

Supplementary Information

BODNs as biocompatible brominating reagents: visible-light photocatalytic tyrosine modification under physiologically favorable conditions

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Instrumentation and Chemicals

¹H and ¹³C Nuclear magnetic resonance spectra were taken on a JEOL ECZ-600R (¹H, 600 MHz; ¹³C, 150 MHz) spectrometer using tetramethylsilane as an internal standard for ¹H NMR (δ = 0 ppm) and CDCl₃ as an internal standard for ¹³C NMR (δ = 77.0 ppm). ¹H NMR data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, sext = sextet, sept = septet, br = broad, m = multiplet), coupling constants (Hz), integration. Mass spectra were recorded on a Thermo Scientific Exactive (ESI, APCI) spectrometer (analyzer type: TOF). Infrared (IR) spectra were determined on an Affinity-1S spectrometer with QATR 10. Melting points were determined using a Stanford Research Systems MPA100. UV-Vis absorption spectra were determined using a SHIMADZU UV-1900i spectrophotometer. Fluorescence spectra were determined using a SHIMADZU RF-6000 fluorospectrophotometer. ASAHI SPECTRA CL-1503/CL-H1-525-7-1-B (525 nm), CL-H1-430-9-1-B (430 nm) were used for light irradiation. TLC analyses were performed by means of Merck Kieselgel 60 F₂₅₄ (0.25 mm) Plates. Visualization was accomplished with UV light (254 nm) and/or such as an aqueous alkaline KMnO₄ solution followed by heating. Flash column chromatography was carried out using Kanto Chemical silica gel (spherical, 40–50 μ m). Unless otherwise noted, commercially available reagents were used without purification.

Experimental Procedure

General procedure for photocatalytic tyrosine bromination

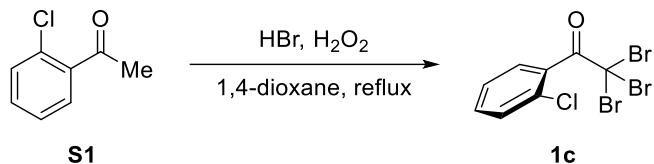
To a 10-mL round-bottom flask were sequentially added tyrosine derivative **2** (30 mg, 0.10 mmol), a solution of a photocatalyst (0.0010 mmol) in MeCN (0.50 mL), and H₂O (0.50 mL) without any special care for excluding air and moisture. The reaction mixture was stirred at 37 °C for 30 min. To the resulting solution was added brominating reagent **1** (0.12 mmol), and the mixture was irradiated with LEDs (525 nm). After being stirred for 3 h, the reaction was quenched with saturated aqueous Na₂S₂O₃ (1.5 mL) and H₂O (15 mL), and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. Purification of the crude product by flash silica gel column chromatography using hexane/EtOAc (v/v = 9:1) as an eluent gave the product **3**.

Procedure for photochemical tyrosine bromination

To a 10-mL round-bottom flask were sequentially added tyrosine derivative **2** (30 mg, 0.10 mmol), MeCN (0.50 mL), and H₂O (0.50 mL) without any special care for excluding air and moisture. The reaction mixture was stirred at 37 °C for 30 min. To the resulting solution was added brominating reagent **1e** (39 mg, 0.12 mmol), and the mixture was irradiated with LEDs (430 nm). After being stirred for 3 h, the reaction was quenched with saturated aqueous Na₂S₂O₃ (1.5 mL) and H₂O (15 mL), and the aqueous layer was extracted with CH₂Cl₂ (15 mL × 3). The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. Purification of the crude product by flash silica gel column chromatography using hexane/EtOAc (v/v = 2:1) as an eluent gave the product **3**.

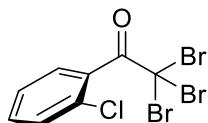
Brominating reagents **1a** and **1b** were commercially available.

Procedure for preparation of **1c**¹



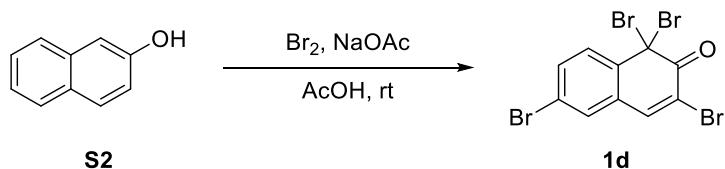
To a solution of 1-(2-chlorophenyl)ethan-1-one (**S1**, 0.77 g, 5.0 mmol) in 1,4-dioxane (14 mL) was added 48% aqueous HBr (13 mL, 0.12 mol), and the mixture was refluxed. To the mixture was slowly added a solution of 30% aqueous H₂O₂ (2.1 mL, 21 mmol) in 1,4-dioxane (1.4 mL). After being stirred for 2 h, the reaction mixture was cooled to ambient temperature, diluted with H₂O (30 mL), and extracted with CH₂Cl₂ (30 mL × 4). The combined organic layers were washed with H₂O (15 mL × 4), dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using hexane/EtOAc (v/v = 94:6) as an eluent gave **1c**.

2,2,2-Tribromo-1-(2-chlorophenyl)ethan-1-one (1c): CAS RN [296281-78-2].



Slightly yellow solid; 58% yield (1.1 g). ¹H NMR (CDCl₃) δ 7.88 (dd, *J* = 7.8, 1.2 Hz, 1H), 7.50 (dd, *J* = 8.4, 1.2 Hz, 1H), 7.45 (ddd, *J* = 8.4, 7.8, 1.2 Hz, 1H), 7.34 (ddd, *J* = 7.8, 7.8, 1.2 Hz, 1H). ¹³C NMR (CDCl₃) δ 185.7, 133.4, 132.1, 131.8, 130.1, 129.0, 126.3, 41.7. TLC: R_f 0.40 (hexane/EtOAc = 94:6).

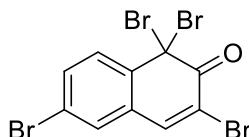
Procedure for preparation of **1d**²



To a solution of sodium acetate (11 g, 0.14 mol) and 2-naphthol (**S2**, 2.9 g, 20 mmol) in acetic acid (80 mL) was added bromine (26 g, 0.16 mol) portionwise over 10 min at 0 °C. The reaction mixture was stirred at ambient temperature for 2 h. Ice was added

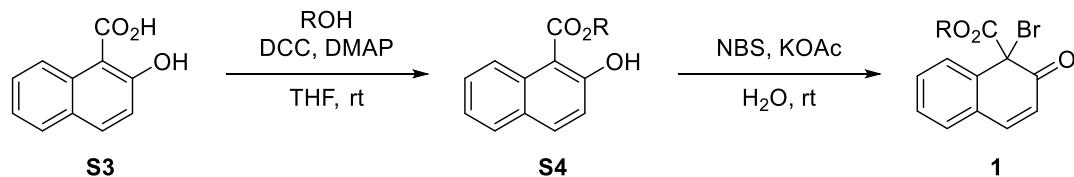
to the reaction mixture, forming slight yellow precipitates, which were collected by filtration with EtOAc and washed with cold water followed by EtOAc. A solution of the crude solid in CH₂Cl₂ was washed with H₂O, dried over Na₂SO₄, and concentrated in vacuo. Purification by recrystallization from toluene gave **1d**.

1,3,3,6-Tetrabromonaphthalen-2-(1H)-one (1d**): CAS RN [858023-30-0].**



Slightly yellow solid; 29% yield (2.6 g). ¹H NMR (CDCl₃) δ 7.99 (d, *J* = 8.8, 1H), 7.78 (s, 1H), 7.65 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.40 (d, *J* = 2.4 Hz, 1H). ¹³C NMR (CDCl₃) δ 179.7, 143.9, 140.3, 134.2, 133.4, 131.2, 127.4, 124.8, 119.2, 57.4. TLC: R_f 0.35 (hexane/EtOAc = 50:1).

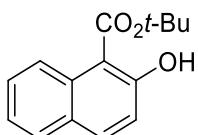
General procedure for preparation of BODN **1e–1g**



General procedure for preparation of **S4e–S4g³**

To a solution of 2-hydroxy-1-naphthoic acid (**S3**, 9.4 g, 50 mmol) in THF (50 mL) was sequentially added an alcohol (0.50 mol), 4-dimethylaminopyridine (0.61 g, 5.0 mmol), and *N,N'*-dicyclohexylcarbodiimide (15 g, 75 mmol), and the mixture was stirred at ambient temperature overnight. The mixture was filtered through a celite pad, which was washed with CH₂Cl₂, and the combined filtrate was concentrated in vacuo. Purification by flash silica gel column chromatography using hexane/EtOAc (v/v = 20/1) as an eluent gave **S4**.

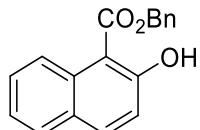
tert-Butyl 2-hydroxy-1-naphthoate (S4e**): CAS RN [2222802-20-0].**



Colorless oil; 99% yield (13 g). ¹H NMR (CDCl₃) δ 12.47 (s, 1H), 8.80 (dd, *J* = 8.8, 1.2 Hz, 1H), 7.86 (d, *J* = 9.2 Hz, 1H), 7.74 (d, *J* = 9.2 Hz, 1H), 7.53 (ddd, *J* = 8.8, 6.8, 1.6

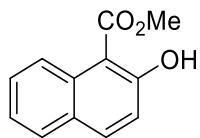
Hz, 1H), 7.35 (ddd, J = 8.4, 6.8, 1.2 Hz, 1H), 7.15 (dd, J = 8.4, 1.6 Hz, 1H), 1.74 (s, 9H). ^{13}C NMR (CDCl_3) δ 171.7, 164.1, 136.2, 132.0, 129.0, 128.7, 128.1, 125.2, 123.4, 119.4, 105.9, 84.1, 28.5. TLC: R_f 0.35 (hexane/EtOAc = 20:1).

