

SUPPORTING INFORMATION

Scalable organocatalytic one-pot asymmetric Strecker reaction *via* camphor sulfonyl functionalized crown-ether-tethered calix[4]arene

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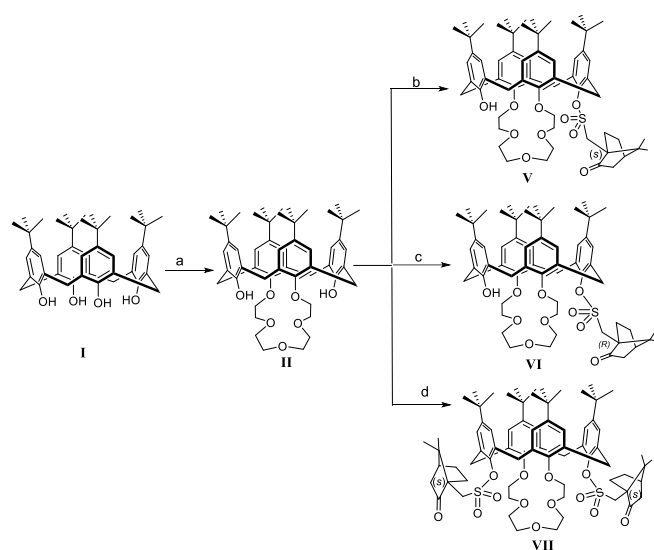
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1. General Information

Starting materials and reagents were used without purification unless otherwise mentioned and were purchased from commercial suppliers. Reaction temperatures were measured externally; reactions were monitored by analytical thin layer chromatography (TLC), Merk 60 F₂₅₄ Aluminium coated plates and visualized by UV light. Column chromatography was performed on silica gel (60-120 mesh) and the solvents employed were of analytical grade purified before Use. The enantiomeric excess (ee) of the cyanated products was determined by HPLC measurements carried out using a thermoscientific Vanquish detector Model: VC-D11-A equipped with chiral columns DAICEL 4.6 mm × 250 mm Chiralpak- AD-H using condition n-Hexane/isopropanol solvent system. Optical rotation values were measured on a JASCO P-1010 polarimeter at $\lambda = 589$ nm, corresponding to the sodium D line, at the temperatures indicated. NMR spectra were recorded on a Bruker 500 spectrophotometer ¹H NMR, ¹³C and ²⁹Si spectra were recorded on 500, 126 and 500 MHz respectively. Chemical shifts (δ) are reported in ppm relative to the residual solvent peak (¹H CDCl₃, $\delta = 7.26$; ¹³C CDCl₃, $\delta = 77.0$) and the following abbreviations designate the multiplicity of each signal: s, singlet; d, doublet; t, triplet; m, multiplet; bs, broad singlet; bd, broad doublet. Coupling constants (J) are quoted in Hertz. High-resolution mass spectra (HRMS) were acquired using an Agilent technologies model G6564 QTOF. The samples were ionized in positive ion mode using a MALDI or ESI ionization source. Calix[4] arene was synthesised via reported procedure ¹

2. Synthesis of catalyst



Reaction conditions: (a) dry toluene, p-tert-butylcalix[4]arene (1.46 g, 2.25 mmol), ^tBuOK (0.46 g, 4 mmol), reflux, tetraethylene glycol ditosylate (1.06g, 2 mmol); (b) II, Dry THF/DMF (16:1), NaH (0.06g, 1.62 mmol), (+)-10-Camphorsulfonyl chloride (0.4g, 1.65 mmol). (c) II, Dry THF/DMF (16:1), NaH (0.06g, 1.62 mmol), (-)-10-Camphorsulfonyl chloride (0.4g, 1.65 mmol) (d) II, Dry THF/DMF (16:1), NaH (0.14g, 3.24 mmol), (+)-10-Camphorsulfonyl chloride (0.8g, 3.3 mmol)

Step a: Synthesis of p-tert-butylcalix[4]arene-crown-5 (II) :

A mixture of p-tert-butylcalix[4]arene (I, 1.46 g, 2.25 mmol) and ^tBuOK (0.23 g, 2 mmol) in 50 mL of dry toluene was refluxed (using heating blocks) under nitrogen gas with stirring for 1.5 h, then tetraethylene glycol ditosylate (1.06 g, 2 mmol) was added. After refluxing for 24

h, a second portion of ^tBuOK (0.23 g, 2 mmol) was added, and the reaction mixture was refluxed for an additional 24 h. It was then cooled to room temperature, treated with 1 N HCl (125 mL), and extracted five or six times with 30 mL of diethyl ether each time. The combined organic layers were finally washed three times with water (100 mL each time), and the solvent was removed by rotary evaporation. The crude product was purified by column chromatography using silica as packing material and dichloromethane/ethyl acetate (4:1) as eluent gives white solid, 1.17 g, 65% yield of I, ¹H NMR (500 MHz, CDCl₃) δ 7.07 (s, 4H), 6.75 (s, 4H), 4.38 (d, *J* = 13.0 Hz, 4H), 4.08 (s, 8H), 3.97 (t, *J* = 5.5 Hz, 4H), 3.85 (t, *J* = 5.5 Hz, 4H), 3.30 (d, *J* = 13.0 Hz, 4H), 1.31 (s, 20H), 0.91 (s, 16H); ¹³C NMR (126 MHz, Chloroform-*d*): 150.8, 149.8, 146.8, 141.2, 132.5, 129.8, 127.8, 125.4, 124.9, 71.0, 70.8, 70.3, 33.8, 31.7, 31.0

Step b: Synthesis of calix[4]arene-crown-5 tethered with (+)-10-camphorsulfonyl catalyst (V):

To a solution of calix[4]crown-5 II (1.2g, 1.5 mmol) in DryTHF/DMF mixture (16/1), a 60% suspension of sodium hydride in mineral oil was added (0.06g, 1.62 mmol). After stirring the reaction mixture at room temperature for 2 hours, (+)-10-camphor sulfonyl chloride (0.40g, 1.65 mmol; mp 69°C ([α]²⁸_D = +36° C2, CHCl₃)) in THF (2.5 mL) was added. The reaction mixture was stirred at ambient temperature for 18h. The solvent was removed under vacuum and the oily residue was dissolved into chloroform and washed with saturated NH₄Cl solution. Then the organic layer was dried over Na₂SO₄. The residue was further purified through column chromatography CH₂Cl₂/MeOH = 95/5 to 60/40 as eluent to afford **V** as a white crystalline solid (1.14 g, 76%); [α]²⁸_D = +52° (c 1, CHCl₃); ¹H NMR (500 MHz, Chloroform-*d*) δ 7.19 (s, 1H), 7.07 (s, 4H), 6.75 (s, 4H), 4.60 (m, 1H), 4.37 (d, *J* = 13.0 Hz, 4H), 4.08 (s, 8H), 3.97 (t, *J* = 5.3 Hz, 4H), 3.84 (t, *J* = 5.5 Hz, 4H), 3.29 (d, *J* = 13.0 Hz, 4H), 3.18 (s, 1H), 2.06 – 1.94 (m, 3H), 1.31 (s, 18H), 1.19 (s, 3H), 1.11 (s, 3H), 1.04 (d, *J* = 7.4 Hz, 6H), 0.91 (s, 18H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 214.73, 214.17, 150.82, 149.82, 146.85, 141.23, 135.12, 134.93, 134.83, 134.30, 134.24, 132.51, 127.85, 125.56, 125.35, 125.01, 124.96, 71.26, 71.07, 70.90, 70.69, 70.37, 70.06, 58.66, 58.39, 47.81, 47.70, 43.37, 43.11, 42.49, 33.89, 33.85, 31.68, 31.34, 31.04, 30.97, 20.35, 20.17, 20.07, 19.96, 19.78. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₆₂H₈₄O₁₀S, 1020.5800; found 1020.5805

Step c: Synthesis of calix[4]arene-crown-5 tethered with (-)-10-camphor sulfonyl catalyst(VI):

The synthetic procedure is the same as catalyst **V** using (-)-10-camphor sulfonyl chloride; mp 67°C ([α]²⁸_D = -34° C2, CHCl₃). The residue was purified through column chromatography CH₂Cl₂/MeOH = 95/5 to 60/40 as eluent to afford **VI** as a white crystalline solid (1.11 g, 74%); [α]²⁸_D = -50° (c 1, CHCl₃); ¹H NMR (500 MHz, Chloroform-*d*) δ 7.29 (s, 1H), 7.07 (s, 4H), 6.75 (s, 4H), 4.60 (m, 1H), 4.37 (d, *J* = 13.0 Hz, 4H), 4.07 (s, 8H), 3.96 (t, *J* = 5.4 Hz, 4H), 3.84 (d, *J* = 5.4 Hz, 4H), 3.29 (d, *J* = 13.1 Hz, 4H), 3.18 (m, 1H), 2.00 (m, 1H), 1.31 (s, 20H), 1.11 (s, 4H), 1.04 (s, 4H), 1.03 (s, 4H), 0.91 (s, 16H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 213.73, 213.17, 149.82, 148.82, 145.85, 140.23, 131.511, 126.85, 124.56, 124.35, 124.01, 123.96, 57.66, 57.39,

46.81, 46.70, 46.26, 42.37, 41.49, 32.89, 32.86, 30.88, 30.68, 30.35, 29.97, 18.96, 18.87 HRMS (ESI+) m/z $[M + H]^+$ calcd for $C_{62}H_{84}O_{10}S$, 1020.5800; found 1020.5803

Step d: Synthesis of calix[4]arene-crown-5 tethered with (+)-10-camphorsulfonyl catalyst (VII):

To a solution of calix[4]crown-5 II (1.2g, 1.5 mmol) in DryTHF/DMF mixture (16/1), a 60% suspension of sodium hydride in mineral oil was added (0.14g, 6.0 mmol). After stirring the reaction mixture at room temperature for 2 hours, D-(+)-10-camphor sulfonyl chloride (1.50g, 8.0 mmol mp 69°C ($[\alpha]^{28}_D = +36^\circ$ C2, $CHCl_3$)) in THF (2.5 mL) was added. The reaction mixture was stirred at ambient temperature for 5h followed by heating at 70 °C for 12h. Reaction was monitored via TLC. After completion of reaction, reaction mixture was cooled to room temperature and solvent was removed under vacuum and the oily residue was dissolved into chloroform and washed with saturated NH_4Cl solution. Then the organic layer was dried over Na_2SO_4 . The residue was further purified through column chromatography $CH_2Cl_2/MeOH = 95/5$ to 60/40 as eluent to afford VII as a white crystalline solid (1.47 g, 80%); $[\alpha]^{28}_D = +78^\circ$ (c 1, $CHCl_3$); 1H NMR (600 MHz, Chloroform-*d*) δ 7.07 (s, 4H), 6.75 (s, 4H), 4.38 (d, $J = 13.0$ Hz, 4H), 4.37-4.25 (m, 2H), 4.10 – 4.06 (m, 4H), 3.96 (d, $J = 5.4$ Hz, 4H), 3.85 (d, $J = 5.4$ Hz, 4H), 3.30 (d, $J = 13.1$ Hz, 4H), 2.83 (s, 2H), 2.17 – 1.90 (m, 2H), 1.78 (s, 4H), 1.34 (s, 2H), 1.31 (s, 20H), 1.21 (s, 4H), 0.94 (s, 4H), 0.91 (s, 16H). ^{13}C NMR (126 MHz, Chloroform-*d*) δ 214.41, 214.15, 150.82, 149.81, 146.85, 141.22, 132.51, 127.84, 125.44, 124.97, 71.07, 70.90, 70.37, 43.26, 42.82, 42.52, 42.4, 37.12, 34.15, 33.92, 33.89, 33.85, 33.82, 33.06, 31.76, 31.50, 31.35, 31.27, 31.04, 30.99, 29.72, 26.90, 24.93, 22.72, 20.25, 20.25, 19.86, 19.71, 14.14. HRMS (ESI+) m/z $[M + H]^+$ calcd for $C_{72}H_{98}O_{13}S_2$ 1234.6400; found 1234.6406

3. General procedure for one-pot three-component Strecker reaction.

In an inert environment, benzaldehyde (84 mg, 0.8 mmol), aniline (74 mg, 0.8 mmol), trimethylsilyl cyanide (79 mg, 0.8 mmol), and catalyst (2 mol%) were introduced in a dry glass tube with dichloromethane (0.4ml) as a solvent. The reaction mixture was stirred continuously for 15 minutes and monitored through TLC. The reaction mixture was treated with chloroform and washed with a saturated solution of sodium bisulphite. The organic phase was dried over anhydrous sodium sulphate. The solvent was then evaporated under reduced pressure and the crude was purified through column chromatography (hexane/Ethyl acetate 80/20). The enantiomeric excess was determined by chiral stationary-phase HPLC analysis.

Table S1-Effects of the ratio of aldehyde/amine to TMSCN on enantioselective Strecker reaction*

Entry ^[a]	Ratio of benzaldehyde/aniline /TMSCN	Time[<i>min</i>]	Yield ^[b]	ee ^[c]
1	1/1/1	15	99.9	99.2
2	1/0.9/1.2	15	99.9	99.1
3	1/0.9/1.5	30	99.9	98.9

*Reaction Conditions: 1 equiv of catalyst **V**, benzaldehyde, aniline and TMSCN in DCM at 25°C ^[b] Isolated yield.^[c]Determined by HPLC analysis on Daicel chiralpak AD-H

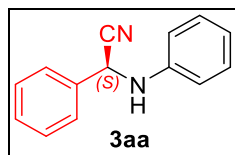
Table S2- Limitation Table for Substrate scope using catalyst **V***

Entry ^[a]	aldehyde	Time[min]	Yield ^[b]	$[\alpha]^{28}_D$ ^[c]
1	Propionaldehyde	15	90	+51 (C 0.5, MeOH)
2	butyraldehyde	15	94	+23 (C 0.5, CHCl ₃)
3	Hexanal	15	97	+46 (C 0.5, CHCl ₃)

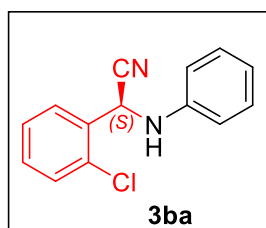
*Reaction Conditions: 1 equiv of catalyst **V**, aldehyde, aniline and TMSCN in DCM at 25°C ^[b] Isolated yield.

^[c] Optical rotation values were measured on a JASCO P-1010 polarimeter at $\lambda = 589$ nm.

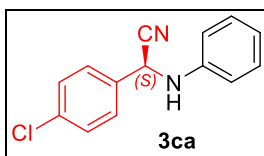
4. Characterisation of Strecker products



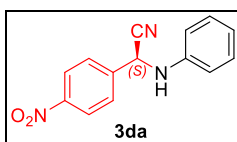
(S)-2-phenyl-2-(phenylamino)acetonitrile(3aa). Colourless solid; 164.3 mg, 99.9% yield; 99.2% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) =12.5min, tR (minor) = 18.9 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.61 (dd, $J = 7.5, 1.9$ Hz, 2H), 7.47 (d, $J = 7.4$ Hz, 3H), 7.31 – 7.27 (m, 2H), 7.17 (dd, $J = 8.5, 7.4$ Hz, 1H), 6.96 – 6.88 (m, 1H), 6.82 – 6.77 (m, 2H), 6.73 – 6.69 (m, 1H), 5.44 (s, 1H), 4.00 (s, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 143.62, 132.87, 128.54, 128.51, 128.31, 126.23, 119.24, 113.09, 49.16. HRMS (ESI+) m/z $[M + H]^+$ calcd for C₁₄H₁₂N₂, 208.1000 ; found 208.1009.



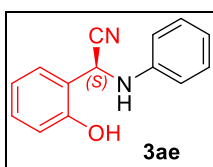
(S)-2-(2-chlorophenyl)-2-(phenylamino)acetonitrile (3ba). Colourless solid; 171.8 mg, 99.9% yield; 89.6% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 4.1 min, tR (minor) = 7.1 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.41 – 7.35 (m, 5H), 7.31 – 7.28 (m, 4H), 5.75 (s, 1H), 4.07 (s, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 144.39, 135.64, 132.43, 129.65, 128.63, 126.25, 120.60, 117.86, 114.33, 49.69. HRMS (ESI+) m/z $[M + H]^+$ calcd for C₁₄H₁₁ClN₂, 242.0600; found 242.0602.



(S)-2-(4-chlorophenyl)-2-(phenylamino)acetonitrile (3ca). Colourless solid; 151.4 mg, 92% yield; 88.4% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 4.2 min, tR (minor) = 4.4 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.48 – 7.39 (m, 5H), 7.29 (d, *J* = 7.9 Hz, 1H), 7.24 – 7.20 (m, 2H), 6.93 (t, *J* = 7.4 Hz, 1H), 5.42 (s, 1H), 4.10 (s, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 157.92, 150.67, 143.37, 136.41, 133.69, 131.42, 128.98, 128.65, 128.57, 128.23, 128.11, 127.62, 125.24, 119.87, 119.62, 116.82, 113.32, 48.68, 28.73 HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₁₄H₁₁ClN₂, 242.0600; found 242.0603

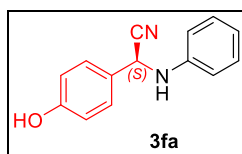


(S)-2-(4-nitrophenyl)-2-(phenylamino)acetonitrile(3da). Yellow solid; 207.3 mg, 99.9% yield; 47.0% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 26.3 min, tR (minor) = 39.3 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 8.30 (d, *J* = 8.7 Hz, 2H), 8.09 – 8.06 (m, 1H), 7.82 (d, *J* = 8.4 Hz, 2H), 7.27 (dd, *J* = 13.4, 6.9 Hz, 3H), 6.94 (t, *J* = 7.4 Hz, 1H), 6.76 (d, *J* = 7.9 Hz, 2H), 5.64 – 5.48 (m, 1H), 4.22 (s, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 141.59, 138.75, 129.73, 129.51, 129.43, 129.36, 129.15, 128.68, 128.24, 127.11, 124.49, 124.21, 124.04, 120.98, 120.79, 114.62, 49.76. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₁₄H₁₁N₃O₂, 253.0900; found 253.0907.

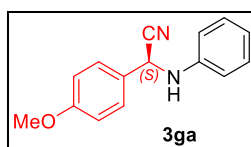


(S)-2-(2-hydroxyphenyl)-2-(phenylamino)acetonitrile (3ea). Colourless solid; 157.9 mg, 92% yield; 89.1% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 8.9 min, tR (minor) = 18.8 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 8.63 (s, 1H), 7.56 (dd, *J* = 7.8, 1.6 Hz, 1H), 7.45 – 7.38 (m, 3H), 7.31 – 7.27 (m, 3H), 7.03 (dd, *J* = 7.7, 3.6 Hz, 3H), 6.92 (d, *J* = 8.0 Hz, 2H), 5.63 (d, *J* = 9.1 Hz, 1H), 4.23 (d, *J* = 9.6 Hz, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 161.66, 153.76,

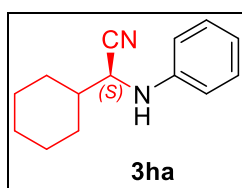
132.15, 131.26, 128.71, 128.40, 127.13, 125.89, 121.23, 120.15, 118.07, 116.30, 115.41, 47.61. HRMS (ESI+) m/z $[M + H]^+$ calcd for $C_{14}H_{12}N_2O$, 224.0900; found 224.0903



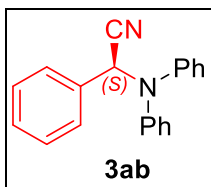
(S)-2-(4-hydroxyphenyl)-2-(phenylamino)acetonitrile (3fa). Colourless solid; 171.3 mg, 95% yield; 95.5% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 26.9 min, tR (minor) = 33.3 min; 1H NMR (500 MHz, Chloroform-*d*) δ 7.46 (d, J = 8.5 Hz, 2H), 7.31 – 7.27 (m, 2H), 6.91 (t, J = 8.1 Hz, 3H), 6.78 (dt, J = 7.7, 3.4 Hz, 2H), 5.36 (d, J = 7.5 Hz, 1H), 4.01 (d, J = 8.5 Hz, 1H). ^{13}C NMR (126 MHz, Chloroform-*d*) δ 155.65, 143.66, 128.56, 127.85, 124.99, 119.22, 117.38, 115.13, 113.08, 48.66. HRMS (ESI+) m/z $[M + H]^+$ calcd for $C_{14}H_{12}N_2O$, 224.0900; found 224.0905



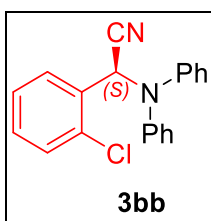
(S)-2-(4-methoxyphenyl)-2-(phenylamino)acetonitrile (3ga). Colourless solid; 151.5 mg, 97% yield; 97.9% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 11.8 min, tR (minor) = 12.5 min; 1H NMR (500 MHz, Chloroform-*d*) δ 7.51 (d, J = 8.7 Hz, 2H), 7.28 (dd, J = 8.5, 7.3 Hz, 2H), 6.97 (d, J = 8.7 Hz, 2H), 6.90 (t, J = 7.4 Hz, 1H), 6.80 – 6.75 (m, 2H), 5.37 (s, 1H), 3.84 (s, 3H). ^{13}C NMR (126 MHz, Chloroform-*d*) δ 159.38, 143.69, 128.52, 127.60, 124.88, 119.13, 117.39, 113.59, 113.04, 54.39, 48.60. HRMS (ESI+) m/z $[M + H]^+$ calcd for $C_{15}H_{14}N_2O$, 238.1100; found 238.1107.



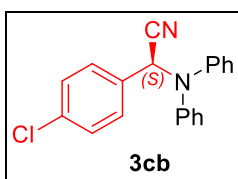
(S)-2-cyclohexyl-2-(phenylamino)acetonitrile (3ha). Colourless solid; 141.1 mg, 99.9% yield; 98.5% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 22.9 min, tR (minor) = 25.6 min; 1H NMR (500 MHz, Chloroform-*d*) δ 7.29 – 7.27 (m, 2H), 7.20 – 7.17 (m, 1H), 6.90 – 6.87 (m, 1H), 6.73 (ddt, J = 11.0, 8.5, 1.1 Hz, 3H), 3.88 – 3.79 (m, 1H), 1.36 – 1.21 (m, 12H). ^{13}C NMR (126 MHz, Chloroform-*d*) δ 145.19, 129.55, 119.89, 118.86, 114.06, 51.75, 40.80, 29.69, 28.94, 25.94, 25.66, 25.59. HRMS (ESI+) m/z $[M + H]^+$ calcd for $C_{14}H_{18}N_2$, 214.1500; found 214.1504



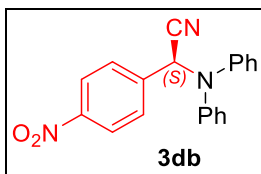
(S)-2-(diphenylamino)-2-phenylacetonitrile (3ab). Colourless solid; 217.1 mg, 98% yield; 93.1% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 18.8 min, tR (minor) = 34.2 min; ¹H NMR (500 MHz, CDCl₃) δ 7.53 – 7.50 (m, 2H), 7.43 (s, 3H), 7.07 (d, *J* = 7.9 Hz, 9H), 6.93 (d, *J* = 7.4 Hz, 5H), 5.50 (s, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 143.19, 135.38, 129.41, 126.81, 126.63, 121.12, 121.11, 121.01, 118.89, 117.88, 63.76. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₂₀H₁₆N₂, 284.1300; found 284.1308.



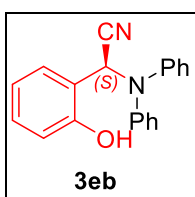
(S)-2-(2-chlorophenyl)-2-(diphenylamino)acetonitrile (3bb). Colourless solid; 225.8 mg, 99.9% yield; 95.5% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 9.1 min, tR (minor) = 19.1 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.30 (t, *J* = 7.8 Hz, 6H), 7.10 (d, *J* = 8.0 Hz, 6H), 6.96 (t, *J* = 7.4 Hz, 3H), 5.85 (d, *J* = 4.6 Hz, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 144.17, 142.09, 130.13, 130.03, 129.59, 129.10, 128.85, 128.31, 127.41, 126.69, 125.78, 122.95, 121.98, 119.97, 116.78, 59.99. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₂₀H₁₅ClN₂, 318.0900; found 318.0902.



