

Supporting Information

Electrochemical hydrocarboxylation of enol derivatives with CO₂: access to β-acetoxycarboxylic acids

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General materials and methods

^1H , ^{13}C , ^{19}F NMR spectra were recorded on Bruker AVneo-300 and Bruker Fourier 300 HD spectrometers (300.13, 75.48, 282.5 MHz, respectively) in CDCl_3 and DMSO-d_6 . Chemical shifts were reported in parts per million (ppm), and the residual solvent peak was used as an internal reference: ^1H (CDCl_3 δ = 7.26 ppm), ^{13}C (CDCl_3 δ = 77.16 ppm), ^1H (DMSO-d_6 δ = 2.50 ppm), ^{13}C (DMSO-d_6 δ = 39.52 ppm). Multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), quintet, m (multiplet), brs (broad singlet).¹ 1,4-dinitrobenzene has been used as a standard for determining ^1H NMR yields.

High resolution mass spectra (HR-MS) were measured on a Bruker micrOTOF II instrument using electrospray ionization (ESI).² The measurements were performed in a positive ion mode (interface capillary voltage – 4500 V); mass range from m/z 50 to m/z 3000 Da; external calibration with Electrospray Calibrant Solution (Fluka). A syringe injection was used for all acetonitrile solutions (flow rate 3 $\mu\text{L}/\text{min}$). Nitrogen was applied as a dry gas; interface temperature was set at 180 °C.

IR spectra were recorded on a «InfraLUM FT-08» by Lumex. Method of measurement is ATR.

Elemental analyzes were performed on a PerkinElmer 2400 CHN analyzer. Method of measurement is automatic.

Acetophenone, triethanolamine, Et_3N , K_2CO_3 , pyridine, NaOH , monoethanolamine, imidazole, 1,3-bis(methylamino)propan-2-ol, N,N,N',N' -tetraethylmethanediamine, LiClO_4 , $n\text{-Bu}_4\text{NClO}_4$, $n\text{-Bu}_4\text{NBF}_4$, $n\text{-Bu}_4\text{NBr}$, $n\text{-Bu}_4\text{NI}$, phenanthrene, 1-methylnaphthalene, anthracene, phenantrolin, graphene, *p*-terphenyl, LiOH , PEG, *o*-phenylenediamine, methyl iodide were purchased from commercial sources and were used as is. All solvents were distilled according to standard procedures.³

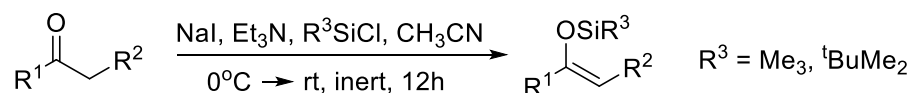
Electrochemical synthesis was carried out using the IKA ElectraSyn 2.0 cell. During the work we used electrodes from the ElectraSyn Starter Kit. At the end of the experiment, all electrodes were washed with water, acetone and then dried. In the case of carbon and platinum electrodes, they were placed in a 1M solution of HCl and allowed to soak for 2 hours, washed with water, acetone and dried in an oven overnight.

The TLC analysis was carried out on standard silica gel chromatography plates (DC-Fertigfolien ALUGRAMR Xtra SIL G/UV254). Column chromatography was performed using silica gel (0.060-0.200 mm, 60 A, CAS 7631-86-9, Acros).

Synthesis of the starting enol derivatives

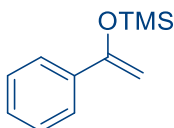
Silyl enol ethers.

Starting silyl enol ethers **S2**, **S3** were prepared according to literature.⁴



NaI (1.4 mmol, 210.0 mg, 1.4 eq.) was placed in a round bottom flask and dried under vacuum (15-20 mmHg) using a heat gun (gun temperature 100-150 °C) for 5 min. After cooling to room temperature, the flask was filled with argon. Then, CH₃CN (1 mL), ketone (1.0 mmol, 1.0 eq.), and Et₃N (1.5 mmol, 151.8 mg, 1.5 eq.) were successively added. The mixture was cooled with an ice/water bath, and R³SiCl (1.3 mmol, 1.3 eq.) was added at 0 °C. The cooling bath was removed, and the mixture was stirred for 12 h at room temperature. Then, volatile components were evaporated under reduced pressure using a rotary evaporator (15-20 mmHg), (bath temperature, ca. 40–45 °C). The solid residue was washed with hexane (3×15 mL) (the hexane layers were decanted and filtered through a cotton plug). The combined filtrates were concentrated under reduced pressure using a rotary evaporator (15-20 mmHg), (bath temperature, ca. 20–25 °C), furnishing the silyl enol ether which was used without purification.

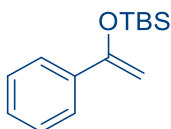
Trimethyl((1-phenylvinyl)oxy)silane, **S2**⁴



¹H NMR (300.13 MHz, CDCl₃, δ): 7.74 – 7.70 (m, 2H), 7.46 – 7.37 (m, 3H), 5.05 – 5.03 (m, 1H), 4.57 – 4.55 (m, 1H), 0.40 (s, 9H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 155.8, 137.6, 128.2, 128.1 (2C), 125.3 (2C), 91.0, 0.1 (3C).

Tert-butyldimethyl((1-phenylvinyl)oxy)silane, **S3**⁵

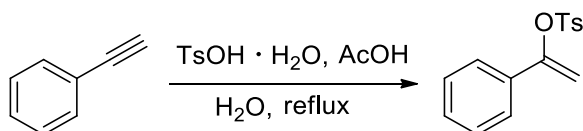


¹H NMR (300.13 MHz, CDCl₃, δ): 7.68 – 7.59 (m, 2H), 7.42 – 7.27 (m, 3H), 4.91 (d, *J* = 1.7 Hz, 1H), 4.44 (d, *J* = 1.7 Hz, 1H), 1.03 (s, 9H), 0.23 (s, 6H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 156.1, 138.0, 128.3, 128.2 (2C), 125.4 (2C), 91.0, 26.1 (3C), 18.5, -4.5 (2C).

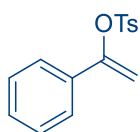
1-phenylvinyl 4-methylbenzenesulfonate.

Starting 1-phenylvinyl 4-methylbenzenesulfonate **S4** was prepared according to literature.⁶



The corresponding alkyne (19.6 mmol, 2.00 g) was added to a solution of *p*-toluenesulfonic acid monohydrate (19.6 mmol, 3.72 g), acetic acid (9.8 mL) and H₂O (3.9 mL). The reaction mixture was stirred and refluxed at 60 °C for 9 hours. The mixture was poured into saturated NaHCO₃ (70 mL), and extracted with CH₂Cl₂ (3 x 30 mL). The organic layer was dried over Na₂SO₄ and concentrated in vacuo. The crude mixture was purified by column chromatography on silica gel. (PE:EA, 20:1).

1-Phenylvinyl 4-methylbenzenesulfonate, **S4**⁶



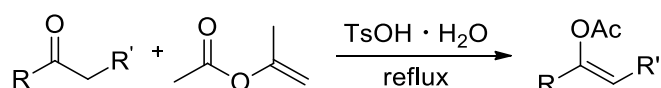
¹H NMR (300.13 MHz, CDCl₃, δ): 7.86 – 7.77 (m, 2H), 7.49 – 7.38 (m, 2H), 7.33 – 7.23 (m, 5H), 5.39 (d, *J* = 2.9 Hz, 1H), 5.10 (d, *J* = 2.9 Hz, 1H), 2.41 (s, 3H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 153.0, 145.3, 133.5, 133.3, 129.8 (2C), 129.4, 128.6 (2C), 128.5 (2C), 125.6 (2C), 103.1, 21.8.

Enol acetates.

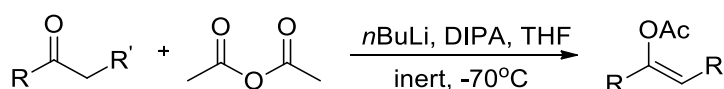
Starting enol acetates **1a-v** were prepared according to literature.⁷⁻⁸

Method 1.⁷



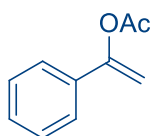
To a mixture of ketone (42.5 mmol) and 2-propenyl acetate (0.213 mol, 5 eq.) was added *p*-toluenesulfonic acid monohydrate (3.925 mmol, 0.09 eq.). The resulting mixture was refluxed in a 100 ml flask equipped with a condenser and a drying tube. The procedure of reaction was controlled by TLC. After achieving the optimal reaction conversion, reaction solution was cooled to room temperature, and the solvent was evaporated in vacuo. Ether was added (100 ml), and the resulting solution was washed with water (3 x 50 ml) and dried over Na₂SO₄. The solvent was evaporated in vacuo to give a dark orange/red oily residue. This residue was purified by column chromatography on silica gel to give target enol acetate.

Method 2.⁸



*n*Butyllithium (9.6 mmol) was added to a solution of diisopropylamine (9.6 mmol) in dried THF (40 mL) in a flame-dried round-bottom flask under inert atmosphere at $-70\text{ }^{\circ}\text{C}$. The mixture was stirred for 30 min, and then ketone (8 mmol) was added. The resulting mixture was stirred for 45 min, and then acetic anhydride (16 mmol) was added. The reaction was stirred 30 min at $-70\text{ }^{\circ}\text{C}$ and another 30 min at room temperature. The mixture was poured into saturated NaHCO_3 (100 mL), and extracted thrice with EtOAc (60 mL). The combined organic layers were washed with 10mL brine and dried over MgSO_4 , and the solvent was removed under reduced pressure. The crude mixture was purified by column chromatography on silica gel.

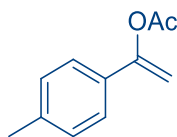
1-Phenylvinyl acetate, **1a**⁹



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.51 – 7.43 (m, 2H), 7.40 – 7.30 (m, 3H), 5.48 (d, $J = 2.0$ Hz, 1H), 5.02 (d, $J = 2.0$ Hz, 1H), 2.28 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.2, 153.1, 134.4, 129.1, 128.6 (2C), 125.0 (2C), 102.2, 21.1.

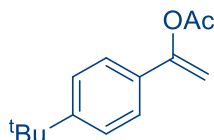
1-(*p*-Tolyl)vinyl acetate, **1b**¹⁰



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.39 – 7.34 (m, 2H), 7.18 – 7.13 (m, 3H), 5.48 (d, $J = 2.2$ Hz, 1H), 4.98 (d, $J = 2.2$ Hz, 1H), 2.35 (s, 3H), 2.27 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.2, 153.2, 139.1, 131.6, 129.3 (2C), 124.9 (2C), 101.3, 21.3, 21.1.

1-(4-(*Tert*-butyl)phenyl)vinyl acetate, **1c**

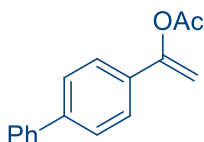


^1H NMR (300.13 MHz, CDCl_3 , δ): 7.43 – 7.35 (m, 4H), 5.43 (d, $J = 2.2$ Hz, 1H), 4.99 (d, $J = 2.2$ Hz, 1H), 2.28 (s, 3H), 1.32 (s, 9H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.3, 153.1, 152.2, 131.5, 125.6 (2C), 124.7 (2C), 101.5, 34.8, 31.3 (3C), 21.1.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{14}\text{H}_{18}\text{O}_2\text{Na}]^+$ 241.1199; Found 241.1203.

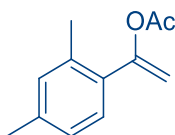
1-([1,1'-Biphenyl]-4-yl)vinyl acetate, **1d**¹¹



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.62 – 7.51 (m, 6H), 7.49 – 7.35 (m, 3H), 5.53 (d, J = 2.3 Hz, 1H), 5.03 (d, J = 2.3 Hz, 1H), 2.31 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.3, 152.8, 141.9, 140.5, 133.3, 129.0 (2C), 127.7, 127.4 (2C), 127.2 (2C), 125.4 (2C), 102.3, 21.2.

1-(2,4-Dimethylphenyl)vinyl acetate, 1e

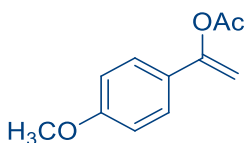


^1H NMR (300.13 MHz, CDCl_3 , δ): 7.29 – 7.24 (m, 1H), 7.02 – 6.95 (m, 2H), 5.15 (d, J = 1.6 Hz, 1H), 5.00 (d, J = 1.6 Hz, 1H), 2.38 (s, 3H), 2.31 (s, 3H), 2.13 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 168.9, 154.0, 138.8, 135.9, 132.7, 129.2, 126.5, 126.5, 105.4, 21.3, 21.2, 20.4.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{12}\text{H}_{14}\text{O}_2\text{NH}_4]^+$ 208.1332; Found 208.1338.

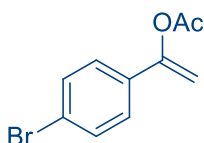
1-(4-Methoxyphenyl)vinyl acetate, 1f⁹



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.43 – 7.36 (m, 2H), 6.90 – 6.84 (m, 2H), 5.35 (d, J = 2.2 Hz, 1H), 4.91 (d, J = 2.2 Hz, 1H), 3.81 (s, 3H), 2.26 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.3, 160.3, 152.9, 129.3, 126.4 (2C), 114.0 (2C), 100.4, 55.5, 21.1.

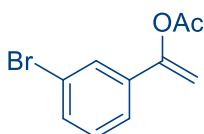
1-(4-Bromophenyl)vinyl acetate, 1r¹²



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.51 – 7.44 (m, 2H), 7.36 – 7.29 (m, 2H), 5.45 (d, J = 2.4 Hz, 1H), 5.05 (d, J = 2.4 Hz, 1H), 2.27 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.0, 152.1, 133.4, 131.8 (2C), 126.6 (2C), 123.2, 102.9, 21.1.

1-(3-Bromophenyl)vinyl acetate, 1s

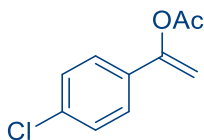


^1H NMR (300.13 MHz, CDCl_3 , δ): 7.61 – 7.57 (m, 1H), 7.48 – 7.36 (m, 2H), 7.25 – 7.17 (m, 1H), 5.47 (d, $J = 2.4$ Hz, 1H), 5.07 (d, $J = 2.4$ Hz, 1H), 2.28 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.0, 151.6, 136.6, 132.0, 130.2, 128.1, 123.7, 122.9, 103.6, 21.1.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{10}\text{H}_9\text{BrO}_2\text{Na}]^+$ 262.9678; Found 262.9683.

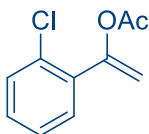
1-(4-Chlorophenyl)vinyl acetate, **1u**¹³



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.42 – 7.36 (m, 2H), 7.34 – 7.28 (m, 2H), 5.45 (d, $J = 2.3$ Hz, 1H), 5.05 (d, $J = 2.3$ Hz, 1H), 2.27 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.0, 152.1, 135.0, 133.0, 128.9 (2C), 126.3 (2C), 102.8, 21.1.

1-(2-Chlorophenyl)vinyl acetate, **1v**

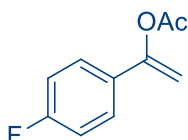


^1H NMR (300.13 MHz, CDCl_3 , δ): 7.48 – 7.43 (m, 1H), 7.40 – 7.35 (m, 1H), 7.28 – 7.23 (m, 2H), 5.27 (d, $J = 1.9$ Hz, 1H), 5.23 (d, $J = 1.9$ Hz, 1H), 2.17 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 168.9, 151.5, 134.6, 132.2, 130.9, 130.2, 130.0, 126.8, 107.2, 21.1.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{10}\text{H}_9\text{ClO}_2\text{Na}]^+$ 219.0183; Found 219.0188.

1-(4-Fluorophenyl)vinyl acetate, **1g**¹⁴

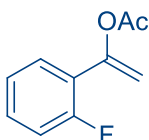


^1H NMR (300.13 MHz, CDCl_3 , δ): 7.48 – 7.40 (m, 2H), 7.07 – 6.98 (m, 2H), 5.40 (d, $J = 2.2$ Hz, 1H), 5.01 (d, $J = 2.2$ Hz, 1H), 2.27 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.1, 163.2 (d, $^1J_{\text{CF}} = 248.8$ Hz), 152.2, 131.1 (d, $^3J_{\text{CF}} = 9.3$ Hz), 130.7 (d, $^4J_{\text{CF}} = 3.4$ Hz), 126.9 (d, $^3J_{\text{CF}} = 8.3$ Hz, 2C), 115.7 (d, $^2J_{\text{CF}} = 21.9$ Hz, 2C), 102.1 (d, $^6J_{\text{CF}} = 1.6$ Hz), 21.1.

^{19}F NMR (282.5 MHz, CDCl_3 , δ): -112.35 (tt, $J = 8.4, 5.2$ Hz).

1-(2-Fluorophenyl)vinyl acetate, **1h**



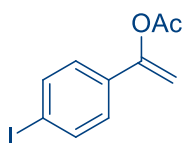
^1H NMR (300.13 MHz, CDCl_3 , δ): 7.41 – 7.24 (m, 2H), 7.16 – 7.02 (m, 2H), 7.39 (td, $J = 7.7, 1.7$ Hz, 1H), 7.32 (tdd, $J = 7.6, 5.1, 1.8$ Hz, 1H), 7.15 (td, $J = 7.6, 1.2$ Hz, 1H), 7.10 (ddd, $J = 11.5, 8.1, 0.9$ Hz, 1H), 5.56 (d, $J = 1.9$ Hz, 1H), 5.25 (t, $J = 1.9$ Hz, 1H), 2.24 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.1, 160.0 (d, $^1J_{\text{CF}} = 251.9$ Hz), 148.0 (d, $^3J_{\text{CF}} = 3.3$ Hz), 130.4 (d, $^3J_{\text{CF}} = 8.6$ Hz), 128.1 (d, $^3J_{\text{CF}} = 2.7$ Hz), 124.2 (d, $^4J_{\text{CF}} = 3.7$ Hz), 122.6 (d, $^2J_{\text{CF}} = 11.2$ Hz), 116.5 (d, $^2J_{\text{CF}} = 22.7$ Hz), 107.5 (d, $^4J_{\text{CF}} = 9.0$ Hz), 21.0.

^{19}F NMR (282.5 MHz, CDCl_3 , δ): -113.42 (dddd, $J = 11.3, 7.2, 5.1, 1.7$ Hz)

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{10}\text{H}_9\text{FO}_2\text{Na}]^+$ 203.0479; Found 203.0475.

1-(4-Iodophenyl)vinyl acetate, 1t

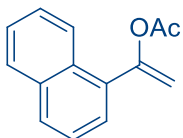


^1H NMR (300.13 MHz, CDCl_3 , δ): 7.71 – 7.64 (m, 2H), 7.23 – 7.15 (m, 2H), 5.47 (d, $J = 2.3$ Hz, 1H), 5.04 (d, $J = 2.3$ Hz, 1H), 2.26 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.1, 152.2, 137.8 (2C), 134.0, 126.7 (2C), 103.0, 94.9, 21.1.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{10}\text{H}_9\text{IO}_2\text{Na}]^+$ 310.9539; Found 310.9534.

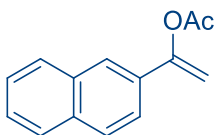
1-(Naphthalen-1-yl)vinyl acetate, 1i¹⁵



^1H NMR (300.13 MHz, CDCl_3 , δ): 8.30 – 8.20 (m, 1H), 7.91 – 7.79 (m, 2H), 7.65 – 7.59 (m, 1H), 7.57 – 7.41 (m, 3H), 5.36 (d, $J = 1.5$ Hz, 1H), 5.24 (d, $J = 1.5$ Hz, 1H), 2.13 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.1, 153.5, 133.7, 133.7, 130.9, 129.6, 128.5, 127.4, 126.6, 126.1, 125.6, 125.2, 107.0, 21.2.

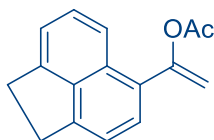
1-(Naphthalen-2-yl)vinyl acetate, 1j⁹



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.93 – 7.77 (m, 4H), 7.66 – 7.56 (m, 1H), 7.54 – 7.45 (m, 2H), 5.62 (d, $J = 2.3$ Hz, 1H), 5.13 (d, $J = 2.3$ Hz, 1H), 2.35 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.3, 153.1, 133.6, 133.2, 131.7, 128.6, 128.5, 127.8, 126.8, 126.6, 124.2, 122.8, 102.8, 21.2.

1-(1,2-Dihydroacenaphthylen-5-yl)vinyl acetate, 1k

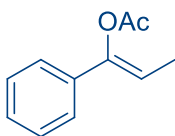


^1H NMR (300.13 MHz, DMSO- d_6 , δ): 7.91 – 7.84 (m, 1H), 7.59 – 7.46 (m, 2H), 7.37 – 7.26 (m, 2H), 5.33 (d, J = 1.6 Hz, 1H), 5.27 (d, J = 1.6 Hz, 1H), 3.37 – 3.31 (m, 4H), 2.15 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.1, 153.5, 147.8, 146.4, 139.5, 129.1, 129.0, 128.6, 128.1, 120.7, 119.7, 118.9, 105.5, 30.6, 30.3, 21.2.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{16}\text{H}_{14}\text{O}_2\text{Na}]^+$ 261.0886; Found 261.0888.

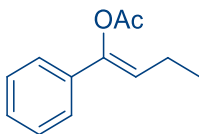
(Z)-1-Phenylprop-1-en-1-yl acetate, 1l⁷



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.43 – 7.23 (m, 5H), 5.90 (q, J = 7.0 Hz, 1H), 2.31 (s, 3H), 1.73 (d, J = 7.0 Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 168.7, 147.1, 135.1, 128.6 (2C), 128.1, 124.4 (2C), 112.8, 20.7, 11.7.

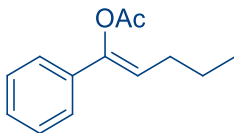
(Z)-1-Phenylbut-1-en-1-yl acetate, 1m⁸



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.44 – 7.22 (m, 5H), 5.82 (t, J = 7.5 Hz, 1H), 2.29 (s, 3H), 2.15 (quintet, 2H), 1.08 (t, J = 7.5 Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 168.9, 145.7, 135.1, 128.6 (2C), 128.2, 124.5 (2C), 119.9, 20.8, 19.8, 13.6.

(Z)-1-Phenylpent-1-en-1-yl acetate, 1n

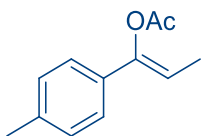


^1H NMR (300.13 MHz, CDCl_3 , δ): 7.44 – 7.23 (m, 5H), 5.83 (t, J = 7.4 Hz, 1H), 2.30 (s, 3H), 2.11 (q, J = 7.4 Hz, 2H), 1.50 (quintet, J = 7.4 Hz, 2H), 0.97 (t, J = 7.4 Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 168.9, 146.3, 135.2, 128.6 (2C), 128.1, 124.5 (2C), 118.3, 28.4, 22.3, 20.7, 14.0.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{13}\text{H}_{16}\text{O}_2\text{Na}]^+$ 227.1043; Found 227.1042.

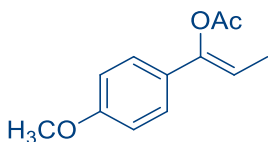
(Z)-1-(p-Tolyl)prop-1-en-1-yl acetate, 1o⁸



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.31 – 7.24 (m, 2H), 7.17 – 7.05 (m, 2H), 5.84 (q, J = 7.0 Hz, 1H), 2.33 (s, 3H), 2.29 (s, 3H), 1.70 (d, J = 7.0 Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 168.8, 147.2, 138.0, 132.4, 129.3 (2C), 124.3 (2C), 111.8, 21.3, 20.8, 11.6.

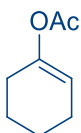
(Z)-1-(4-Methoxyphenyl)prop-1-en-1-yl acetate, 1p⁸



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.43 – 7.28 (m, 2H), 6.90 – 6.79 (m, 2H), 5.76 (q, J = 7.0 Hz, 1H), 3.80 (s, 3H), 2.30 (s, 3H), 1.69 (d, J = 7.0 Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 168.9, 159.6, 146.8, 127.9, 125.7 (2C), 114.0 (2C), 110.8, 55.4, 20.8, 11.6.

Cyclohex-1-en-1-yl acetate, 1q¹⁶



^1H NMR (300.13 MHz, CDCl_3 , δ): 5.37 – 5.30 (m, 1H), 2.16 – 2.04 (m, 7H), 1.76 – 1.65 (m, 2H), 1.62 – 1.52 (m, 2H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 169.6, 148.5, 114.1, 26.9, 23.7, 22.7, 21.8, 21.2.

Reaction setup.



Picture S1. Electrochemical reaction setup

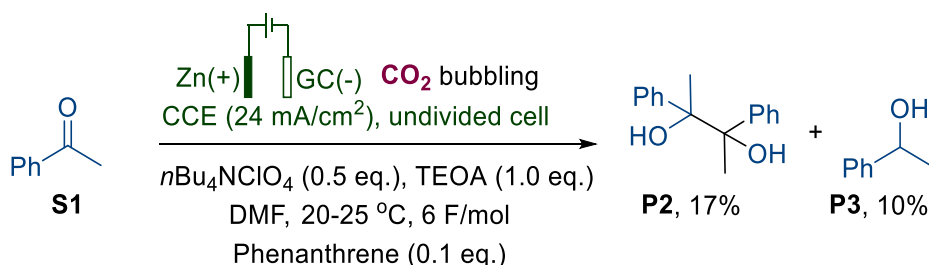


Picture S2. Typical mixture during a reaction

General Experimental Procedure for Scheme 2a.

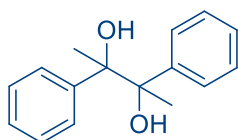
A 5 mL electrolysis cell IKA ElectraSyn 2.0 containing a glassy carbon cathode and a zinc anode (30 mm × 7 mm × 0.1 mm) was charged with a solution of enol derivative **S** (1.0 mmol), $n\text{Bu}_4\text{NClO}_4$ (0.5 mmol), TEOA (1.0 mmol) and phenanthrene (0.1 eq.) in DMF (3 mL). The resulting mixture was bubbled with CO_2 (at a flow rate of 2 mL/min) and electrolysed at a constant current of 50 mA ($j = 24 \text{ mA/cm}^2$) with stirring at 20–25 °C for 194 min. Then, aq. HCl (5 M, 4 mL) was added to the reaction mixture. Extraction with PE/EtOAc (1:1, 5 × 5 mL) afforded the organic phase, which was washed with 10 mL brine and dried over Na_2SO_4 , filtered and concentrated under reduced pressure using a rotary evaporator (15–20 mmHg), (bath temperature, ca. 20–25 °C). The crude mixture was purified by column chromatography on silica gel.

Electrolysis of S1



Products **P2** (17%, 40 mg, 0.17 mmol) and **P3** (10%, 12 mg, 0.10 mmol) were isolated by chromatography on SiO_2 (PE:EA = from 8:1 to 1:1).

2,3-Diphenylbutane-2,3-diol, **P2**¹⁷



White powder, m.p. = 128–130 °C (lit. mp = 127–130 °C).¹⁷

The mixture of DL and meso isomers (ratio = 2/1).

DL isomer¹⁷:

$^1\text{H NMR}$ (300.13 MHz, CDCl_3 , δ): 7.34 – 7.13 (m, 10H), 2.61 (s, 2H), 1.54 (s, 6H).

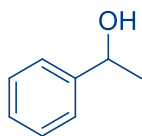
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 143.5 (2C), 127.5 (4C), 127.3 (4C), 127.2 (2C), 79.0 (2C), 25.2 (2C).

Meso isomer¹⁷:

$^1\text{H NMR}$ (300.13 MHz, CDCl_3 , δ): 7.34 – 7.13 (m, 10H), 2.31 (s, 2H), 1.59 (s, 6H)

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 143.9 (2C), 127.4 (4C), 127.1 (4C), 127.0 (2C), 78.7 (2C), 25.2 (2C).

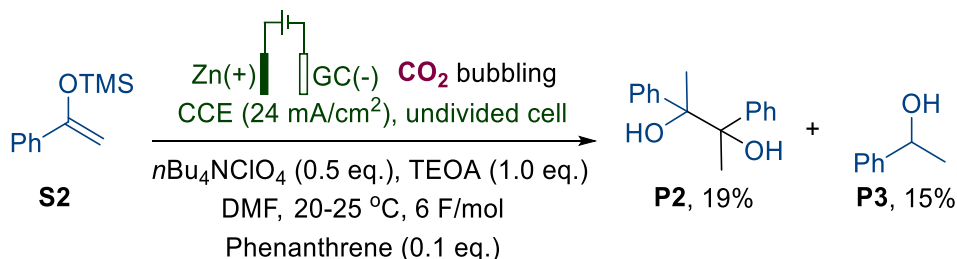
1-Phenylethanol, **P3**¹⁸



^1H NMR (300.13 MHz, CDCl_3 , δ): 7.43 – 7.20 (m, 5H), 4.87 (q, J = 6.4 Hz, 1H), 1.49 (d, J = 6.4 Hz, 3H).

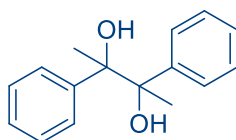
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 145.9, 128.6 (2C), 127.5, 125.5 (2C), 70.4, 25.2.

Electrolysis of S2



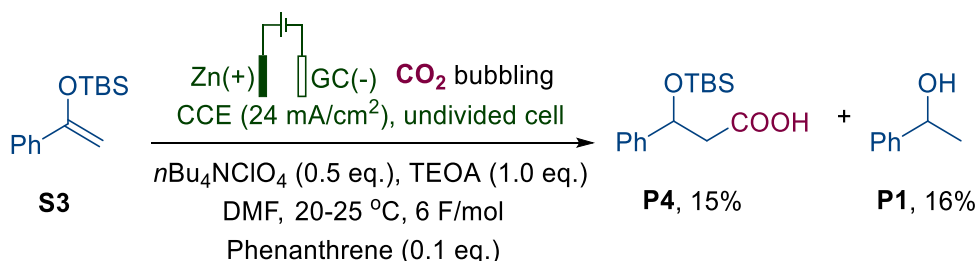
Products **P2** (19%, 45 mg, 0.19 mmol) and **P3** (15%, 18 mg, 0.15 mmol) were isolated by chromatography on SiO_2 (PE:EA = from 5:1 to 1:1).

2,3-Diphenylbutane-2,3-diol, **P2**¹⁷



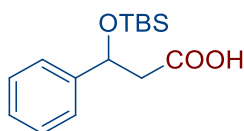
The mixture of DL and meso isomers (ratio = 2/1).

Electrolysis of S3



Products **P4** (15%, 41 mg, 0.15 mmol) and **P1** (16%, 20 mg, 0.16 mmol) were isolated by chromatography on SiO_2 (PE:EA = from 4:1 to 1:1).

3-((*Tert*-butyldimethylsilyloxy)-3-phenylpropanoic acid, **P4**¹⁹



Colorless oil.

^1H NMR (300.13 MHz, CDCl_3 , δ): 10.68 (brs, 1H), 7.40 – 7.22 (m, 5H), 5.15 (dd, J = 8.8, 4.2 Hz, 1H), 2.77 (dd, J = 15.0, 8.8 Hz, 1H), 2.64 (dd, J = 15.0, 4.2 Hz, 1H), 0.86 (s, 9H), 0.04 (s, 3H), -0.16 (s, 3H).

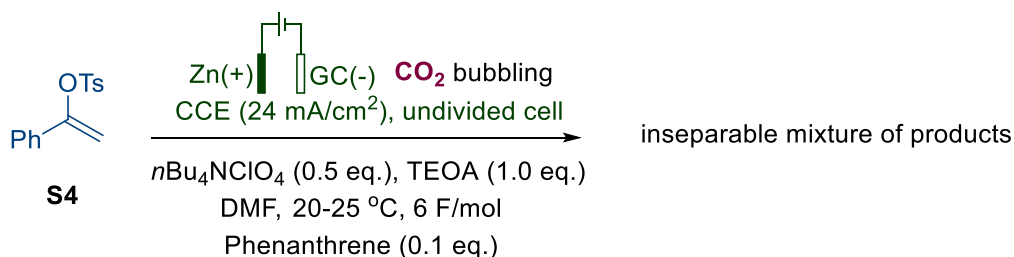
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 176.4, 143.6, 128.5 (2C), 127.9, 125.9 (2C), 72.1, 46.0, 25.8 (3C), 18.2, -4.6, -5.2.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{15}\text{H}_{24}\text{O}_3\text{SiNa}]^+$ 303.1387; Found 303.1387.

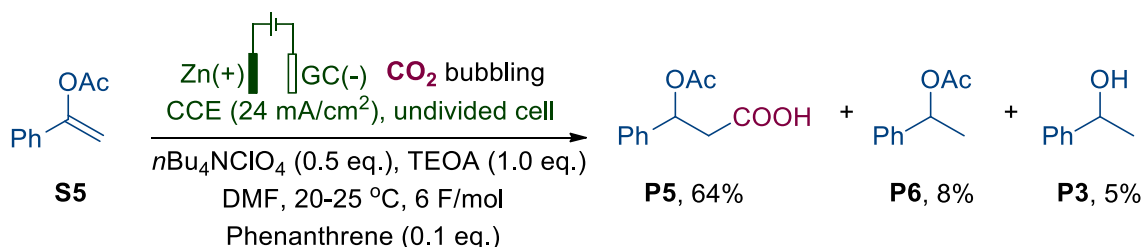
Anal. Calcd for $\text{C}_{15}\text{H}_{24}\text{SiO}_3$: C, 64.24; H, 8.63. Found: C, 64.54; H, 8.59.

IR (ATR): 2955, 2928, 2896, 2857, 1708, 1251, 1091, 828, 776, 698 cm^{-1} .

Electrolysis of S4



Electrolysis of S5

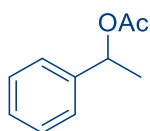


Products **P5** (64%, 133.1 mg, 0.65 mmol), **P3** (8%, 13.8 mg, 0.08 mmol), and **P6** (5%, 6 mg, 0.05 mmol) were isolated by chromatography on SiO_2 (PE:EA = from 5:1 to 1:1).

Further in the text, **S5** is **1a**, **P5** is **2a**.

The characteristic data of the acids **P5** (**2a**) is given below.

1-Phenylethyl acetate, **P6**²⁰



Yellow liquid.

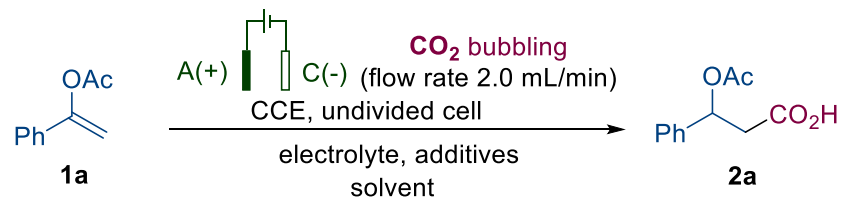
^1H NMR (300.13 MHz, CDCl_3 , δ): 7.41 – 7.26 (m, 5H), 5.89 (q, $J = 6.6$ Hz, 1H), 2.08 (s, 3H), 1.54 (d, $J = 6.6$ Hz, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 170.5, 141.8, 128.6 (2C), 128.0, 126.2 (2C), 72.4, 22.3, 21.5.

Experimental procedure for Table 1.

A 5 mL electrolysis cell IKA ElectraSyn 2.0 containing a cathode (Zn, Pt, GC) and an Zn anode (30 mm × 7 mm × 0.1 mm) was charged with a solution of enol acetate **1a** (1.0 mmol), electrolyte (0.5 mmol), amine (TEOA, DABCO, Et₃N, HOCH₂CH₂NH₂) (1.0 mmol), and redox mediator (phenanthrene, *p*-terphenyl) (0.1 eq.) in DMF (3 mL). The resulting mixture was bubbled with CO₂ (at a flow rate of 2 mL/min) and electrolyzed at a constant current of 30-70 mA ($j = 14\text{-}33 \text{ mA/cm}^2$) or without electricity with stirring at 20-25°C for 103-242 min. Then, aq. HCl (5M, 4 mL) was added to the reaction mixture. Extraction with PE/EtOAc (1:1, 5 × 5 mL) afforded the organic phase, which was washed with 10mL brine and dried over Na₂SO₄, filtered and concentrated under reduced pressure using a rotary evaporator (15-20 mmHg), (bath temperature, ca. 20–25 °C). The crude mixture was purified by column chromatography on silica gel or analyzed by ¹H NMR spectroscopy.

Table S1. The detailed optimization of electrochemical hydrocarboxylation of enol acetate 1a.



Entry	Anode/Cathode	Electrolyte (0.5 mmol)	Additives (base/amine 1.0 mmol, mediator 0.1 mmol)	Solvent (3 mL)	Current density, mA/cm ²	Electricity passed per 1a , F/mol	Yield 2a , %
1	C/C	ⁿ Bu ₄ NI	TEOA, -	DMF	7	4	traces ^b
2	C/C	ⁿ Bu ₄ NI	TEOA, -	DMF	48	4	11 ^b
3	C/C	ⁿ Bu ₄ NI	TEOA, -	CH₃CN	24	6	traces ^b
4	C/C	ⁿ Bu ₄ NI	TEOA, -	DMSO	24	6	19 ^a
5	C/C	ⁿ Bu ₄ NI	TEOA, -	THF	24	6	traces ^b
6	C/C	ⁿBu₄NBr	TEOA, -	DMF	24	6	14 ^a
7	C/C	ⁿBu₄NBF₄	TEOA, -	DMF	24	6	33 ^a
8	C/C	ⁿMe₄NBr	TEOA, -	DMF	24	6	n.d. ^a
9	C/C	ⁿBu₄NCIO₄	TEOA, -	DMF	24	6	36 ^b
10	C/C	LiClO₄	TEOA, -	DMF	24	6	9 ^a
11	C/C	KI	TEOA, -	DMF	24	6	traces ^b
12	C/C	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	21 ^a
13	C/Ni	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	6 ^b
14	C/SS	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	26 ^a
15	C/Zn	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	40 ^a
16	C/Mg	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	traces ^a
17	C/RVC_{foam}	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	15 ^a
18	C/Ni_{foam}	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	traces ^a
19	C/Cu	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	traces ^b
20	Zn/C	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	50 ^b
21	Mg/C	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	45 ^a
22	SS/Zn	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	37 ^a
23	Zn/Zn	ⁿ Bu ₄ NI	TEOA, -	DMF	24	6	48 ^a

24	Zn/Zn	ⁿ Bu ₄ Nl	-, -	DMF	24	6	33 ^a
25	Zn/Zn	ⁿ Bu ₄ NPF ₆	TEOA, -	DMF	24	6	34 ^a
26	Zn/Zn	ⁿ Bu ₄ NBF ₄	TEOA, -	DMF	24	6	51 ^a
27	Zn/Zn	LiClO ₄	TEOA, -	DMF	24	6	traces ^a
28	Zn/Zn	ⁿ Bu ₄ NBr	TEOA, -	DMF	24	6	55 ^a
29	Zn/Zn	[PyH]ClO ₄	TEOA, -	DMF	24	6	traces ^a
30	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA, -	DMF	24	6	57 ^a
31	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA, phenanthrene	DMF	24	6	63 ^a
32	Zn/Zn	ⁿ Bu ₄ NClO ₄	DABCO , -	DMF	24	6	31 ^a
33	Zn/Zn	ⁿ Bu ₄ NClO ₄	Et₃N , -	DMF	24	6	24 ^a
34	Zn/Zn	ⁿ Bu ₄ NClO ₄	K₂CO₃ , -	DMF	24	6	27 ^a
35	Zn/Zn	ⁿ Bu ₄ NClO ₄	Py , -	DMF	24	6	22 ^a
36	Zn/Zn	ⁿ Bu ₄ NClO ₄	NaOH , -	DMF	24	6	19 ^a
37	Zn/Zn	ⁿ Bu ₄ NClO ₄	H₂O , -	DMF	24	6	25 ^a
38	Zn/Zn	ⁿ Bu ₄ NClO ₄	HOCH₂CH₂NH₂ , -	DMF	24	6	0 ^a
39	Zn/Zn	ⁿ Bu ₄ NClO ₄	Imidazole , -	DMF	24	6	30 ^a
40	Zn/Zn	ⁿ Bu ₄ NClO ₄	1,3-bis(methylamino)propan-2-ol , -	DMF	24	6	20 ^a
41	Zn/Zn	ⁿ Bu ₄ NClO ₄	N,N,N',N'-tetraethylmethanedi-amine , -	DMF	24	6	12 ^a
42	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA , anthracene	DMF	24	6	48 ^a
43	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA , phenantroline	DMF	24	6	44 ^a
44	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA , graphene	DMF	24	6	44 ^a
45	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA , 1-methylnaphthalene	DMF	24	6	50 ^a
46	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA , p-terphenyl	DMF	24	6	53 ^b
47	Zn/Zn	ⁿ Bu ₄ NClO ₄	TEOA , phenanthrene	DMF	24	4	38 ^b

48 ^c	Zn/Zn	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	6	37 ^b
49 ^d	Zn/Zn	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	6	6 ^b
50 ^e	Zn/Zn	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	6	48 ^b
51	Pt/Pt	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	6	11 ^a
52	Zn/Pt	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	6	46 ^a
53	Zn/GC	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	6	64 ^a
54	Zn/GC	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	4	63 ^a
55	Zn/GC	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	24	2	14 ^b
56	Zn/GC	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	14	4	35 ^a
57	Zn/GC	ⁿ Bu ₄ NCIO ₄	TEOA, phenanthrene	DMF	33	4	61 ^a

^a isolated **2a** yield.

^b yield **2a** by ¹H NMR

^c at 0°C

^d under 5 bar

^e CO₂ flow rate of 0.5 mL/min

General Experimental Procedure for Table S1.

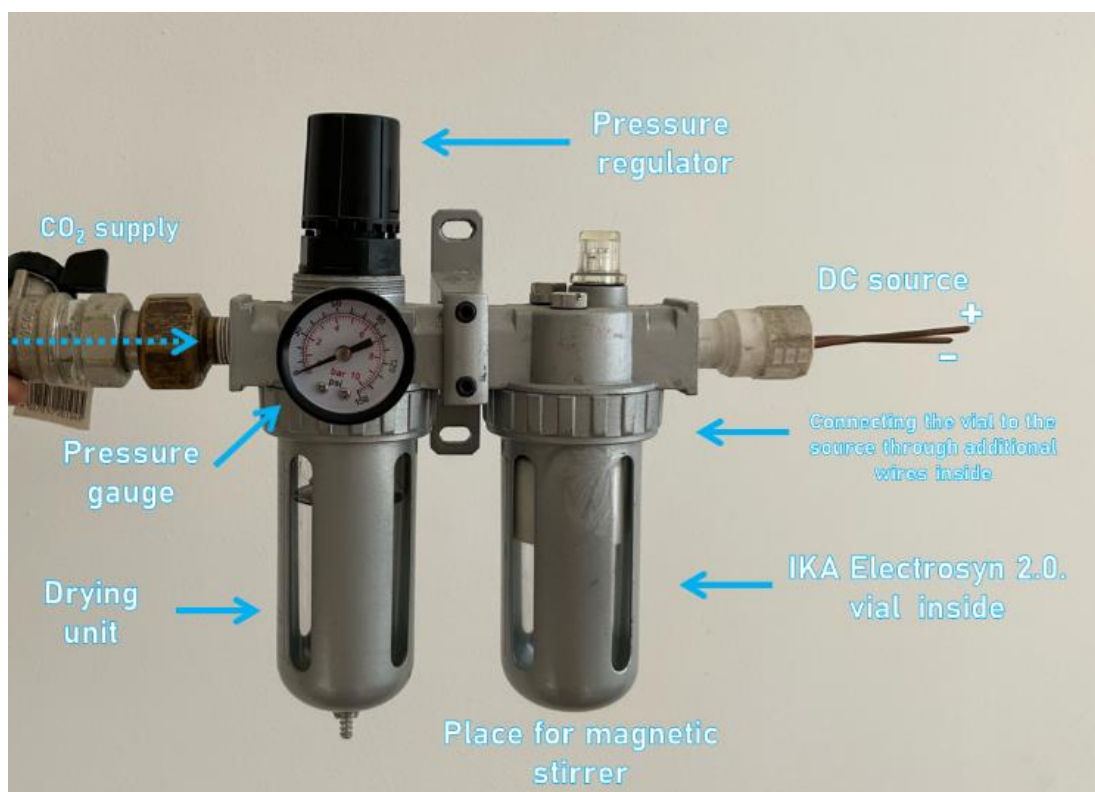
Isolation method A

A 5 mL electrolysis cell IKA ElectraSyn 2.0 containing a plate cathode and plate anode (30 mm × 7 mm × 0.1 mm) was charged with a solution of enol acetate **1a** (1.0 mmol), supporting electrolyte (0.5 mmol) and additives (base/amine 1.0 mmol; mediator 0.1 mmol) in solvent (3 mL). The resulting mixture was bubbled with CO₂ (at a flow rate of 2 mL/min) and electrolyzed at a constant current of 15 – 100 mA ($j = 7 - 48 \text{ mA/cm}^2$) with stirring at 20-25 °C for 64-428 min. Then, aq. HCl (5M, 4 mL) was added to the crude reaction mixture. Extraction with Et₂O (1:1, 5×5 mL) afforded the organic phase, which was washed with 10 mL brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure using a rotary evaporator (15-20 mmHg), (bath temperature, ca. 25–30 °C).

In ^a cases, the crude mixture was purified by column chromatography on silica gel (PE:EA = from 5:1 to 1:1) to give the target product.

In ^b cases, the **2a** yield was determined by ¹H NMR from the crude reaction mixture.

entries 3,5: CH₃CN and THF are volatile compounds, during the reaction the solvent was carried away by the CO₂ flow. Therefore, as the volume of solvent was reduced, extra portion solvent was added to the reaction (about 20 mL in total for each entry).



Picture S3. Electrochemical reaction setup for the experiment 52 (Table S1).

General Experimental Procedure for Scheme 3.

A 5 mL electrolysis cell IKA ElectraSyn 2.0 containing zinc cathode and glassy carbon anode (30 mm × 7 mm × 0.1 mm) was charged with a solution of enol acetate **1** (1.0 mmol), *n*Bu₄NClO₄ (0.5 mmol, 171 mg), TEOA (1.0 mmol, 149 mg) and phenanthrene (0.1 mmol, 17.8 mg) in DMF (3 mL). The resulting mixture was bubbled with CO₂ (at a flow rate of 2 mL/min) and electrolyzed at a constant current of 50 mA ($j = 24 \text{ mA/cm}^2$) with stirring at 20–25 °C for 194 min (for **1e** – 478 min, **1o** – 487 min, for **1p** – 362 min.). Then, aq. HCl (5M, 4 mL) was added to the crude reaction mixture. Extraction with PE/EtOAc (1:1, 5×5 mL) afforded the organic phase, which was washed with 10 mL brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure using a rotary evaporator (15–20 mmHg), (bath temperature, ca. 25–30 °C). The crude mixture was purified by column chromatography on silica gel (PE:EA = from 5:1 to 1:1) to give the target product **2**.

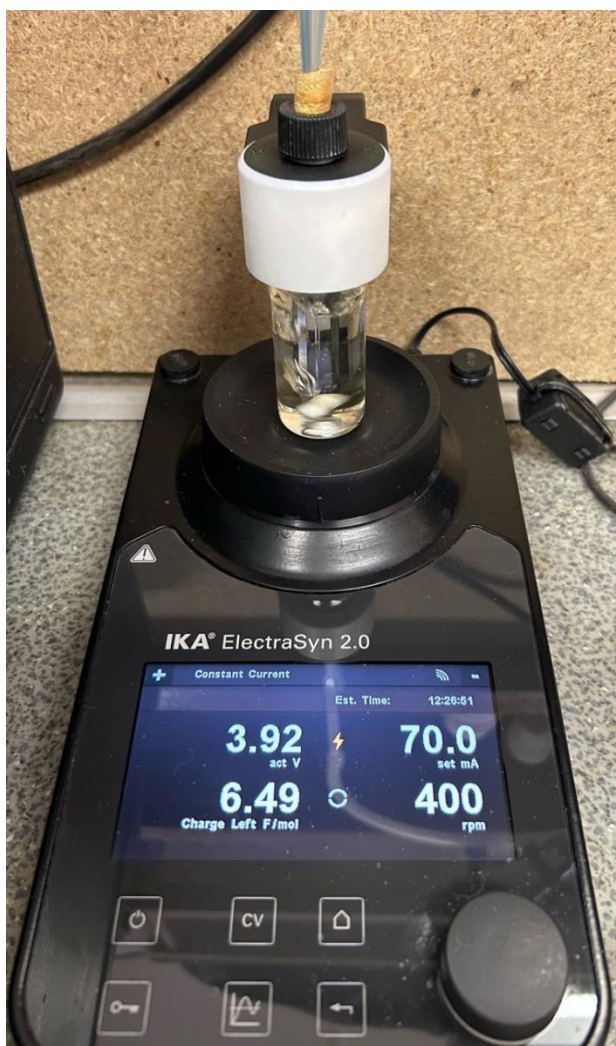
If column chromatography is unsuccessful, pure acids can be isolated by the following procedure:

NaHCO₃ (20 mmol), H₂O (10 mL), EtOAc (10 mL) was added to the crude reaction mixture and the resulting mixture were stirred for 4 hours at room temperature. After that water (15 mL) was added and aqueous layer was washed with EtOAc (4×5 mL). The resulting aqueous layer was washed with 0.7M solution of HCOOH in Et₂O (3×15 mL). Combined organic layers were dried over Na₂SO₄, filtered and concentrated under reduced pressure using rotary evaporator (15–20 mmHg), (bath temperature, ca. 40–45 °C) to give the target product **2**.

Gram-scale experiment

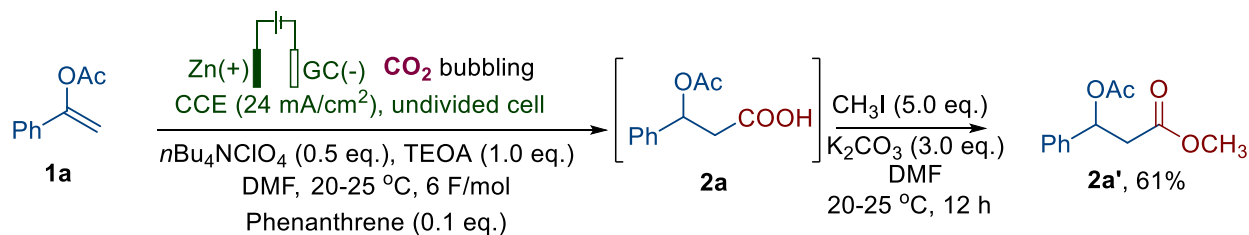
A 20 mL electrolysis cell IKA ElectraSyn 2.0 containing a glassy carbon cathode and zinc anode (30 mm × 7 mm × 0.1 mm) was charged with a solution of enol acetate **1a** (5.0 mmol, 810 mg), *n*Bu₄NClO₄ (2.5 mmol, 855 mg), TEOA (5.0 mmol, 745 mg) and phenanthrene (0.5 mmol, 89 mg) in DMF (15 mL). The resulting mixture was bubbled with CO₂ (at a flow rate of 4 mL/min) and electrolyzed at a constant current of 70 mA ($j = 33 \text{ mA/cm}^2$) with stirring at 20–25° for 804 min. Then, aq. HCl (5M, 10 mL) was added to the crude reaction mixture. Extraction with PE/EtOAc (1:1, 5×30 mL) afforded the organic phase, which was washed with 2×30 mL brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure using a rotary evaporator (15–20 mmHg), (bath temperature, ca. 25–30 °C). Then the crude reaction mixture was added NaHCO₃ (50 mmol), H₂O (70 mL), EtOAc (70 mL) and stirred for 8 hours at room temperature. After that H₂O (15 mL) was added and aqueous layer was washed EtOAc (4×10 mL). The resulting aqueous layer was washed 0.7M solution of HCOOH in Et₂O (3×50 mL). Combined organic layers

were dried over Na_2SO_4 , filtered and concentrated under reduced pressure using rotary evaporator (15-20 mmHg), (bath temperature, ca. 40–45 °C) to give the target product **2a**.



Picture S4. Electrochemical reaction setup for Gram-scale experiment.

The synthesis of methyl ester **2a'**

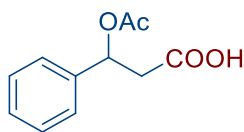


A 5 mL electrolysis cell IKA ElectraSyn 2.0 containing zinc cathode and glassy carbon anode (30 mm × 7 mm × 0.1 mm) was charged with a solution of enol acetate **1a** (1.0 mmol), *n*Bu₄NClO₄ (0.5 mmol, 171 mg), TEOA (1.0 mmol, 149 mg) and phenanthrene (0.1 mmol, 17.8 mg) in DMF (3 mL). The resulting mixture was bubbled with CO₂ (at a flow rate of 2 mL/min) and electrolyzed at a constant current of 50 mA ($j = 24 \text{ mA/cm}^2$) with stirring at 20-25 °C for 194 min.

Later, DMF (2.5 mL), K₂CO₃ (3.0 mmol, 414 mg), methyl iodide (5.0 mmol, 709.5 mg) were added to the reaction mixture. The mixture was stirred at 20-25 °C for 12 h. Then, water (10 mL) was added to the crude reaction mixture. Extraction with PE/EtOAc (1:1, 5×10 mL) afforded the organic phase, which was washed with 10mL brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure using a rotary evaporator (15-20 mmHg), (bath temperature, ca. 25–30 °C). The crude mixture was purified by column chromatography on silica gel (PE:EtOAc = from 10:1 to 5:1) to give the target product **2a'**.

The characterization data of the synthesized products.

3-Acetoxy-3-phenylpropanoic acid, 2a²¹



Yield 64% (133.1 mg, 0.64 mmol). For gram-scale experiment yield was 66% (0.690 g, 3.31 mmol)

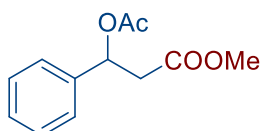
Brown crystals, m.p. = 99-100 °C (lit. mp = 100-101 °C).²⁴

¹H NMR (300.13 MHz, CDCl₃, δ): 11.20 (brs, 1H), 7.50 – 7.27 (m, 5H), 6.17 (dd, *J* = 9.0, 5.0 Hz, 1H), 3.03 (dd, *J* = 16.3, 9.0 Hz, 1H), 2.81 (dd, *J* = 16.3, 5.0 Hz, 1H), 2.07 (s, 3H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 176.0, 170.1, 139.1, 128.8 (2C), 128.6, 126.6 (2C), 71.9, 41.1, 21.1.

HRMS (ESI/TOF) m/z: [M + Na]⁺ Calcd for [C₁₁H₁₂O₄Na]⁺ 231.0628; Found 231.0623.

Methyl 3-acetoxy-3-phenylpropanoate, 2a'²²



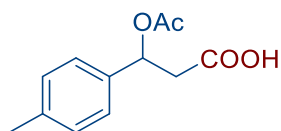
Yield 61% (135.2 mg, 0.61 mmol). Colorless oil.

¹H NMR (300.13 MHz, CDCl₃, δ): 7.42 – 7.27 (m, 5H), 6.17 (dd, *J* = 8.9, 5.2 Hz, 1H), 3.67 (s, 3H), 2.98 (dd, *J* = 15.8, 8.9 Hz, 1H), 2.76 (dd, *J* = 15.8, 5.2 Hz, 1H), 2.05 (s, 3H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 170.3, 169.9, 139.4, 128.8 (2C), 128.5, 126.6 (2C), 72.2, 52.0, 41.4, 21.2.

HRMS (ESI/TOF) m/z: [M + Na]⁺ Calcd for [C₁₂H₁₄O₄NH₄]⁺ 240.1230; Found 240.1232.

3-Acetoxy-3-(p-tolyl)propanoic acid, 2b



Yield 55% (122.0 mg, 0.55 mmol). Brownish powder, m.p. = 122-124 °C.

¹H NMR (300.13 MHz, CDCl₃, δ): 10.65 (brs, 1H), 7.35 – 7.08 (m, 4H), 6.14 (dd, *J* = 8.9, 5.1 Hz, 1H), 3.03 (dd, *J* = 16.2, 8.9 Hz, 1H), 2.80 (dd, *J* = 16.2, 5.1 Hz, 1H), 2.35 (s, 3H), 2.05 (s, 3H).

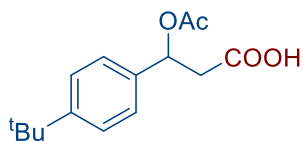
¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 176.1, 170.1, 138.5, 136.1, 129.5 (2C), 126.7 (2C), 71.8, 41.1, 21.3, 21.2.

HRMS (ESI/TOF) m/z: [M + Na]⁺ Calcd for [C₁₂H₁₄O₄Na]⁺ 245.0784; Found 245.0773.

Anal. Calcd for C₁₂H₁₄O₄: C, 64.85; H, 6.35. Found: C, 65.12; H, 6.45.

IR (ATR): 3184, 3050, 2971, 2952, 2873, 1734, 1701, 1519, 1437, 1220, 1208, 1181, 1013, 942 cm⁻¹.

3-Acetoxy-3-(4-(*tert*-butyl)phenyl)propanoic acid, 2c²¹



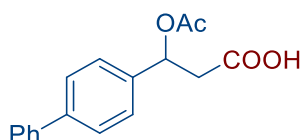
Yield 52% (145.3 mg, 0.52 mmol). Yellow oil.

¹H NMR (300.13 MHz, CDCl₃, δ): 9.72 (brs, 1H), 7.42 – 7.24 (m, 4H), 6.16 (dd, *J* = 9.2, 4.9 Hz, 1H), 3.03 (dd, *J* = 16.3, 9.2 Hz, 1H), 2.80 (dd, *J* = 16.3, 4.9 Hz, 1H), 2.05 (s, 3H), 1.31 (s, 9H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 176.1, 170.2, 151.6, 136.0, 126.4 (2C), 125.7 (2C), 71.7, 41.1, 34.7, 31.4 (3C), 21.2.

HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₁₅H₂₀O₄Na]⁺ 287.1254; Found 287.1260.

3-([1,1'-Biphenyl]-4-yl)-3-acetoxypropanoic acid, 2d²¹



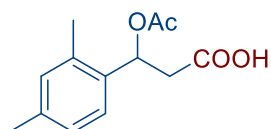
Yield 53% (157.8 mg, 0.53 mmol). White powder, m.p. = 136-138 °C.

¹H NMR (300.13 MHz, CDCl₃, δ): 10.30 (brs, 1H), 7.63 – 7.54 (m, 4H), 7.49 – 7.30 (m, 5H), 6.22 (dd, *J* = 9.0, 5.1 Hz, 1H), 3.08 (dd, *J* = 16.3, 9.0 Hz, 1H), 2.86 (dd, *J* = 16.3, 5.1 Hz, 1H), 2.09 (s, 3H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 175.7, 170.1, 141.6, 140.7, 138.0, 128.9 (2C), 127.6, 127.6 (2C), 127.3 (2C), 127.2 (2C), 71.7, 41.0, 21.2.

HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₁₇H₁₆O₄Na]⁺ 307.0941; Found 307.0938.

3-Acetoxy-3-(2,4-dimethylphenyl)propanoic acid, 2e



Yield 45% (106.1 mg, 0.45 mmol). Yellow powder, m.p. = 80-82 °C.

¹H NMR (300.13 MHz, CDCl₃, δ): 10.30 (brs, 1H), 7.28 – 7.19 (m, 1H), 7.06– 6.96 (m, 2H), 6.33 (dd, *J* = 9.2, 4.7 Hz, 1H), 2.97 (dd, *J* = 16.3, 9.2 Hz, 1H), 2.75 (dd, *J* = 16.3, 4.7 Hz, 1H), 2.42 (s, 3H), 2.30 (s, 3H), 2.05 (s, 3H).

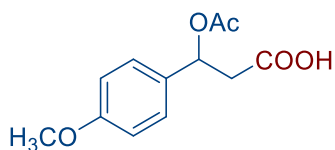
¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 175.9, 170.0, 138.1, 135.3, 134.7, 131.5, 127.2, 125.9, 68.9, 40.7, 21.2, 19.1.

HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₁₃H₁₆O₄Na]⁺ 259.0941; Found 259.0942.

Anal. Calcd for C₁₃H₁₆O₄: C, 66.09; H, 6.83. Found: C, 66.38; H, 6.97.

IR (ATR): 3040, 2949, 2923, 2858, 1746, 1707, 1435, 1371, 1225, 1212, 1187 cm⁻¹.

3-Acetoxy-3-(4-methoxyphenyl)propanoic acid, 2f



Yield 25% (60.1 mg, 0.25 mmol). Yellow powder, 114-116 °C.

¹H NMR (300.13 MHz, CDCl₃, δ): 9.86 (brs, 1H), 7.34 – 7.27 (m, 2H), 6.90– 6.84 (m, 2H), 6.11 (dd, *J* = 8.8, 5.3 Hz, 1H), 3.80 (s, 4H), 3.03 (dd, *J* = 16.2, 8.8 Hz, 1H), 2.79 (dd, *J* = 16.2, 5.3 Hz, 1H), 2.04 (s, 3H).

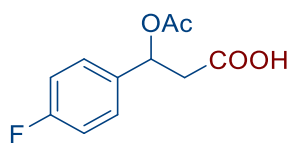
¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 175.6, 170.1, 159.8, 131.2, 128.2 (2C), 114.2 (2C), 71.6, 55.4, 40.9, 21.3.

HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₁₂H₁₄O₅Na]⁺ 261.0733; Found 261.0742.

Anal. Calcd for C₁₂H₁₄O₅: C, 60.50; H, 5.92. Found: C, 60.79; H, 6.00.

IR (ATR): 3041, 2955, 2925, 2837, 1733, 1706, 1240, 1206, 1174, 1023, 822 cm⁻¹.

3-Acetoxy-3-(4-fluorophenyl)propanoic acid, 2g



Yield 49% (110.2 mg, 0.49 mmol). Yellow powder, m.p. = 72-74 °C.

¹H NMR (300.13 MHz, CDCl₃, δ): 7.38 – 7.29 (m, 2H), 7.08– 6.94 (m, 2H), 6.12 (dd, *J* = 8.8, 5.3 Hz, 1H), 3.02 (dd, *J* = 16.3, 8.8 Hz, 1H), 2.78 (dd, *J* = 16.3, 5.3 Hz, 1H), 2.05 (s, 3H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 175.6, 170.0, 162.8 (d, ¹*J*_{CF} = 247.4 Hz), 134.9 (d, ⁴*J*_{CF} = 3.2 Hz), 128.6 (d, ³*J*_{CF} = 8.2 Hz, 2C), 115.8 (d, ²*J*_{CF} = 21.8 Hz, 2C), 71.2, 41.0, 21.2.

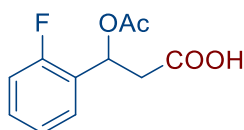
¹⁹F NMR (282 MHz, CDCl₃, δ): -113.13 (tt, *J* = 8.6, 5.2 Hz).

HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₁₁H₁₁FO₄Na]⁺ 249.0534; Found 249.0542.

Anal. Calcd for C₁₁H₁₁FO₄: C, 58.41; H, 4.90. Found: C, 58.74; H, 4.87.

IR (ATR): 3130, 3070, 2934, 2858, 2798, 1734, 1698, 1604, 1511, 1267, 1222, 1175, 1159, 883 cm⁻¹.

3-Acetoxy-3-(2-fluorophenyl)propanoic acid, 2h



Yield 52% (117.3 mg, 0.52 mmol). Brownish powder, m.p. = 66-68 °C.

^1H NMR (300.13 MHz, CDCl_3 , δ): 7.39 (td, $J = 7.5, 1.8$ Hz, 1H), 7.31 (tdd, $J = 7.3, 5.3, 1.9$, 1H), 7.16 (td, $J = 7.5, 1.2$ Hz, 1H), 7.07 (ddd, $J = 10.5, 8.2, 1.2$ Hz, 1H), 6.39 (dd, $J = 9.2, 4.6$ Hz, 1H), 3.04 (dd, $J = 16.5, 9.2$ Hz, 1H), 2.88 (dd, $J = 16.5, 4.6$ Hz, 1H), 2.08 (s, 3H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 175.8, 170.0, 159.9 (d, $^1J_{\text{CF}} = 248.0$ Hz), 130.2 (d, $^3J_{\text{CF}} = 8.4$ Hz), 128.0 (d, $^3J_{\text{CF}} = 3.8$ Hz), 126.2 (d, $^2J_{\text{CF}} = 13.1$ Hz), 124.4 (d, $^4J_{\text{CF}} = 3.8$ Hz), 115.9 (d, $^2J_{\text{CF}} = 21.5$ Hz), 66.9 (d, $^3J_{\text{CF}} = 2.5$ Hz), 39.7 (d, $^4J_{\text{CF}} = 1.9$ Hz), 20.9.

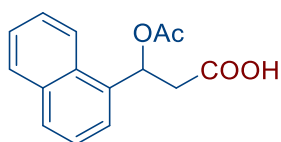
^{19}F NMR (282 MHz, CDCl_3 , δ): -117.23 (ddd, $J = 10.5, 7.3, 5.3$ Hz).

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{11}\text{H}_{11}\text{FO}_4\text{Na}]^+$ 249.0534; Found 249.0534.

Anal. Calcd for $\text{C}_{11}\text{H}_{11}\text{FO}_4$: C, 58.41; H, 4.90. Found: C, 58.21; H, 4.83.

IR (ATR): 3110, 3052, 2924, 2696, 1738, 1702, 1491, 1434, 1376, 1247, 1233, 1211, 1196, 1032, 765 cm^{-1} .

3-Acetoxy-3-(naphthalen-1-yl)propanoic acid, 2i



Yield 62% (159.1 mg, 0.62 mmol). Yellow powder, m.p. = 96-98 $^{\circ}\text{C}$.

^1H NMR (300.13 MHz, CDCl_3 , δ): 9.53 (brs, 1H), 8.19 – 8.13 (m, 1H), 7.90 – 7.80 (m, 2H), 7.61 – 7.42 (m, 4H), 6.94 (dd, $J = 9.2, 4.2$ Hz, 1H), 3.12 (dd, $J = 16.4, 9.2$ Hz, 1H), 3.01 (dd, $J = 16.4, 4.2$ Hz, 1H), 2.14 (s, 3H).

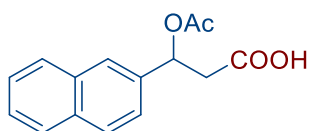
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 175.9, 170.0, 135.1, 134.0, 130.0, 129.2, 129.2, 126.9, 126.1, 125.4, 123.9, 123.0, 69.6, 41.0, 21.2.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{15}\text{H}_{14}\text{O}_4\text{Na}]^+$ 281.0784; Found 281.0779.

Anal. Calcd for $\text{C}_{15}\text{H}_{14}\text{O}_4$: C, 69.76; H, 5.46. Found: C, 69.74; H, 5.42.

IR (ATR): 3060, 2924, 1729, 1693, 1512, 1377, 1255, 1239, 1159, 1021, 778 cm^{-1} .

3-Acetoxy-3-(naphthalen-2-yl)propanoic acid, 2j



Yield 64% (165.3 mg, 0.64 mmol). White powder, m.p. = 124-126 $^{\circ}\text{C}$.

^1H NMR (300.13 MHz, CDCl_3 , δ): 7.87 – 7.79 (m, 3H), 7.53 – 7.44 (m, 4H), 6.33 (dd, $J = 9.0, 5.1$ Hz, 1H), 3.13 (dd, $J = 16.3, 9.0$ Hz, 1H), 2.90 (dd, $J = 16.3, 5.1$ Hz, 1H), 2.08 (s, 3H).

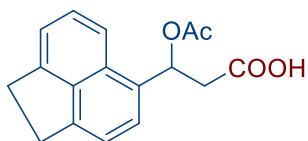
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3 , δ): 175.7, 170.1, 136.4, 133.4, 133.2, 128.8, 128.3, 127.8, 126.6, 126.1, 124.1, 72.1, 41.1, 21.2.

HRMS (ESI/TOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $[\text{C}_{15}\text{H}_{14}\text{O}_4\text{Na}]^+$ 281.0784; Found 281.0792.

Anal. Calcd for C₁₅H₁₄O₄: C, 69.76; H, 5.46. Found: C, 69.66; H, 5.66.

IR (ATR): 3031, 2967, 2917, 2683, 1736, 1698, 1426, 1375, 1241, 1222, 1186, 1033, 833, 751 cm⁻¹.

3-Acetoxy-3-(1,2-dihydroacenaphthylen-5-yl)propanoic acid, 2k



Yield 51% (144.8 mg, 0.51 mmol). White powder, m.p. = 126-128 °C.

¹H NMR (300.13 MHz, CDCl₃, δ): 7.90 – 7.83 (m, 1H), 7.54 – 7.47 (m, 2H), 7.35 – 7.22 (m, 2H), 6.78 (dd, *J* = 9.2, 4.7 Hz, 1H), 3.44 – 3.25 (m, 4H), 3.19 (dd, *J* = 16.3, 9.2 Hz, 1H), 2.99 (dd, *J* = 16.3, 4.7 Hz, 1H), 2.09 (s, 3H).

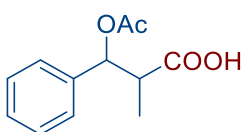
¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 175.8, 170.1, 147.2, 146.9, 139.8, 130.7, 128.7, 128.6, 126.1, 119.7, 119.1, 119.0, 70.1, 40.7, 30.6, 30.1, 21.2.

HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₁₇H₁₆O₄Na]⁺ 307.0941; Found 307.0933.

Anal. Calcd for C₁₇H₁₆O₄: C, 71.82; H, 5.67. Found: C, 71.80; H, 5.68.

IR (ATR): 3046, 2960, 2945, 2908, 2825, 1751, 1707, 1418, 1364, 1250, 1222, 1194, 841, 769 cm⁻¹.

3-Acetoxy-2-methyl-3-phenylpropanoic acid, 2l²³



Yield of two diastereomers was 52% (115.4 mg, 0.52 mmol). The diastereomer ratio was 5:2. Yellow oil.

HRMS (ESI/TOF) *m/z*: [M + K]⁺ Calcd for [C₁₂H₁₄O₄K]⁺ 261.0524; Found 261.0529.

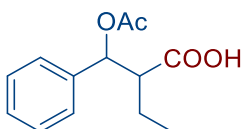
Major: ¹H NMR (300.23 MHz, CDCl₃, δ): 9.80 (brs, 1H), 7.36 – 7.26 (m, 5H), 5.82 (d, *J* = 10.1 Hz, 1H), 3.07 – 2.91 (m, 1H), 2.01 (s, 3H), 1.01 (d, *J* = 7.2 Hz, 3H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 179.1, 169.9, 138.3, 128.5 (2C), 128.2, 126.7 (2C), 75.5, 45.5, 21.0, 12.2.

Minor: ¹H NMR (300.23 MHz, CDCl₃, δ): 9.80 (brs, 1H), 7.36 – 7.26 (m, 5H), 6.10 (d, *J* = 6.4 Hz, 1H), 3.07 – 2.91 (m, 1H), 2.10 (s, 3H), 1.24 (d, *J* = 7.1 Hz, 3H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): 180.2, 169.9, 137.7, 128.7, 128.7 (2C), 127.6 (2C), 77.2, 45.3, 21.1, 14.0.

2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



Yield of mixture was 49% (115.63 mg, 0.49 mmol). The diastereomer ratio was 10:7. Yellow oil. HRMS (ESI/TOF) m/z : $[M + K]^+$ Calcd for $[C_{13}H_{16}O_4K]^+$ 275.0675; Found 275.0680.

Anal. Calcd for $C_{13}H_{16}O_4$: C, 66.09; H, 6.83. Found: C, 65.95; H, 6.73.

IR (ATR): 3035, 2974, 2937, 2880, 1740, 1707, 1378, 1358, 1227, 1023, 869, 699 cm^{-1} .

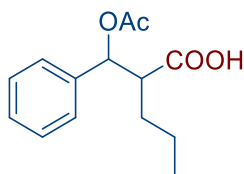
Major: 1H NMR (300.23 MHz, $CDCl_3$, δ): 10.19 (brs, 1H), 7.40 – 7.26 (m, 5H), 5.97 (d, $J = 8.4$ Hz, 1H), 2.91– 2.78 (m, 1H), 2.08 (s, 3H), 1.76 (quintet, $J = 7.4$ Hz, 2H), 0.96 (t, $J = 7.4$ Hz, 3H).

$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$, δ): 178.6, 170.1, 138.3, 128.5 (2C), 128.5, 127.1 (2C), 76.7, 53.3, 21.8, 21.1, 11.7.

Minor: 1H NMR (300.23 MHz, $CDCl_3$, δ): 10.19 (brs, 1H), 7.40 – 7.26 (m, 5H), 5.82 (d, $J = 10.4$ Hz, 1H), 2.91 – 2.78 (m, 1H), 1.98 (s, 3H), 1.55 – 1.38 (m, 1H), 1.33 – 1.18 (m, 1H), 0.86 (t, $J = 7.4$ Hz, 3H).

$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$, δ): 179.6, 169.9, 137.9, 128.7, 128.7 (2C), 127.6 (2C), 75.5, 52.9, 22.0, 21.0, 11.3.

2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n



Yield of mixture was 58% (145.1 mg, 0.58 mmol). The diastereomer ratio was 10:6.5. Colorless oil. HRMS (ESI/TOF) m/z : $[M + Na]^+$ Calcd for $[C_{14}H_{18}O_4Na]^+$ 273.1097; Found 273.1107.

Anal. Calcd for $C_{14}H_{18}O_4$: C, 67.18; H, 7.25. Found: C, 67.47; H, 7.28.

IR (ATR): 3035, 2963, 2934, 2874, 1741, 1707, 1373, 1228, 1022, 870, 700 cm^{-1} .

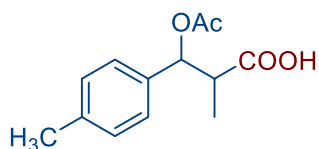
Major: 1H NMR (300.13 MHz, $CDCl_3$): 10.09 (brs, 1H), 7.37 – 7.26 (m, 5H), 5.95 (d, $J = 8.3$ Hz, 1H), 2.98 – 2.86 (m, 1H), 2.08 (s, 3H), 1.79 – 1.55 (m, 2H), 1.52 – 1.29 (m, 2H), 0.91 (t, $J = 7.3$ Hz, 3H).

$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$): δ 177.5, 170.0, 138.4, 128.5 (2C), 128.5, 127.2 (2C), 75.7, 51.6, 30.7, 21.2, 20.6, 14.0.

Minor: 1H NMR (300.13 MHz, $CDCl_3$): 10.09 (brs, 1H), 7.37 – 7.26 (m, 5H), 5.79 (d, $J = 10.4$ Hz, 1H), 2.98 – 2.86 (m, 1H), 1.98 (s, 3H), 1.52 – 1.03 (m, 4H), 0.79 (t, $J = 7.2$ Hz, 3H).

$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$): δ 178.7, 169.7, 138.0, 128.8, 128.8 (2C), 127.7 (2C), 76.9, 51.2, 30.8, 21.1, 20.3, 13.9.

3-Acetoxy-2-methyl-3-(p-tolyl)propanoic acid, 2o



Yield of mixture was 56% (132.2 mg, 0.56 mmol). The diastereomer ratio was 10:1.5. Colorless oil. HRMS (ESI/TOF) m/z : $[M + Na]^+$ Calcd for $[C_{13}H_{16}O_4Na]^+$ 259.0941; Found 259.0941.

Anal. Calcd for $C_{13}H_{16}O_4$: C, 66.09; H, 6.83. Found: C, 66.09; H, 6.80.

IR (ATR): 2979, 2960, 2924, 1742, 1702, 1222, 1201, 1018, 813 cm^{-1} .

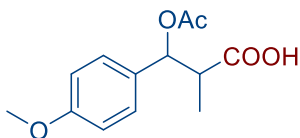
Major: 1H NMR (300.13 MHz, $CDCl_3$): 7.25 – 7.08 (m, 4H), 6.05 (d, $J = 6.6$ Hz, 1H), 2.99 – 2.87 (m, 1H), 2.33 (s, 3H), 2.08 (s, 3H), 1.23 (d, $J = 7.1$ Hz, 3H).

$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$): 179.3, 170.1, 138.1, 135.4, 129.2 (2C), 126.6 (2C), 75.5, 45.6, 21.3, 21.2, 12.4.

Minor: 1H NMR (300.13 MHz, $CDCl_3$): 7.25 – 7.08 (m, 4H), 5.78 (d, $J = 10.1$ Hz, 1H), 3.03 – 2.91 (m, 1H), 2.33 (s, 3H), 1.99 (s, 3H), 0.99 (d, $J = 7.2$ Hz, 3H).

$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$): 180.4, 169.9, 138.6, 134.6, 129.4 (2C), 127.5 (2C), 77.1, 45.2, 21.4, 21.2, 14.1.

3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p



Yield of mixture was 35% (88.4 mg, 0.35 mmol). The diastereomer ratio was 10:3.9. Brownish oil. HRMS (ESI/TOF) m/z : $[M + NH_4]^+$ Calcd for $[C_{13}H_{16}O_5NH_4]^+$ 270.1336; Found 270.1337.

Anal. Calcd for $C_{13}H_{16}O_5$: C, 61.90; H, 6.39. Found: C, 61.82; H, 6.29.

IR (ATR): 2985, 2940, 2915, 2839, 1736, 1707, 1515, 1227, 1175, 1022, 825 cm^{-1} .

Major: 1H NMR (300.13 MHz, $CDCl_3$): 10.02 (brs, 1H), 7.33 – 7.19 (m, 2H), 6.92 – 6.81 (m, 2H), 6.00 (d, $J = 7.1$ Hz, 1H), 3.78 (s, 3H), 2.97 (quintet, $J = 7.1$ Hz, 1H), 2.07 (s, 3H), 1.25 (d, $J = 7.1$ Hz, 3H).

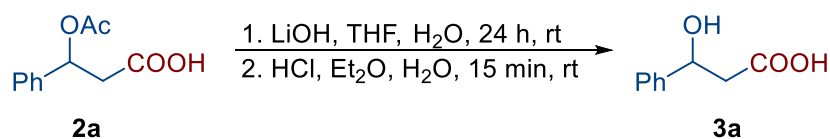
$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$): 179.2, 170.1, 159.5, 130.5, 128.2 (2C), 113.9 (2C), 75.5, 55.3, 45.7, 21.1, 12.9.

Minor: 1H NMR (300.13 MHz, $CDCl_3$): 10.02 (brs, 1H), 7.33 – 7.19 (m, 2H), 6.92 – 6.81 (m, 2H), 5.77 (d, $J = 10.1$ Hz, 1H), 3.79 (s, 3H), 3.04 – 2.89 (m, 1H), 1.98 (s, 3H), 0.99 (d, $J = 7.2$ Hz, 3H).

$^{13}C\{^1H\}$ NMR (75.48 MHz, $CDCl_3$): 180.2, 169.9, 159.8, 129.7, 128.9 (2C), 114.1 (2C), 76.9, 55.4, 45.3, 21.1, 14.1.

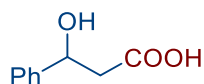
Experimental Procedures for Scheme 4.

Synthesis of 3-Hydroxy-3-phenylpropanoic acid, **3a**²⁴



LiOH (1.15 g, 5.55 mmol) in H₂O (15 mL) was added to a solution of acid **2a** (300.0 mg, 1.44 mmol) in THF (180 mL) at 20–25 °C. The mixture was stirred at room temperature 24 h. Et₂O (200 mL) and dilute aqueous HCl (1M, 200 mL) were added and the mixture was stirred for 15 min before separating the layers. The aqueous layer was extracted with Et₂O (3 x 70 mL). The combined organic layer was washed with water (2 x 70 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure using rotary evaporator (15–20 mmHg), (bath temperature, ca. 25–30 °C) to give the target product **3a**.

3-Hydroxy-3-phenylpropanoic acid **3a**²⁵



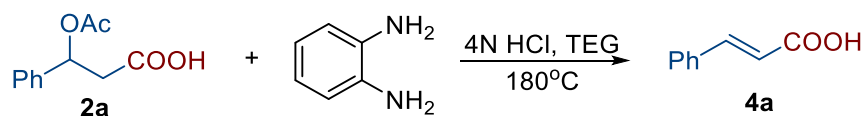
Yield 89% (212.4 mg, 1.28 mmol). Brownish oil.

¹H NMR (300.13 MHz, CDCl₃, δ): 7.44 – 7.27 (m, 5H), 6.66 – 5.45 (brs, 1H), 5.16 (dd, *J* = 8.9, 4.0 Hz, 1H), 2.85 (dd, *J* = 16.6, 8.9 Hz, 1H), 2.76 (dd, *J* = 16.6, 4.0 Hz, 1H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): δ 177.2, 142.2, 128.8 (2C), 128.2, 125.8 (2C), 70.4, 43.2.

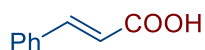
HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₉H₁₀O₃Na]⁺ 189.0522; Found 189.0535.