Benzyl 2-hydroxy-1-naphthoate (S4f): CAS RN [86170-47-0].



White solid; 36% yield (5.0 g). ^1H NMR (CDCl_3) δ 12.26 (s, 1H), 8.79 (d, J = 9.0 Hz, 1H), 7.90 (d, J = 9.0 Hz, 1H), 7.75 (d, J = 9.0 Hz, 1H), 7.54–7.49 (m, 3H), 7.44–7.34 (m, 4H), 7.17 (d, J = 9.0 Hz, 1H), 5.58 (s, 2H). ^{13}C NMR (CDCl_3) δ 172.2, 164.6, 137.0, 135.2, 131.8, 129.1, 128.8, 128.65, 128.57, 128.5, 128.4, 125.3, 123.6, 119.3, 104.6, 67.5. TLC: R_f 0.35 (hexane/EtOAc = 20:1).

Methyl 2-hydroxy-1-naphthoate (S4g): CAS RN [947-65-9].

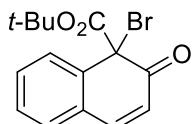


White solid; 43% yield (4.3 g). ^1H NMR (CDCl_3) δ 12.27 (s, 1H), 8.75 (d, J = 8.5 Hz, 1H), 7.90 (d, J = 9.0 Hz, 1H), 7.76 (d, J = 8.0 Hz, 1H), 7.56 (dd, J = 8.5, 8.0 Hz, 1H), 7.38 (dd, J = 8.0, 8.0 Hz, 1H), 7.17 (d, J = 9.0 Hz, 1H), 4.12 (s, 3H). ^{13}C NMR (CDCl_3) δ 172.8, 164.4, 136.9, 131.7, 129.1, 128.6, 128.5, 125.3, 123.6, 119.3, 104.7, 52.4. TLC: R_f 0.45 (hexane/EtOAc = 20:1).

General procedure for preparation of **1e–1g**⁴

To a solution of **S4** (1.0 equiv) and potassium acetate (1.5 equiv) in H_2O (0.13 M) was added *N*-bromosuccinimide (1.2 equiv), and the mixture was stirred at ambient temperature for 1 h. The mixture was extracted with CH_2Cl_2 (0.15 L \times 3), and the organic layers were washed with H_2O , dried over Na_2SO_4 , and concentrated in vacuo. Purification by flash silica gel column chromatography using hexane or hexane/EtOAc (v/v = 5:1) as an eluent gave **1**.

tert-Butyl 1-bromo-2-oxo-1,2-dihydroronaphthalene-1-carboxylate (1e).



Yellow solid; 77% yield (12 g). ^1H NMR (CDCl_3) δ 7.50–7.35 (m, 5H), 6.28 (d, $J = 9.5$ Hz, 1H), 1.40 (s, 9H). ^{13}C NMR (CDCl_3) δ 190.3, 164.6, 144.7, 138.7, 130.8, 129.9, 129.5, 128.8, 127.9, 123.3, 84.9, 61.2, 27.6. TLC: R_f 0.33 (hexane/EtOAc = 5:1). Mp. 60.5–61.5 °C. IR (neat): 2983, 1750, 1662, 1367, 1239, 1146, 994, 745, 620, 514 cm^{-1} . HRMS (ESI) Calcd for $\text{C}_{15}\text{H}_{15}\text{BrO}_3\text{Na}$: $[\text{M}+\text{Na}]^+$, 345.0097, 347.0076. Found: m/z 345.0097, 347.0072.

Benzyl 1-bromo-2-oxo-1,2-dihydroronaphthalene-1-carboxylate (1f).



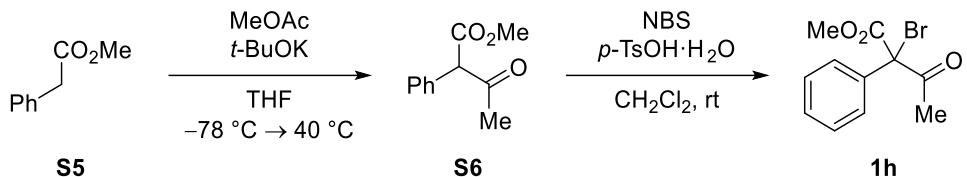
Yellow solid; 84% yield (5.4 g). ^1H NMR (CDCl_3) δ 7.46 (d, $J = 10.0$ Hz, 1H), 7.40–7.34 (m, 4H), 7.30–7.28 (m, 3H), 7.22–7.19 (m, 2H), 6.30 (d, $J = 10.0$ Hz, 1H), 5.25 (s, 2H). ^{13}C NMR (CDCl_3) δ 189.7, 165.9, 144.9, 137.9, 134.7, 130.8, 130.0, 129.8, 129.1, 128.5, 128.4, 128.1, 127.9, 123.2, 69.1, 60.3. TLC: R_f 0.25 (hexane/EtOAc = 5:1). Mp. 87.5–88.5 °C. IR (neat): 2928, 1753, 1659, 1229, 1204, 945, 752, 698, 623, 500 cm^{-1} . HRMS (ESI) Calcd for $\text{C}_{18}\text{H}_{13}\text{BrO}_3\text{Na}$: $[\text{M}+\text{Na}]^+$, 378.9940, 380.9920. Found: m/z 378.9945, 380.9912.

Methyl 1-bromo-2-oxo-1,2-dihydroronaphthalene-1-carboxylate (1g): CAS RN [1799904-02-1].



Yellow solid; 76% yield (4.8 g). ^1H NMR (CDCl_3) δ 7.49–7.37 (m, 5H), 6.31 (d, $J = 10.0$ Hz, 1H), 3.81 (s, 3H). ^{13}C NMR (CDCl_3) δ 189.8, 166.6, 144.9, 138.0, 131.0, 130.1, 129.9, 129.1, 127.9, 123.2, 60.1, 54.4. TLC: R_f 0.25 (hexane/EtOAc = 5:1).

Procedure for preparation of **1h**



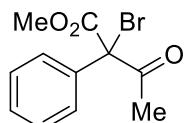
Procedure for preparation of **S6⁵**

To a solution of methyl phenylacetate (**S5**, 3.8 g, 25 mmol) and methyl acetate (7.3 mL, 75 mmol) in THF (25 mL) were slowly added a suspension of *t*-BuOK (4.2 g, 38 mmol) in THF (25 mL) at -78 $^\circ\text{C}$ under argon atmosphere. After being stirred for 4 h, the reaction mixture was warmed to 40 $^\circ\text{C}$. After being stirred for additional 11 h, the reaction was quenched with 1.0 M HCl (ca. 50 mL). The aqueous layer was extracted with EtOAc (50 mL \times 3), and the combined organic layers were washed with brine (50 mL \times 2), dried over Na₂SO₄, and concentrated in vacuo. Purification by distillation gave methyl 2-phenylacetate (**S6**) as a colorless oil in 74% yield (2.9 g).

Procedure for preparation of **1h⁶**

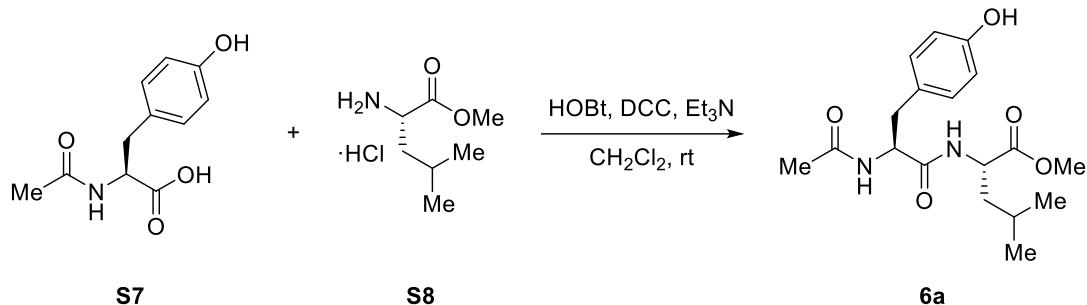
To a solution of **S6** (1.5 g, 8.0 mmol) in CH₂Cl₂ (40 mL) was sequentially added *p*-toluenesulfonic acid monohydrate (0.30 g, 1.6 mmol) and *N*-bromosuccinimide (1.6 g, 8.8 mmol), and the mixture was stirred at ambient temperature for 2 h. H₂O (30 mL) was added to the reaction mixture, and the aqueous layer was extracted with CH₂Cl₂ (40 mL \times 3). The combined organic layers were washed with H₂O (40 mL \times 2), dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using hexane/EtOAc (v/v = 10:1) as an eluent gave **1h**.

Methyl 2-bromo-3-oxo-2-phenylbutanoate (1h**).**



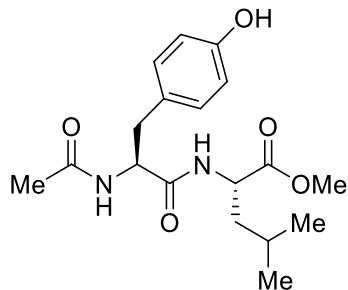
Colorless oil; 67% yield (1.8 g). ¹H NMR (CDCl₃) δ 7.51–7.49 (m, 2H), 7.41–7.37 (m, 3H), 3.86 (s, 3H), 2.40 (s, 3H). ¹³C NMR (CDCl₃) δ 196.8, 167.8, 134.1, 129.2, 128.6, 128.5, 70.9, 54.0, 26.3. TLC: R_f 0.35 (hexane/EtOAc = 10:1). IR (neat): 2955, 1724, 1433, 1356, 1240, 1180, 1030, 739, 692, 556 cm⁻¹. HRMS (ESI) Calcd for C₁₁H₁₁BrO₃Na: [M+Na]⁺, 292.9784, 294.9769. Found: *m/z* 292.9777, 294.9756.