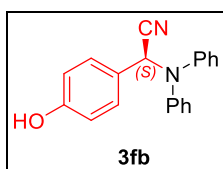
(S)-2-(4-chlorophenyl)-2-(diphenylamino)acetonitrile (3cb). Colourless solid; 209.8 mg, 97% yield; 98.4% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 17.2 min, tR (minor) = 22.4 min; ¹H NMR (500 MHz, CDCl₃) δ 7.07 (d, *J* = 7.9 Hz, 9H), 6.92 (t, *J* = 7.4 Hz, 5H), 5.51 (s, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 143.12, 133.73, 131.09, 129.38, 127.98, 121.09, 120.97, 118.38, 117.81, 63.08. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₂₀H₁₅ClN₂, 318.0900; found 318.0909.



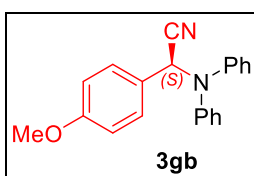
(S)-2-(diphenylamino)-2-(4-nitrophenyl)acetonitrile (3db). yellow solid; 248.1 mg, 92% yield; 66.4 % ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) =17.0 min, tR (minor) = 18.6 min; ^1H NMR (500 MHz, Chloroform-*d*) δ 8.33 (d, J = 8.8 Hz, 1H), 7.80 – 7.73 (m, 2H), 7.29 (s, 1H), 7.28 (d, J = 1.2 Hz, 2H), 7.27 (s, 3H), 7.11 – 7.06 (m, 4H), 6.94 (s, 2H), 5.71 (s, 1H). ^{13}C NMR (126 MHz, Chloroform-*d*) δ 143.13, 141.44, 129.36, 127.51, 124.37, 121.03, 117.83, 62.54. HRMS (ESI+) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{20}\text{H}_{15}\text{N}_3\text{O}$, 329.1200; found 329.1204.



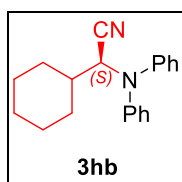
(S)-2-(diphenylamino)-2-(2-hydroxyphenyl)acetonitrile (3eb). Colourless solid; 218.4 mg, 95% yield; 98% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) =18.1 min, tR (minor) = 20.8 min; ^1H NMR (500 MHz, Chloroform-*d*) δ 7.52 – 7.48 (m, 1H), 7.27 (d, J = 7.8 Hz, 4H), 7.08 (d, J = 7.9 Hz, 7H), 6.94 (t, J = 7.4 Hz, 3H), 5.70 (s, 1H). ^{13}C NMR (126 MHz, Chloroform-*d*) δ 163.07, 141.56, 130.46, 128.39, 127.77, 126.75, 119.42, 116.24, 112.97, 112.76, 61.82, 54.03. HRMS (ESI+) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{20}\text{H}_{16}\text{N}_2\text{O}$, 300.1300; found 300.1304.



(S)-2-(diphenylamino)-2-(4-hydroxyphenyl)acetonitrile (3fb) Colourless solid; 241.3 mg, 99.9% yield; 94.6% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 18.8 min, tR (minor) =28.3 min; ^1H NMR (500 MHz, Chloroform-*d*) δ 7.81 (d, J = 8.2 Hz, 1H), 7.28 (t, J = 7.8 Hz, 5H), 7.08 (d, J = 8.0 Hz, 5H), 6.95 (q, J = 8.0 Hz, 3H), 5.73 (s, 1H); ^{13}C NMR (126 MHz, Chloroform-*d*) 163.59, 142.07, 131.00, 128.91, 128.31, 127.29, 119.95, 116.75, 113.50, 113.29, 62.36 HRMS (ESI+) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{20}\text{H}_{16}\text{N}_2\text{O}$, 300.1300; found 300.1305.

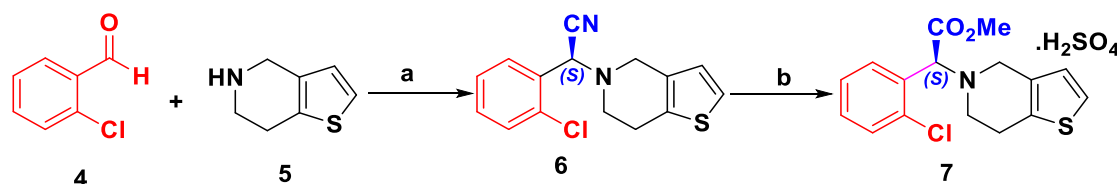


(S)-2-(diphenylamino)-2-(4-methoxyphenyl)acetonitrile (3gb). Colourless solid; 189.5 mg, 92% yield; 96.2% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) =5.0 min, tR (minor) = 5.6 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.30 – 7.26 (m, 4H), 7.11 – 7.07 (m, 6H), 7.02 (d, *J* = 8.8 Hz, 2H), 6.96 – 6.90 (m, 3H), 5.76 (s, 1H), 3.90 (s, 3H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 163.59, 142.07, 131.00, 128.31, 127.29, 119.95, 116.75, 113.50, 113.29, 62.36, 54.56. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₂₁H₁₈N₂O, 314.1400; found 314.1402.



(S)-2-cyclohexyl-2-(diphenylamino)acetonitrile (3hb). Colourless solid; 176.1 mg, 92% yield; 96.1% ee; determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 90/10, flow rate: 1 mL/min, wavelength=210 nm: tR (major) = 9.5 min, tR (minor) = 13.0 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.28 (d, *J* = 7.8 Hz, 3H), 7.08 (d, *J* = 7.9 Hz, 5H), 6.93 (t, *J* = 7.4 Hz, 2H), 4.27 (d, *J* = 6.2 Hz, 1H), 1.89 (s, 1H), 1.85 – 1.80 (m, 2H), 1.75 – 1.70 (m, 3H), 1.27 (q, *J* = 5.0 Hz, 5H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 141.55, 127.77, 119.43, 116.24, 64.84, 40.72, 26.52, 26.20, 24.35, 23.87, 23.84. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₂₀H₂₂N₂, 290.1800; found 290.1800.

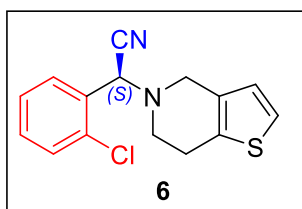
5. Procedure for the Synthesis of S-Clopidogrel*



*Reaction Conditions (a) **4** (3 g, 21mmol), **5** (2.9 g, 21 mmol), TMSCN (2.0 g, 21 mmol) DCM (20 ml) to give **6** (98% yield, 99.3% ee) ;(b) MeOH, H₂SO₄ to give **7** (73.2% yield, 98.5% ee) ;(c) % ee determined by HPLC equipped with AD-H chiral columns and configuration was determined by relative retention time and specific rotation with literature data

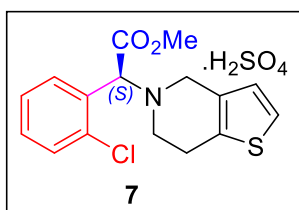
In an inert environment 2-Chlorobenzaldehyde (3 g, 21mmol), 4,5,6,7-tetrahydrothieno[3,2-c]pyridine (2.9 g, 21 mmol) TMSCN (2.0 g, 21 mmol), were introduced in a round bottom flask with dichloromethane (20 ml) as a solvent and catalyst V (0.2 mol%) The reaction mixture was stirred continuously for 15 minutes and monitored through TLC. The reaction mixture was treated with chloroform and washed with a saturated solution of sodium bisulphite. The organic phase was dried over anhydrous sodium sulphate. The solvent was then evaporated under reduced pressure and the crude was purified through column chromatography (n-hexane/ethyl acetate 80:20) to give (S)-2-(2-chlorophenyl)-2-(6,7-dihydrothieno[3,2-c]pyridin-5(4H)-yl)acetonitrile (5.9 g, 98% yield). To this compound **6** esterification was done with MeOH (40 ml), and H₂SO₄ (1 ml) and the reaction was continued for 4 days and monitored through TLC. The reaction mixture was treated with water followed by the addition of sodium

carbonate and dichloromethane. After drying over Na₂SO₄ solvent was evaporated to obtain S-Clopidogrel (4.8 g, 73.2% yield). The enantiomeric excess was determined by chiral stationary-phase HPLC analysis using AD-H chiral column.



(S)-2-(2-chlorophenyl)-2-(6,7-dihydrothieno[3,2-c]pyridin-5(4H)-yl)acetonitrile(6):

colourless solid 5.9 g, 98% yield; 99.3% ee determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 60/40, flow rate: 1 mL/min, wavelength=210 nm : t_R (major) = 10.6 min, t_R (minor) = 12.2 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.71 (s, 1H), 7.48 – 7.33 (m, 3H), 7.09 (d, *J* = 5.2 Hz, 1H), 6.71 (d, *J* = 5.2 Hz, 1H), 5.33 (s, 1H), 3.81 (d, *J* = 13.7 Hz, 1H), 3.67 (d, *J* = 13.7 Hz, 1H), 3.03 – 2.90 (m, 3H), 2.86 (s, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 133.06, 131.36, 130.95, 129.02, 128.96, 128.47, 125.29, 123.51, 121.57, 113.65, 57.69, 47.88, 46.24, 24.04. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₁₅H₁₃ClN₂S, 288.0500; found. 288.0502



methyl(S)-2-(2-chlorophenyl)-2-(6,7-dihydrothieno[3,2-c]pyridin-5(4H)-yl)acetate(7):

colourless solid 4.8 g, 73.2% yield; 98.5% ee determined by HPLC analysis (Daicel Chiralpak AD-H column, hexane/2-propanol = 60/40, flow rate: 1 mL/min, wavelength=210 nm : t_R (major) = 12.3 min, t_R (minor) = 13.7 min; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.70 (dd, *J* = 7.3, 2.2 Hz, 1H), 7.41 (dd, *J* = 7.4, 1.9 Hz, 1H), 7.30 (d, *J* = 1.8 Hz, 1H), 7.28 (d, *J* = 2.3 Hz, 1H), 7.06 (d, *J* = 5.2 Hz, 1H), 6.67 (d, *J* = 5.2 Hz, 1H), 4.92 (s, 1H), 3.76 (d, *J* = 14.0 Hz, 1H), 3.73 (s, 3H), 3.63 (d, *J* = 14.1 Hz, 1H), 2.88 (s, 4H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 129.98, 129.82, 129.46, 127.21, 125.27, 122.78, 67.90, 52.24, 50.74, 48.34, 29.73, 25.53. HRMS (ESI+) *m/z* [M + H]⁺ calcd for C₁₆H₁₆ClN₂S, 321.0600; found 321.0608

6. Recyclability Test (under scale-up condition)

The reaction was performed in a 250 mL round bottom flask at 25 °C in an inert environment, benzaldehyde **1a** (8.4 g, 80 mmol), aniline **2a** (7.4 g, 80 mmol), trimethylsilyl cyanide (7.9 g, 80 mmol), and catalyst **V** (0.2 mol%) were introduced in a dry glass tube with dichloromethane (120 ml) as a solvent. The reaction mixture was stirred continuously for 15 minutes and monitored through TLC. After completion of reaction, solvent was removed *in-vacuo*. The reaction mixture was dissolved in cold diethyl ether (100x3 times) and organic layer was

washed with saturated solution of sodium bisulphite. The organic phase was dried over anhydrous sodium sulphate. The solvent was then evaporated under reduced pressure to afford 3aa (16.4g; 99.2% yield). For the next cycle, in the same flask (containing catalyst), 120 mL dichloromethane was charged and stirred for 15 min, followed by subsequent catalytic cycles under similar conditions as described above.

7. Time-dependent NMR:

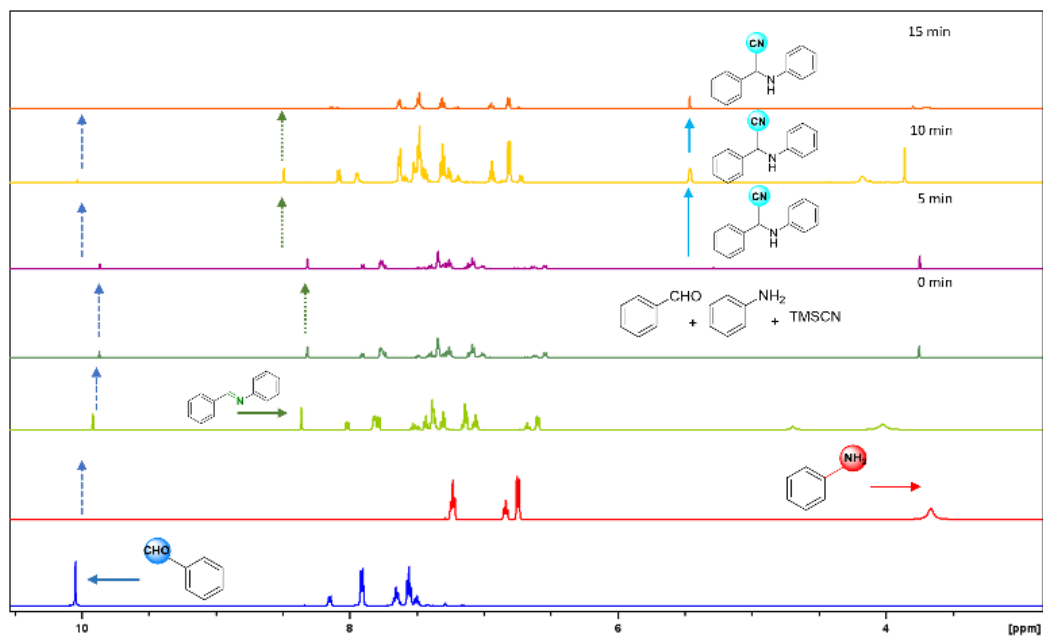


Figure 1

In Figure S1 The time dependent NMR showed the formation of imine intermediate and it was ascertained by the downfield value of chemical shift i.e. 8.91 ppm. As the reaction progresses the α -aminonitrile moiety starts forming as the upfield value of 5.02 ppm of Hydrogen attached to quaternary carbon of α -aminonitrile starts appearing. Within the due course of time eventually, the chemical shift value of the imine intermediate and aldehyde would disappear completely within 15 minutes of time

8. IR and UV-visible spectroscopic study-

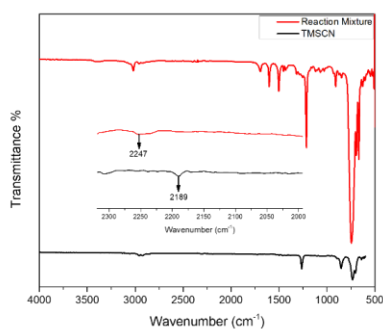


Figure 2

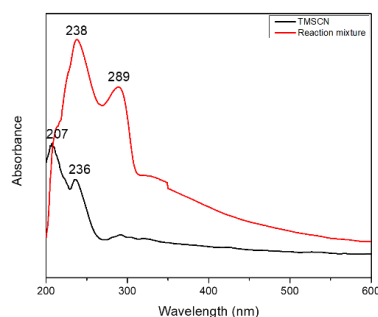


Figure 3

In figure S2: IR spectrum, the peak intensities at 1697cm^{-1} (CO), 2189cm^{-1} (CN) for aldehyde and TMSCN respectively were diminished in the cyanated product. The characteristic peaks at 3021cm^{-1} (N-H) and 2247cm^{-1} (CN) indicate the formation of the α -aminonitrile moiety.

In **Figure S3**: of the UV absorbance spectrum, TMSCN shows a low-intensity band at 207 nm and a high-intensity band at 236 nm due to $n-\pi^*$ and $\pi-\pi^*$ transition respectively. In a similar way, the formation of α -aminonitrile moiety was confirmed by the shift in $n-\pi^*$ (238 nm) and $\pi-\pi^*$ transition (289 nm). The shift in absorbance maxima is due to the presence of a benzene chromophore.

9. ^{29}Si -NMR

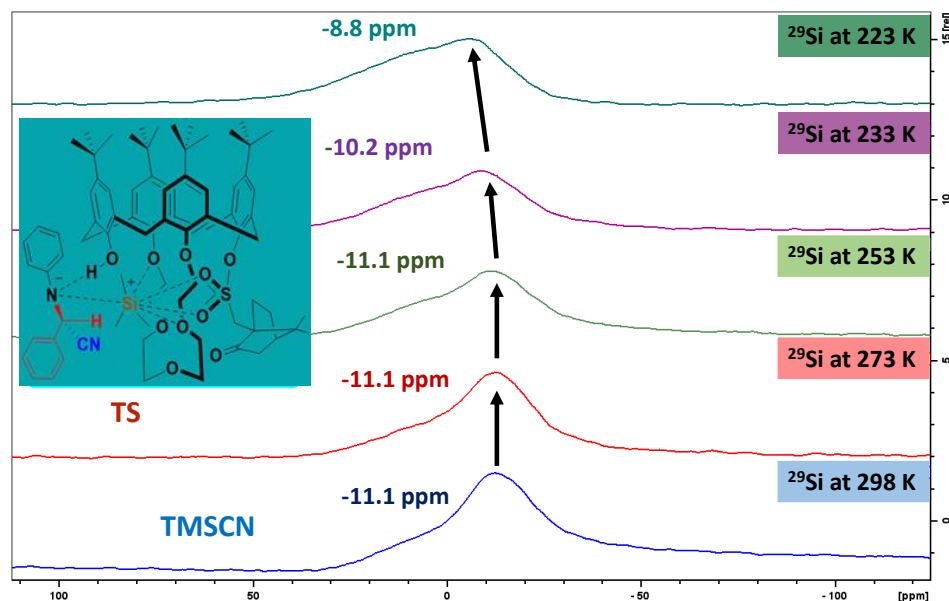


Figure S4: ^{29}Si (500MHz) analysis was carried out at a temperature range of rt to $-50\text{ }^\circ\text{C}$. All experiments were performed in CDCl_3 . ^{29}Si -NMR Spectra were recorded and scaled from TMS^2