Synthesis 3-phenylacrylic acid, **4a**



A solution *o*-phenylenediamine (108.0 mg, 1.0 mmol) in 4N solution HCl (8 mL) was added to a solution of acid **2a** (250.0 mg, 1.2 mmol) in triethylene glycol (3 mL). The resulting mixture was refluxed at 180 °C for 6 h. Later the reaction mixture was cooled at 20 °C and adjusted to pH ~ 6 with aq. NaOH (5N). Extraction with CH₂Cl₂ (1:1, 5x5 mL) afforded the organic phase, which was washed with 10 mL brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure using a rotary evaporator (15–20 mmHg), (bath temperature, ca. 25–30 °C). The crude mixture was purified by column chromatography on silica gel (PE:EA = 2:1) to give the target product.

3-Phenylacrylic acid, **4a**²⁶



Yield 91% (161.4 mg, 1.09 mmol). Brownish powder.

¹H NMR (300.13 MHz, CDCl₃, δ): 11.22 (brs, 1H), 7.81 (d, *J* = 16.0 Hz, 1H), 7.57 (m, 2H), 7.49 – 7.35 (m, 3H), 6.47 (d, *J* = 16.0 Hz, 1H).

¹³C{¹H} NMR (75.48 MHz, CDCl₃, δ): δ 172.3, 147.2, 134.2, 130.9, 129.1 (2C), 128.5 (2C), 117.4.

HRMS (ESI/TOF) *m/z*: [M + Na]⁺ Calcd for [C₉H₈O₂Na]⁺ 171.0417; Found 171.0425.

Experimental Procedures for Scheme 5b.

A 5 mL electrolysis cell IKA ElectraSyn 2.0 containing zinc cathode and glassy carbon anode (30 mm × 7 mm × 0.1 mm) was charged with a solution of enol acetate **1a** (1.0 mmol, 162 mg), *n*Bu₄NClO₄ (0.5 mmol, 171 mg), TEOA (1.0 mmol, 149 mg) and phenanthrene (0.1 mmol, 17.8 mg) in DMF (3 mL). The resulting mixture was stirred and degassed by bubbling argon through the solvent for ca. 5 minutes. Then mixture electrolyzed at a constant current of 50 mA (*j* = 24 mA/cm²) with stirring at 20–25 °C for 32, 64, 129 or 194 min. Then, aq. HCl (5M, 4 mL) was added to the crude reaction mixture. Extraction with PE/EtOAc (1:1, 5×5 mL) afforded the organic phase, which was washed with 10mL brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure using a rotary evaporator (15–20 mmHg), (bath temperature, ca. 25–30 °C). The crude reaction mixture was studied by ¹H NMR.

CV study.

Cyclic voltammetry (CV) was implemented on an IPC-Pro M computer-assisted potentiostat manufactured by «Econix» (scan rate error 1.0%). The starting potential was set to 0.25 mV, and the initial sweep was carried out in the negative (cathode) region at a rate of 100 mV/s. The experiments were performed in a 10 mL fiveneck glass conic electrochemical cell with a water jacket for thermostating. CV curves were recorded using a three-electrode scheme. In a typical case, 2 mL of a solution was utilized. The working electrode was a glassy-carbon disk electrode ($d = 3$ mm, surface area ~ 0.07 cm²). A platinum wire served as an auxiliary electrode. An Ag/AgNO₃ electrode was used as the reference electrode and was linked to the solution by a porous glass diaphragm. The solutions were kept under thermally controlled conditions at 25 ± 0.5 °C and deaerated by bubbling argon. Electrochemical experiments were performed under an argon atmosphere. The working electrode was polished with figure-eight motions on a synthetic chamois leather pad using a Cr₂O₃-based polishing paste (~ 5 μm particle size) down to the mirror-like surface, and rinsed with acetonitrile. Polishing was carried before each recording of CV curve. In the case of testing CV with carbon dioxide, CO₂ was used instead of argon.

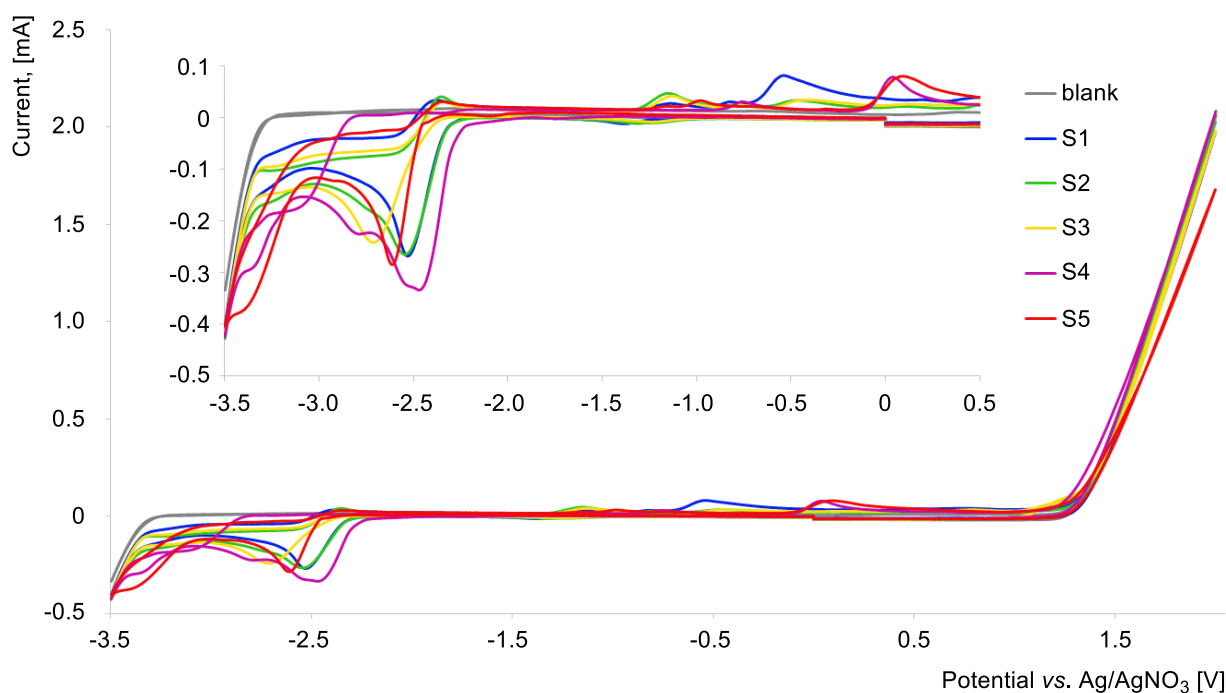


Figure S1. CV curves on a working glassy-carbon electrode ($d = 3$ mm) under a scan rate of 0.1 V/s for (grey) blank, (blue) acetophenone **S1** (0.02 M), TMS-enol ether **S2** (0.02 M), TBS-enol ether **S3** (0.02 M), vinyl tosylate **S4** (0.02 M), enol acetate **S5** (0.02 M) in 0.1 M *n*Bu₄NClO₄ solution in DMF.

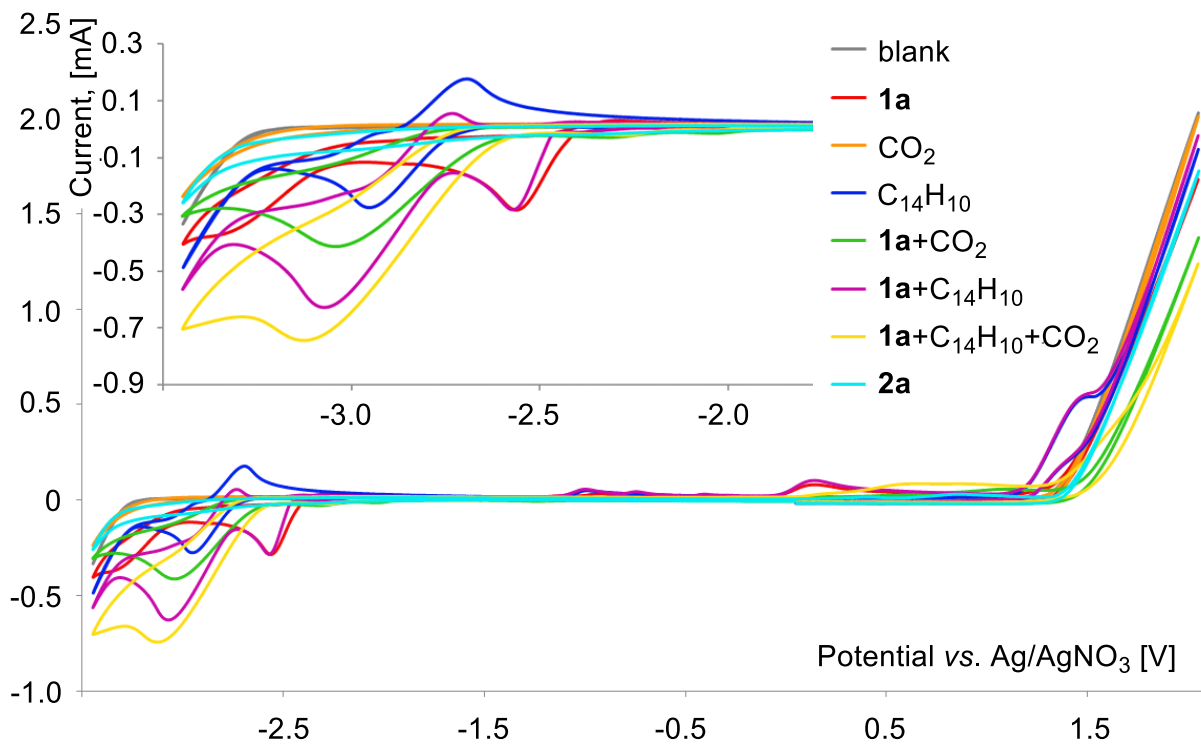


Figure S2. CV curves on a working glassy-carbon electrode ($d = 3$ mm) under a scan rate of 0.1 V/s for (grey) blank, (red) enol acetate **1a** (0.02 M), (orange) CO₂, (blue) Phenanthrene (0.02 M), (green) enol acetate **1a** (0.02 M) with CO₂, (purple) enol acetate **1a** (0.02 M) with phenanthrene (0.02 M), (yellow) enol acetate **1a** (0.02 M) with CO₂ and phenanthrene (0.02 M), (turquoise) 3-acetoxy-3-phenyl-propanoic acid **2a** (0.02 M) in 0.1 M $n\text{Bu}_4\text{NClO}_4$ solution in DMF.

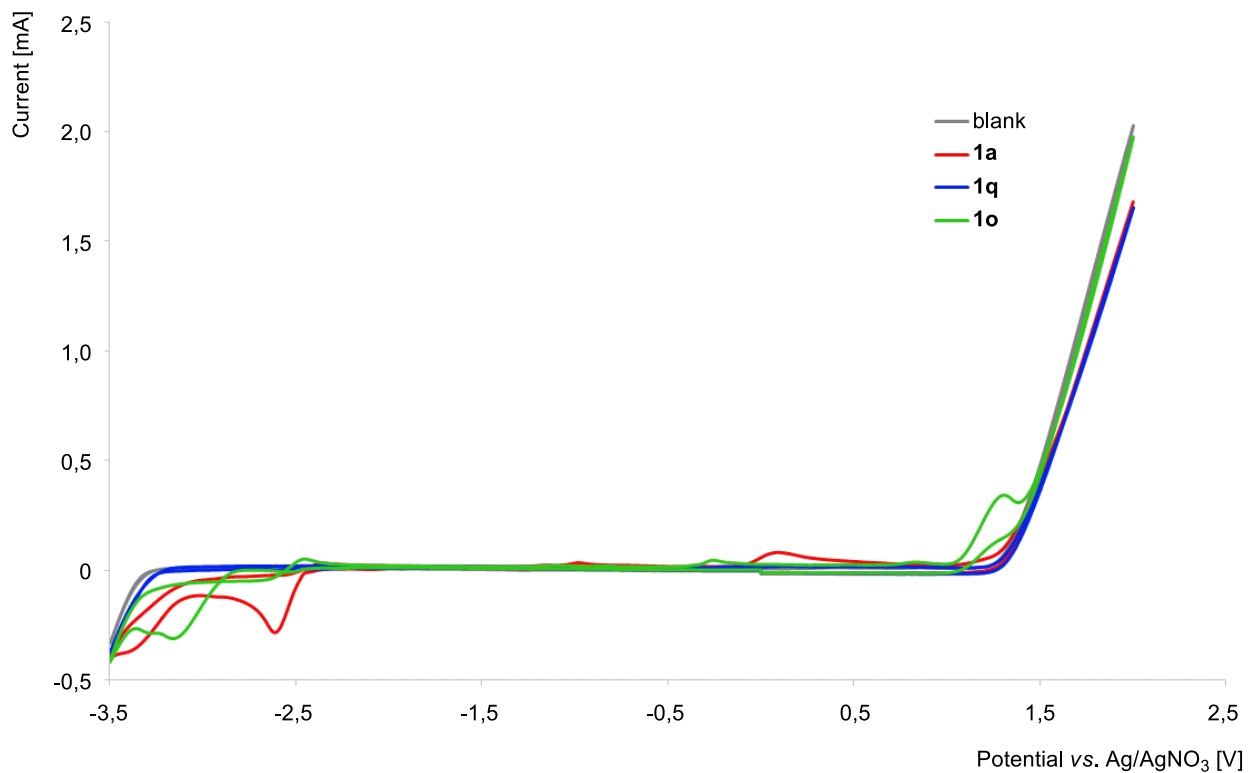


Figure S3. CV curves on a working glassy-carbon electrode ($d = 3$ mm) under a scan rate of 0.1 V/s for (grey) blank, (red) enol acetate **1a** (0.02 M), (blue) enol acetate **1q** (0.02 M), (green) enol acetate **1o** (0.02 M) in 0.1 M $n\text{Bu}_4\text{NClO}_4$ solution in DMF.

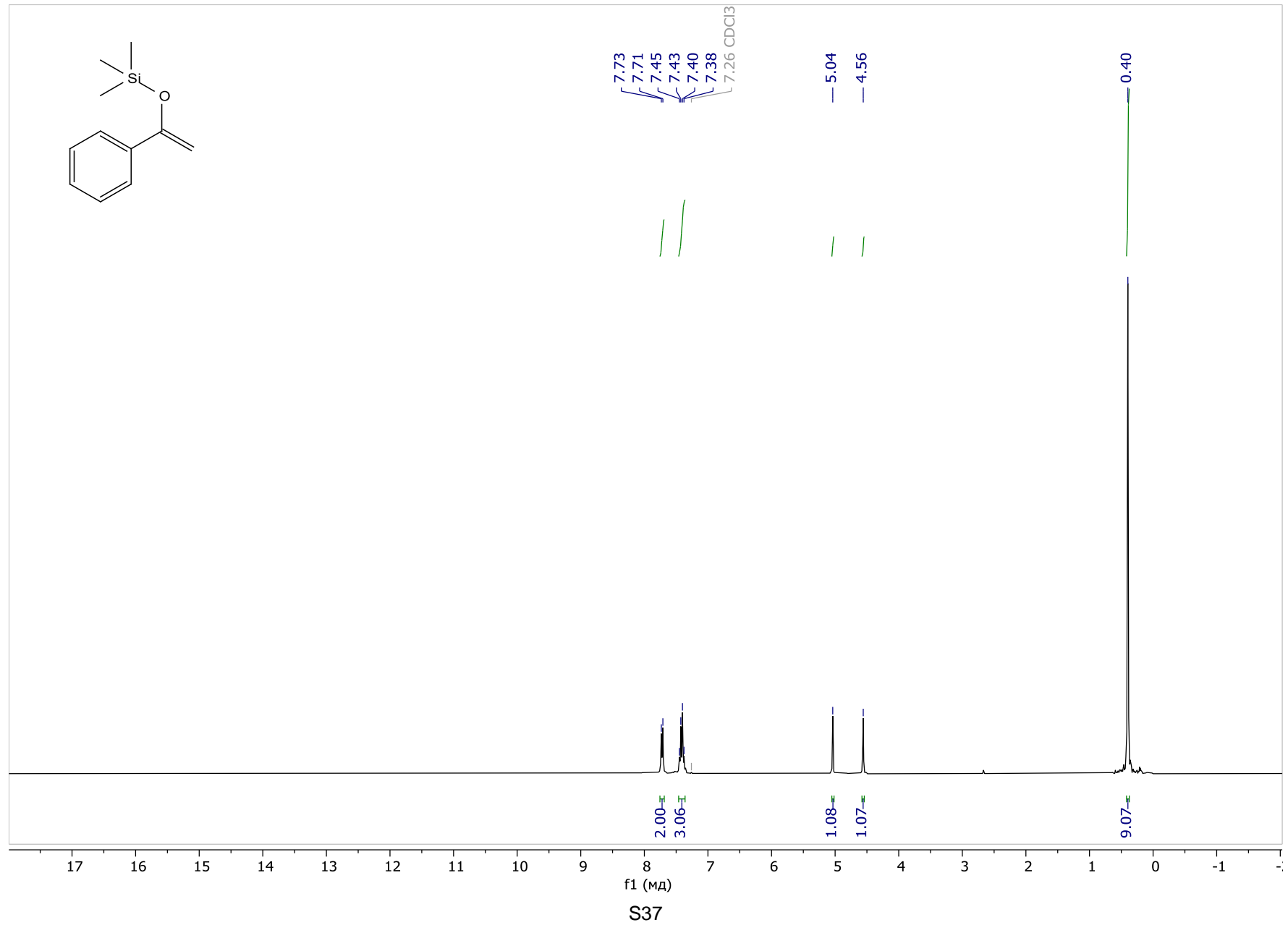
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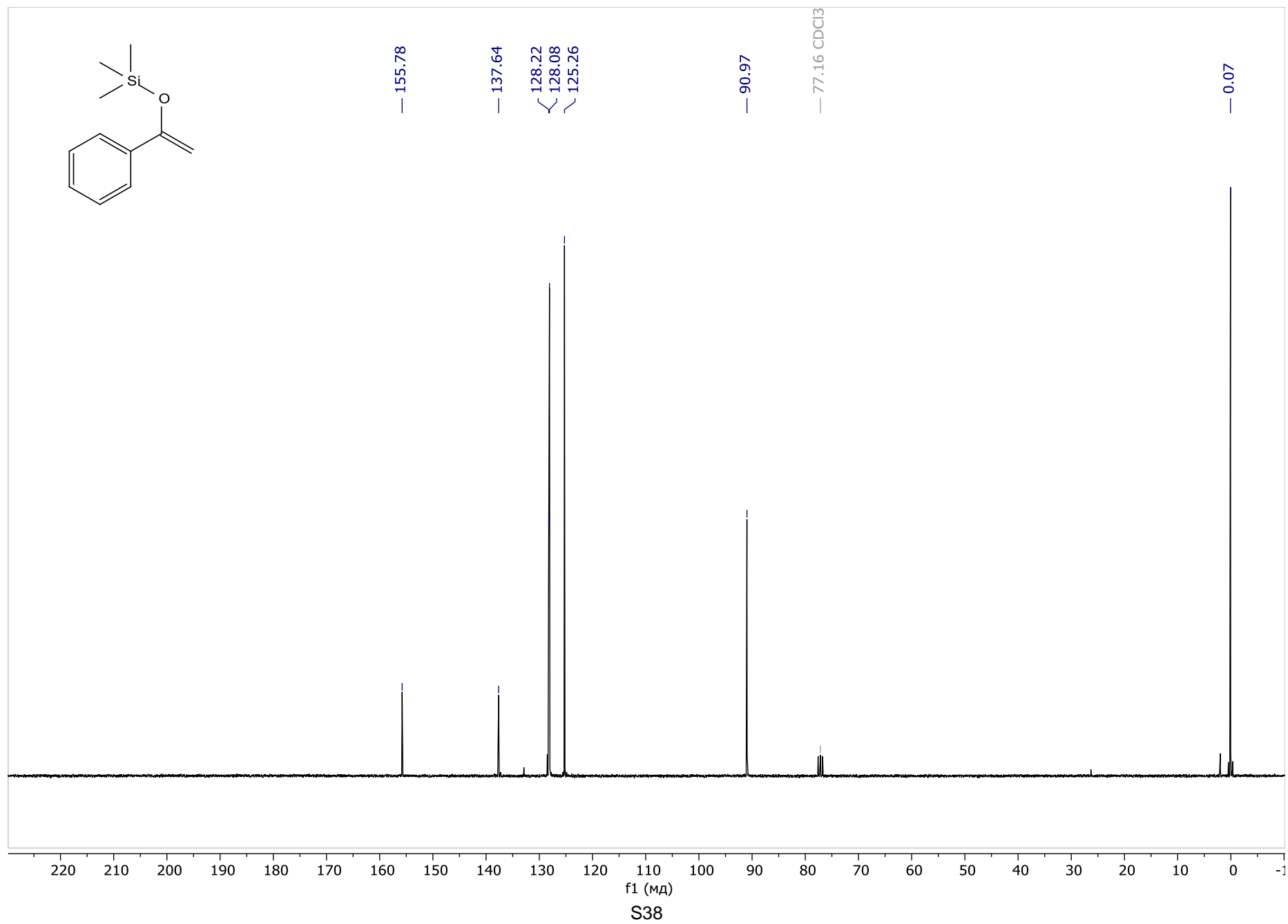
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NMR spectra of the starting enol derivatives.

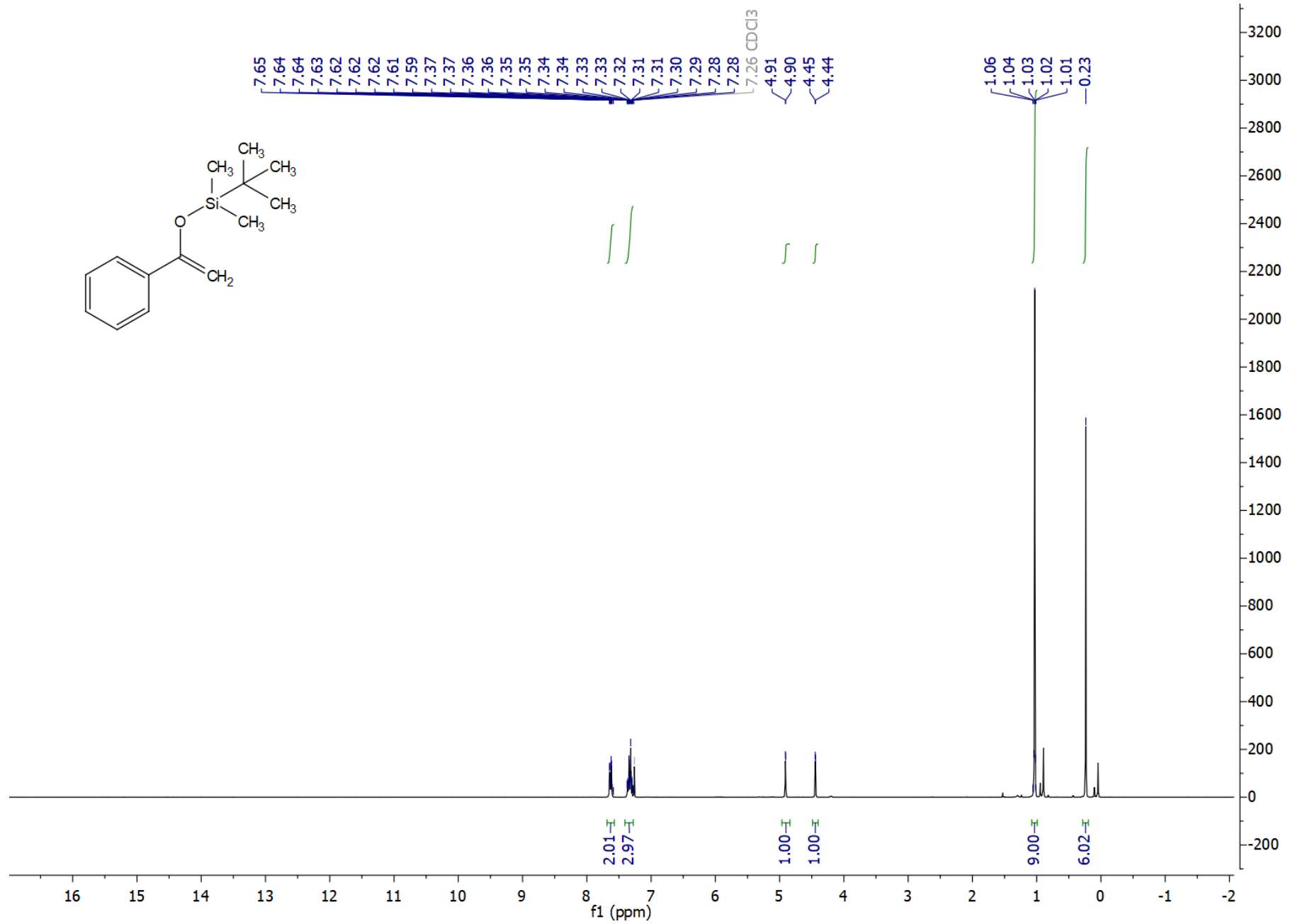
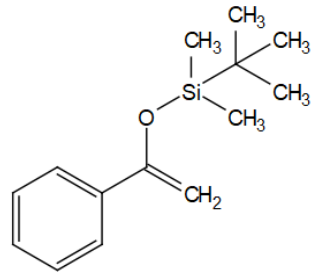
^1H NMR (300.13 MHz, CDCl_3), Trimethyl((1-phenylvinyl)oxy)silane, S2



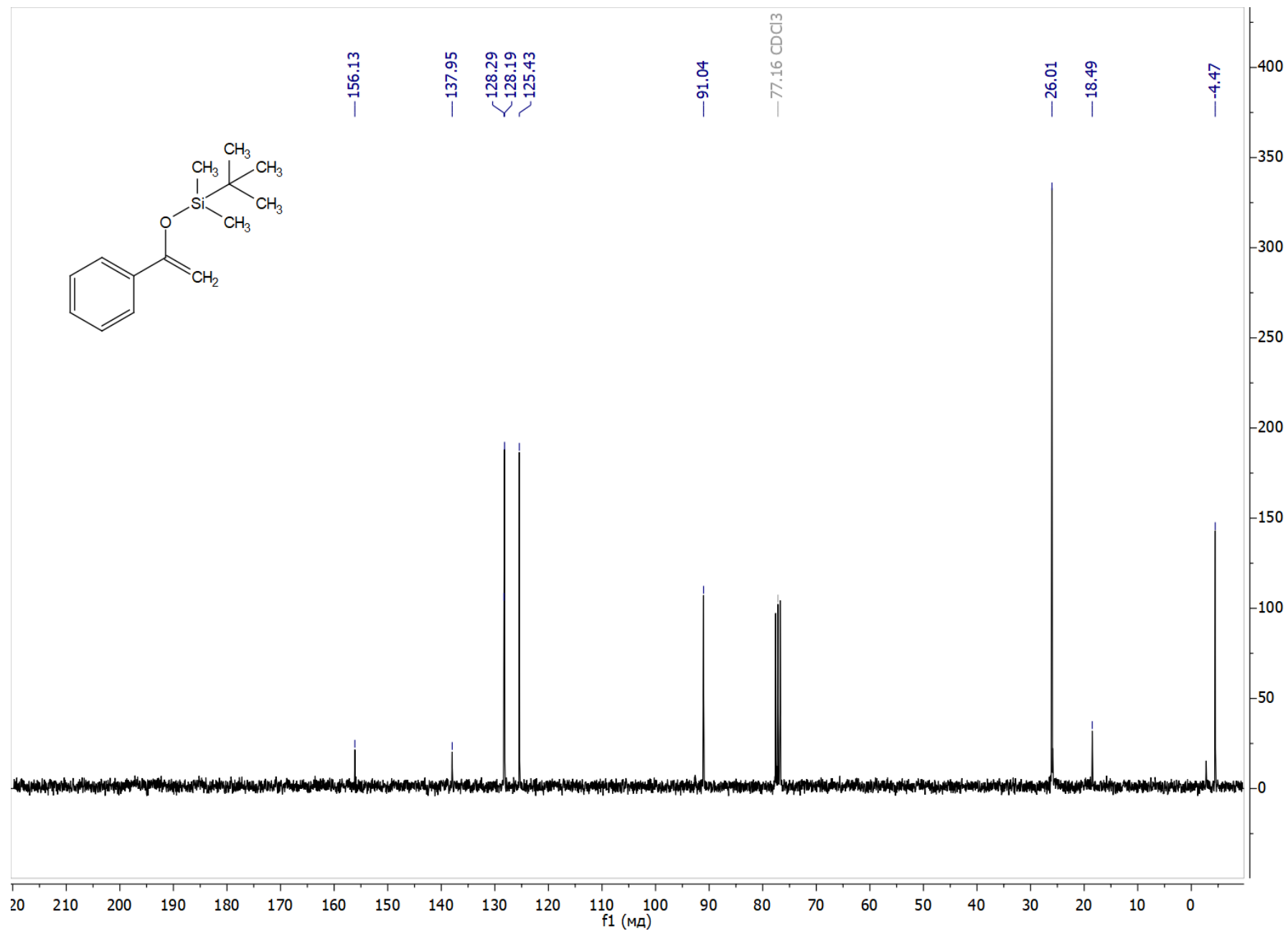
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), Trimethyl((1-phenylvinyl)oxy)silane, S2



¹H NMR (300.13 MHz, CDCl₃), *Tert*-butyldimethyl((1-phenylvinyl)oxy)silane, S3

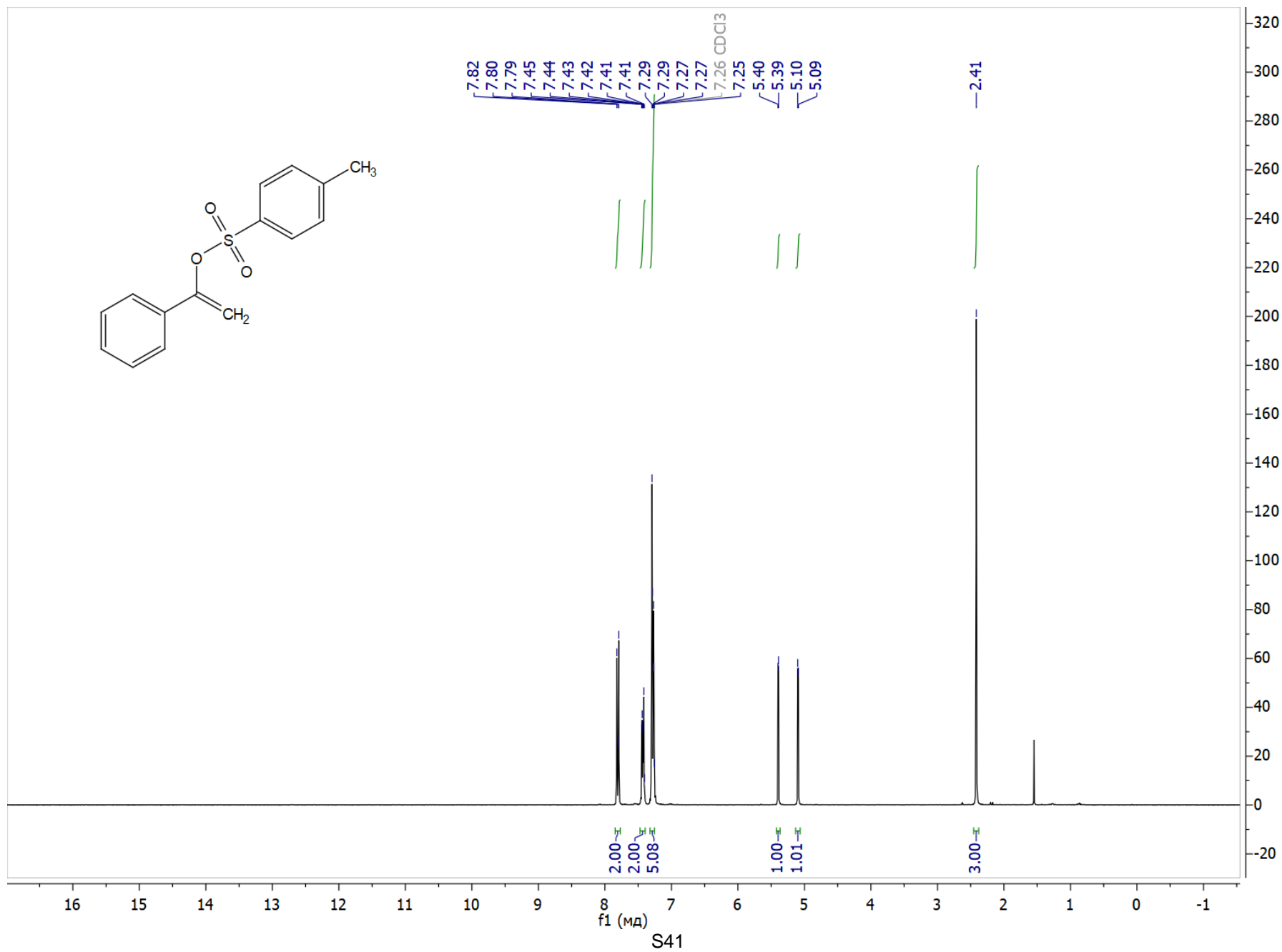


$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), ***Tert*-butyldimethyl((1-phenylvinyl)oxy)silane, S3**

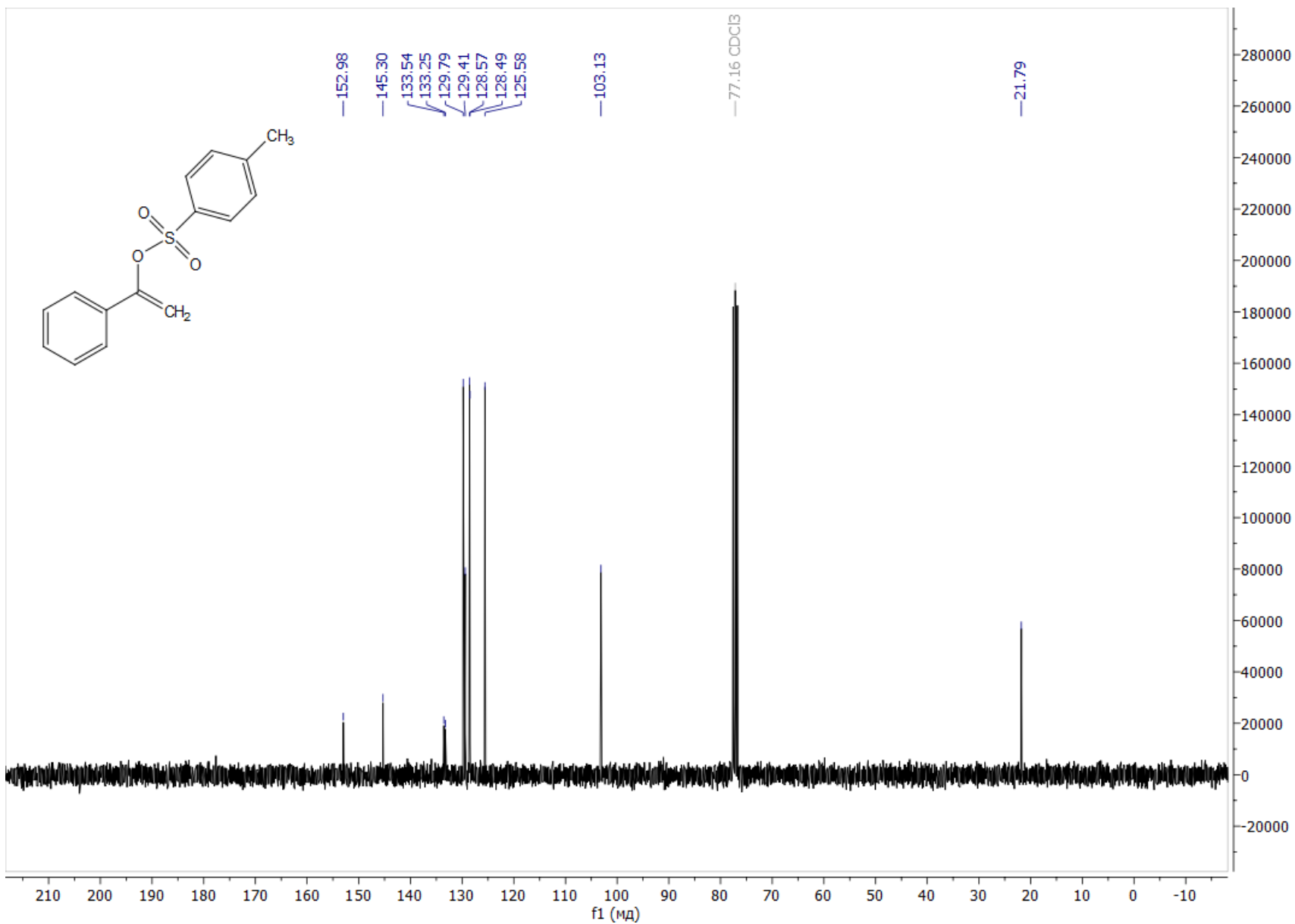


S40

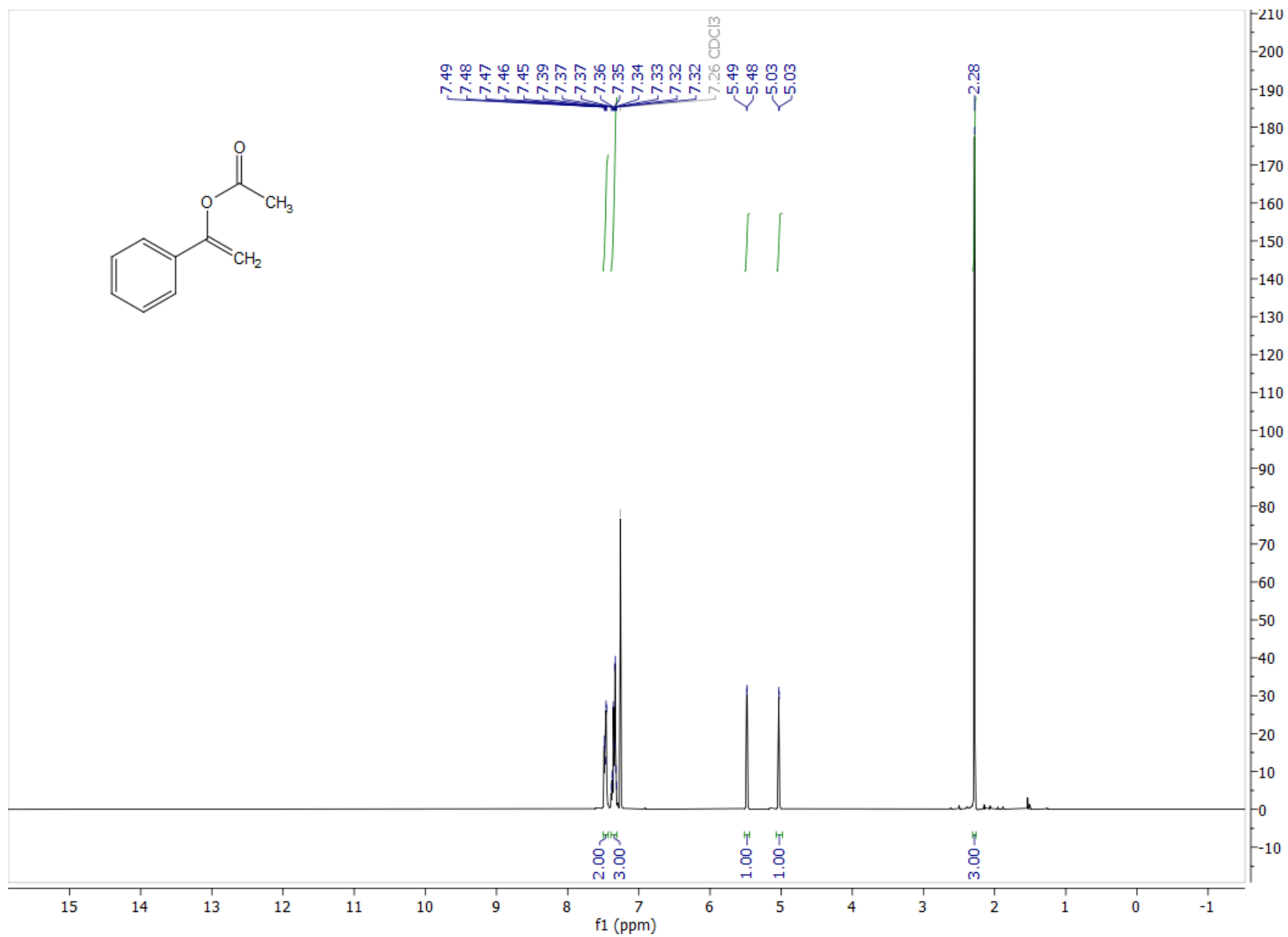
¹H NMR (300.13 MHz, CDCl₃), 1-Phenylvinyl 4-methylbenzenesulfonate, S4



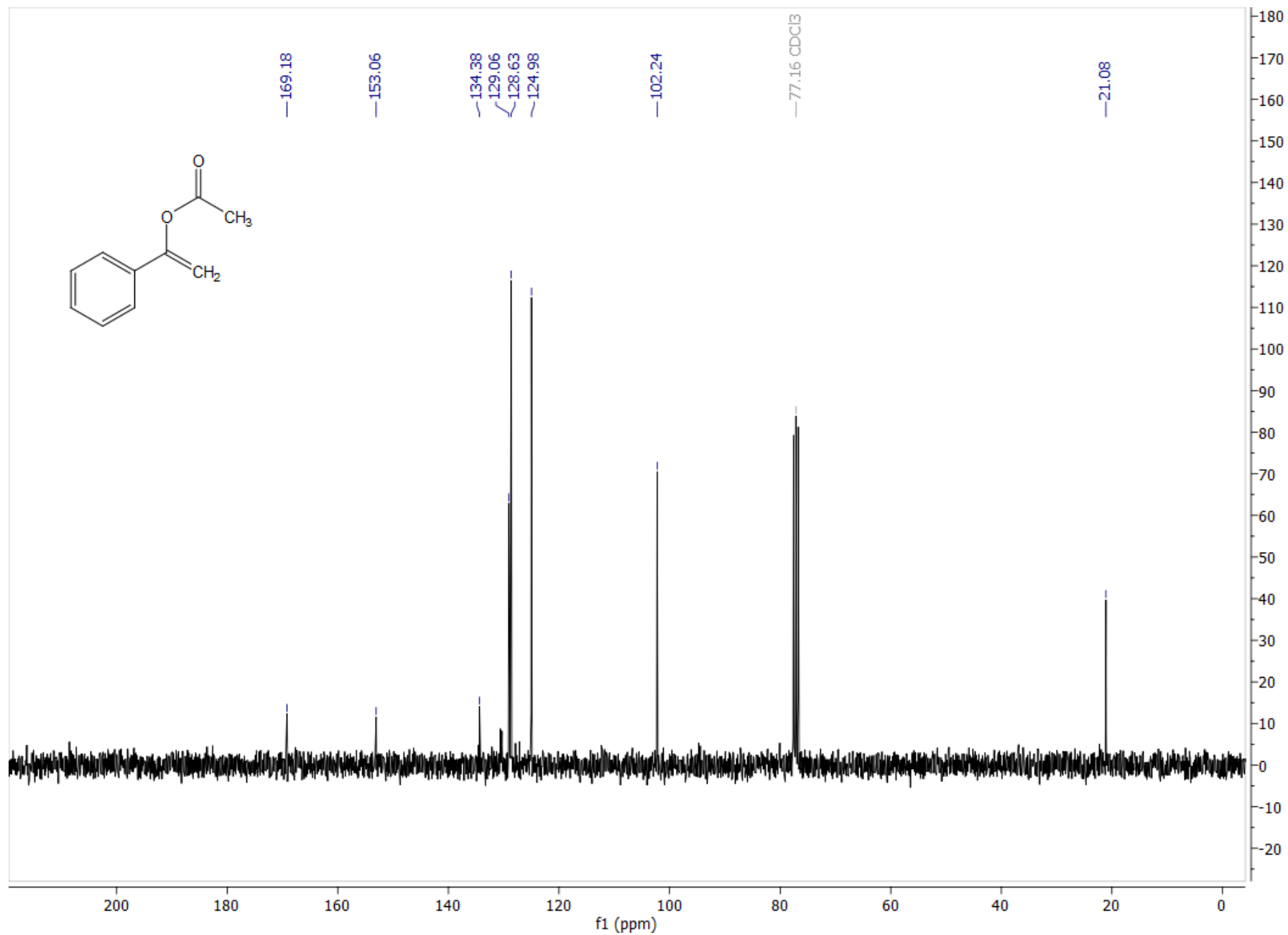
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-Phenylvinyl 4-methylbenzenesulfonate, S4



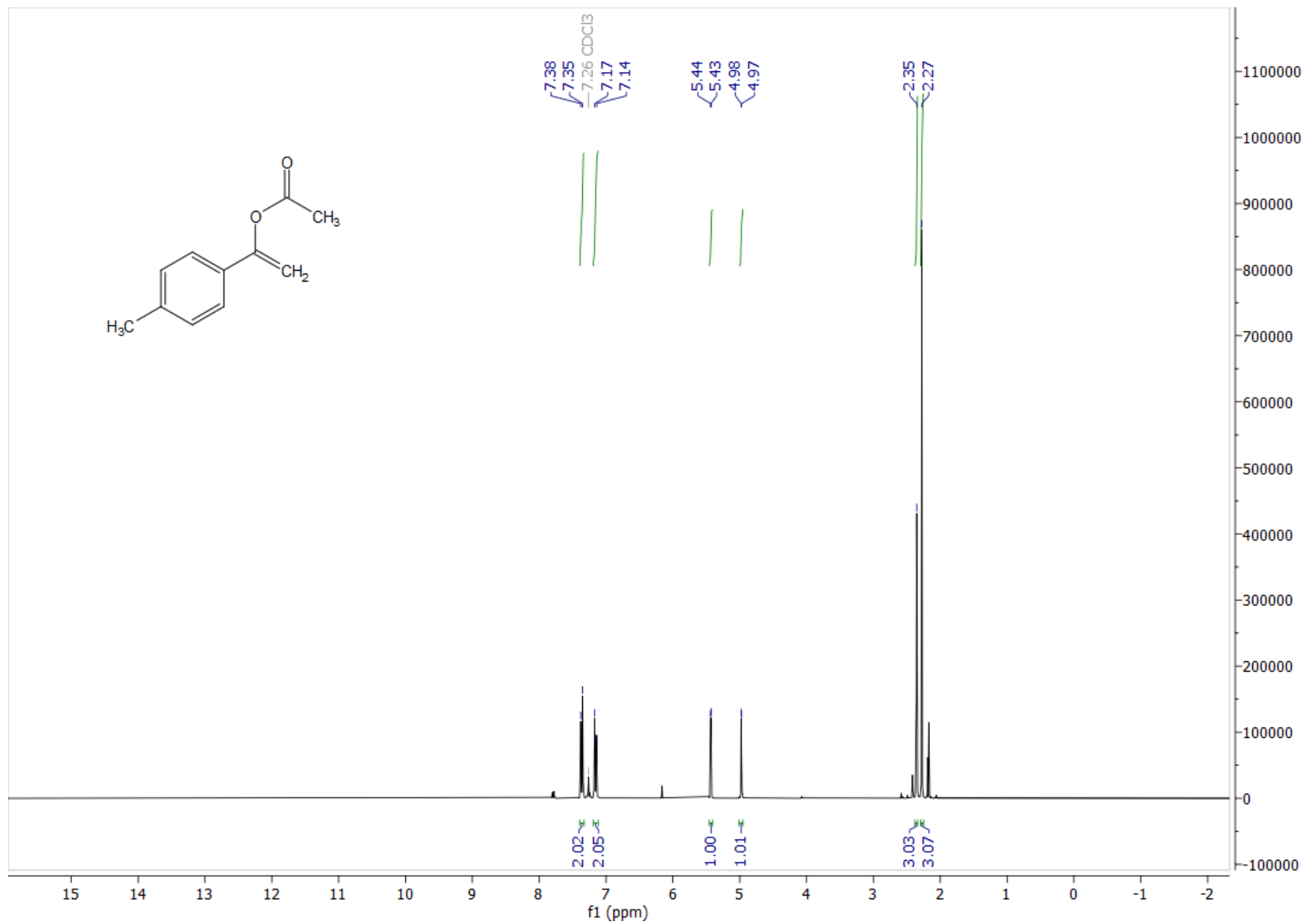
¹H NMR (300.13 MHz, CDCl₃), 1-Phenylvinyl acetate, 1a



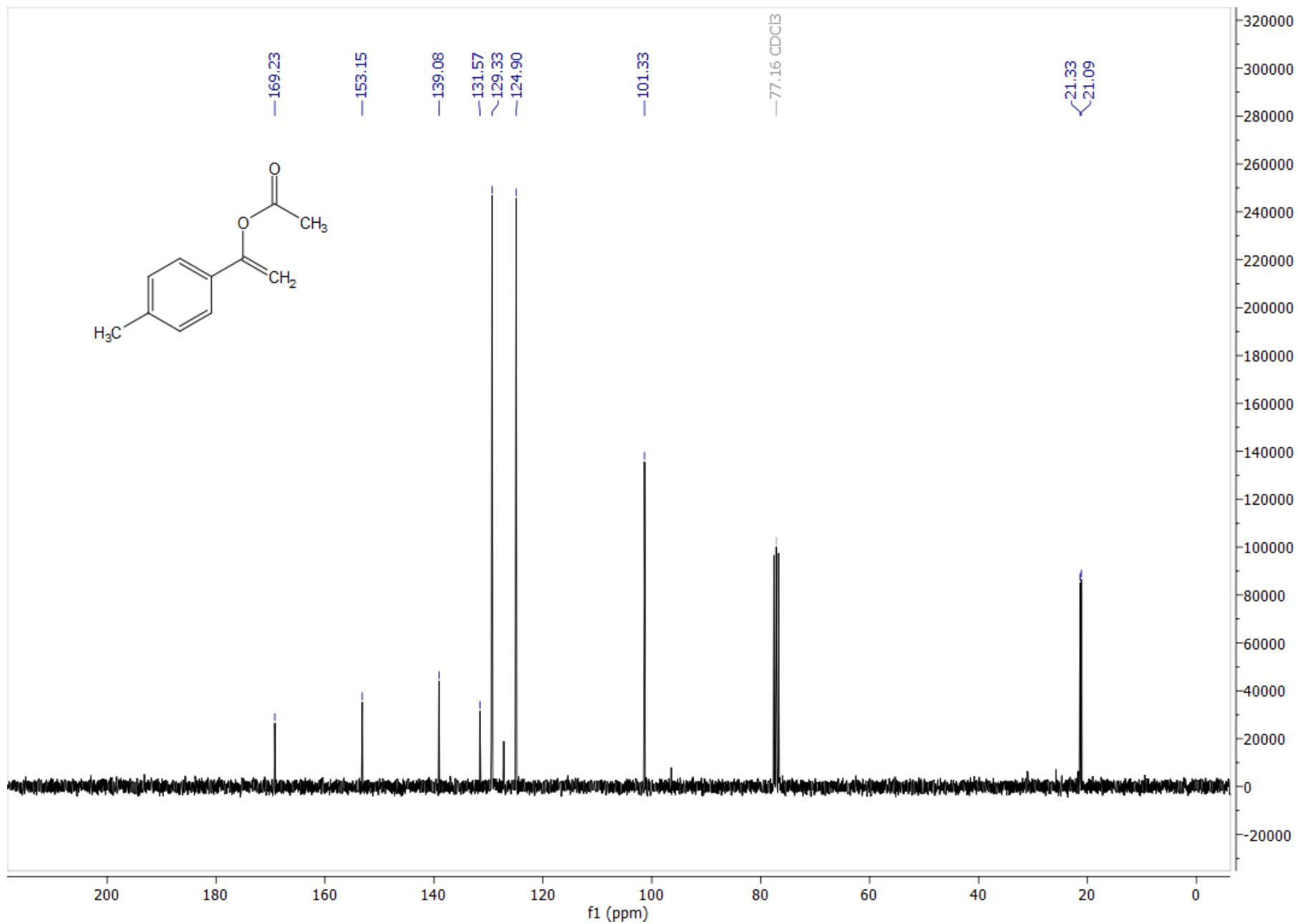
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-Phenylvinyl acetate, 1a



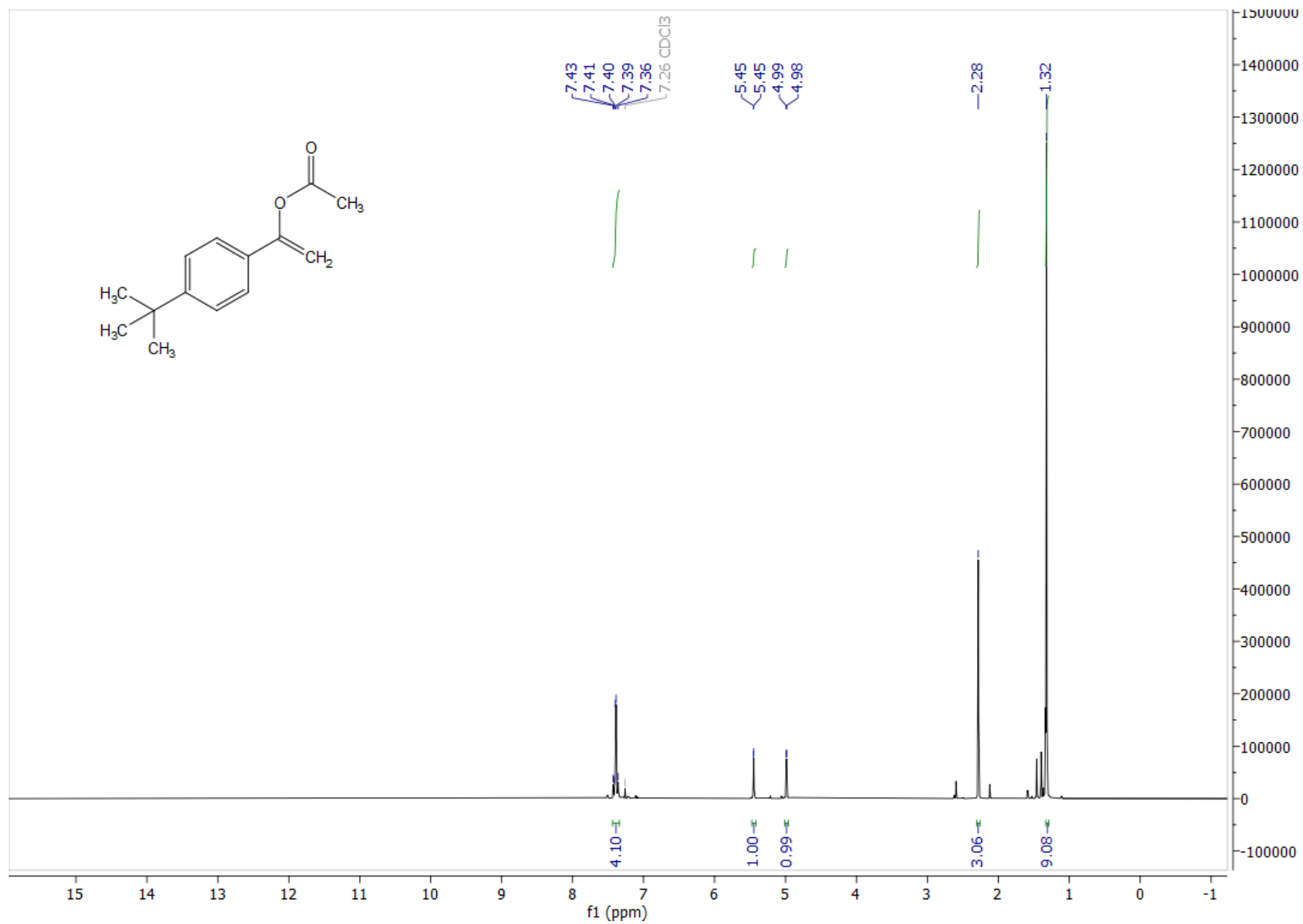
¹H NMR (300.13 MHz, CDCl₃), 1-(*p*-Tolyl)vinyl acetate, 1b



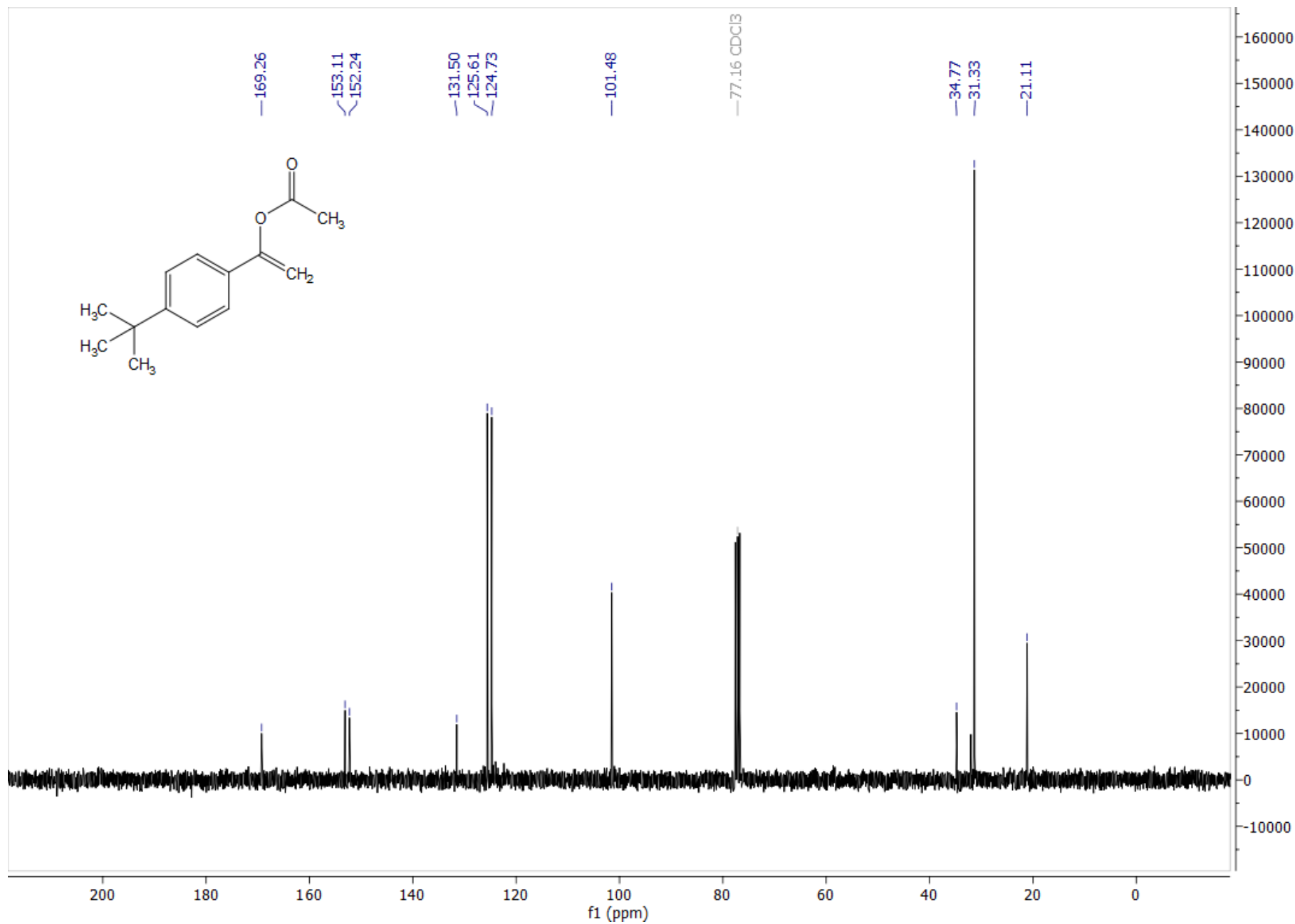
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(*p*-Tolyl)vinyl acetate, **1b**



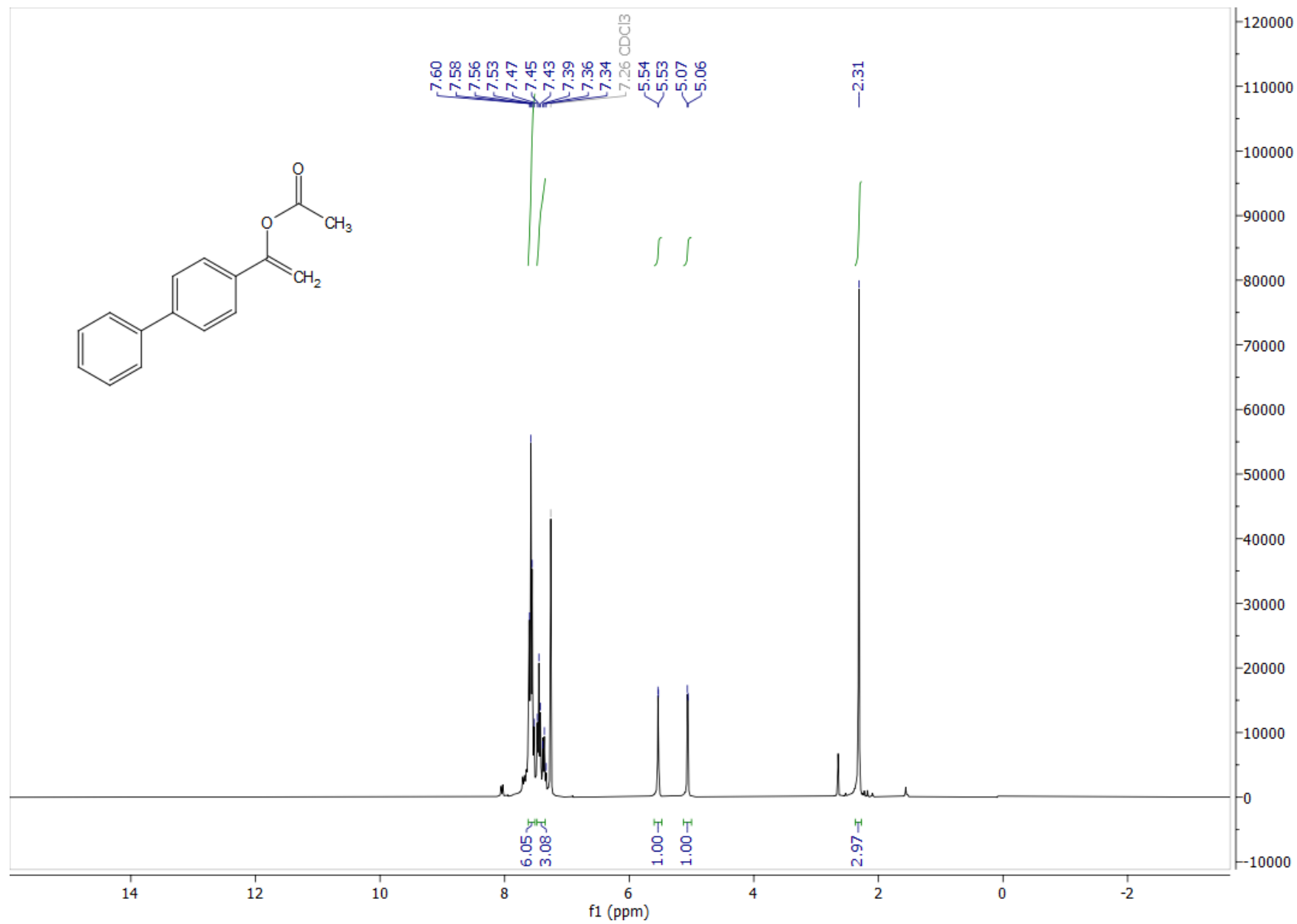
¹H NMR (300.13 MHz, CDCl₃), 1-(4-(*Tert*-butyl)phenyl)vinyl acetate, **1c**



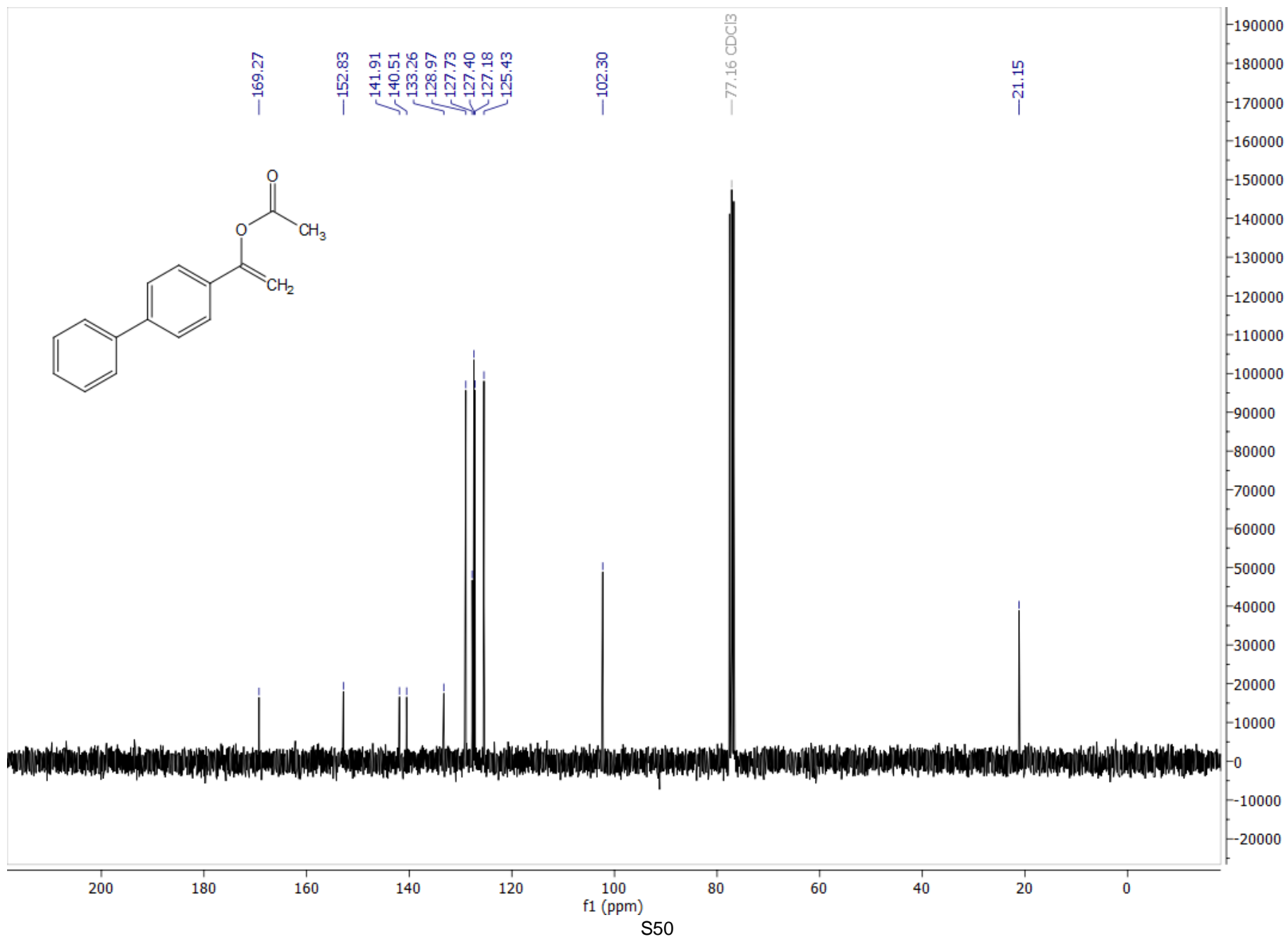
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(4-(*Tert*-butyl)phenyl)vinyl acetate, 1c



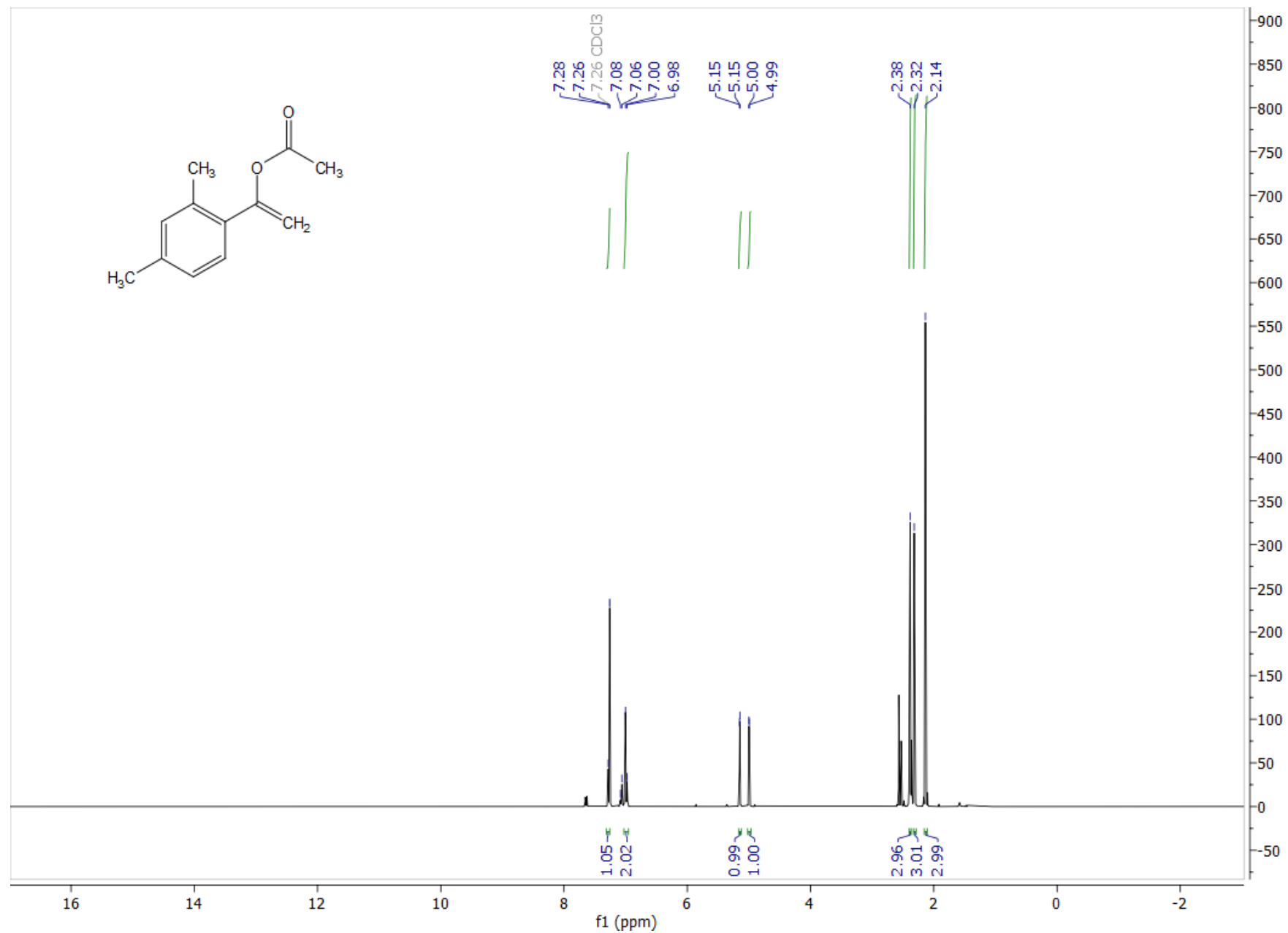
¹H NMR (300.13 MHz, CDCl₃), 1-([1,1'-Biphenyl]-4-yl)vinyl acetate, 1d



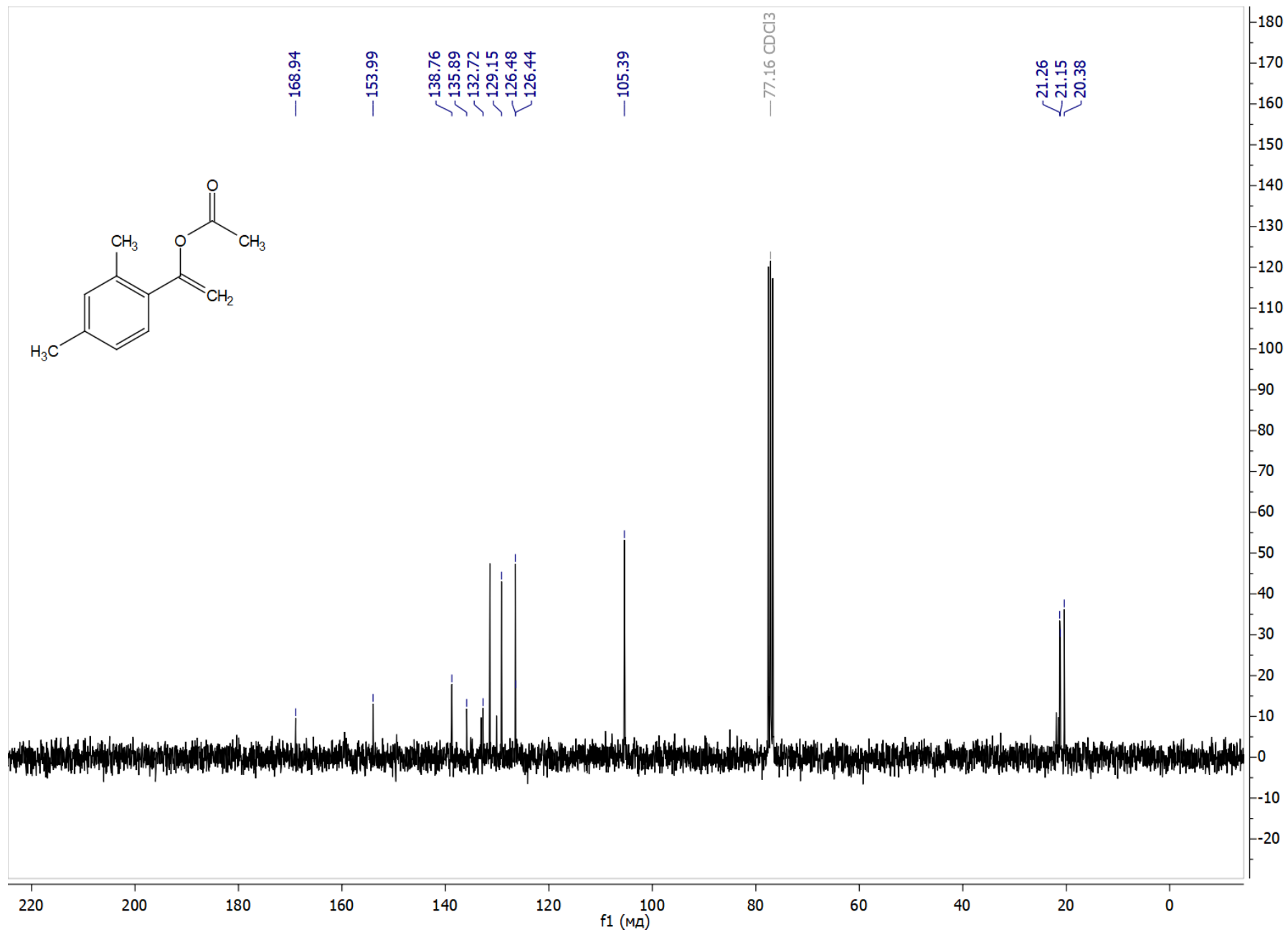
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-([1,1'-Biphenyl]-4-yl)vinyl acetate, 1d



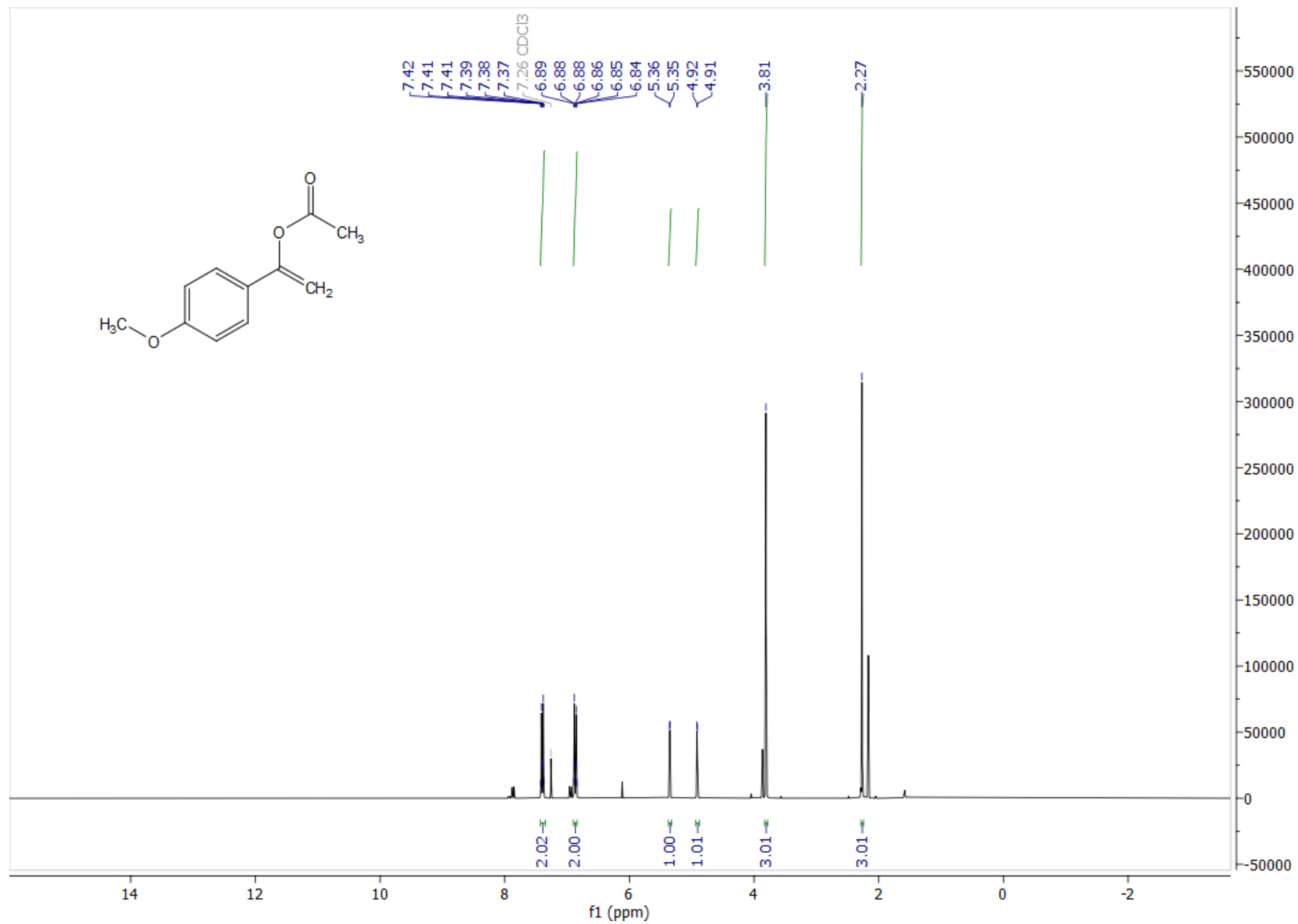
¹H NMR (300.13 MHz, CDCl₃), 1-(2,4-Dimethylphenyl)vinyl acetate 1e



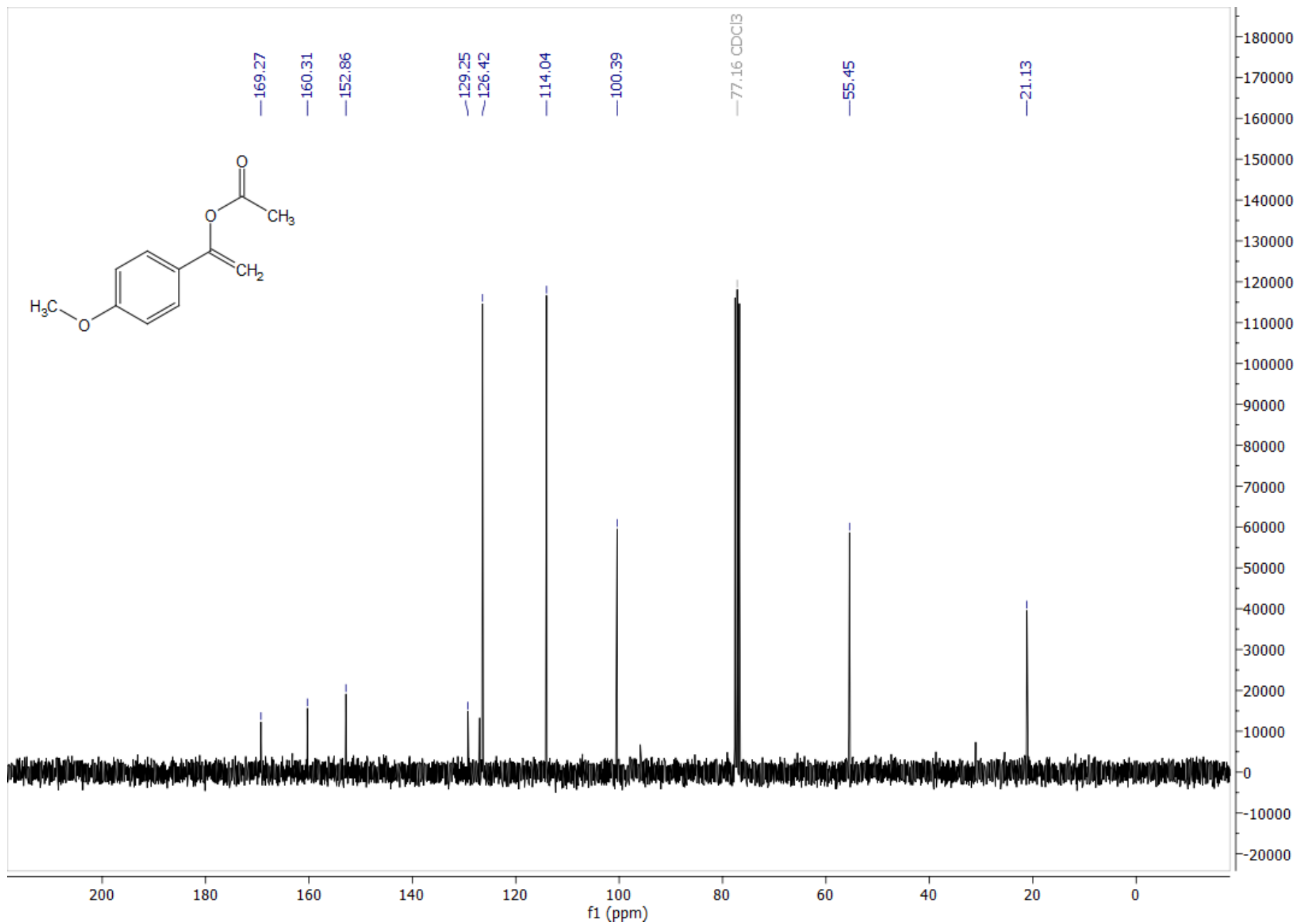
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(2,4-Dimethylphenyl)vinyl acetate, **1e**