Procedure for preparation of 6a



To a 200-mL round-bottom flask were sequentially added *N*-acetyl-L-tyrosine **S7** (2.2 g, 10 mmol), L-leucine methyl ester hydrochloride **S8** (1.8 g, 10 mmol), DMF (50 mL), 1-hydroxybenzotriazole (1.4 g, 10 mmol), *N,N'*-dicyclohexylcarbodiimide (2.3 g, 11 mmol), and triethylamine (3.5 mL, 25 mmol). After being stirred at 25 °C overnight, the reaction was quenched with H₂O (30 mL). After white precipitates were removed by filtration, the aqueous layer was extracted with CH₂Cl₂ (50 mL × 3). The combined organic layers were washed with H₂O and brine, dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using CH₂Cl₂/MeOH (v/v = 10:1) as an eluent gave **6a**.

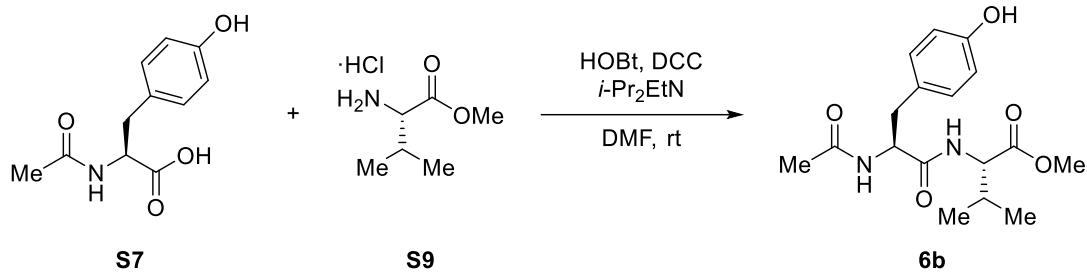
N-Acetyl-L-tyrosyl-L-leucin methyl ester (6a): CAS RN [33049-03-5].



White solid; 47% yield (1.6 g).

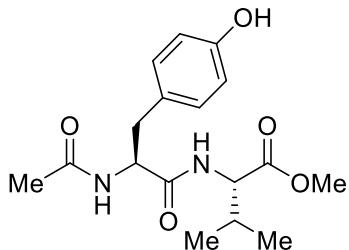
¹H NMR (CD₃OD) δ 7.03 (d, *J* = 8.4 Hz, 2H), 6.66 (d, *J* = 8.4 Hz, 2H), 4.53 (dd, *J* = 8.4, 6.0 Hz, 1H), 4.42 (dd, *J* = 9.6, 5.4 Hz, 1H), 3.64 (s, 3H), 2.98 (dd, *J* = 13.8, 6.0 Hz, 1H), 2.73 (dd, *J* = 13.8, 8.4 Hz, 1H), 1.87 (s, 3H), 1.66–1.52 (m, 3H), 0.91 (d, *J* = 6.6 Hz, 3H), 0.87 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (CD₃OD) δ 172.9, 172.6, 155.9, 130.2, 129.7, 127.6, 114.8, 54.9, 54.7, 50.8, 48.2, 40.2, 24.4, 21.9, 20.9, 20.5. TLC: R_f 0.58 (CH₂Cl₂/MeOH = 10:1).

Procedure for preparation of 6b



To a 100-mL round-bottom flask were sequentially added *N*-acetyl-L-tyrosine **S7** (1.6 g, 6.0 mmol), L-valine methyl ester hydrochloride **S9** (1.0 g, 6.0 mmol), DMF (20 mL), 1-hydroxybenzotriazole (1.0 g, 6.6 mmol), *N,N*'-dicyclohexylcarbodiimide (1.2 g, 6.0 mmol), and *N,N*-diisopropylethylamine (2.2 mL, 13 mmol). After being stirred at 25 °C overnight, the reaction was quenched with H₂O (30 mL). After white precipitates were removed by filtration, the aqueous layer was extracted with EtOAc (30 mL × 3). The combined organic layers were washed with H₂O and brine, dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using CH₂Cl₂/MeOH (v/v = 10:1) as an eluent gave **6b**.

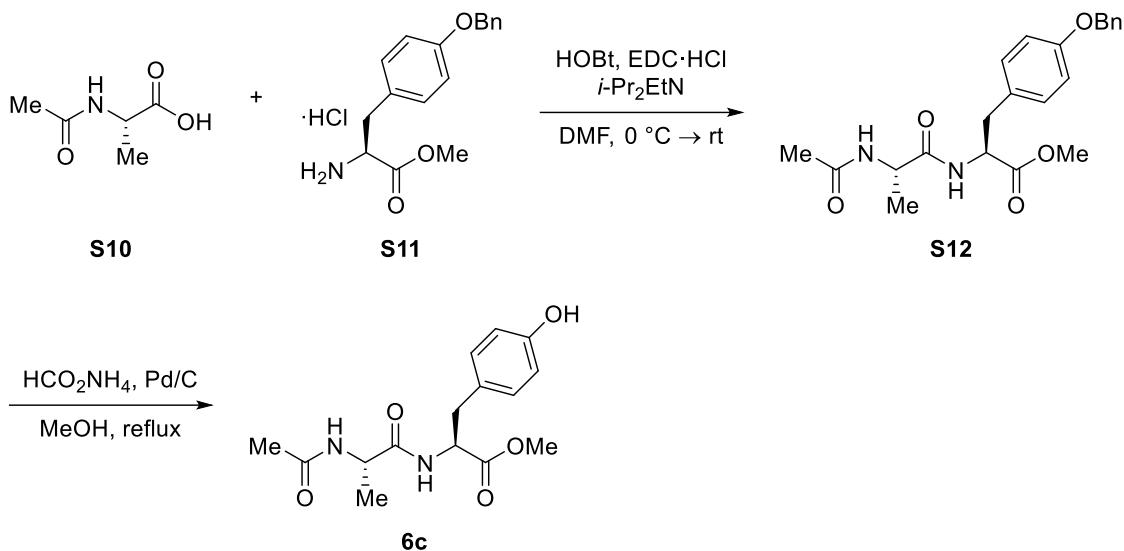
N-Acetyl-L-tyrosyl-L-valine methyl ester (6b): CAS RN [573968-42-0].



White solid; 56% yield (1.1 g).

¹H NMR (CDCl₃) δ 7.04 (d, *J* = 8.4 Hz, 2H), 6.72 (d, *J* = 8.4 Hz, 2H), 6.62 (s, 1H), 6.39 (d, *J* = 8.4 Hz, 1H), 6.35 (d, *J* = 7.8 Hz, 1H), 4.68 (dt, *J* = 7.8, 7.2 Hz, 1H), 4.42 (dd, *J* = 8.4, 5.4 Hz, 1H), 3.71 (s, 3H), 2.97 (d, *J* = 7.2 Hz, 2H), 2.09 (m, 1H), 1.98 (s, 3H), 0.87 (d, *J* = 6.6 Hz, 3H), 0.84 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (CDCl₃) δ 171.8, 171.2, 170.5, 155.3, 130.5, 127.8, 115.7, 57.6, 54.9, 52.3, 37.8, 31.2, 23.2, 18.9, 17.9. TLC: R_f 0.38 (CH₂Cl₂/MeOH = 10:1).

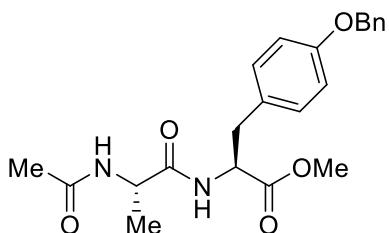
Procedure for preparation of 6c



Procedure for preparation of S12

To a 50-mL round-bottom flask were sequentially added *N*-acetyl-L-alanine **S10** (0.60 g, 4.6 mmol), *O*-(phenylmethyl)-L-tyrosine methyl ester hydrochloride **S11** (1.5 g, 4.6 mmol), DMF (23 mL), and *N,N*-diisopropylethylamine (2.4 mL, 14 mmol) at 25 °C. Subsequently, 1-hydroxybenzotriazole (0.74 g, 5.5 mmol), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (1.1 g, 5.5 mmol) were added at 0 °C. After being stirred at 25 °C overnight, the reaction was quenched with H₂O (30 mL), and the aqueous layer was extracted with EtOAc (30 mL × 4). The combined organic layers were washed with 1.0 M aqueous HCl, saturated aqueous NaHCO₃, and brine, dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using hexane/EtOAc (v/v = 1:3) as an eluent gave **S12**.

Methyl (S)-2-((S)-2-acetamidopropanamido)-3-(4-(benzyloxy)phenyl)propanoate (S12): CAS RN [18828-16-5].



White solid; 56% yield (1.1 g).

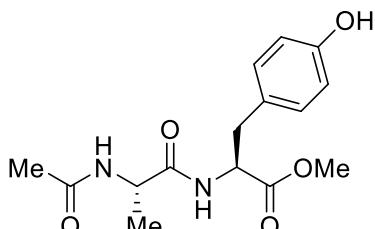
¹H NMR (CDCl₃) δ 7.42 (d, *J* = 7.2 Hz, 2H), 7.38 (dd, *J* = 7.2, 7.2 Hz, 2H), 7.33 (d, *J* = 7.2 Hz, 1H), 7.02 (d, *J* = 8.4 Hz, 2H), 6.89 (d, *J* = 8.4 Hz, 2H), 6.43 (m, 1H), 5.99 (m,

1H), 5.03 (s, 2H), 4.79 (m, 1H), 4.45 (dq, $J = 7.2, 7.2$ Hz, 1H), 3.73 (s, 3H), 3.09 (dd, $J = 14.4, 6.0$ Hz, 1H), 3.01 (dd, $J = 14.4, 6.0$ Hz, 1H), 1.96 (s, 3H), 1.33 (d, $J = 7.2$ Hz, 3H). ^{13}C NMR (CDCl_3) δ 171.83, 171.78, 169.9, 158.0, 137.0, 130.4, 128.7, 128.1, 127.9, 127.6, 115.0, 70.0, 53.4, 52.5, 48.8, 37.0, 23.2, 18.2. TLC: R_f 0.29 (hexane/EtOAc = 1:3).