10. HRMS

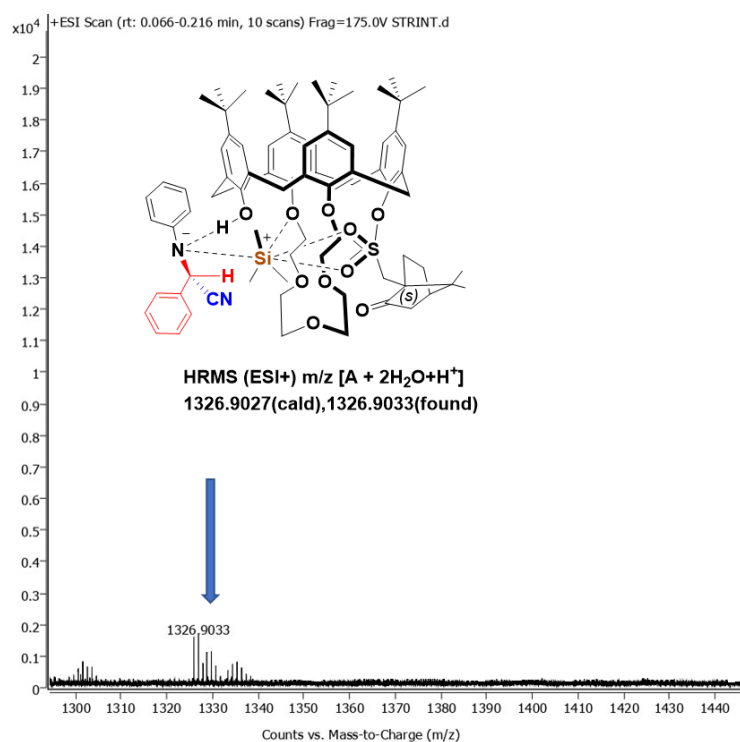
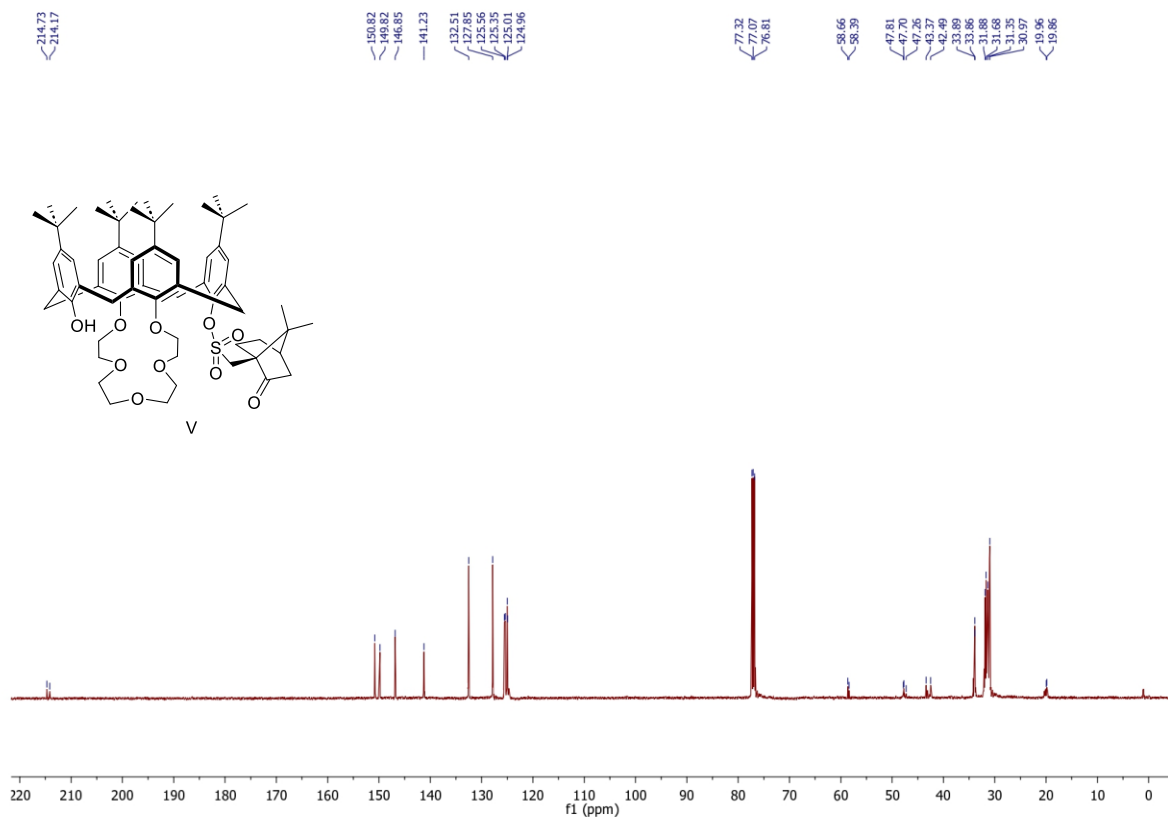
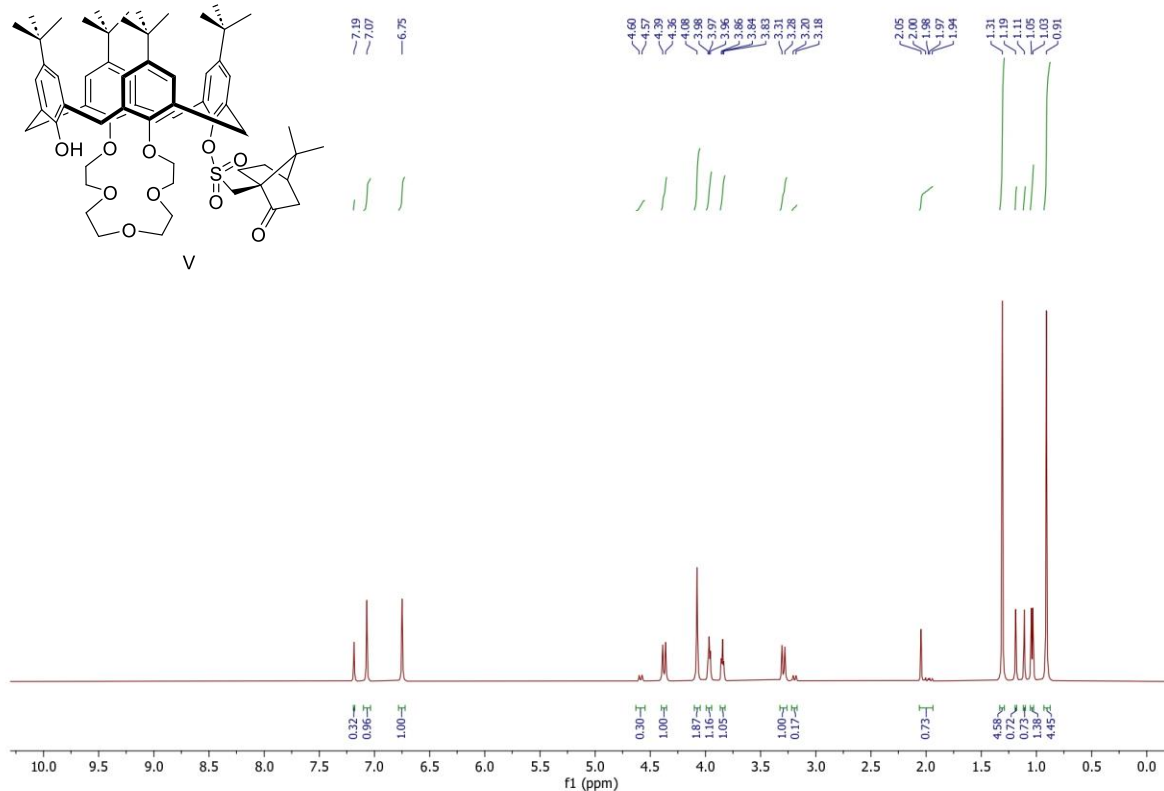
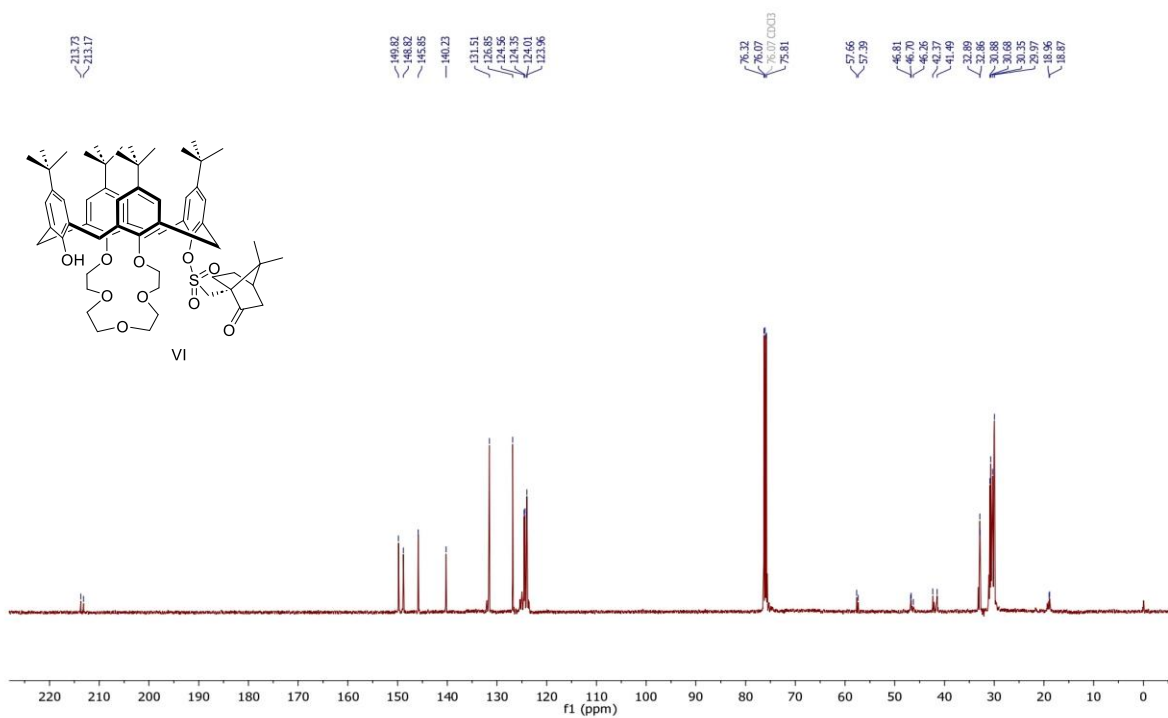
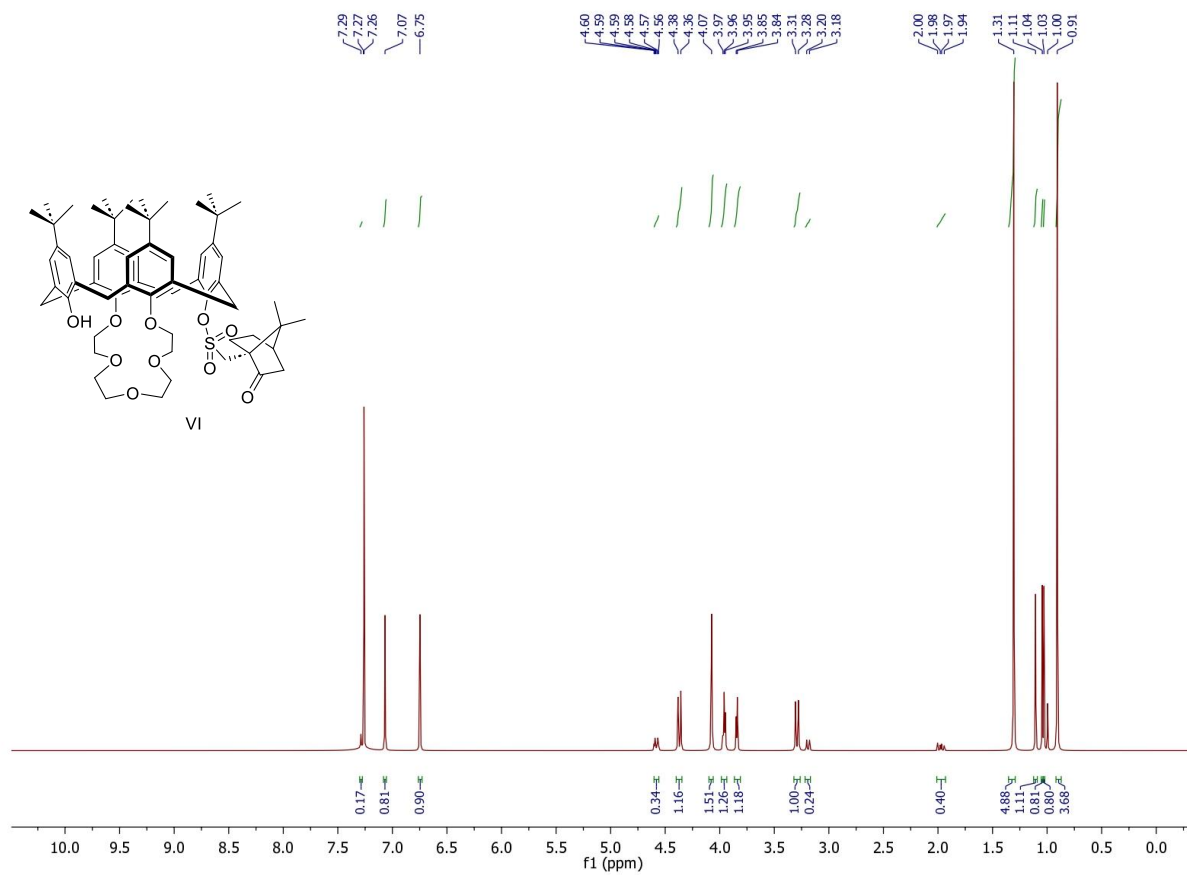


Figure S5: ESI-MS analysis of Transition state

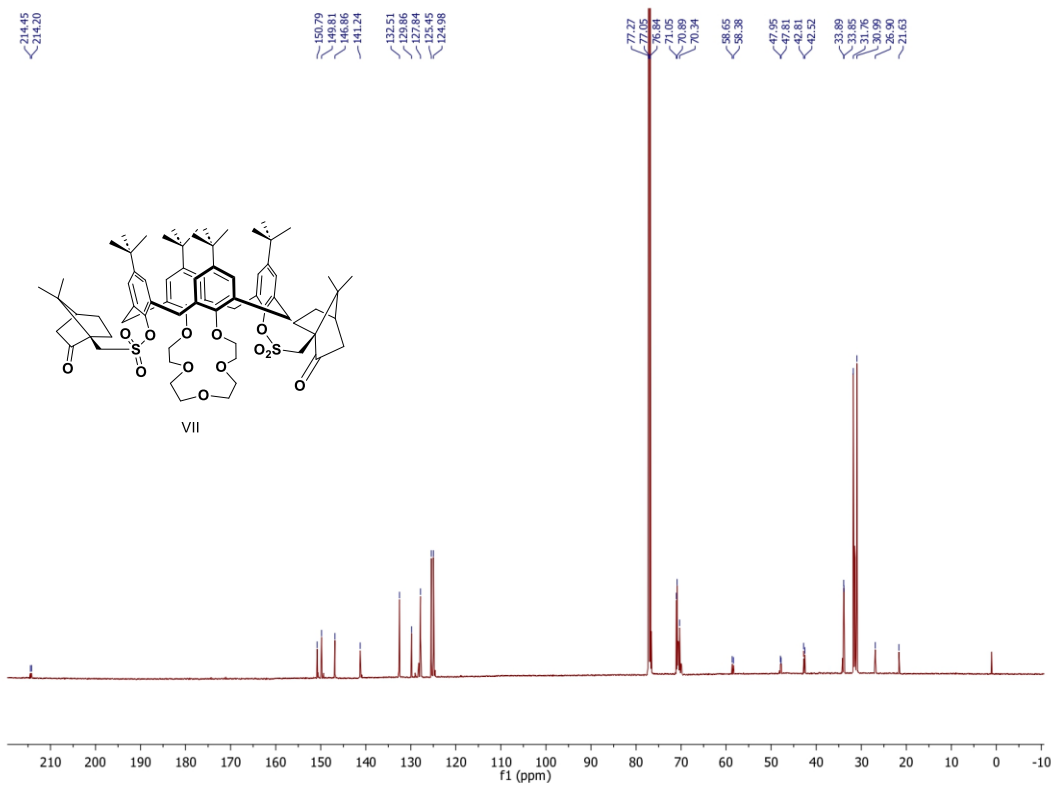
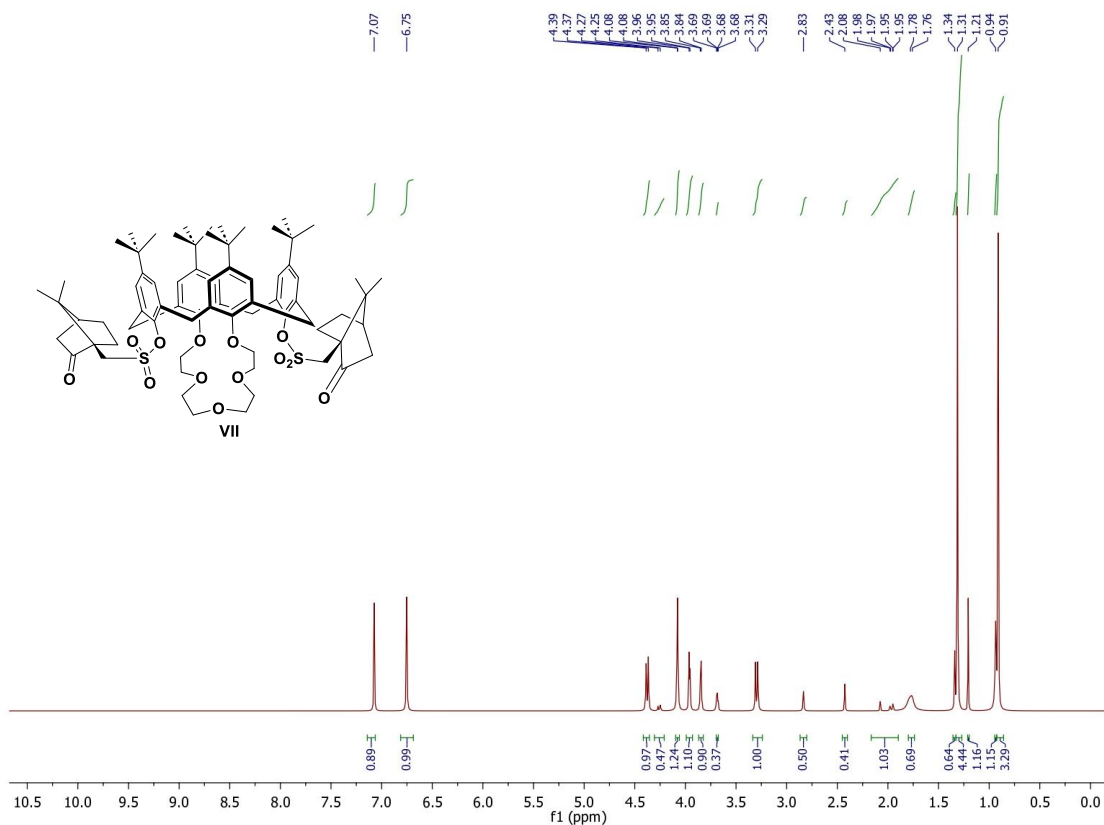
11. NMR (Catalyst-V)



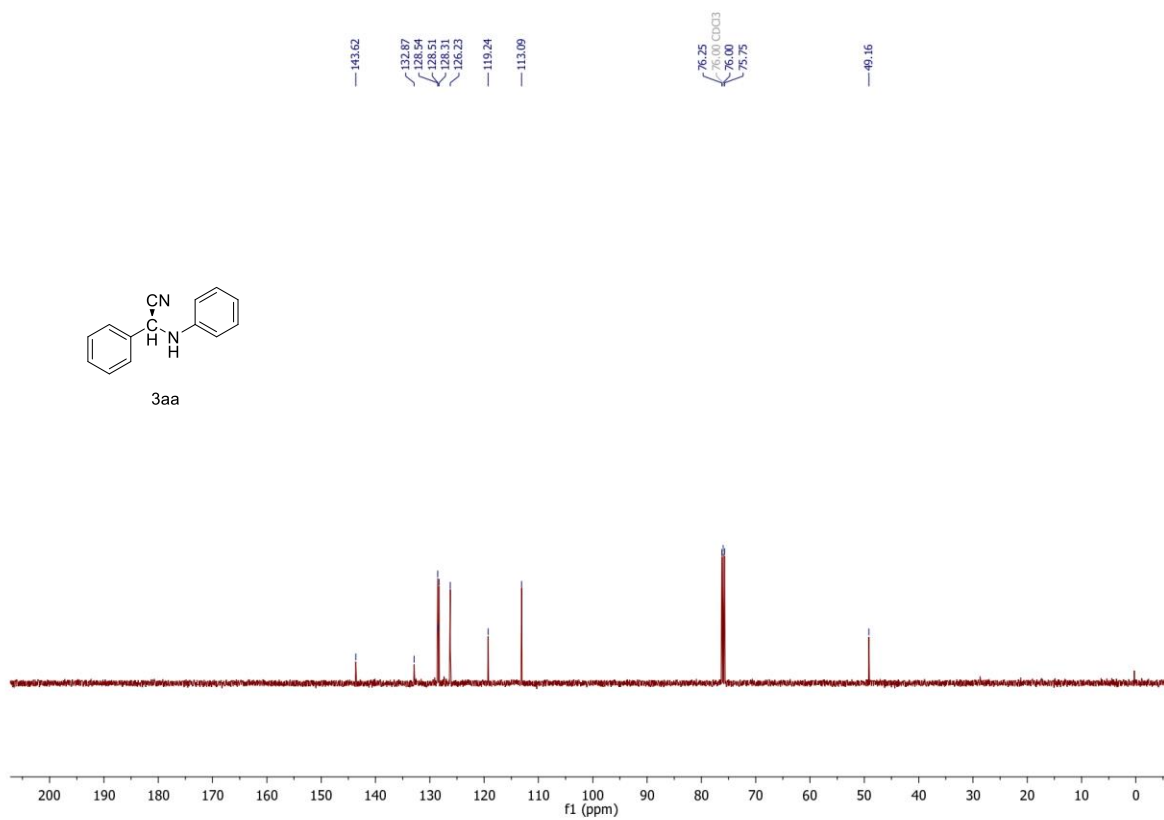
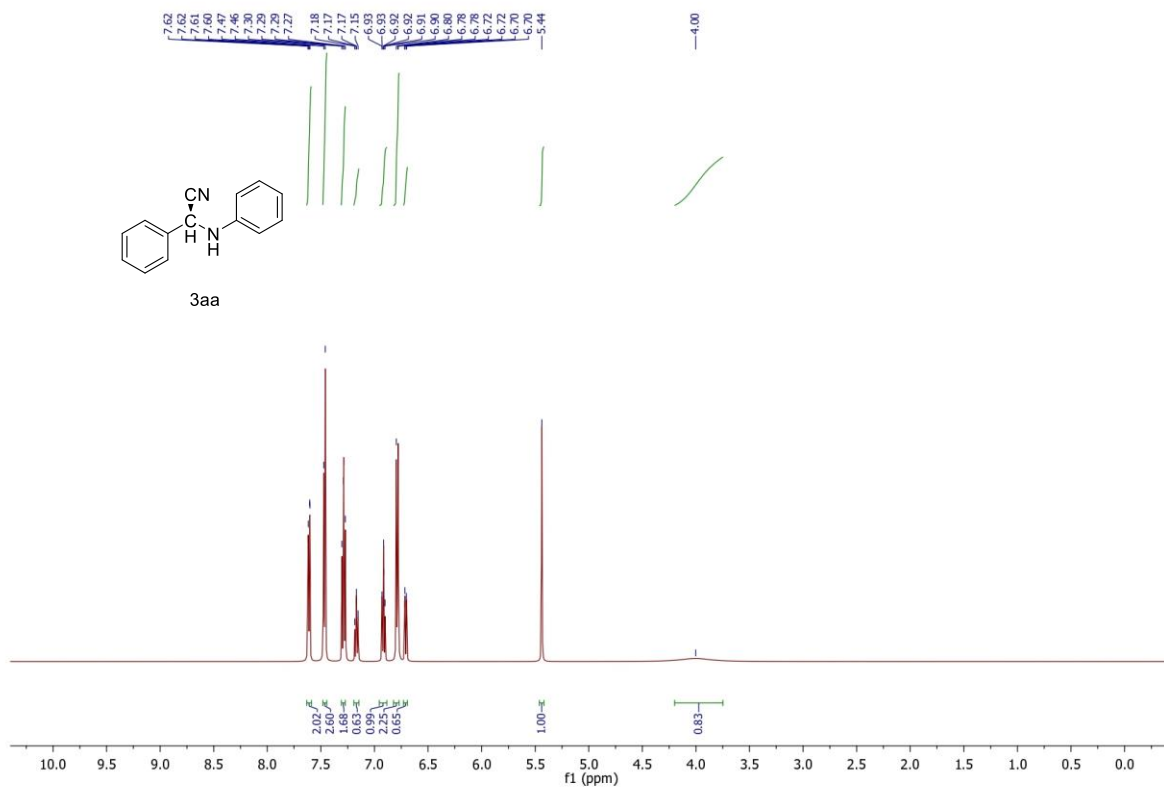
Catalyst-VI



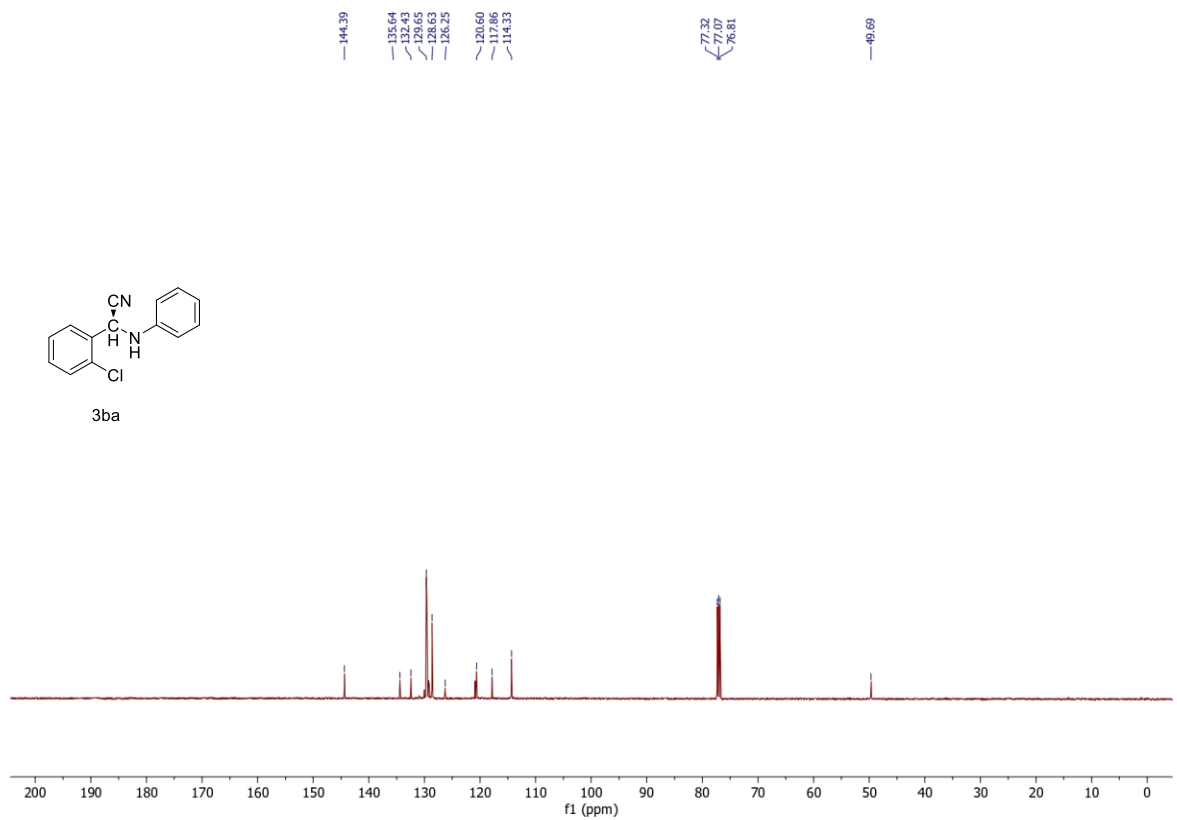
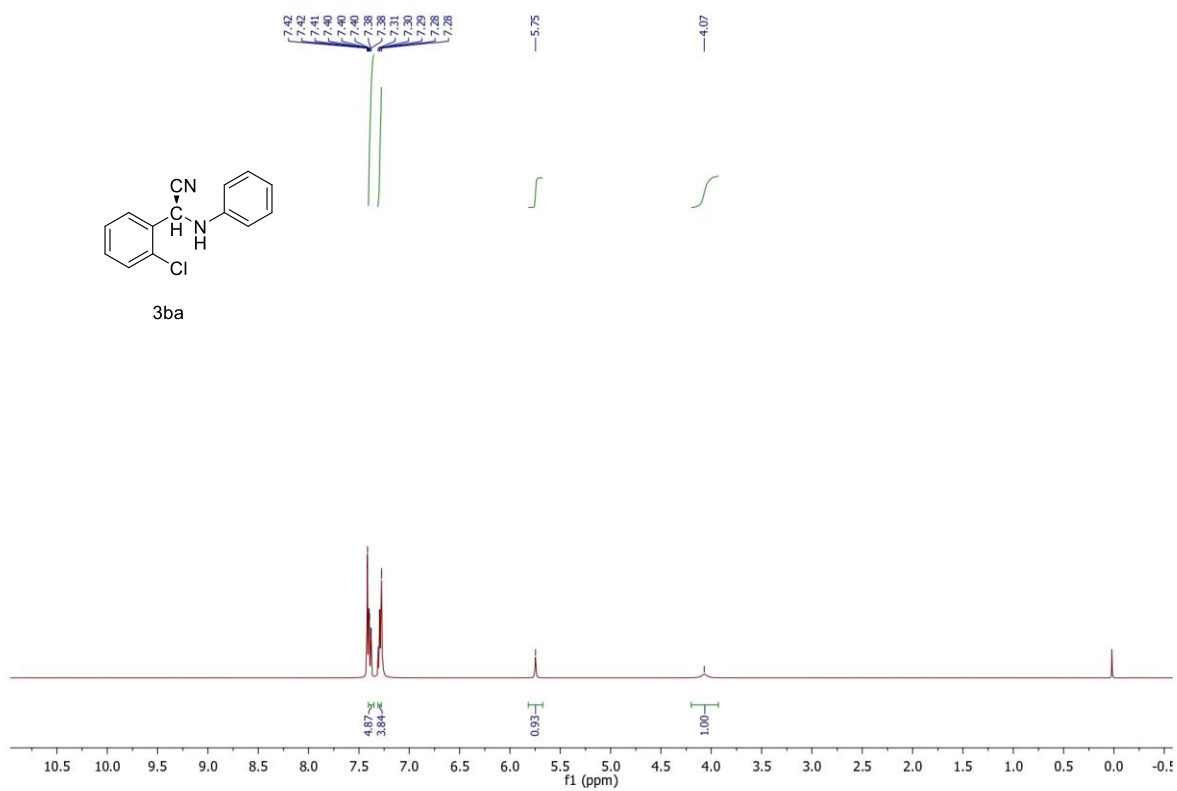
Catalyst VII