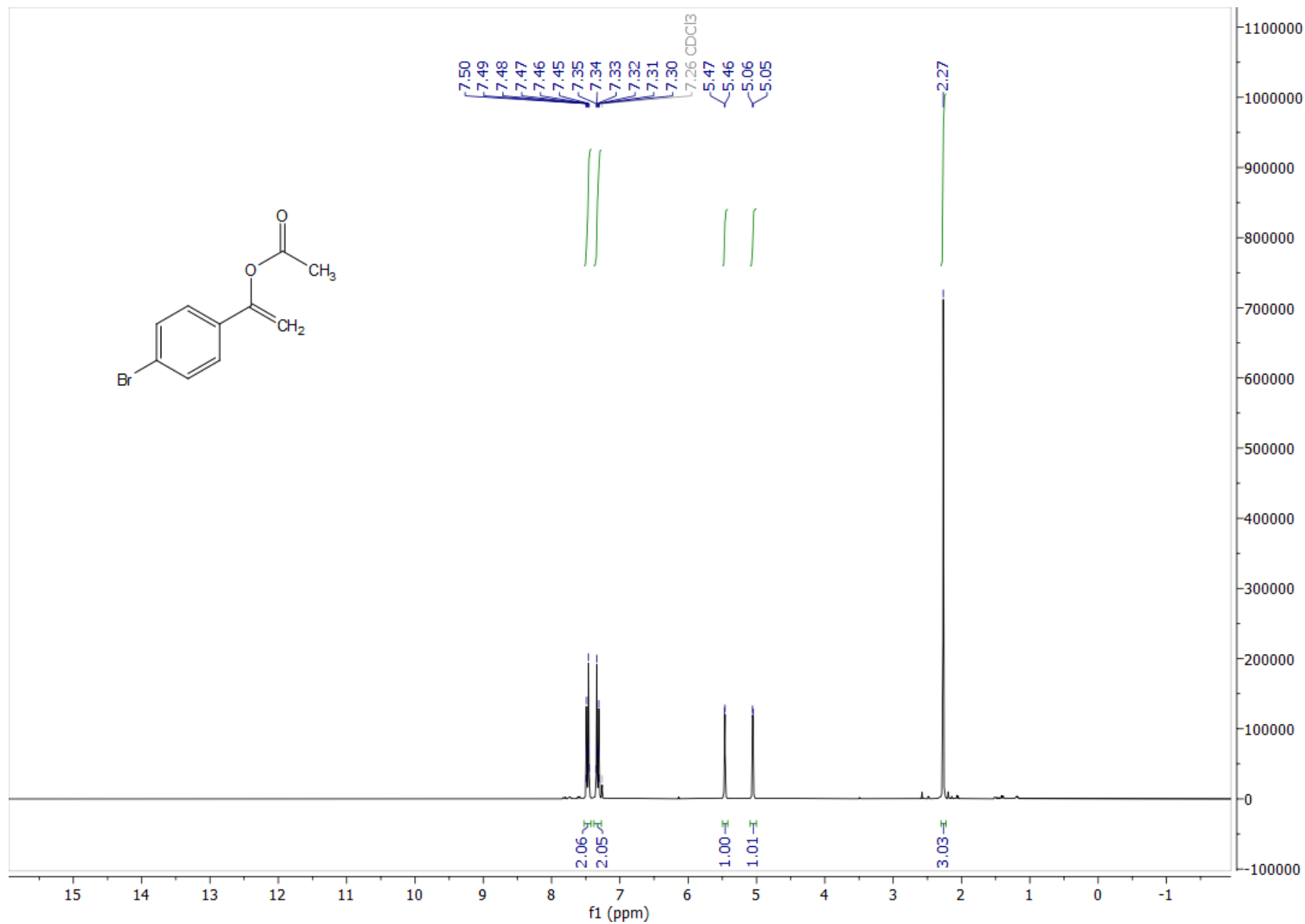
¹H NMR (300.13 MHz, CDCl₃), 1-(4-Methoxyphenyl)vinyl acetate, 1f



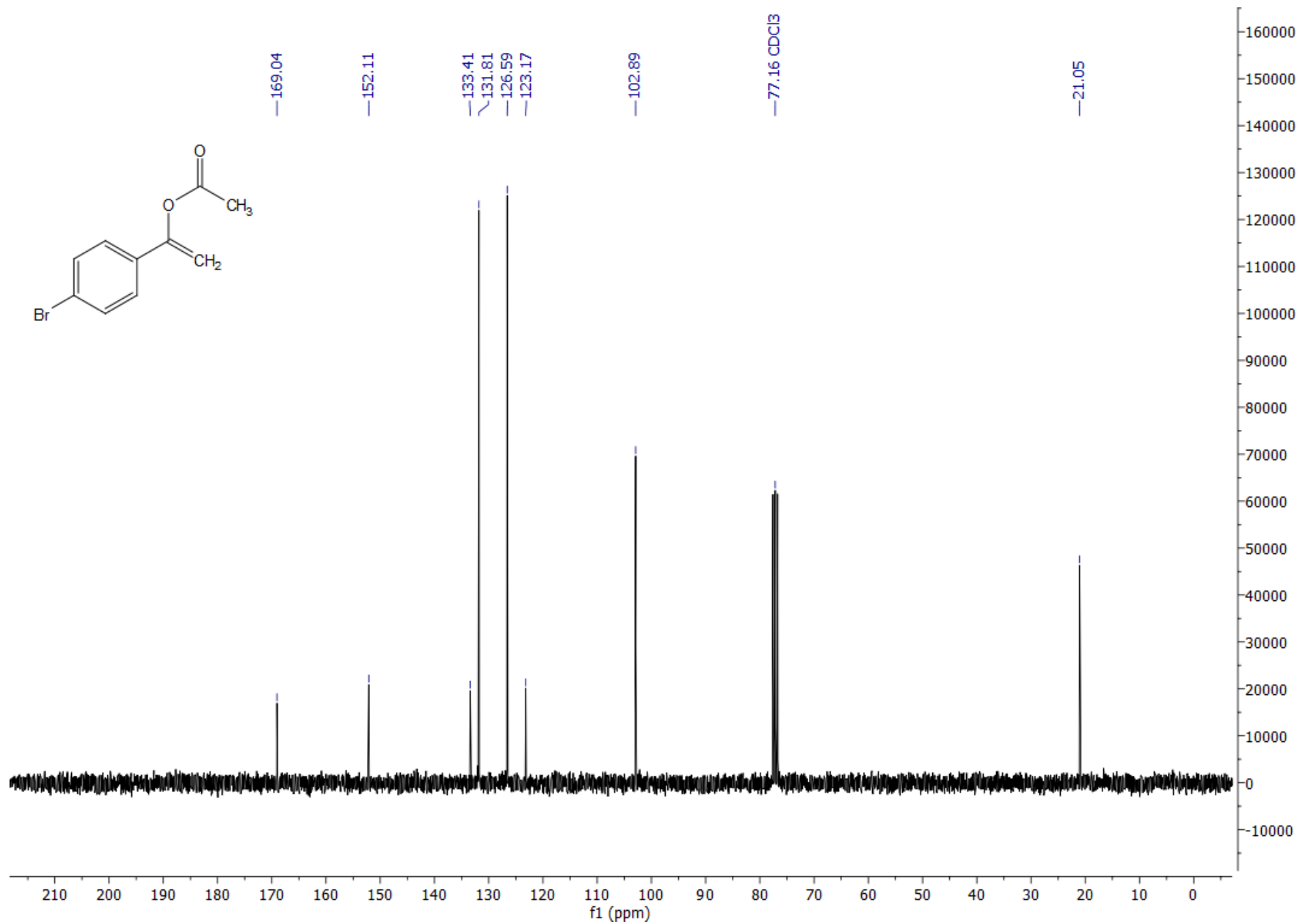
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(4-Methoxyphenyl)vinyl acetate, **1f**



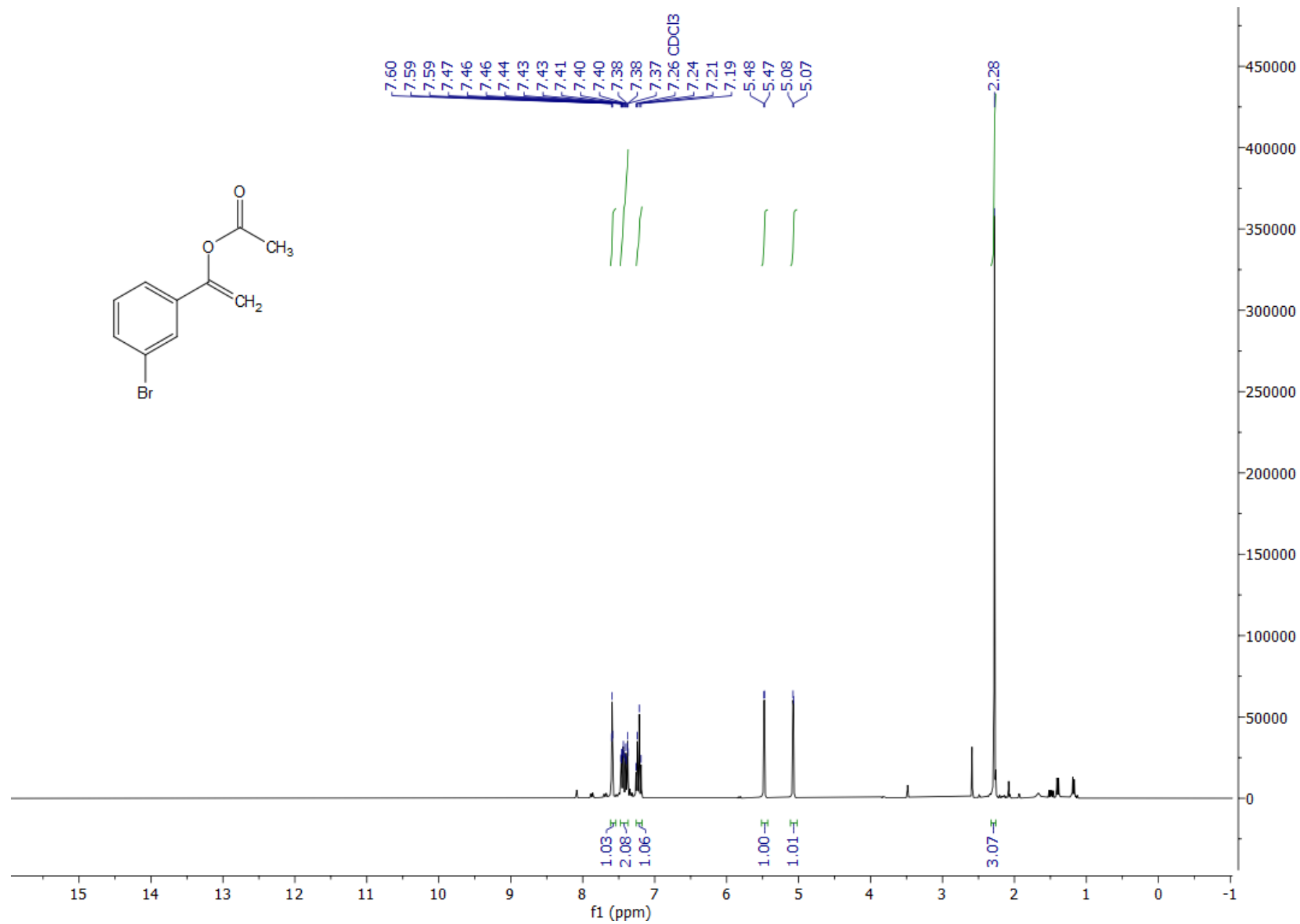
¹H NMR (300.13 MHz, CDCl₃), 1-(4-Bromophenyl)vinyl acetate, 1r



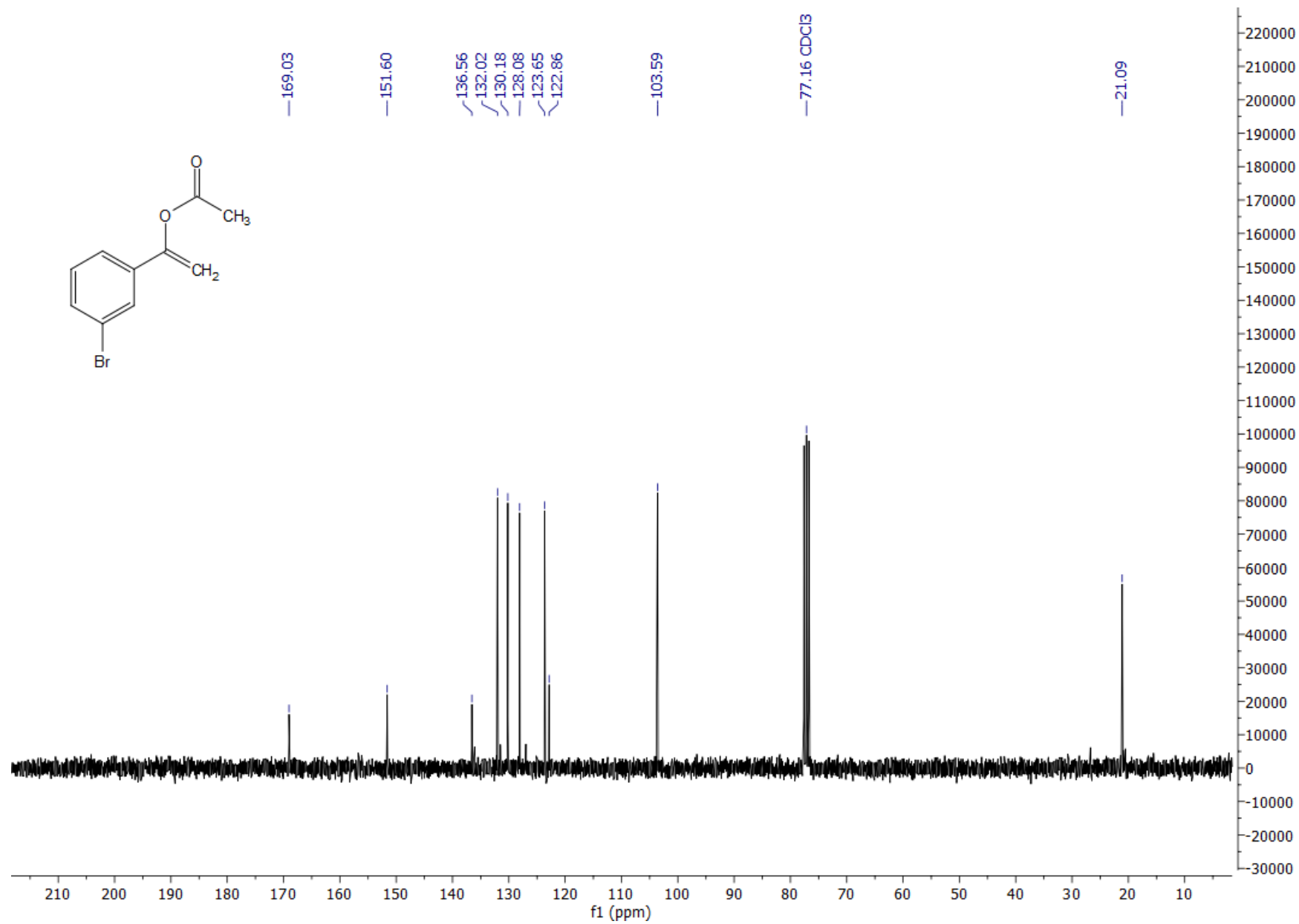
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(4-Bromophenyl)vinyl acetate, 1r



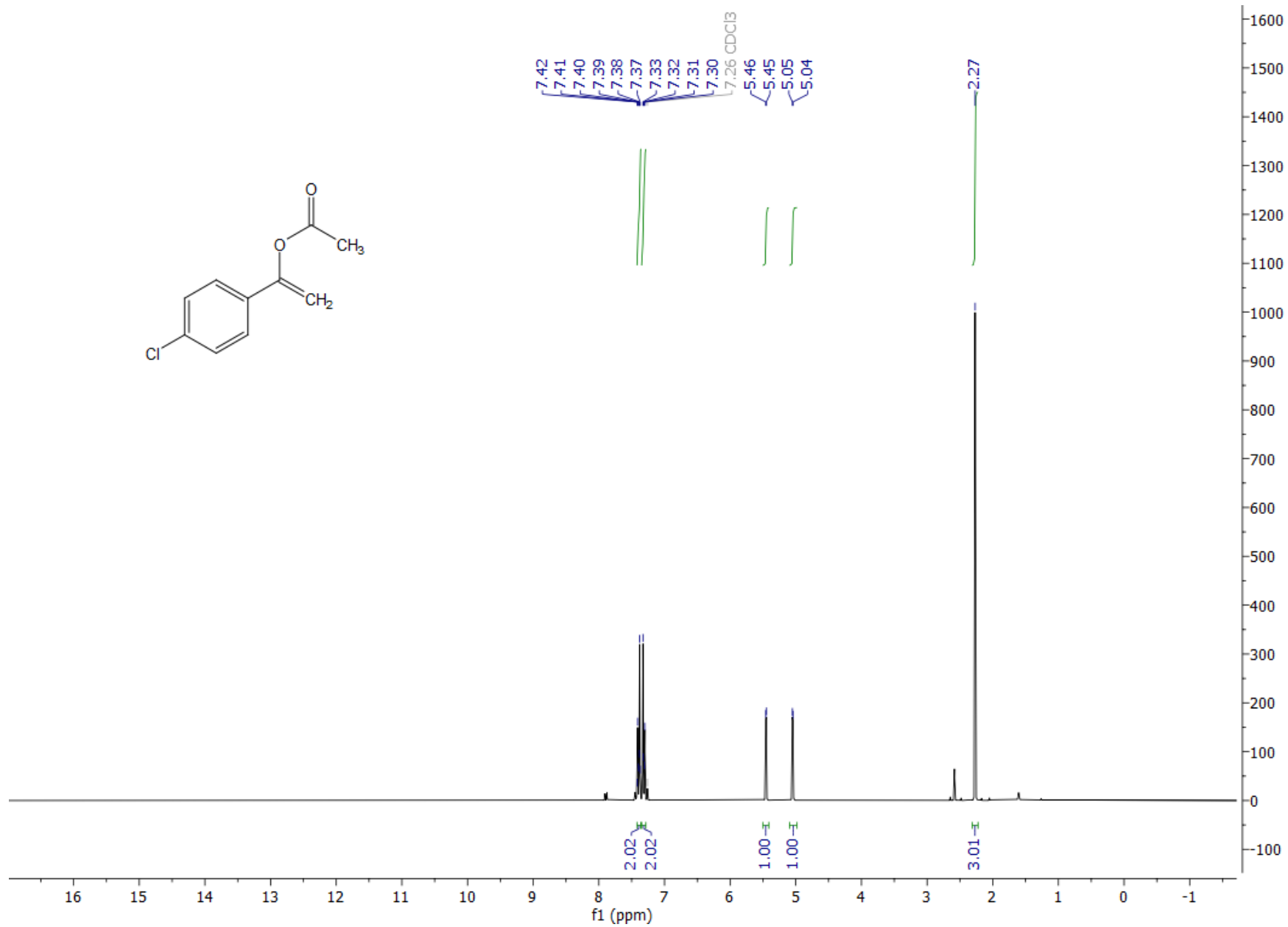
¹H NMR (300.13 MHz, CDCl₃), 1-(3-Bromophenyl)vinyl acetate, 1s



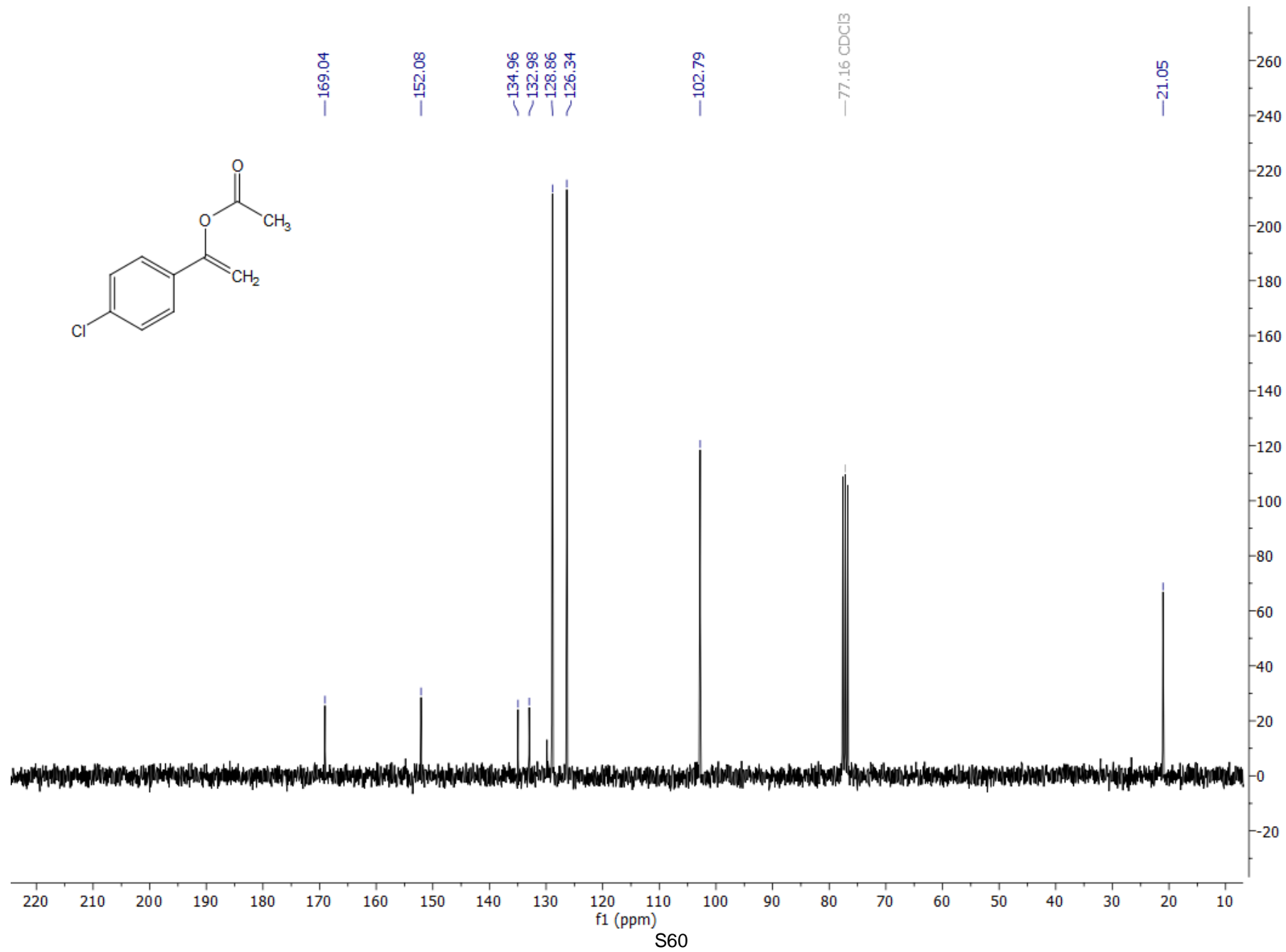
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(3-Bromophenyl)vinyl acetate, **1s**



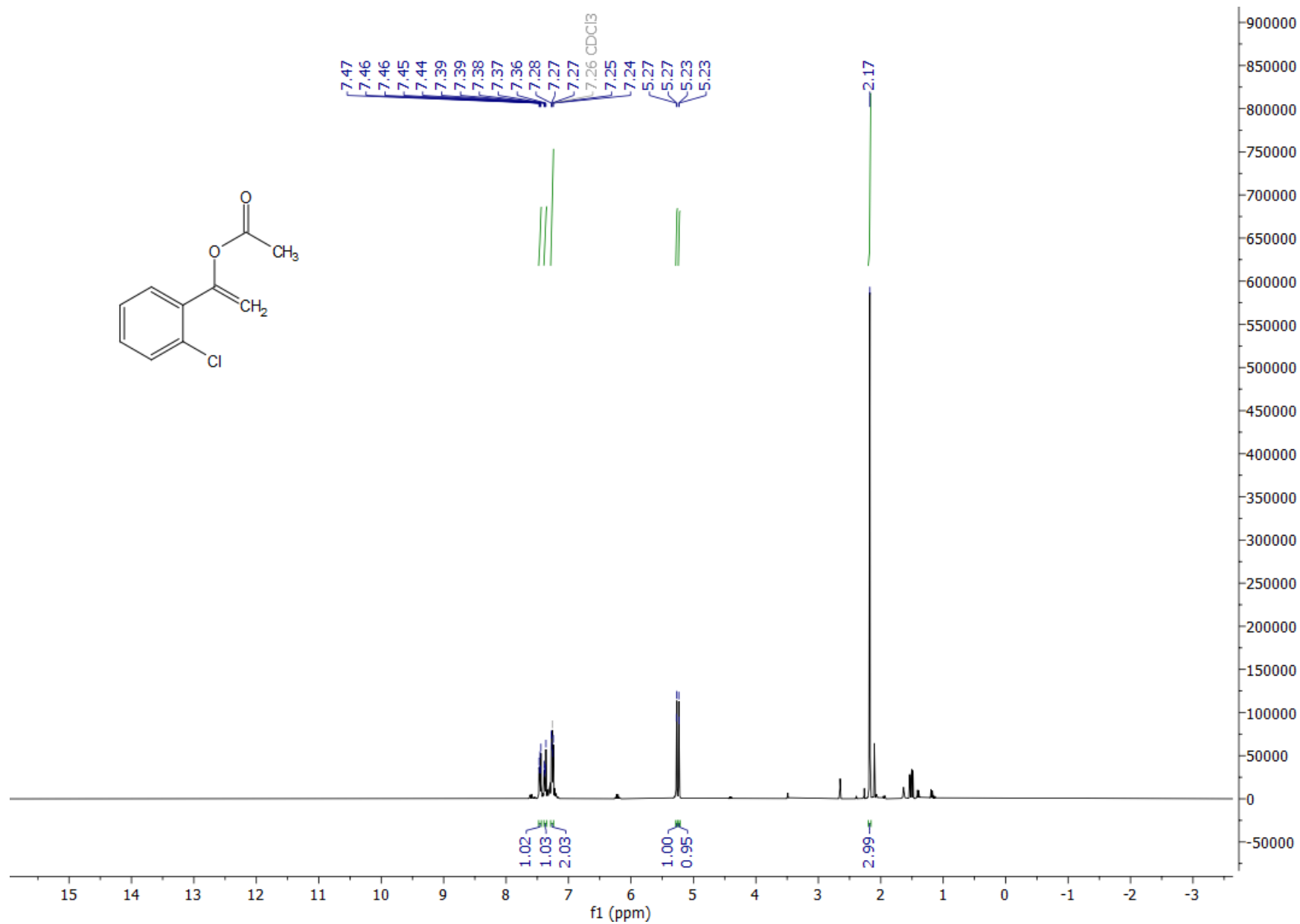
¹H NMR (300.13 MHz, CDCl₃), 1-(4-Chlorophenyl)vinyl acetate, 1u



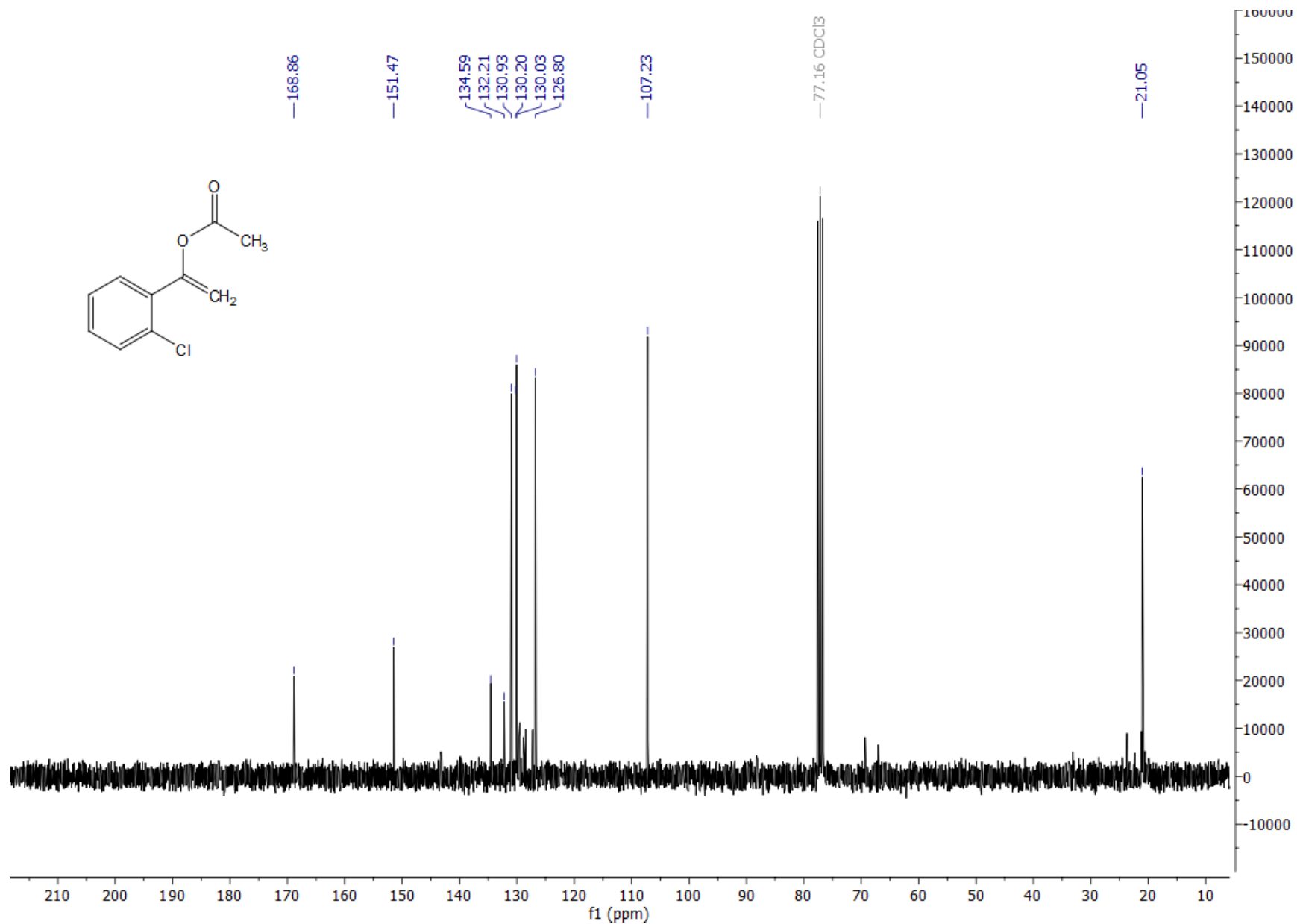
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(4-Chlorophenyl)vinyl acetate, **1u**



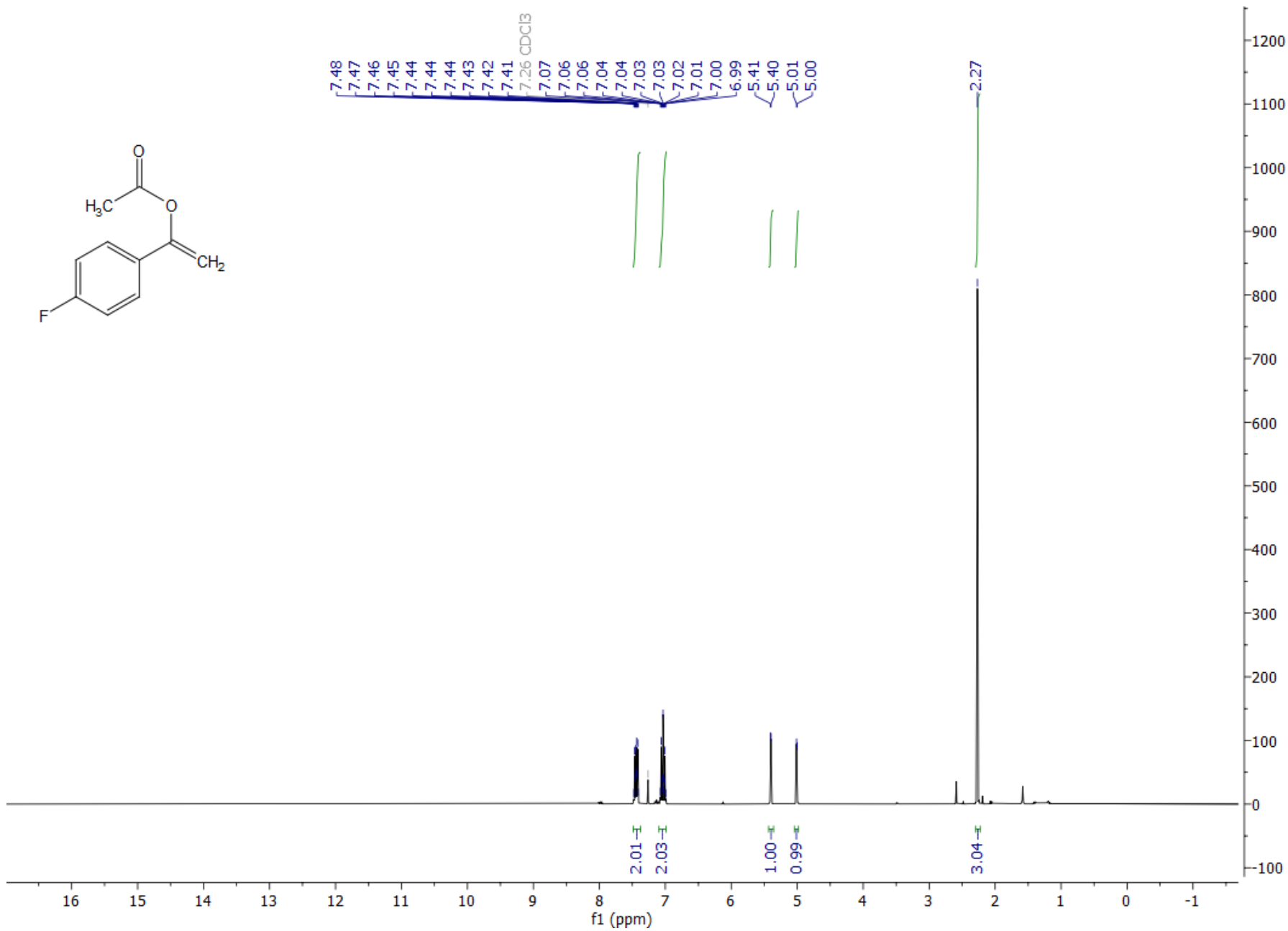
¹H NMR (300.13 MHz, CDCl₃), 1-(2-Chlorophenyl)vinyl acetate, 1v



$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(2-Chlorophenyl)vinyl acetate, 1v

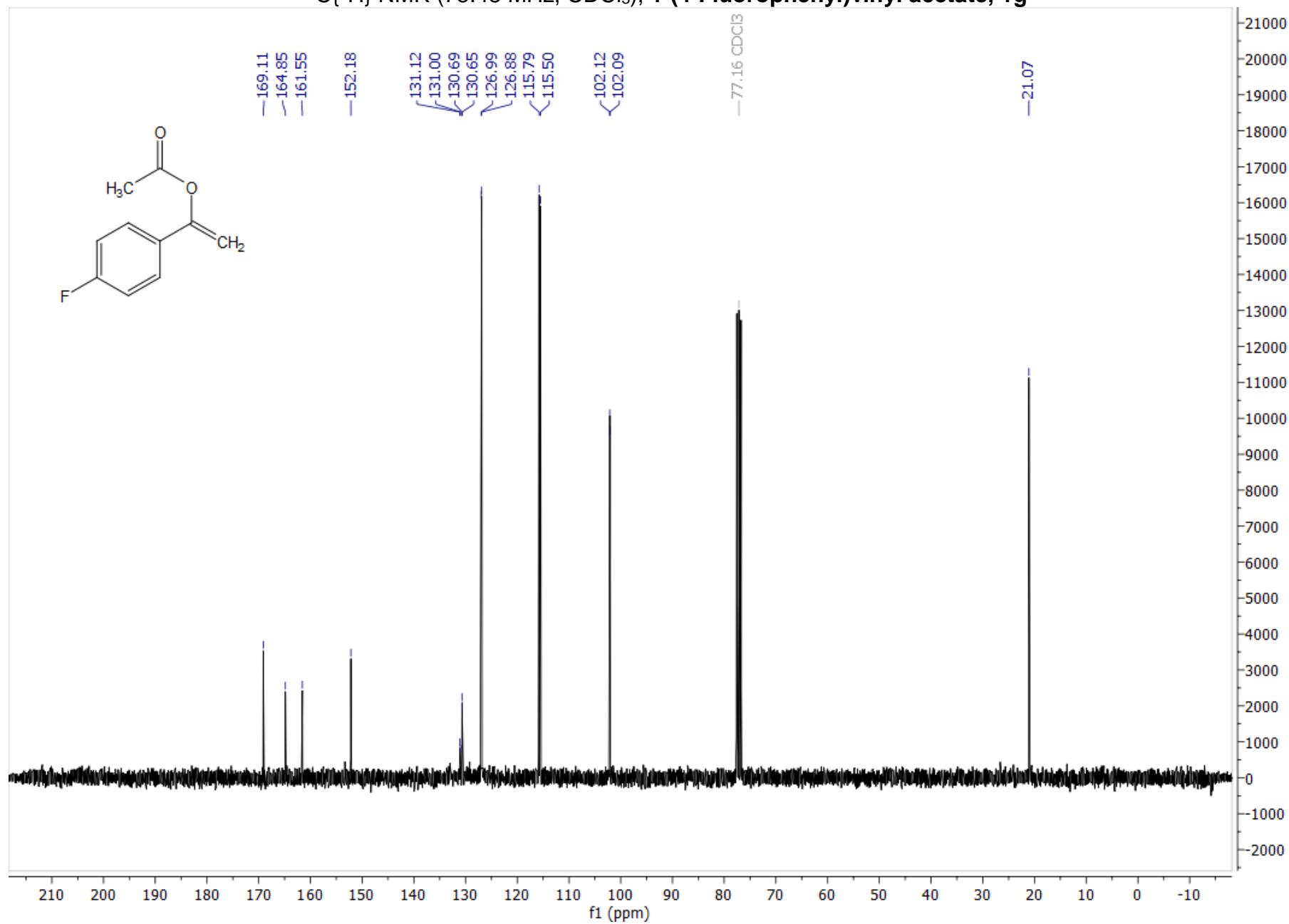


¹H NMR (300.13 MHz, CDCl₃), 1-(4-Fluorophenyl)vinyl acetate, 1g

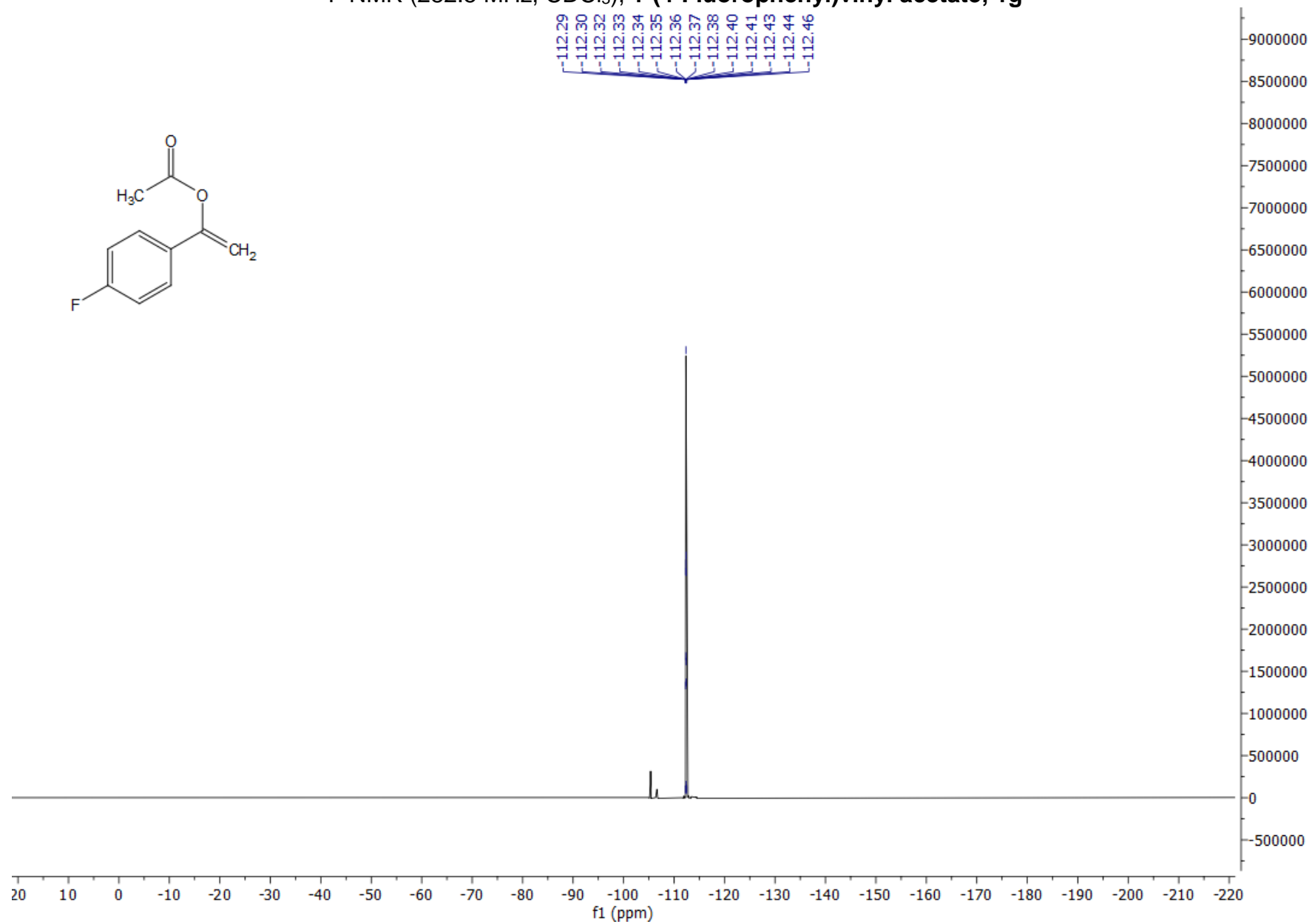


S63

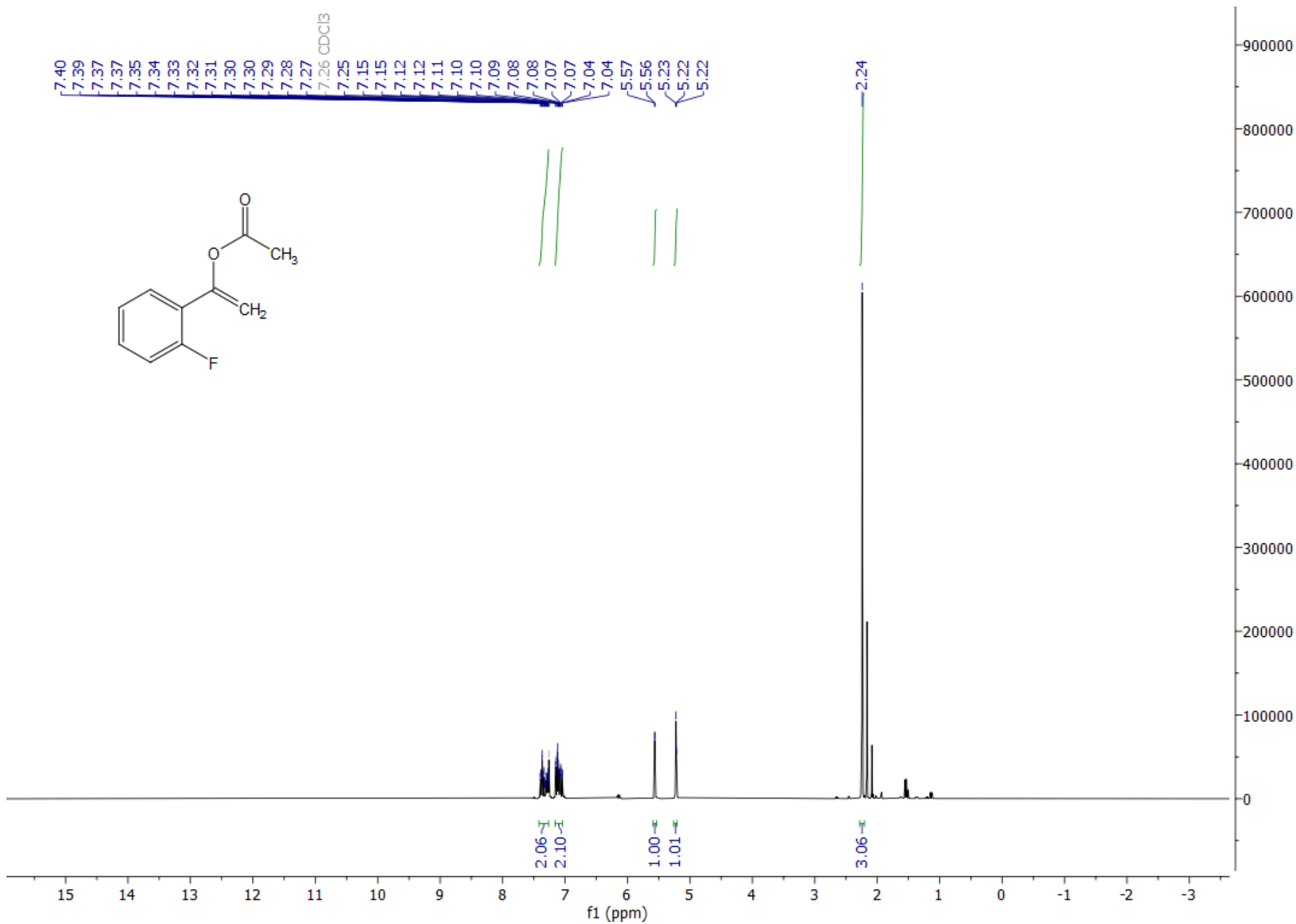
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(4-Fluorophenyl)vinyl acetate, 1g



¹⁹F NMR (282.5 MHz, CDCl₃), 1-(4-Fluorophenyl)vinyl acetate, 1g

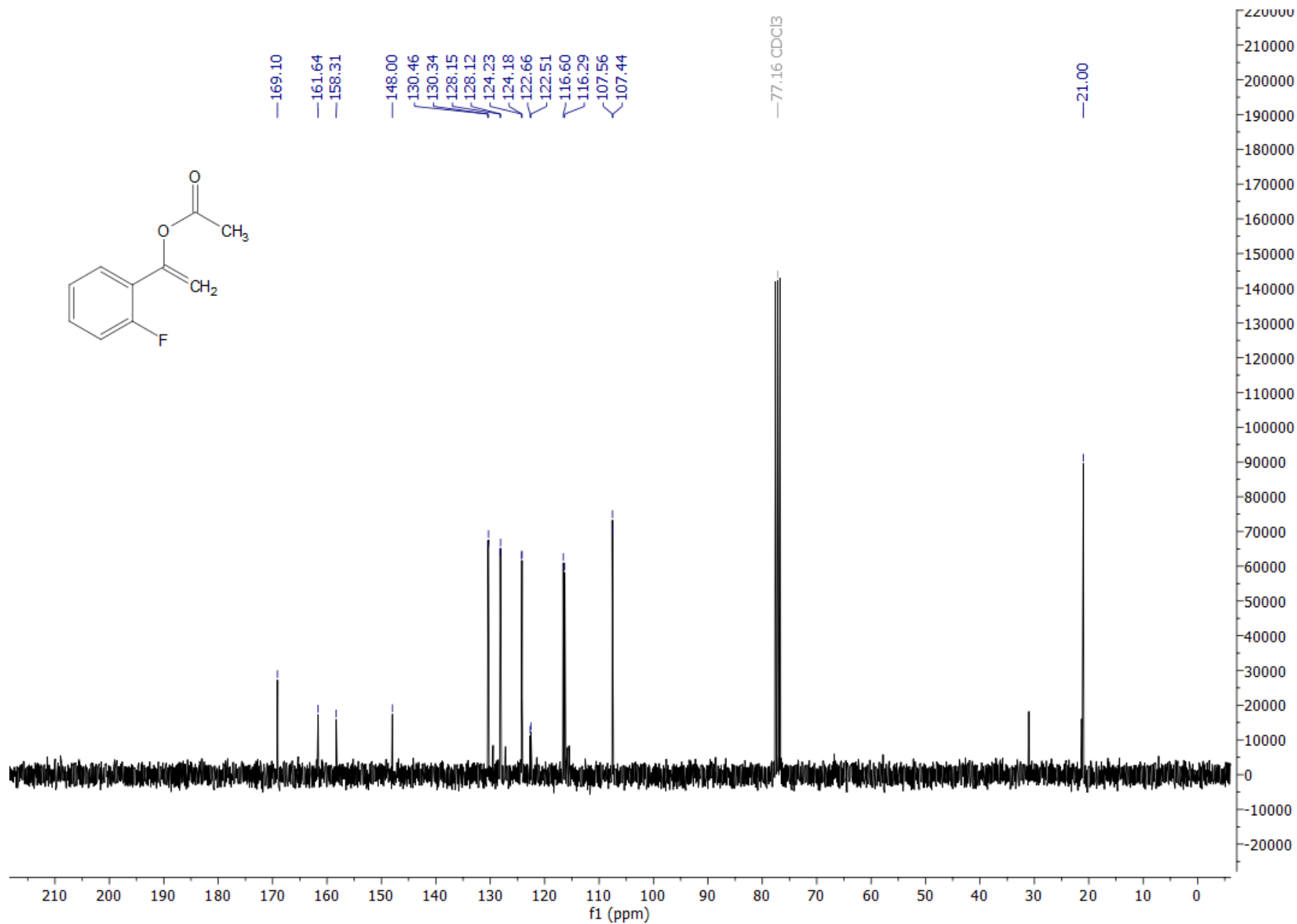


¹H NMR (300.13 MHz, CDCl₃), 1-(2-Fluorophenyl)vinyl acetate, 1h

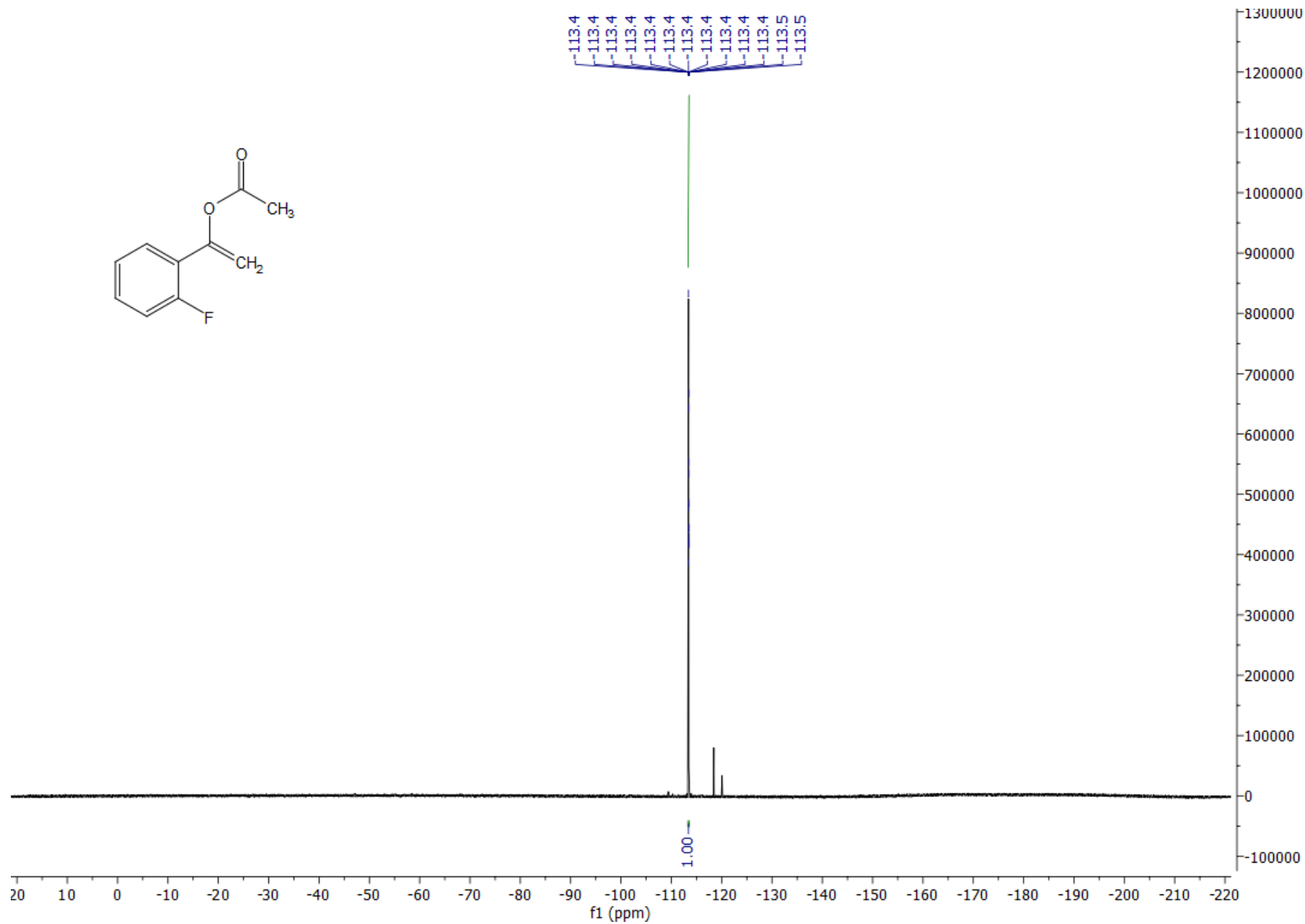


S66

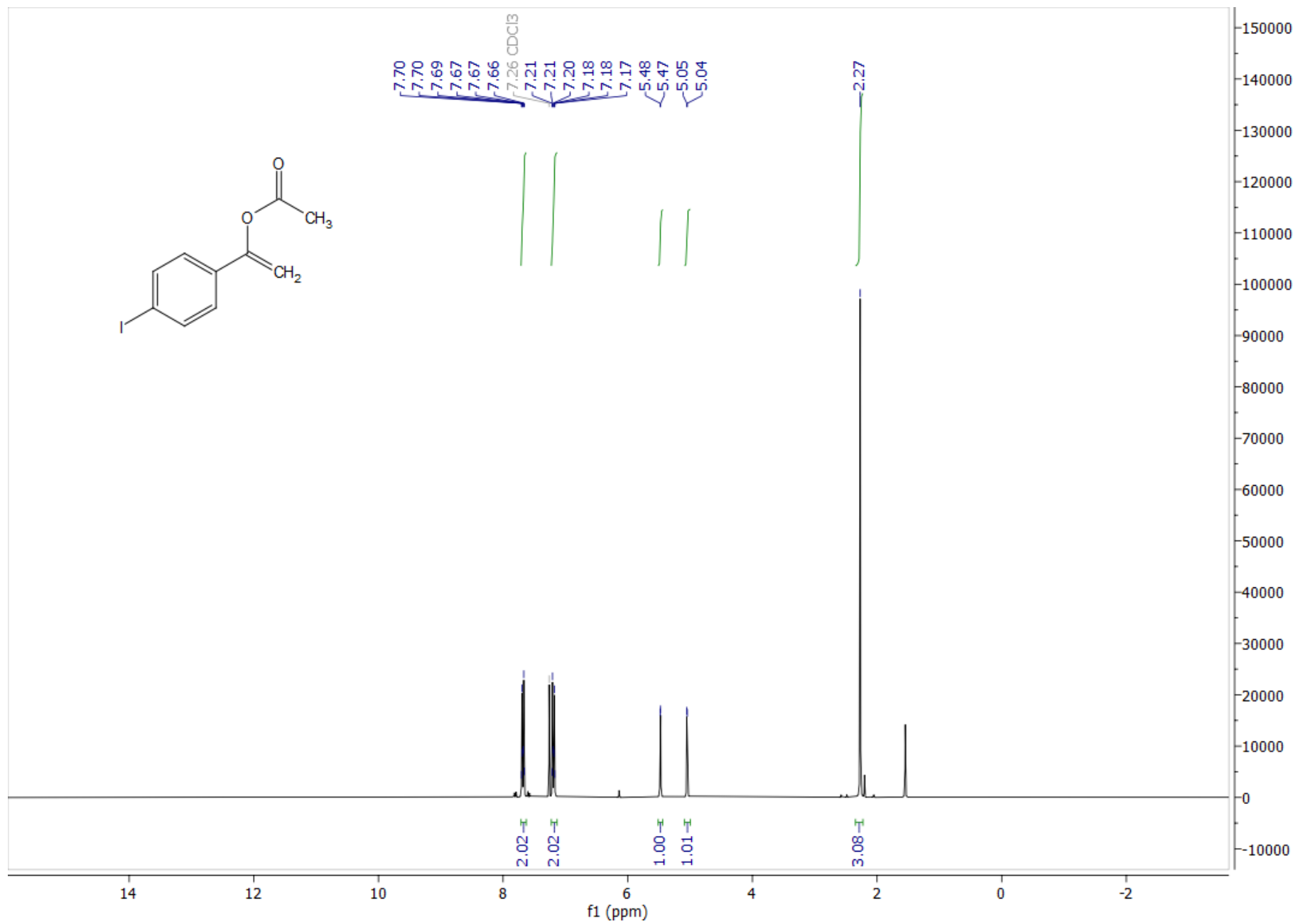
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(2-Fluorophenyl)vinyl acetate, 1h



^{19}F NMR (282,5 MHz, CDCl_3), 1-(2-Fluorophenyl)vinyl acetate, 1h

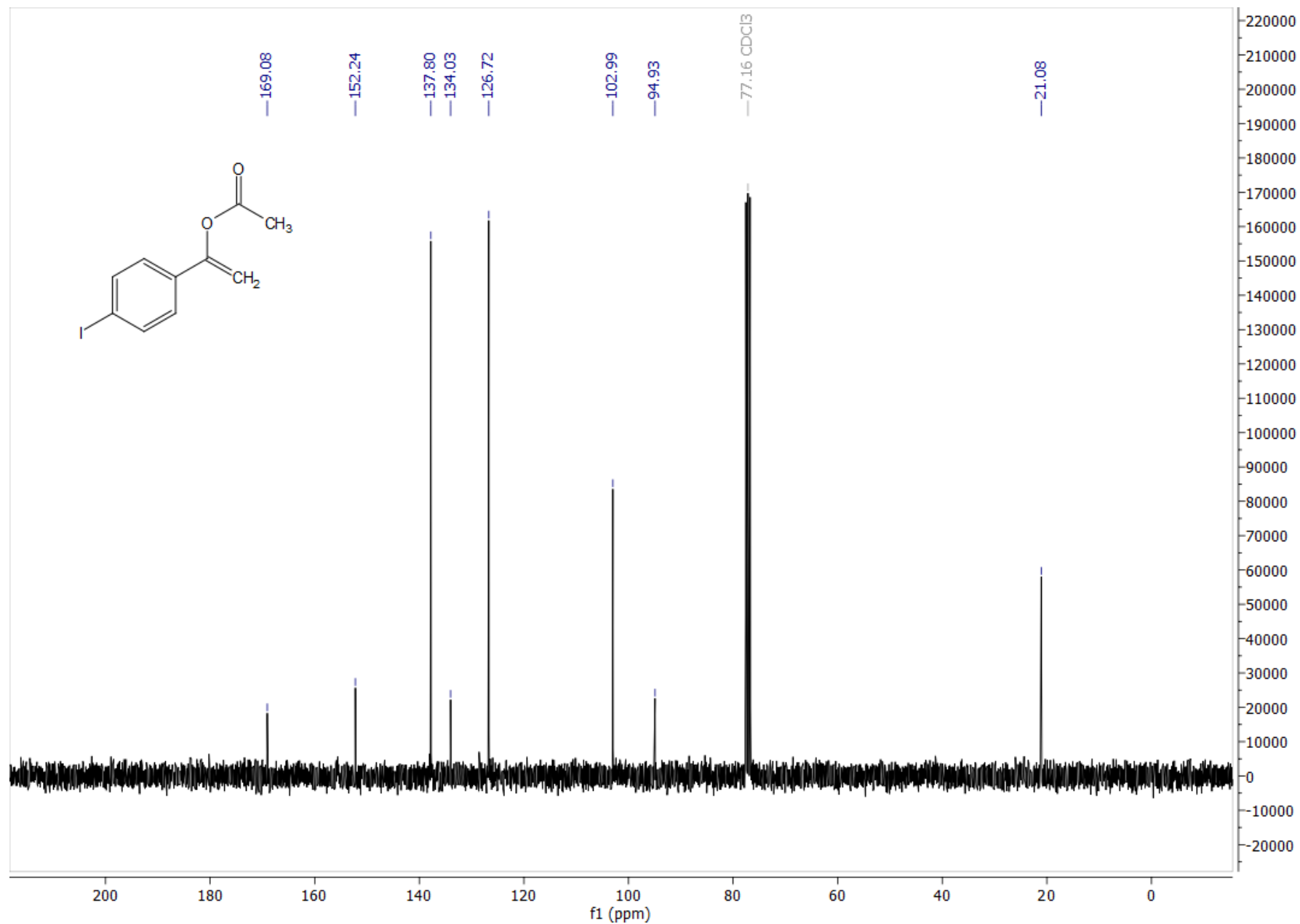


¹H NMR (300.13 MHz, CDCl₃), 1-(4-Iodophenyl)vinyl acetate, 1t

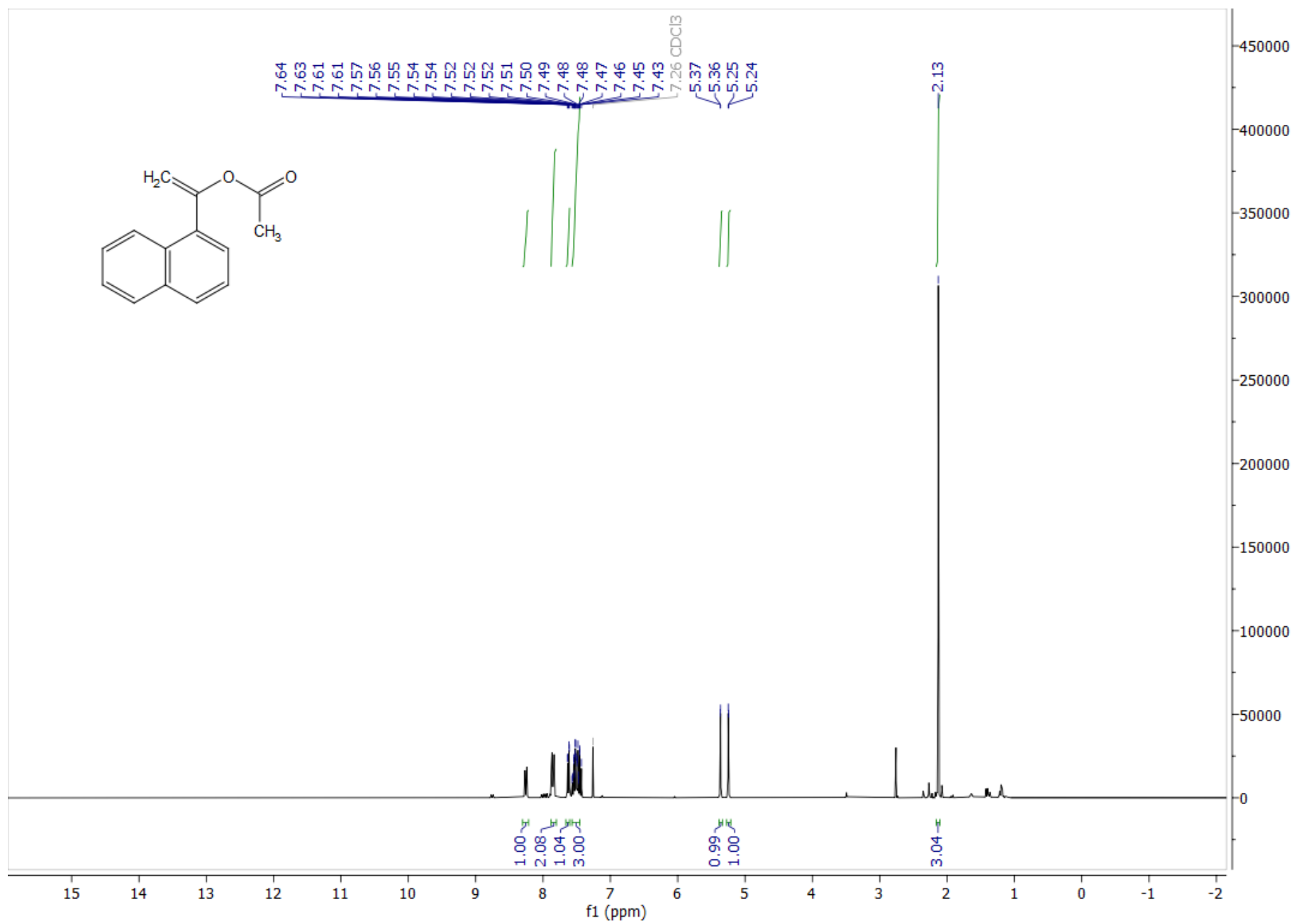


S69

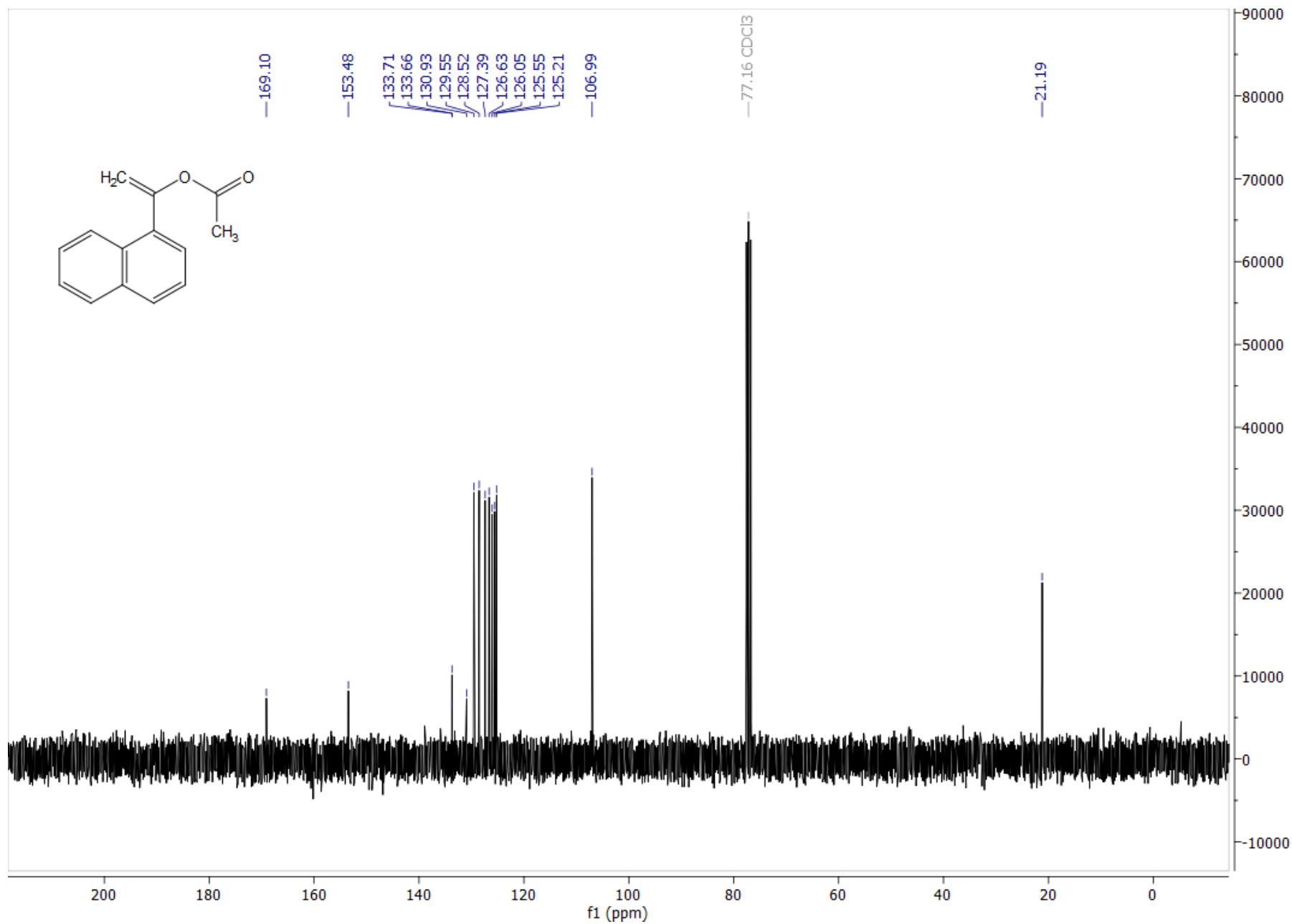
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(4-Iodophenyl)vinyl acetate, 1t



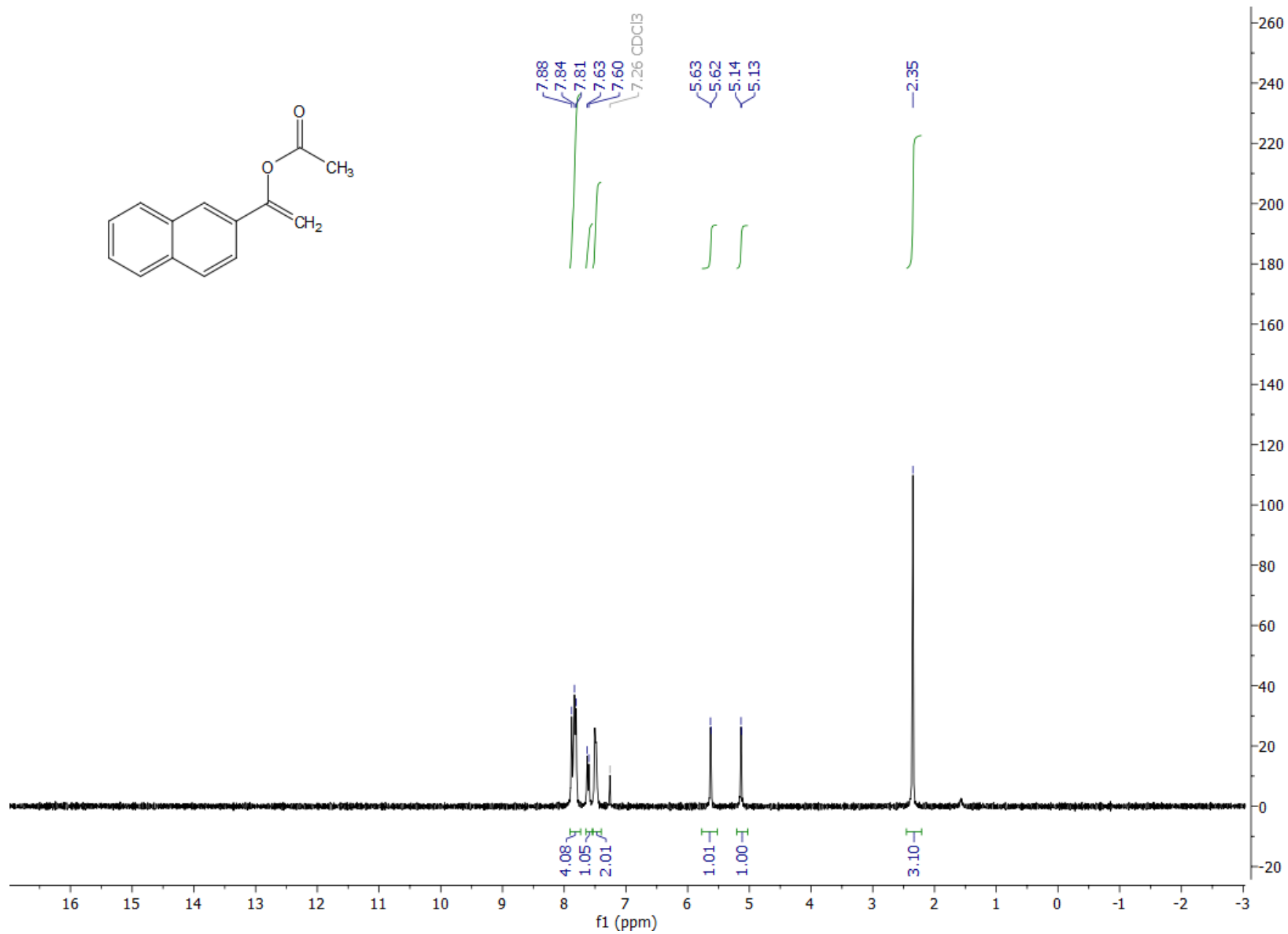
¹H NMR (300.13 MHz, CDCl₃), 1-(Naphthalen-1-yl)vinyl acetate, 1i



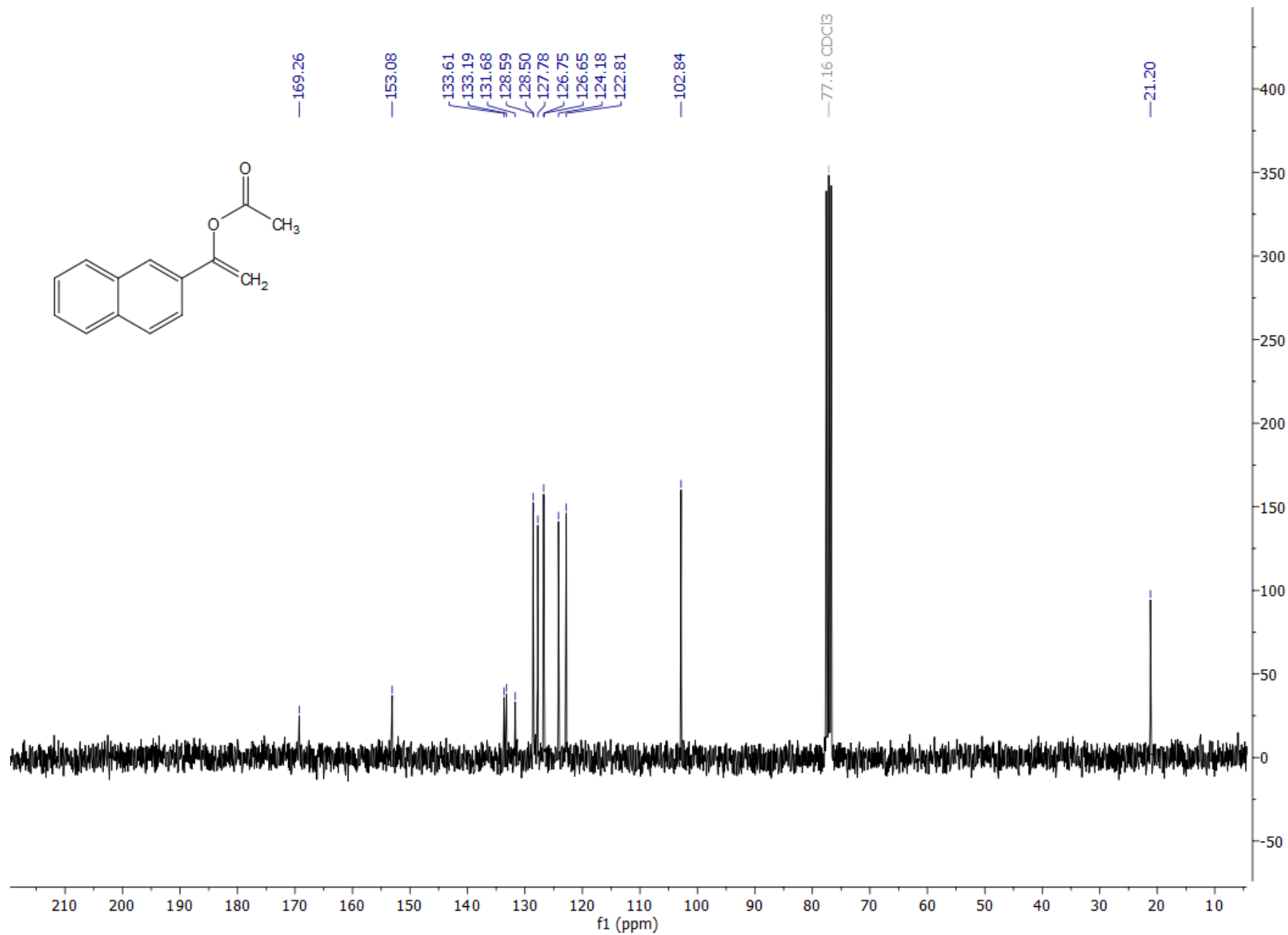
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(Naphthalen-1-yl)vinyl acetate, **1i**



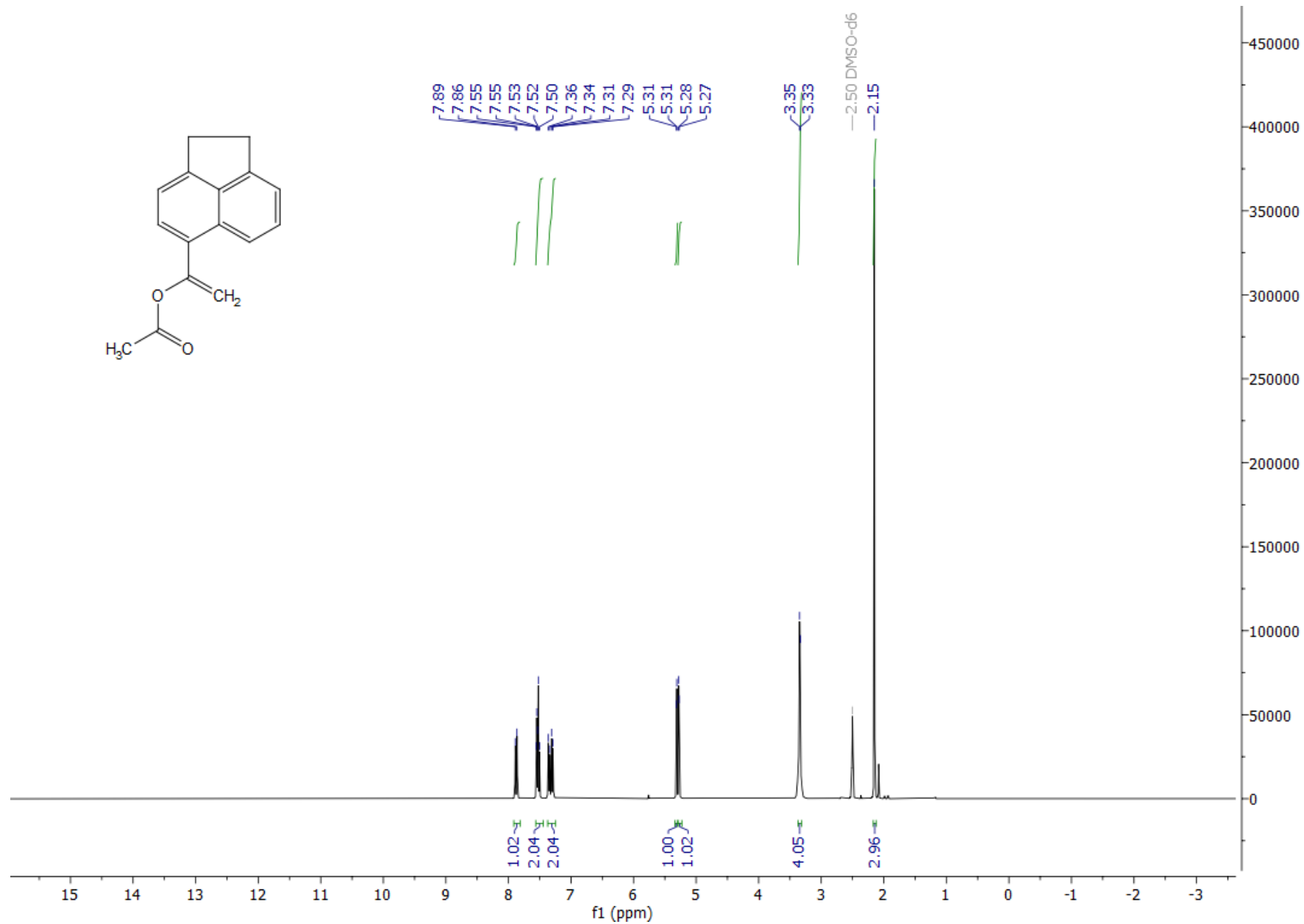
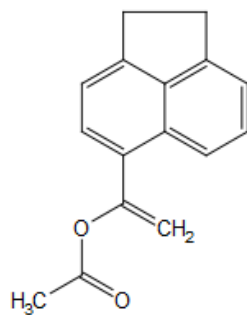
¹H NMR (300.13 MHz, CDCl₃), 1-(Naphthalen-2-yl)vinyl acetate, 1j