Procedure for preparation of **6c**

To a 100 mL round-bottom flask were sequentially added palladium-activated carbon (0.13 g, 0.11 mmol), MeOH (4.0 mL), **S12** (0.88 g, 2.2 mmol), and ammonium formate (2.8 g, 44 mmol) under argon atmosphere. After being refluxed overnight, the mixture was filtered through a celite pad and concentrated in vacuo. H_2O (25 mL) was added, and the aqueous layer was extracted with EtOAc (30 mL \times 3). The combined organic layers were washed with H_2O and brine, dried over Na_2SO_4 , and concentrated in vacuo. Purification by flash silica gel column chromatography using EtOAc as an eluent gave **6c**.

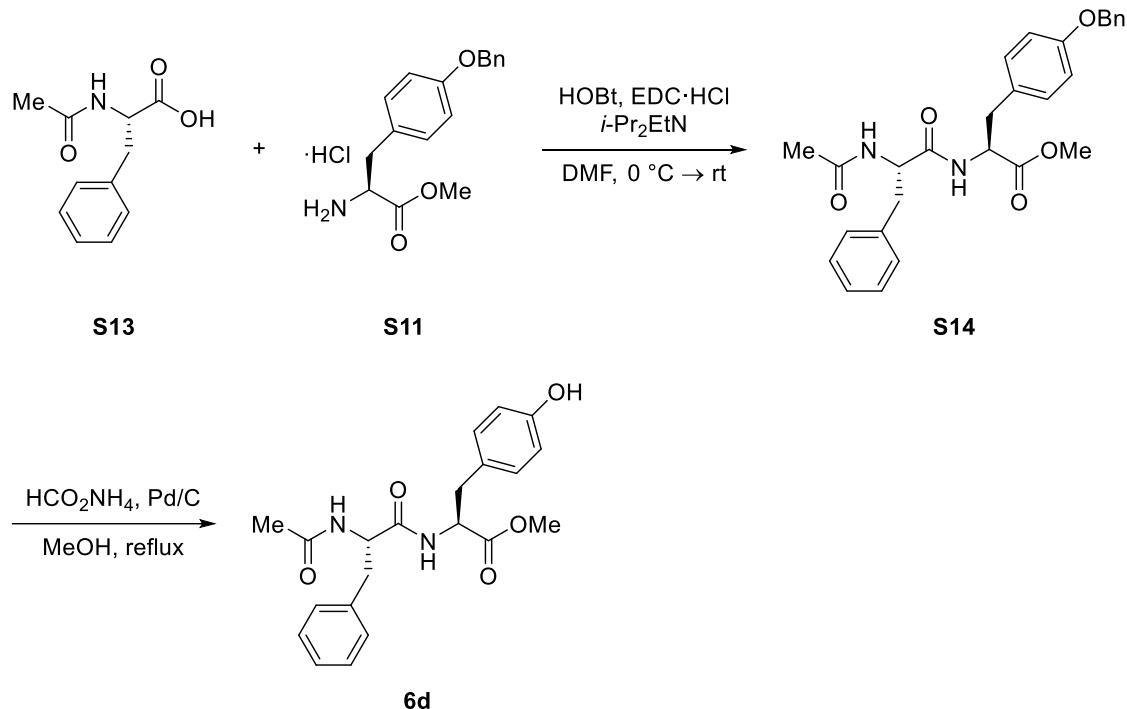
N-Acetyl-L-alanyl-L-tyrosine methyl ester (6c): CAS RN [57328-71-9].



White solid; 50% yield (0.34 g).

^1H NMR (CDCl_3) δ 8.01 (s, 1H), 6.91 (d, $J = 8.4$ Hz, 1H), 6.91 (d, $J = 8.4$ Hz, 2H), 6.69 (d, $J = 8.4$ Hz, 2H), 6.67 (d, $J = 7.8$ Hz, 1H), 4.79 (m, 1H), 4.51 (dq, $J = 7.8, 7.2$ Hz, 1H), 3.72 (s, 3H), 3.05 (dd, $J = 13.8, 4.8$ Hz, 1H), 2.93 (dd, $J = 13.8, 6.0$ Hz), 1.93 (s, 3H), 1.30 (d, $J = 7.2$ Hz, 3H). ^{13}C NMR (CDCl_3) δ 172.4, 171.9, 170.9, 155.7, 130.4, 126.8, 115.7, 53.6, 52.5, 48.8, 37.0, 23.0, 18.2. TLC: R_f 0.17 (EtOAc).

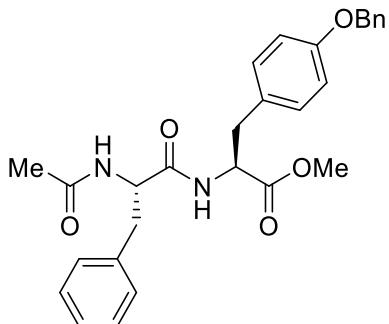
Procedure for preparation of 6d



Procedure for preparation of S14

To a 50-mL round-bottom flask were sequentially added *N*-acetyl-L-phenylalanine **S13** (1.6 g, 5.0 mmol), *O*-(phenylmethyl)-L-tyrosine methyl ester hydrochloride **S11** (1.6 g, 5.0 mmol), DMF (15 mL), and *N,N*-diisopropylethylamine (1.26 mL, 7.5 mmol) at 25 °C. Subsequently, 1-hydroxybenzotriazole (0.81 g, 6.0 mmol), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (1.2 g, 6.0 mmol) were added at 0 °C. After being stirred at 25 °C overnight, the reaction was quenched with H₂O (45 mL), and the aqueous layer was extracted with EtOAc (30 mL × 4). The combined organic layers were washed with 1.0 M aqueous HCl, saturated aqueous NaHCO₃, and brine, dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using hexane/EtOAc (v/v = 1:3) as an eluent gave **S14**.

Methyl (S)-2-((S)-2-acetamido-3-phenylpropanamido)-3-(4-(benzyloxy)phenyl)propanoate (S14).



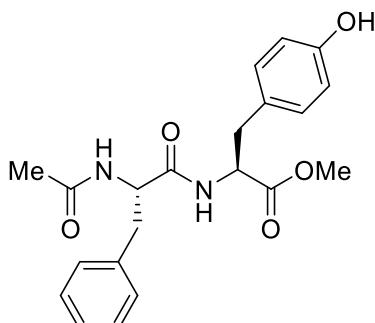
White solid; 86% yield (2.0 g).

¹H NMR (CD₃OD) δ 7.39–7.15 (m, 10H), 7.06 (d, *J* = 8.4 Hz, 2H), 6.87 (d, *J* = 8.4 Hz, 2H), 5.01 (s, 2H), 4.57–4.56 (m, 2H), 3.62 (s, 3H), 3.06–3.02 (m, 2H), 2.89 (dd, *J* = 13.8, 8.6 Hz, 1H), 2.77 (dd, *J* = 13.8, 8.6 Hz, 1H), 1.82 (s, 3H). ¹³C NMR (CD₃OD) δ 173.4, 173.1, 173.0, 159.2, 138.8, 138.4, 131.4, 130.3, 130.1, 129.5, 129.4, 128.8, 128.5, 127.7, 115.9, 70.9, 55.8, 55.3, 52.7, 38.8, 37.6, 22.3. Mp. 172.3–172.7 °C. TLC: R_f 0.68 (hexane/EtOAc = 1:3). IR (neat): 1738, 1635, 1512, 1454, 1246, 1176, 1024, 741, 696, 517 cm⁻¹. HRMS (ESI) Calcd for C₂₈H₃₀N₂O₅Na: [M+Na]⁺, 497.20469. Found: *m/z* 497.20502.

Procedure for preparation of 6d

To a 100 mL round-bottom flask were sequentially added palladium-activated carbon (0.20 g, 0.19 mmol), MeOH (10 mL), **S14** (2.0 g, 4.3 mmol), and ammonium formate (6.3 g, 0.10 mol) under argon atmosphere. After being refluxed overnight, the mixture was filtered through a celite pad and concentrated in vacuo. H₂O (50 mL) was added, and the aqueous layer was extracted with EtOAc (30 mL × 3). The combined organic layers were washed with H₂O and brine, dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using Hexane/EtOAc (v/v = 1:3) as an eluent gave **6d**.