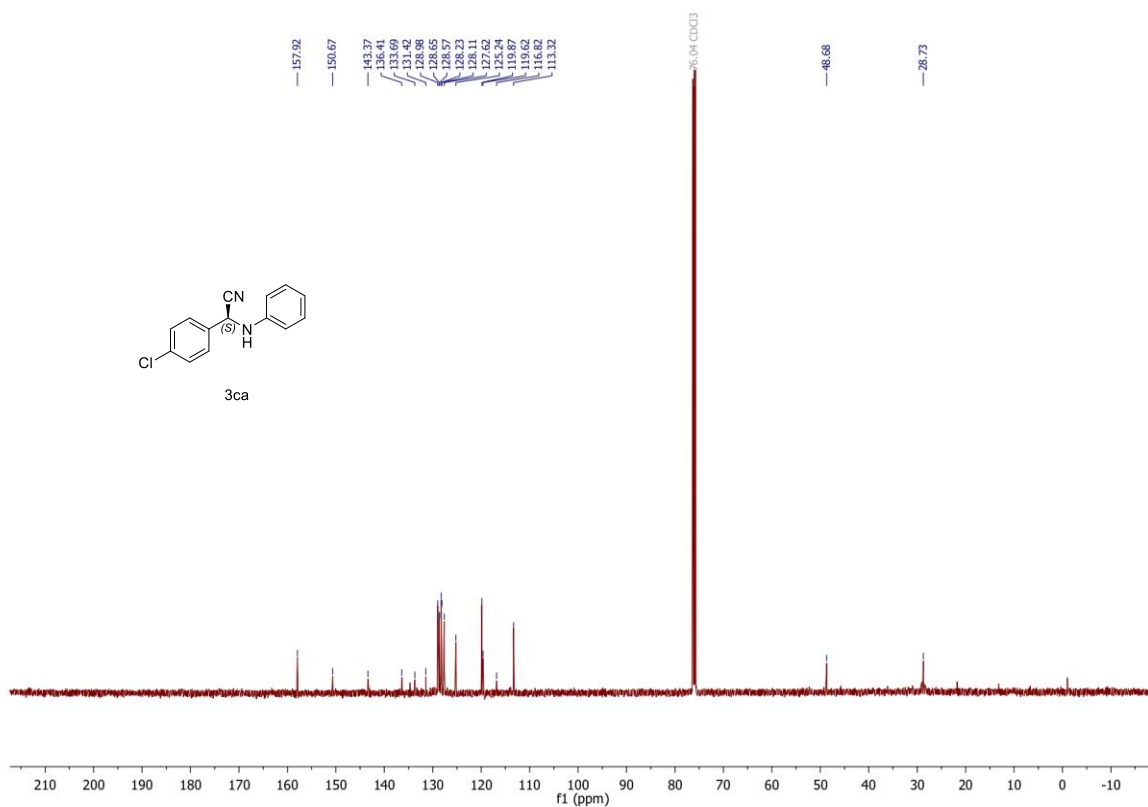
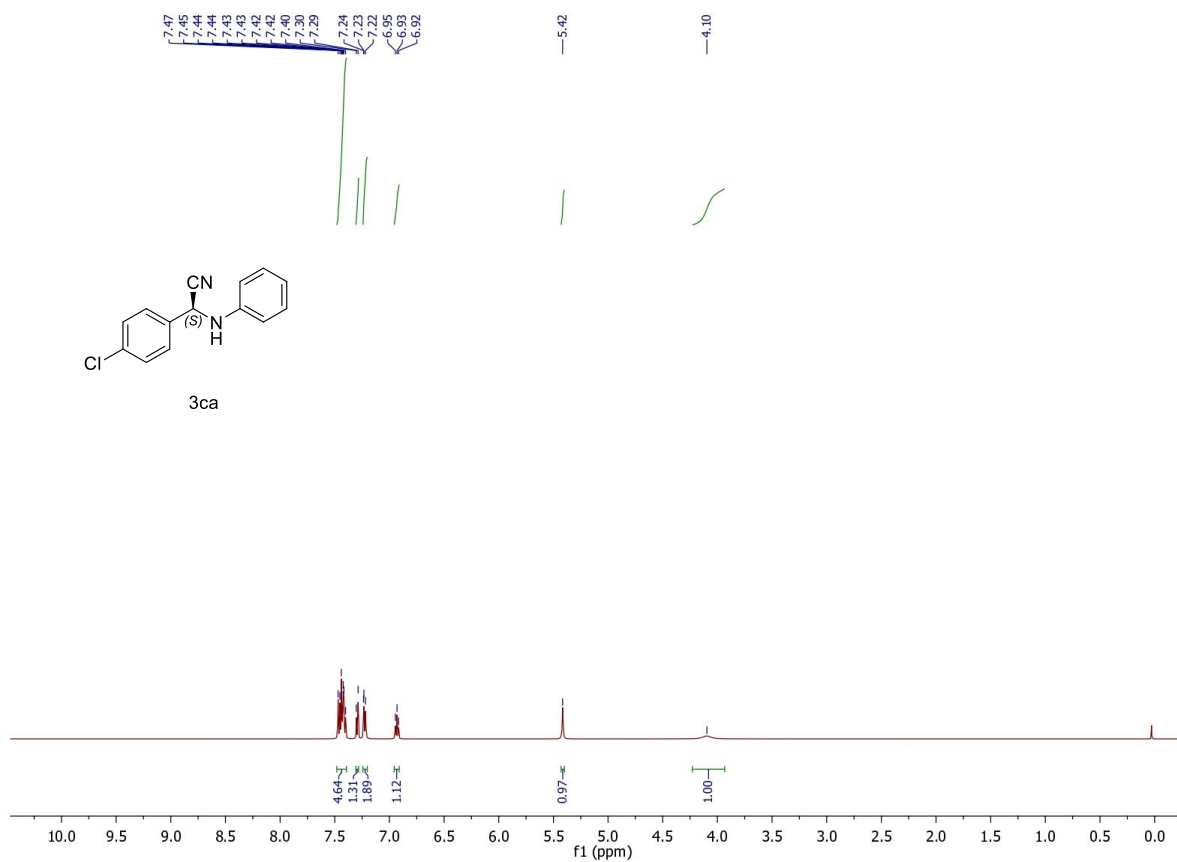
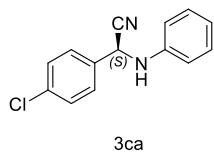
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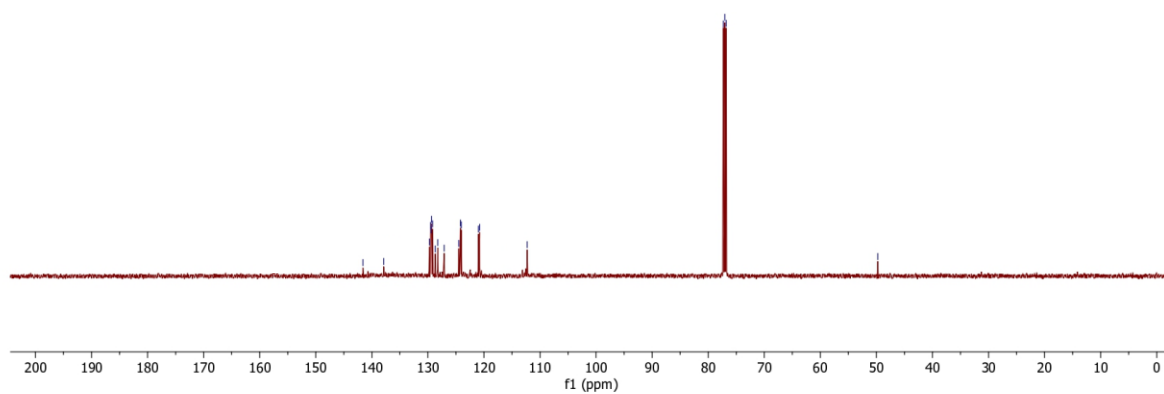
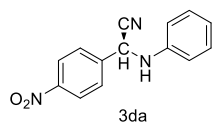
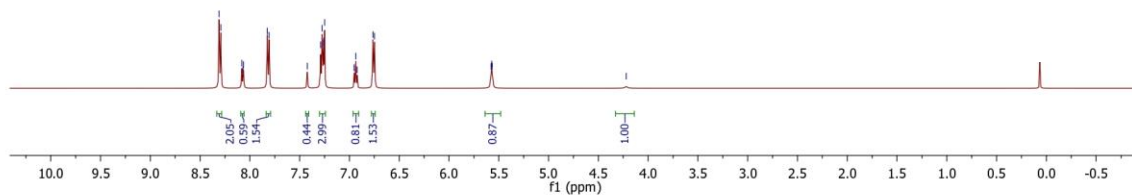
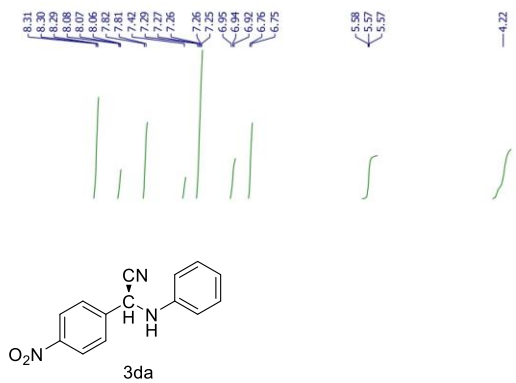
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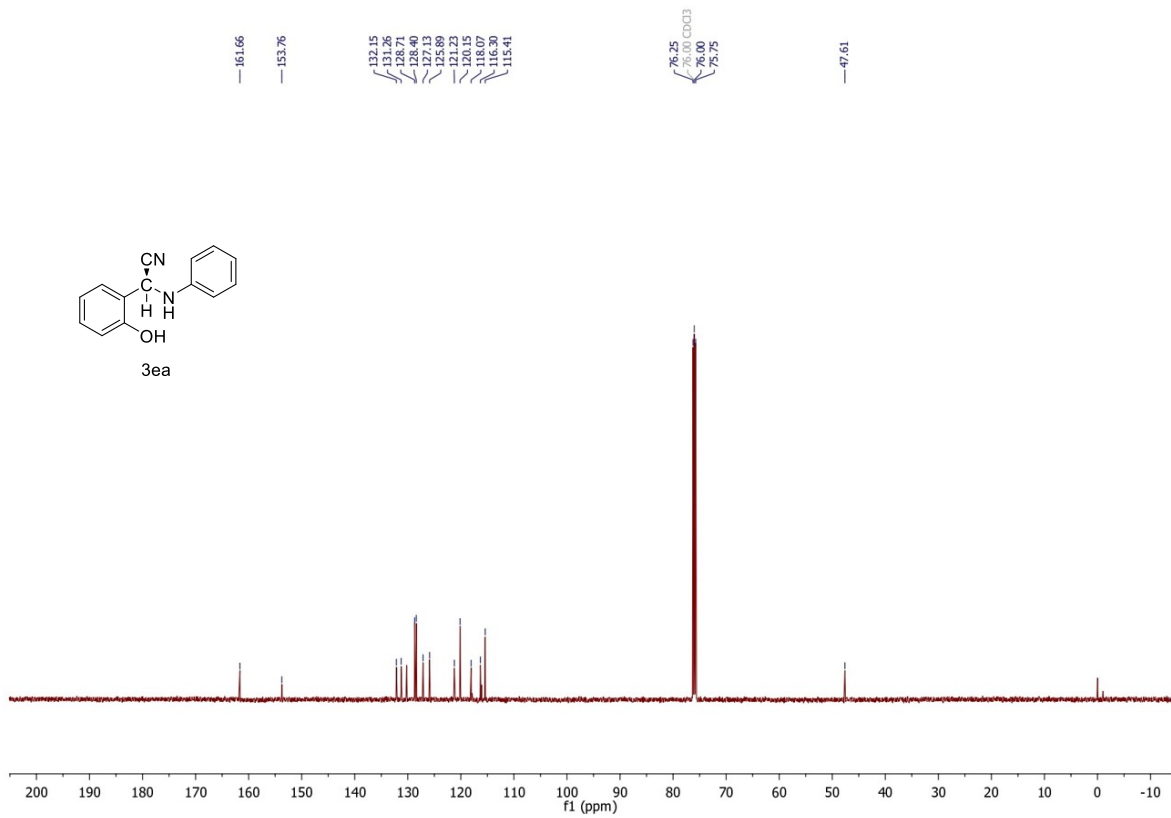
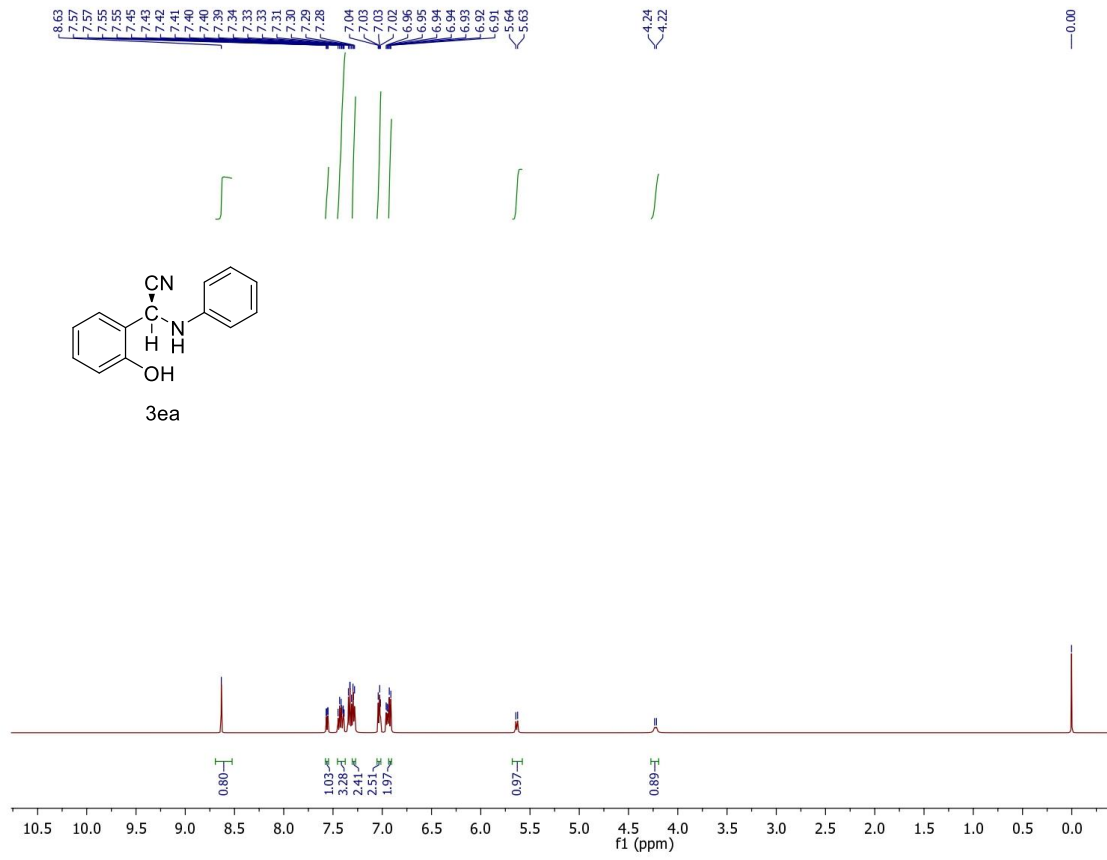
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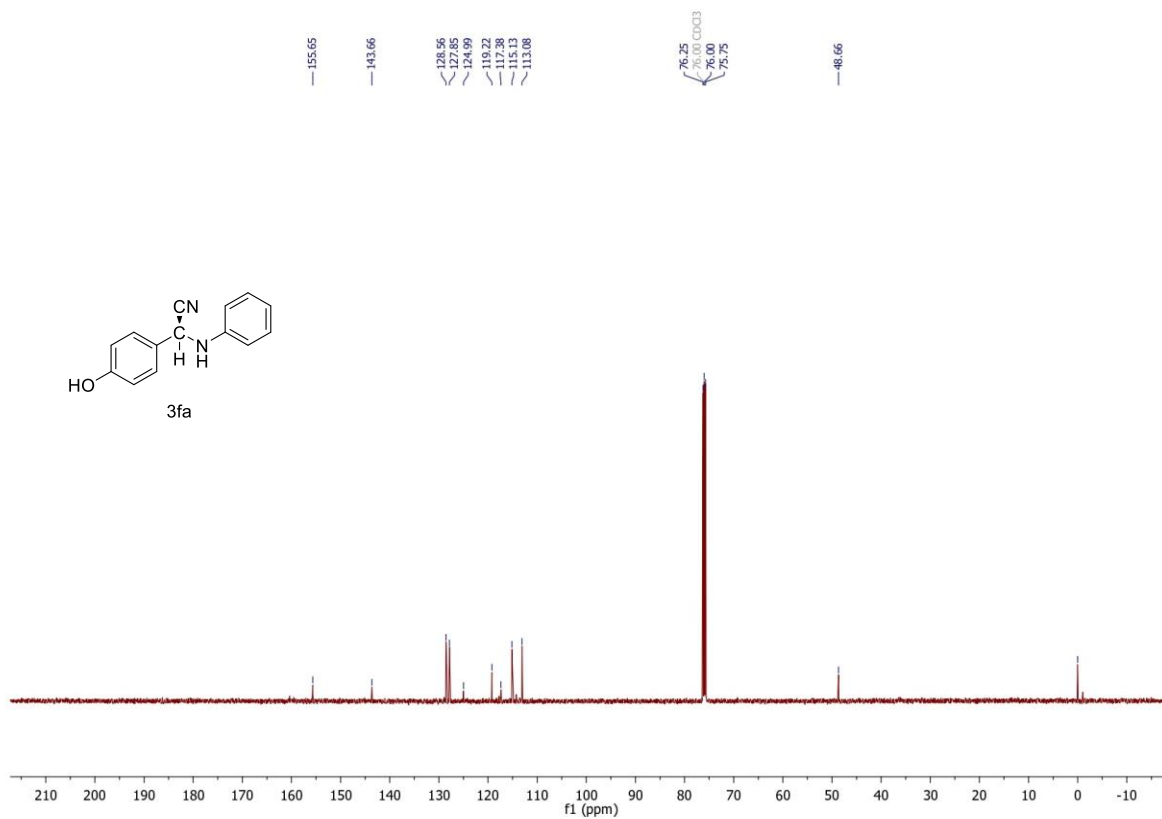
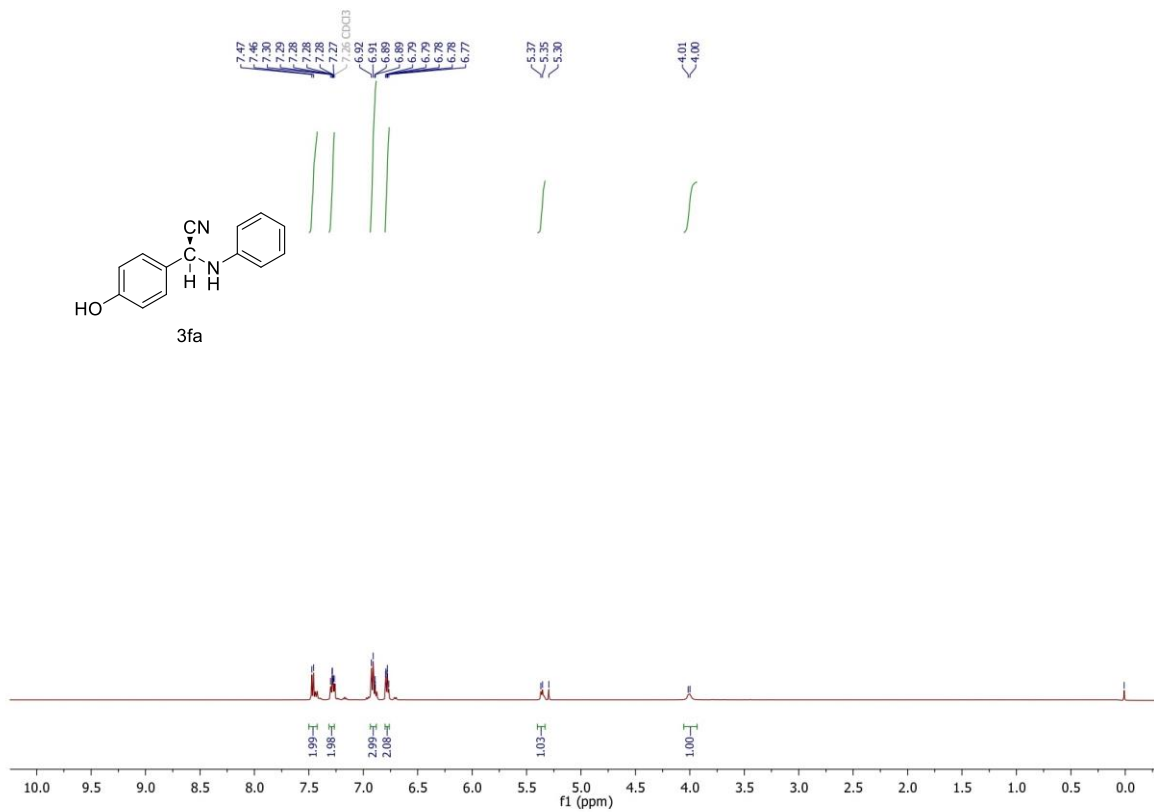
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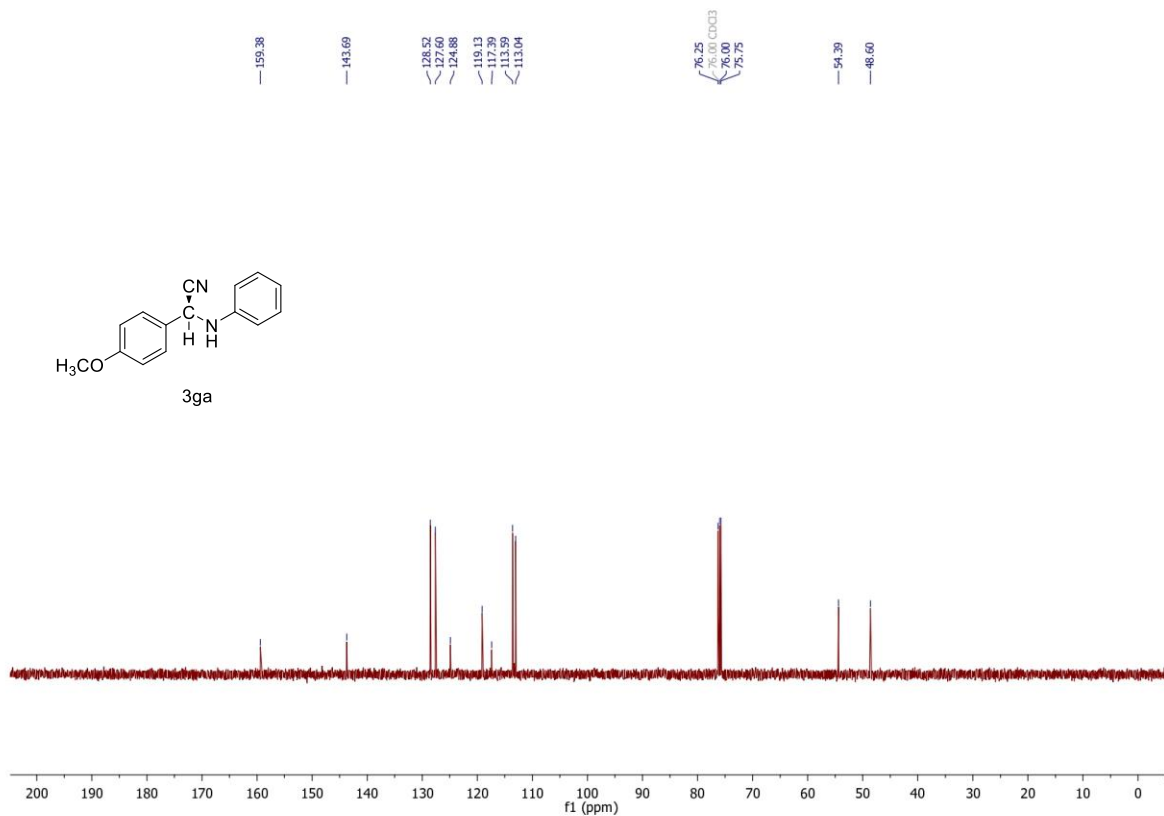
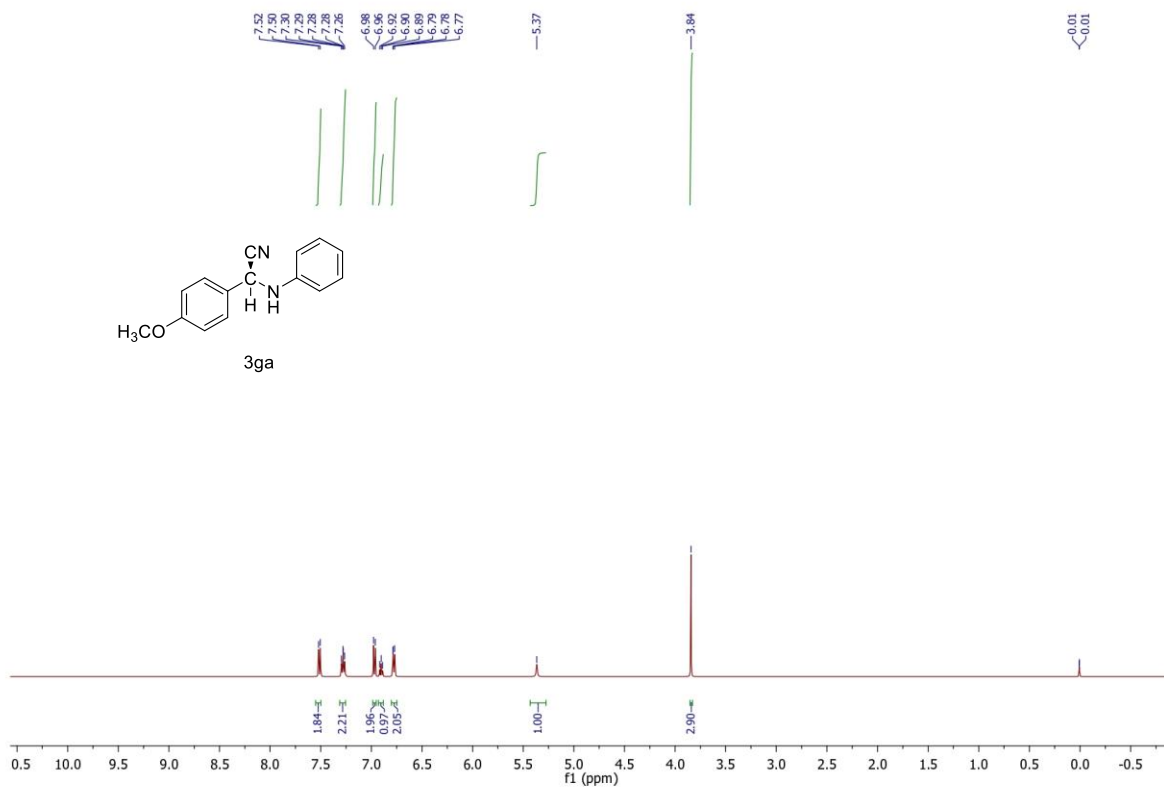
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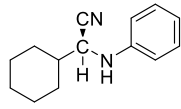
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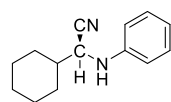
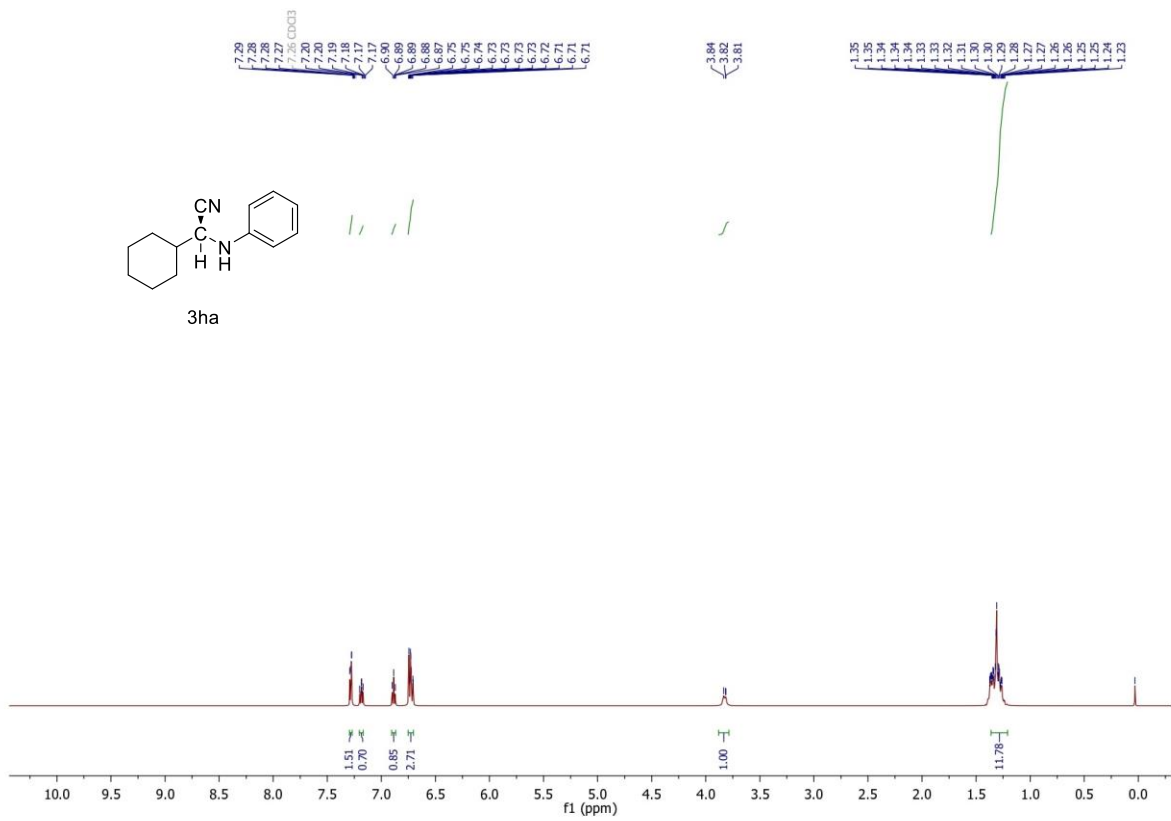