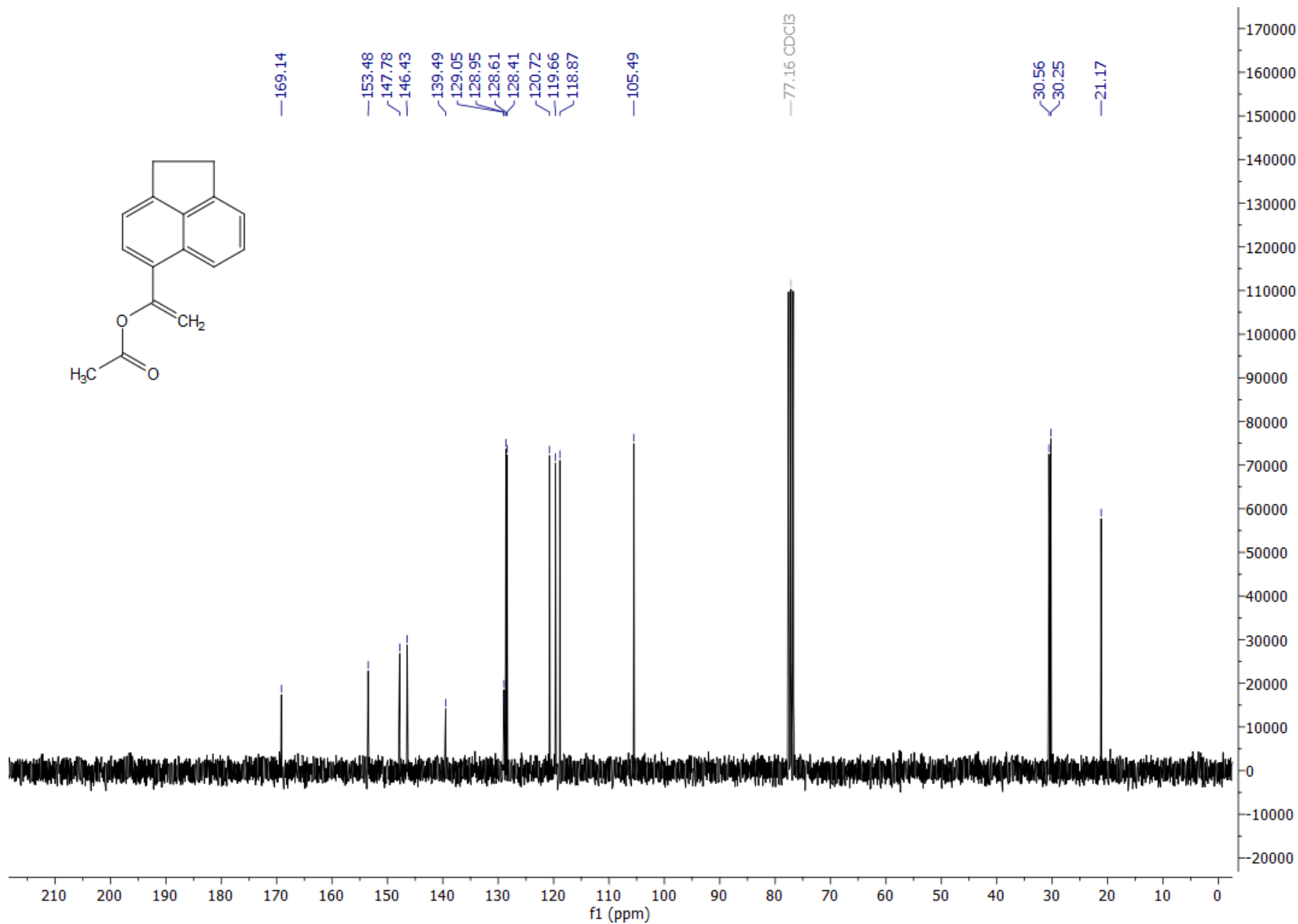
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(Naphthalen-2-yl)vinyl acetate, 1j



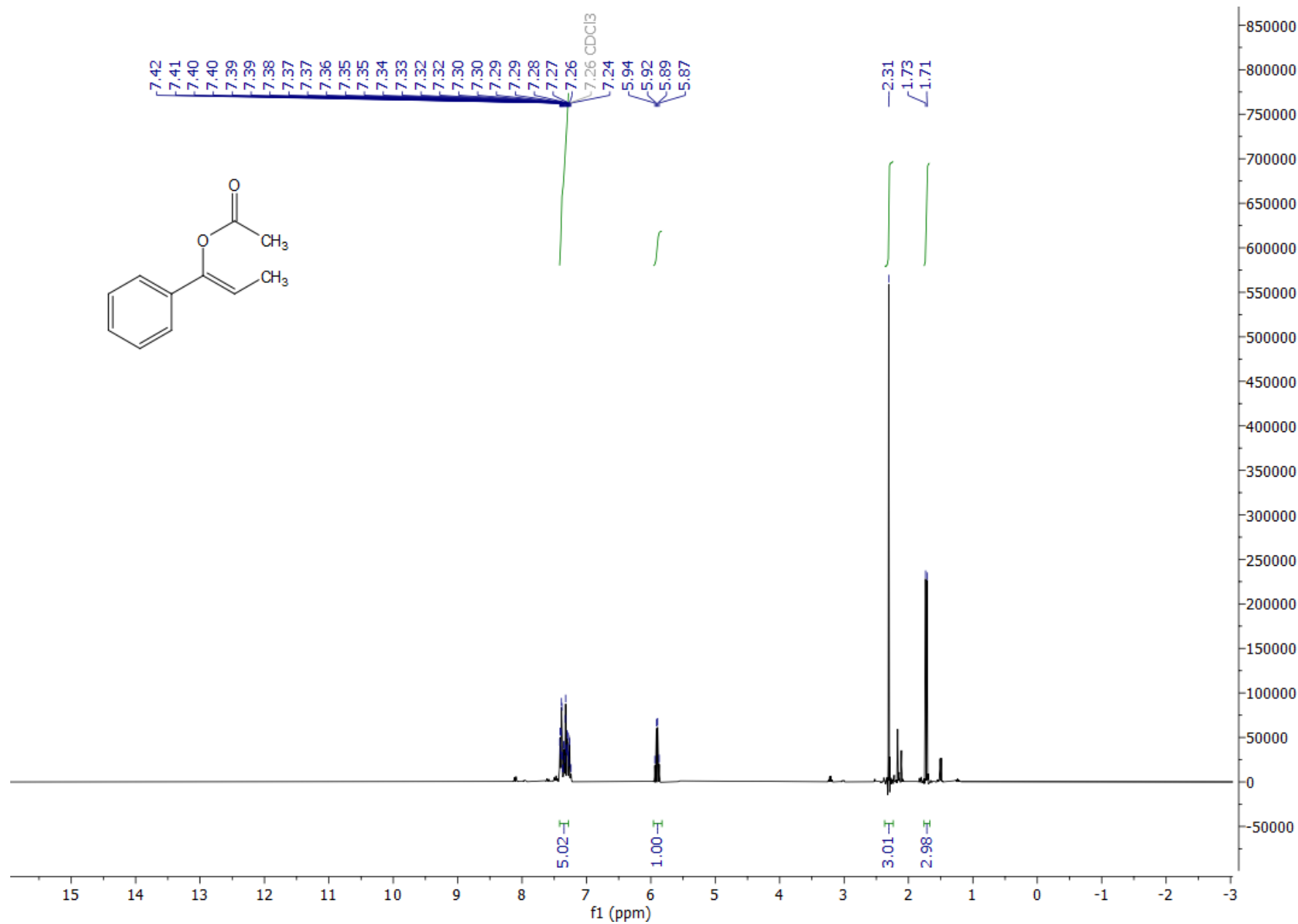
¹H NMR (300.13 MHz, DMSO-d₆), 1-(1,2-Dihydroacenaphthylen-5-yl)vinyl acetate, 1k



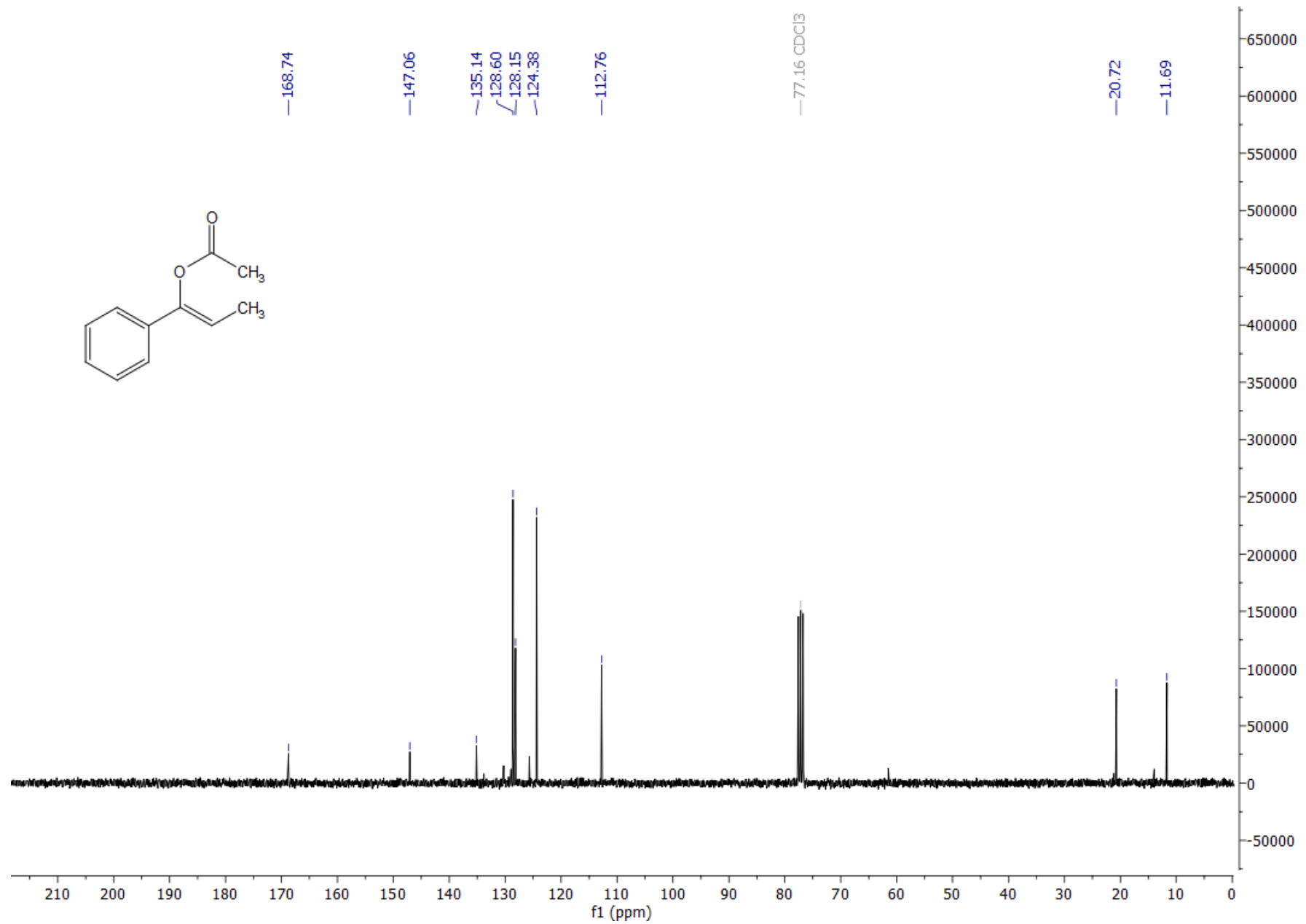
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-(1,2-Dihydroacenaphthylen-5-yl)vinyl acetate, 1k



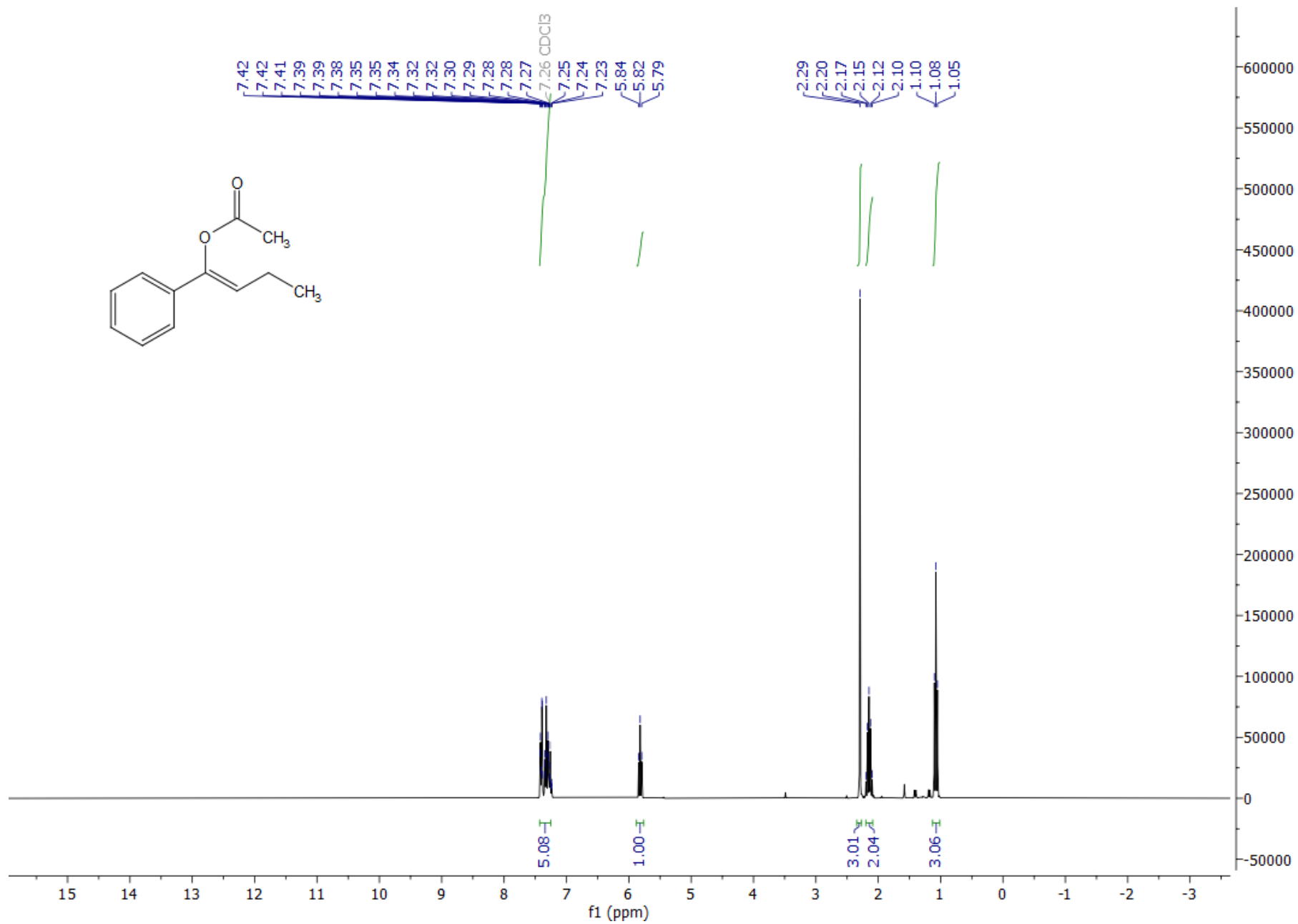
¹H NMR (300.13 MHz, CDCl₃), (Z)-1-Phenylprop-1-en-1-yl acetate, 11



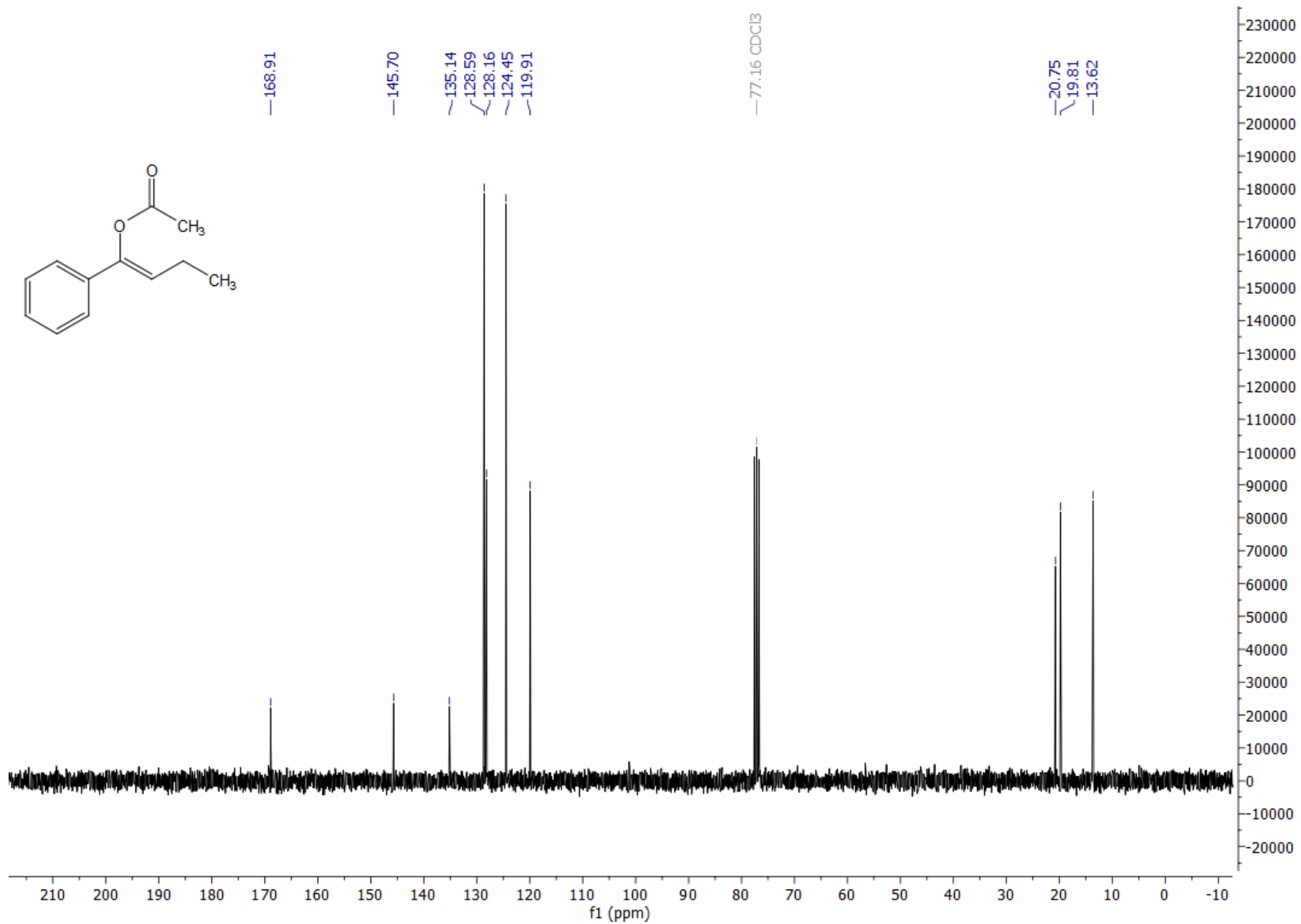
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), (**Z**)-1-Phenylprop-1-en-1-yl acetate, **1l**



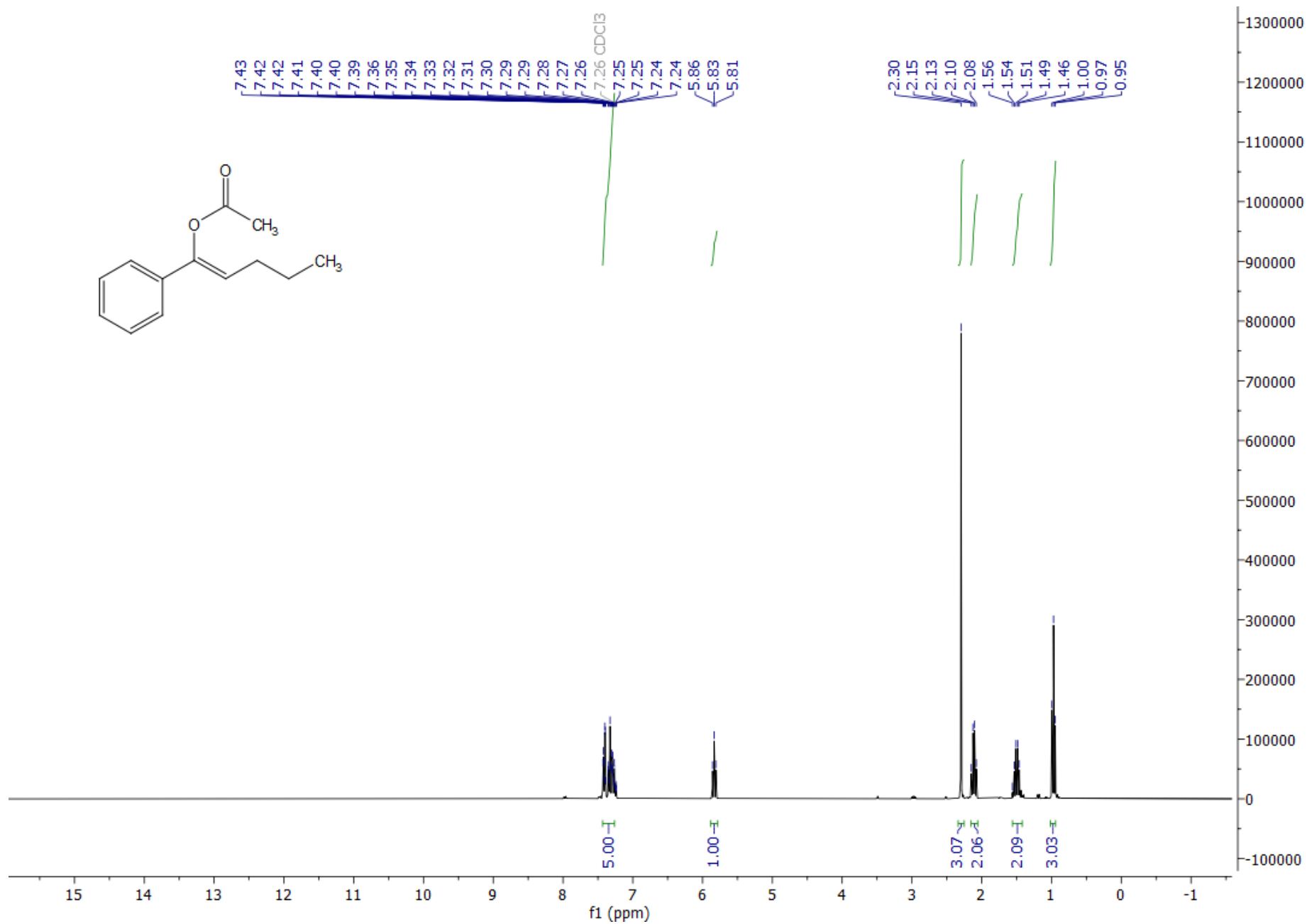
¹H NMR (300.13 MHz, CDCl₃), (**Z**)-1-Phenylbut-1-en-1-yl acetate, 1m



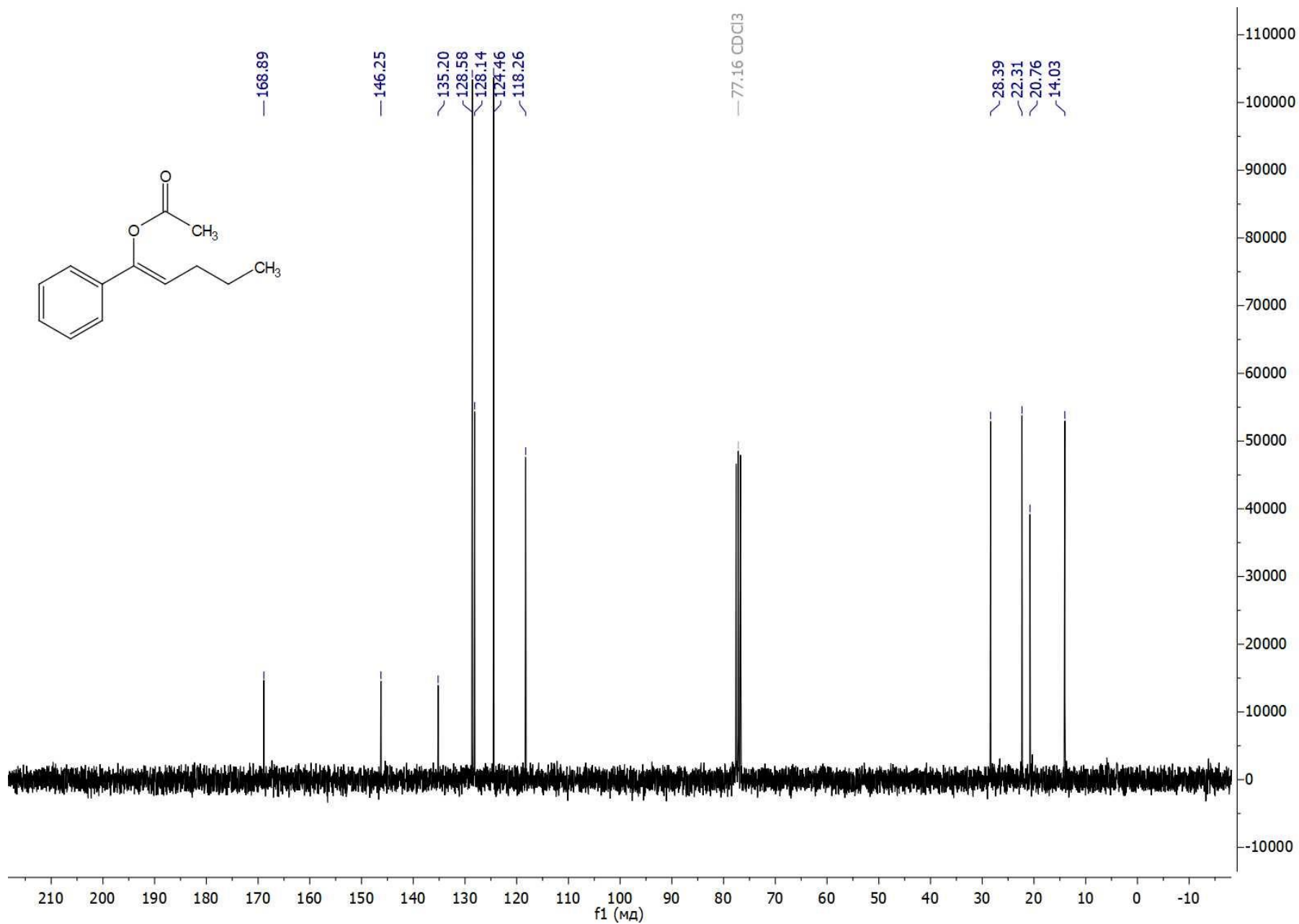
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), (**Z**)-1-Phenylbut-1-en-1-yl acetate, 1m



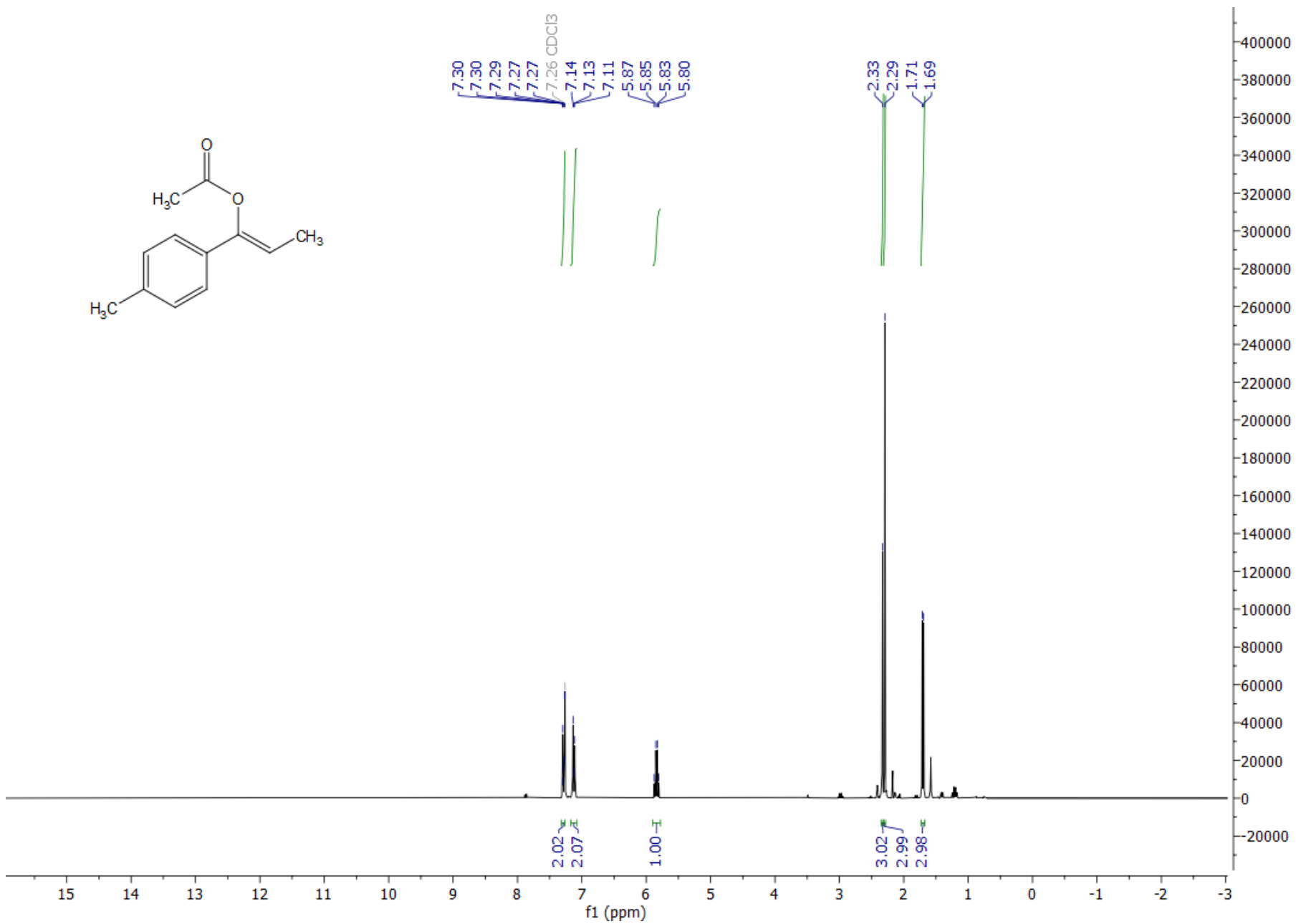
¹H NMR (300.13 MHz, CDCl₃), (Z)-1-Phenylpent-1-en-1-yl acetate, 1n



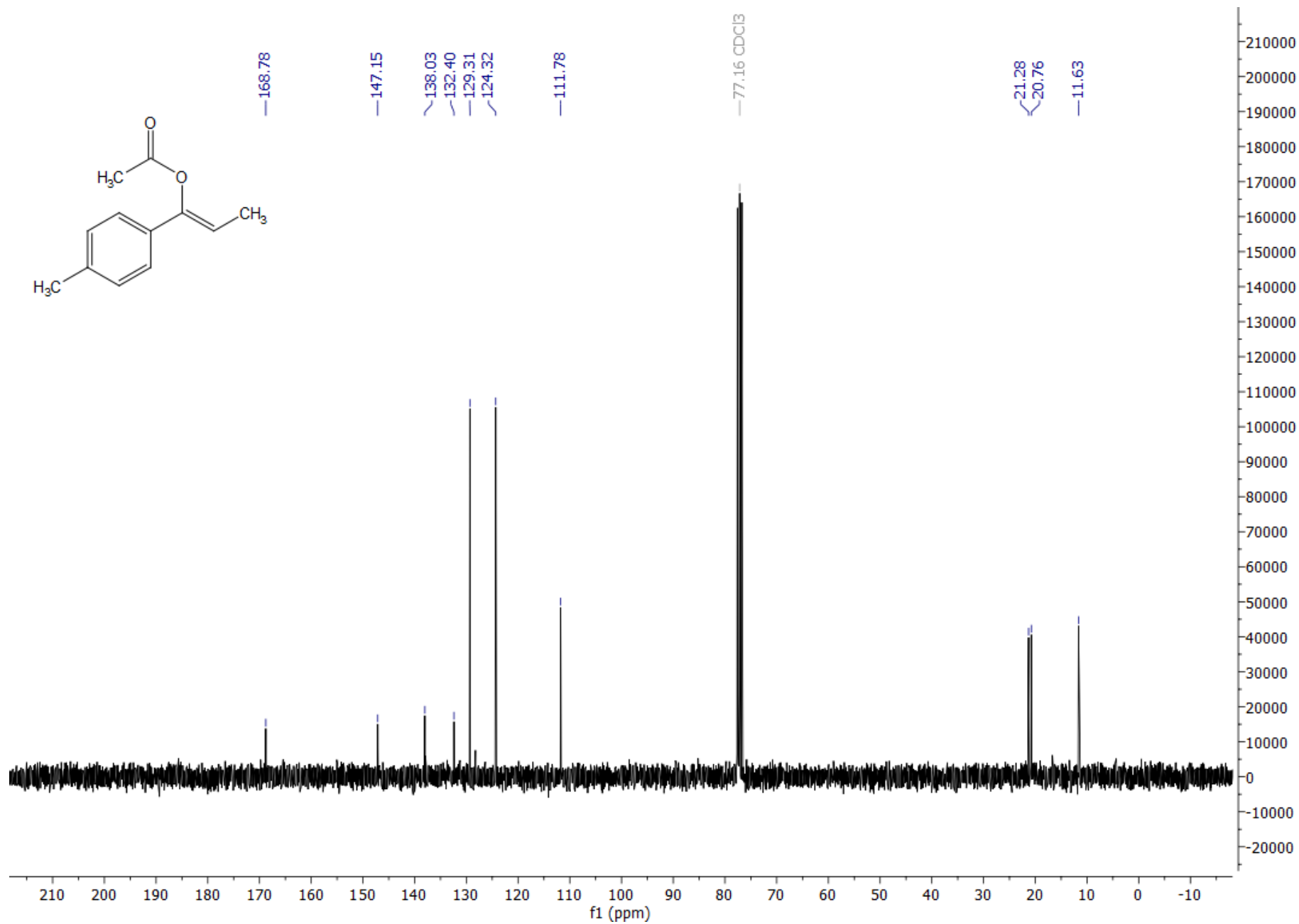
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), (**Z**)-1-Phenylpent-1-en-1-yl acetate, **1n**



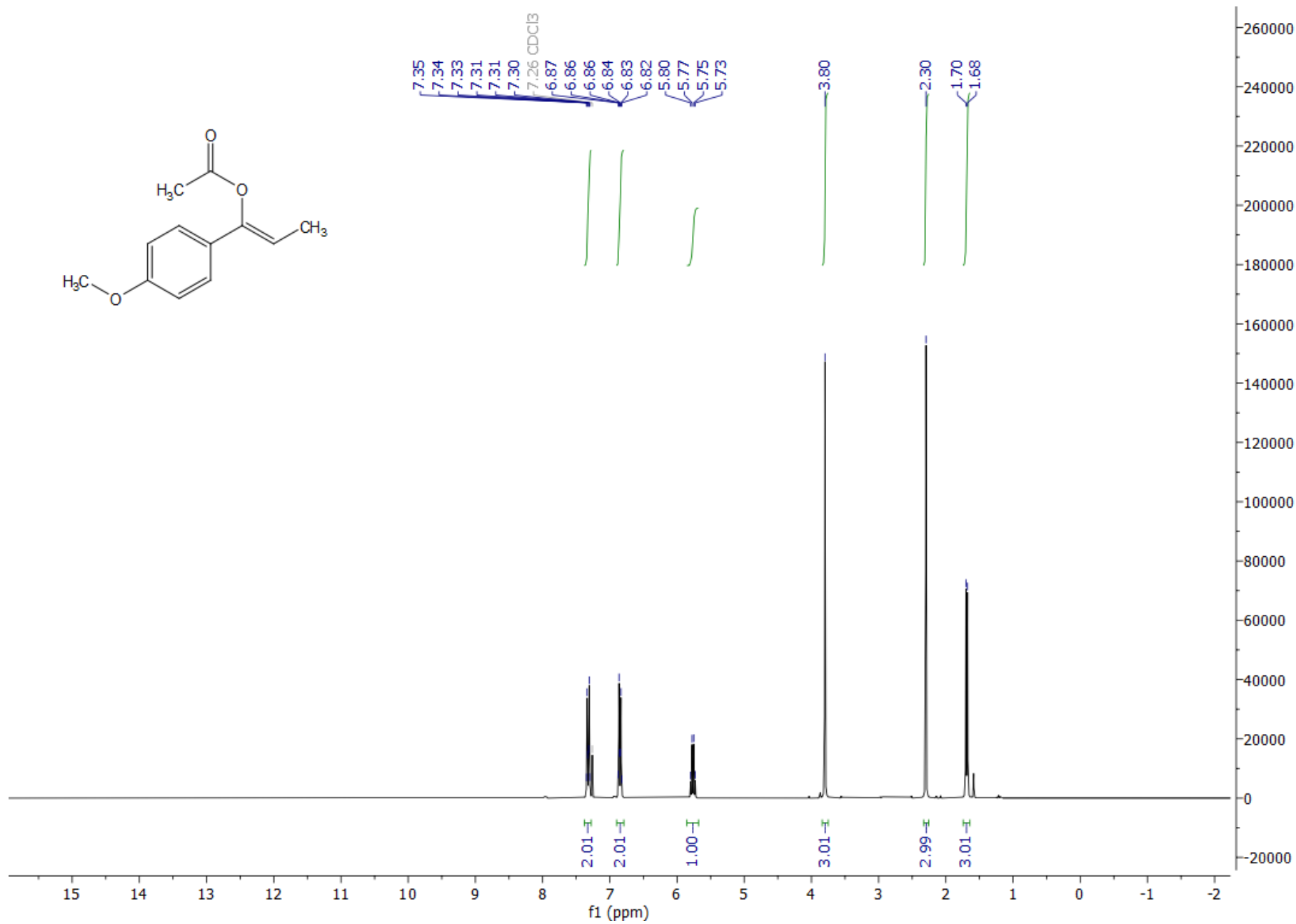
¹H NMR (300.13 MHz, CDCl₃), (Z)-1-(p-Tolyl)prop-1-en-1-yl acetate, 1o



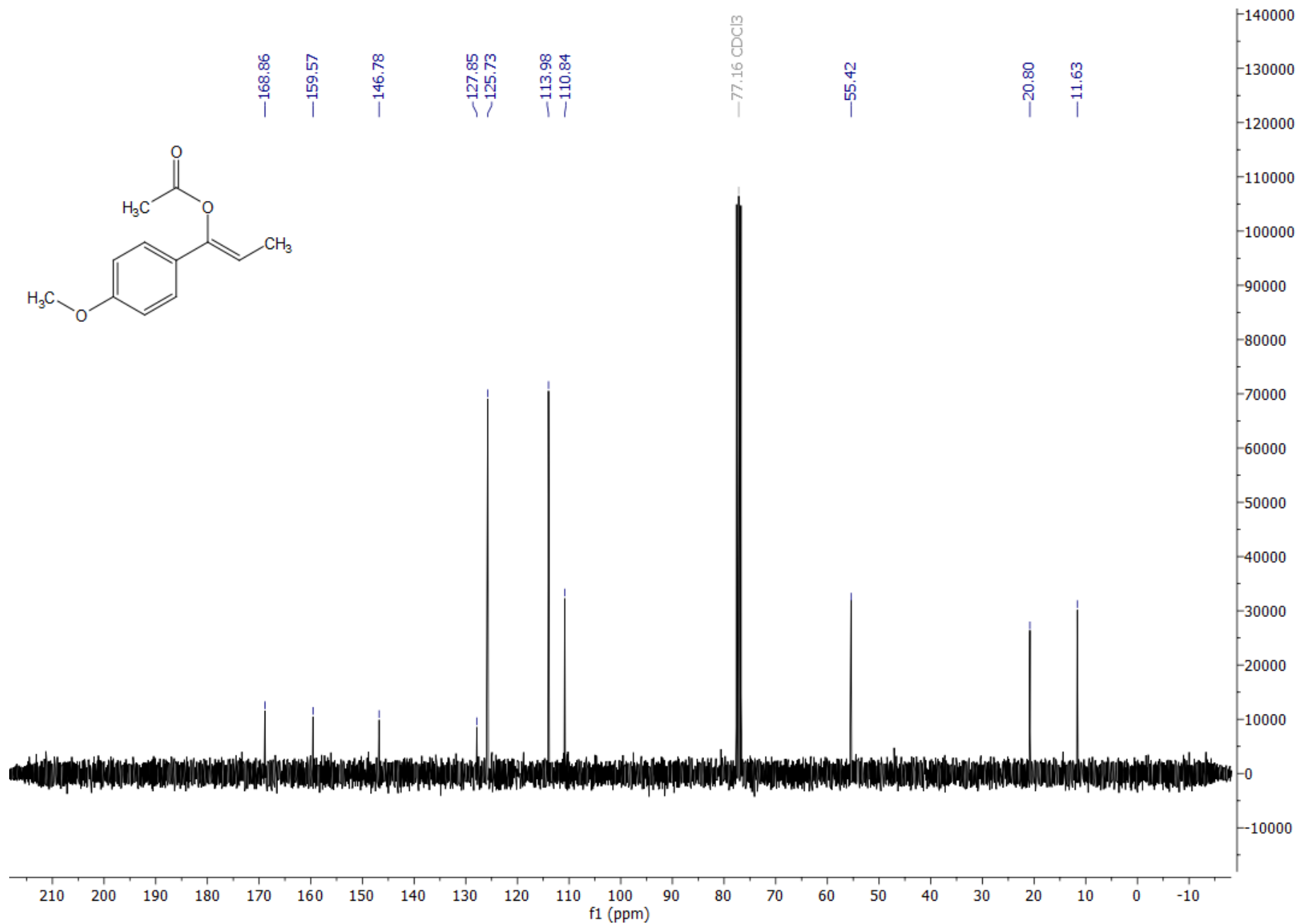
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), (**Z**)-1-(*p*-Tolyl)prop-1-en-1-yl acetate, **1o**



¹H NMR (300.13 MHz, CDCl₃), (Z)-1-(4-Methoxyphenyl)prop-1-en-1-yl acetate, 1p

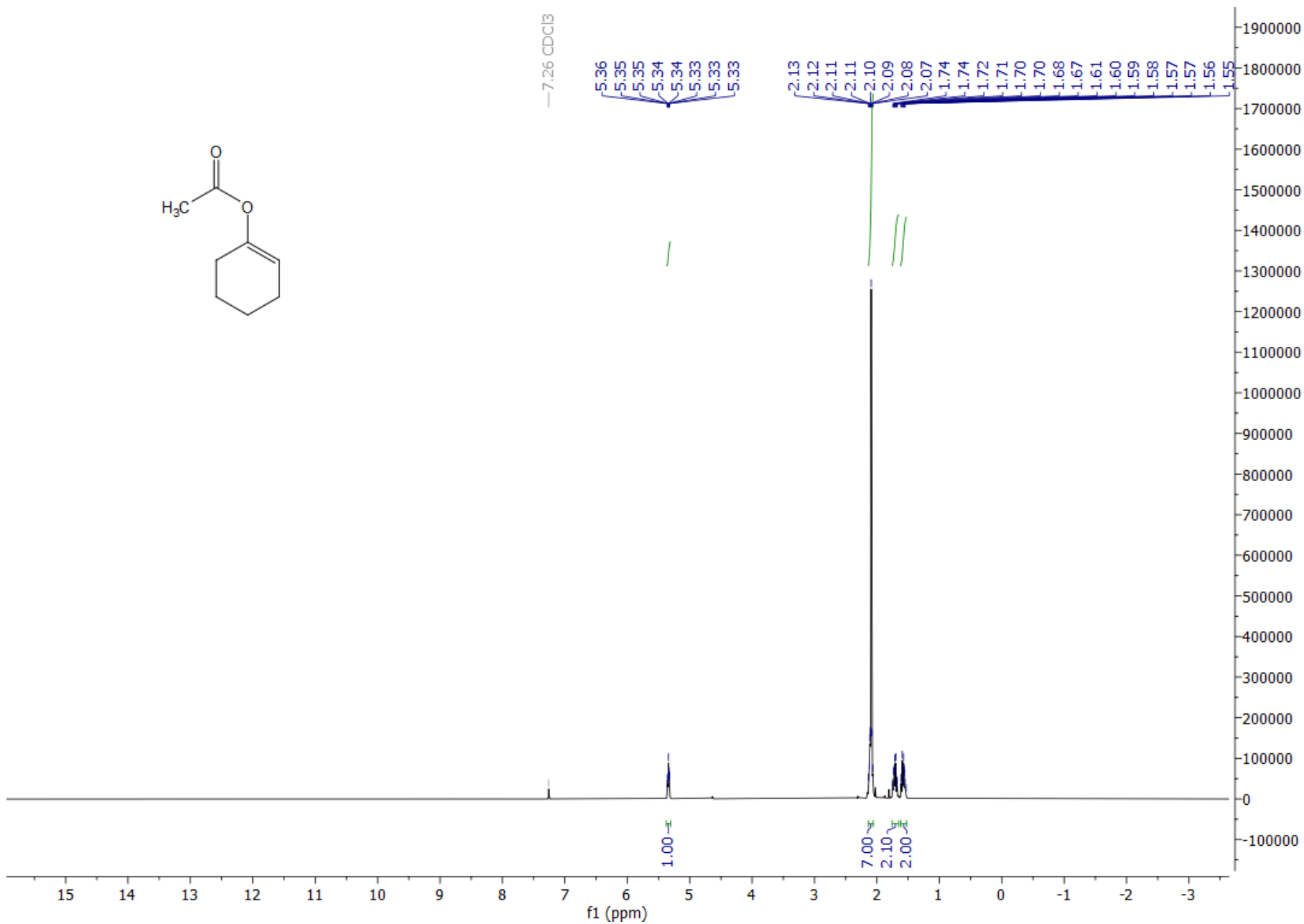
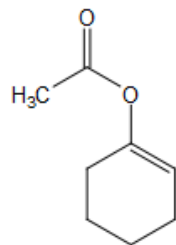


$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), (**Z**)-1-(4-Methoxyphenyl)prop-1-en-1-yl acetate, **1p**

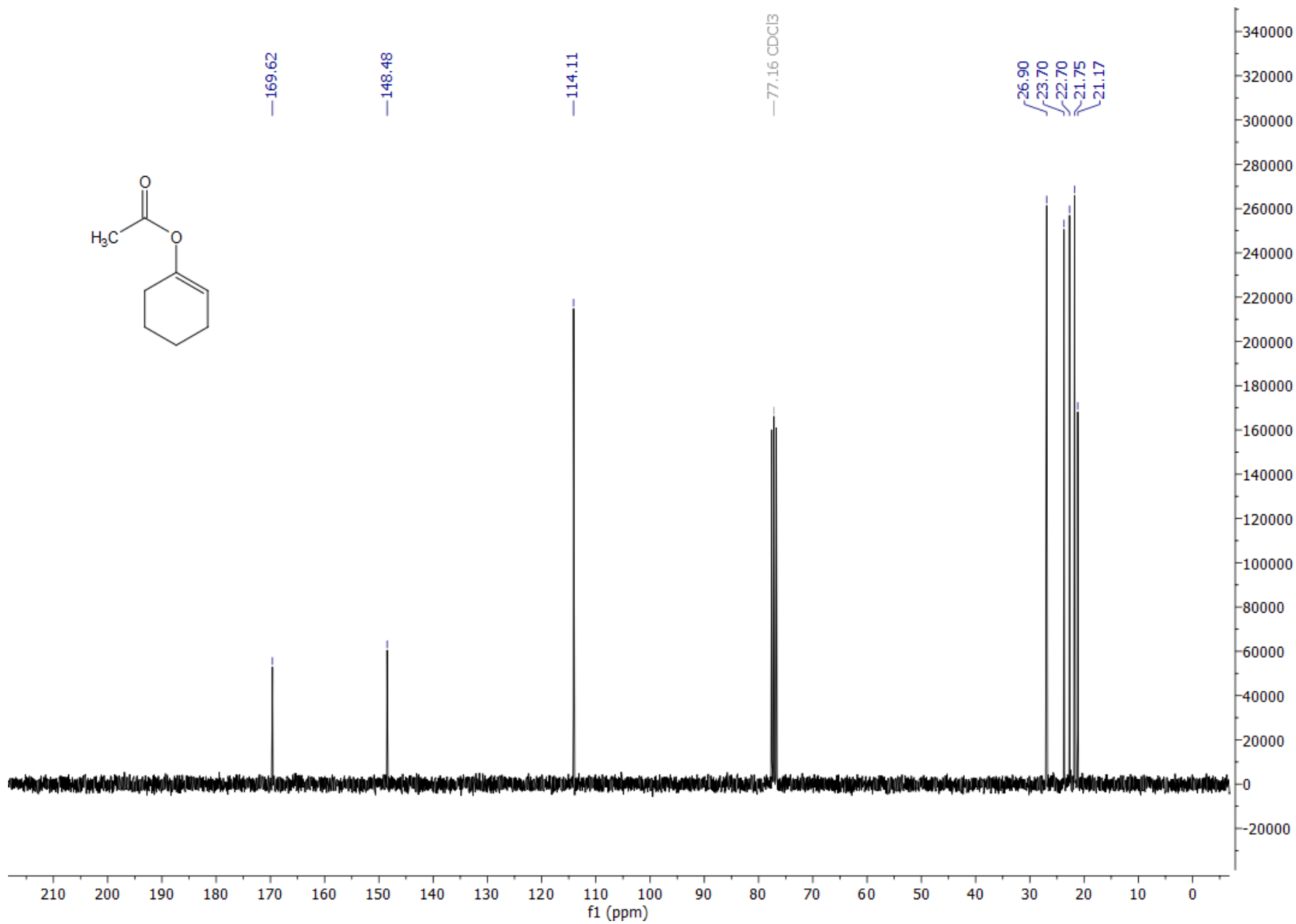


S86

¹H NMR (300.13 MHz, CDCl₃), Cyclohex-1-en-1-yl acetate

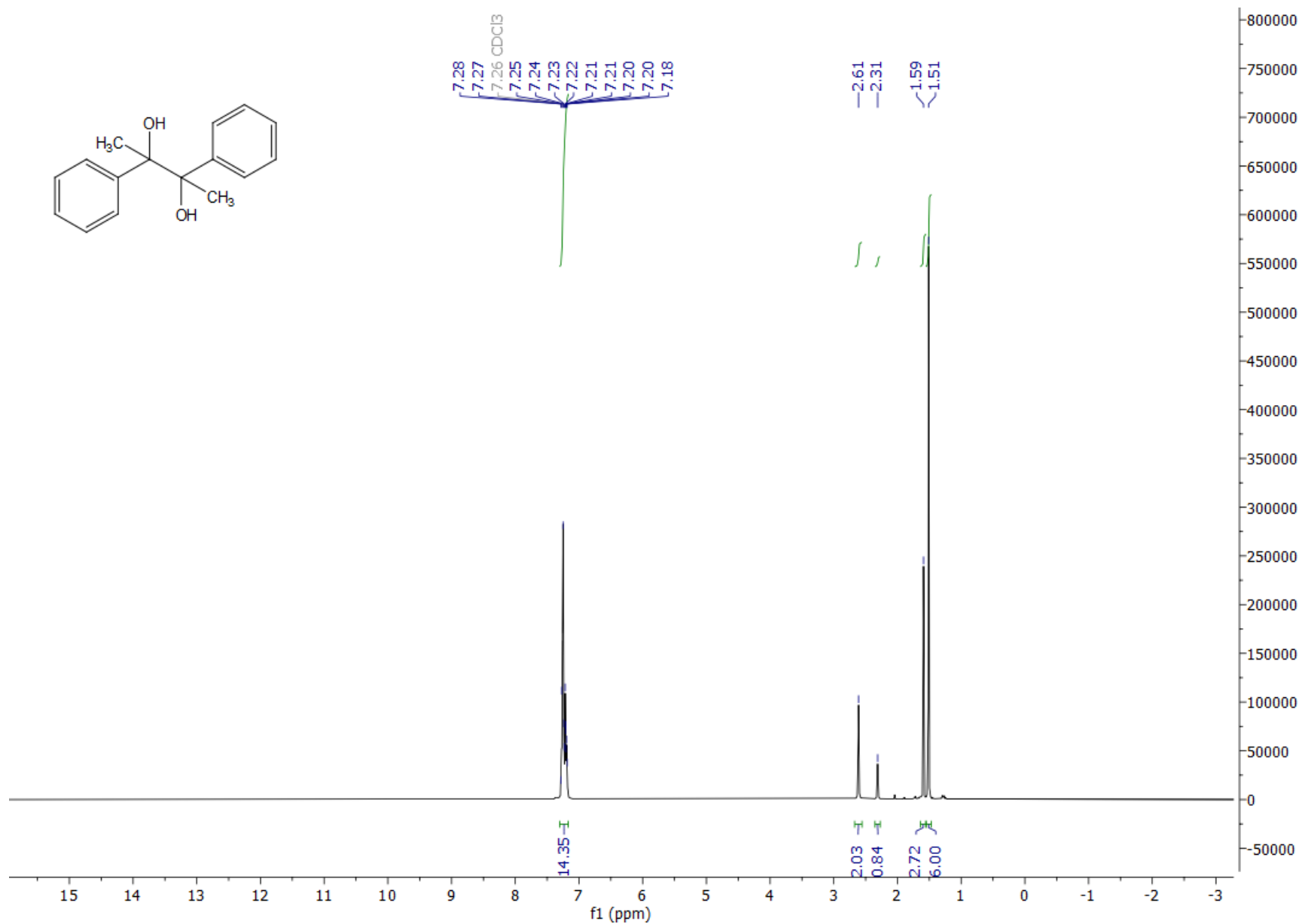


$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), Cyclohex-1-en-1-yl acetate

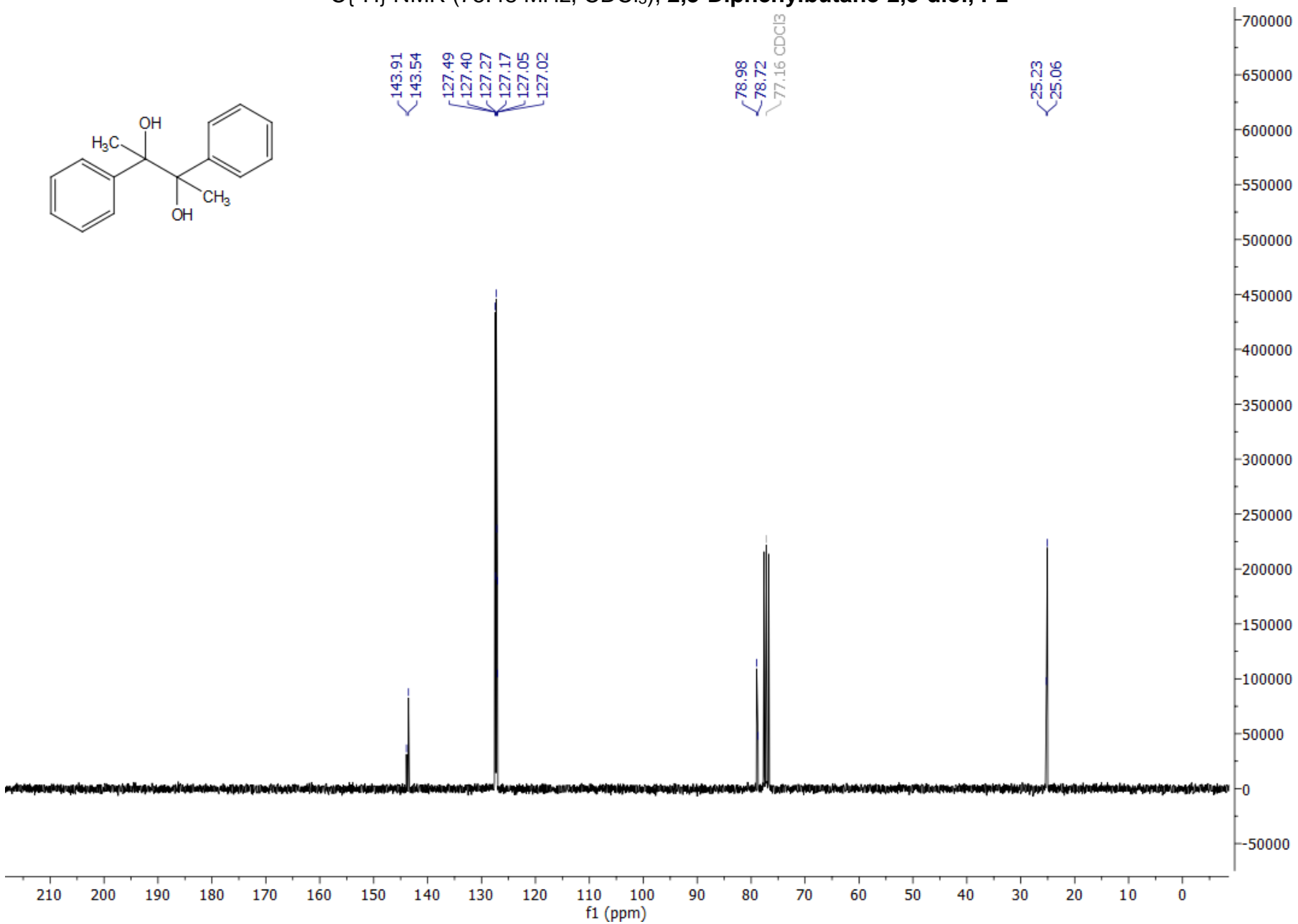


NMR spectra of the obtained products

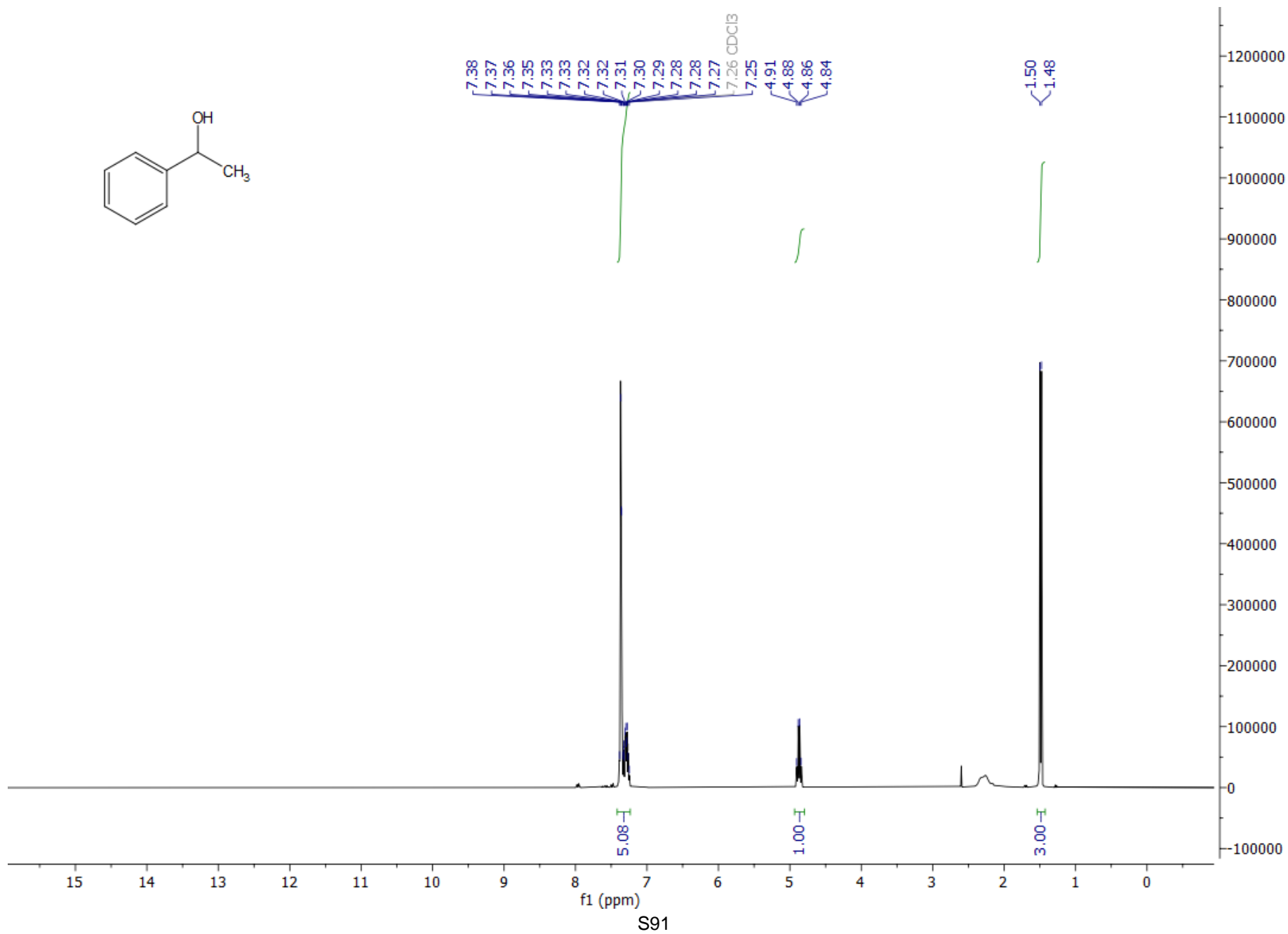
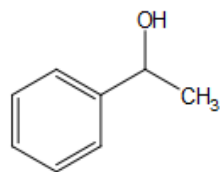
^1H NMR (300.13 MHz, CDCl_3), 2,3-Diphenylbutane-2,3-diol, P2



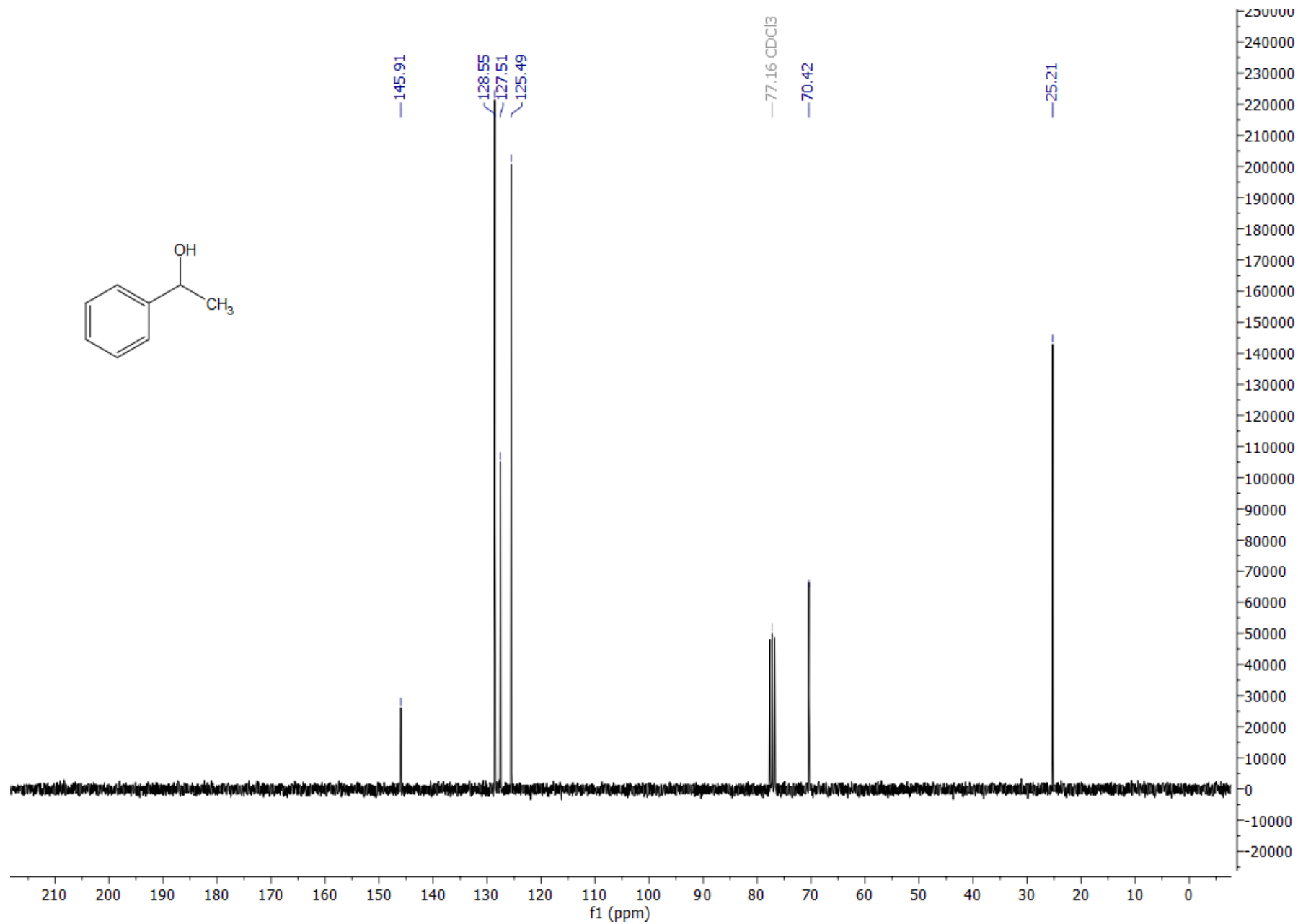
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 2,3-Diphenylbutane-2,3-diol, P2



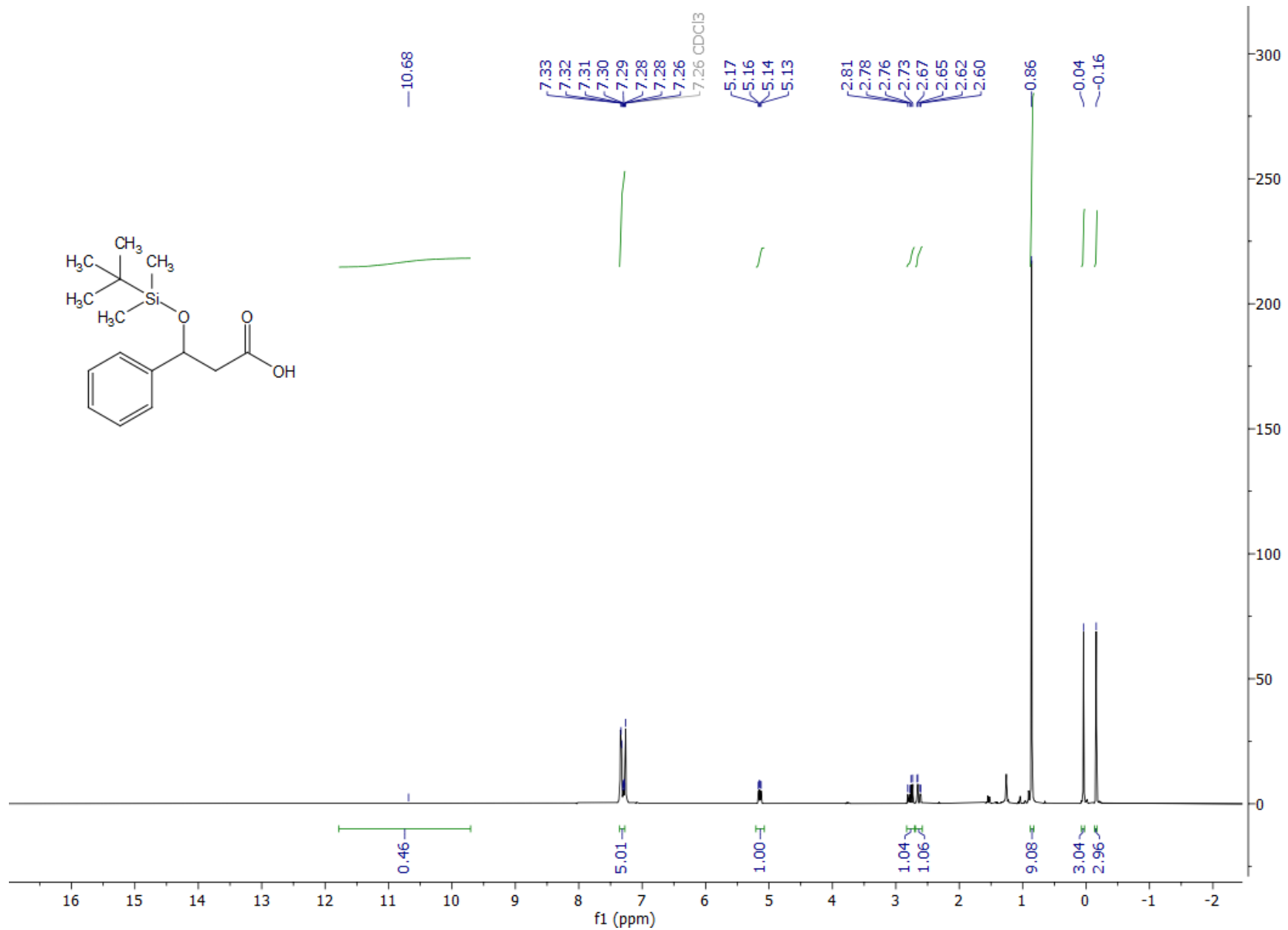
¹H NMR (300.13 MHz, CDCl₃), 1-Phenylethanol, P3



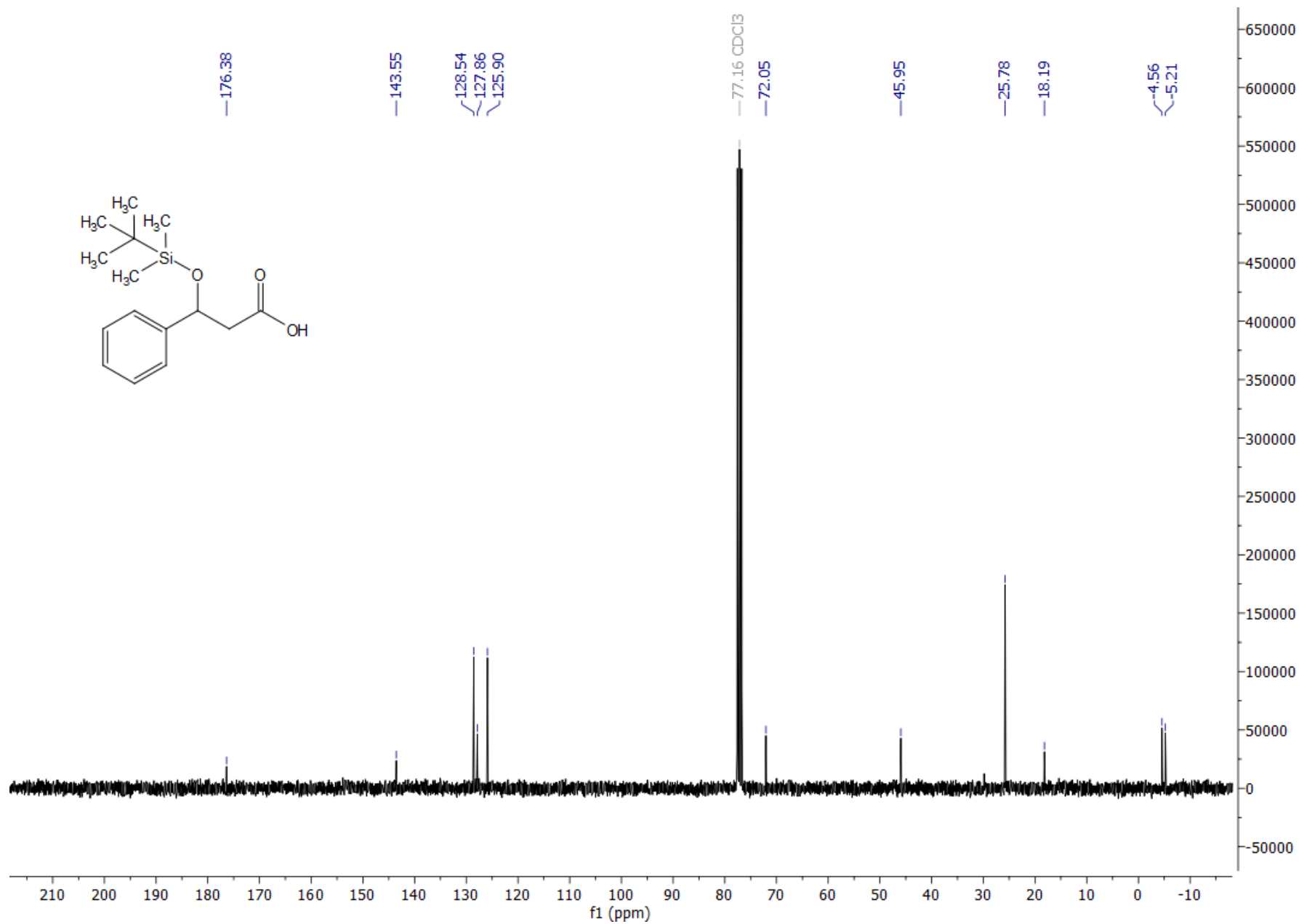
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-Phenylethanol, P3



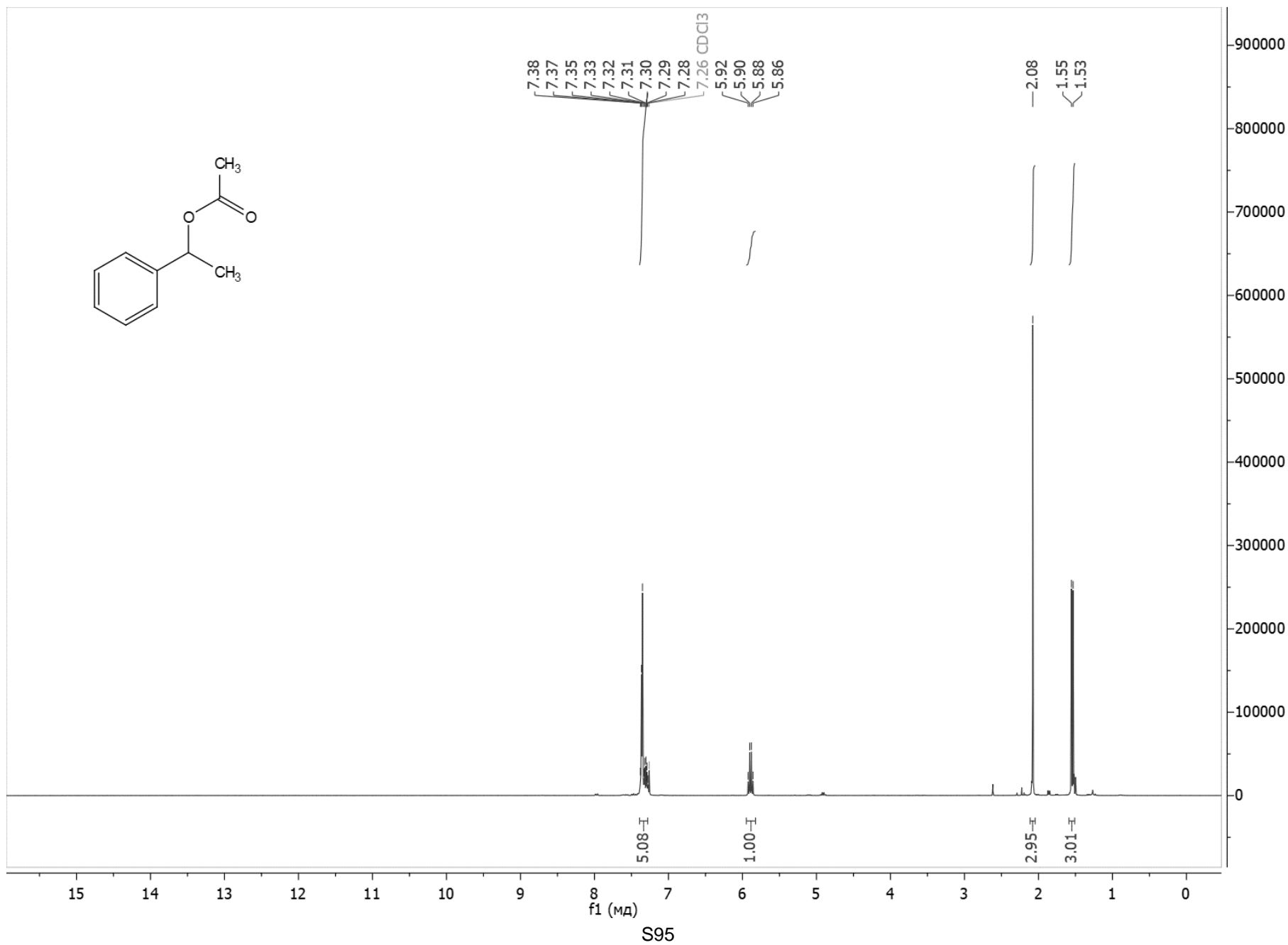
^1H NMR (300.13 MHz, CDCl_3), 3-((*Tert*-butyldimethylsilyloxy)-3-phenylpropanoic acid, P4



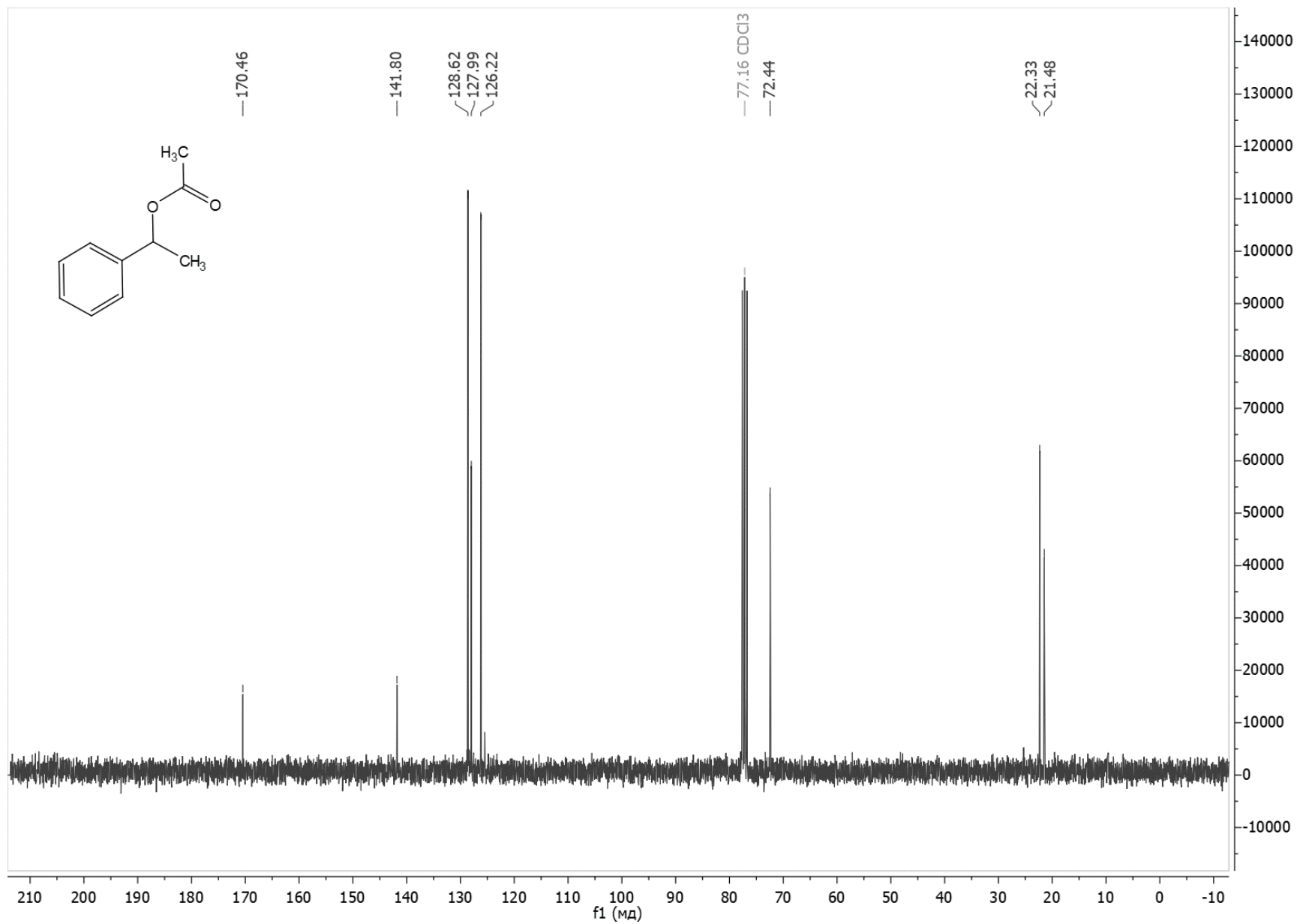
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-((*Tert*-butyldimethylsilyloxy)-3-phenylpropanoic acid, P4