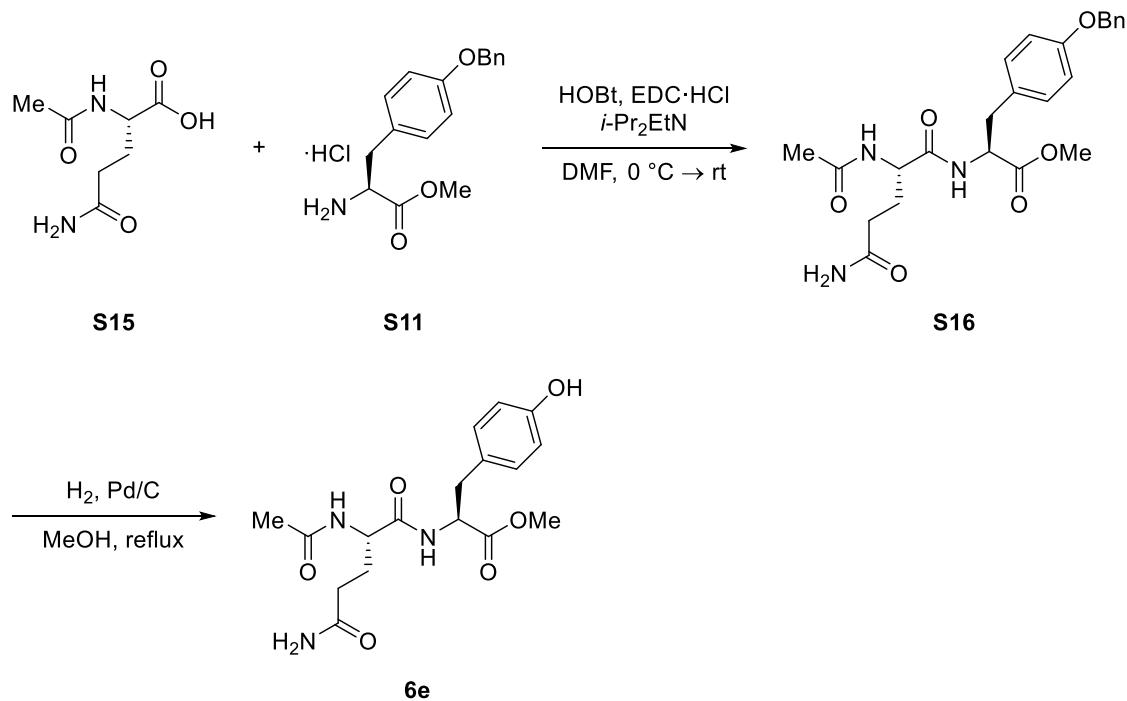
N-Acetyl-L-phenylalanyl-L-tyrosine methyl ester (6d): CAS RN [15852-46-7].



White solid; 85% yield (1.4 g).

¹H NMR (CD_3OD) δ 7.25–7.15 (m, 5H), 6.96 (d, $J = 6.6$ Hz, 2H), 6.66 (d, $J = 6.6$ Hz, 2H), 4.62–4.55 (m, 2H), 3.63 (s, 3H), 3.07–2.98 (m, 2H), 2.88–2.75 (m, 2H), 1.84 (s, 3H).
¹³C NMR (CDCl_3) δ 171.4, 170.73, 170.66, 155.5, 136.1, 130.3, 129.2, 128.6, 127.0, 126.7, 115.5, 54.3, 53.5, 52.4, 38.1, 37.0, 22.9. TLC: R_f 0.48 (Hexane/EtOAc = 1:3).

Procedure for preparation of 6e

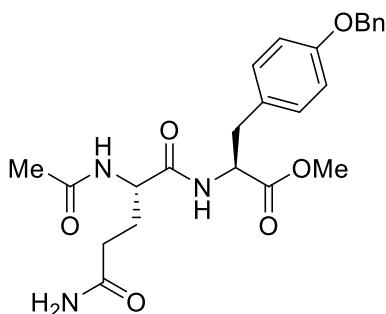


Procedure for preparation of **S16**

To a 100-mL round-bottom flask were sequentially added *N*-acetyl-L-glutamine **S15** (1.3 g, 7.0 mmol), *O*-(phenylmethyl)-L-tyrosine methyl ester hydrochloride **S11** (2.2 g, 7.0 mmol), DMF (35 mL), and *N,N*-diisopropylethylamine (3.7 mL, 22 mmol) at 25 °C.

Subsequently, 1-hydroxybenzotriazole (1.1 g, 8.4 mmol), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (1.6 g, 8.4 mmol) were added at 0 °C. After being stirred at 25 °C overnight, the reaction was quenched with H₂O (45 mL), and the aqueous layer was extracted with EtOAc (40 mL × 5). The combined organic layers were washed with 1.0 M aqueous HCl, saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, and concentrated in vacuo. Purification by flash silica gel column chromatography using CH₂Cl₂/MeOH (v/v = 10:1) as an eluent gave **S16**.

Methyl (S)-2-((S)-2-acetamido-5-amino-5-oxopentanamido)-3-(4-(benzyloxy)phenyl)propanoate (S16).



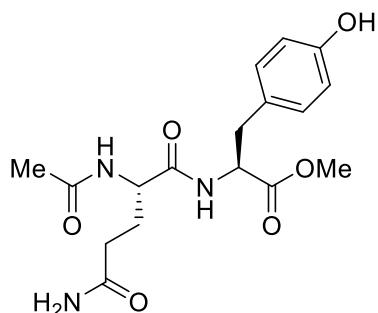
White solid; 16% yield (0.50 g).

¹H NMR (DMSO-*d*₆) δ 8.27 (d, *J* = 6.0 Hz, 1H), 7.92 (d, *J* = 8.4 Hz, 1H), 7.41–7.27 (m, 5H), 7.23 (s, 1H), 7.09 (d, *J* = 6.0 Hz, 2H), 6.87 (d, *J* = 6.0 Hz, 2H), 6.73 (s, 1H), 5.02 (s, 2H), 4.35 (m, 1H), 4.22 (dd, *J* = 14.1, 8.4 Hz, 1H), 3.54 (s, 3H), 2.90 (dd, *J* = 13.2, 6.0 Hz, 1H), 2.82 (dd, *J* = 13.2, 9.0 Hz, 1H), 2.02 (dd, *J* = 15.0, 8.4 Hz, 2H), 1.81 (m, 1H), 1.78 (s, 3H), 1.59 (m, 1H). ¹³C NMR (DMSO-*d*₆) δ 174.2, 172.4, 172.2, 169.7, 157.6, 138.7, 130.8, 130.6, 129.6, 129.0, 128.2, 114.9, 69.6, 54.3, 52.2, 40.5, 36.3, 31.9, 23.1, 22.9. Mp. 234.0–235.0 °C. TLC: R_f 0.23 (CH₂Cl₂/MeOH = 10:1). IR (neat): 3283, 1659, 1638, 1545, 1514, 1246, 1219, 1178, 729, 598 cm⁻¹. HRMS (ESI) Calcd for C₂₄H₂₉N₃O₆Na: [M+Na]⁺, 478.19486. Found: *m/z* 478.19448.

Procedure for preparation of **6e**

To a 50 mL round-bottom flask were sequentially added palladium-activated carbon (0.17 g, 0.17 mmol), dry MeOH (20 mL), and **S16** (0.50 g, 1.1 mmol) under argon atmosphere, which was then replaced with hydrogen gas, and the mixture was stirred at 60 °C. After being refluxed for 3 h, the solution was filtered through a celite pad and concentrated in vacuo. Purification by flash silica gel column chromatography using CH₂Cl₂/MeOH (v/v = 10:1) as an eluent gave **6e**.

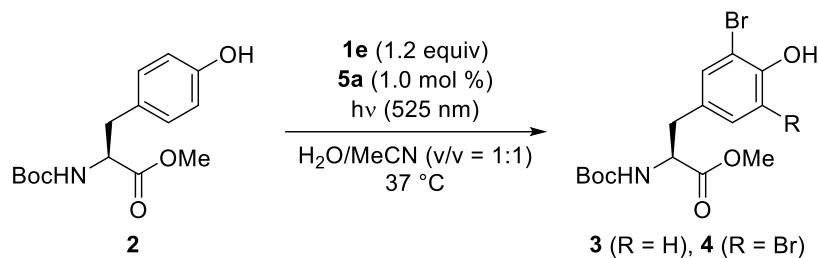
N-Acetyl-L-glutaminyl-L-tyrosine methyl ester (6e).



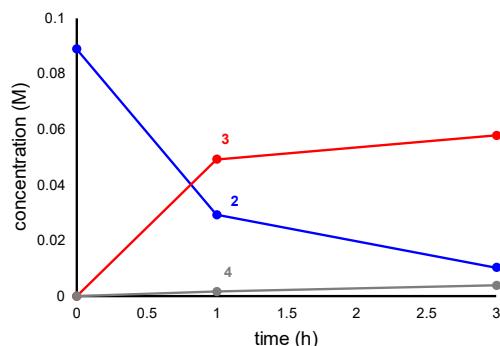
White solid; 94% yield (0.38 g).

^1H NMR (DMSO- d_6) δ 9.20 (s, 1H), 8.23 (d, J = 7.2 Hz, 1H), 7.92 (d, J = 7.8 Hz, 1H), 7.22 (s, 1H), 6.94 (d, J = 7.2 Hz, 2H), 6.72 (s, 1H), 6.60 (d, J = 7.2 Hz, 2H), 4.31 (m, 1H), 4.22 (m, 1H), 3.54 (s, 3H), 2.84 (dd, J = 13.8, 6.0 Hz, 1H), 2.77 (dd, J = 13.8, 7.8 Hz, 1H), 2.03 (m, 2H), 1.82 (m, 1H), 1.78 (s, 3H), 1.60 (m, 1H). ^{13}C NMR (DMSO- d_6) δ 173.7, 171.9, 171.6, 169.1, 156.0, 130.2, 127.0, 115.0, 53.9, 52.0, 51.7, 35.9, 31.3, 27.9, 22.5. Mp. 213.8–214.3 °C. TLC: R_f 0.080 (CH₂Cl₂/MeOH = 10:1). IR (neat): 1724, 1662, 1630, 1544, 1516, 1236, 1223, 604, 592, 530 cm⁻¹. HRMS (ESI) Calcd for C₁₇H₂₃N₃O₆Na: [M+Na]⁺, 388.14791. Found: *m/z* 388.14710.