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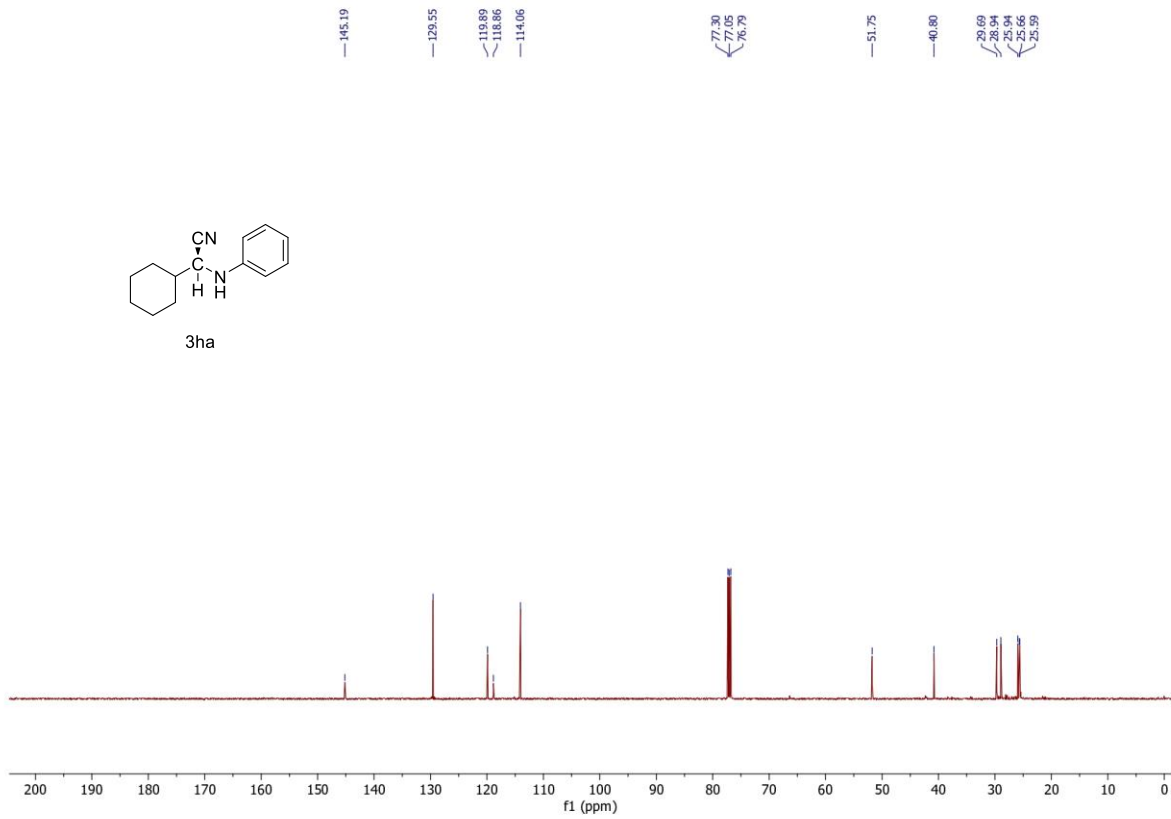
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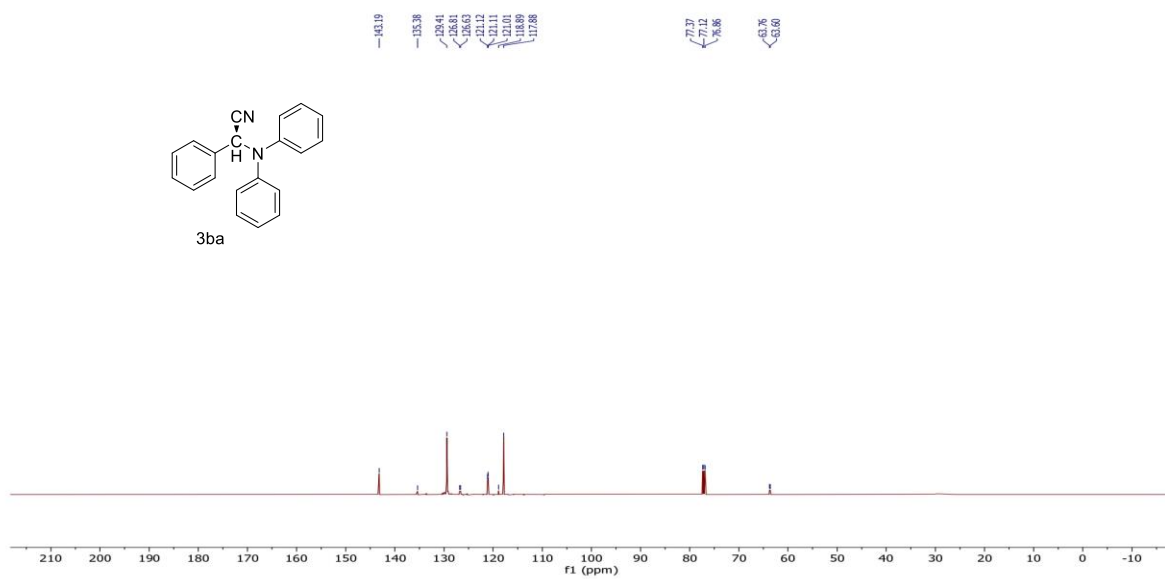
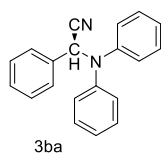
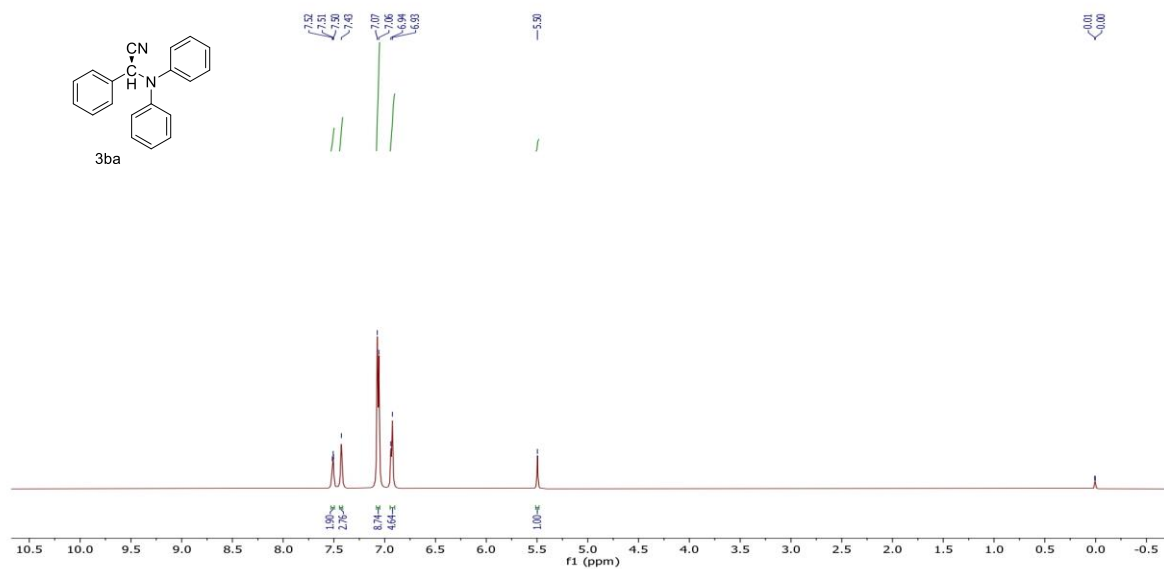
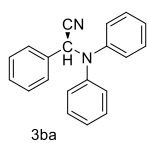
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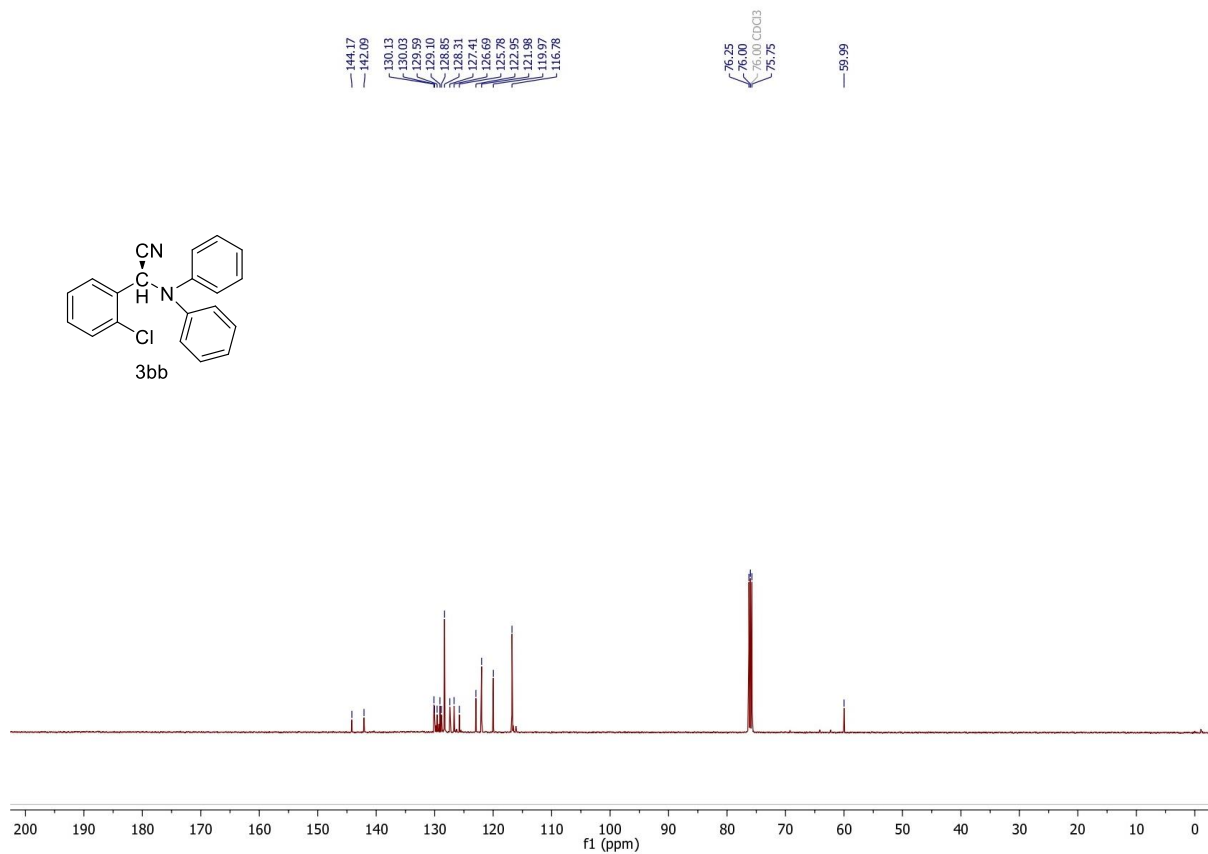
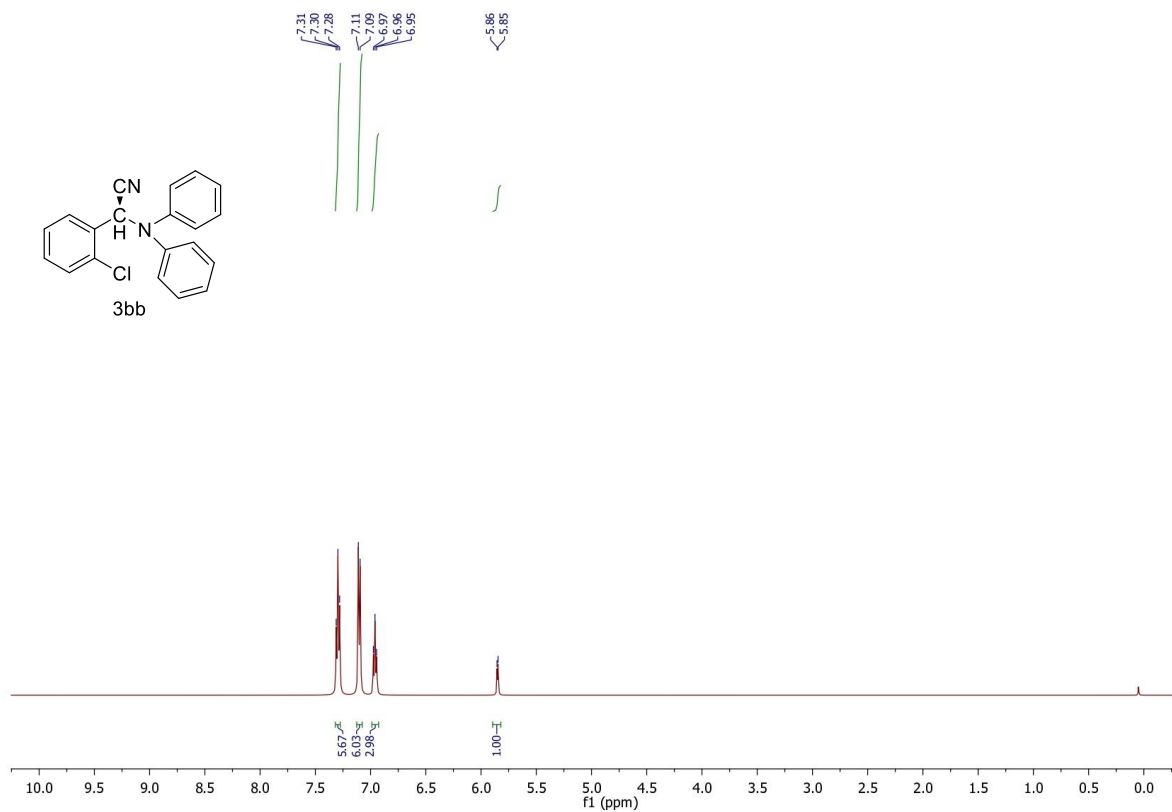
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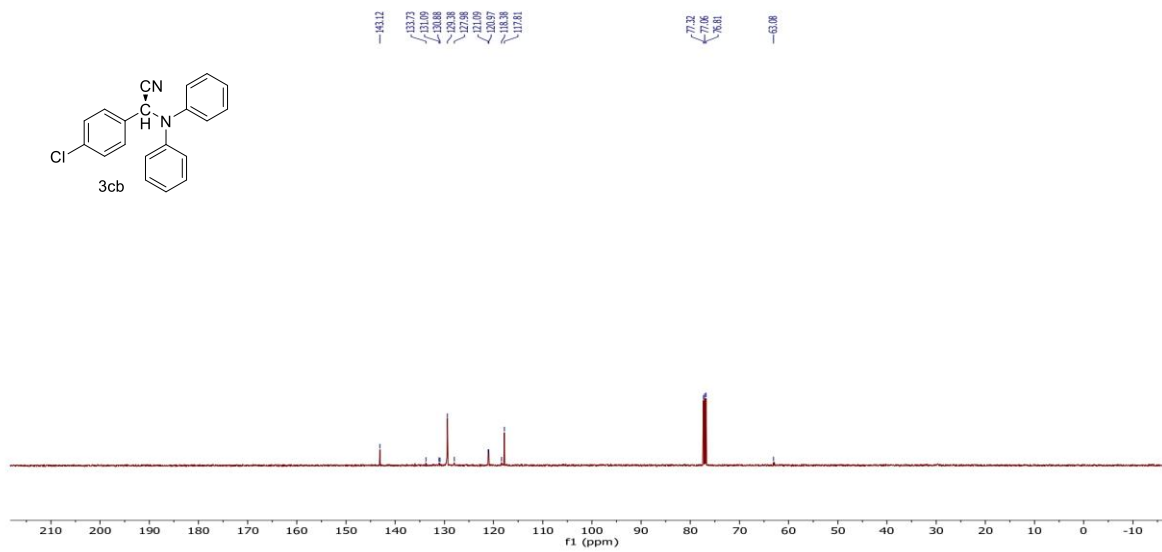
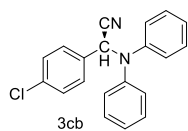
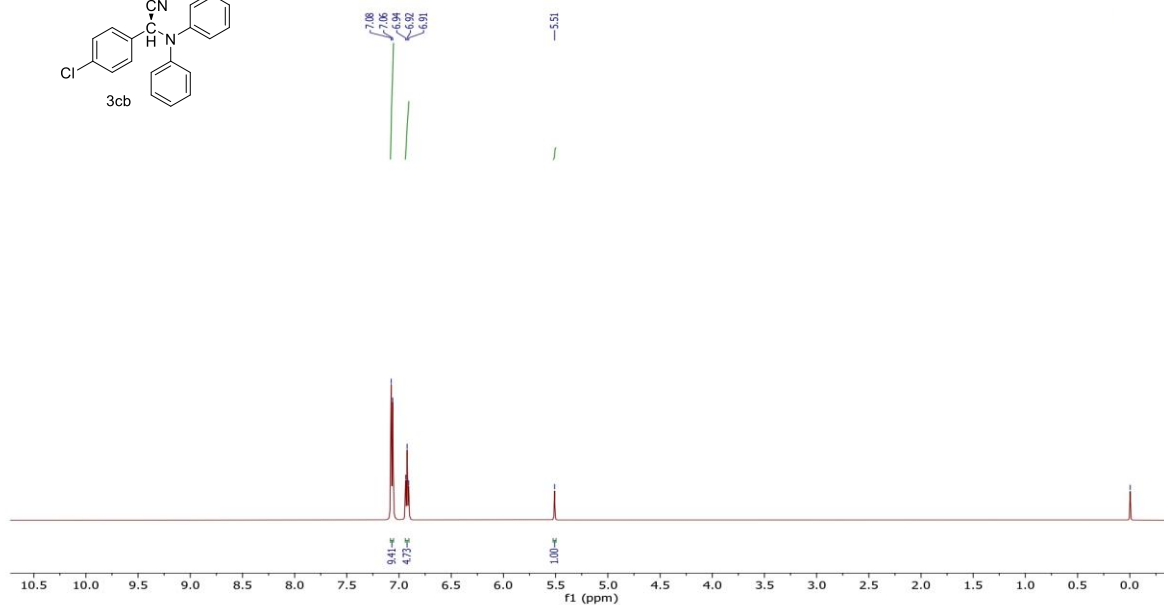
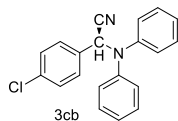
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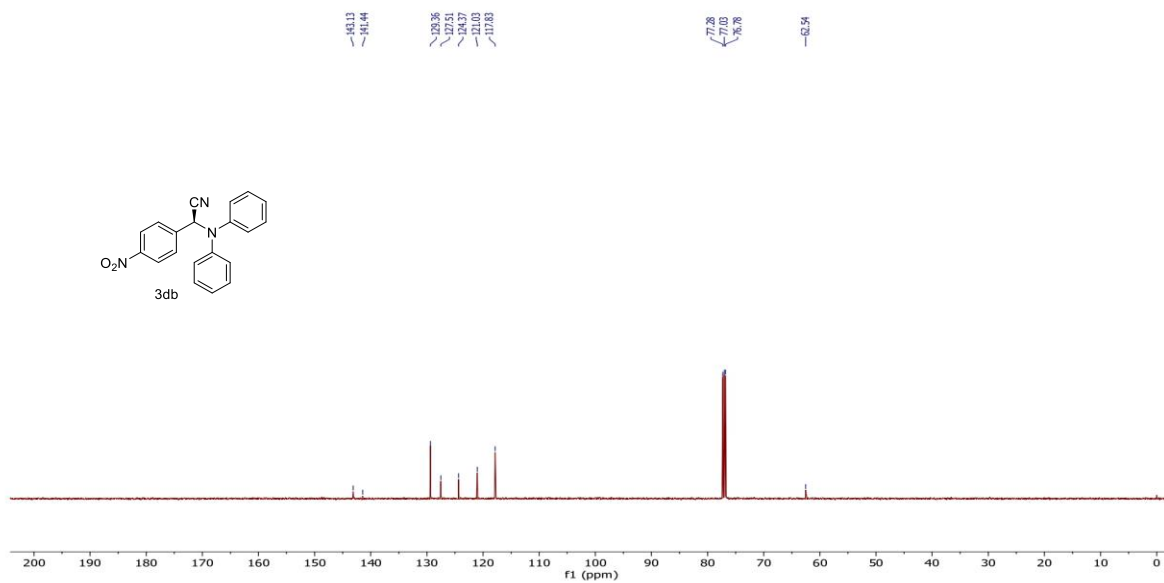
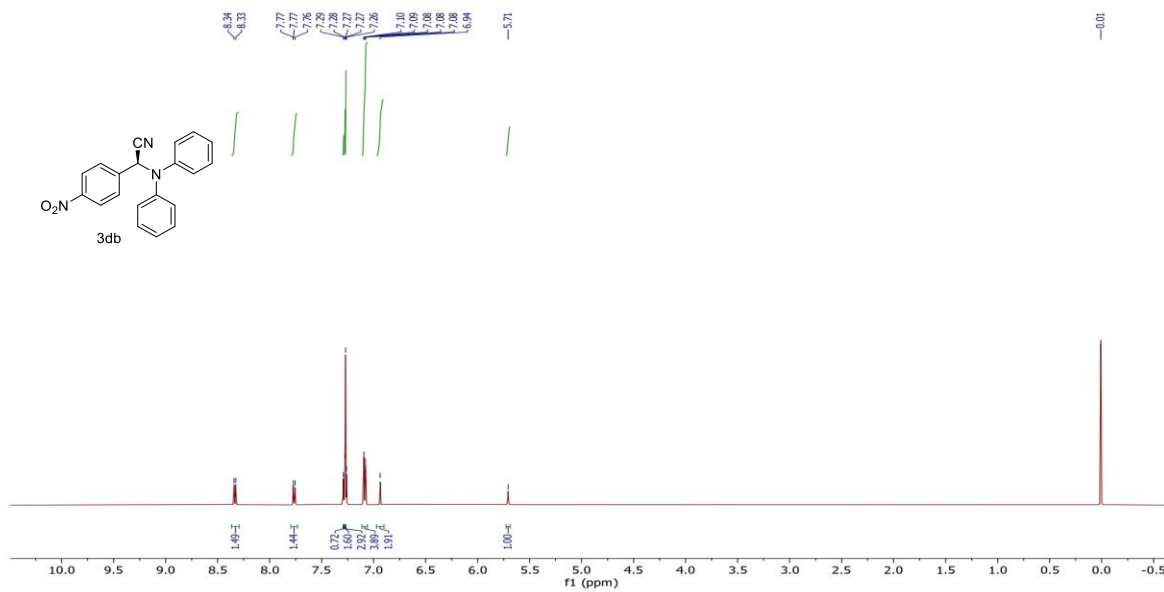
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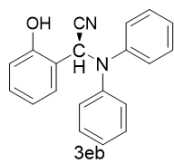
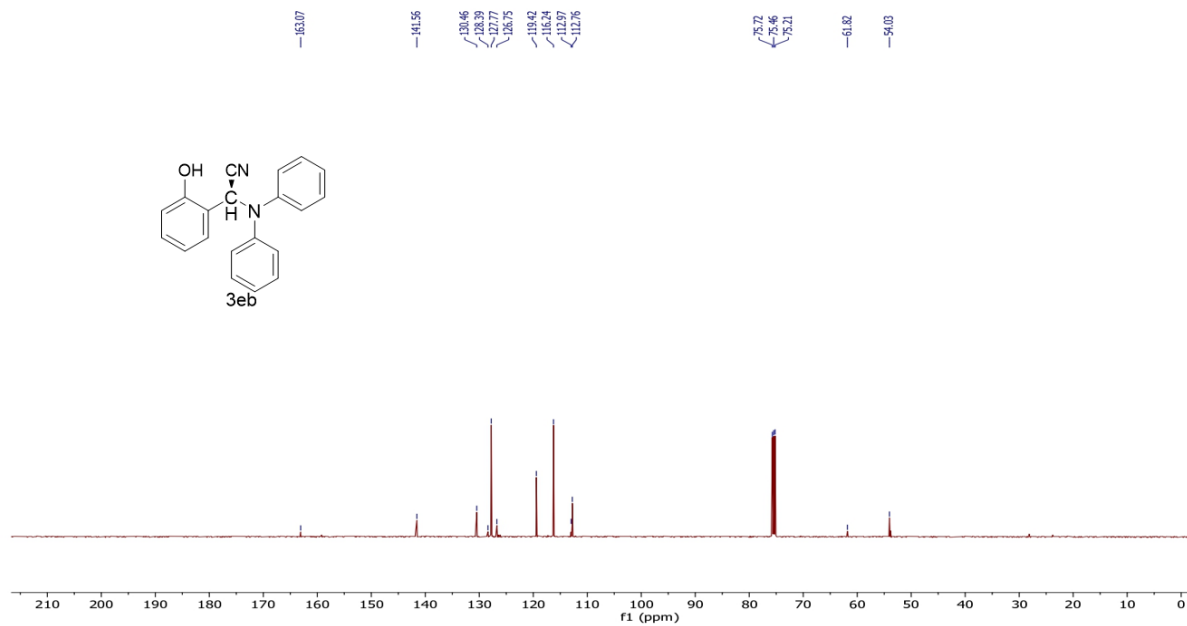
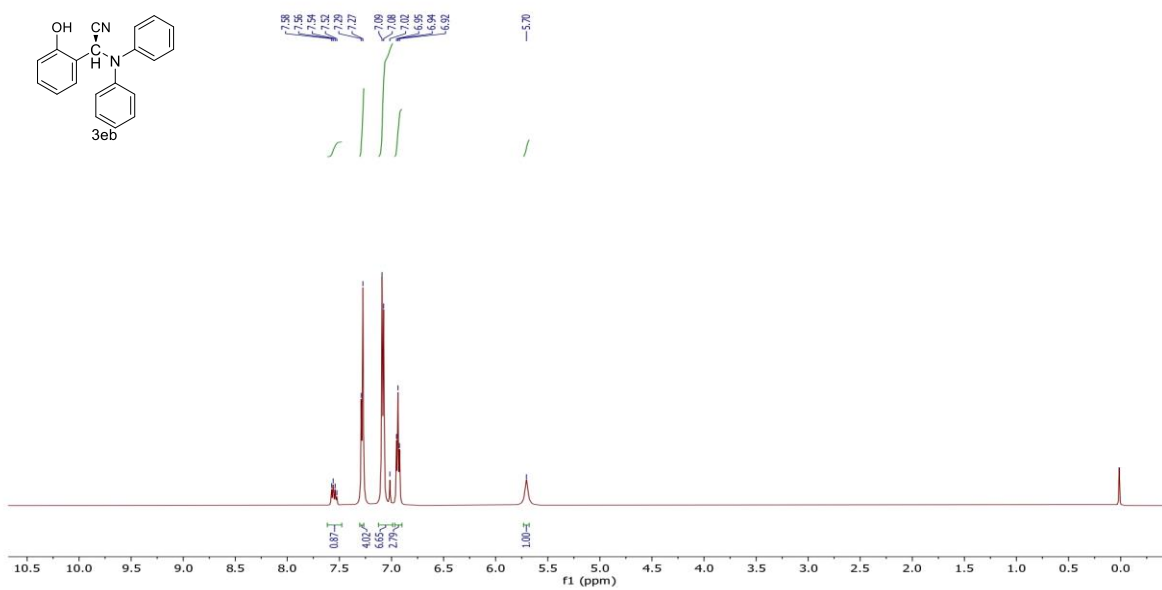