¹H NMR (300.13 MHz, CDCl₃), 1-Phenylethyl acetate, P6

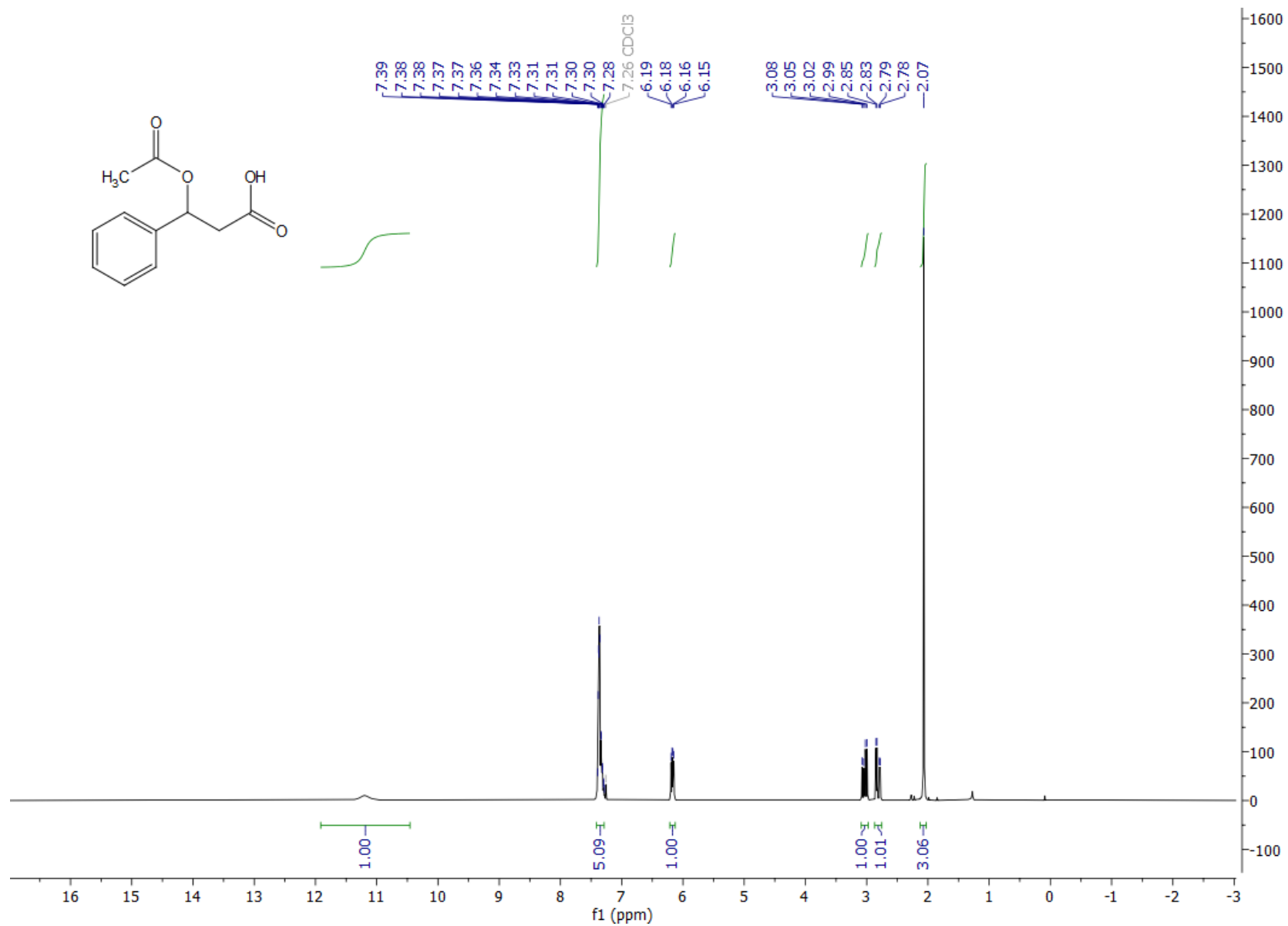


$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 1-Phenylethyl acetate, P6

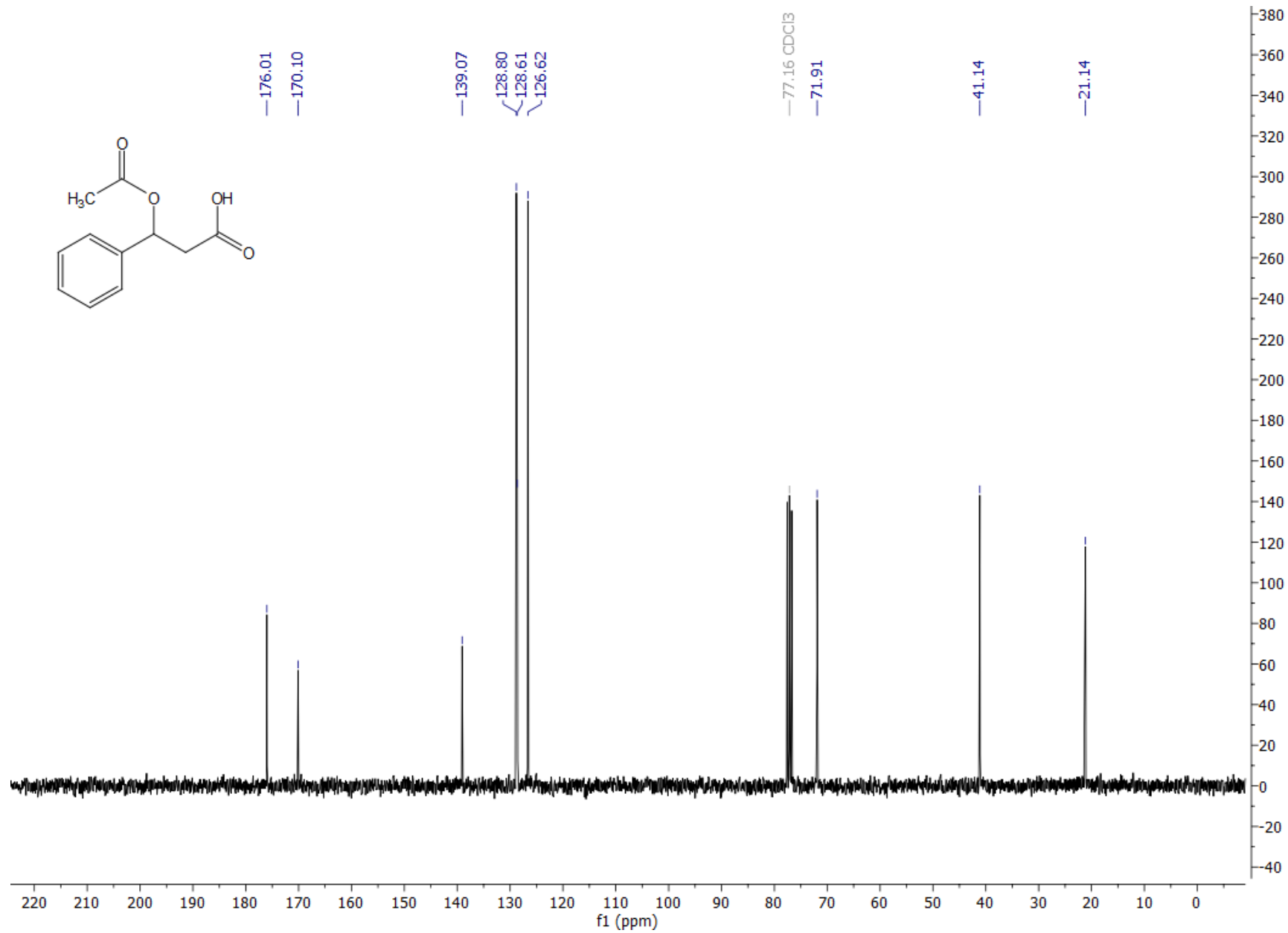


NMR spectra of products 2a-p

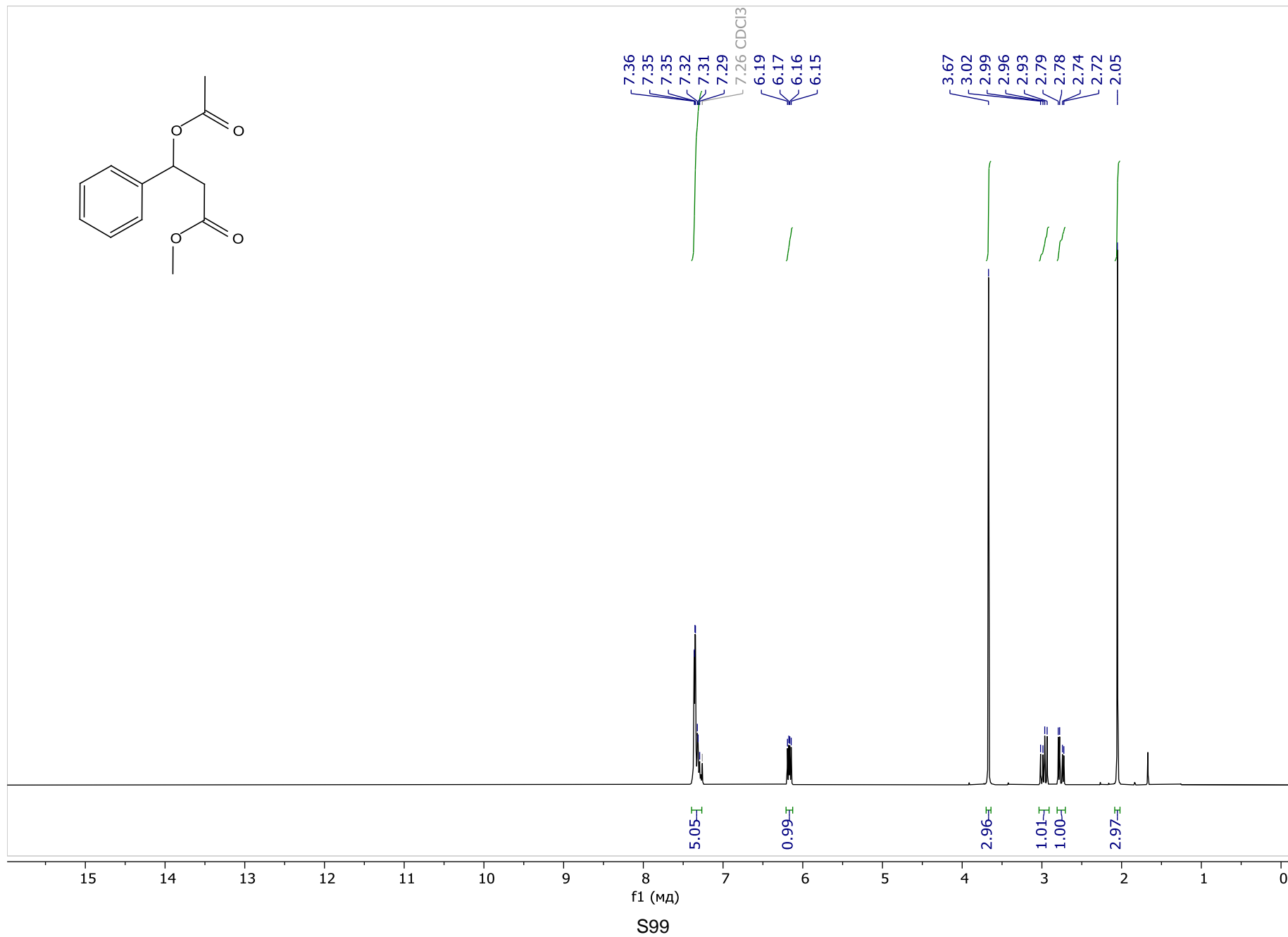
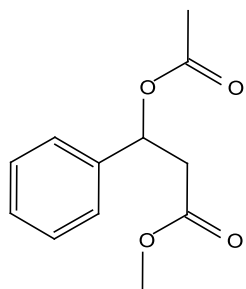
¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-3-phenylpropanoic acid, 2a



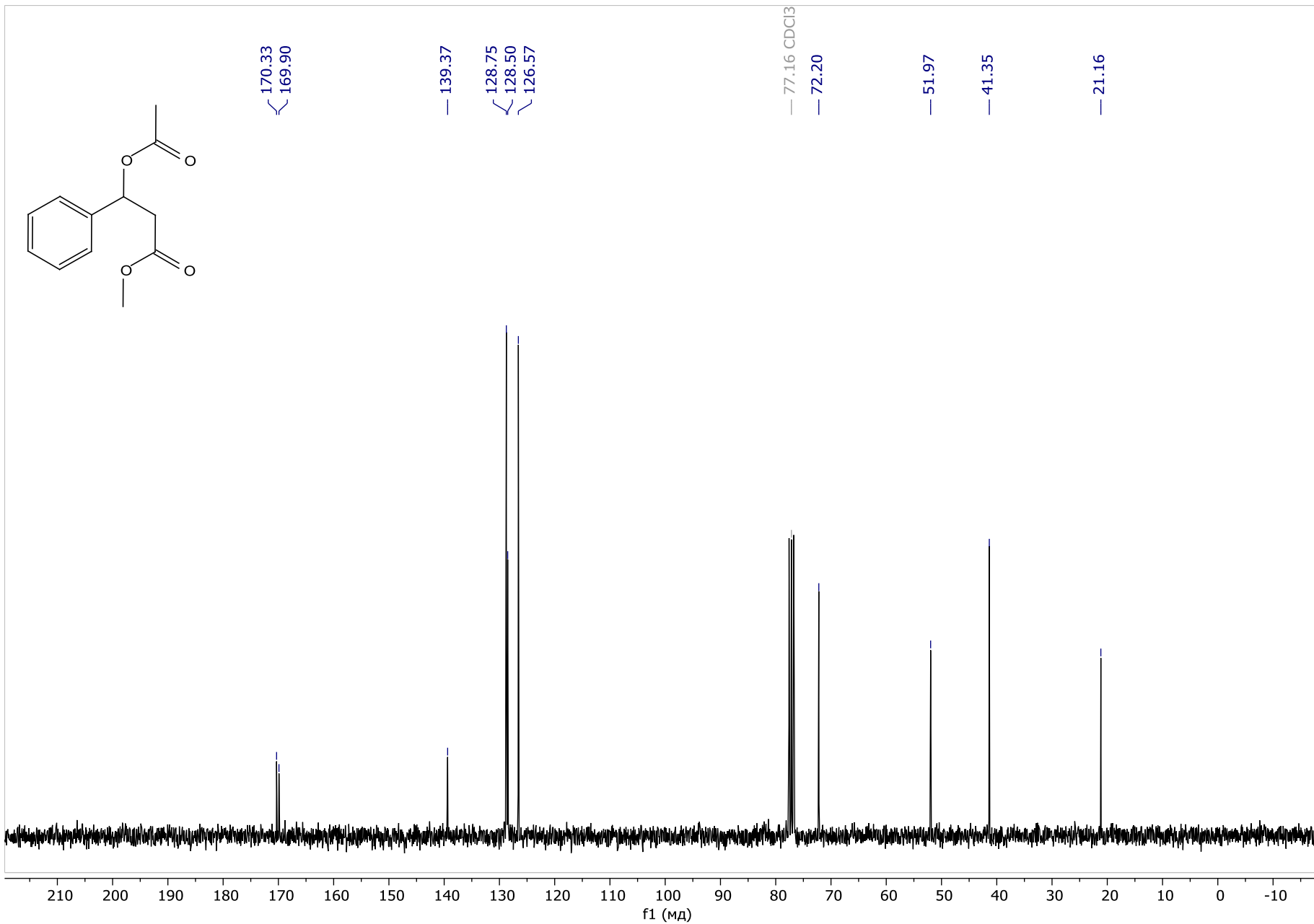
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-3-phenylpropanoic acid, 2a



¹H NMR (300.13 MHz, CDCl₃), **Methyl 3-acetoxy-3-phenylpropanoate, 2a'**

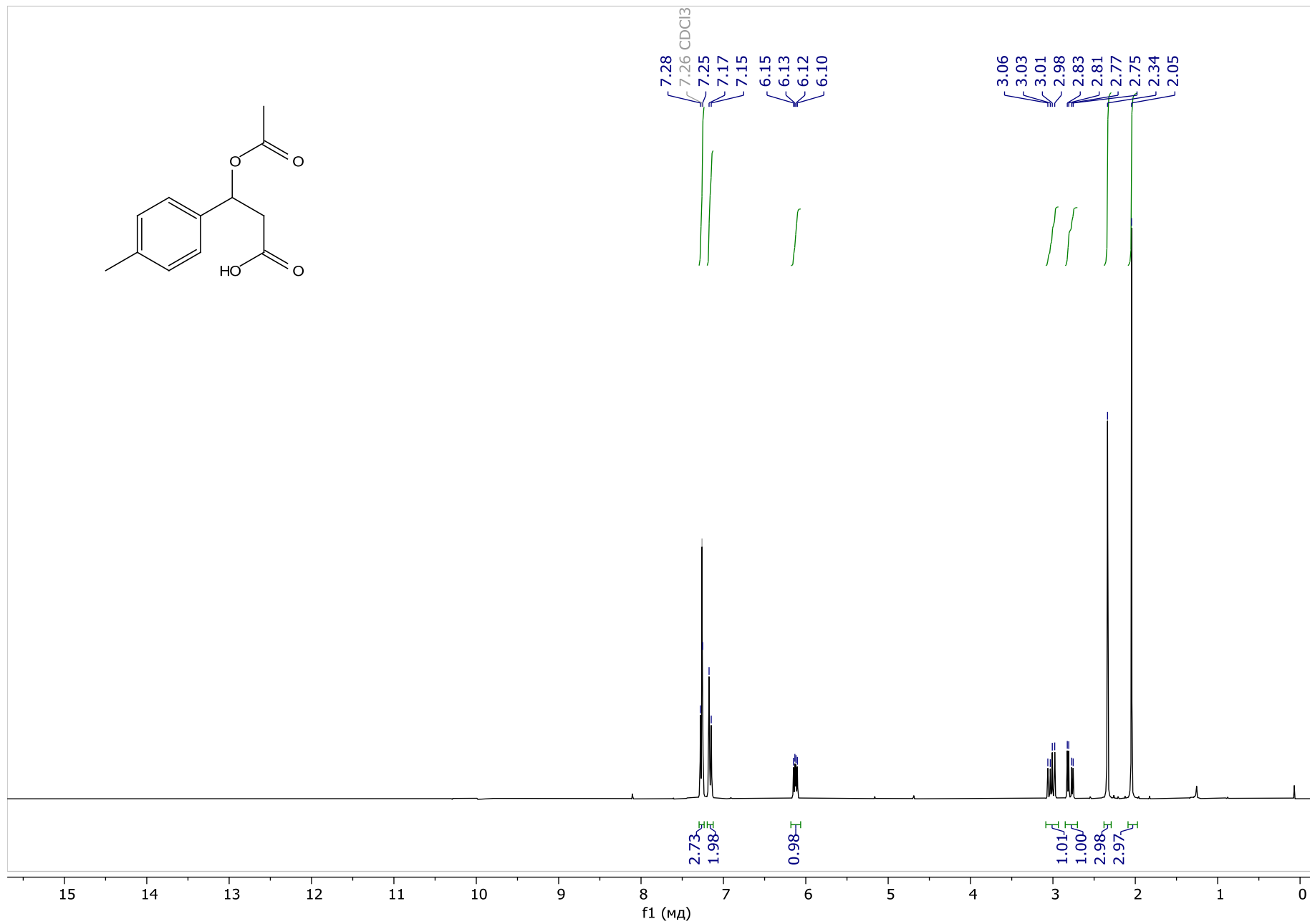
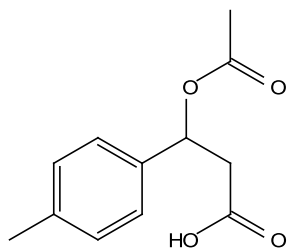


$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), Methyl 3-acetoxy-3-phenylpropanoate, 2a'



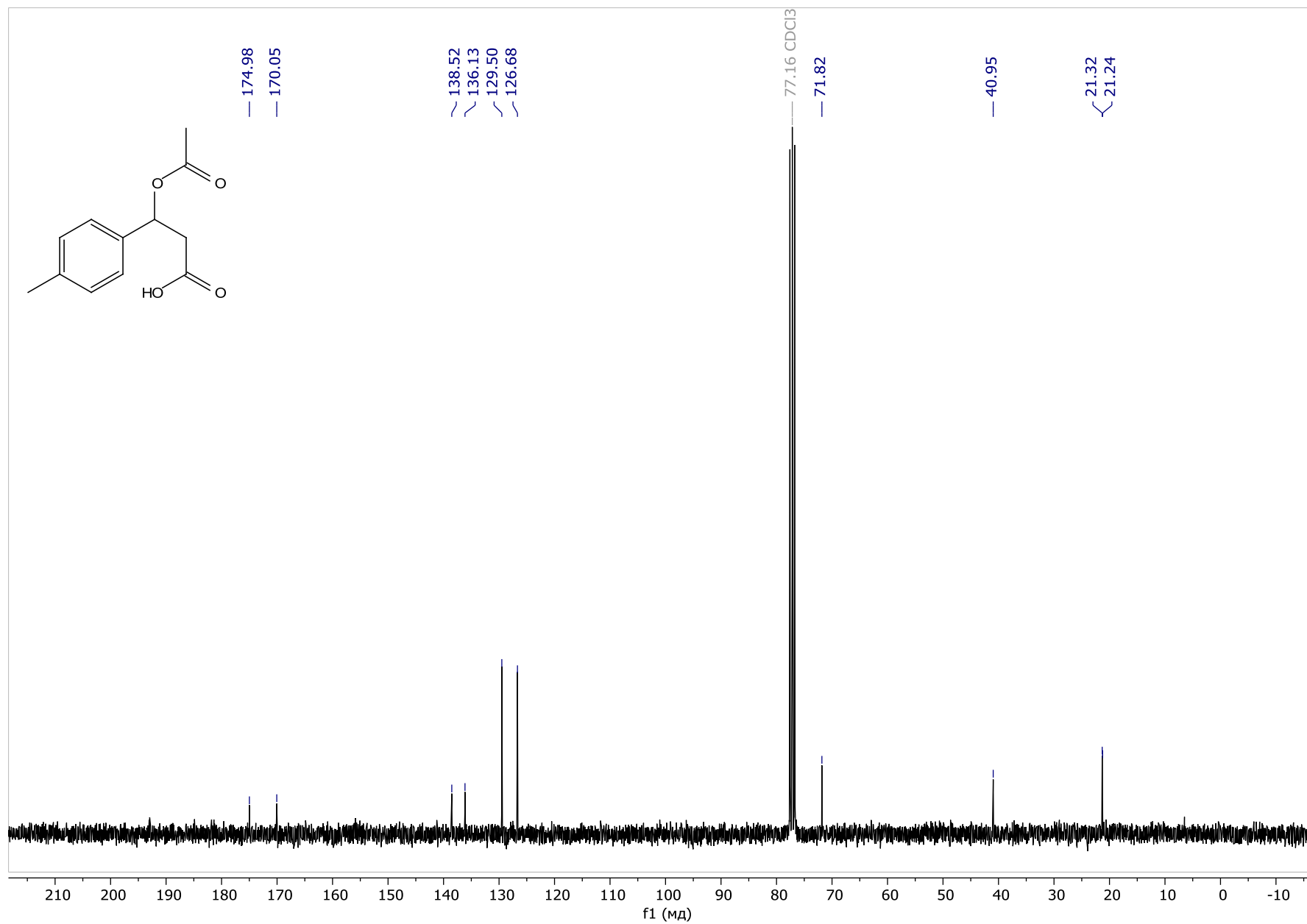
S100

¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-3-(p-tolyl)propanoic acid, 2b

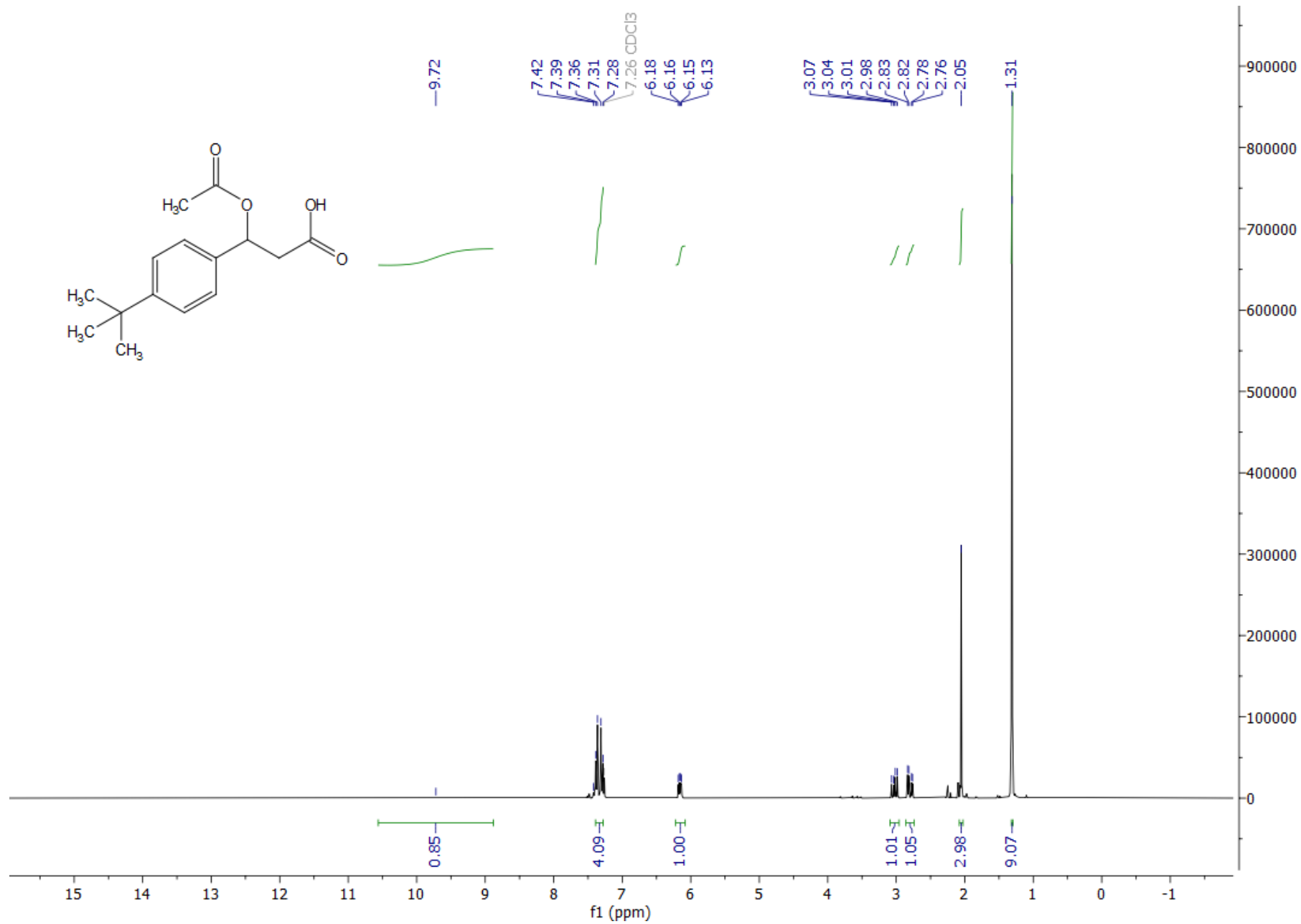


S101

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-3-(*p*-tolyl)propanoic acid, 2b

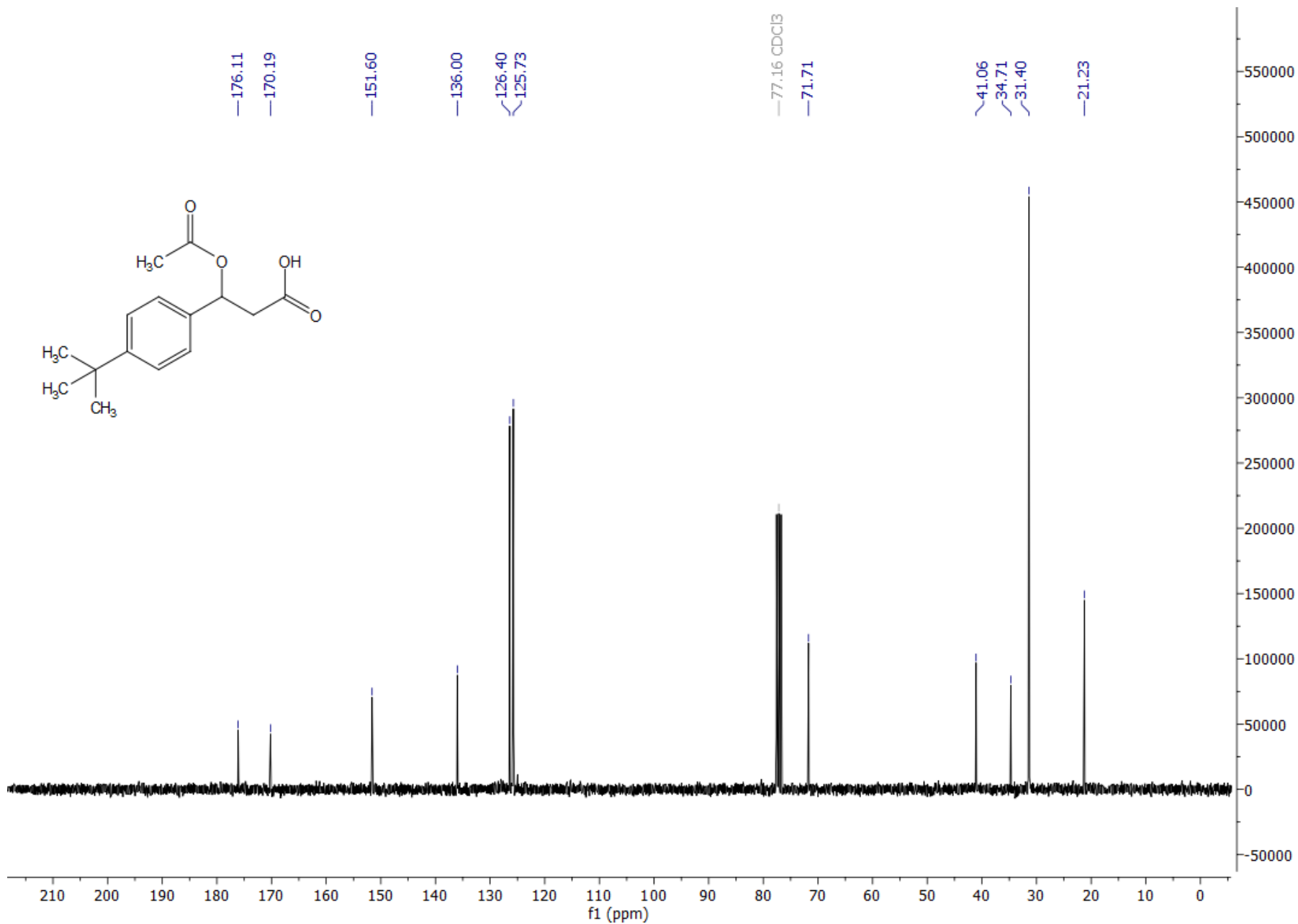


¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-3-(4-(*tert*-butyl)phenyl)propanoic acid, 2c

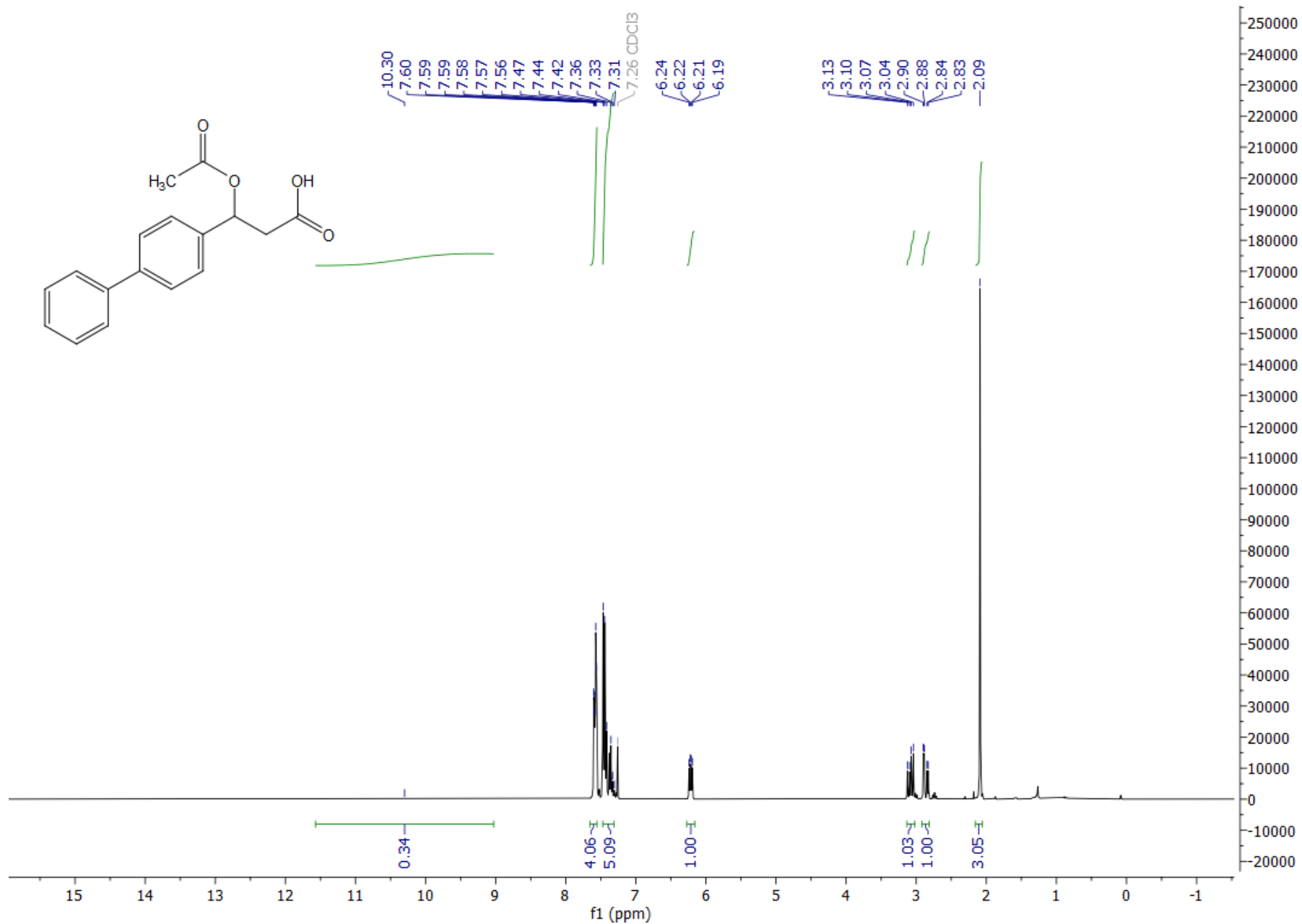


S103

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-3-(4-(*tert*-butyl)phenyl)propanoic acid, 2c

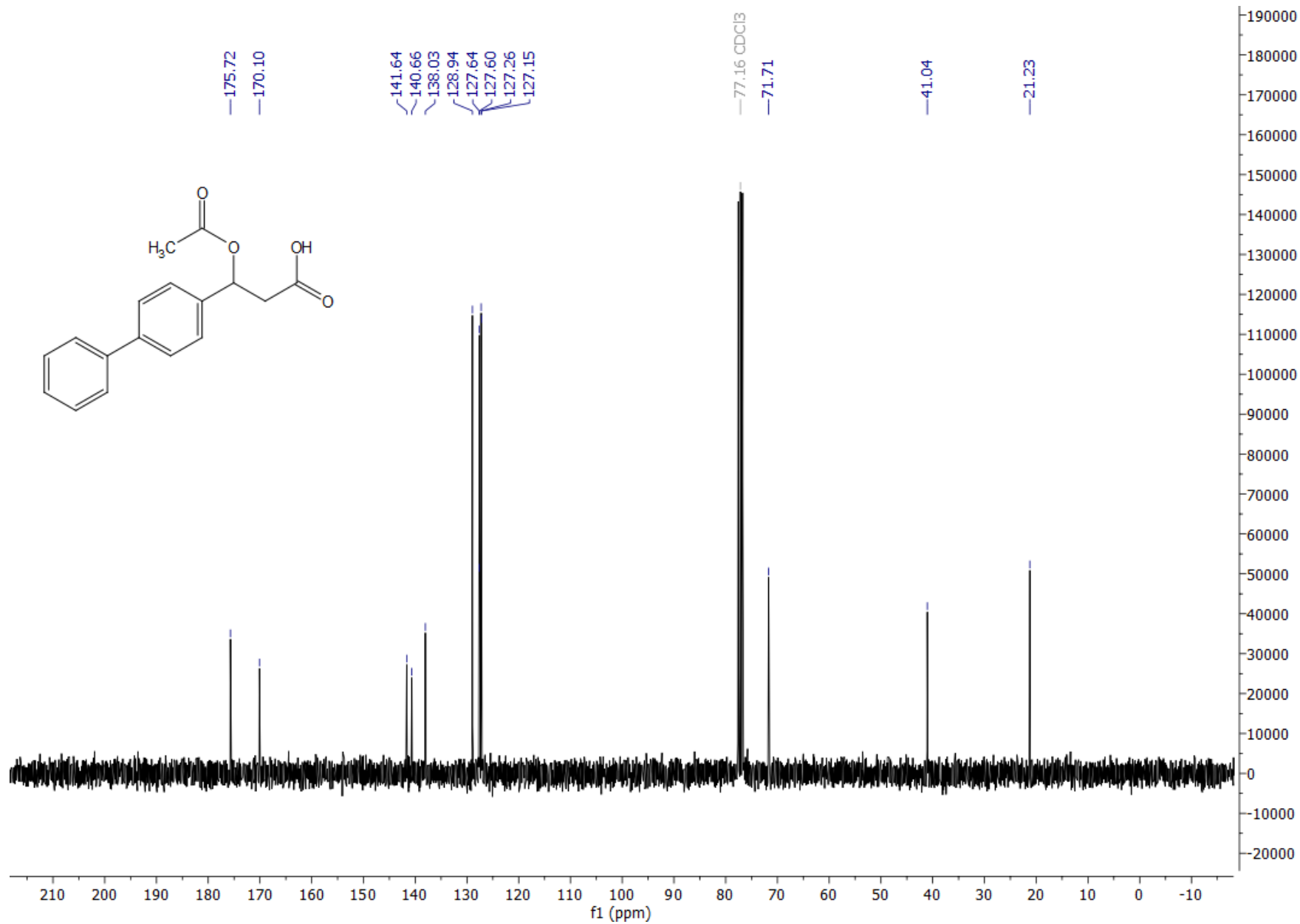


¹H NMR (300.13 MHz, CDCl₃), 3-([1,1'-Biphenyl]-4-yl)-3-acetoxypromanoic acid, 2d

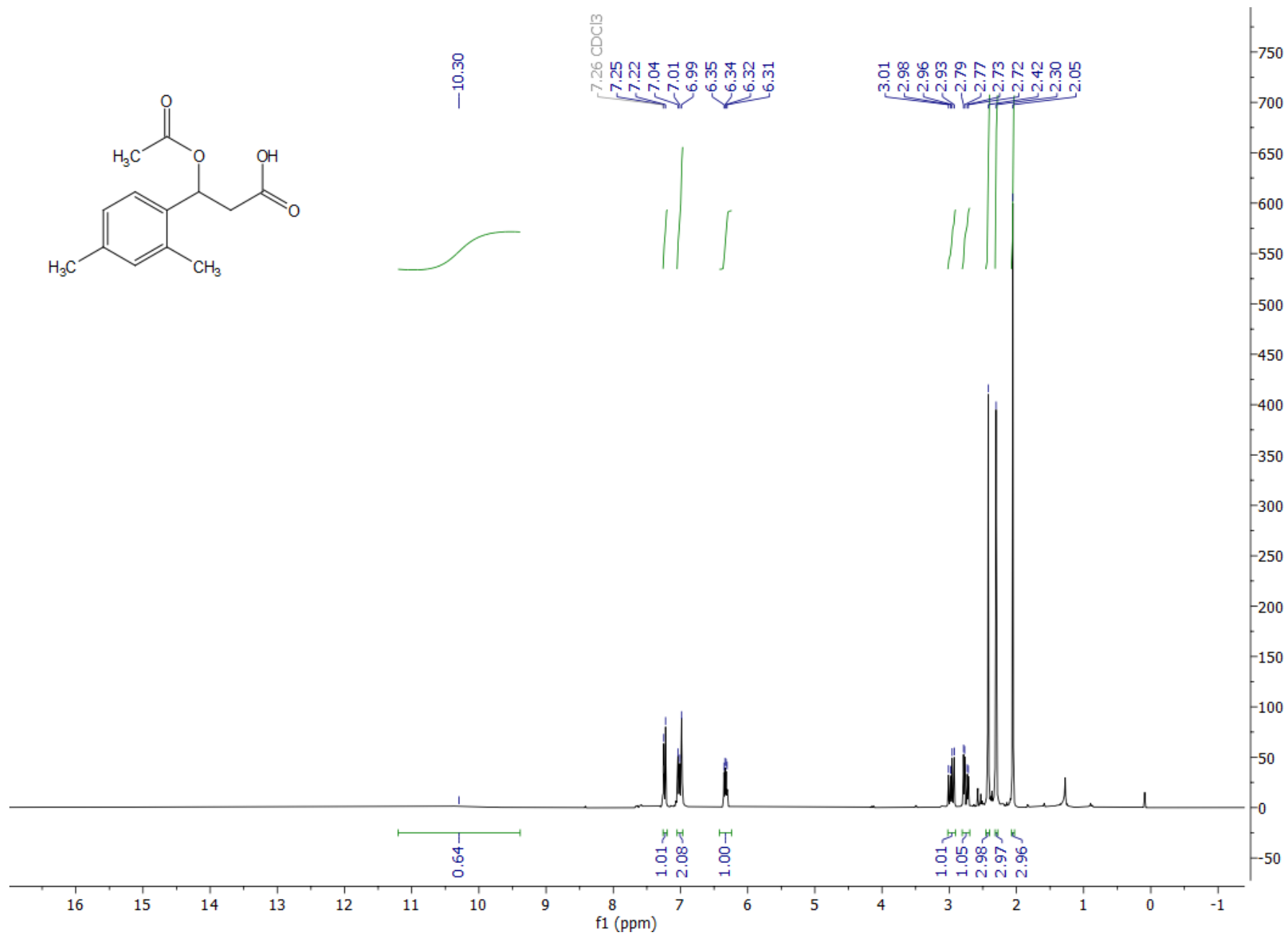
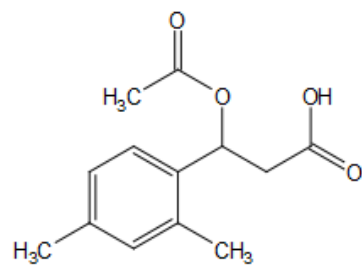


S105

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-([1,1'-Biphenyl]-4-yl)-3-acetoxypropanoic acid, 2d

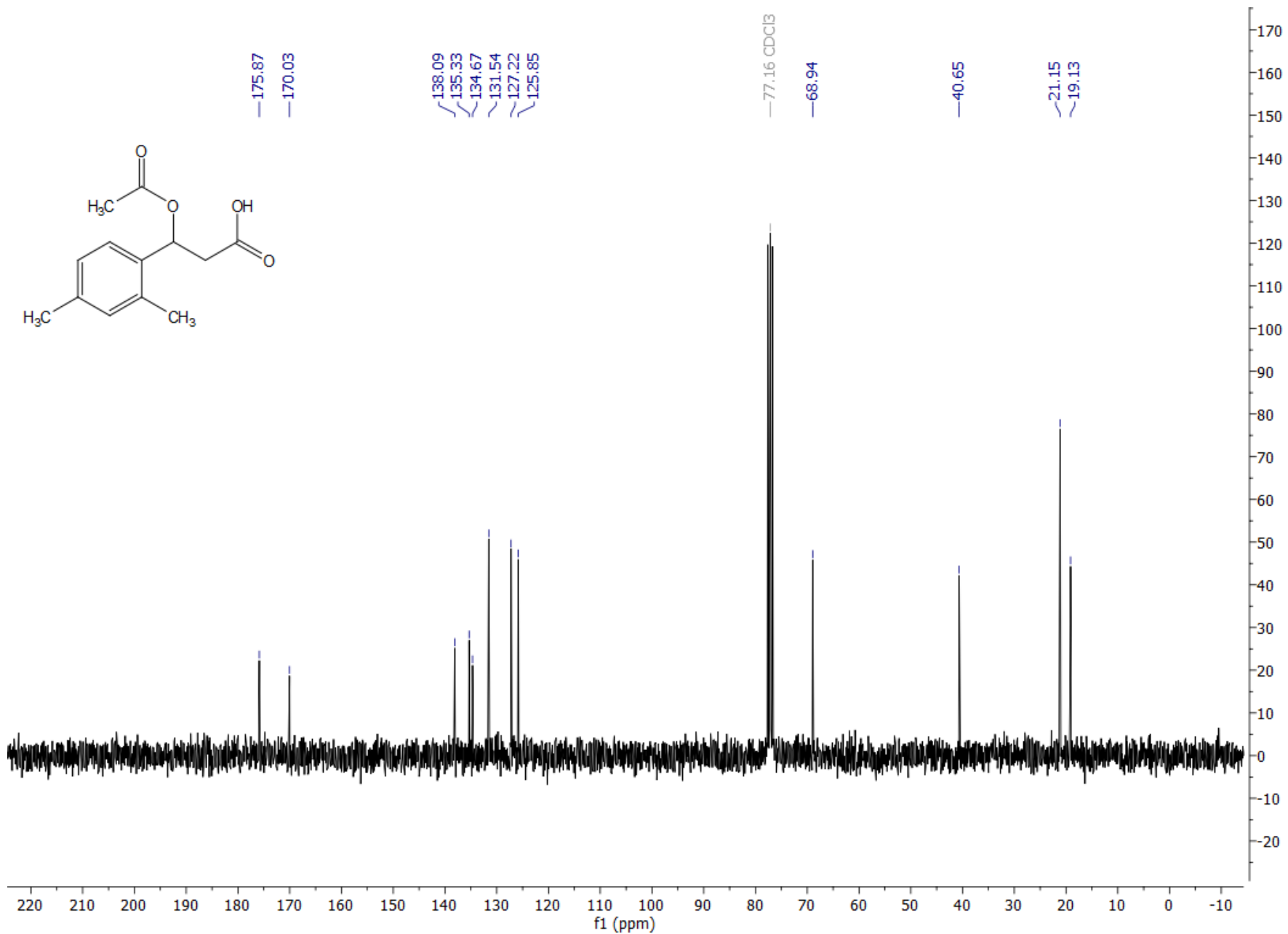


¹H NMR (300.13 MHz, CDCl₃), **3-Acetoxy-3-(2,4-dimethylphenyl)propanoic acid, 2e**

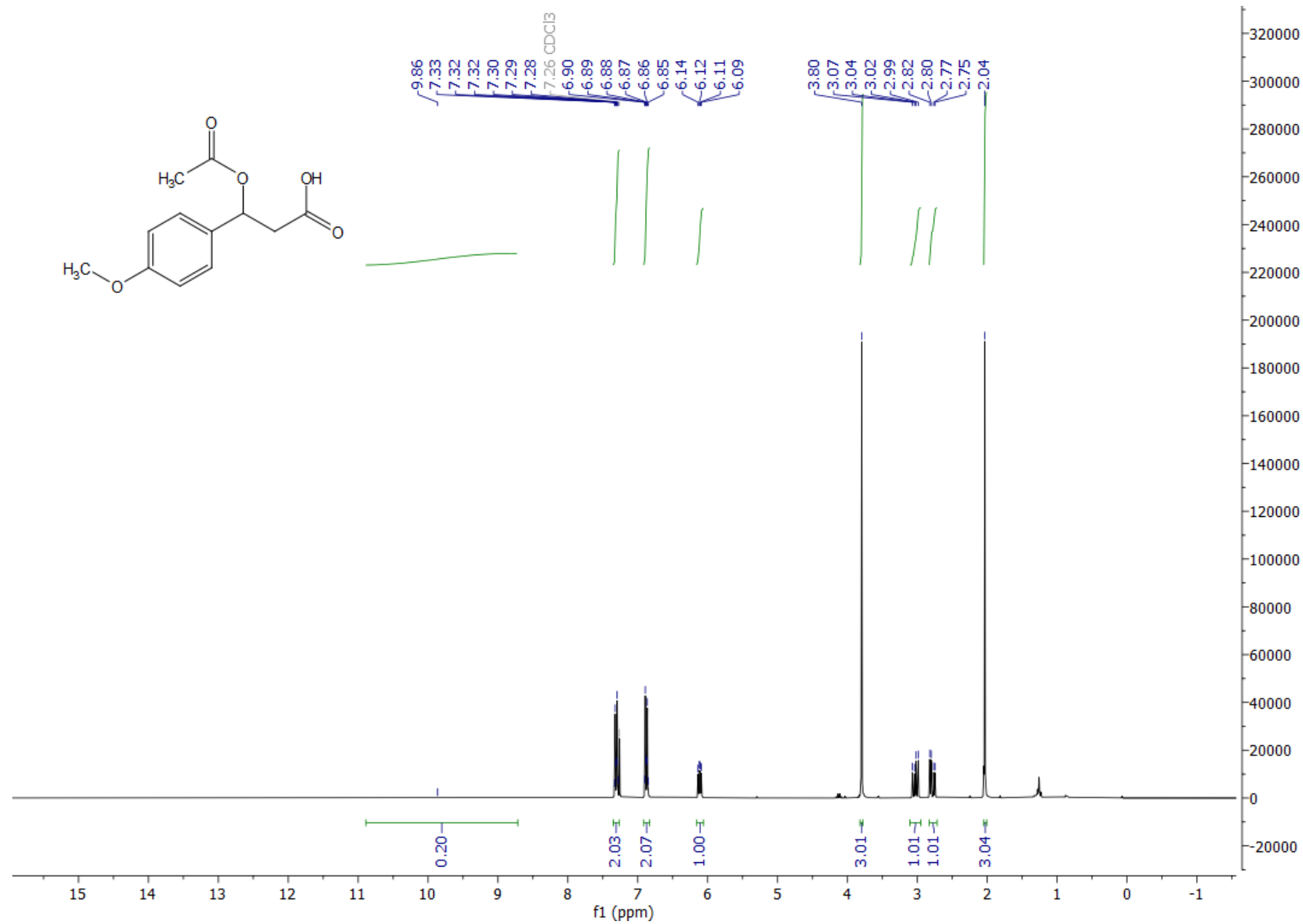


S107

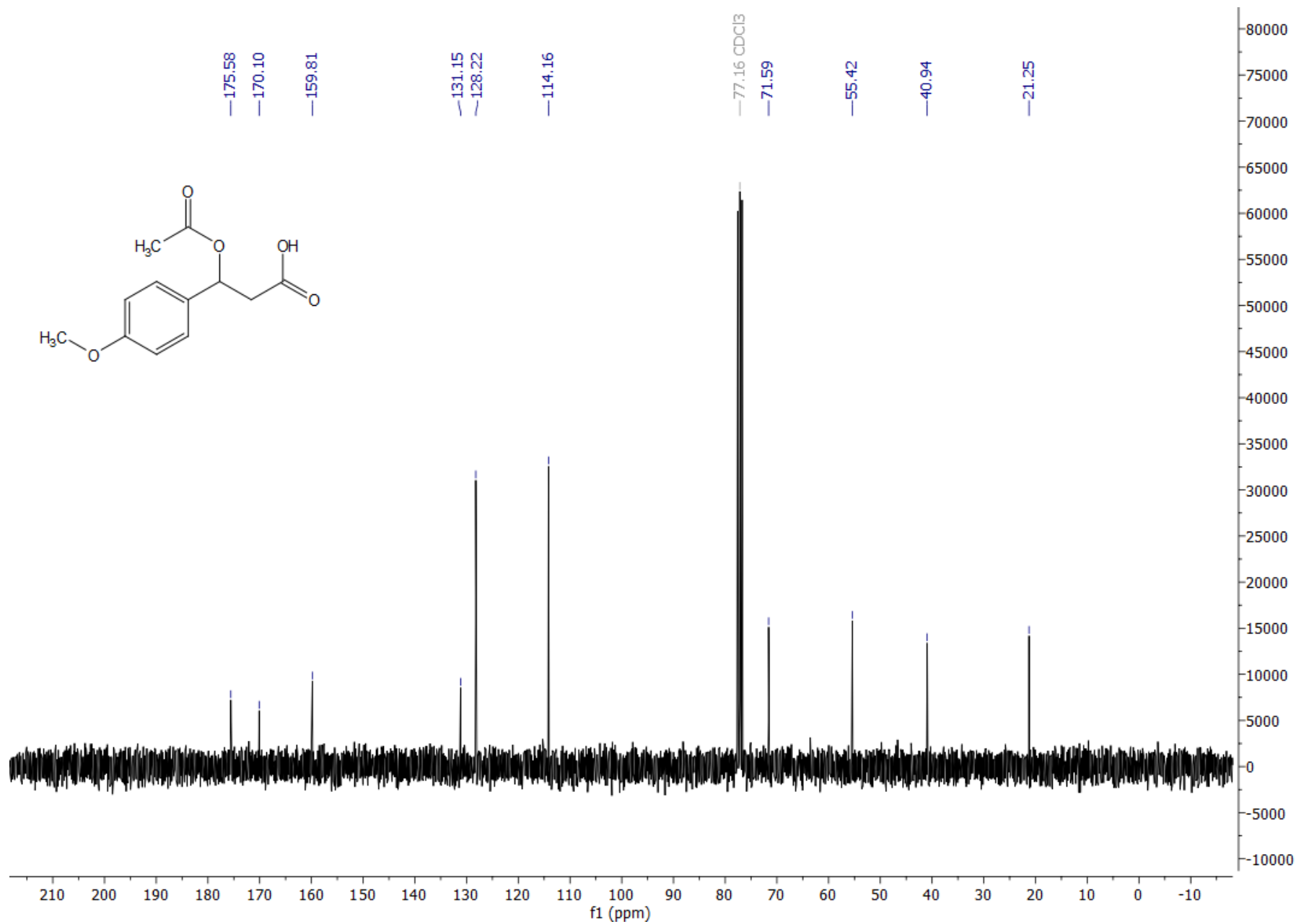
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-3-(2,4-dimethylphenyl)propanoic acid, 2e



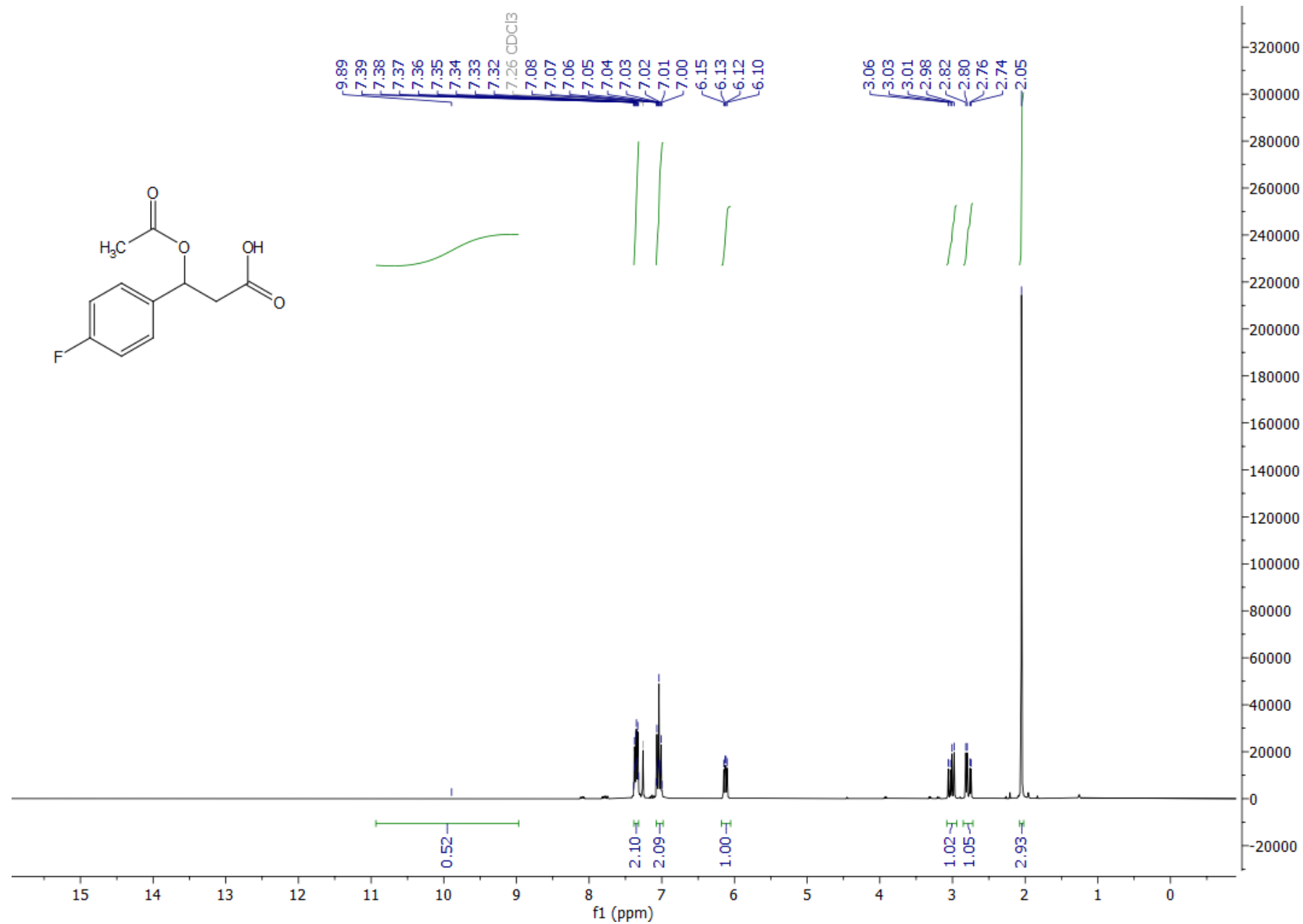
¹H NMR (300.13 MHz, CDCl₃), **3-Acetoxy-3-(4-methoxyphenyl)propanoic acid, 2f**



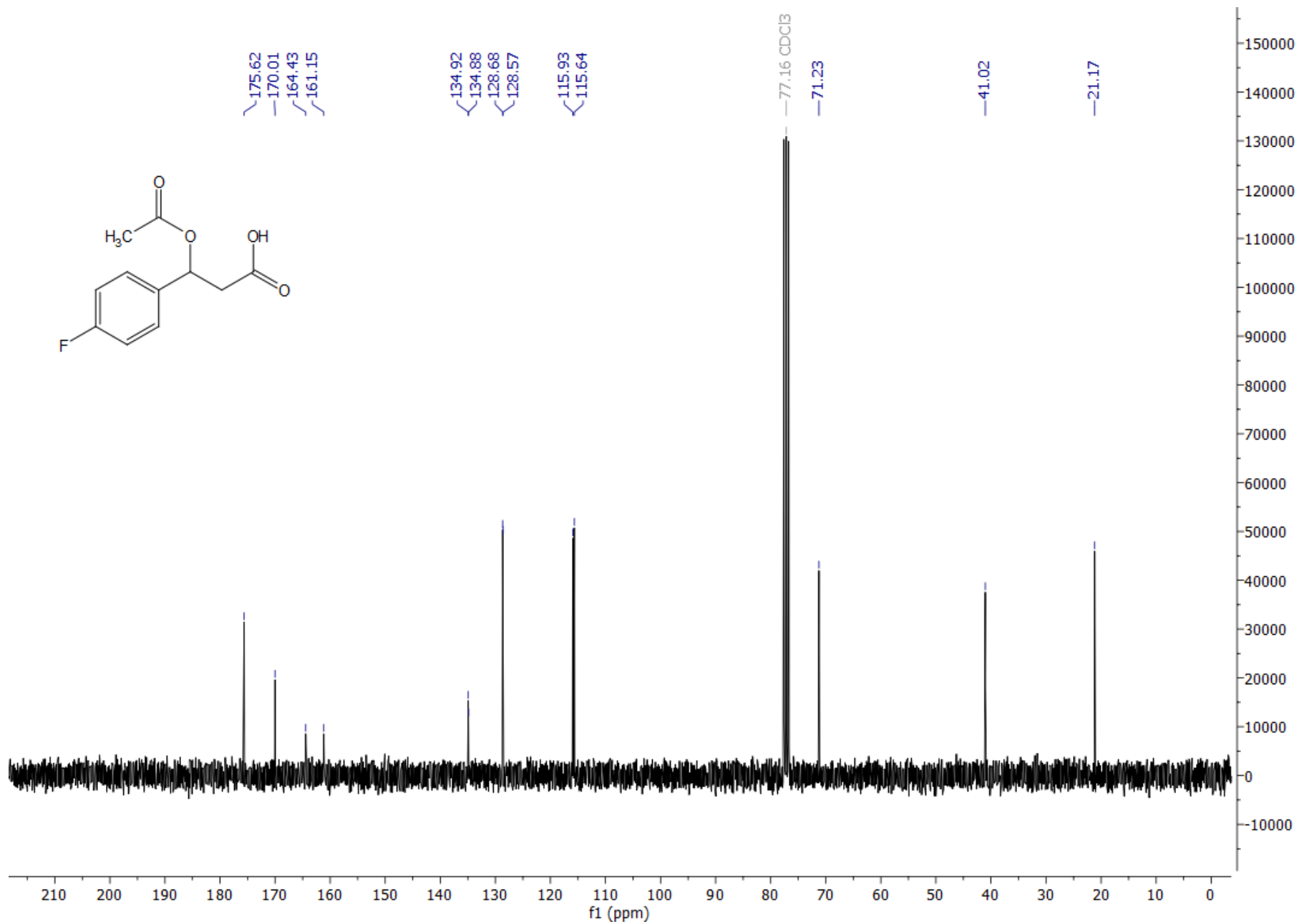
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Acetoxy-3-(4-methoxyphenyl)propanoic acid, 2f**



¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-3-(4-fluorophenyl)propanoic acid, 2g



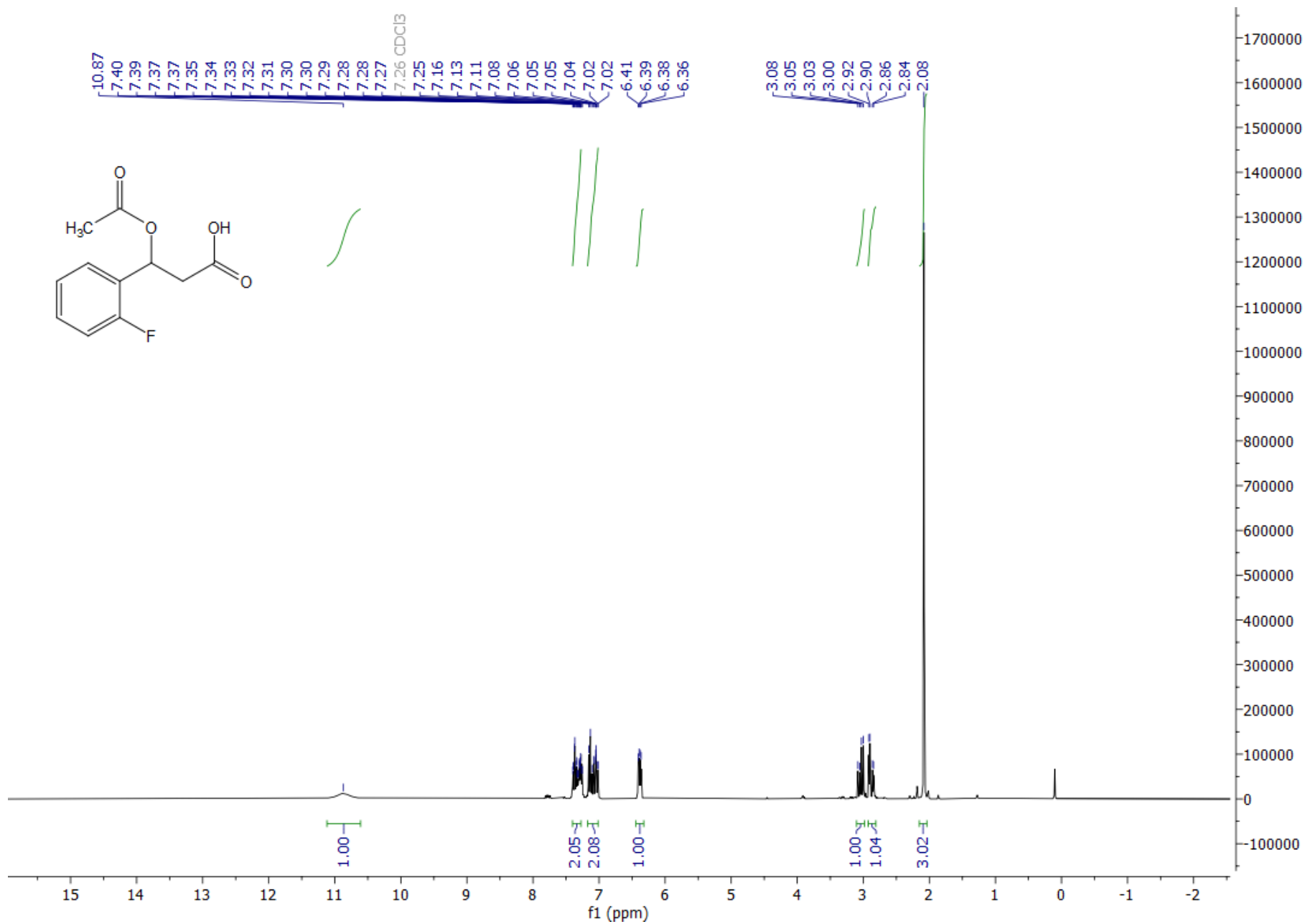
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-3-(4-fluorophenyl)propanoic acid, 2g



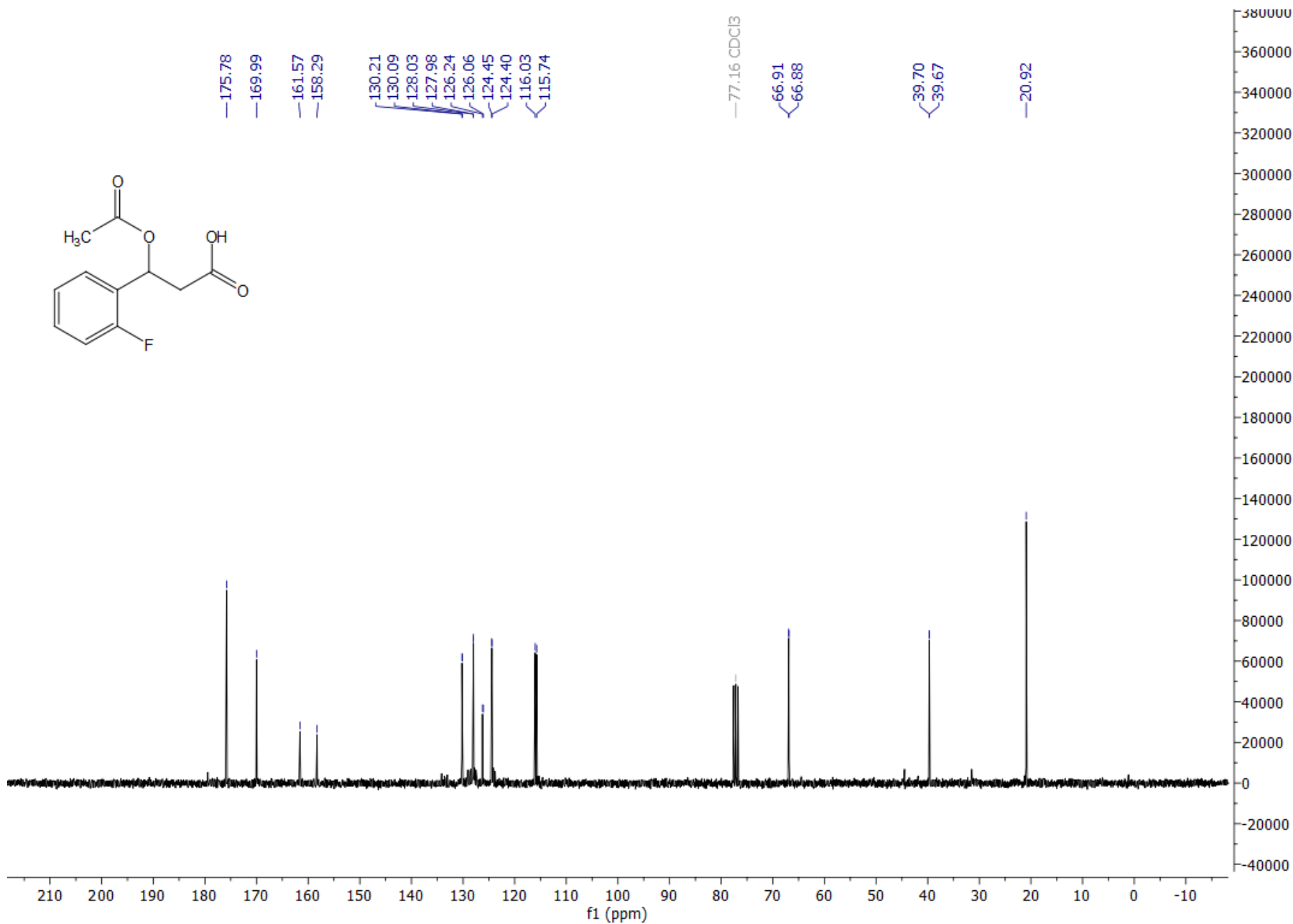
¹⁹F NMR (282,5 MHz, CDCl₃), 3-Acetoxy-3-(4-fluorophenyl)propanoic acid, 2g



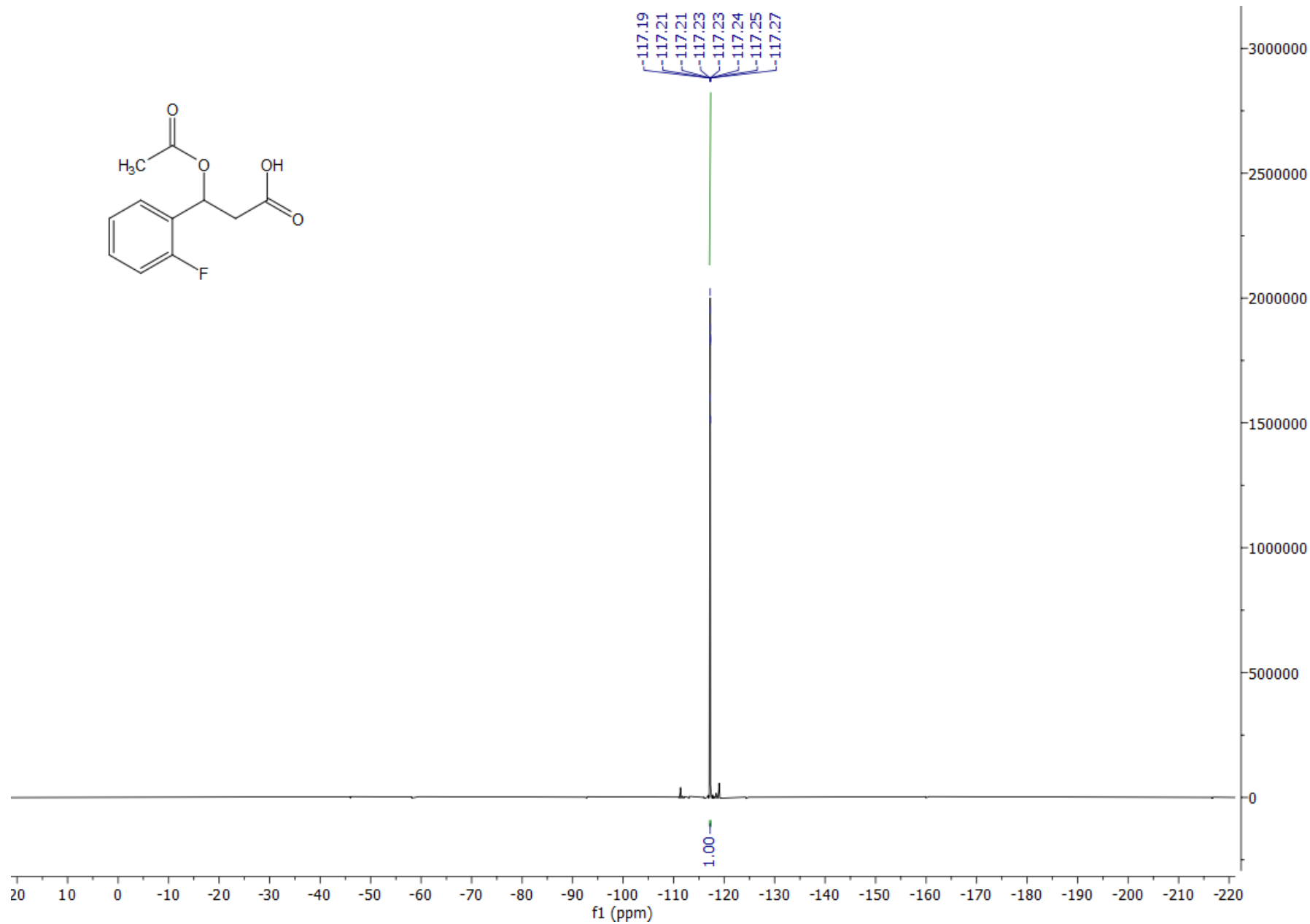
¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-3-(2-fluorophenyl)propanoic acid, 2h



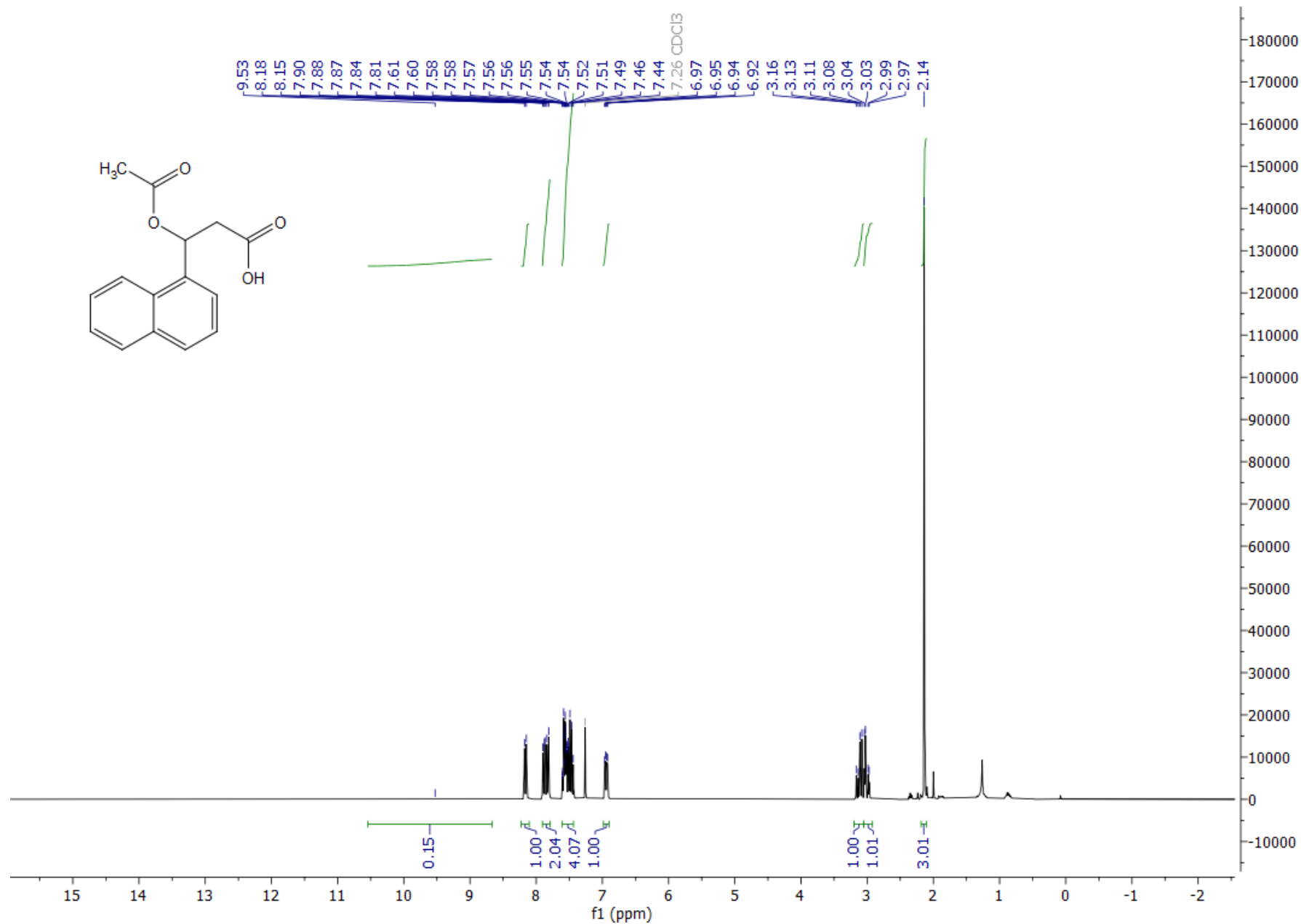
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-3-(2-fluorophenyl)propanoic acid, 2h



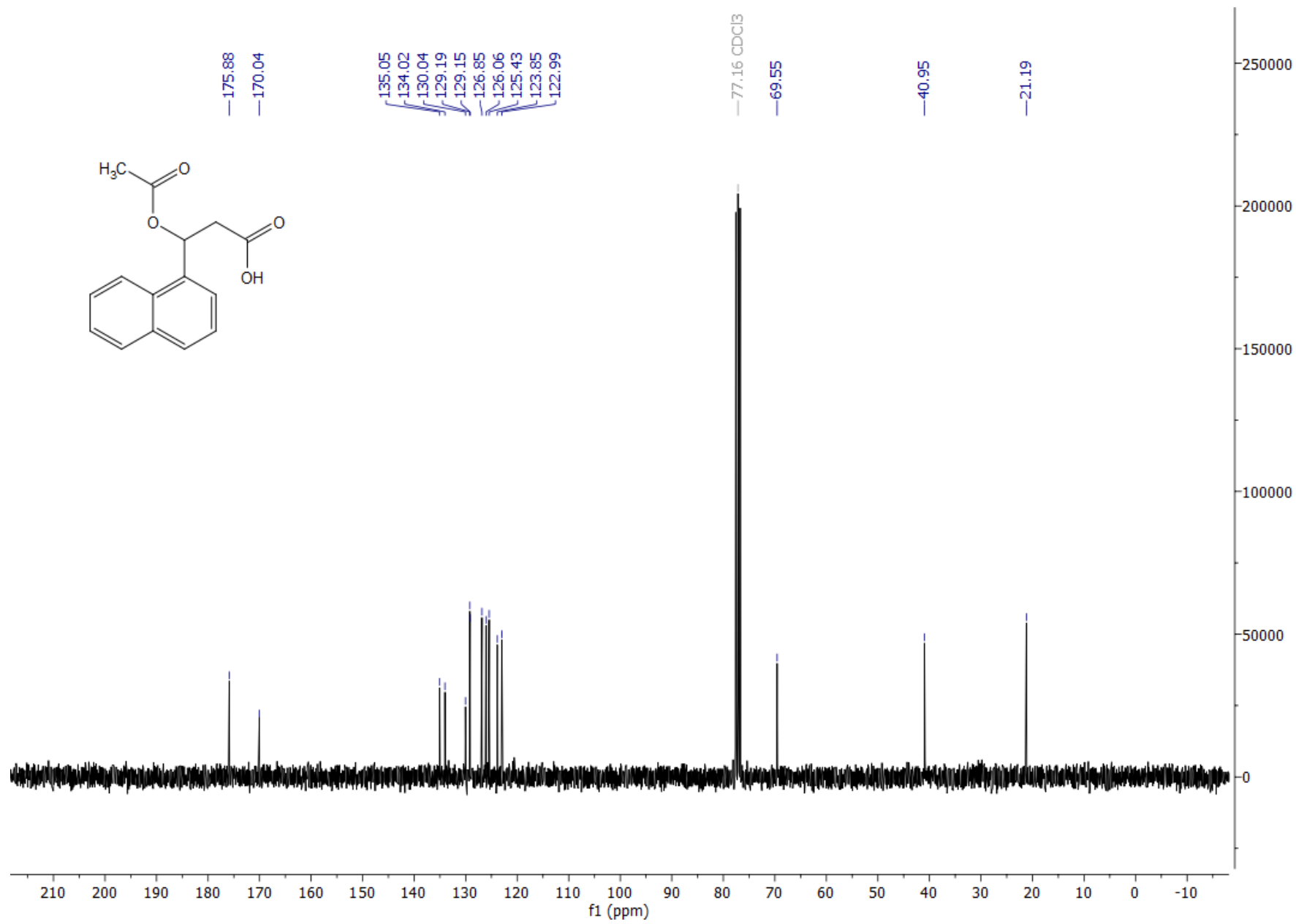
^{19}F NMR (282,5 MHz, CDCl_3), 3-Acetoxy-3-(2-fluorophenyl)propanoic acid, 2h