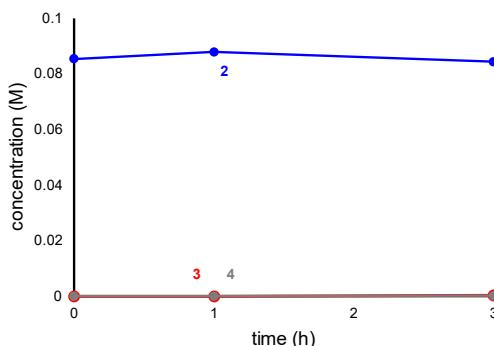
Scheme S1. Photocatalytic Tyrosine Bromination with Green Light



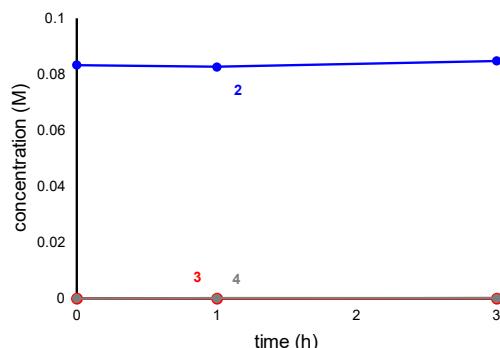
(a) with light and **5a**, 0.10 M



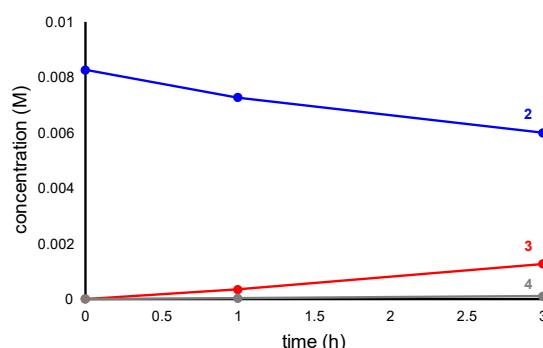
(b) with light, without **5a**, 0.10 M



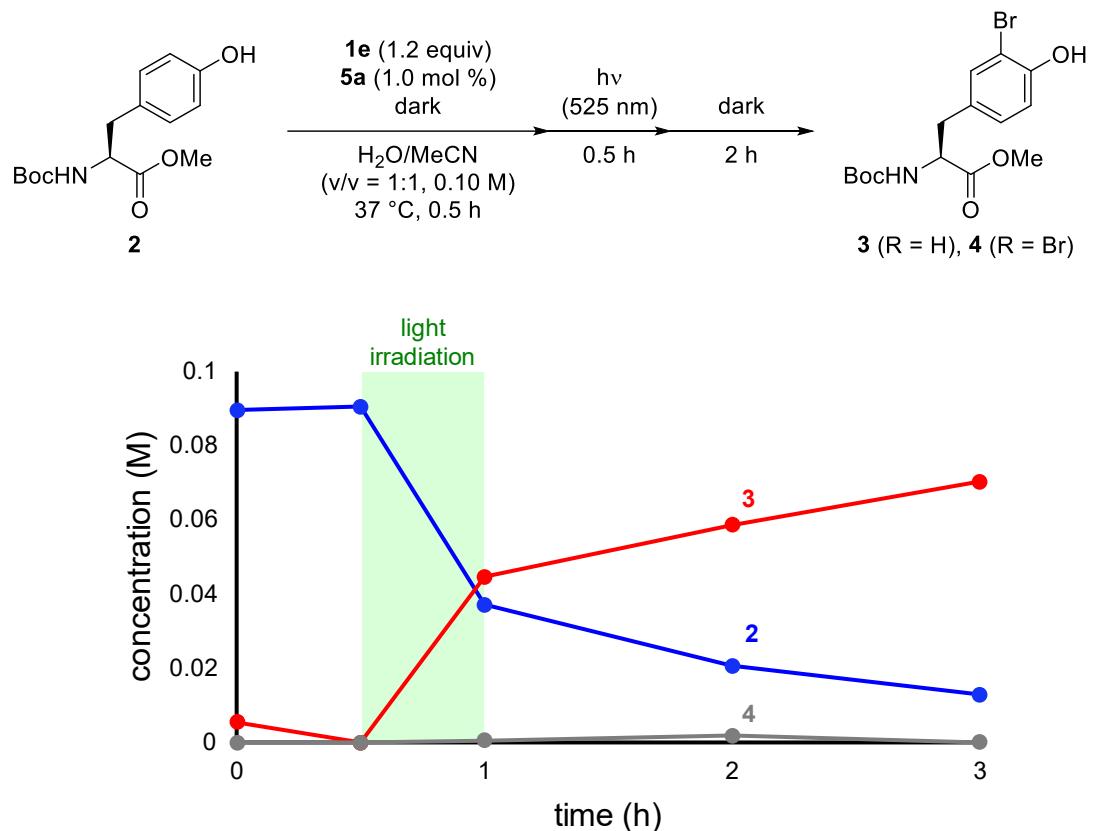
(c) without light, with **5a**, 0.10 M



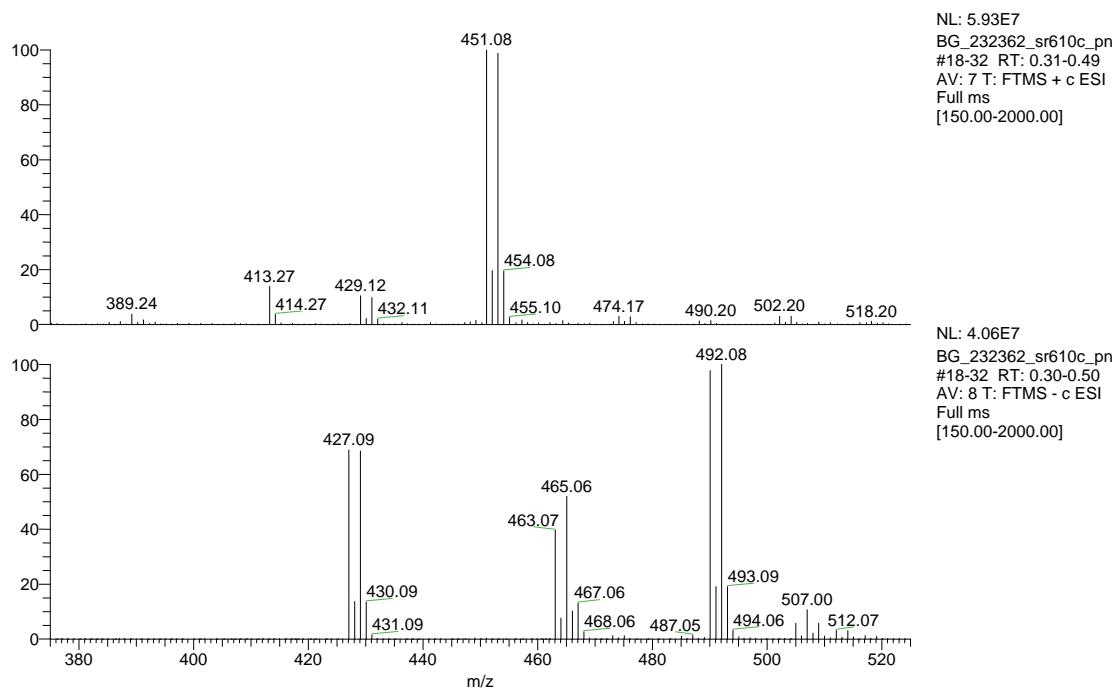
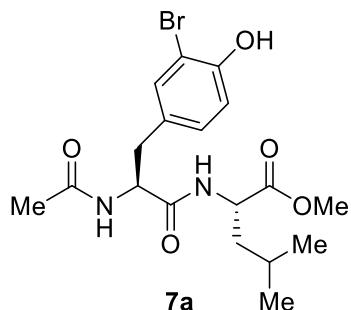
(d) with light and **5a**, 0.010 M

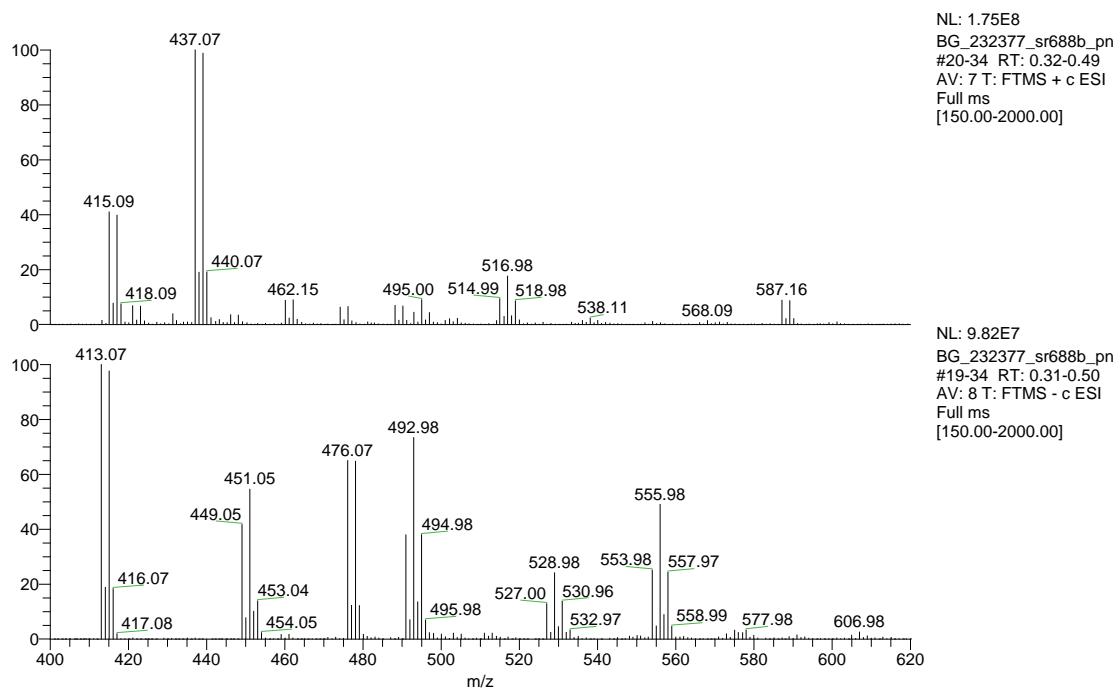
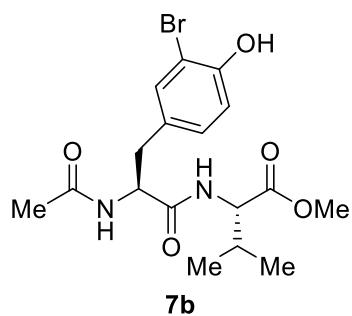