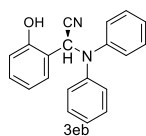
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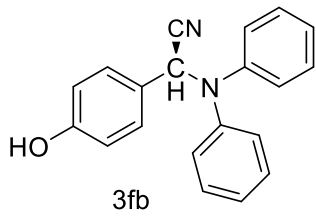
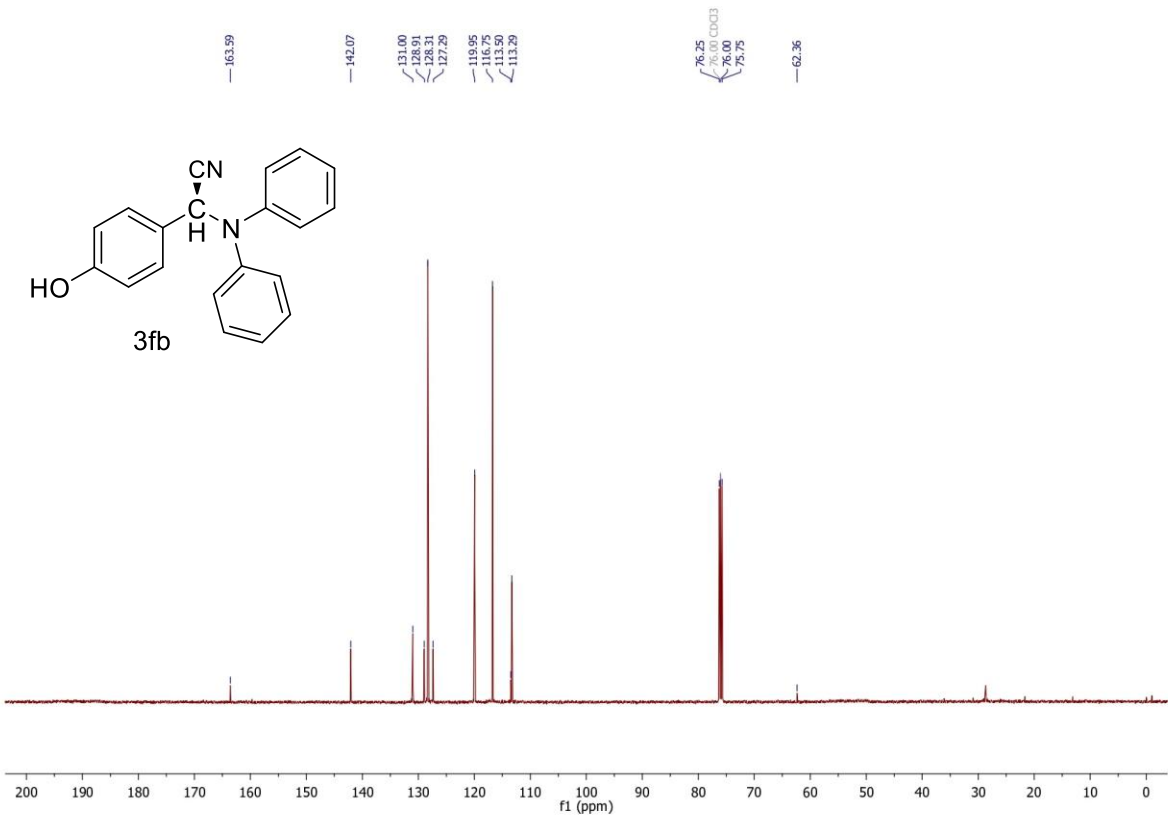
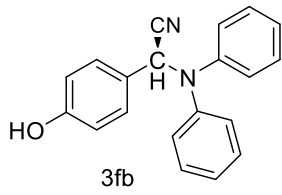
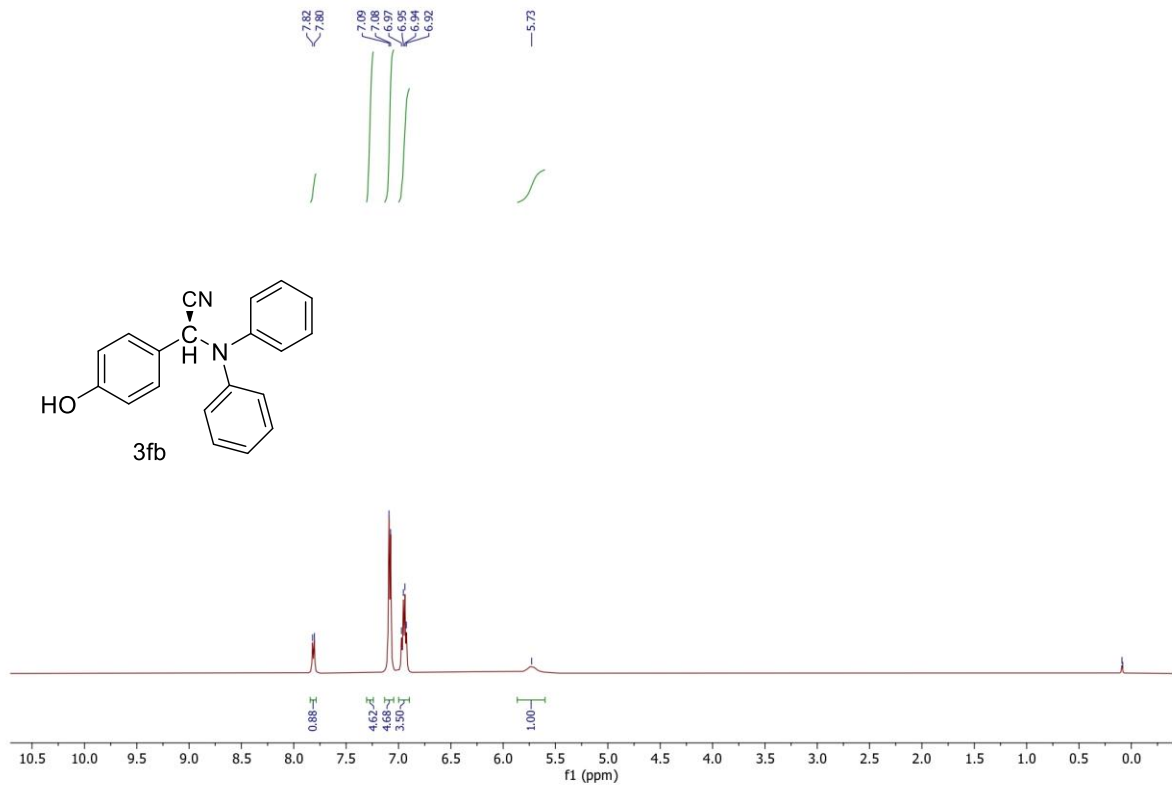
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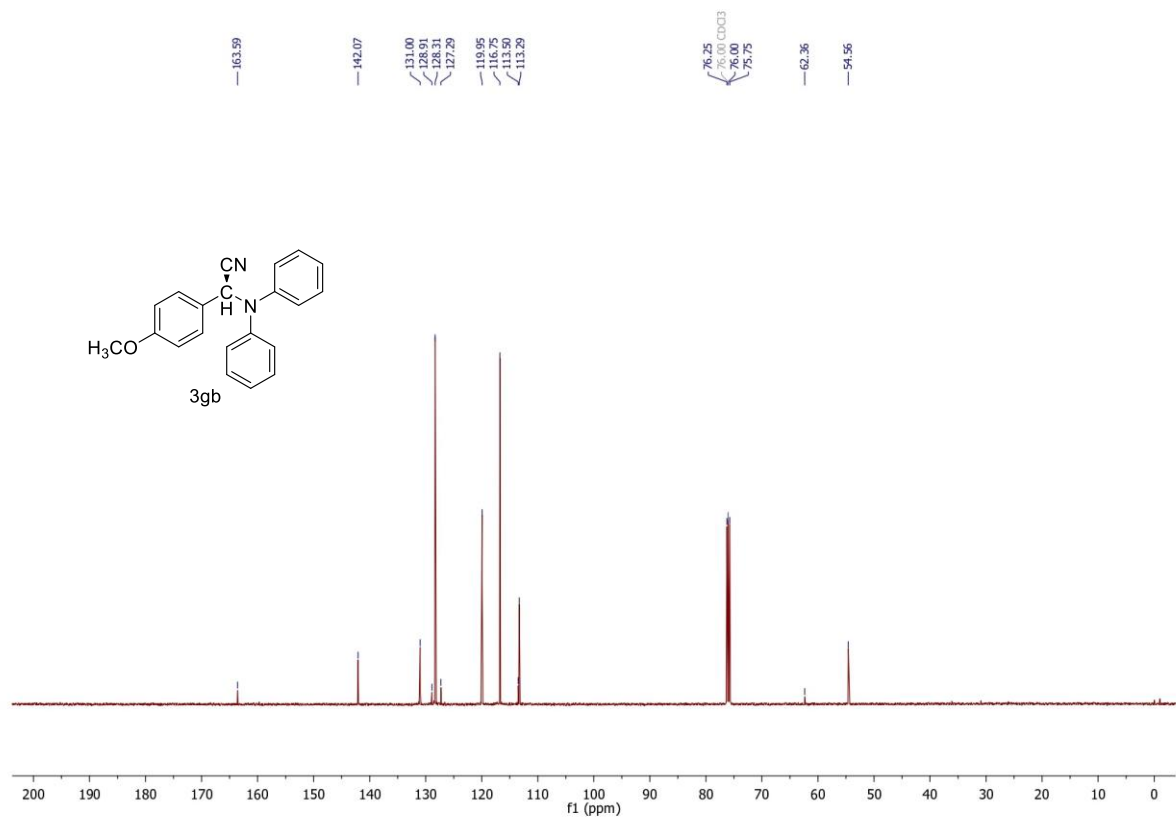
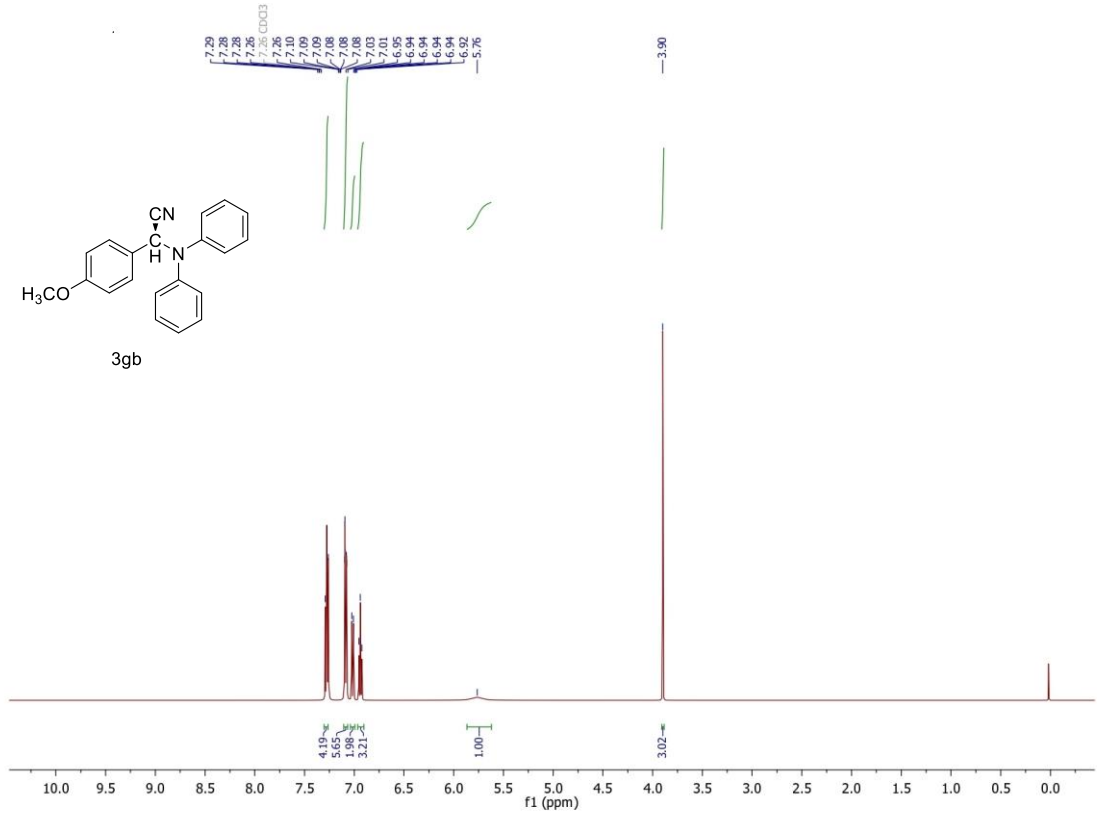
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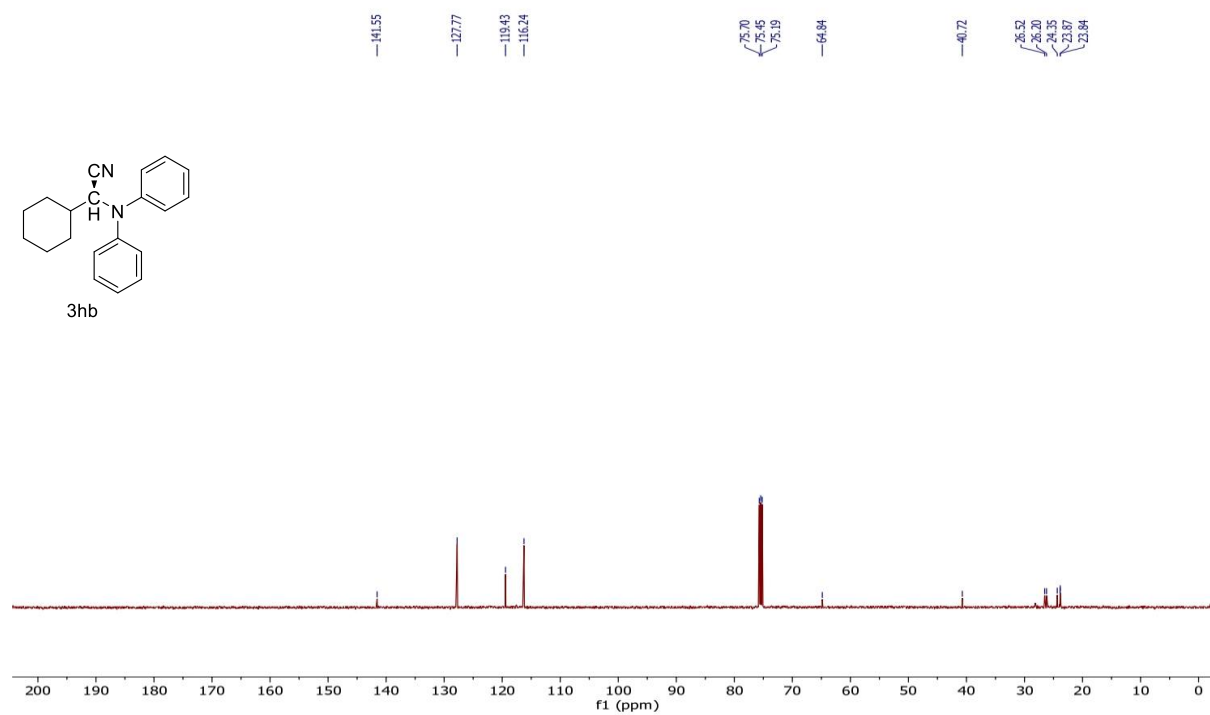
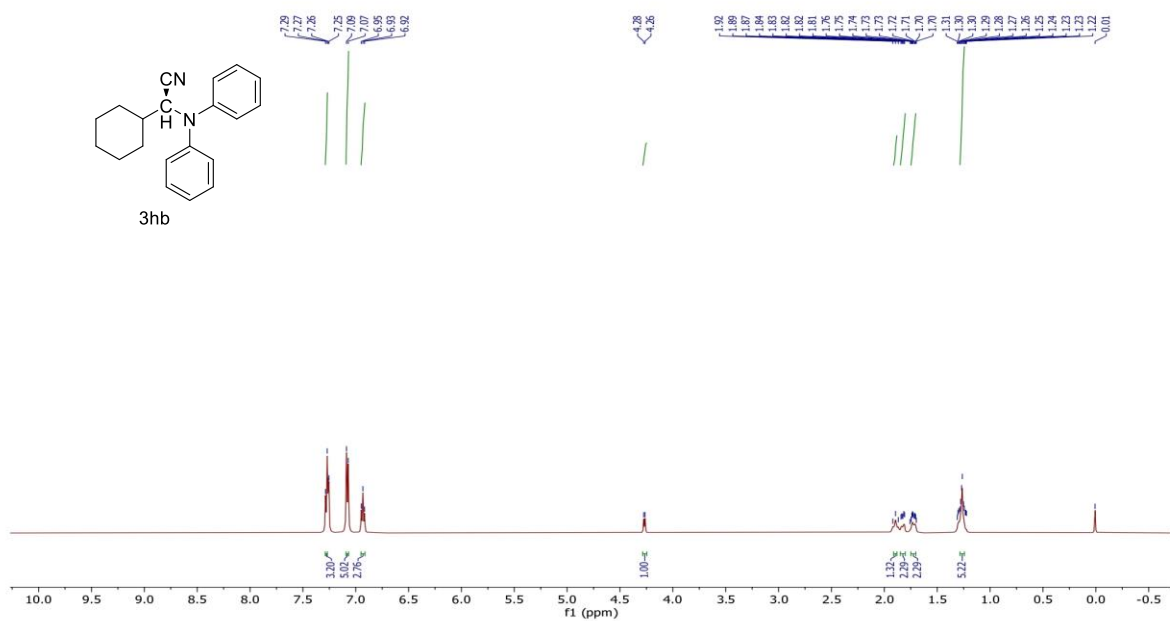
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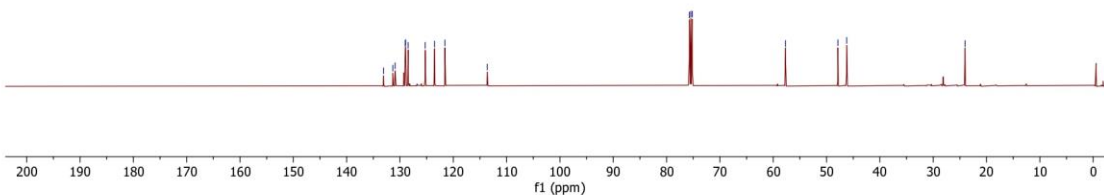
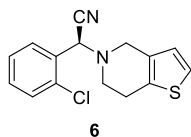
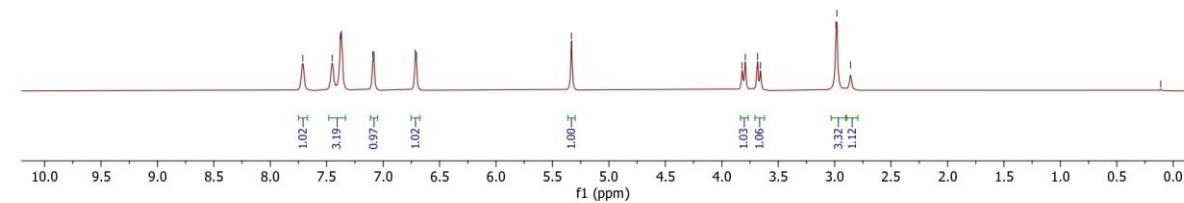
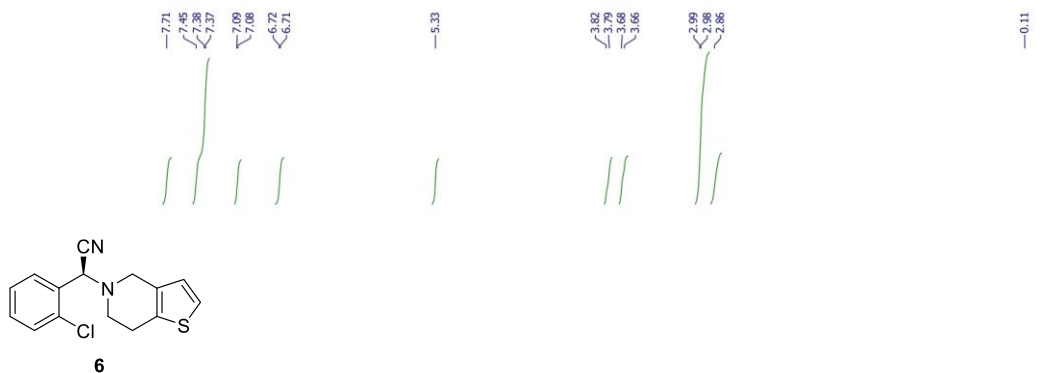
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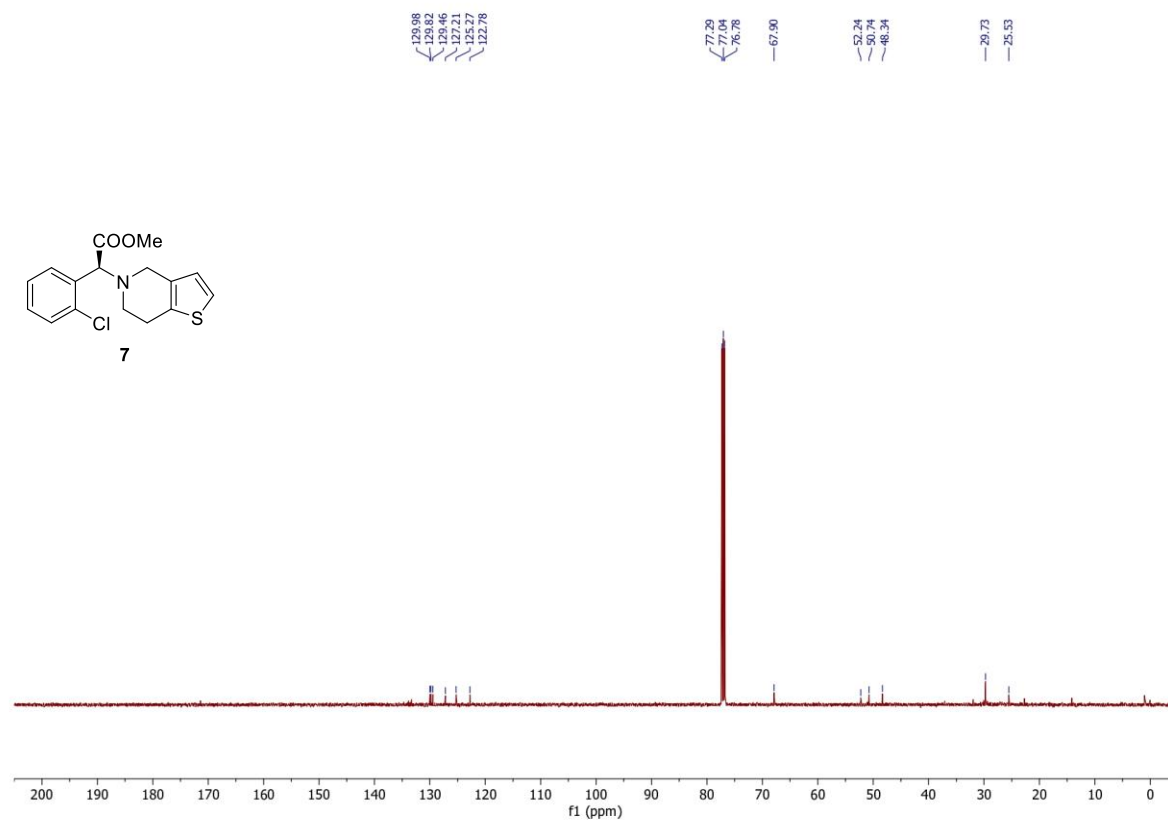
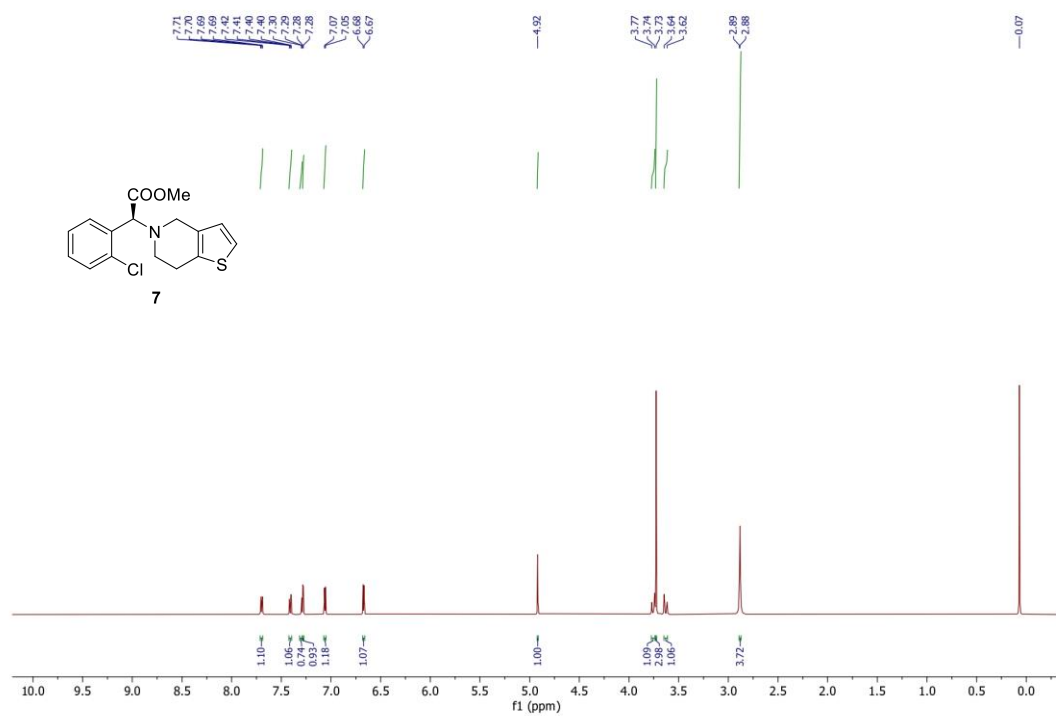
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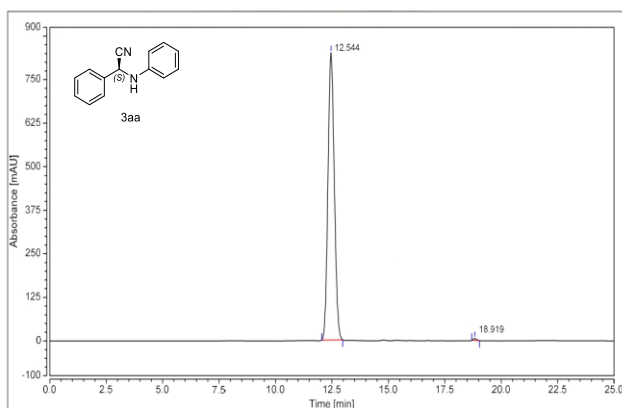
NMR of 6