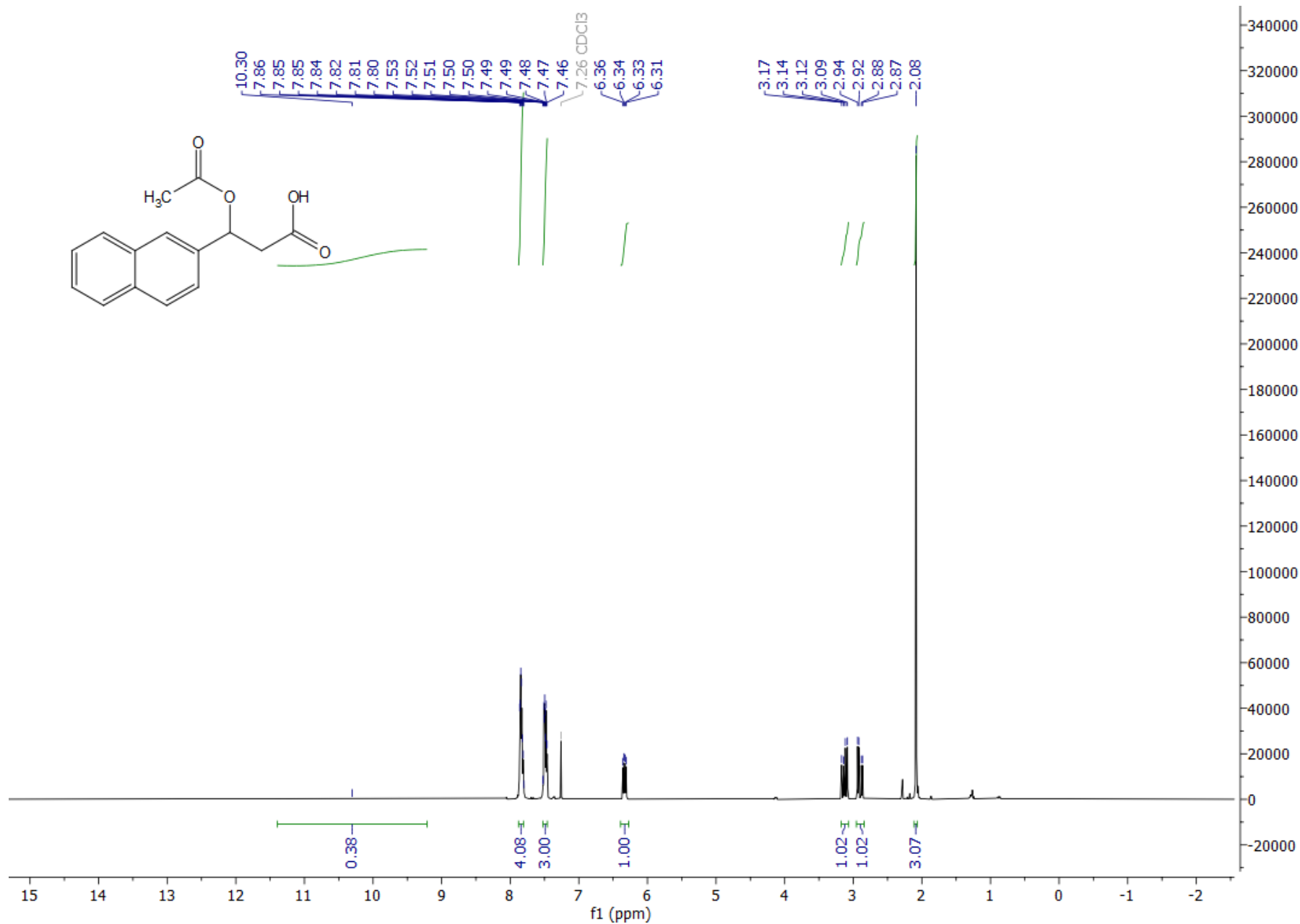
¹H NMR (300.13 MHz, CDCl₃), **3-Acetoxy-3-(naphthalen-1-yl)propanoic acid, 2i**



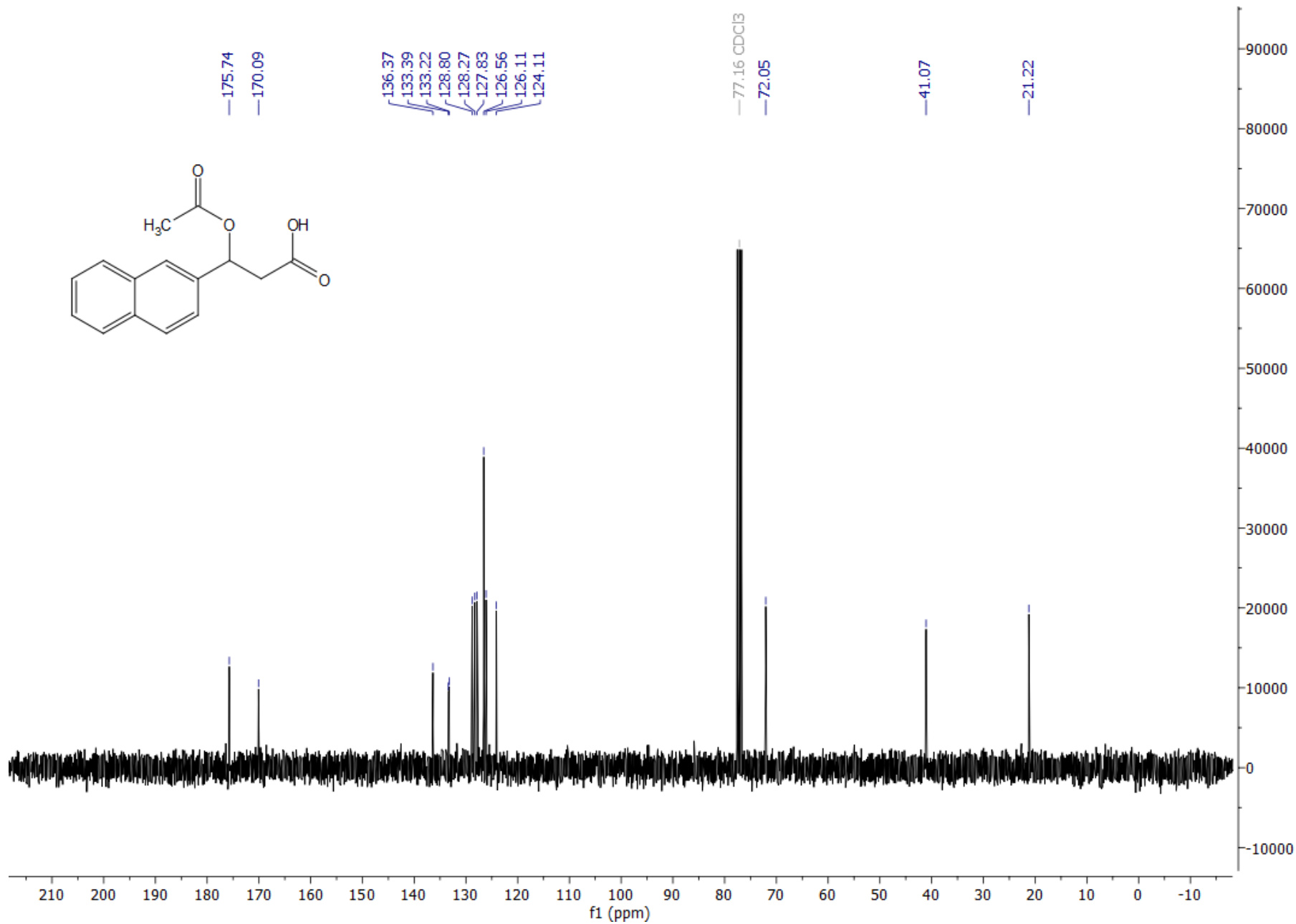
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Acetoxy-3-(naphthalen-1-yl)propanoic acid, 2i**



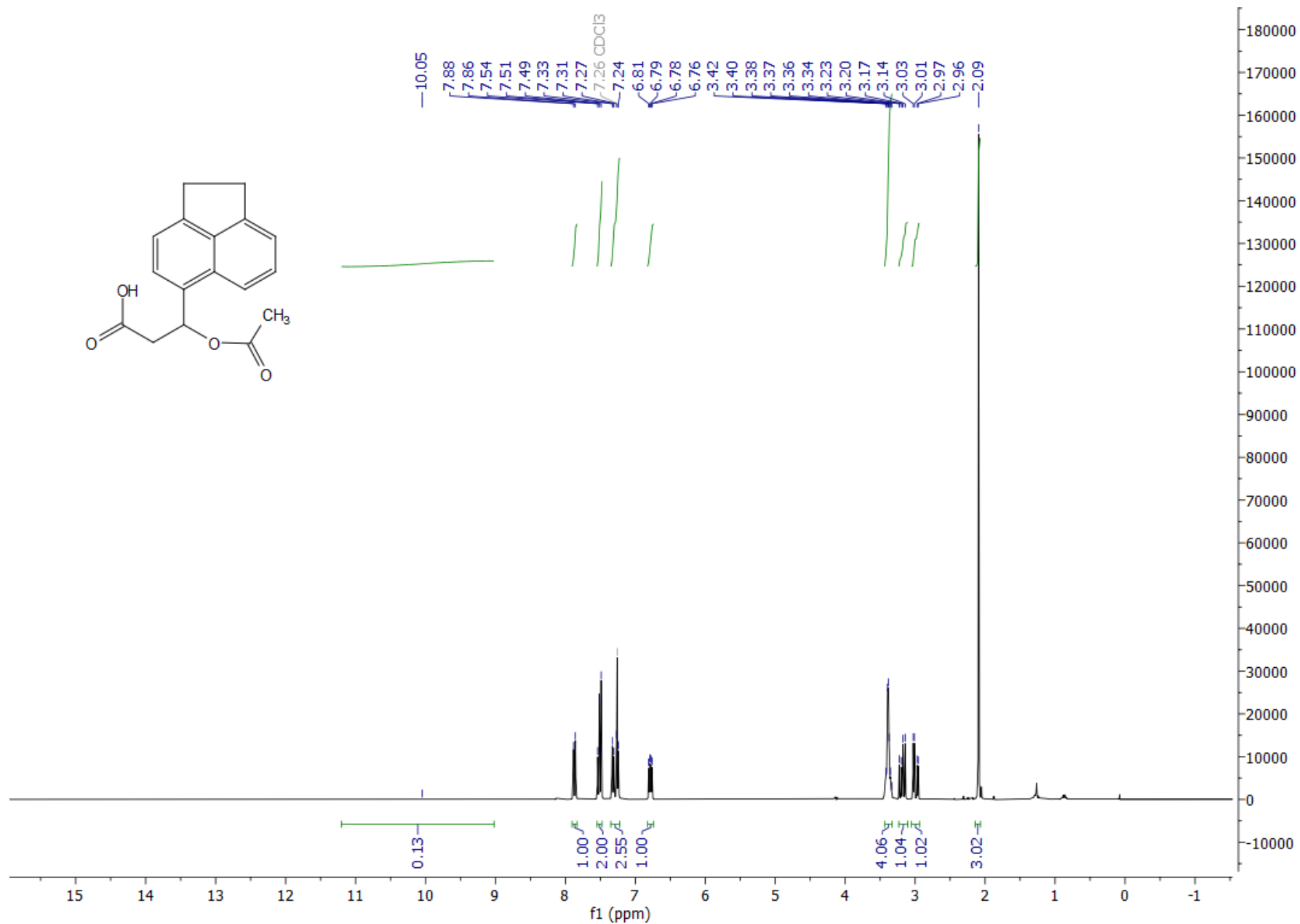
¹H NMR (300.13 MHz, CDCl₃), **3-Acetoxy-3-(naphthalen-2-yl)propanoic acid, 2j**



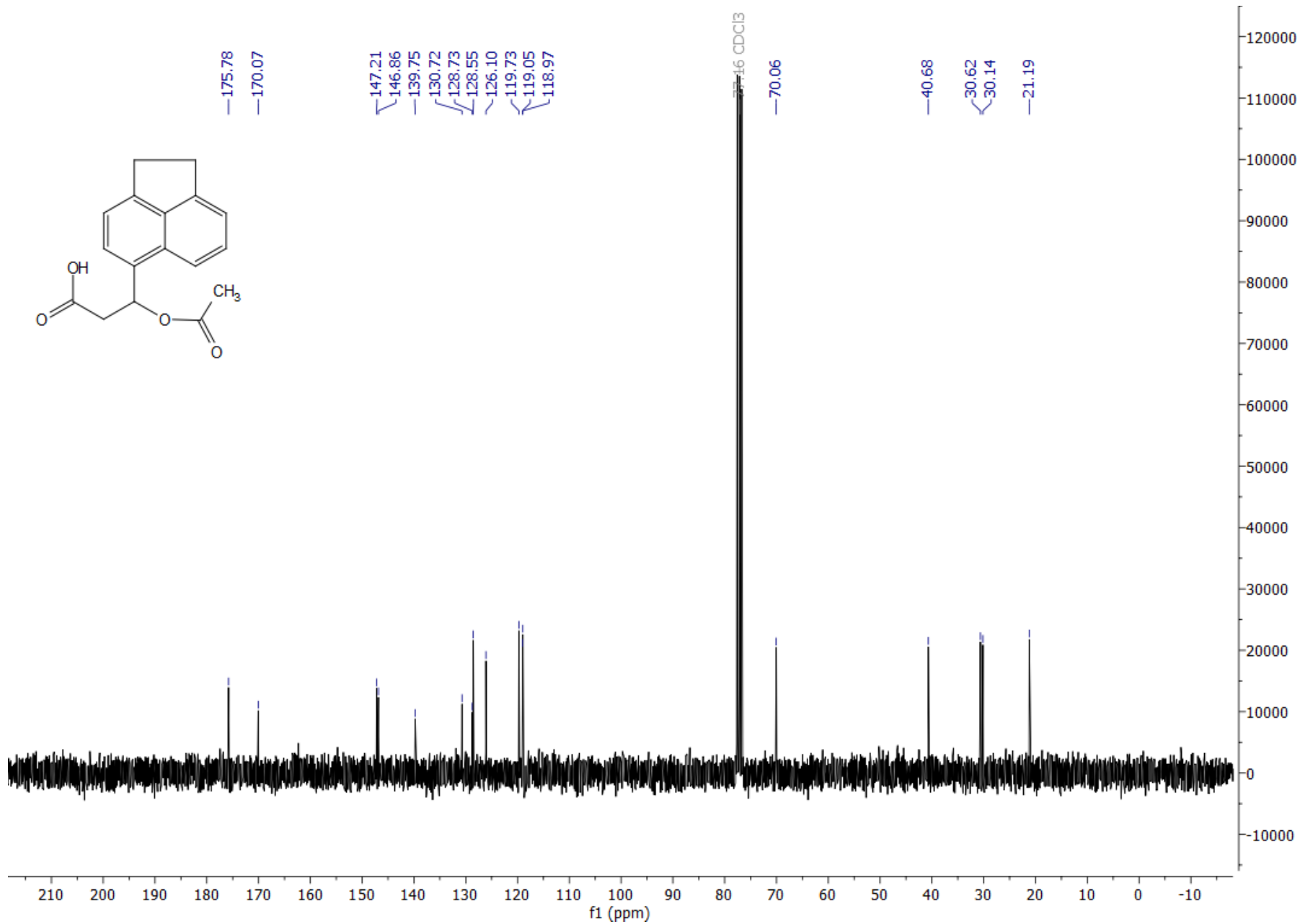
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Acetoxy-3-(naphthalen-2-yl)propanoic acid, 2j**



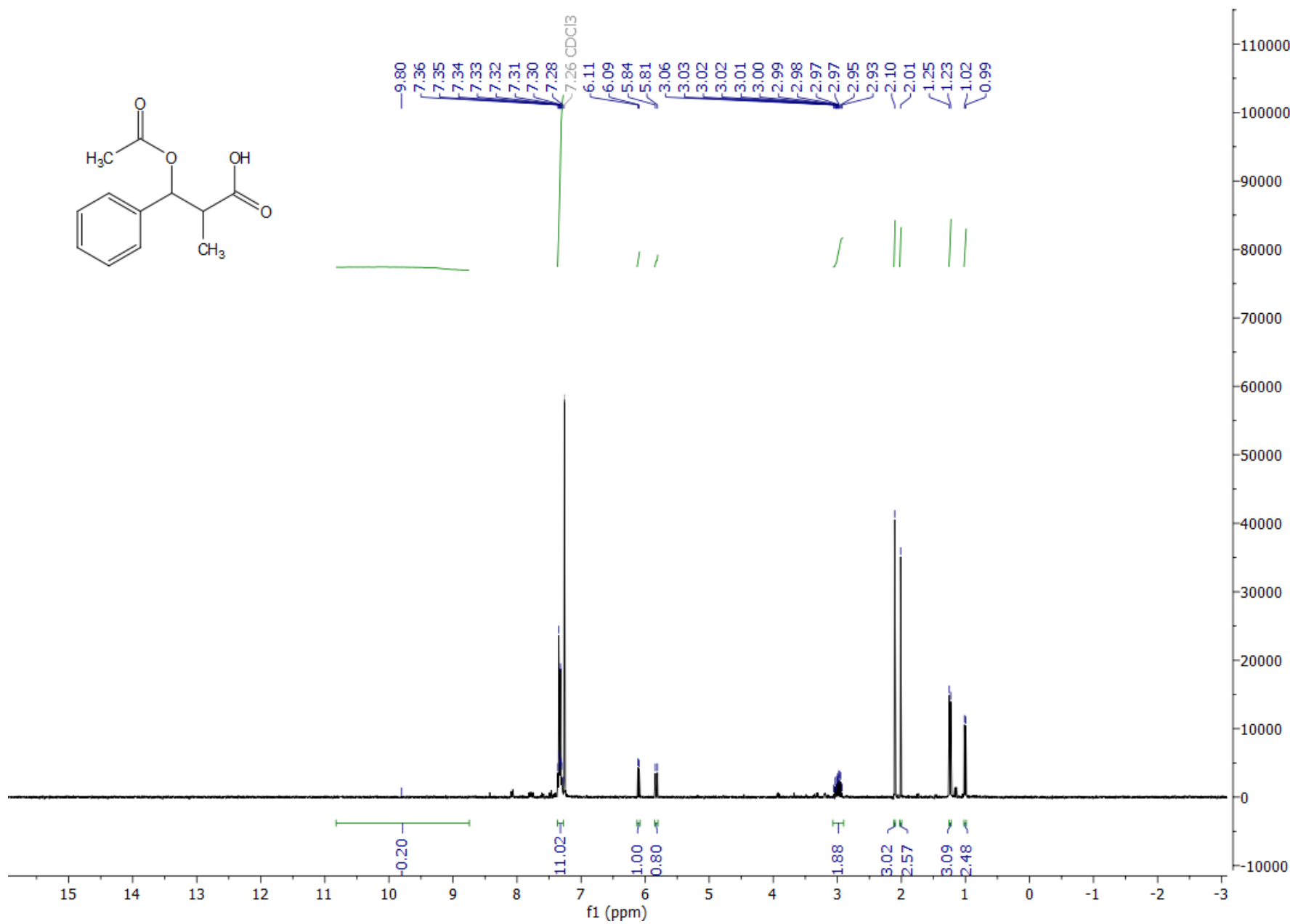
¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-3-(1,2-dihydroacenaphthylen-5-yl)propanoic acid, 2k



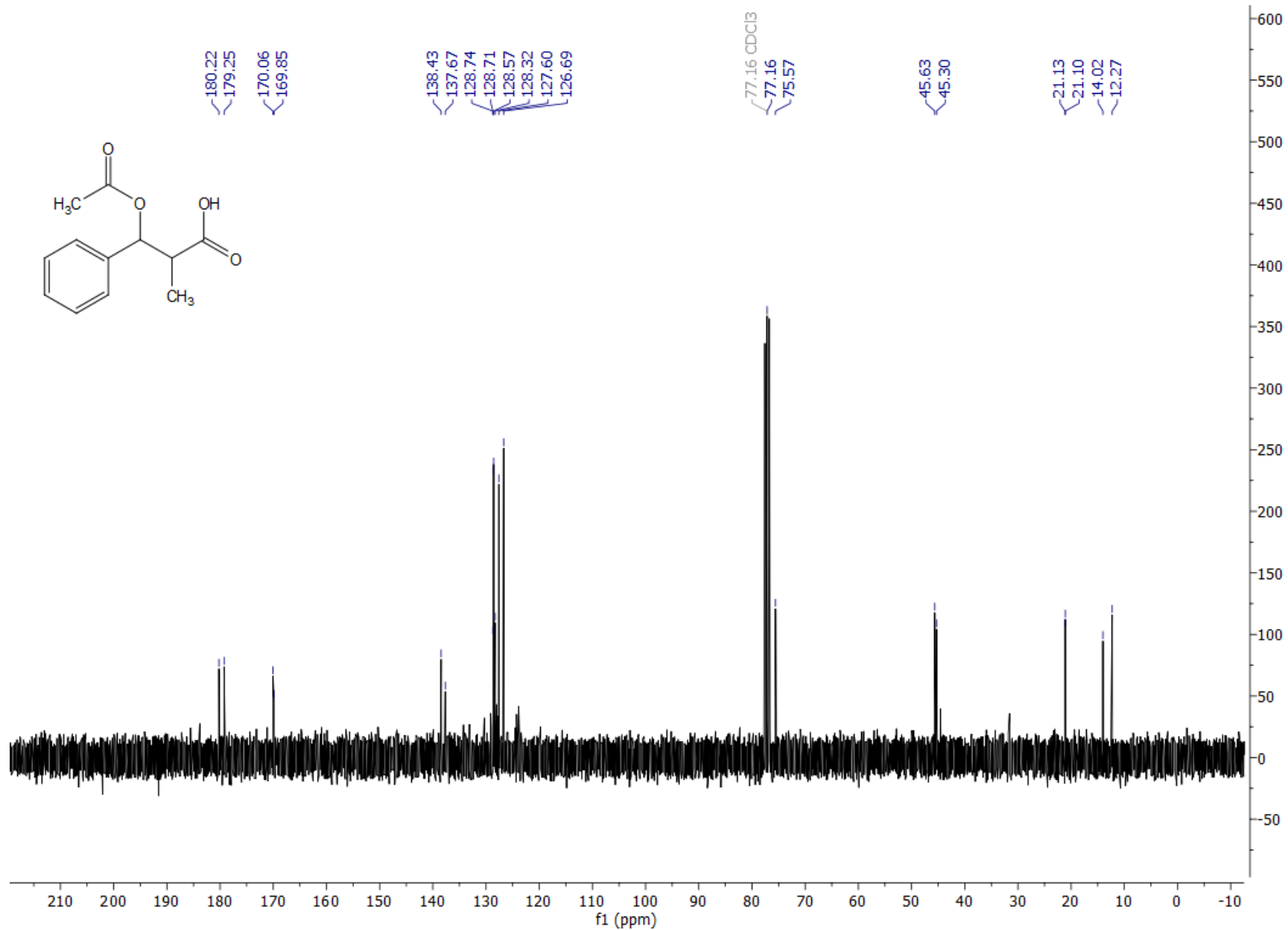
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-3-(1,2-dihydroacenaphthylen-5-yl)propanoic acid, 2k



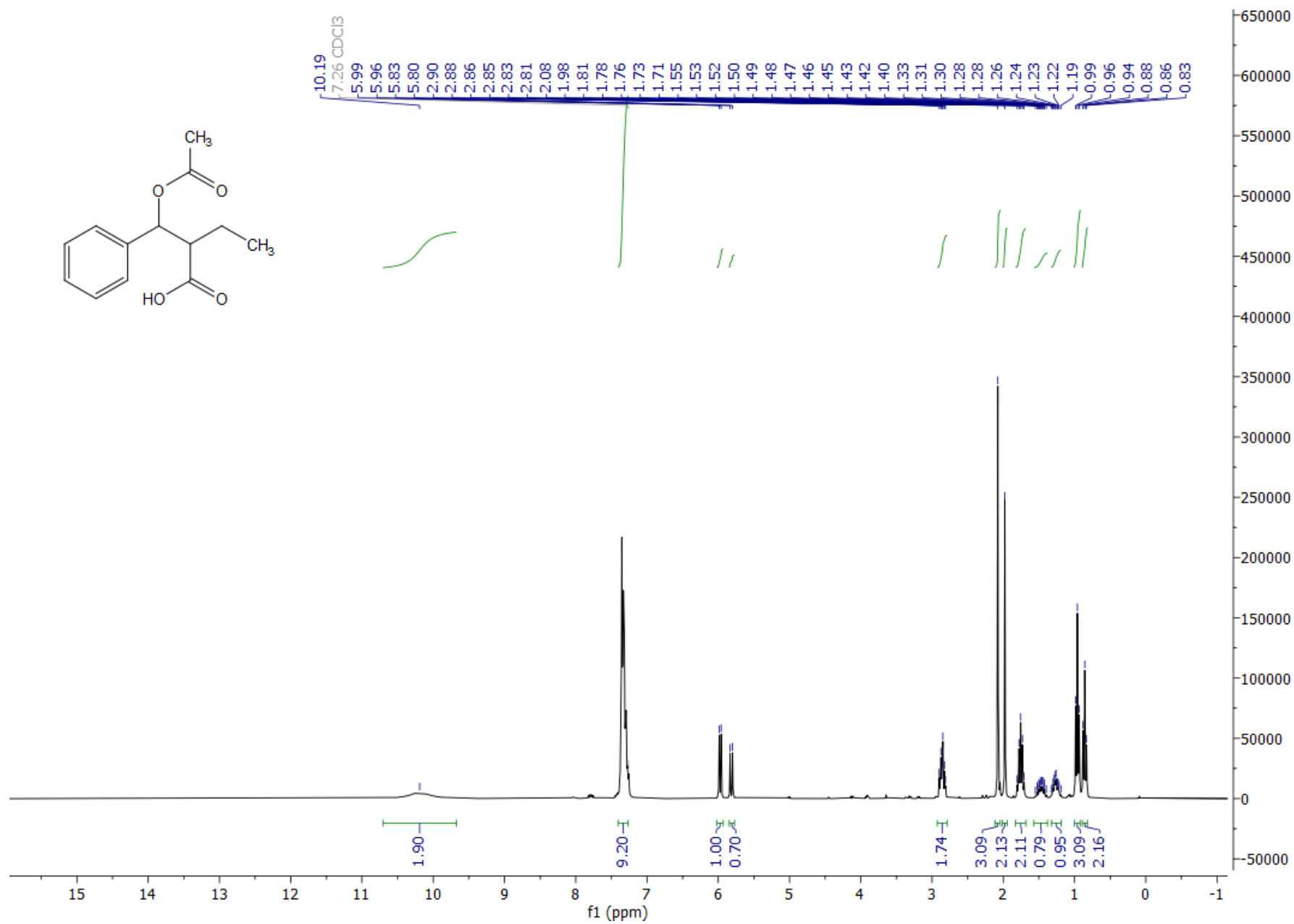
¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-2-methyl-3-phenylpropanoic acid, 2I



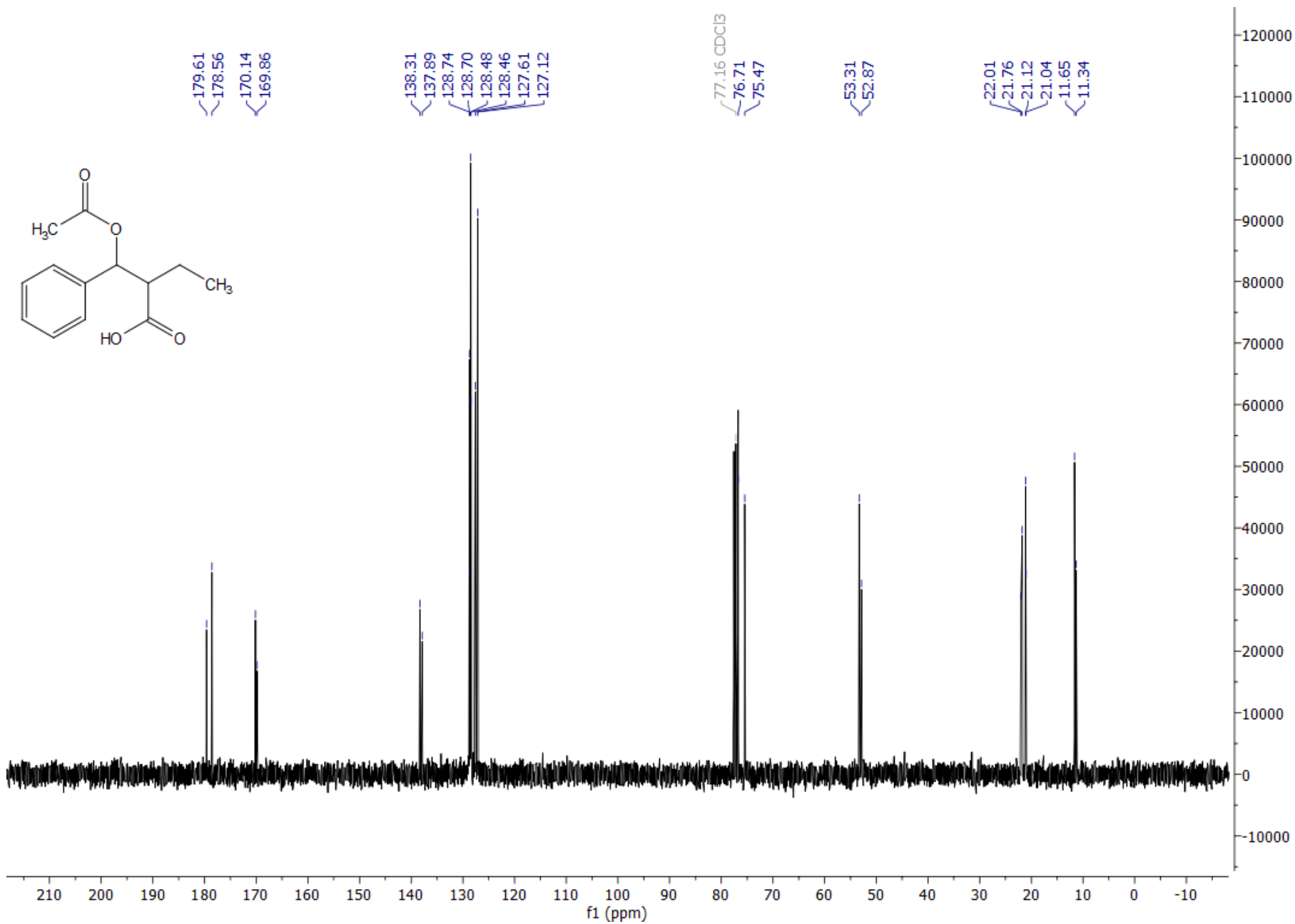
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Acetoxy-2-methyl-3-phenylpropanoic acid, 2l**



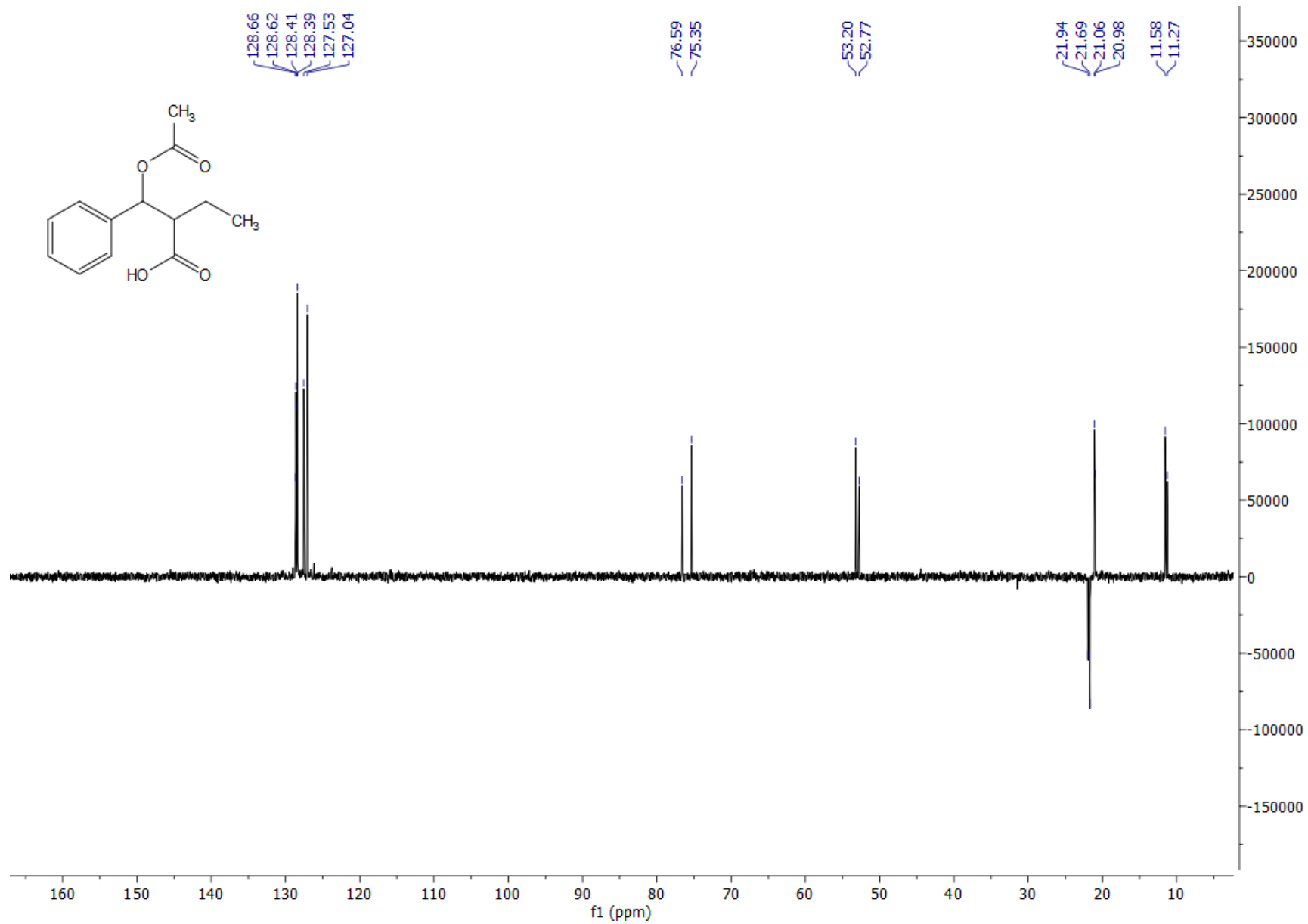
¹H NMR (300.13 MHz, CDCl₃), 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



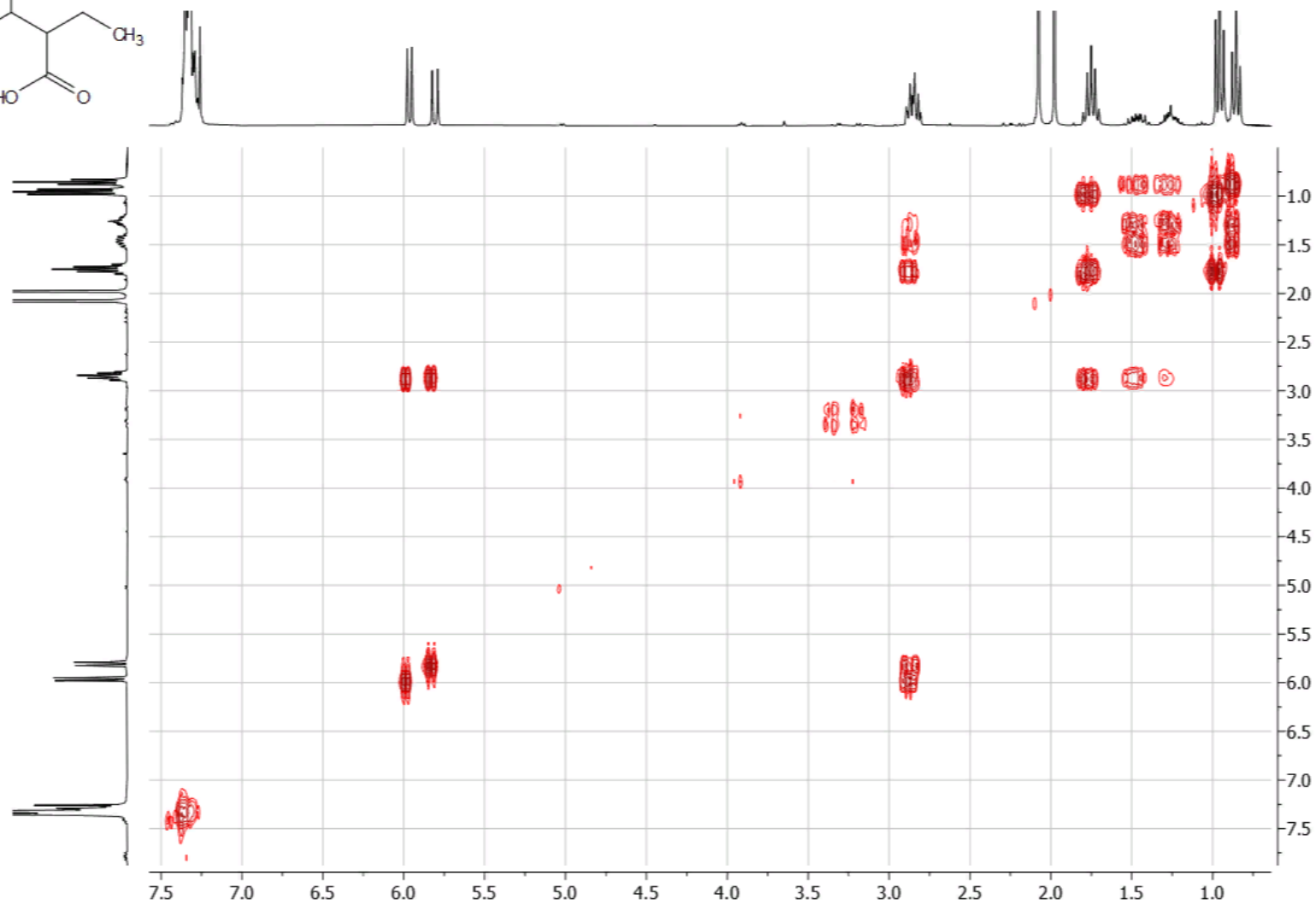
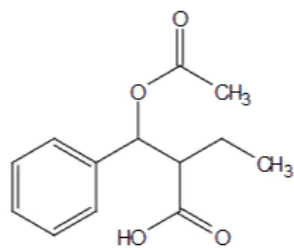
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



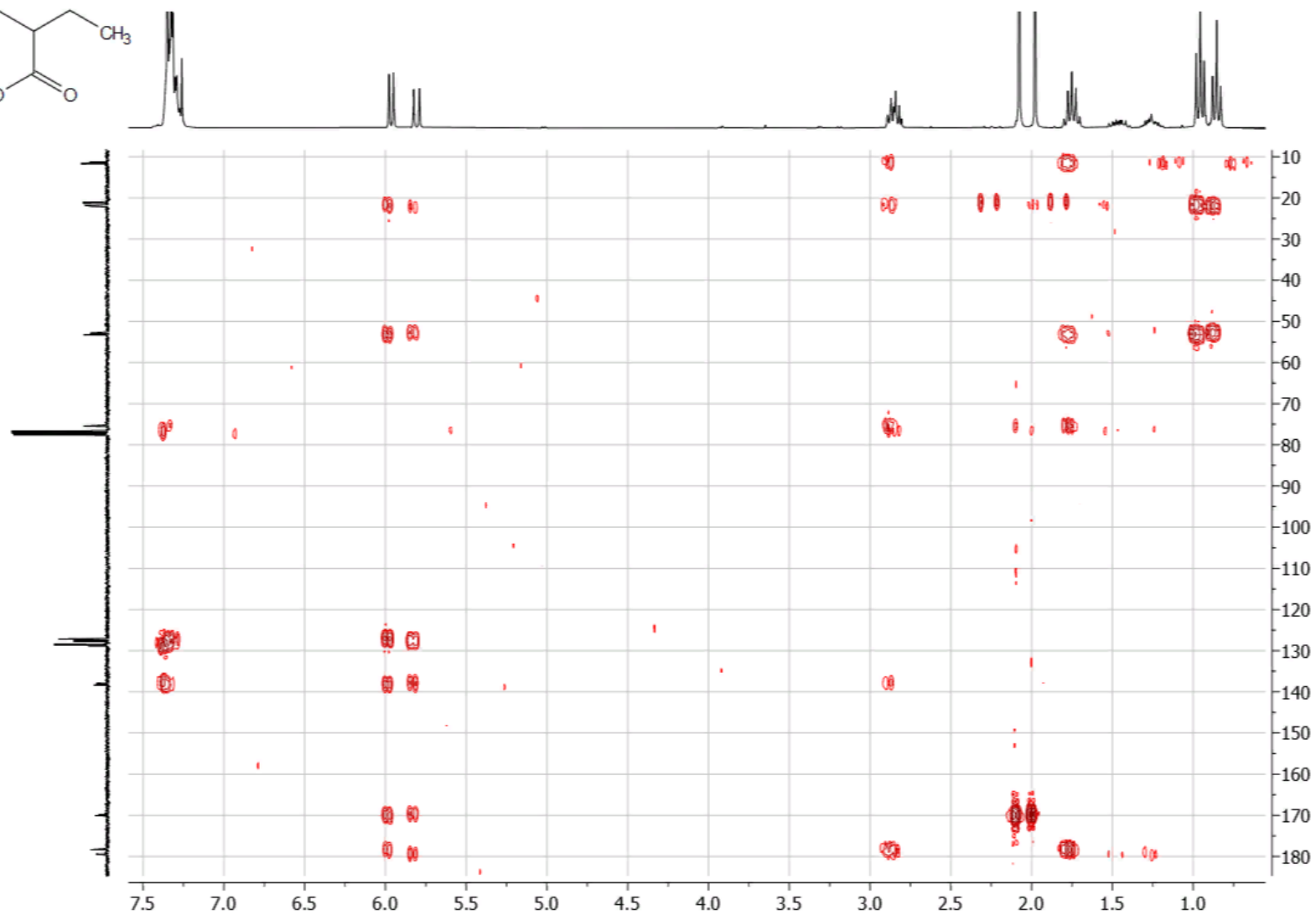
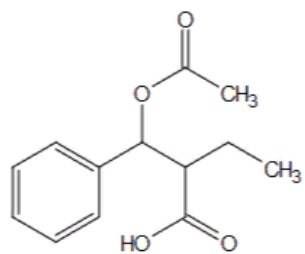
¹³C DEPT135, 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



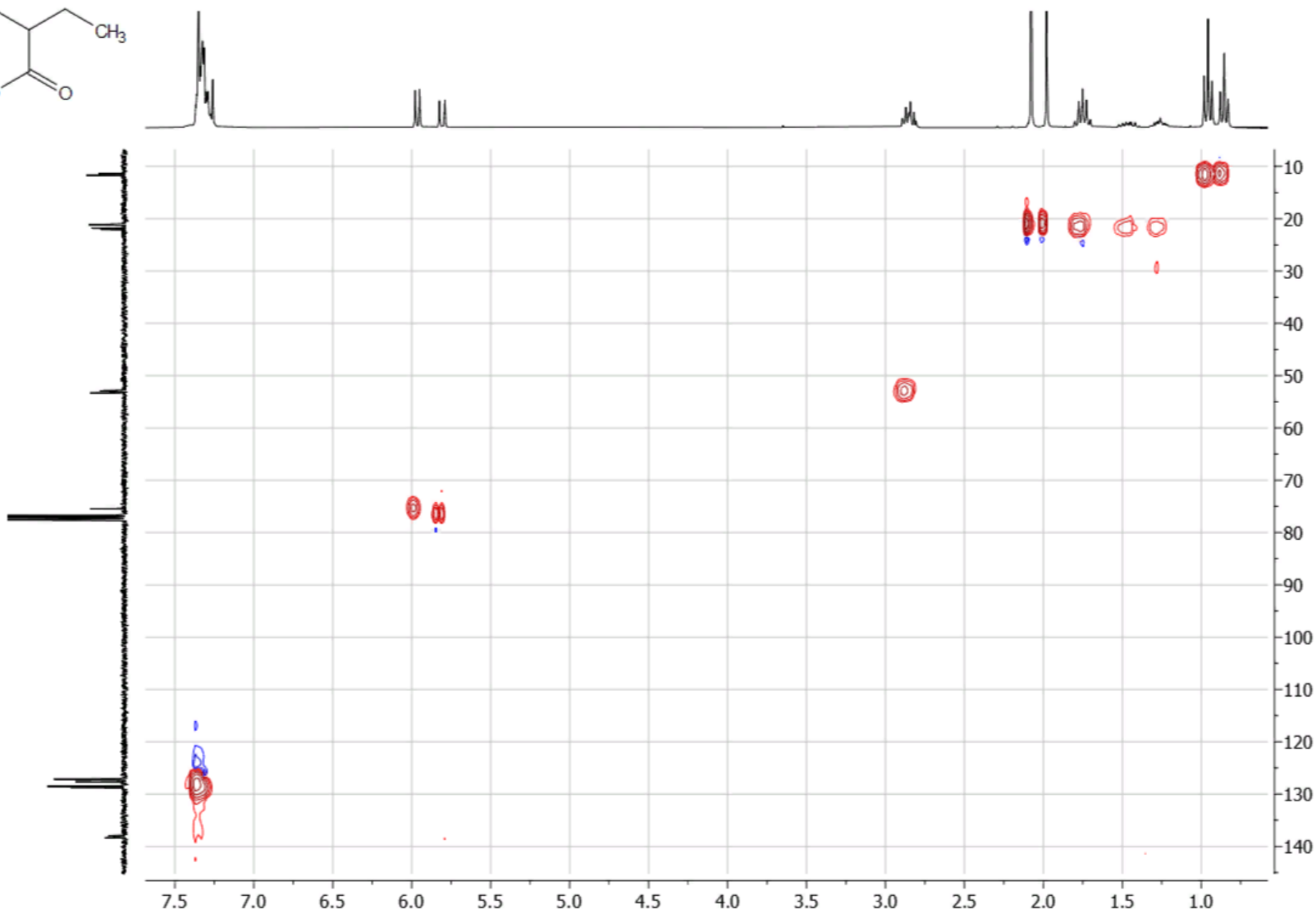
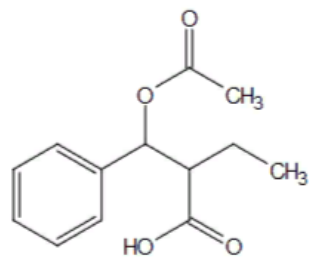
{¹H - ¹H} COSY of 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



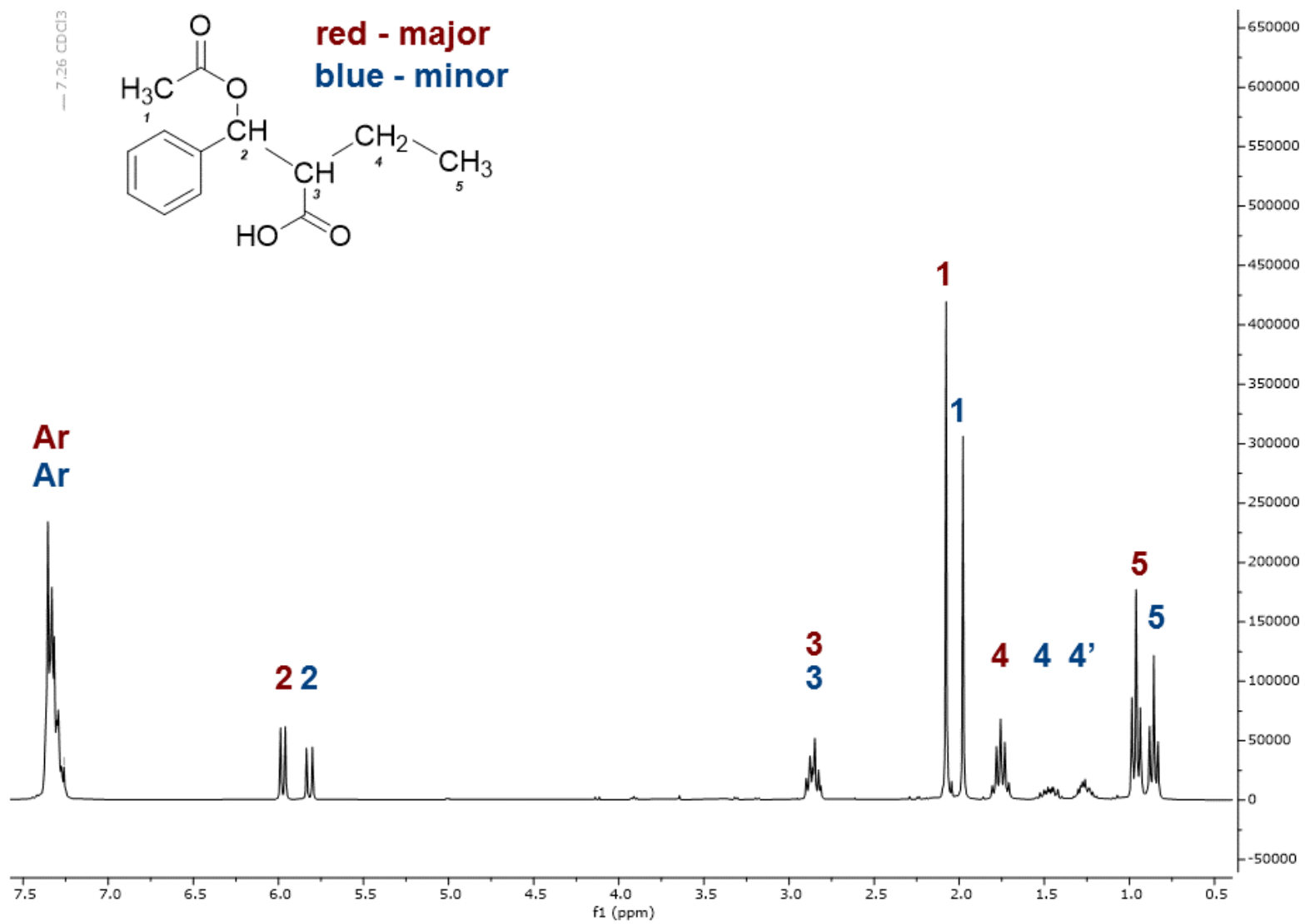
{¹H - ¹³C} HMBC of 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



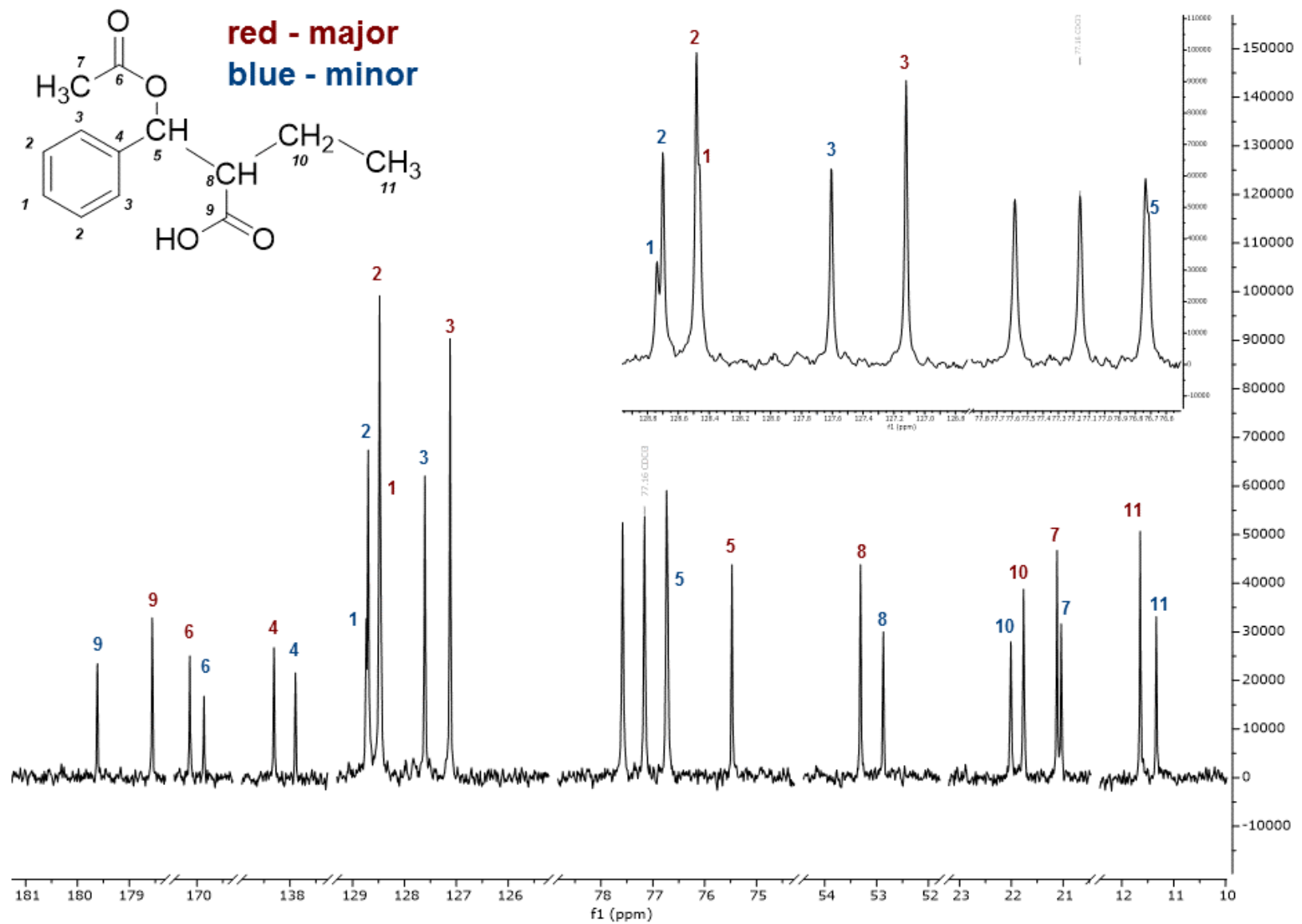
{¹H - ¹³C} HSQC of 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



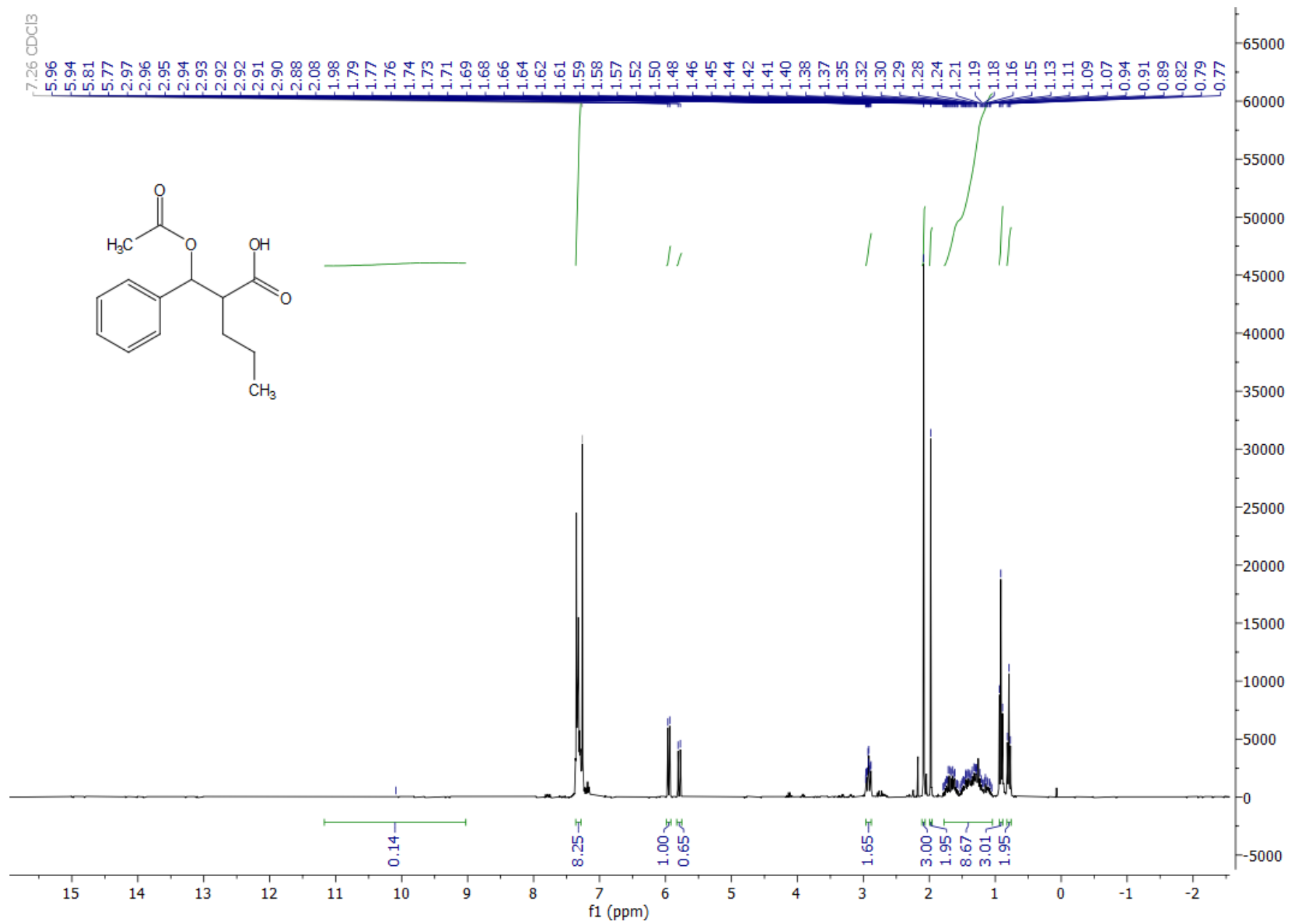
Correlation ^1H NMR (300.13 MHz, CDCl_3), 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



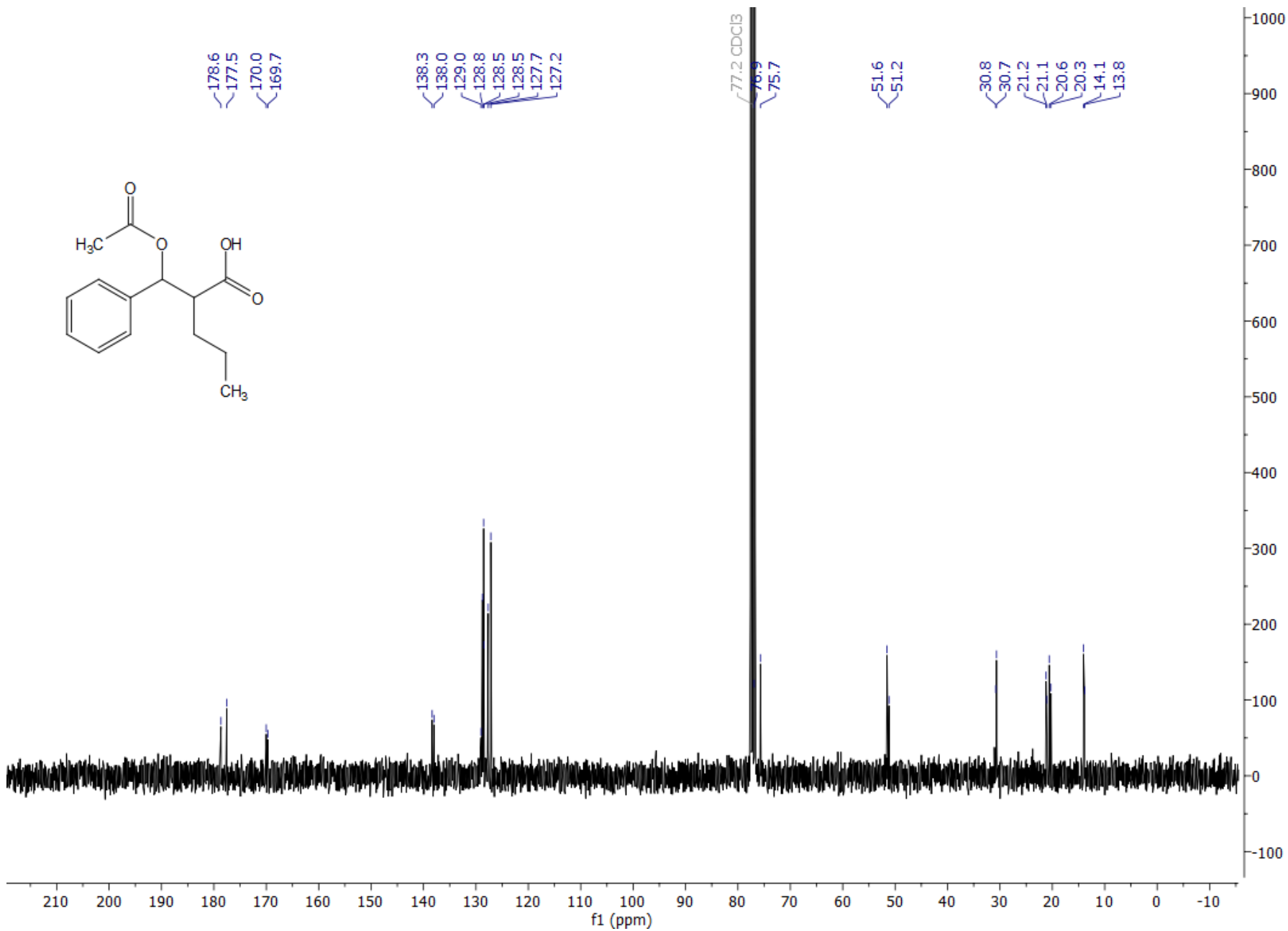
Correlation $^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



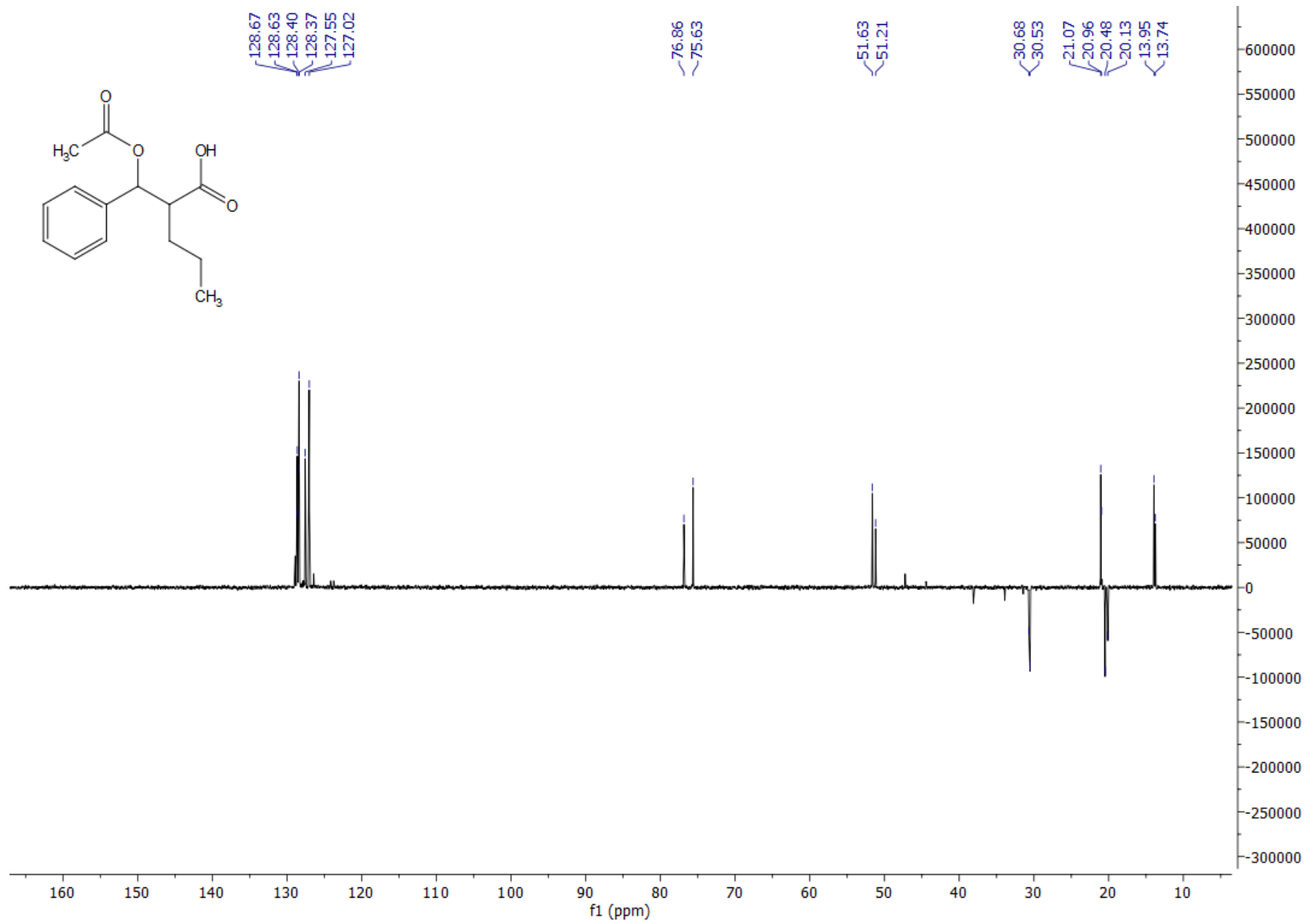
¹H NMR (300.13 MHz, CDCl₃), 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n



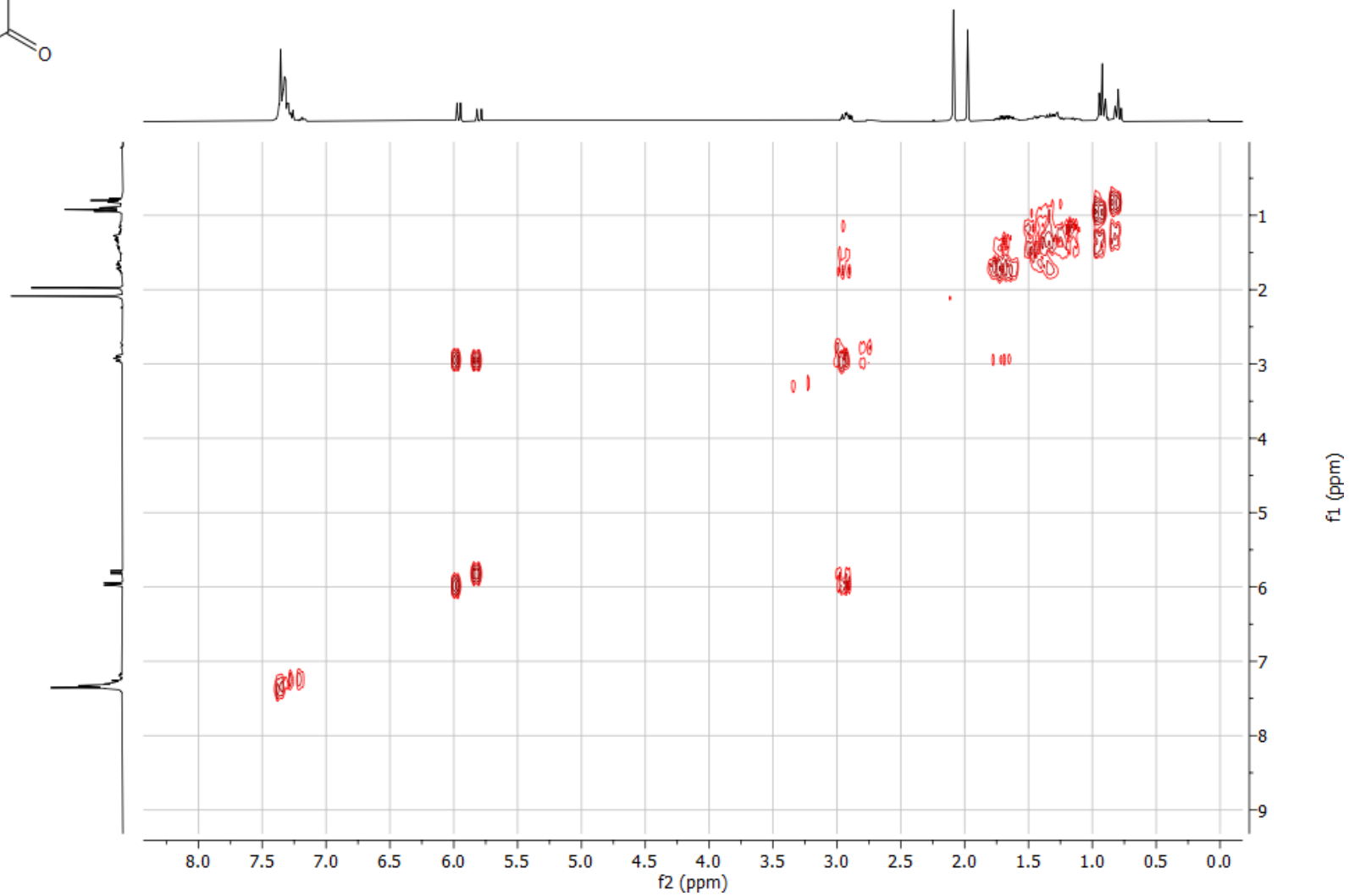
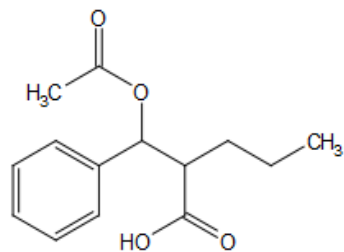
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n



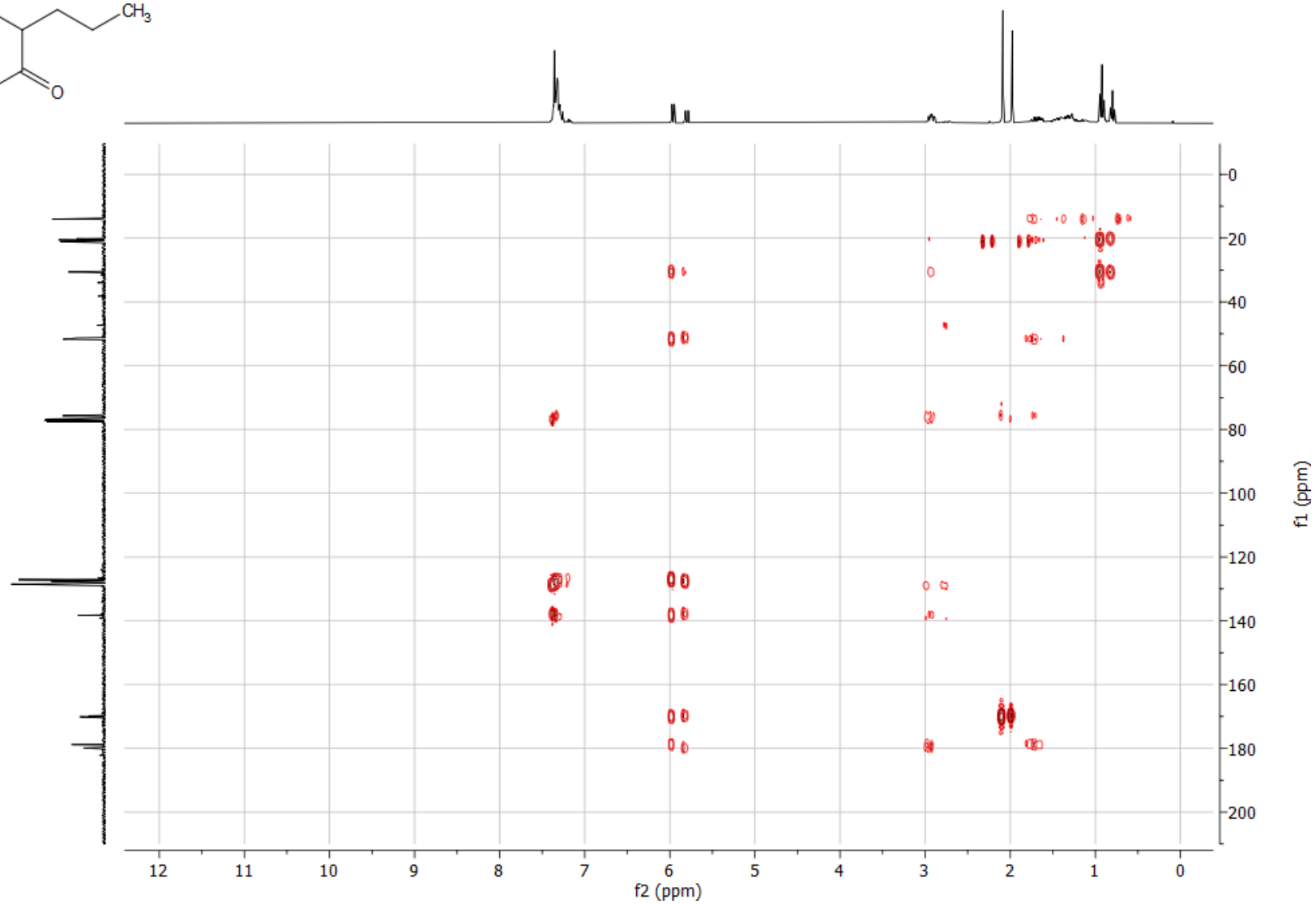
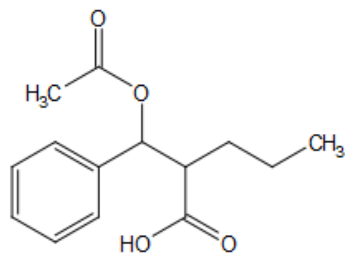
¹³C DEPT135, 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n



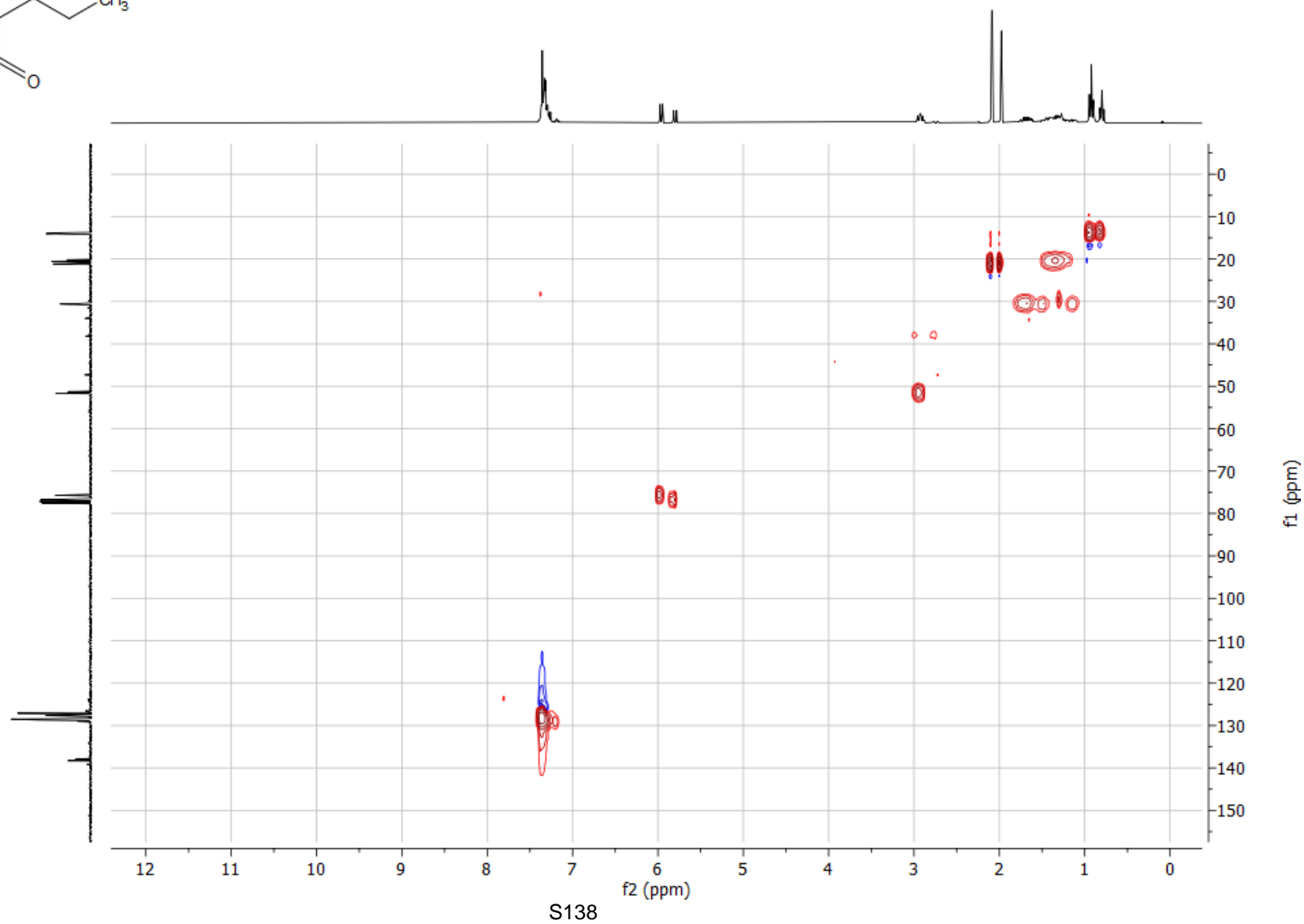
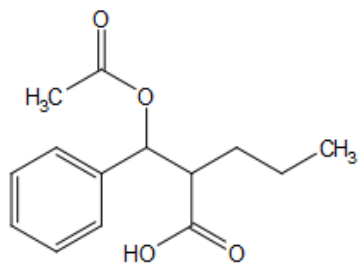
{¹H - ¹H} COSY of 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n