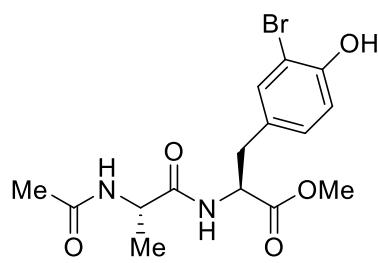
Scheme S2. Temporal Control of Tyrosine Bromination with Light



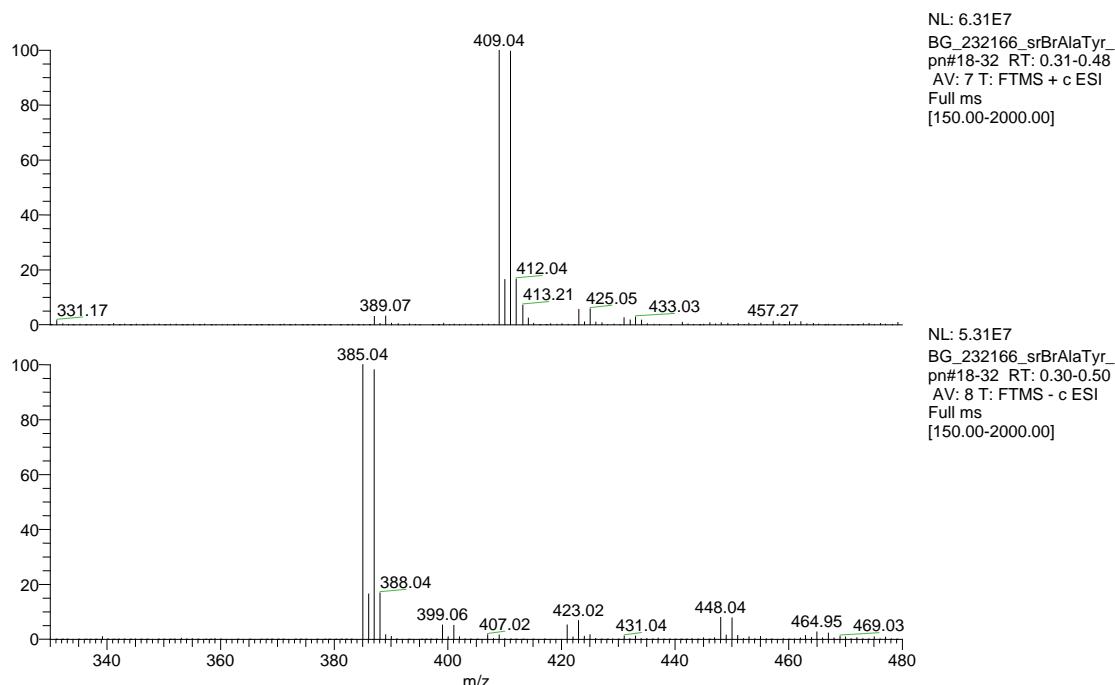
Scheme S3. Mass Spectra Analyses of **7**

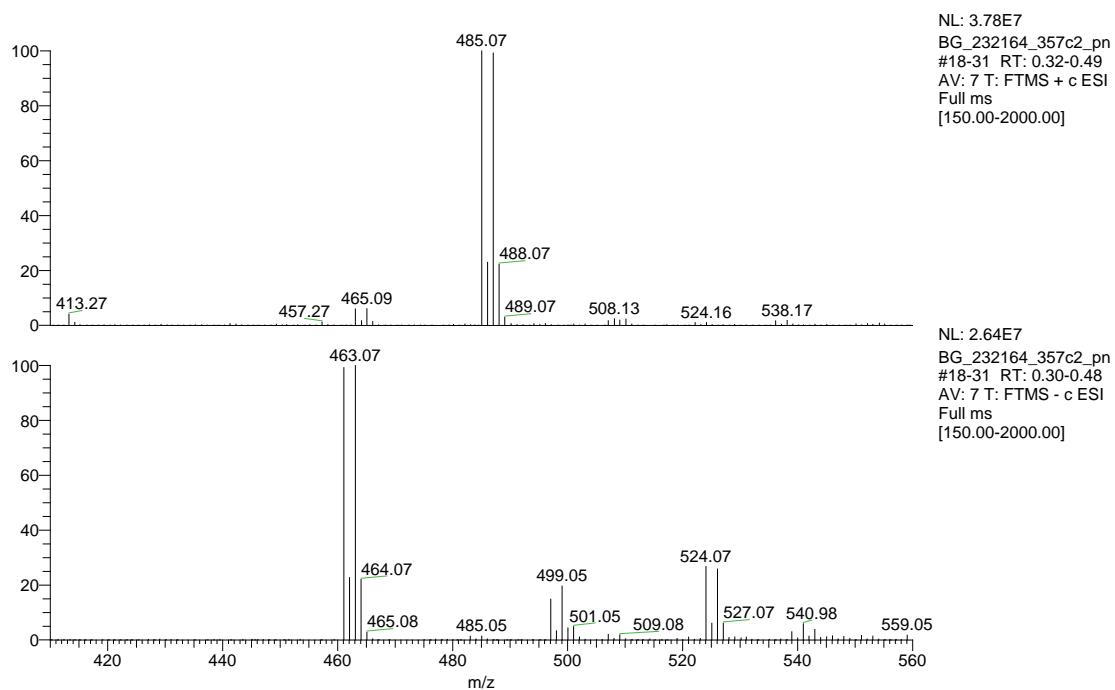
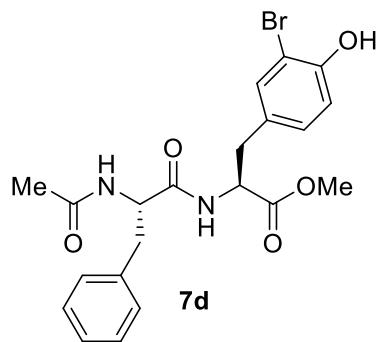


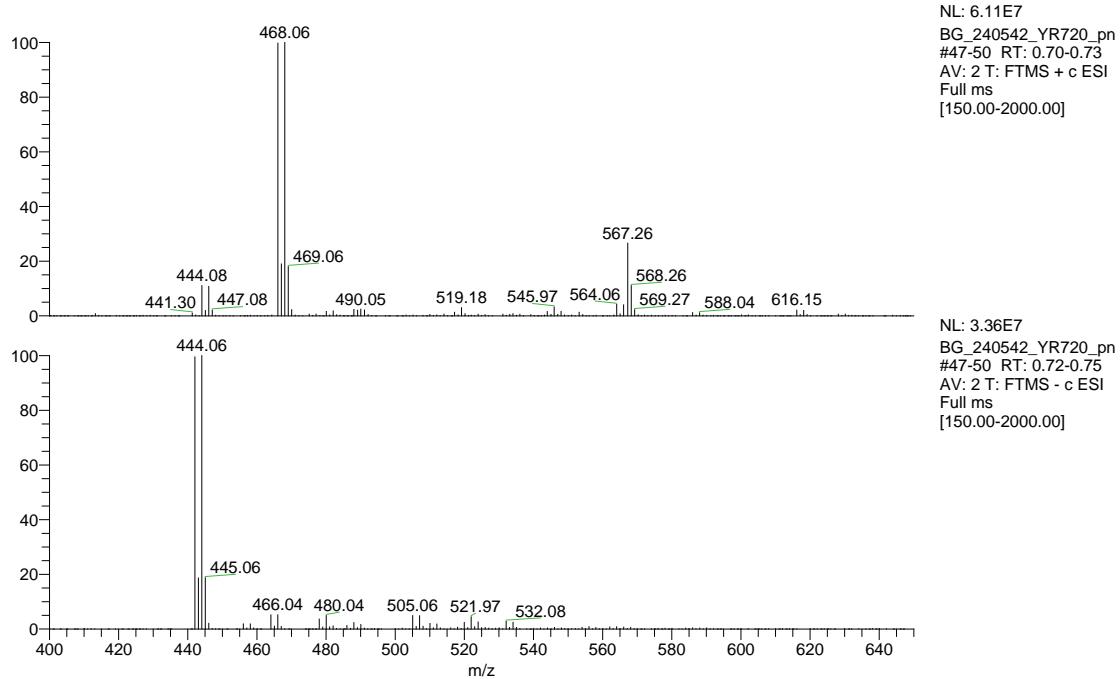
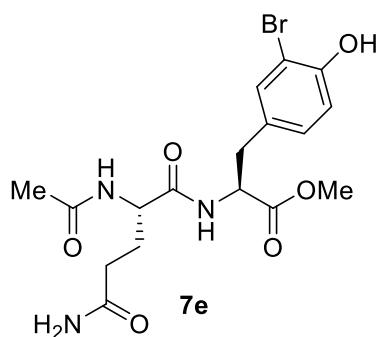




