lactate and H₂ in the absence of CO₂ (Table S6, entry 1). Meanwhile, when glycerol was replaced by H₂, Ru complex **2** was also able to achieve the hydrogenation of CO₂ into formate (Table S6, entries 2-6), with the reaction rate increasing as H₂ pressure increases. Therefore, these two reactions can occur independently.

However, we also found that CO₂ hydrogenation was difficult under low H₂ pressure (Table S6, entries 5-6). The theoretical H₂ yield (Eq. 1; Table S6, entry 1) is 0.0164 mol, corresponding to an H₂ partial pressure of 0.81 MPa (Eq. 2) in our system. To verify this, we measured the pressure of H₂ after the reaction and found that the partial pressure of H₂ was less than 1 MPa, which is disadvantageous for the hydrogenation of CO₂. Therefore, we assume that the active Ru-H intermediate formed during the dehydrogenation of glycerol could directly realize the hydrogenation of CO₂ to formate, without the need to generate free H₂ in advance (Figure S19, Table S6, entry 7). In this case, the dehydrogenation of glycerol would influence the subsequent CO₂ hydrogenation. However, it almost impossible to distinguish between the contributions of the Ru-H pathway and

the free H₂ pathway, as the hydrogenation of CO₂ with H₂ could also form the Ru-H intermediate (Figure S19).

Therefore, we prefer not to mention whether glycerol dehydrogenation and CO₂ hydrogenation are independent processes. Instead, we only emphasize that the Ru complex **2** can serve as the bifunctional catalysts for converting CO₂ and glycerol into formate and lactate simultaneously.

Computational methods of DFT

Gaussian 09¹² calculations were performed with the b3lyp density functional using a def2-SVP basis^{13,14}. The Gibbs free energies, were calculated at T = 473.15 K and 1 atm pressure by Shermo¹⁵. The solvation effect of H₂O was included by performing single-point energy calculations using the SMD solvation model.

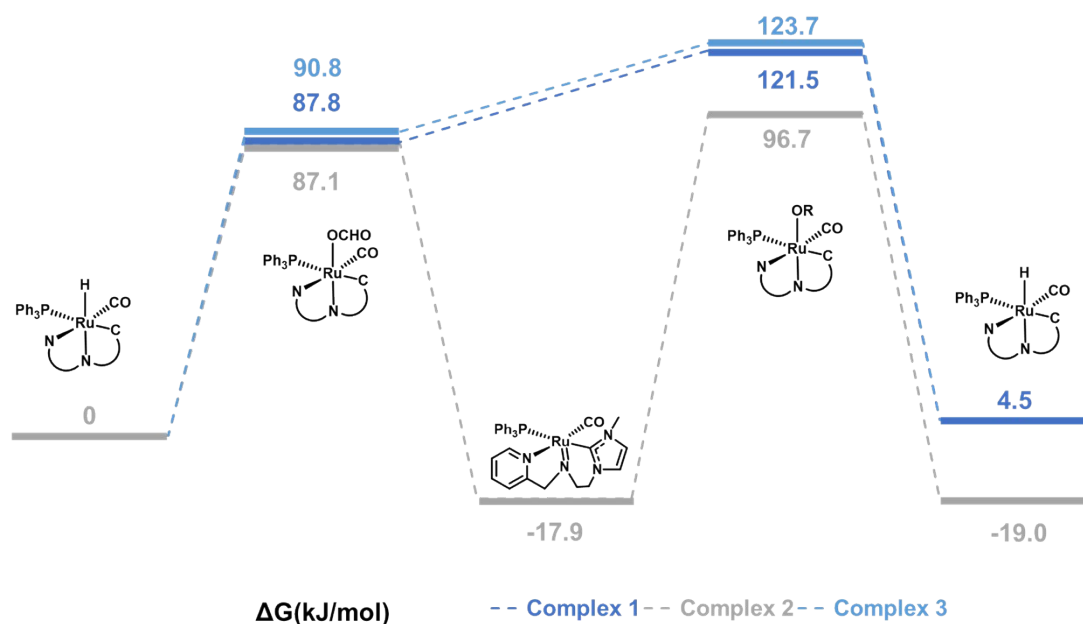
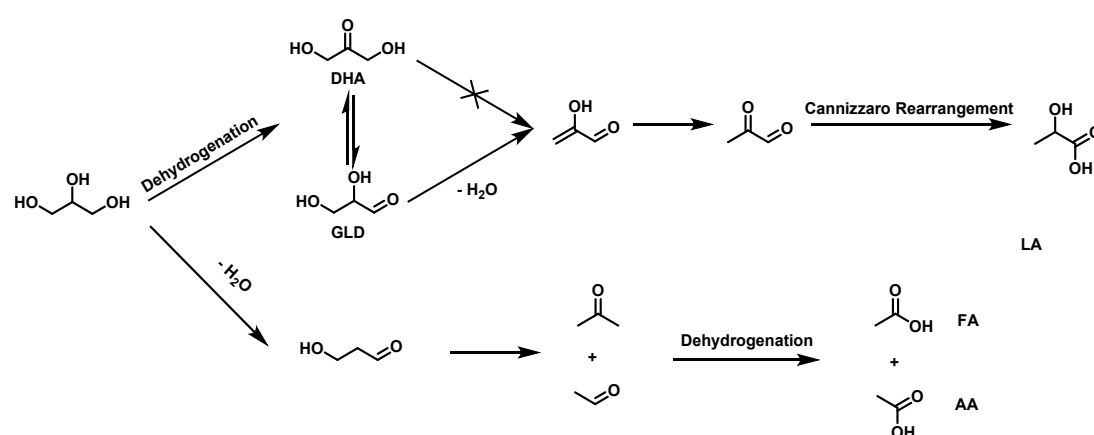