{¹H - ¹³C} HMBC of 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n

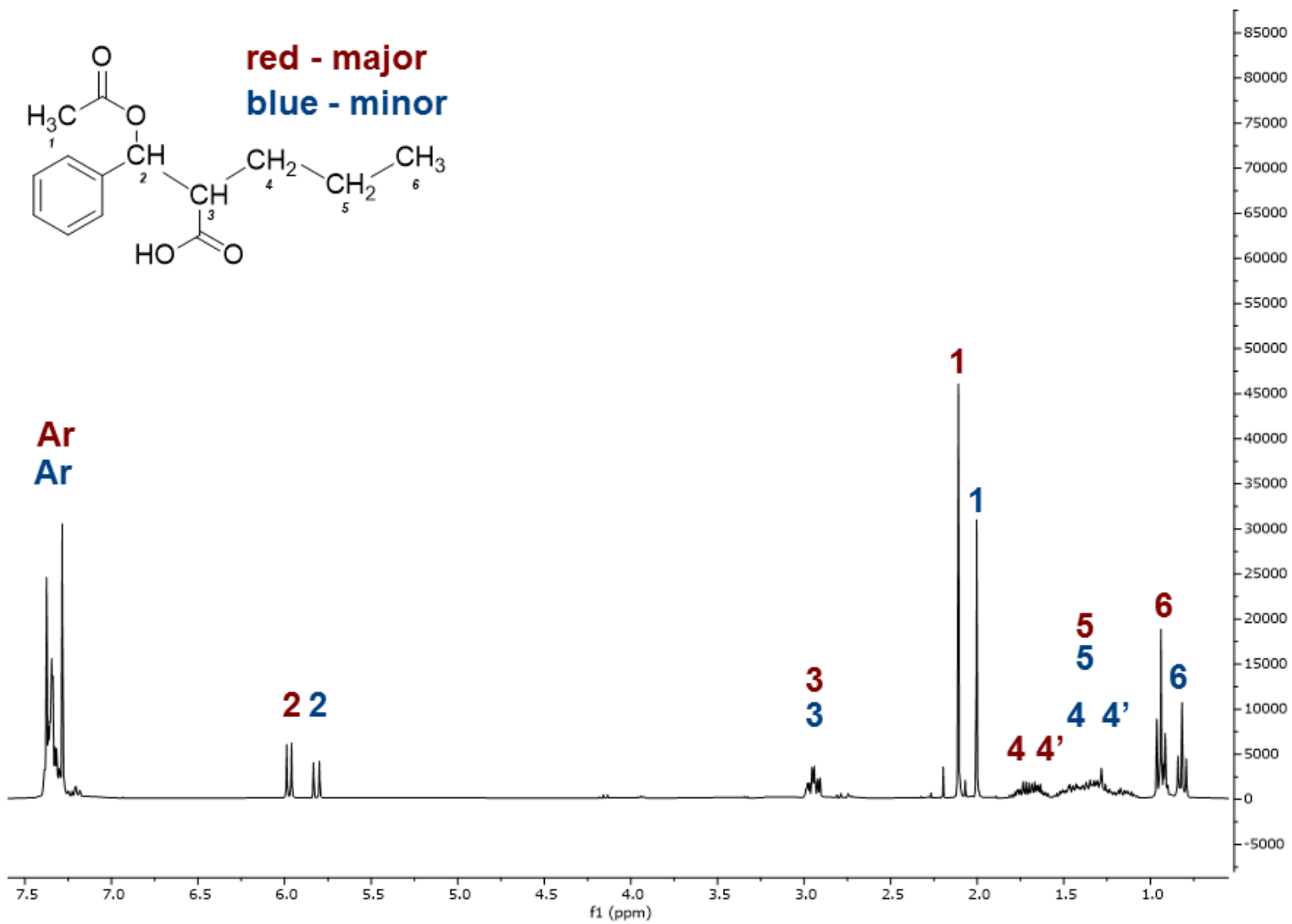


{¹H - ¹³C} HSQC of 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n

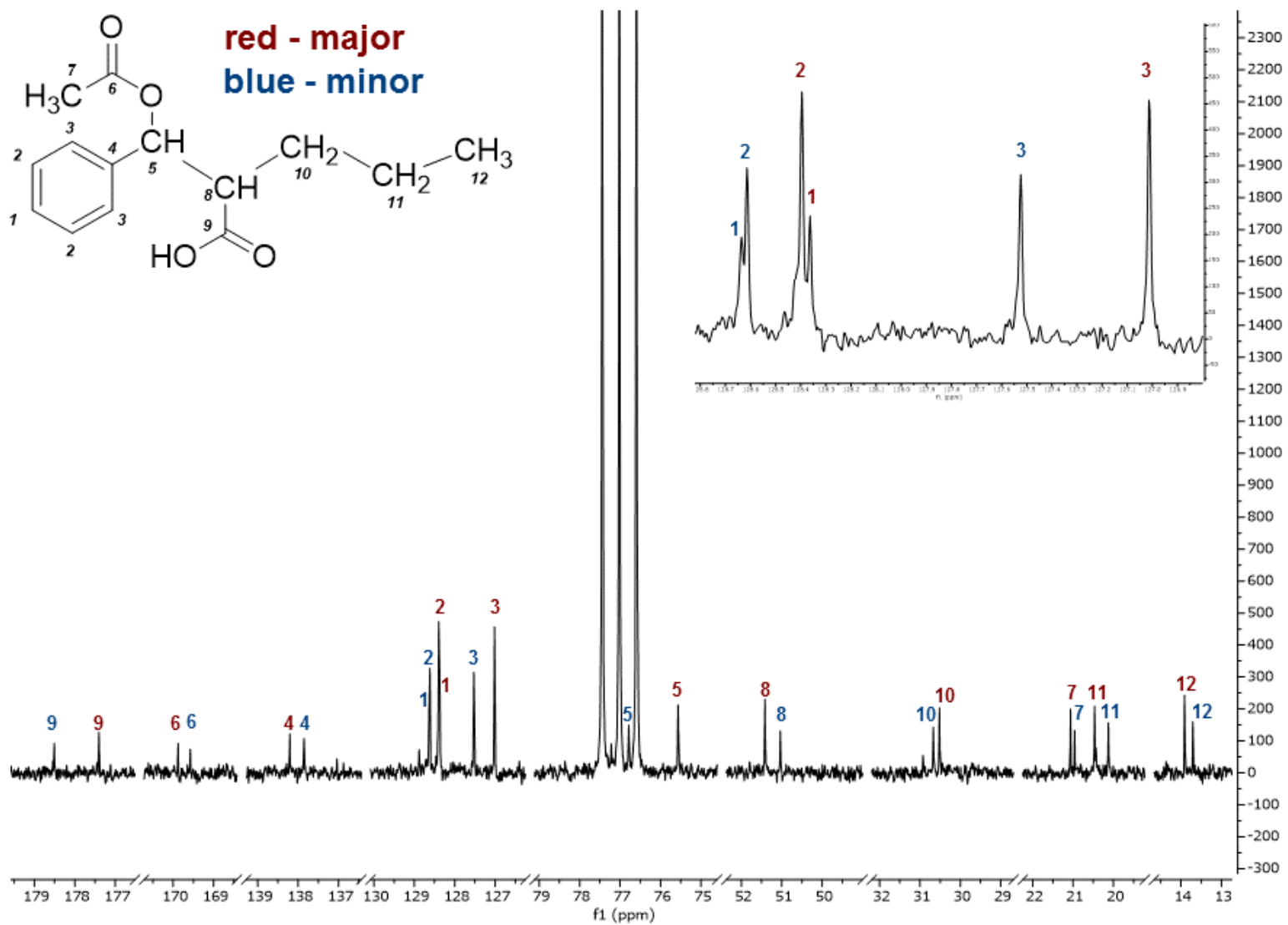


S138

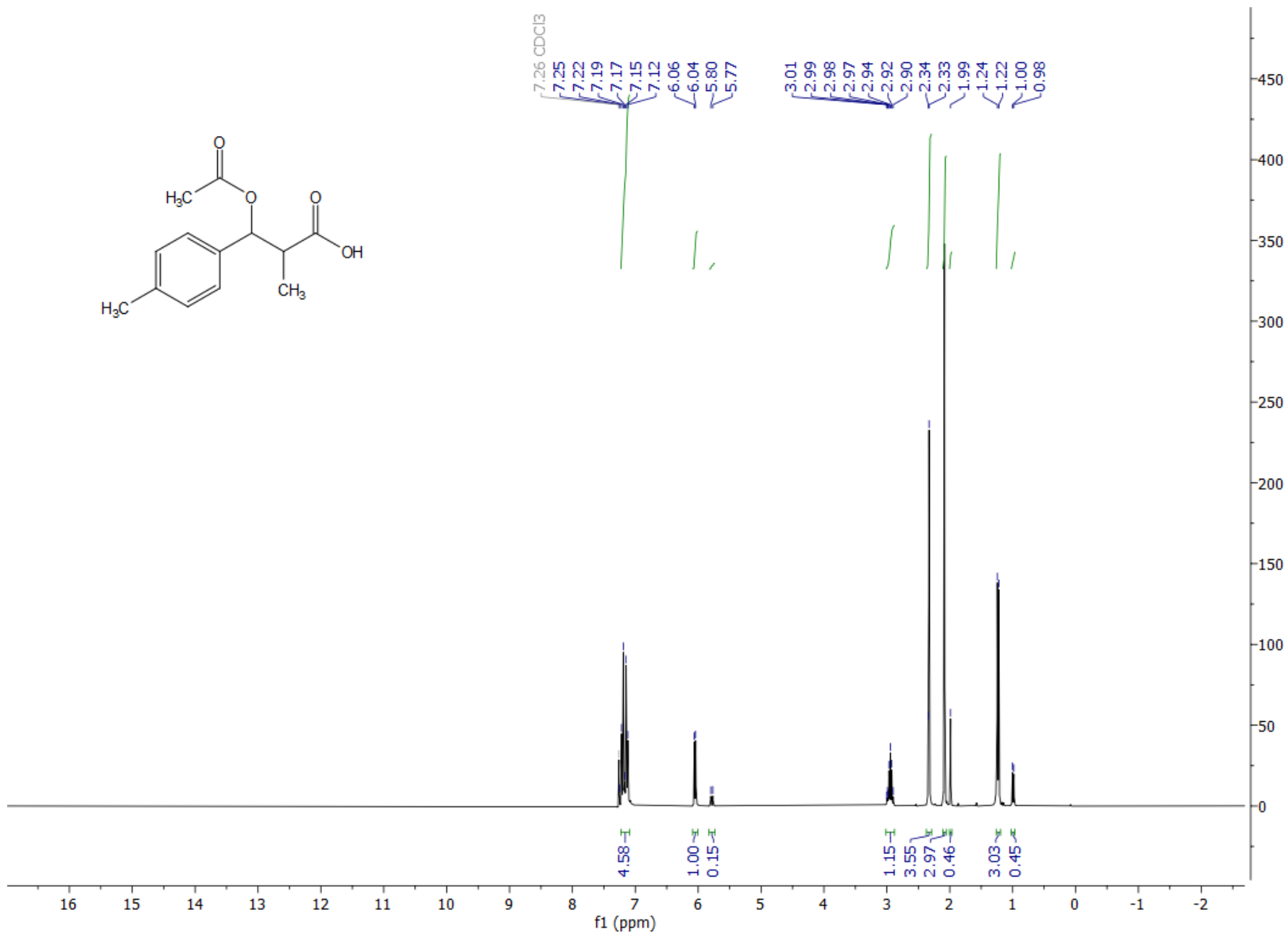
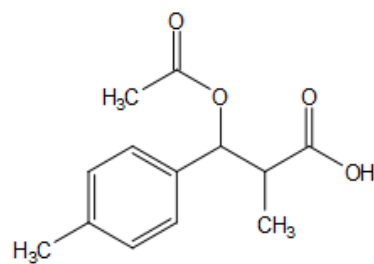
Correlation ^1H NMR (300.13 MHz, CDCl_3), 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n



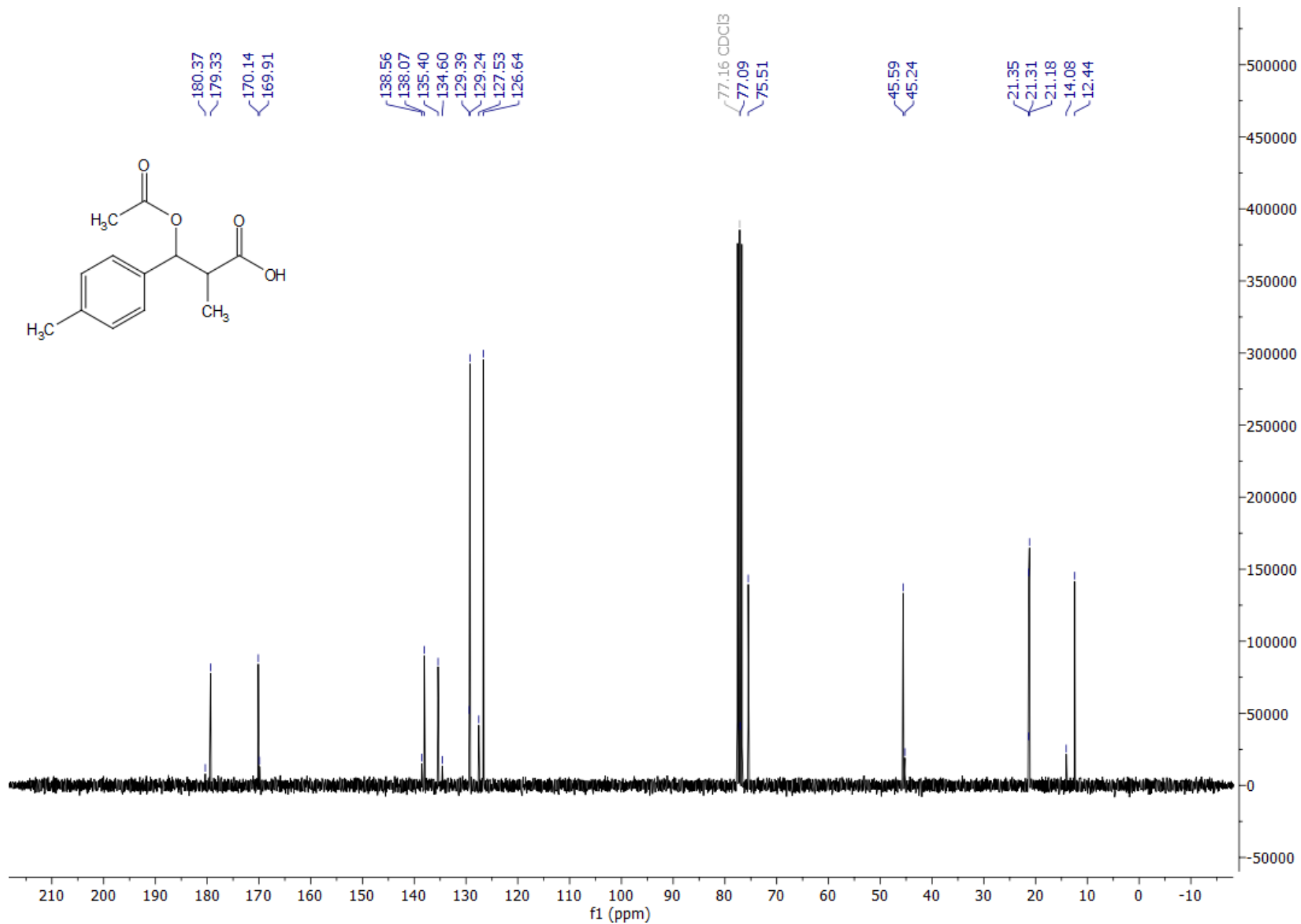
Correlation $^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n**



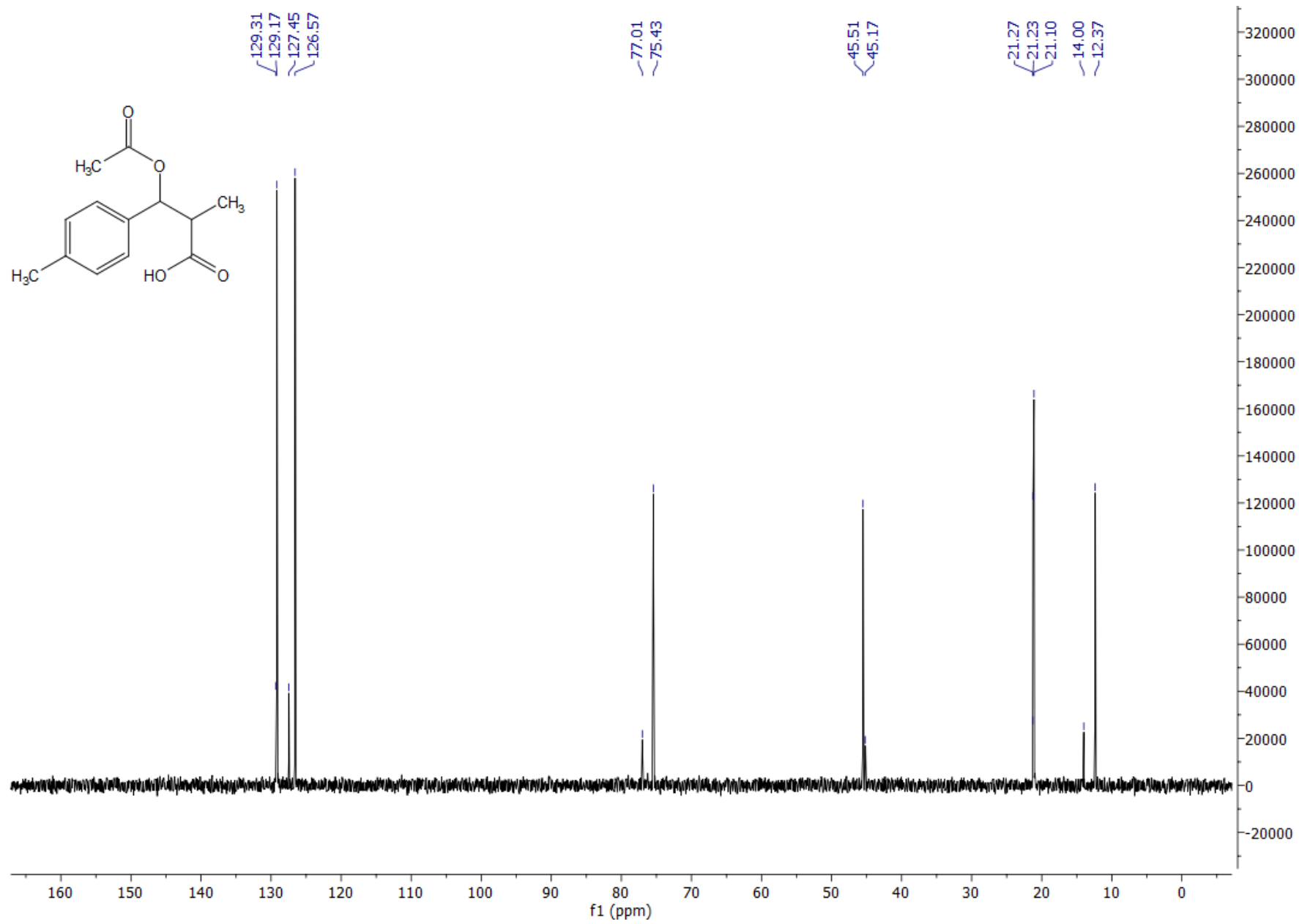
¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o



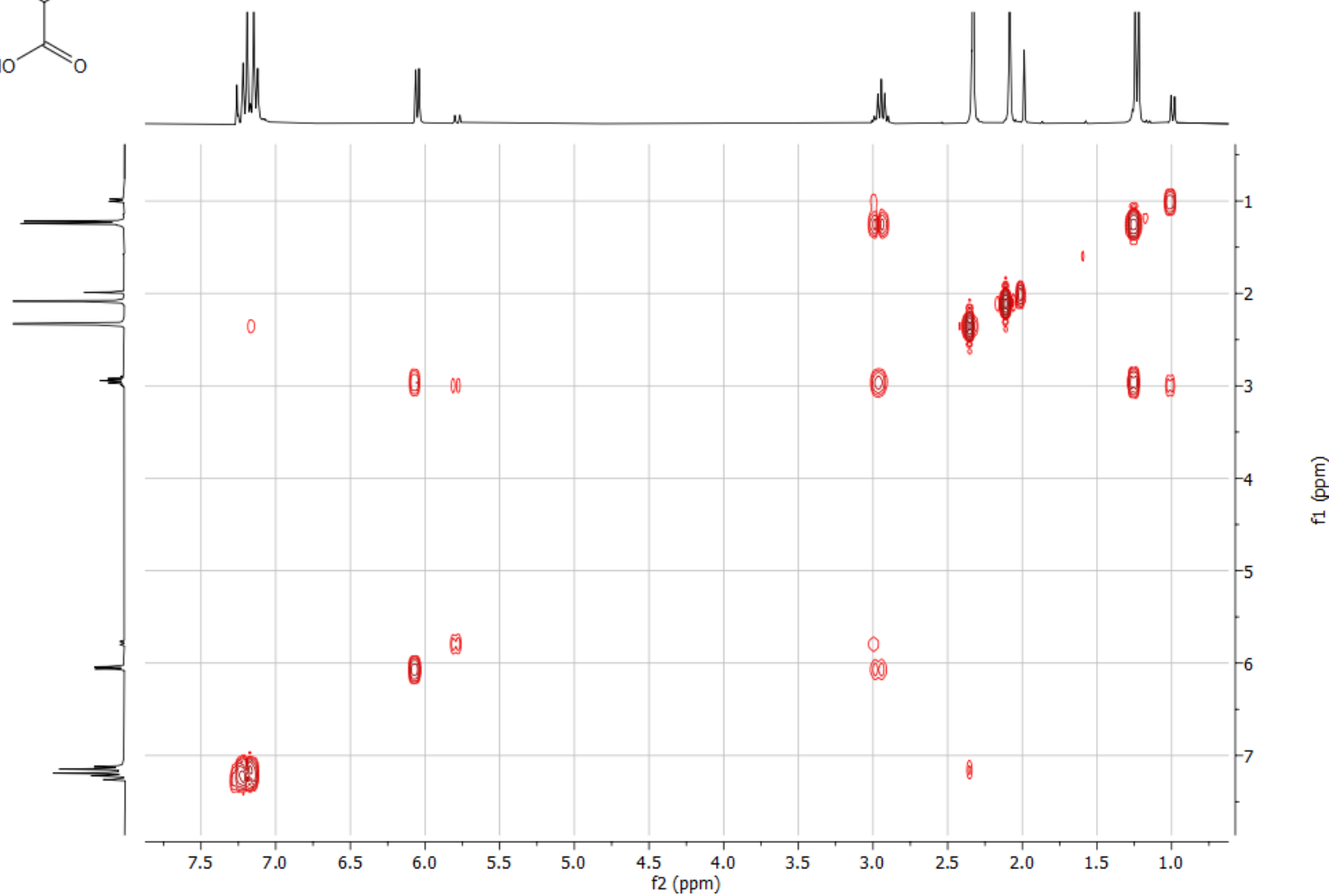
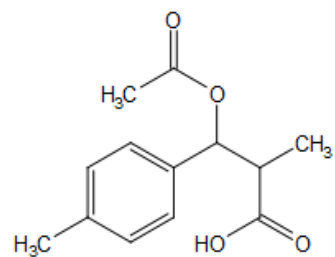
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o**



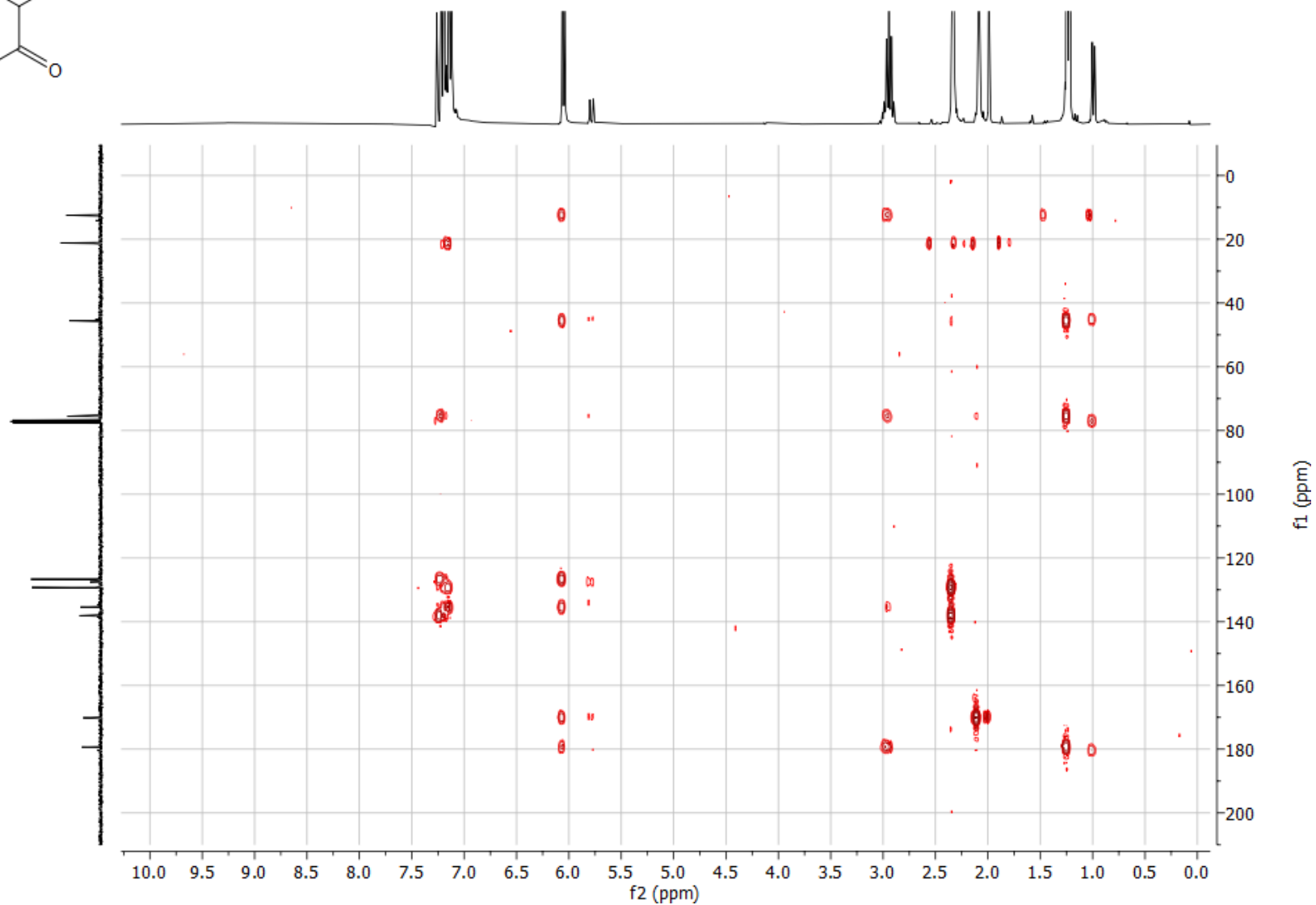
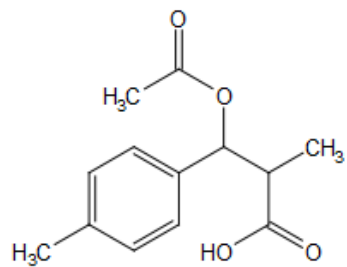
¹³C DEPT135, 3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o



{¹H - ¹H} COSY of 3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o

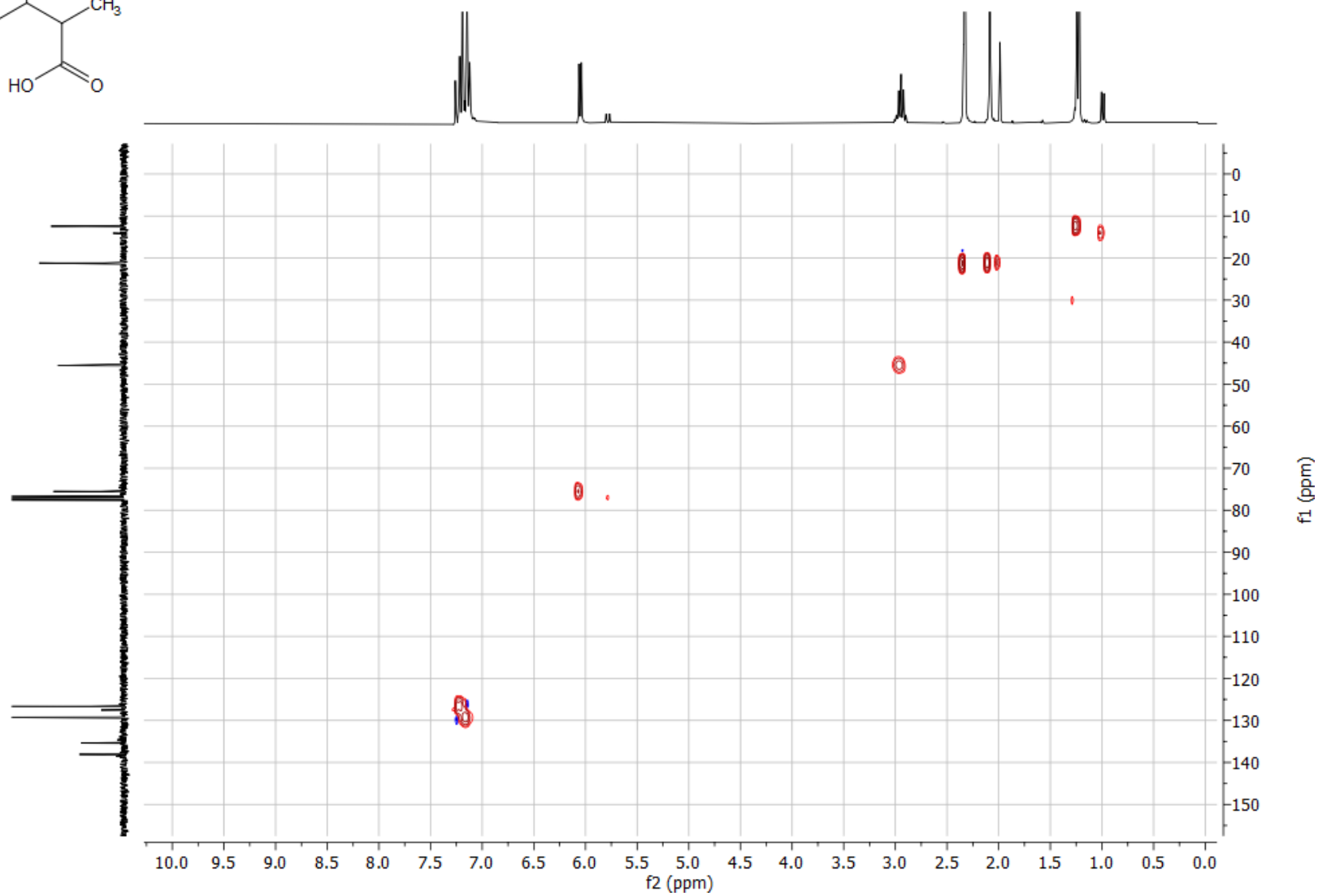
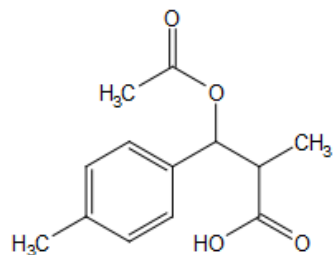


{¹H - ¹³C} HMBC of 3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o

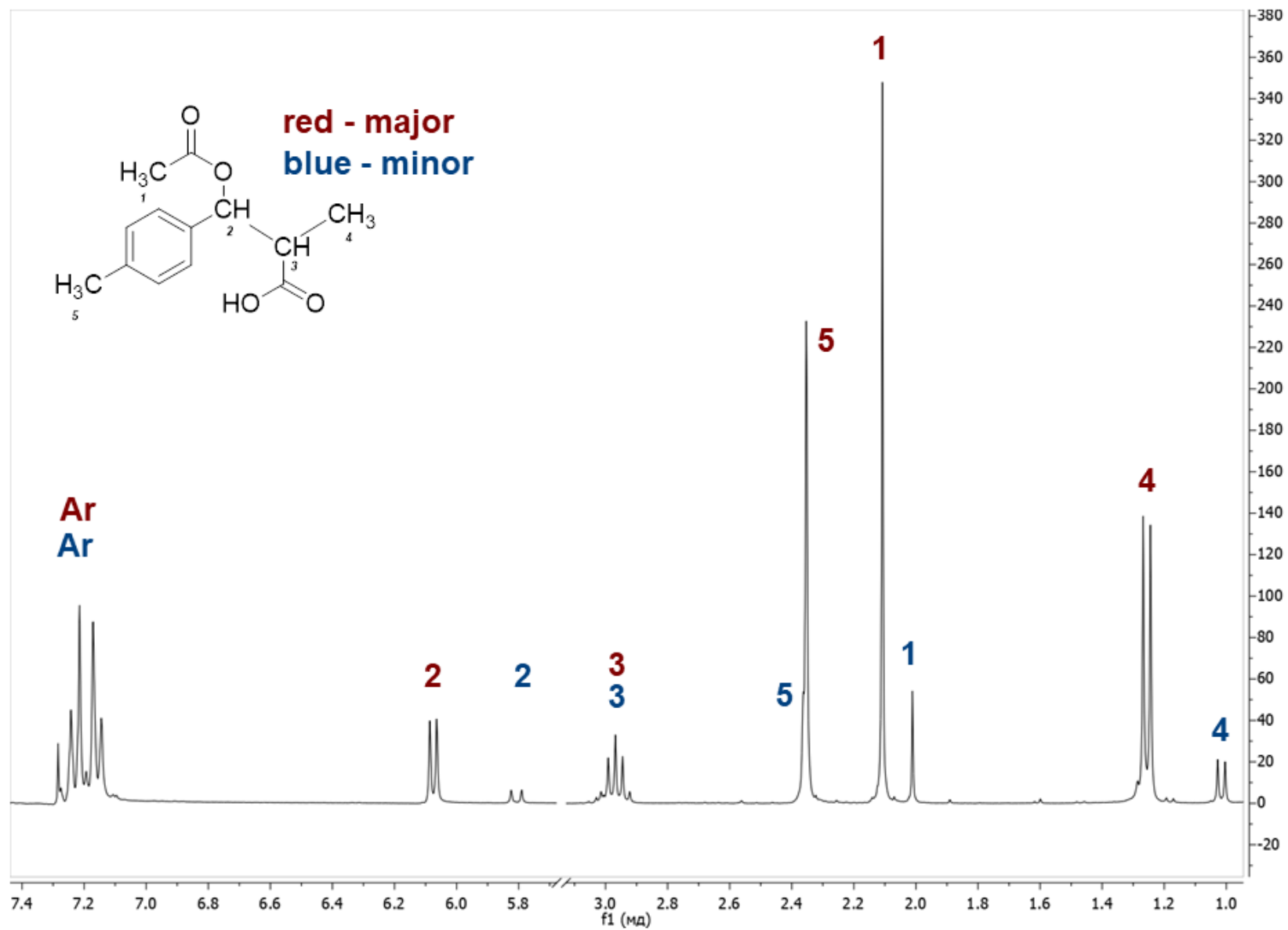


S145

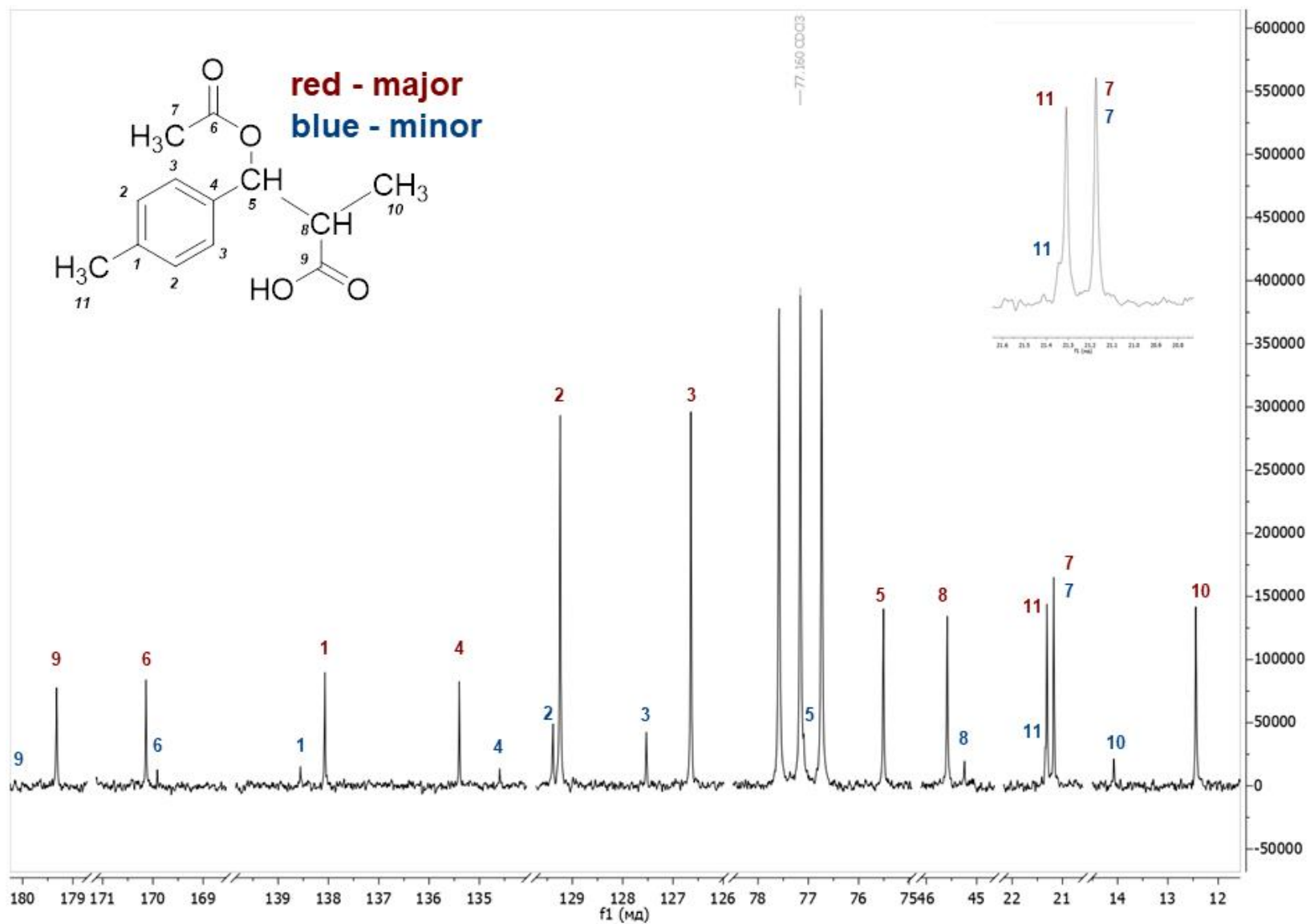
{¹H - ¹³C} HSQC of 3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o



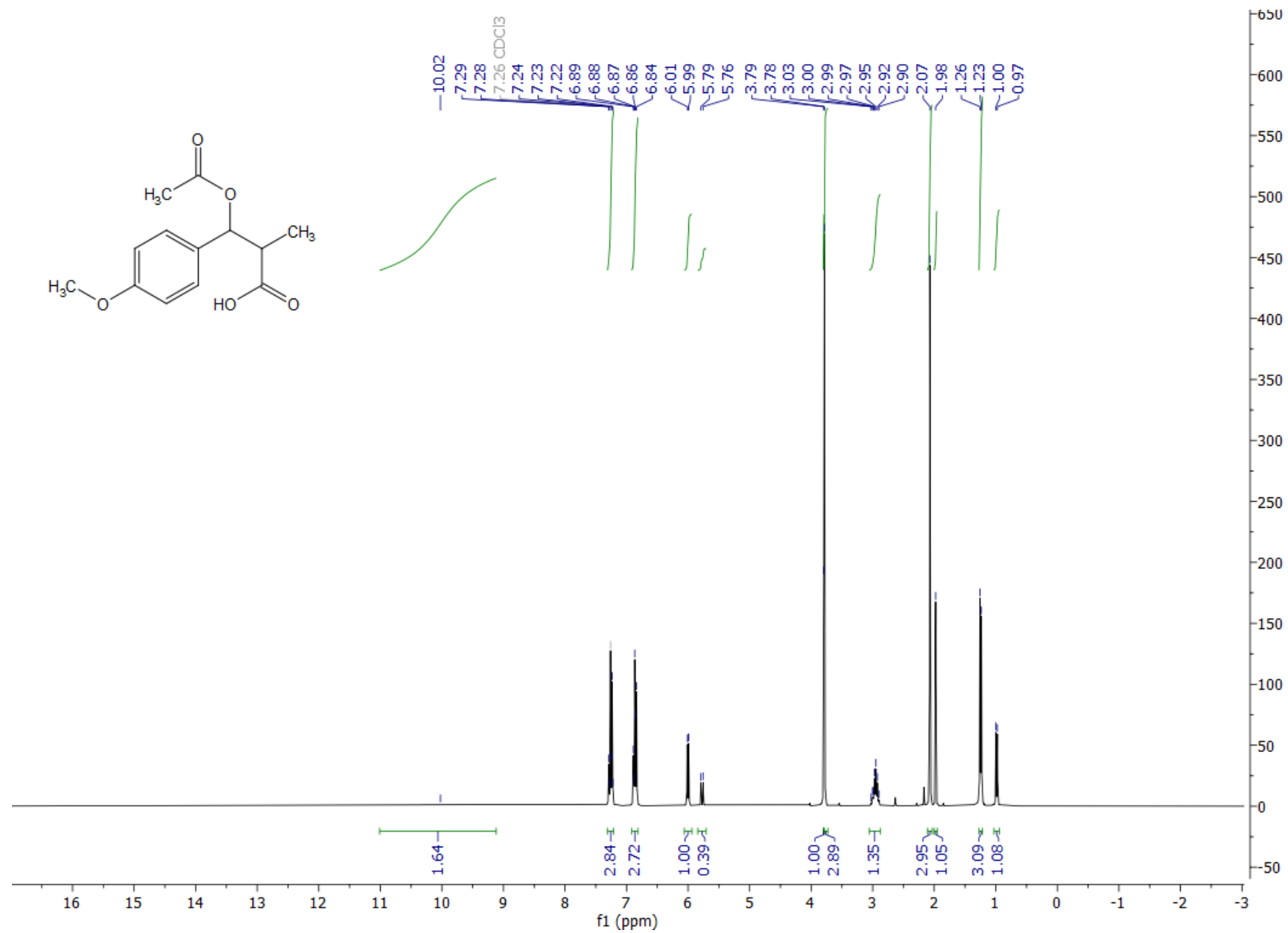
Correlation ^1H NMR (300.13 MHz, CDCl_3), **3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o**



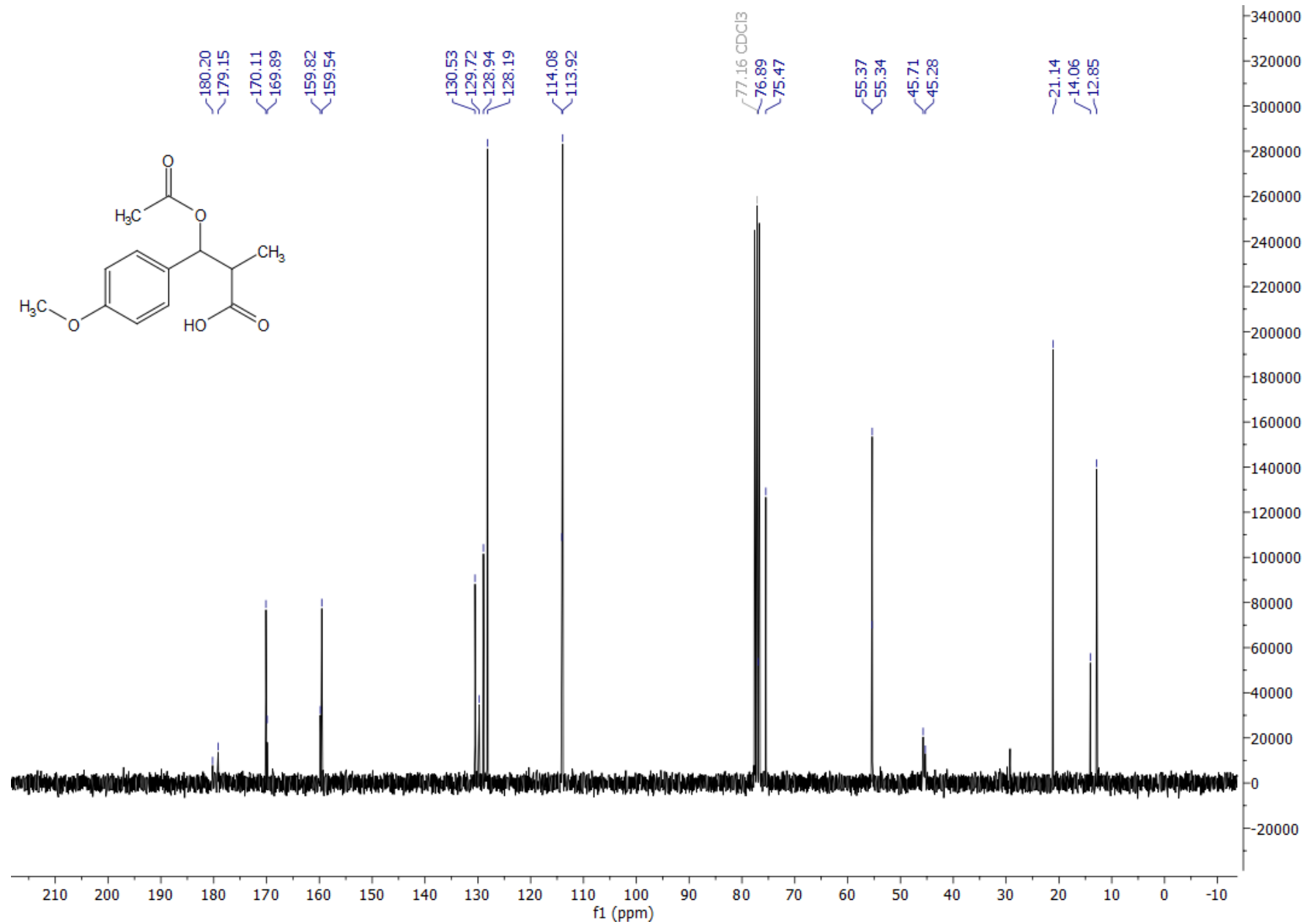
Correlation $^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), 3-Acetoxy-2-methyl-3-(*p*-tolyl)propanoic acid, 2o



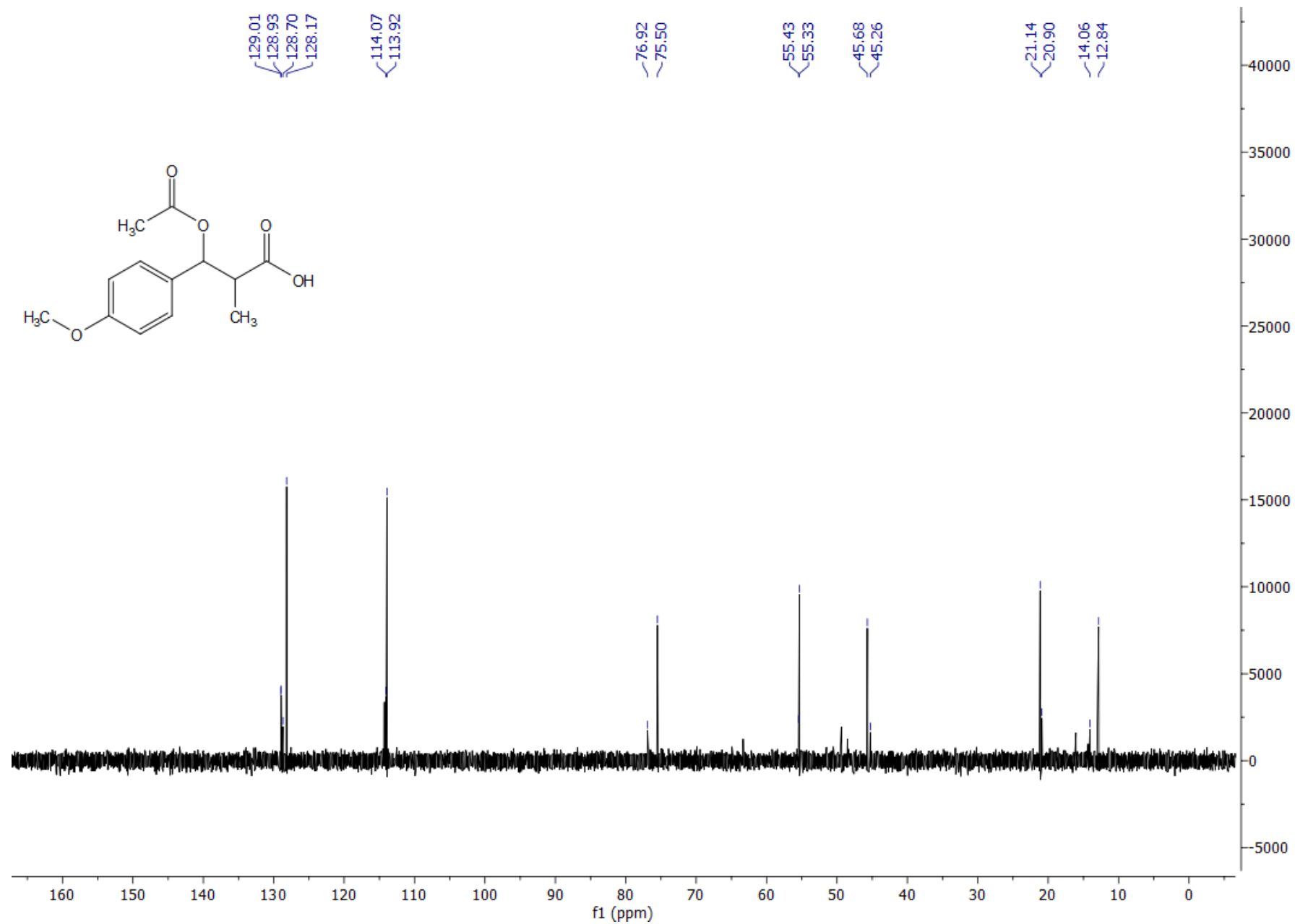
¹H NMR (300.13 MHz, CDCl₃), 3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p



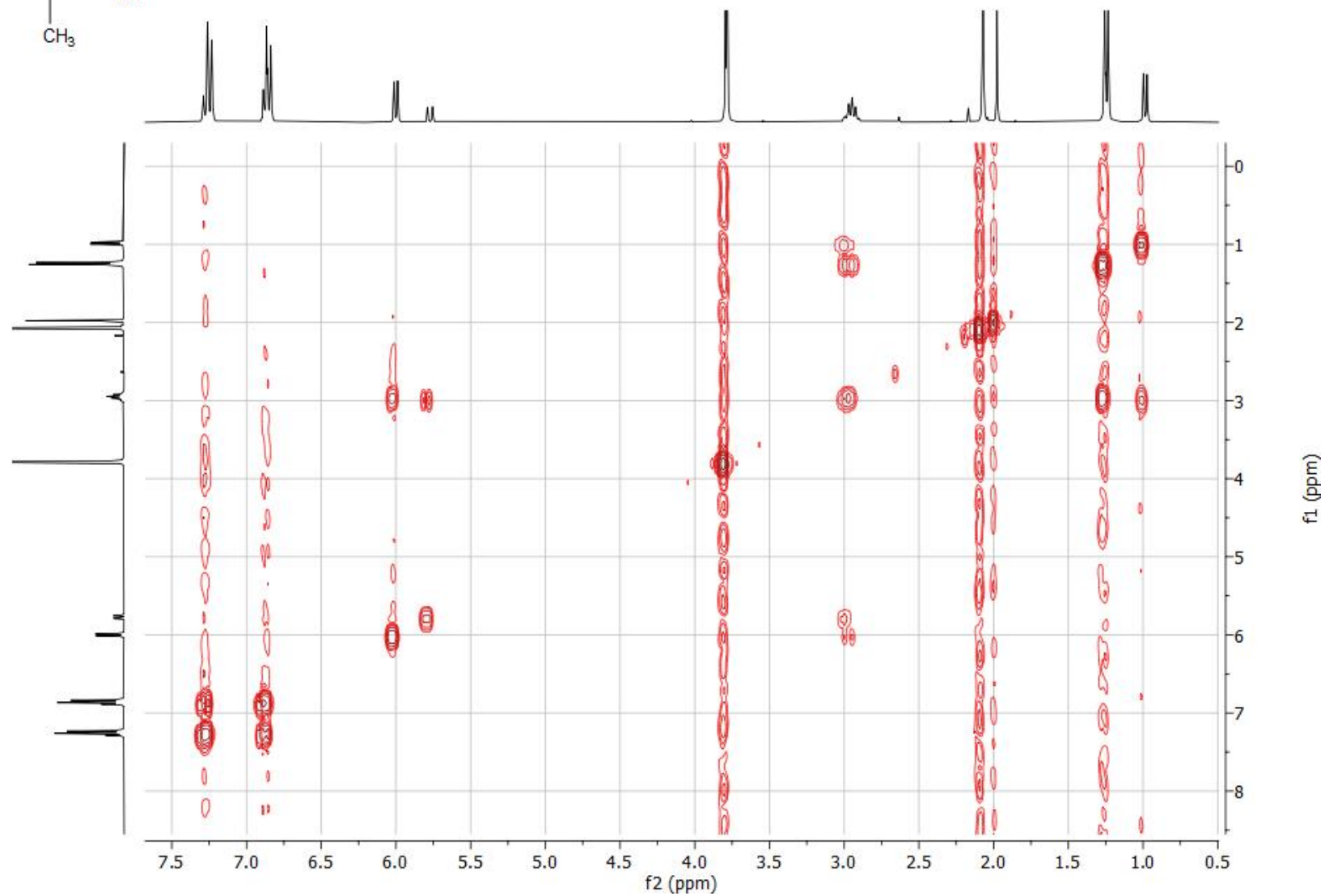
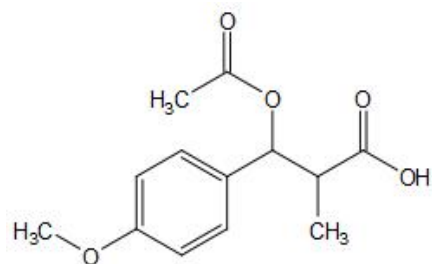
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p**



¹³C DEPT135, 3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p

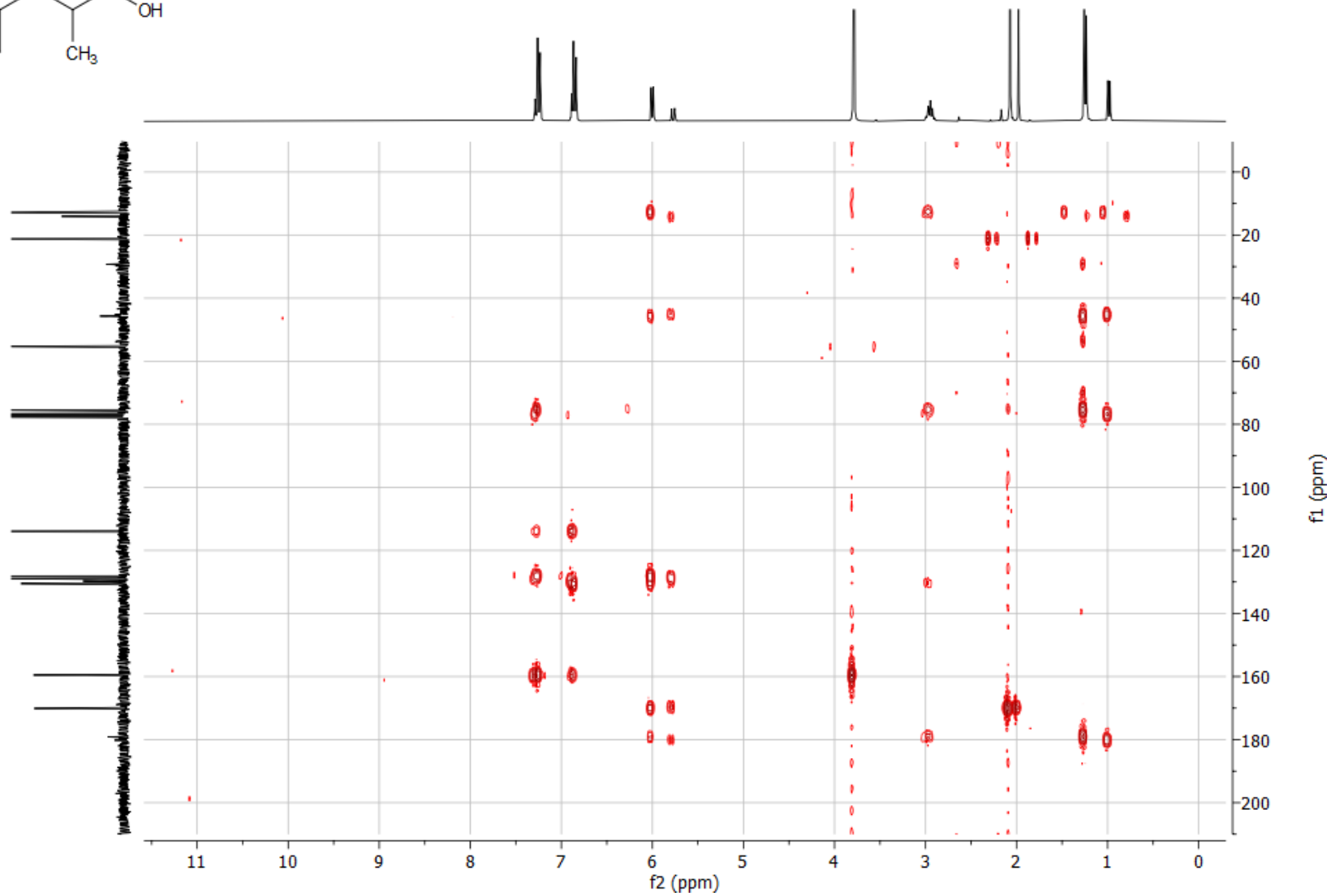
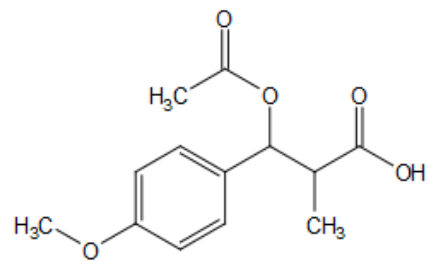


$\{^1\text{H} - ^1\text{H}\}$ COSY of 3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p

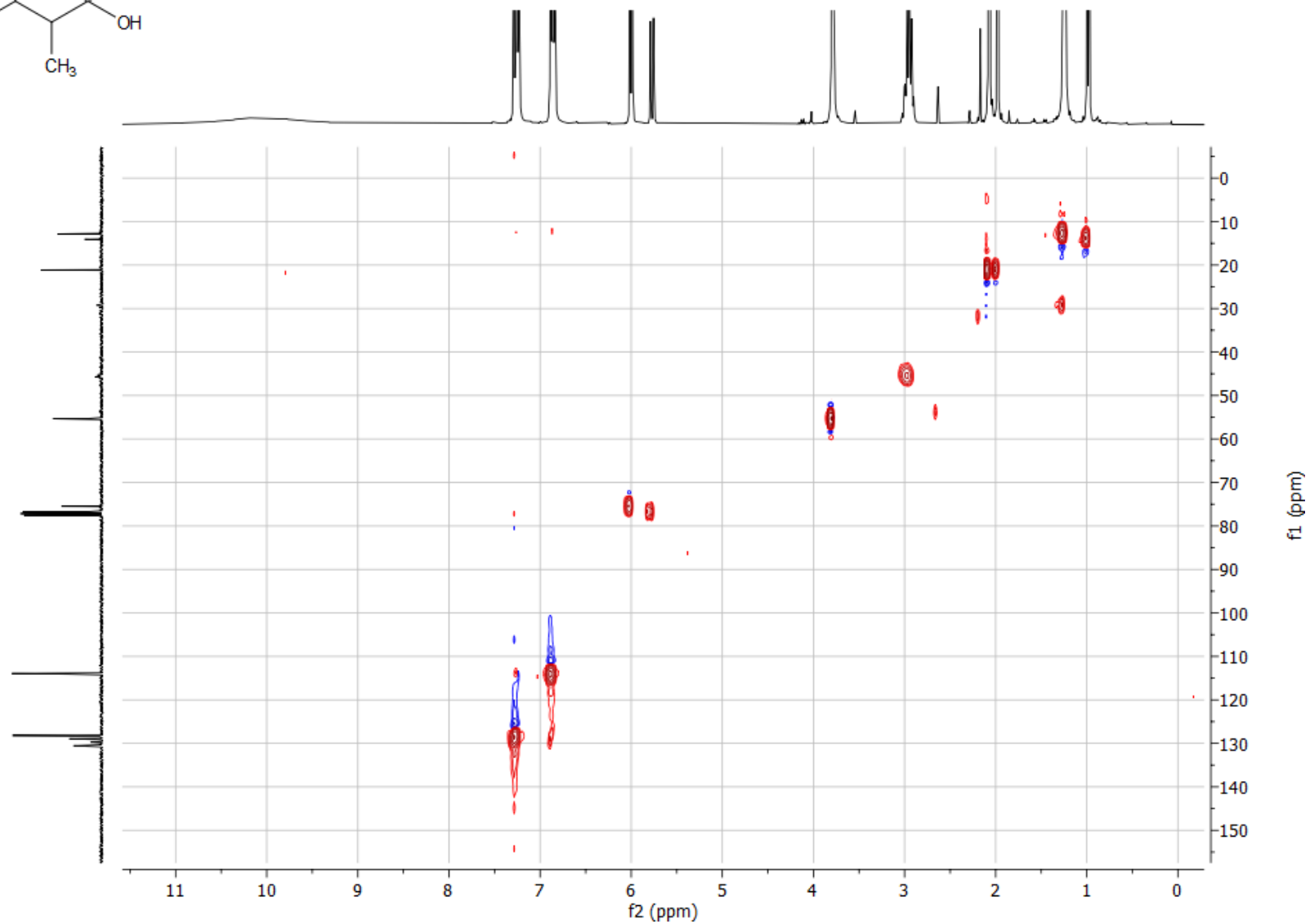
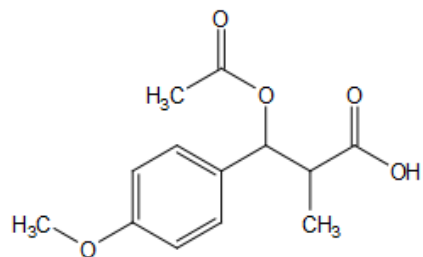


S152

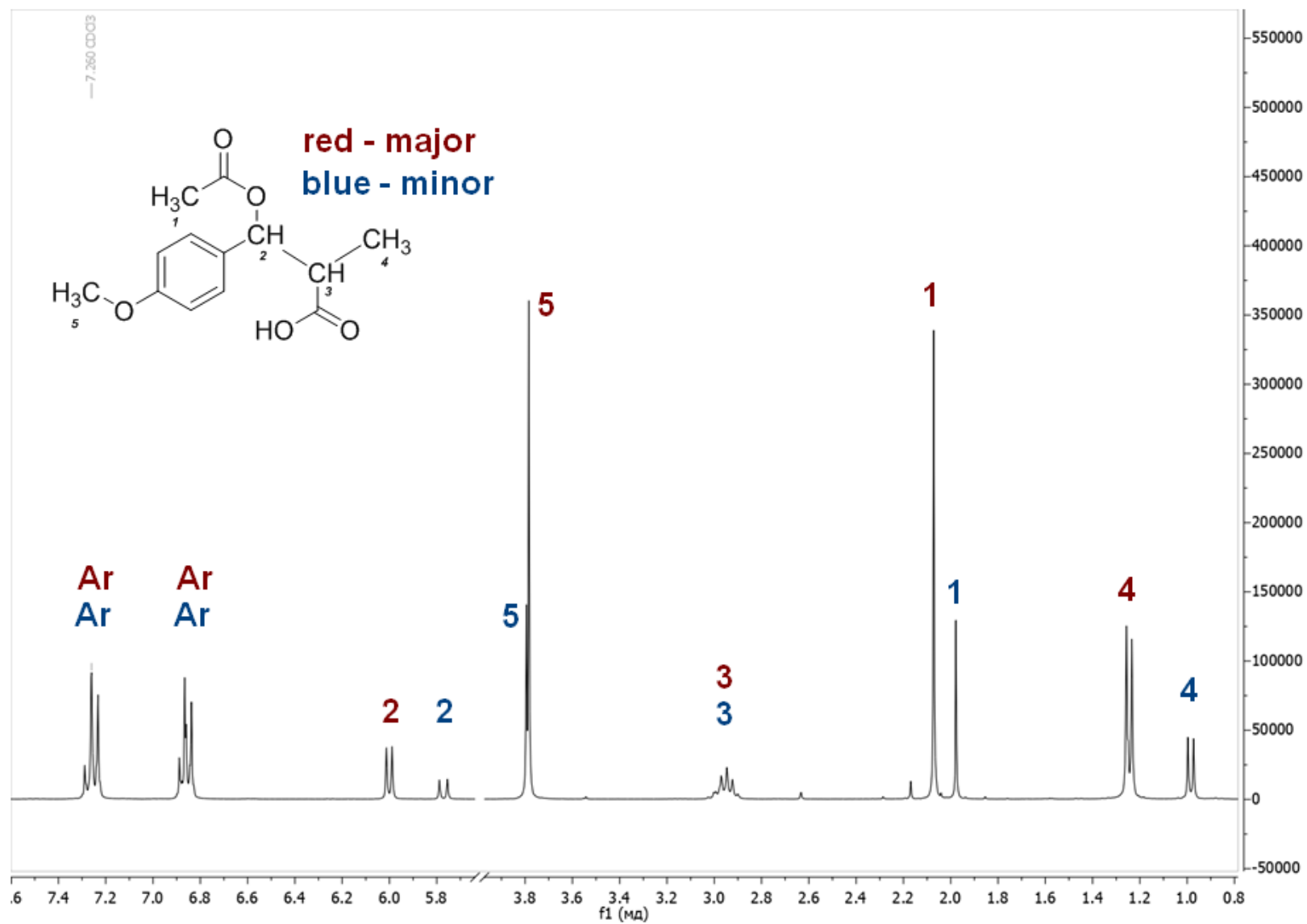
{¹H - ¹³C} HMBC of 3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p



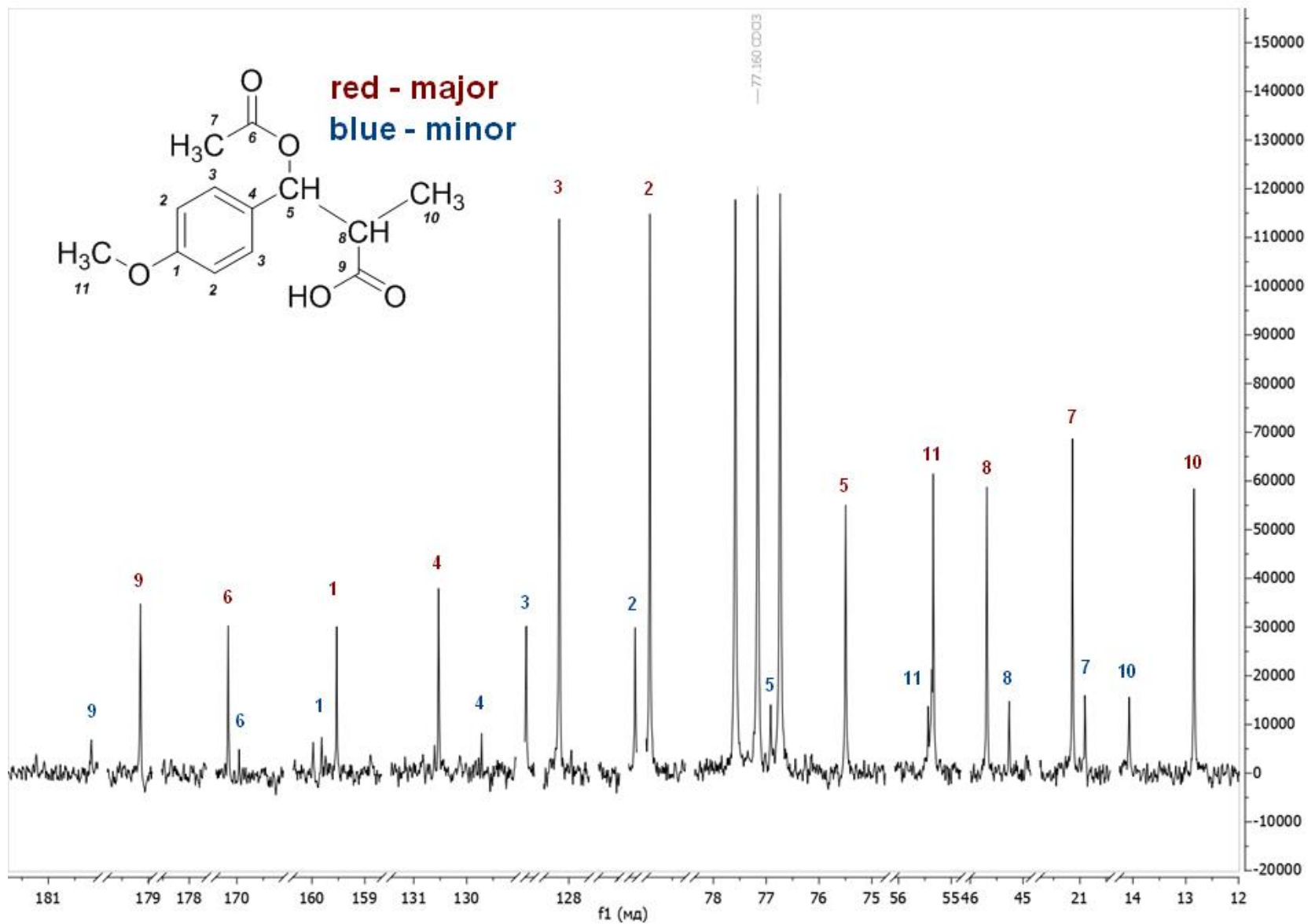
{¹H - ¹³C} HSQC of 3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p



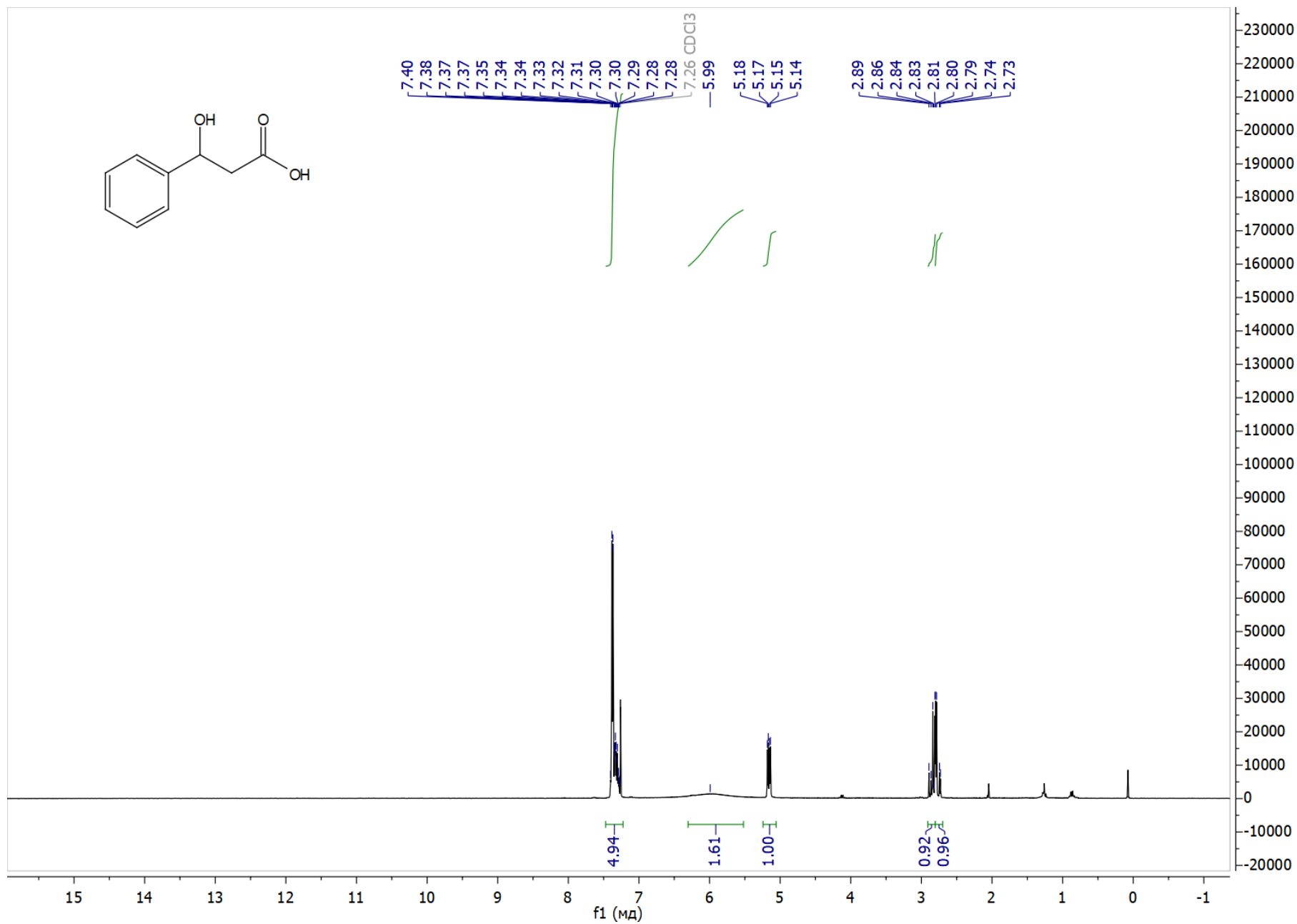
Correlation ^1H NMR (300.13 MHz, CDCl_3), **3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p**



$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Acetoxy-3-(4-methoxyphenyl)-2-methylpropanoic acid, 2p**

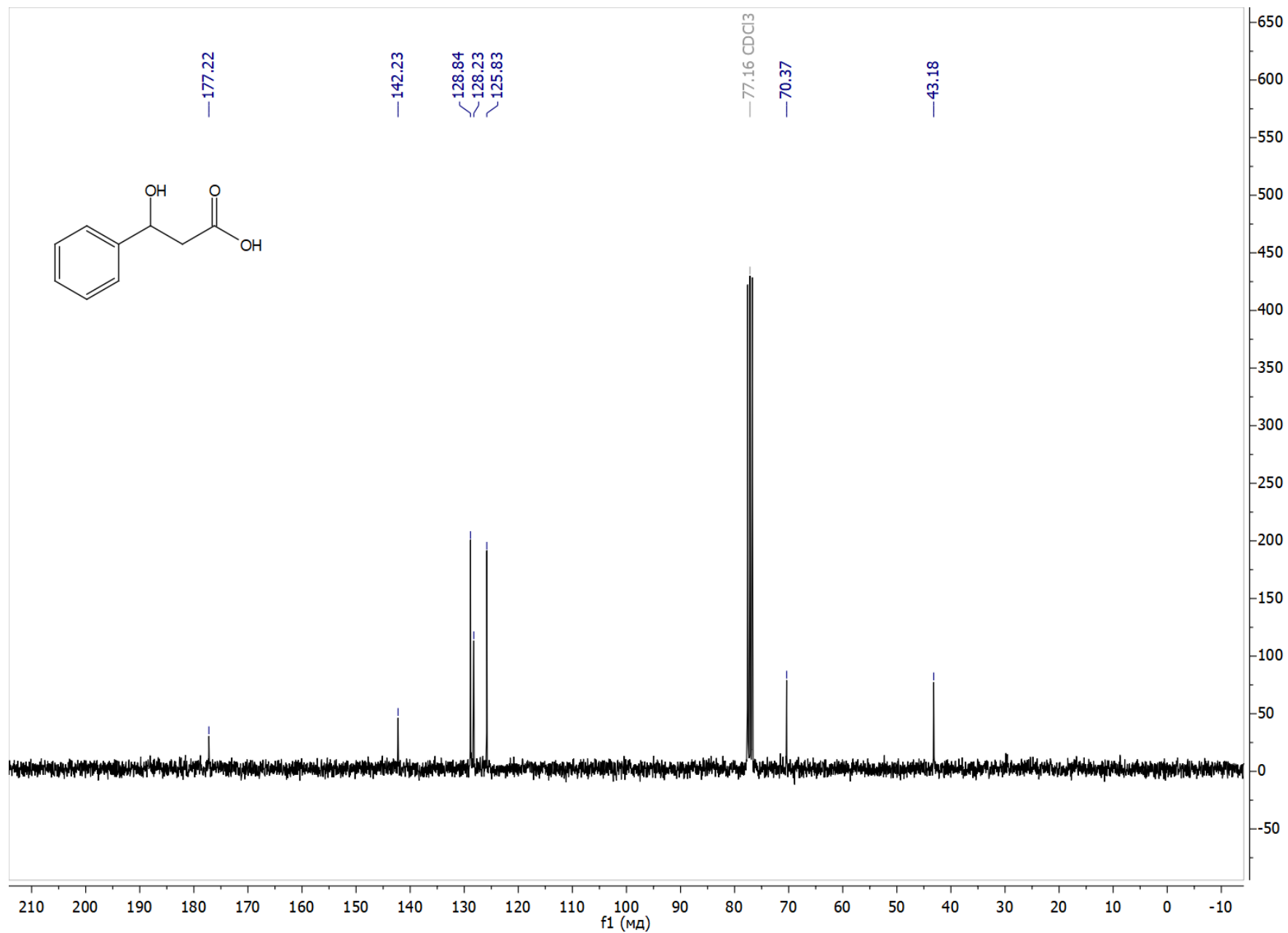


¹H NMR (300.13 MHz, CDCl₃), 3-Hydroxy-3-phenylpropanoic acid, 3a



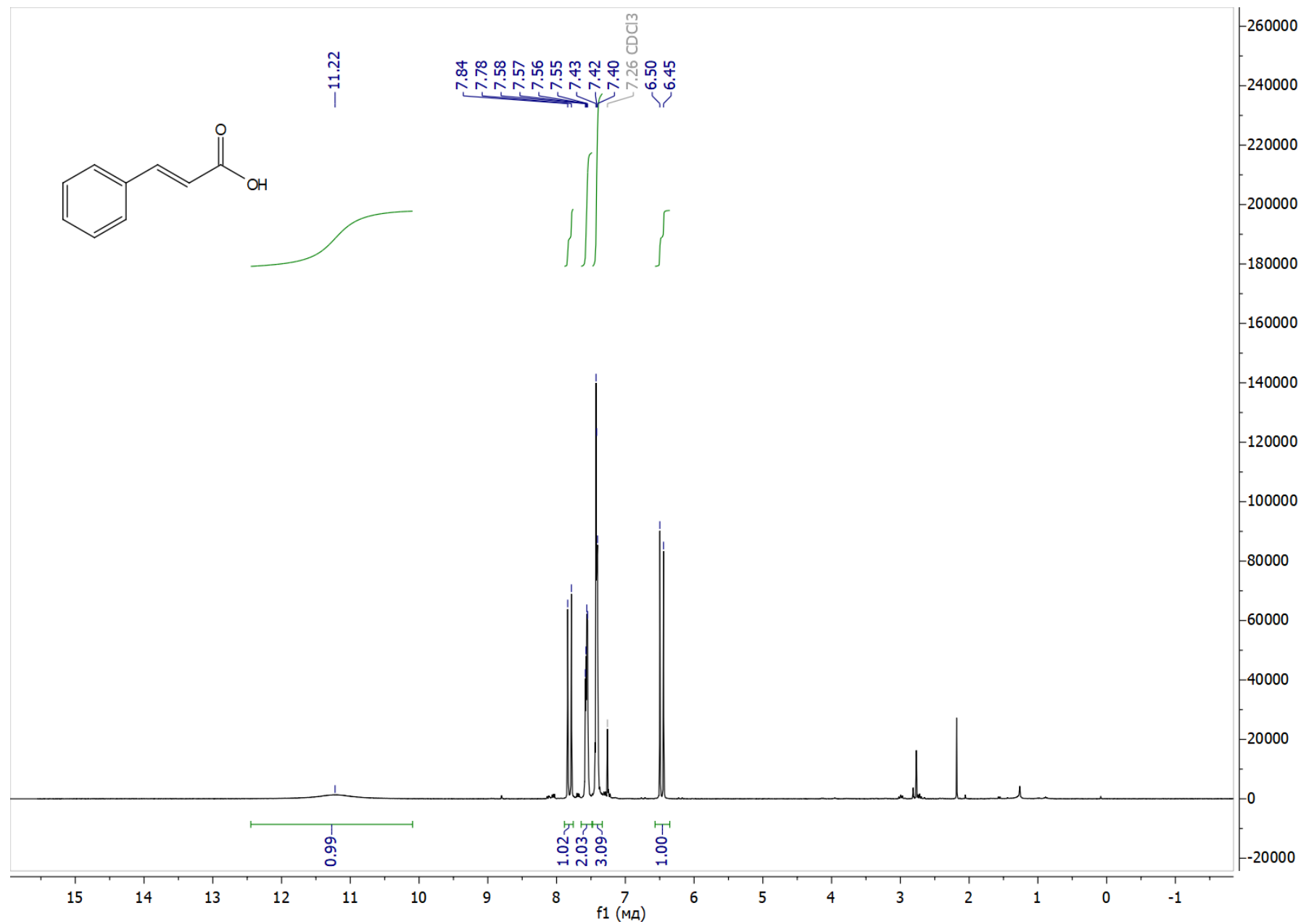
S157

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Hydroxy-3-phenylpropanoic acid, 3a**

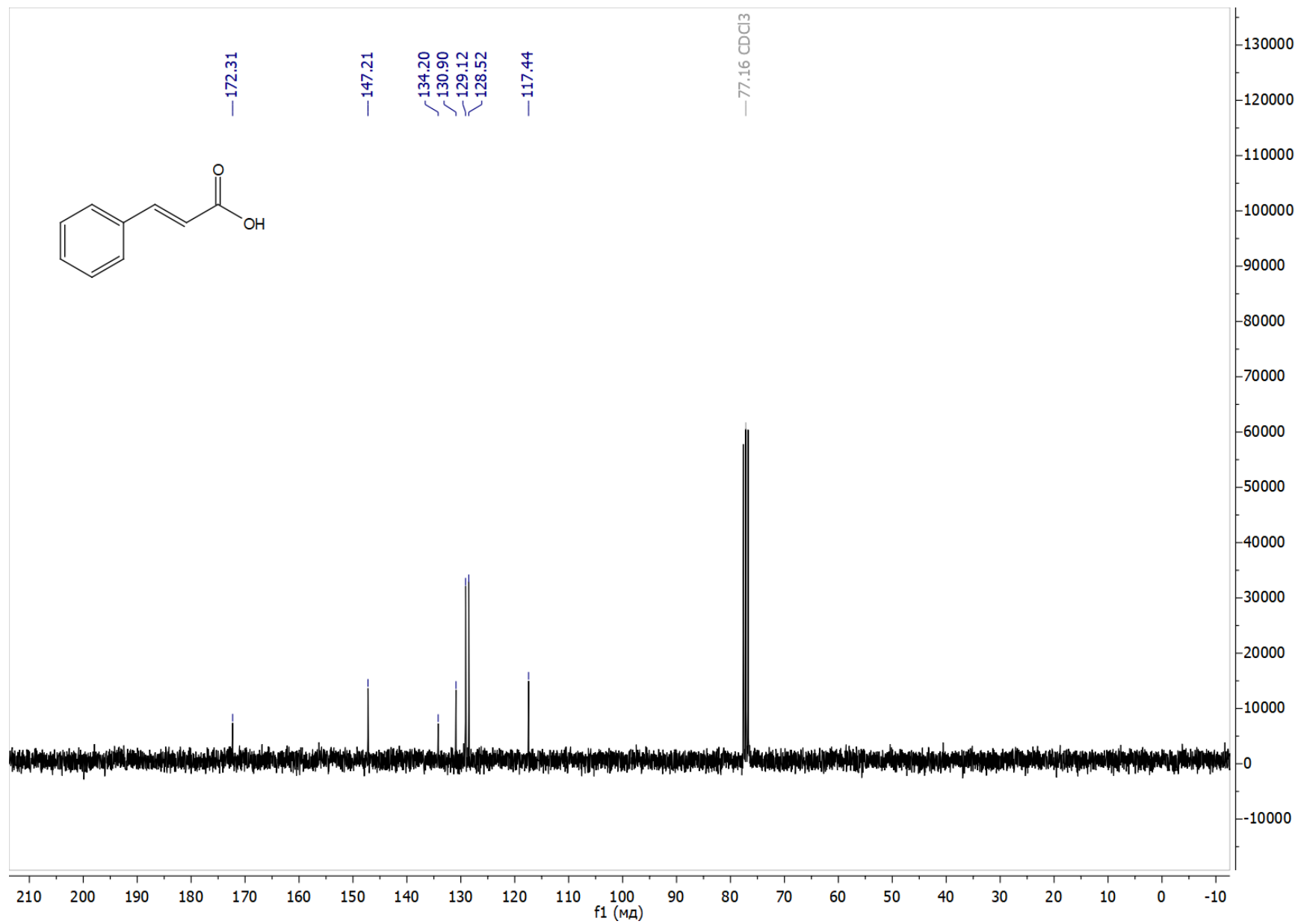


S158

¹H NMR (300.13 MHz, CDCl₃), 3-Phenylacrylic acid, 4a

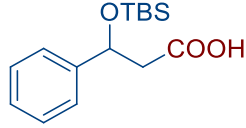


$^{13}\text{C}\{^1\text{H}\}$ NMR (75.48 MHz, CDCl_3), **3-Phenylacrylic acid, 4a**



HRMS spectra of the obtained products.