NMR 7

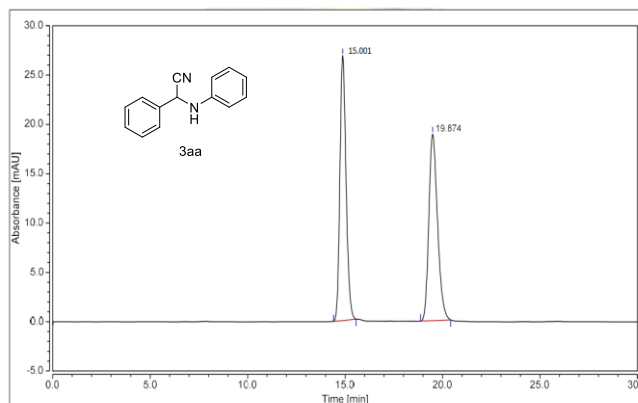


12. HPLC



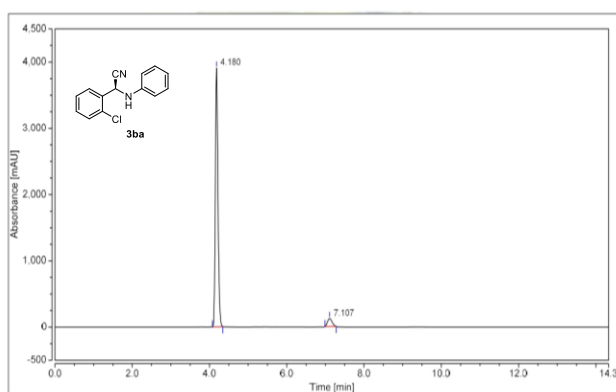
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	12.544	268.550	824.576	99.63	99.32	n.a.
2	18.919	0.996	5.691	0.37	0.68	n.a.
		269.546	830.257	100.00	100.00	



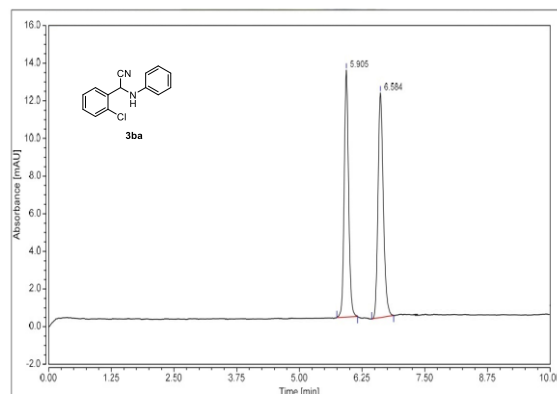
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	15.001	9.830	26.865	50.30	58.65	n.a.
2	19.874	9.714	18.939	49.70	41.35	n.a.
		19.544	45.804	100.00	100.00	



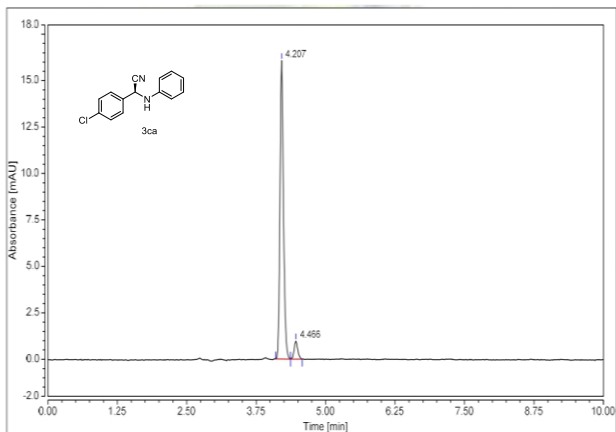
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	4.180	300.203	3903.642	94.80	97.02	n.a.
2	7.107	16.471	119.940	5.20	2.98	n.a.
		316.674	4023.581	100.00	100.00	

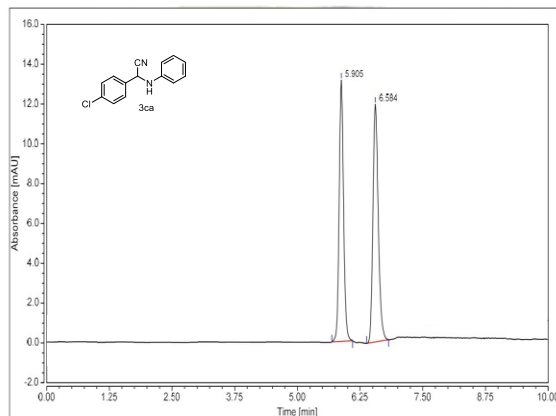


Integration Results

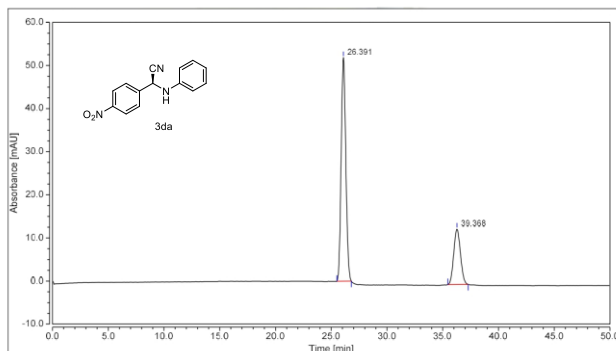
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	6.584	1.470	11.934	52.47	47.63	n.a.
2	5.905	1.332	13.121	47.53	52.37	n.a.
		2.802	25.055	100.00	100.00	



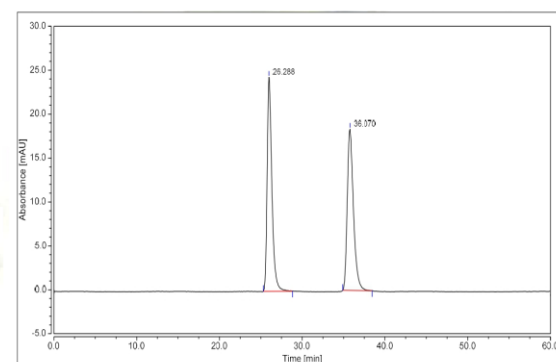
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	4.207	1.184	16.065	94.22	94.28	n.a.
2	4.466	0.073	0.975	5.78	5.72	n.a.
		1.257	17.040	100.00	100.00	



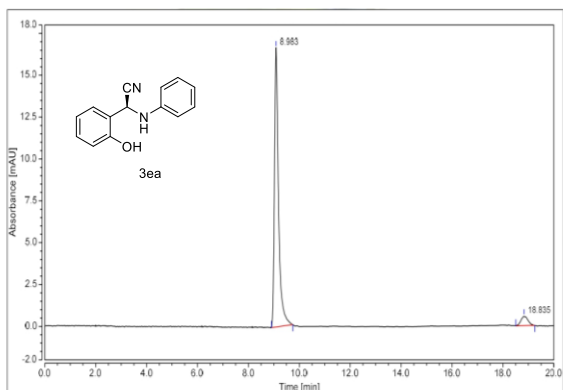
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	5.905	1.470	11.934	52.47	47.63	n.a.
2	6.584	1.332	13.121	47.53	52.37	n.a.
		2.802	25.055	100.00	100.00	



Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	26.391	24.970	51.876	73.53	80.08	n.a.
2	39.368	8.990	12.903	26.47	19.92	n.a.
		33.960	64.778	100.00	100.00	

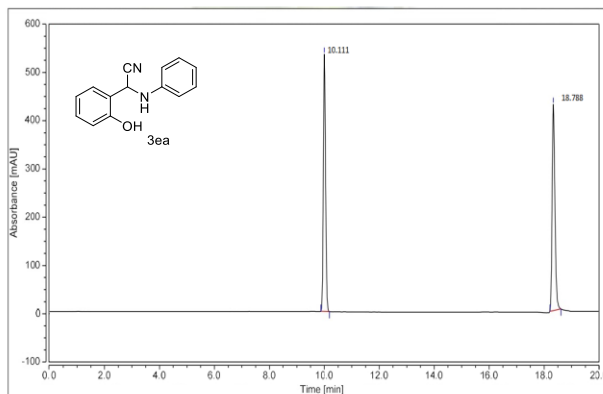


Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	26.288	16.192	24.360	50.47	57.02	n.a.
2	36.070	15.892	18.377	49.53	42.98	n.a.
		32.084	42.737	100.00	100.00	



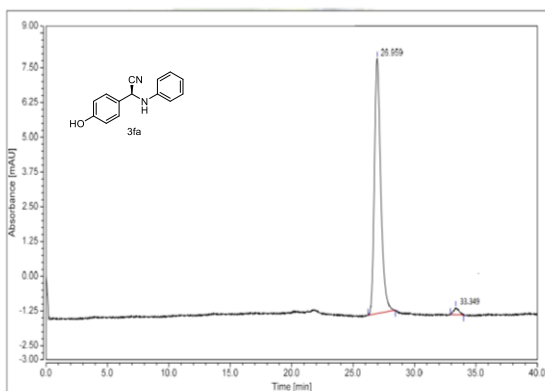
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	8.983	3.084	16.684	94.58	96.64	n.a.
2	18.835	0.177	0.580	5.42	3.36	n.a.
		3.261	17.264	100.00	100.00	