Figure S18. Gibbs free energy profiles at reaction temperature of complex 1-3.



Scheme S3. Pathway for glycerol conversion¹⁶⁻¹⁹

As shown in Scheme N3 and further supported by isotope labeling experiments (Figure 2B), it is suggested that the conversion of glycerol is quite complex, and a small portion of the formate should

also originate from the C-C bond cleavages of glycerol rather than from CO₂ remaining in the water.

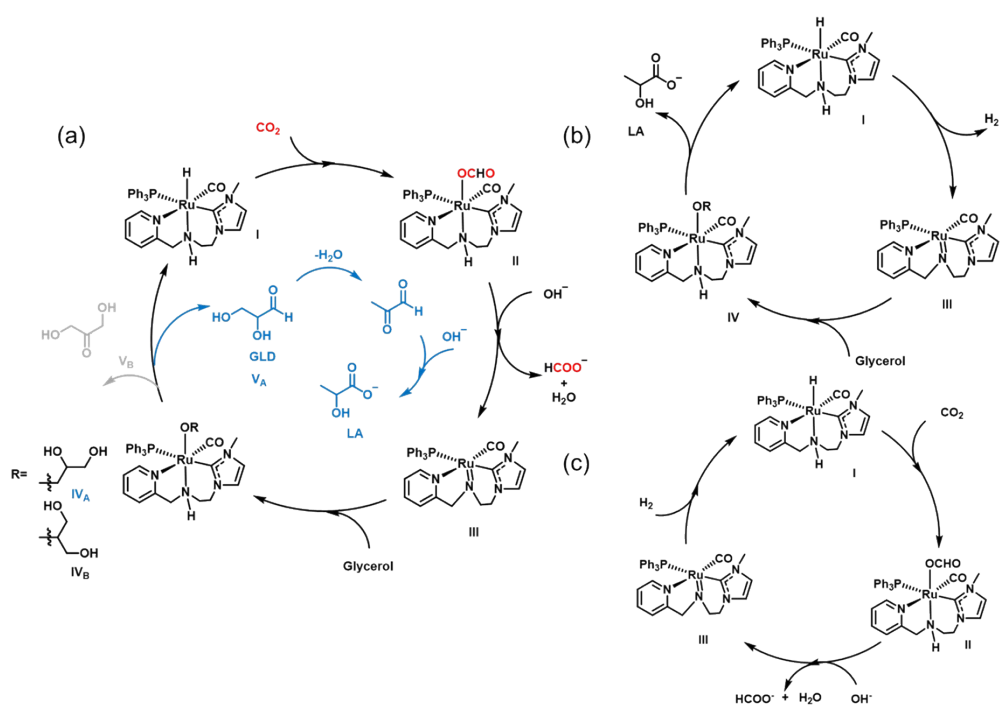


Figure S19. (a) Proposed catalytic cycle for the co-upcycling of CO₂ and glycerol. (b) Proposed catalytic cycle for independent glycerol dehydrogenation and (c) independent CO₂ hydrogenation.

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