HRMS spectrum of 3-((*Tert*-butyldimethylsilyloxy)-3-phenylpropanoic acid, P5



Display Report

Analysis Info

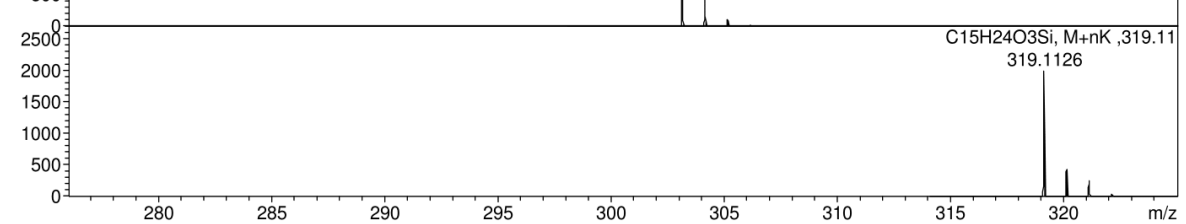
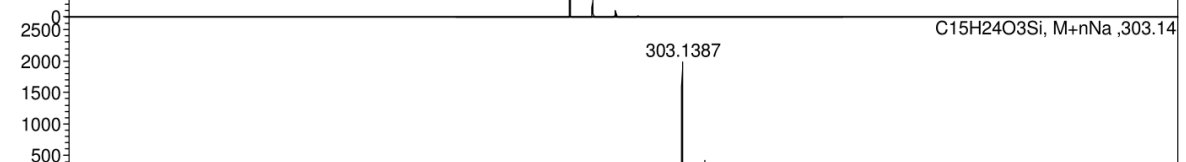
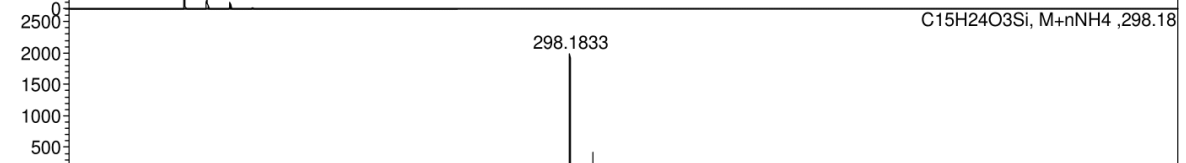
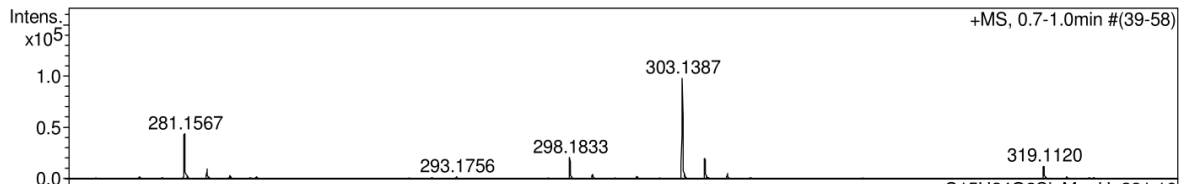
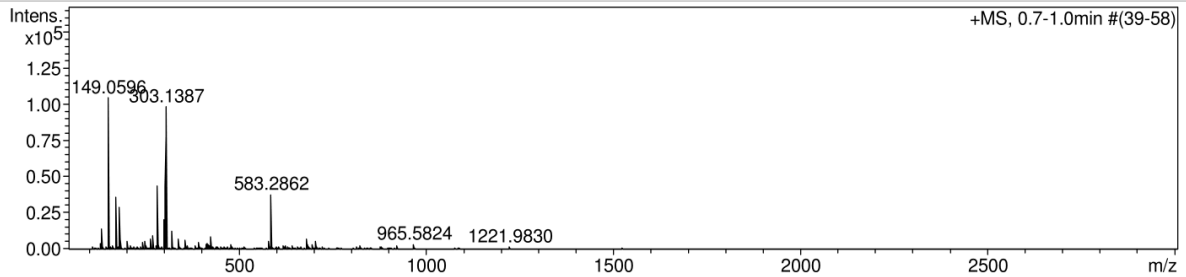
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Sample Name /VILV MG451
Comment C15H24O3Si mH281.1567 calibrant added CH3CN

Acquisition Date 19.03.2024 17:57:12

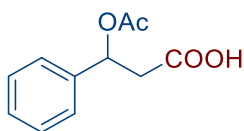
Operator BDAL@DE
Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 1-Phenylvinyl acetate, 2a



Display Report

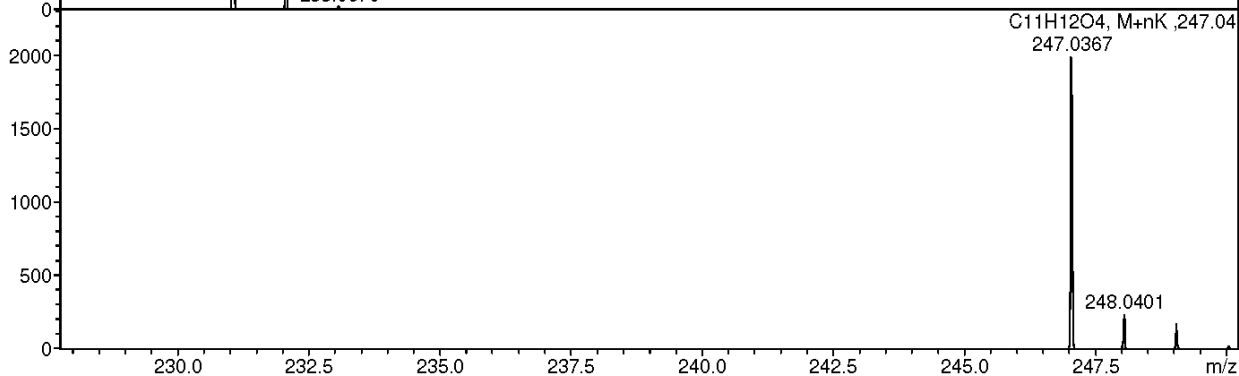
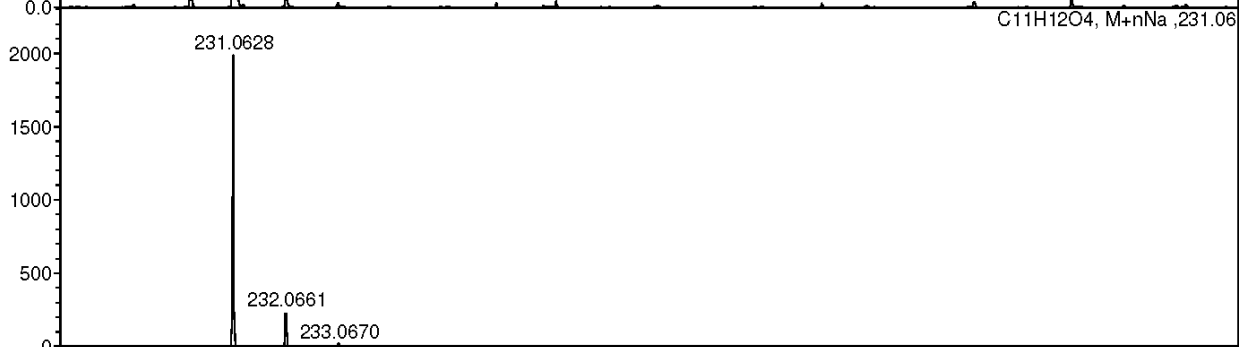
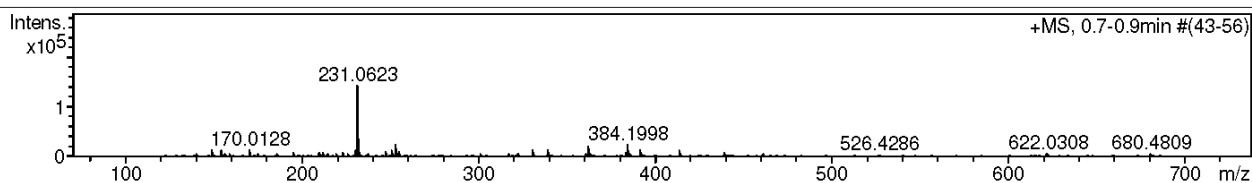
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 Sample Name /TERN MK133
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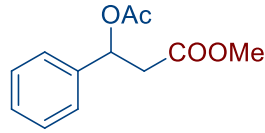
Acquisition Date 24.01.2023 17:34:24
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	2500 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of methyl 3-acetoxy-3-phenylpropanoate, 2a'



Display Report

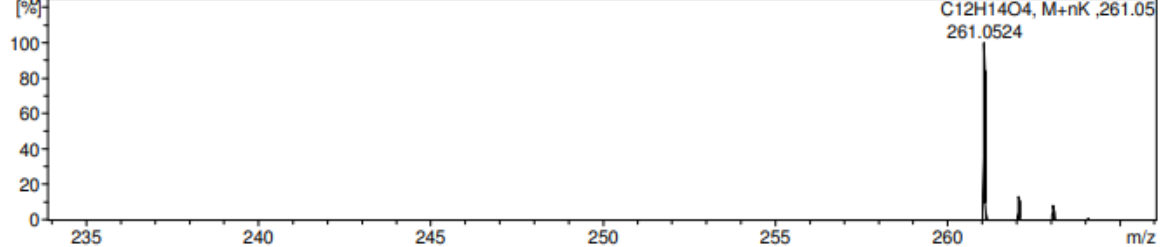
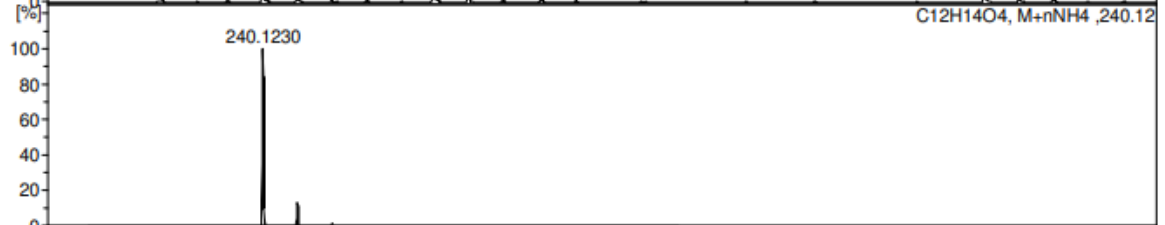
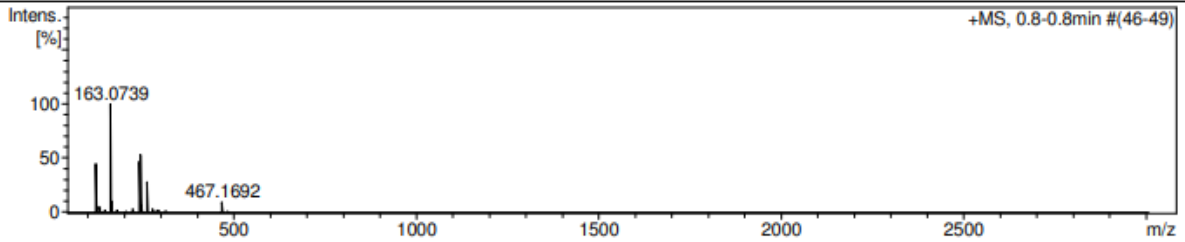
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 Sample Name /VILV MG313
 Comment C12H14O4 mH223.0964 calibrant added CH3CN

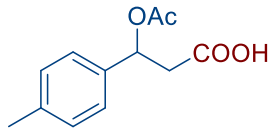
Acquisition Date 27.06.2024 9:30:25
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-3-(p-tolyl)propanoic acid, 2b



Display Report

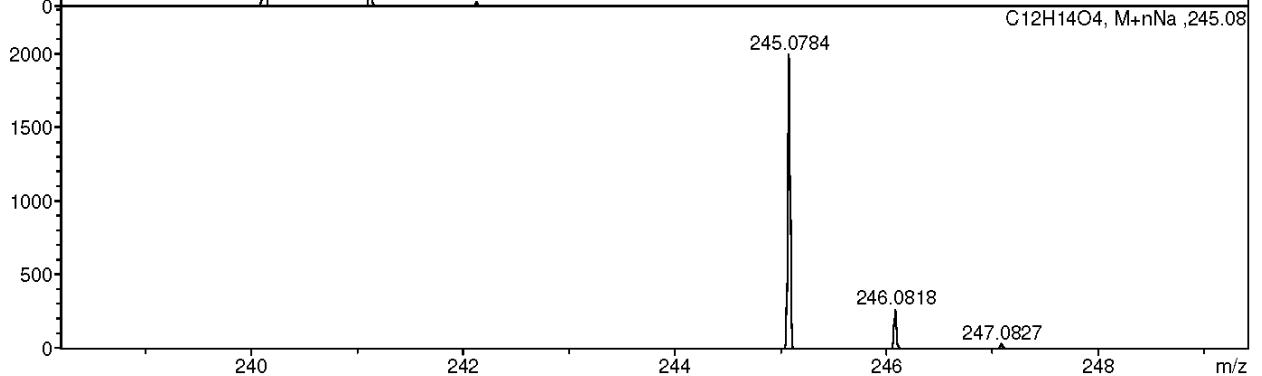
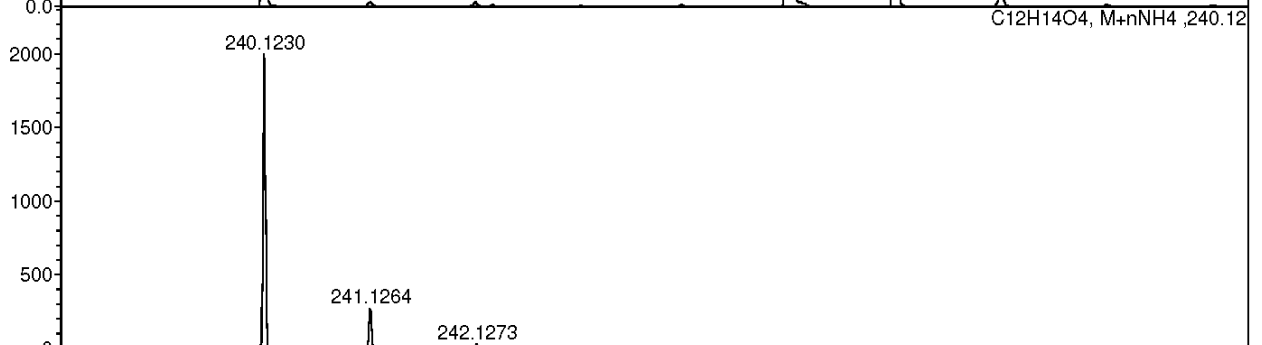
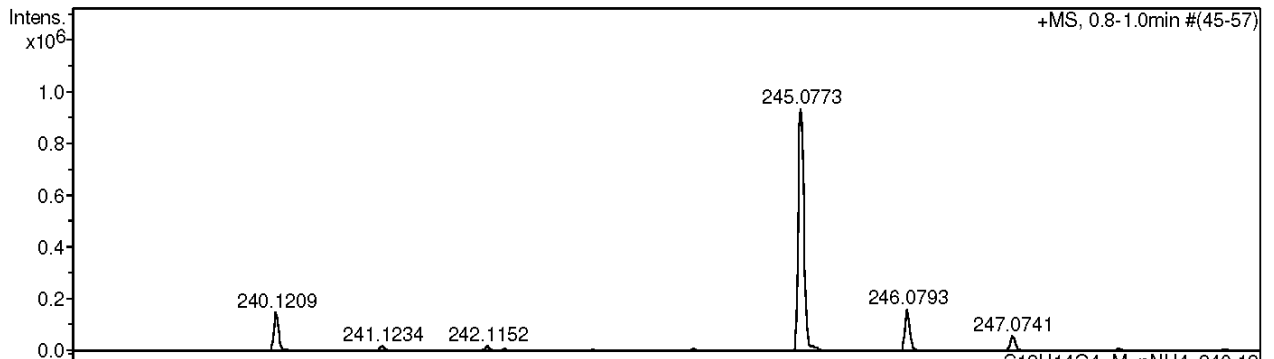
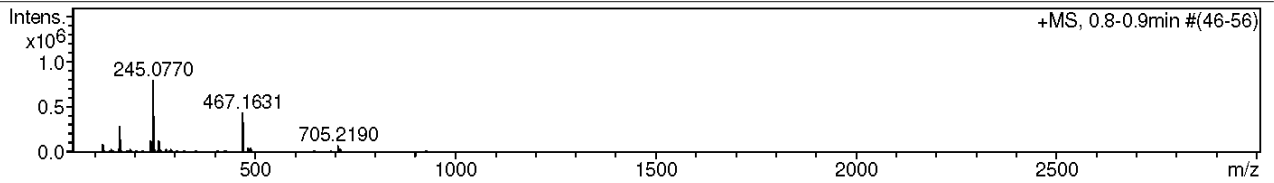
Analysis Info

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 Sample Name /VILV MG322
 Comment C12H14O4 mH223.0964 calibrant added CH3OH

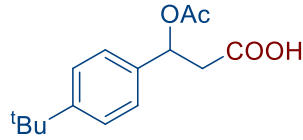
Acquisition Date 29.11.2023 11:59:56
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-3-(4-(tert-butyl)phenyl)propanoic acid, 2c



Display Report

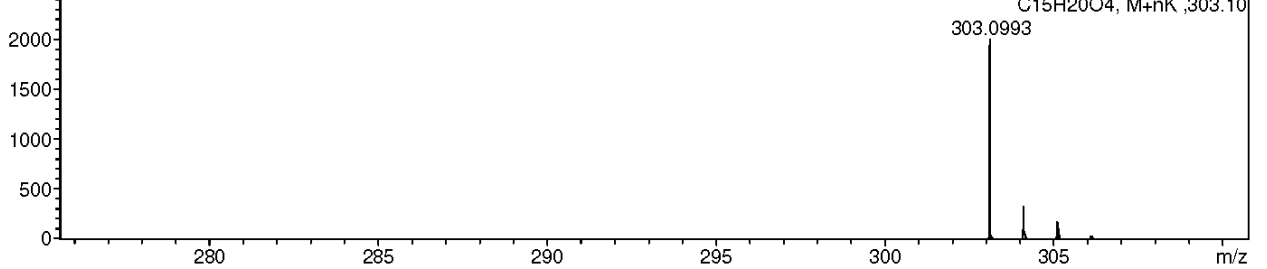
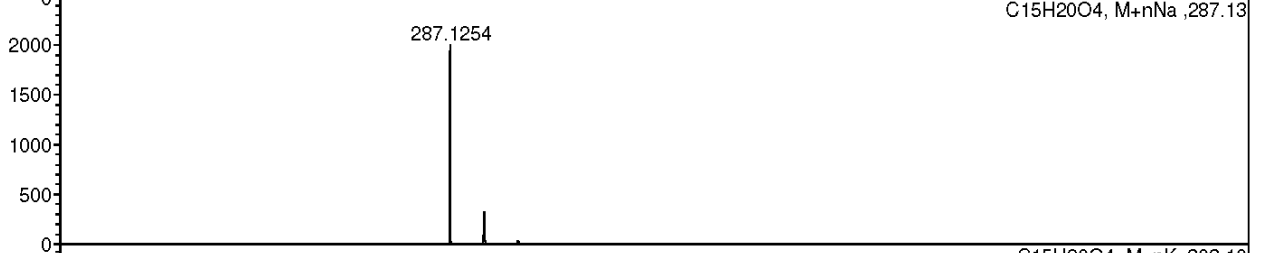
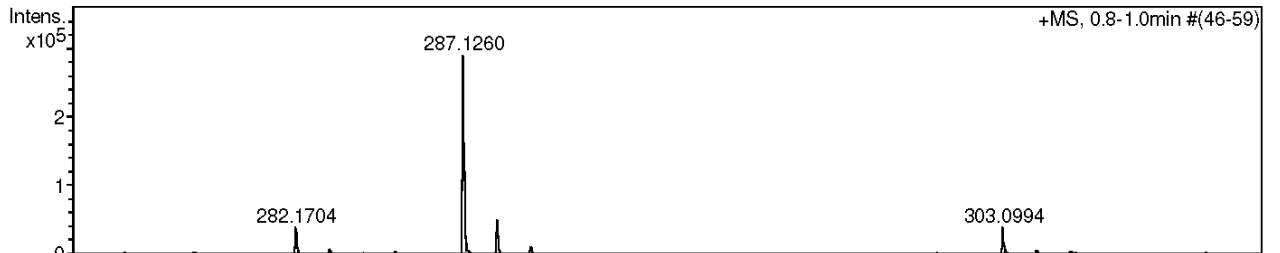
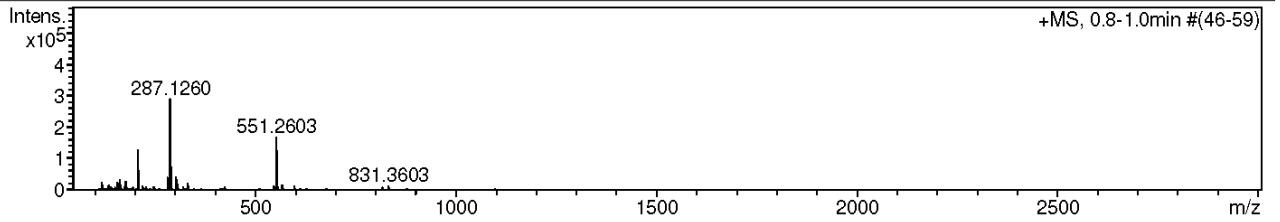
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 Method tune_low.m
 Sample Name /VILV MG390
 Comment C15H20O4 mH265.1434 calibrant added CH3OH

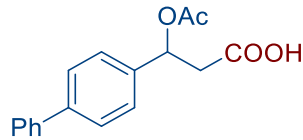
Acquisition Date 08.12.2023 16:56:10
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-([1,1'-Biphenyl]-4-yl)-3-acetoxypropanoic acid, 2d



Display Report

Analysis Info

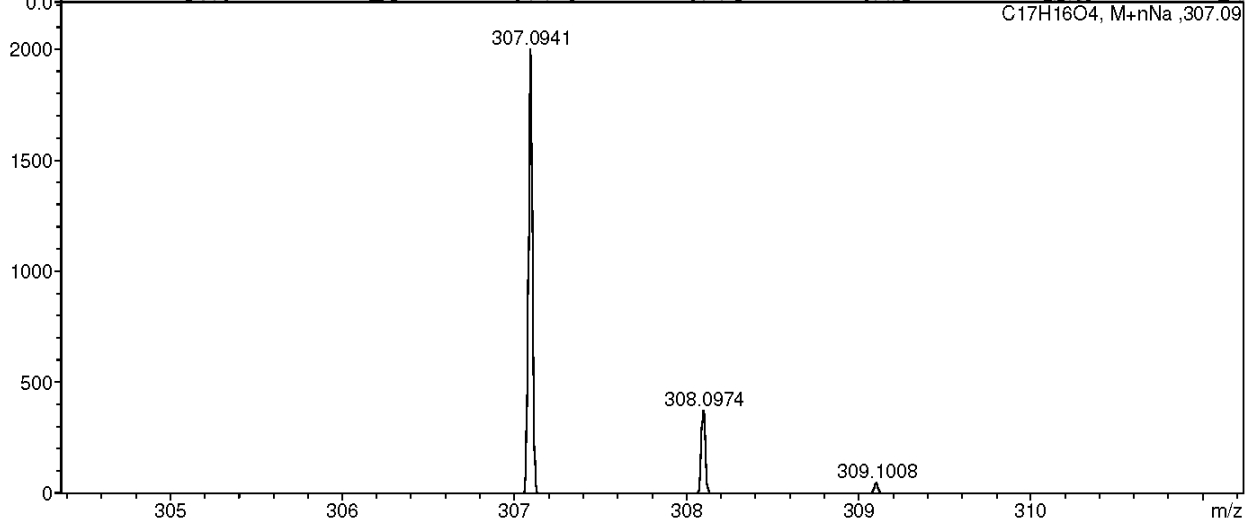
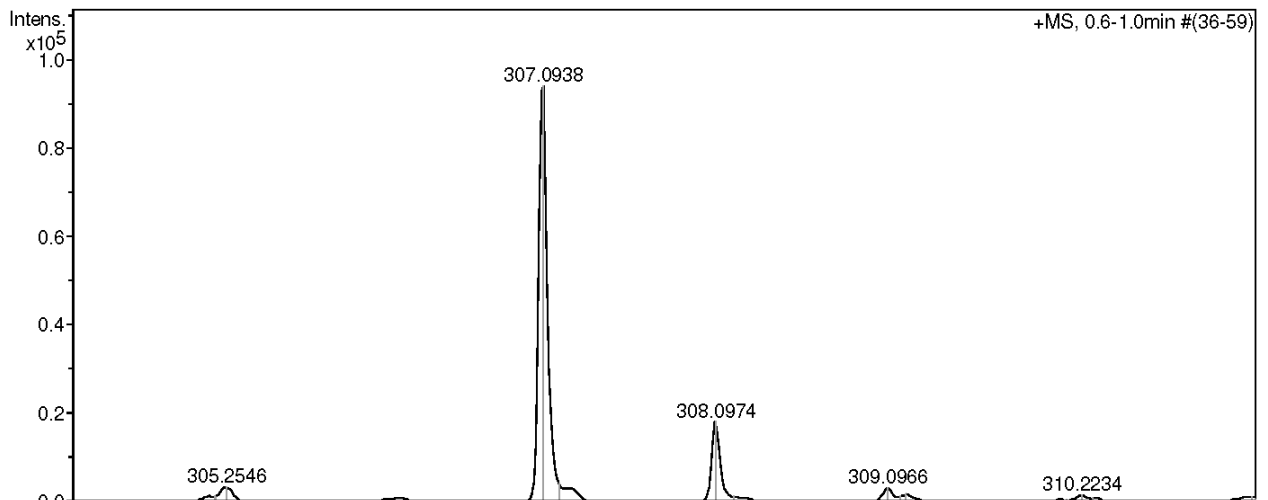
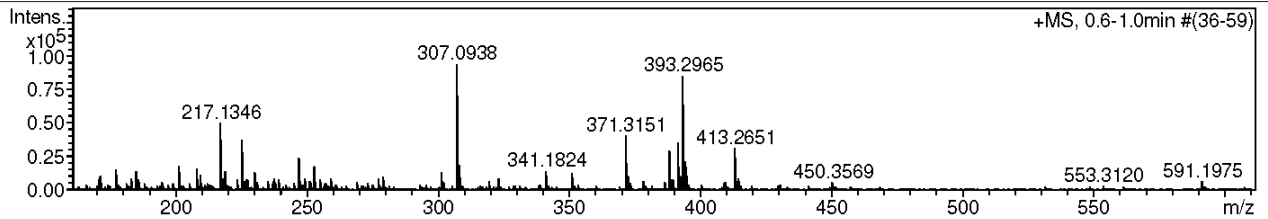
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 Sample Name /VILV MG387
 Comment C17H16O4 mH 285.1121 calibrant added CH3OH

Acquisition Date 07.12.2023 14:12:14

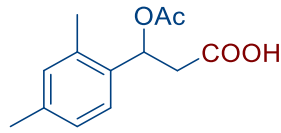
Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-3-(2,4-dimethylphenyl)propanoic acid, 2e



Display Report

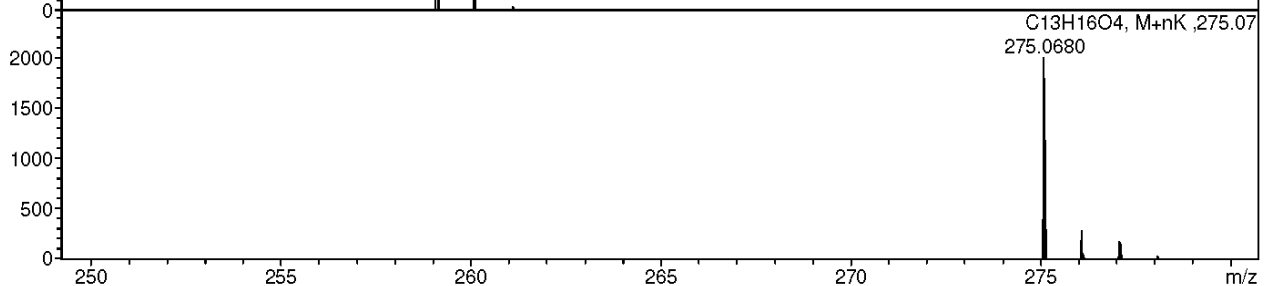
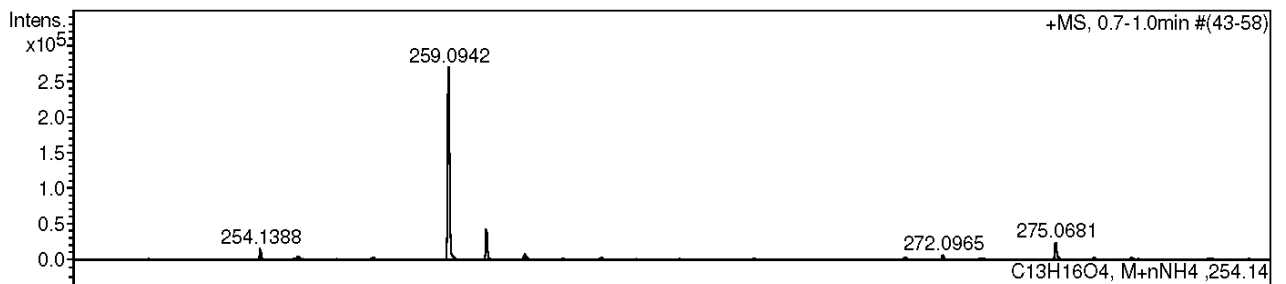
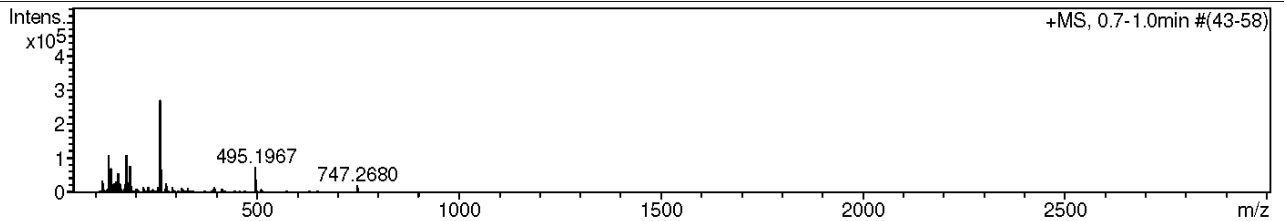
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 Sample Name /VILV MG388
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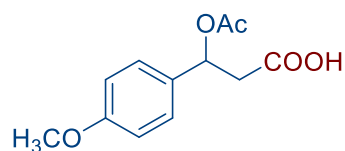
Acquisition Date 08.12.2023 8:42:29
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-3-(4-methoxyphenyl)propanoic acid, 2f



Display Report

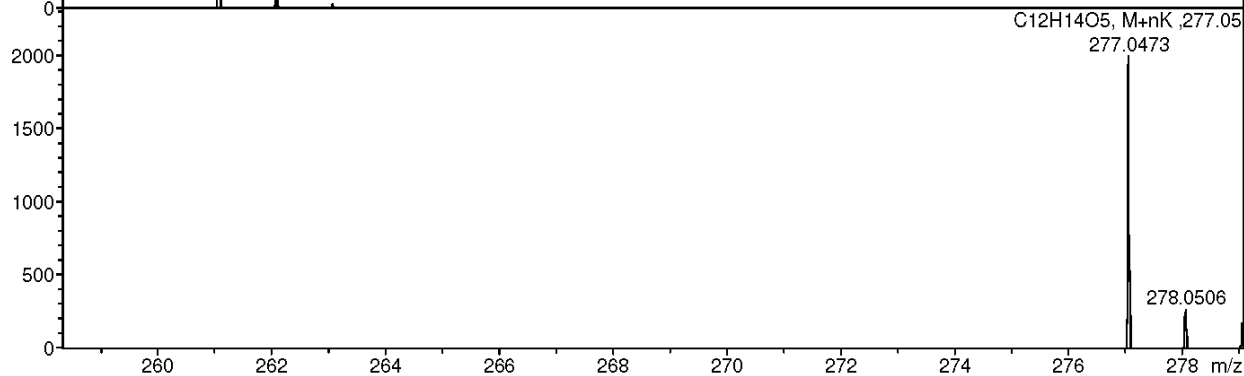
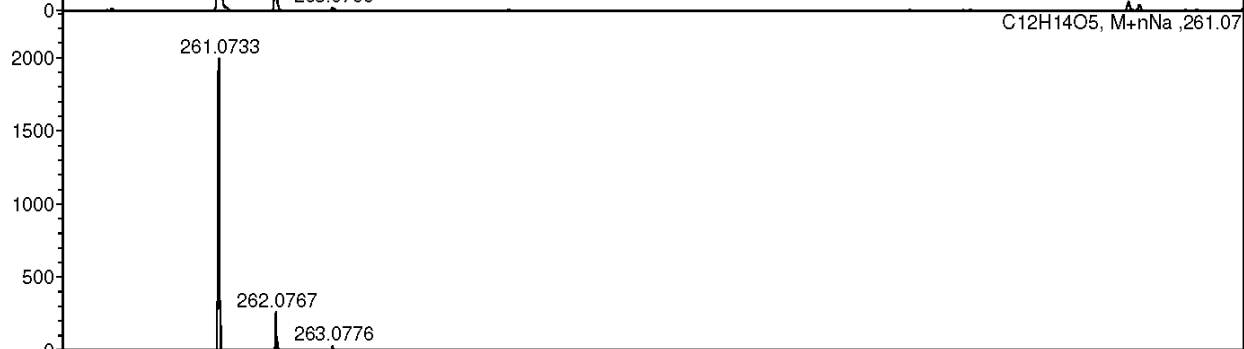
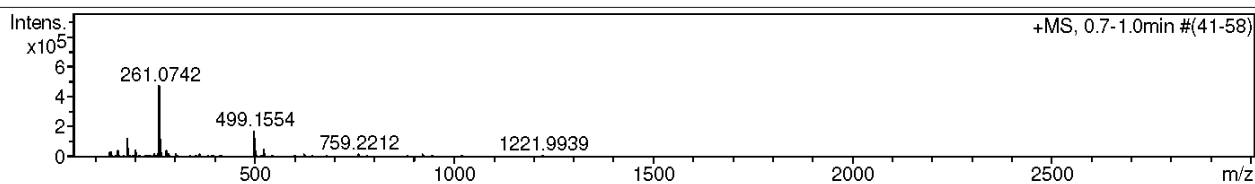
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 Sample Name /VILV MG323
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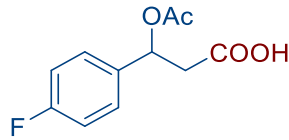
Acquisition Date 29.11.2023 11:27:23
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of **3-Acetoxy-3-(4-fluorophenyl)propanoic acid, 2g**



Display Report

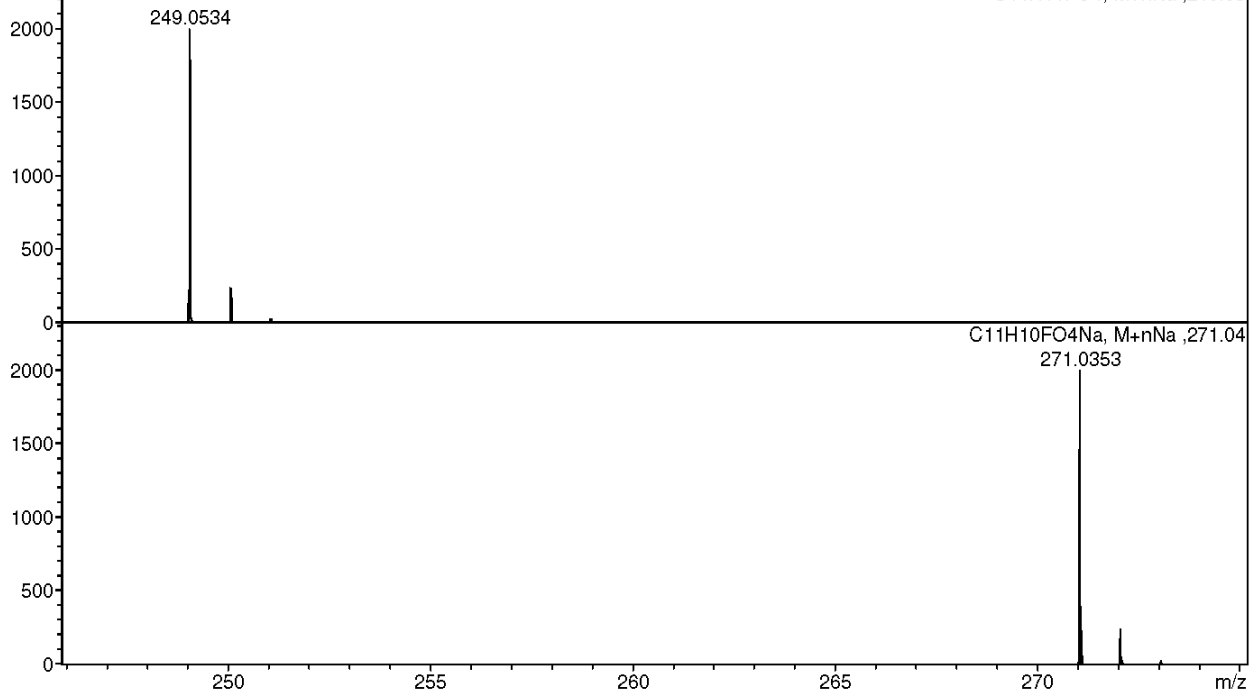
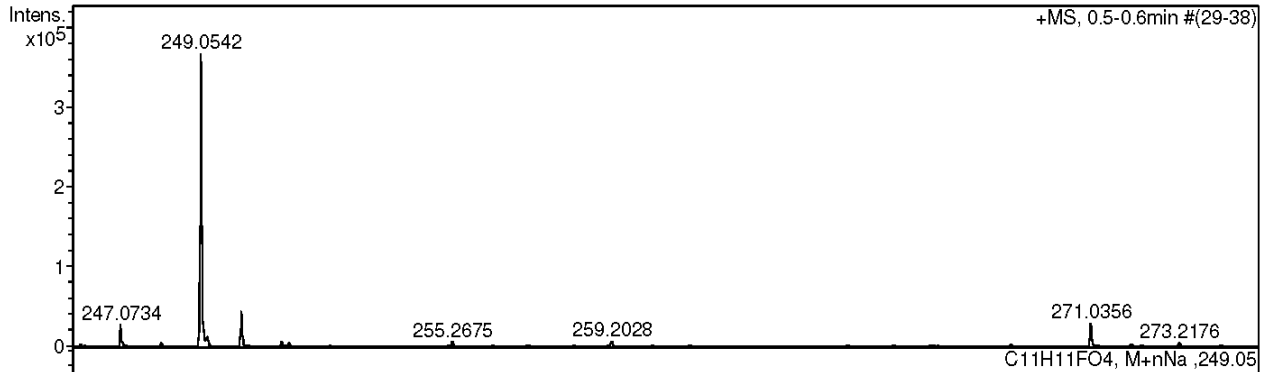
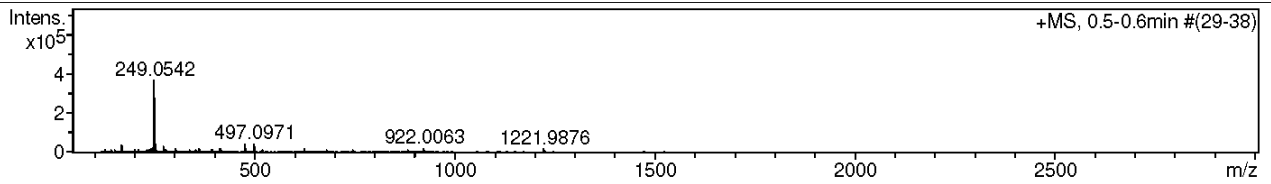
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 Sample Name /VILV MG291
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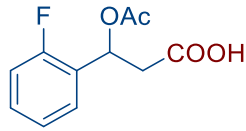
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 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-3-(2-fluorophenyl)propanoic acid, 2h



Display Report

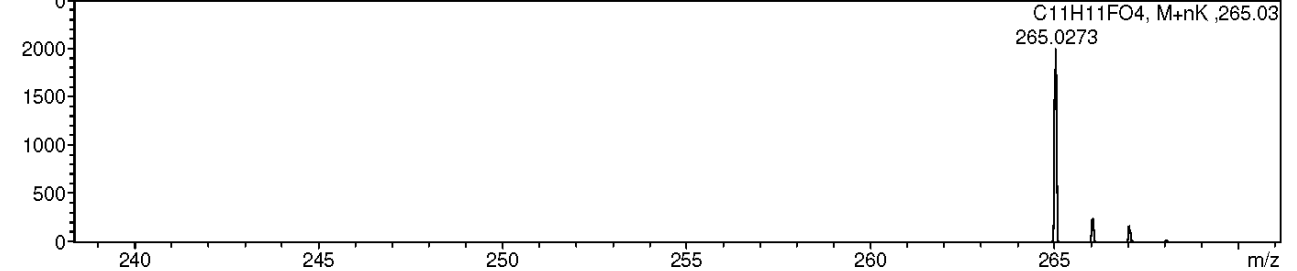
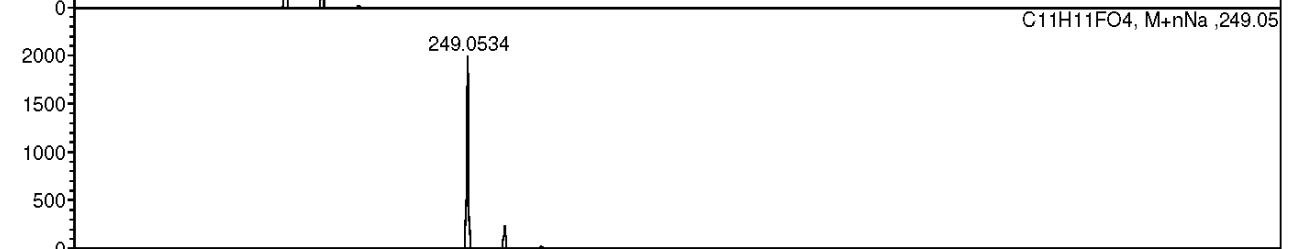
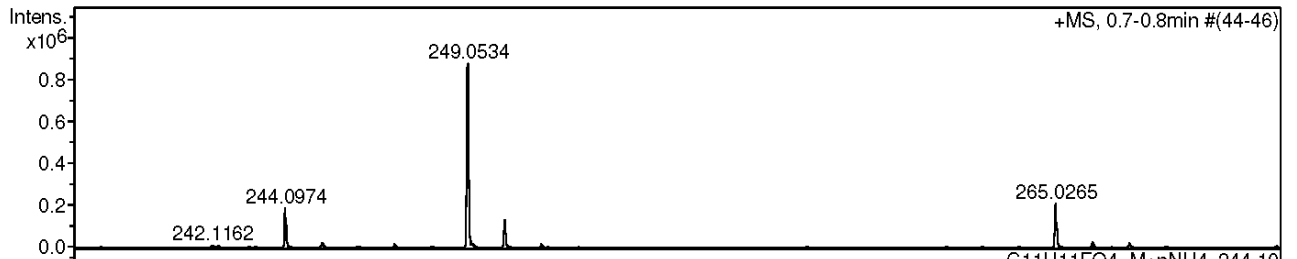
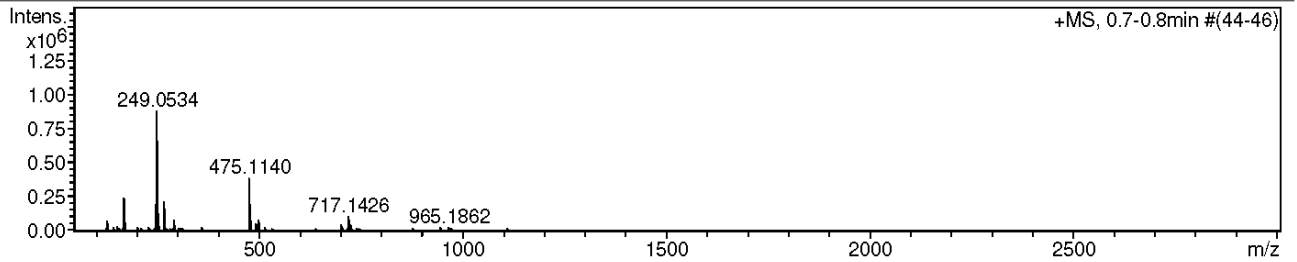
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 Sample Name /TERN MG-394
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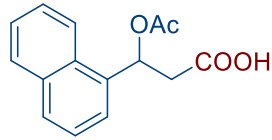
Acquisition Date 31.10.2023 16:17:10
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-3-(naphthalen-1-yl)propanoic acid, 2i



Display Report

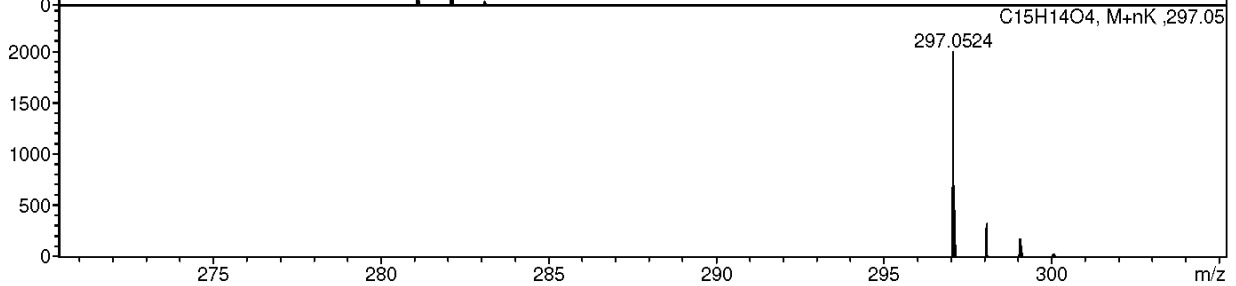
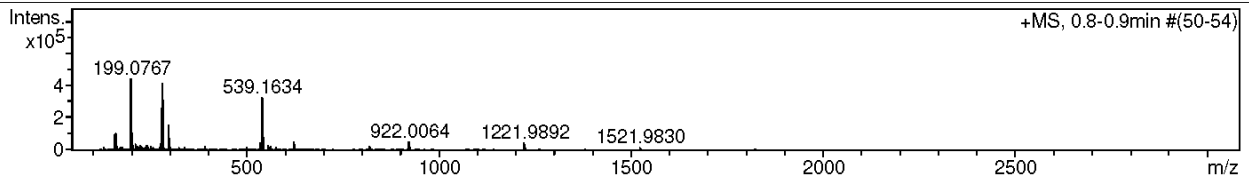
Analysis Info

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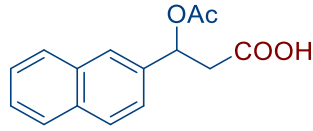
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 Operator BDAL@DE
 Instrument / Ser# microTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of **3-Acetoxy-3-(naphthalen-2-yl)propanoic acid, 2j**



Display Report

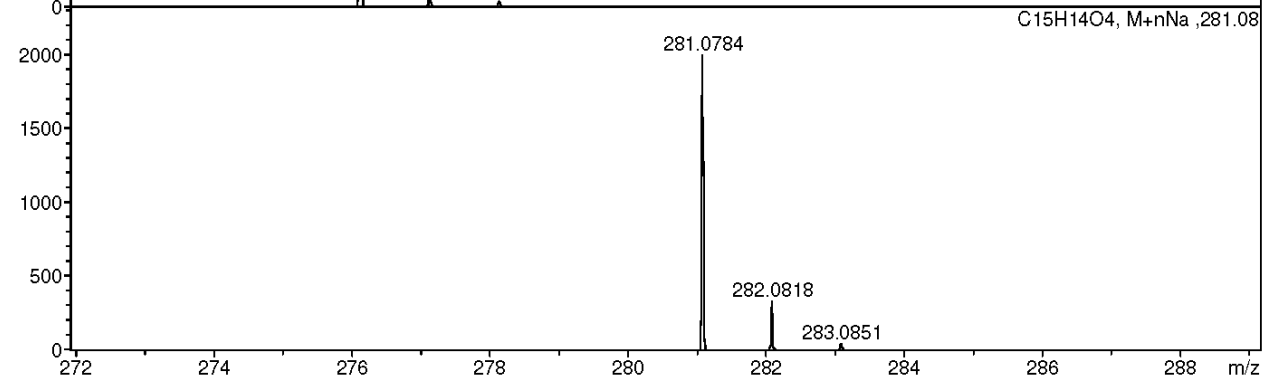
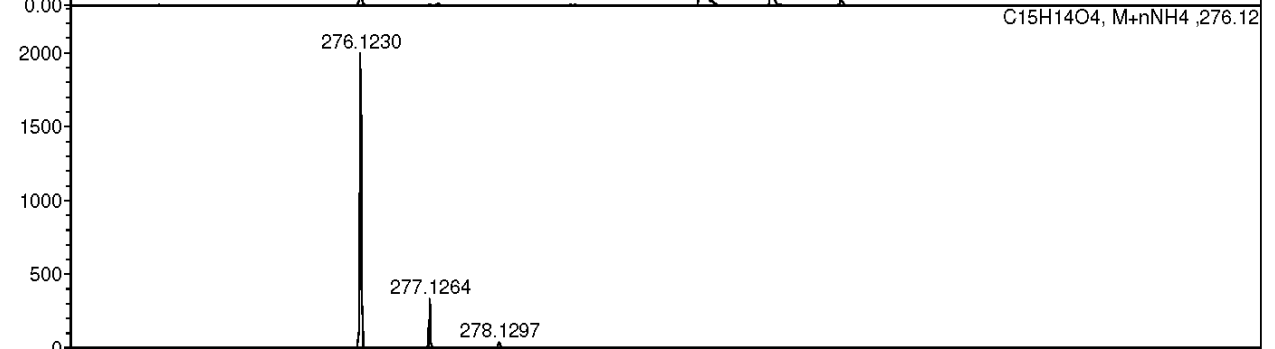
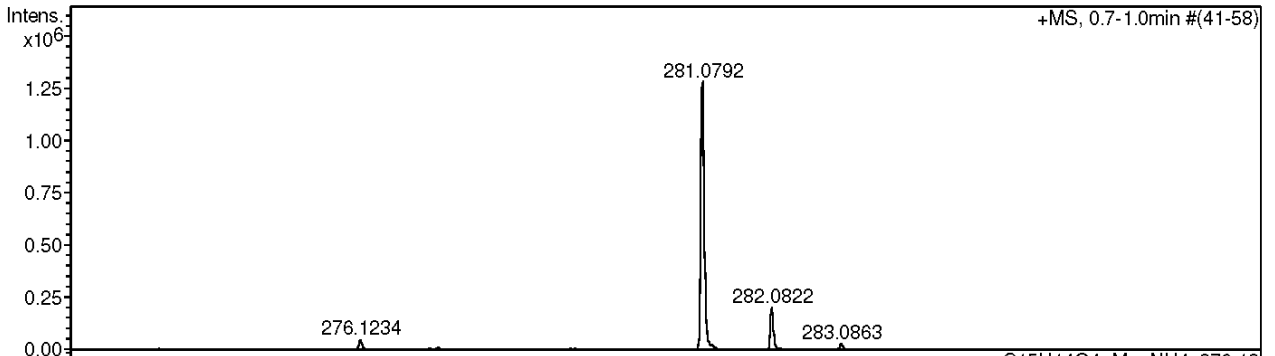
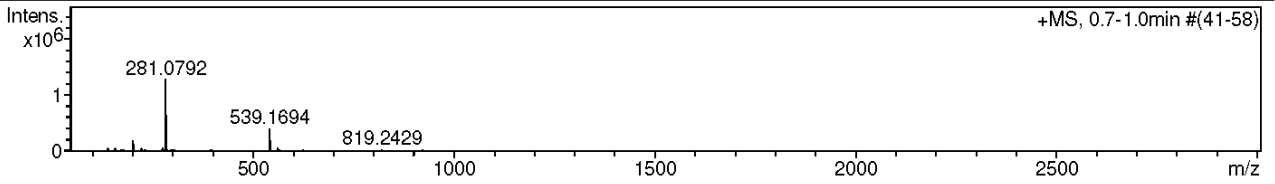
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 Sample Name /MILV MG278
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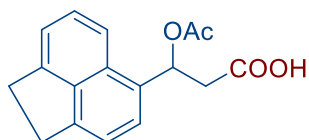
Acquisition Date 30.11.2023 10:30:30
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-3-(1,2-dihydroacenaphthylen-5-yl)propanoic acid, 2k



Display Report

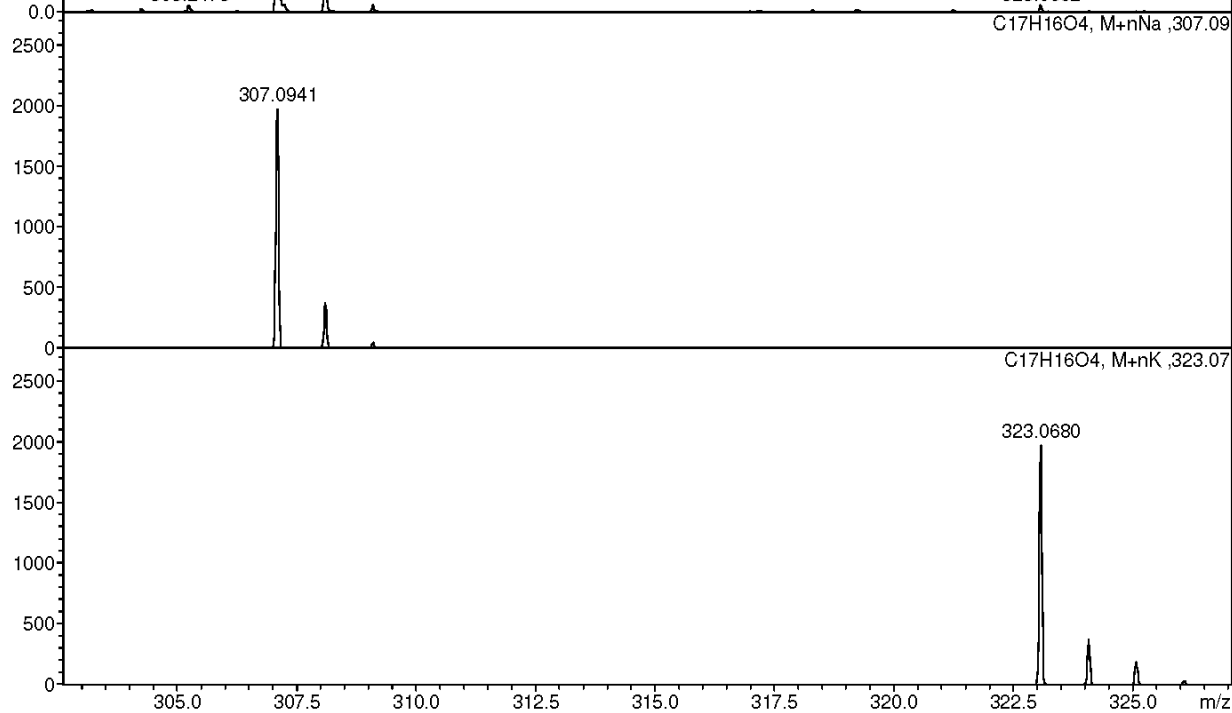
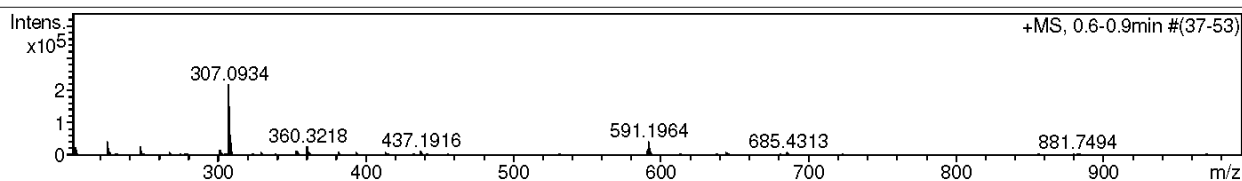
Analysis Info

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 Sample Name /VILV MG-368
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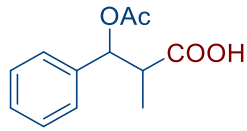
Acquisition Date 22.11.2023 12:45:35
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-2-methyl-3-phenylpropanoic acid, 2l



Display Report

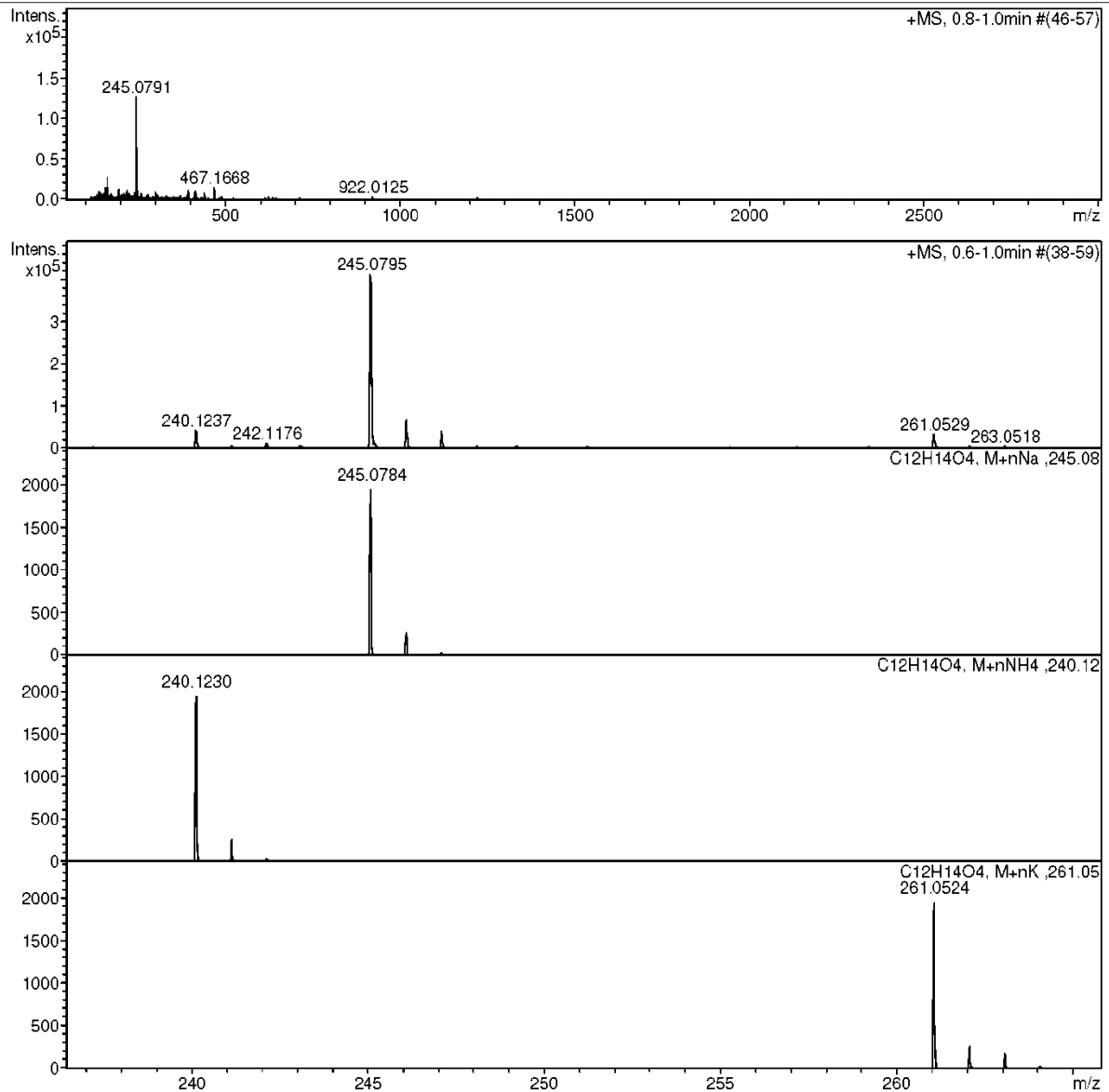
Analysis Info

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 Sample Name /VILV MG295
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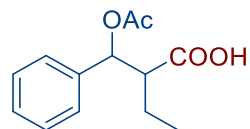
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 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 2-(Acetoxy(phenyl)methyl)butanoic acid, 2m



Display Report

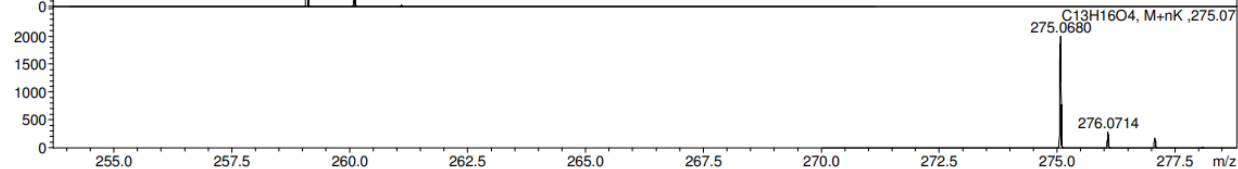
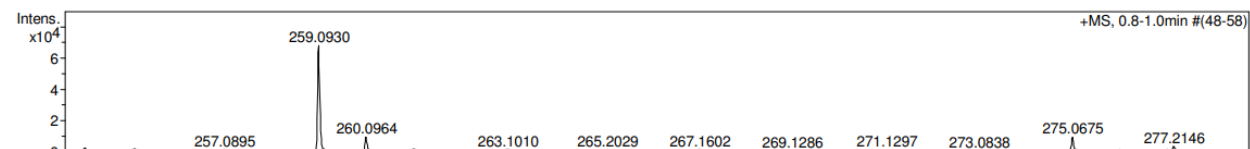
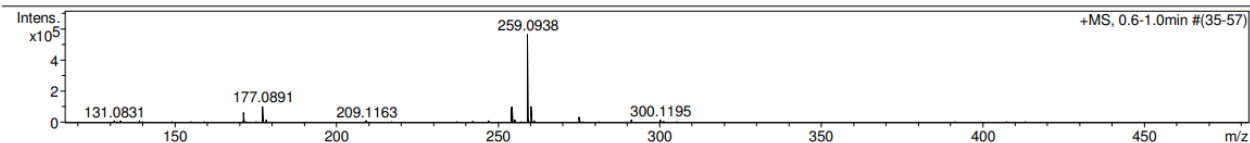
Analysis Info

Analysis Name D:\Data\Kolotyrkina\2023\Ustuzhanin\1130026.d
 Method tune_low.m
 Sample Name /VILV MG366
 Comment C13H16O4 mH 237.1121 calibrant added CH3OH

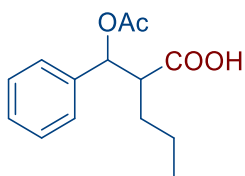
Acquisition Date 30.11.2023 15:02:03
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 2-(Acetoxy(phenyl)methyl)pentanoic acid, 2n



Display Report

Analysis Info

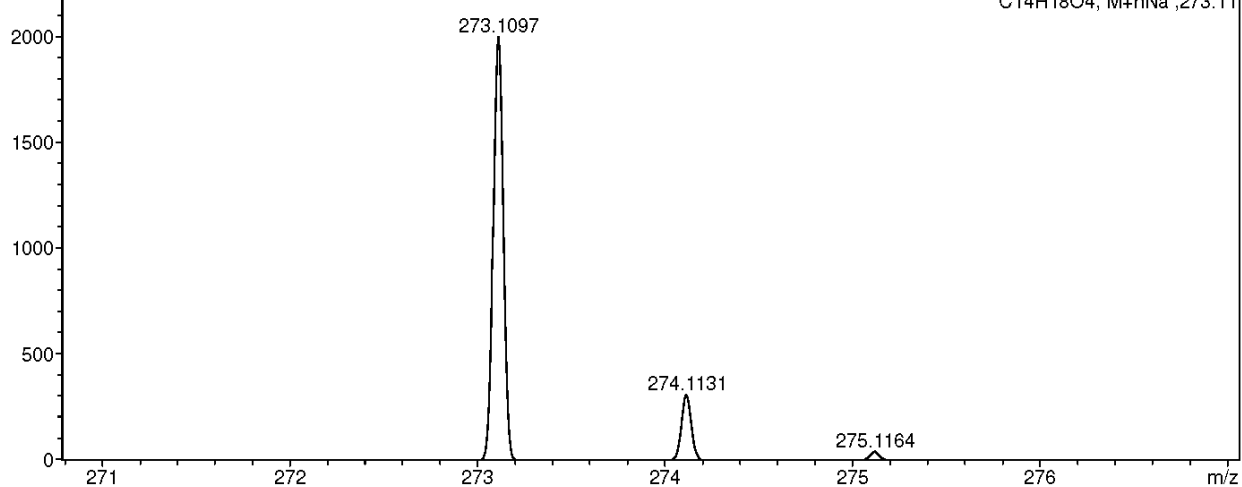
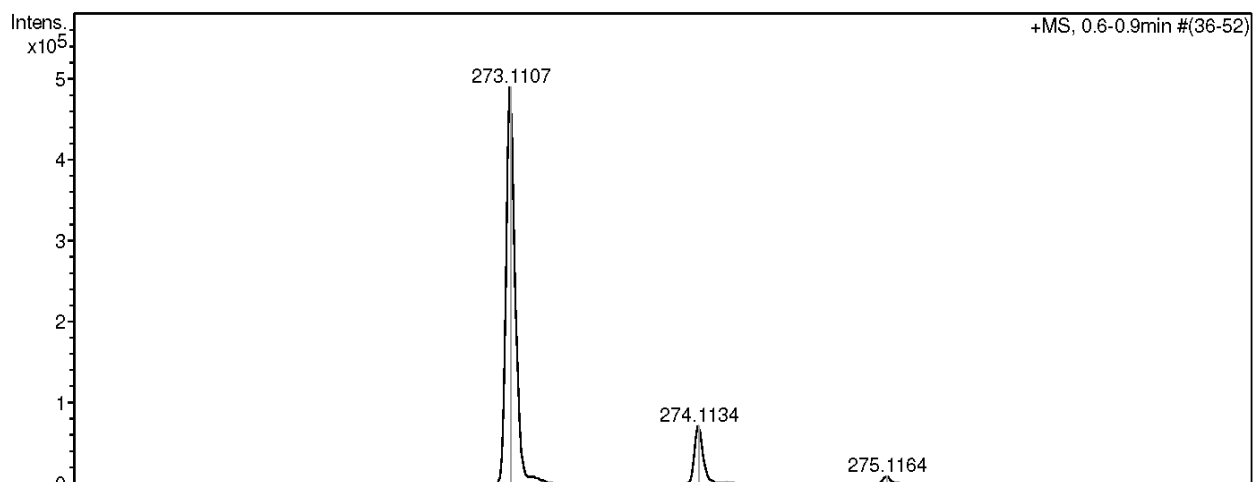
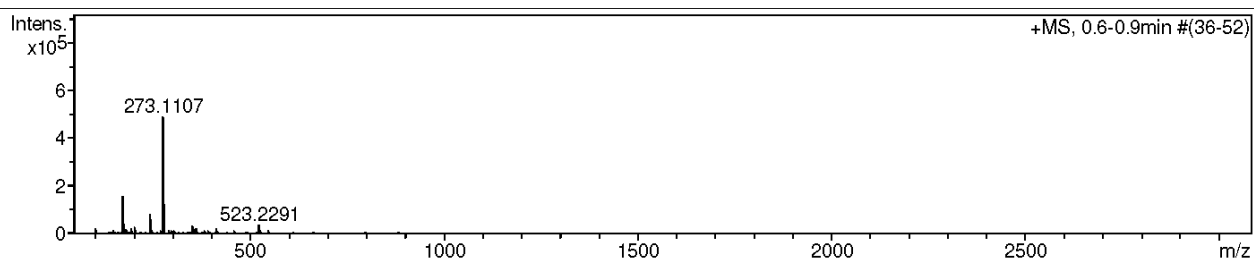
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 Method tune_low.m
 Sample Name /VILV MG398
 Comment C14H18O4 mH 251.1277 clb added CH3OH

Acquisition Date 22.12.2023 10:20:10

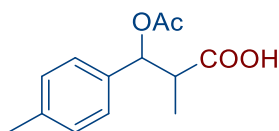
Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Acetoxy-2-methyl-3-(p-tolyl)propanoic acid, 2o



Display Report

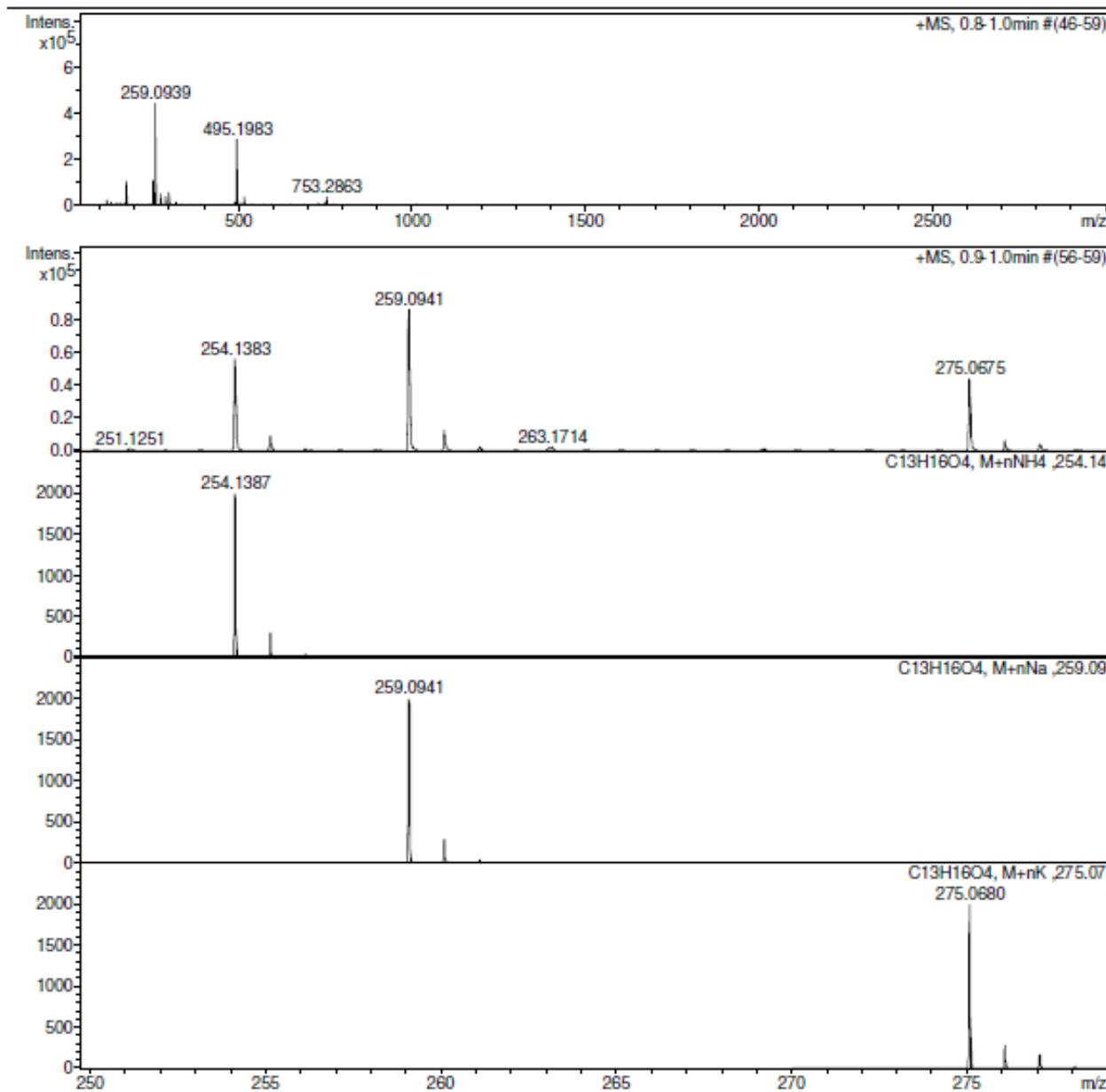
Analysis Info

Analysis Name D:\Data\Kolotyrkina\2024\Ustuzhanin\0306002.d
 Method tune_low.m
 Sample Name /VILV MG445
 Comment C13H16O4 mH237.1121 calibrant added CH3OH

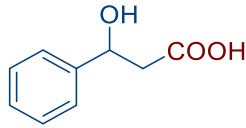
Acquisition Date 06.03.2024 11:00:38
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of **3-Hydroxy-3-phenylpropanoic acid, 3a**



Display Report

Analysis Info

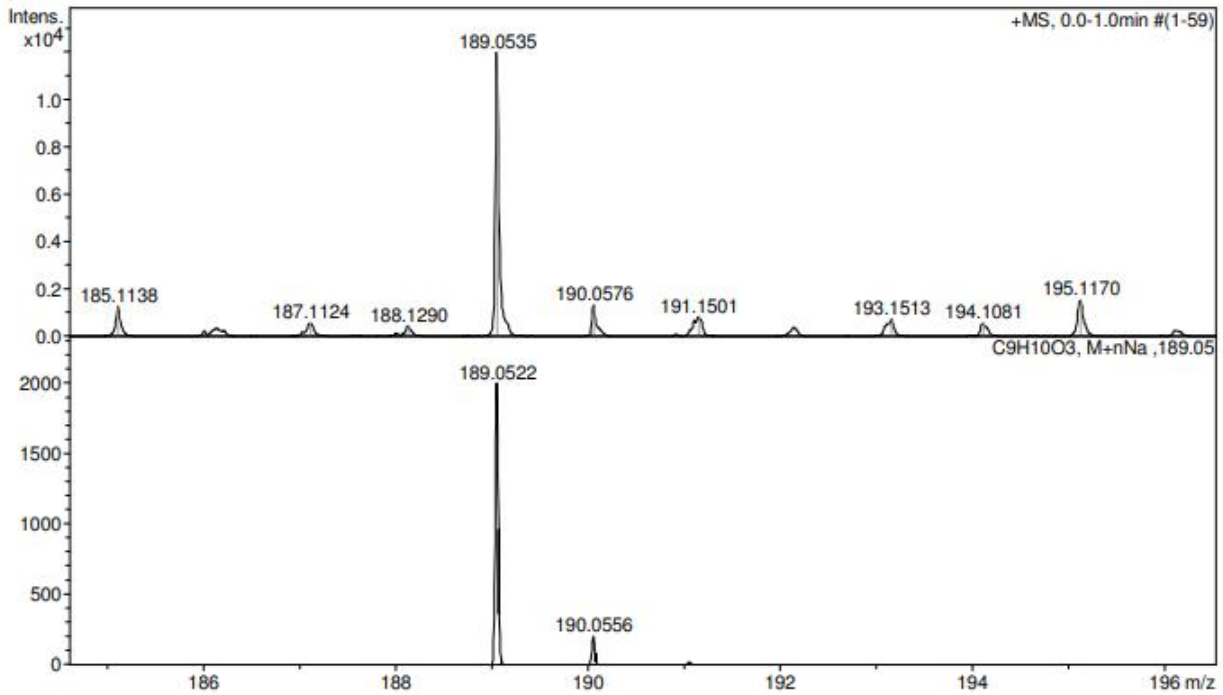
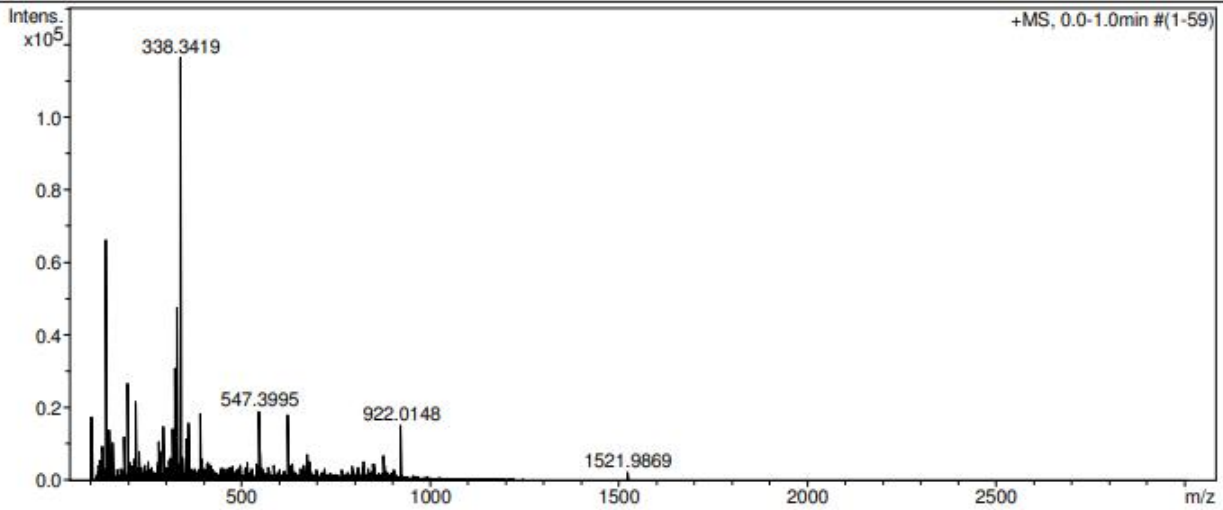
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 Method tune_low.m
 Sample Name /MILV MG296
 Comment CH3CN 100 %, dil. 20, calibrant added

Acquisition Date 16.02.2024 16:44:52

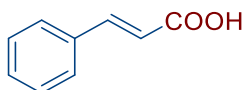
Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste



HRMS spectrum of 3-Phenylacrylic acid, 4a



Display Report

Analysis Info

Analysis Name D:\Data\Kolotyorkina\2024\Ustuzhanin\0423046.d
 Method tune_low.m
 Sample Name /VILV MG486-3
 Comment C9H8O2 mH149.0597 calibrant added CH3CN

Acquisition Date 23.04.2024 17:05:54
 Operator BDAL@DE
 Instrument / Ser# micrOTOF 10248

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Waste