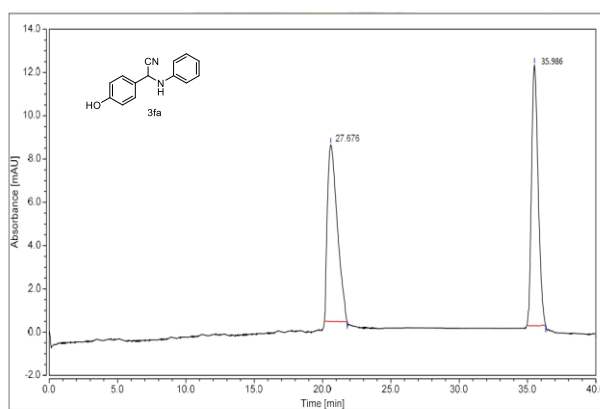
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	10.111	42.543	427.287	50.28	44.50	n.a.
2	18.788	42.074	532.961	49.72	55.50	n.a.
		84.617	960.248	100.00	100.00	



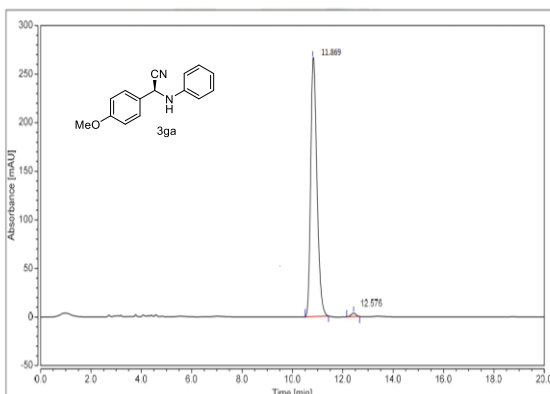
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	26.959	5.522	9.173	97.78	97.38	n.a.
2	33.349	0.125	0.247	2.22	2.62	n.a.
		5.648	9.420	100.00	100.00	



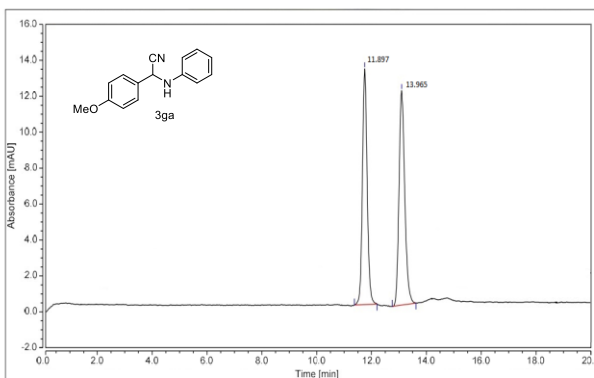
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	27.676	7.063	6.244	51.28	40.70	n.a.
2	35.986	6.730	12.010	48.72	59.30	n.a.
		13.813	20.254	100.00	100.00	



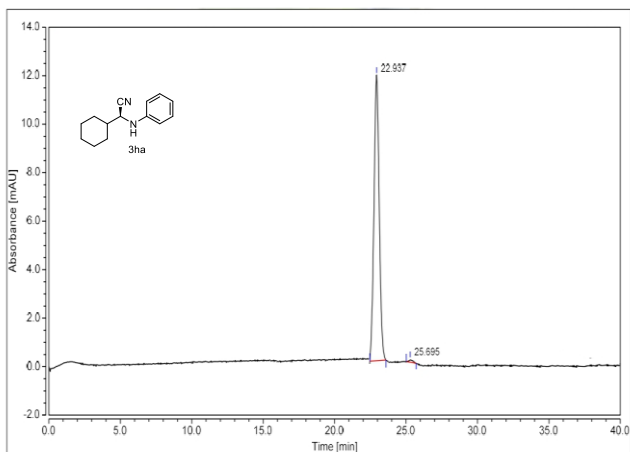
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	11.869	77.155	268.950	98.96	96.72	n.a.
2	12.576	0.808	3.500	1.04	1.28	n.a.
		77.963	272.449	100.00	100.00	



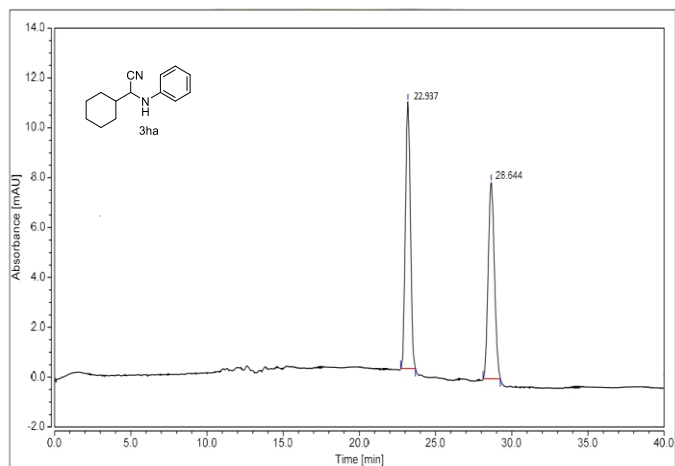
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	11.897	1.470	11.534	62.47	47.63	n.a.
2	13.965	1.332	13.121	47.53	52.37	n.a.
		2.802	25.055	100.00	100.00	



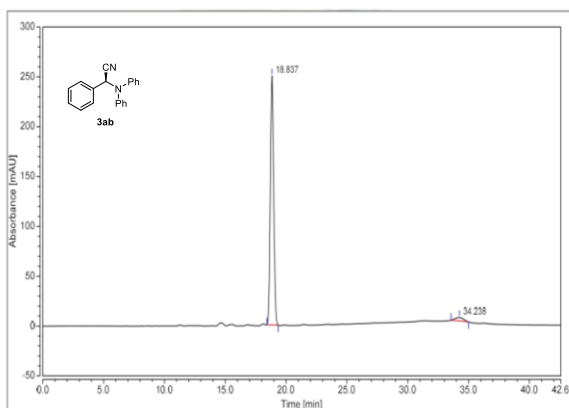
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	22.937	4.974	11.780	99.28	98.92	n.a.
2	25.695	0.036	0.128	0.72	1.08	n.a.
		5.010	11.908	100.00	100.00	



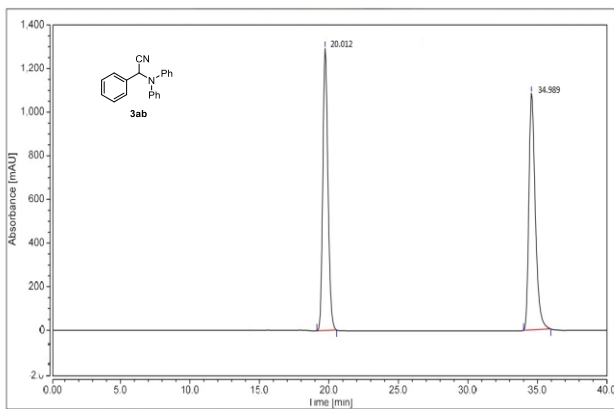
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	22.937	4.007	10.692	51.89	57.57	n.a.
2	28.644	3.715	7.880	48.11	42.43	n.a.
		7.722	18.572	100.00	100.00	



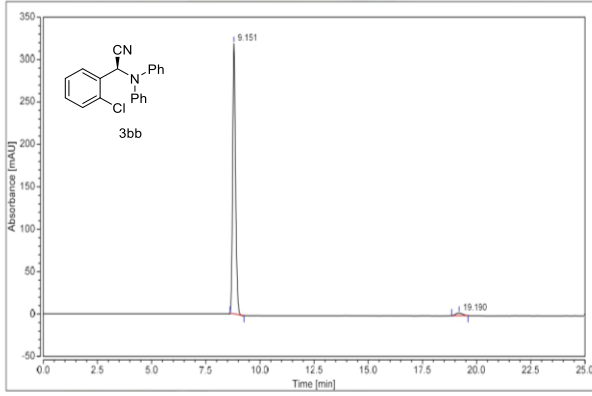
Integration Results

Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	18.837	73.379	249.831	96.58	98.58	n.a.
2	34.238	2.812	3.597	3.42	1.42	n.a.
		82.191	253.427	100.00	100.00	

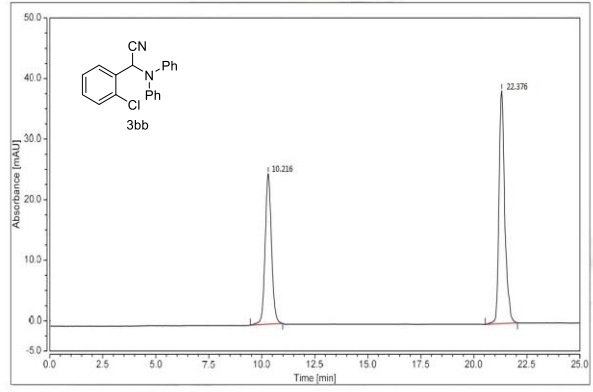


Integration Results

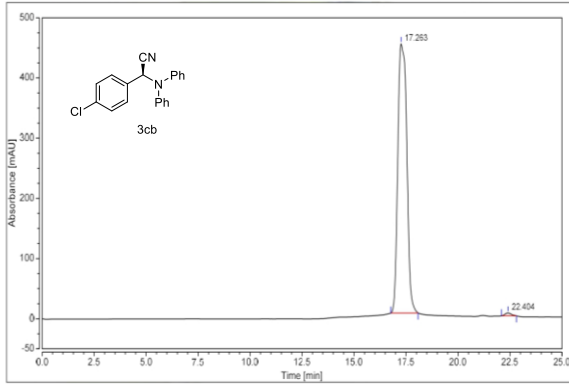
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	20.012	116.913	1084.075	51.17	45.60	n.a.
2	34.989	111.546	1293.323	48.83	54.40	n.a.
		228.459	2377.398	100.00	100.00	



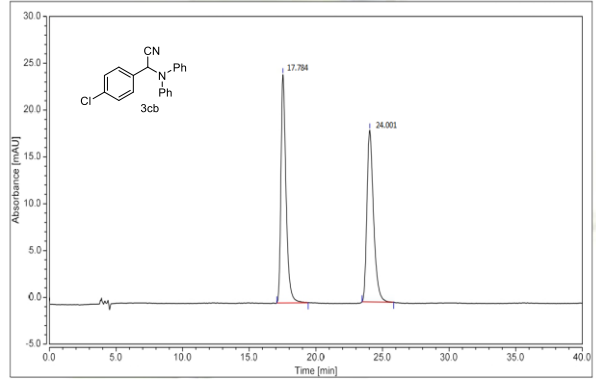
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	9.151	49.735	318.597	97.77	99.07	n.a.
2	19.190	1.135	2.993	2.23	0.93	n.a.
		50.870	321.590	100.00	100.00	



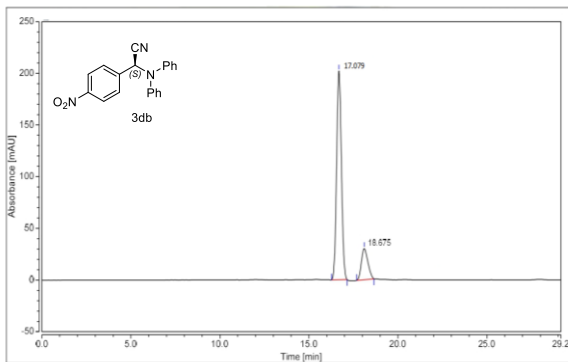
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	10.216	8.639	24.835	50.10	42.03	n.a.
2	22.376	8.605	34.258	49.90	57.97	n.a.
		17.244	59.093	100.00	100.00	



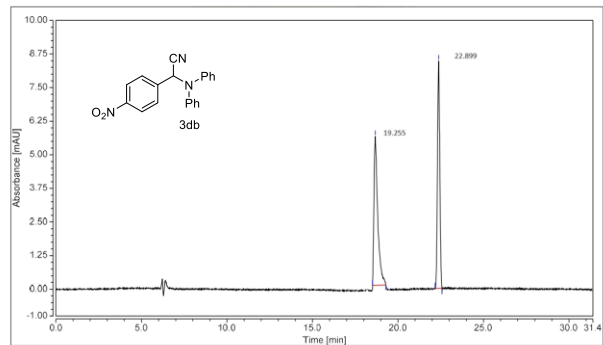
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	17.263	223.887	447.837	99.24	98.92	n.a.
2	22.404	1.703	4.899	0.76	1.08	n.a.
		225.590	452.736	100.00	100.00	



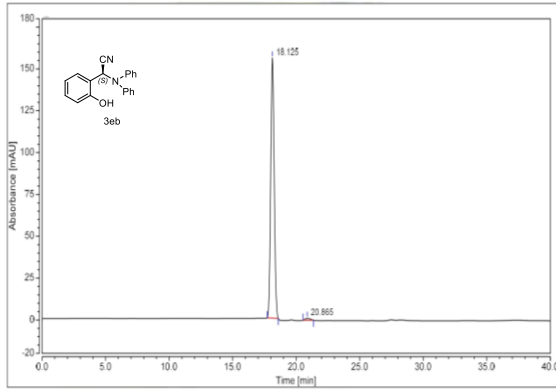
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	17.784	16.192	24.380	50.47	57.02	n.a.
2	24.001	15.892	18.377	49.53	42.98	n.a.
		32.084	42.757	100.00	100.00	



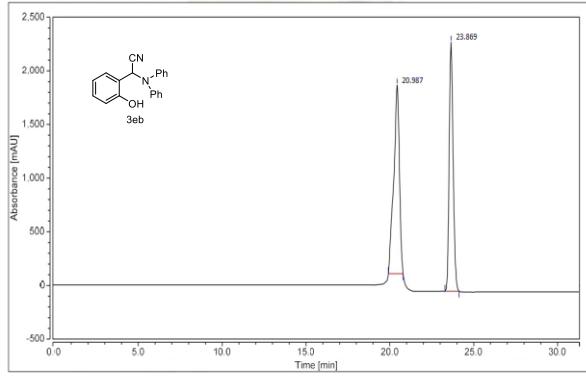
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	17.079	12.811	30.280	83.23	13.04	n.a.
2	18.675	63.581	202.021	16.77	86.96	n.a.
		76.393	232.301	100.00	100.00	



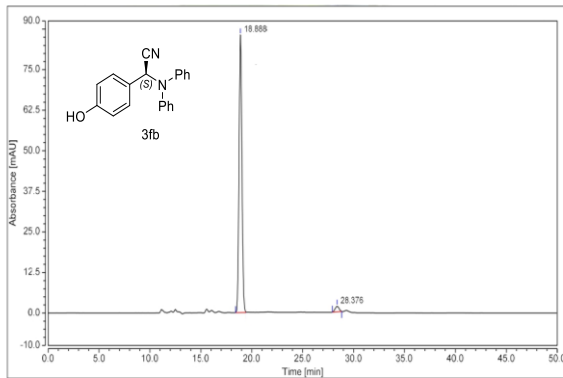
Integration Results						
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	19.255	1.422	5.548	51.44	38.45	n.a.
2	22.899	1.342	8.880	48.56	61.55	n.a.
		2.764	14.428	100.00	100.00	



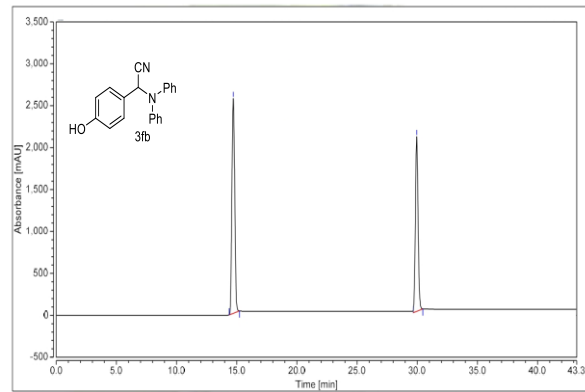
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	18.125	49.142	155.402	99.00	99.28	n.a.
2	20.865	0.498	1.127	1.00	0.72	n.a.
		49.639	156.529	100.00	100.00	



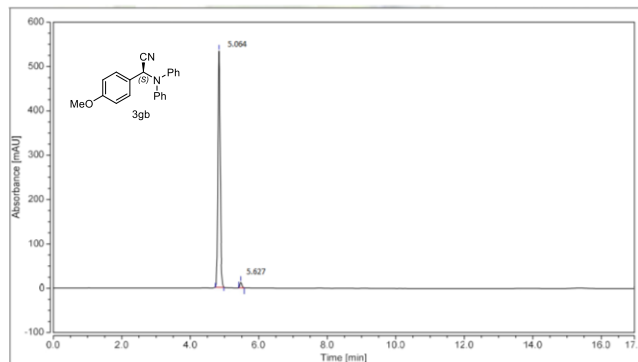
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	20.987	393.531	1757.120	52.01	43.10	n.a.
2	23.869	363.068	2319.919	47.99	56.90	n.a.
		756.599	4077.039	100.00	100.00	



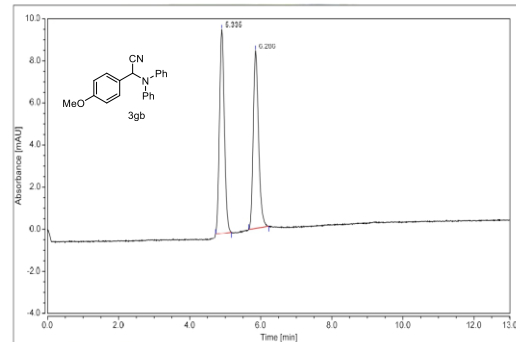
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	18.888	28.123	85.540	97.33	98.01	n.a.
2	28.376	0.771	1.737	2.67	1.99	n.a.
		28.894	87.277	100.00	100.00	



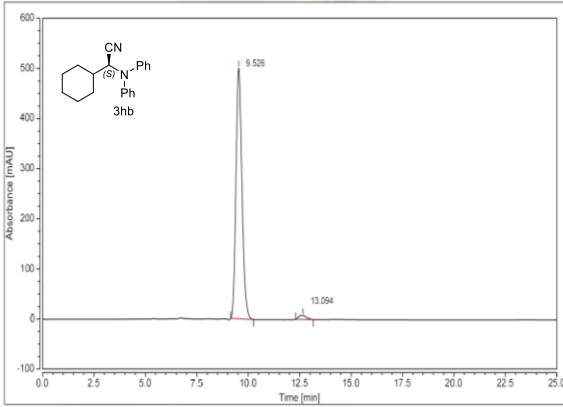
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	19.234	536.389	2084.317	51.29	50.97	n.a.
2	30.115	509.368	2005.074	48.71	49.03	n.a.
		1045.757	4089.391	100.00	100.00	



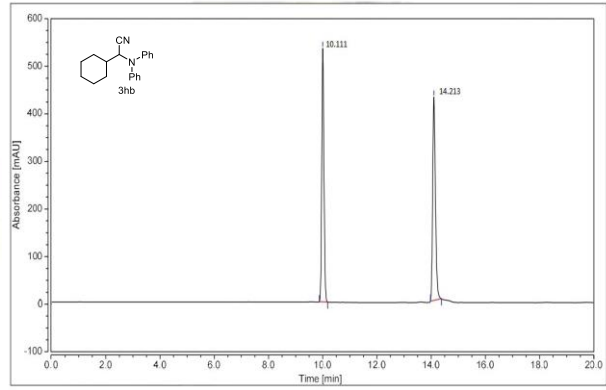
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	5.064	42.011	532.753	96.07	97.76	n.a.
2	5.627	0.828	12.216	1.93	2.24	n.a.
		42.839	544.969	100.00	100.00	



Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	5.335	5.335	1.765	11.009	51.53	n.a.
2	6.286	1.861	9.569	48.47	46.50	n.a.
		3.426	20.578	100.00	100.00	

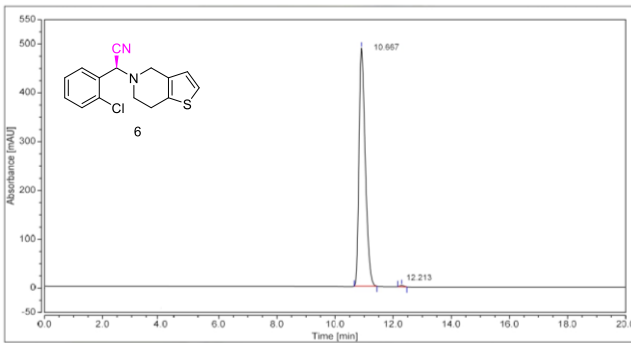


Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	9.526	3.367	7.785	98.05	1.53	n.a.
2	13.094	169.371	500.505	1.95	98.47	n.a.
		172.738	508.290	100.00	100.00	

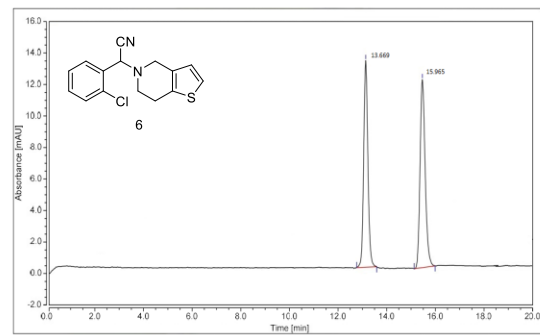


Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	10.111	42.543	427.287	50.28	44.50	n.a.
2	14.213	42.074	532.961	49.72	55.50	n.a.
		84.617	960.248	100.00	100.00	

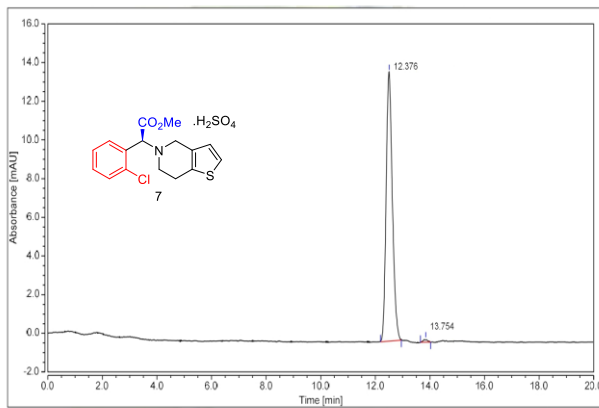
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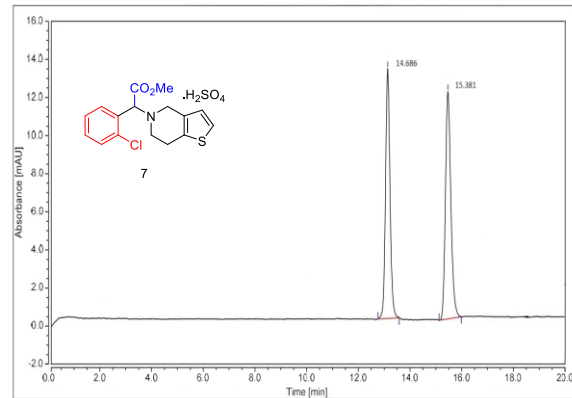
Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	10.667	122.419	487.866	99.67	99.62	n.a.
2	12.213	0.400	2.356	0.33	0.48	n.a.
		122.819	490.222	100.00	100.00	



Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	13.669	1.470	11.934	52.47	47.63	n.a.
2	15.965	1.332	13.121	47.53	52.37	n.a.
		2.802	25.055	100.00	100.00	



Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	12.376	3.673	13.941	99.25	98.84	n.a.
2	13.754	0.028	0.163	0.75	1.16	n.a.
		3.701	14.104	100.00	100.00	



Peak	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount
1	14.686	1.470	11.934	52.47	47.63	n.a.
2	15.381	1.332	13.121	47.53	52.37	n.a.
		2.802	25.055	100.00	100.00	

13. References:

1. C. D. Gutsche, M. I., P-Tert-BUTYLCALIX[4]ARENE. *Organic Synthesis* **1990**, *68*, 234.
2. Jiao, Z.; Feng, X.; Liu, B.; Chen, F.; Zhang, G.; Jiang, Y., Enantioselective Strecker Reactions between Aldimines and Trimethylsilyl Cyanide Promoted by Chiral N,N'-Dioxides. *Eur. J. Org. Chem.* **2003**, *2003*, 3818-3826