7c





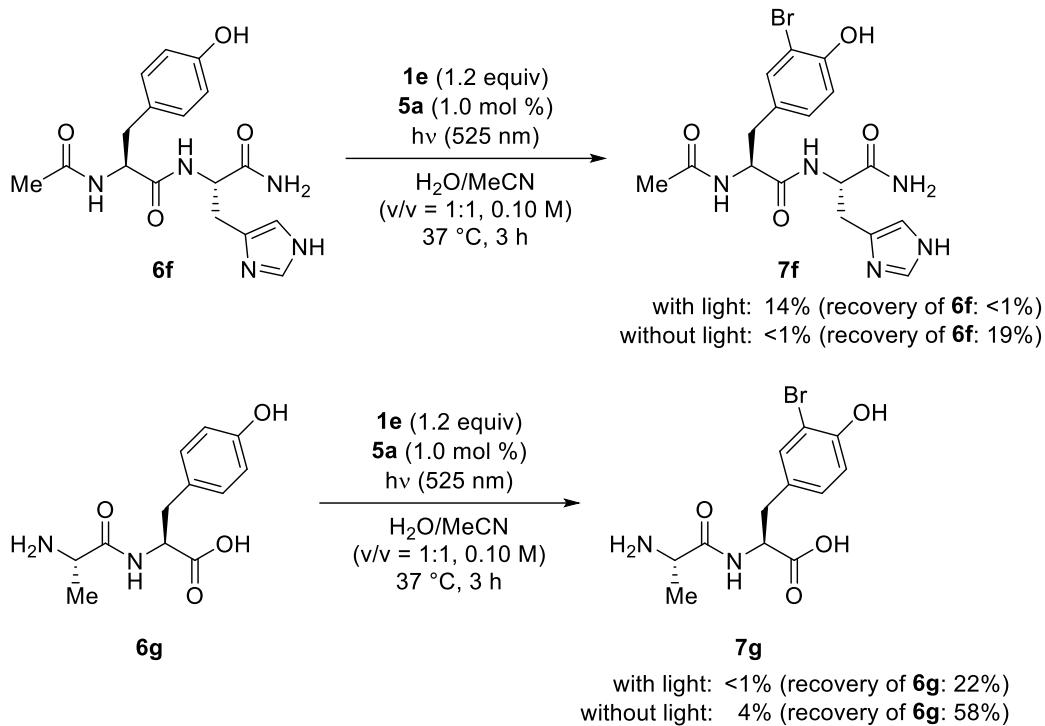


Scheme S4. Bromination of Aromatic Amino Acids^a

entry	amino acid	yields without light [S17 (recovery), S18] (%)	yields with light [S17 (recovery), S18] (%)		
		S17	S18	S17	S18
1	Phe	90	<1	92	<1
2	Trp	19	<1	<1	<1
3	His	70	<1	72	<1

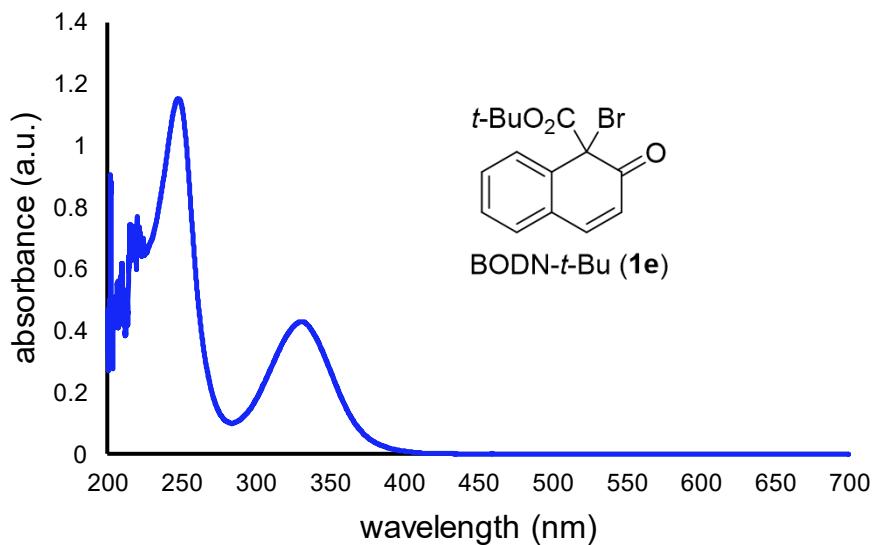
^a Reactions were run using **S17** (0.10 mmol), **1e** (0.12 mmol), and **5a** (0.0010 mmol) in H₂O/MeCN (v/v = 1:1, 1.0 mL). ASAHI SPECTRA CL-1503/CL-H1-450-9-1-B (525 nm) was used as a light source.

Scheme S5. Bromination of Tyrosine-Containing Peptides with Histidine and Unprotected Termini

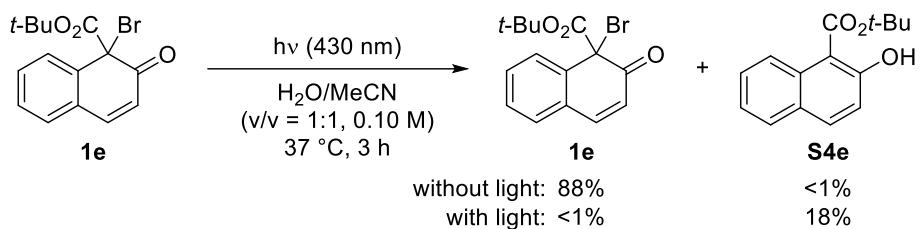


The reactions of **6f** and **6g** resulted in low mass balances while no byproduct was detected. It is probably because the high hydrophilicity of **6/7f** and **6/7g** decreased the efficiency of the extraction during the workup. The reaction of **6g** did not work probably because the substrate was not soluble in the present solvent system containing acetonitrile, which is at this stage necessary as **1e** is not soluble in more water-rich solvents.

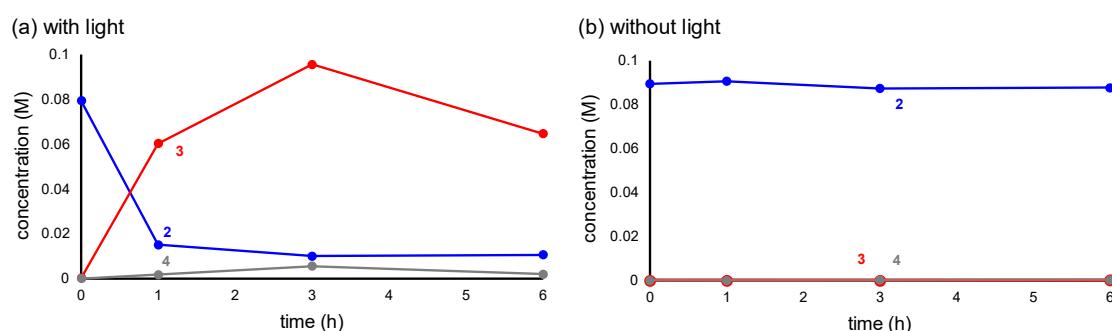
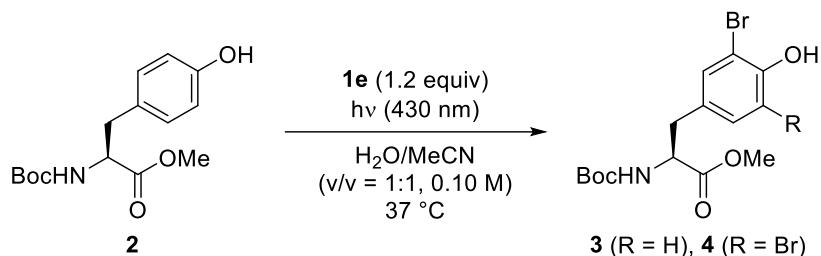
Scheme S6. UV-Vis Absorption of **1e**



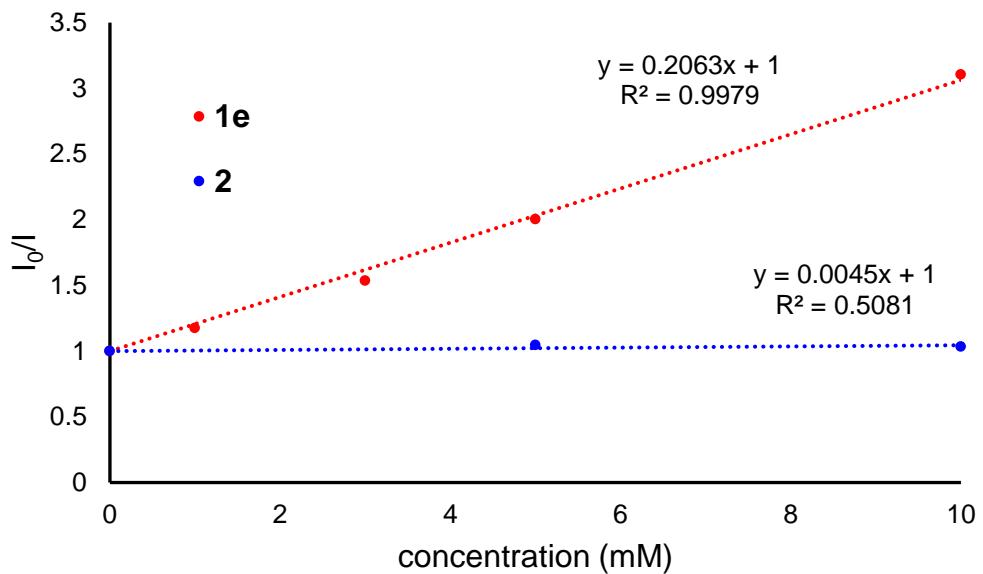
Scheme S7. Irradiation of 430 nm light to **1e**



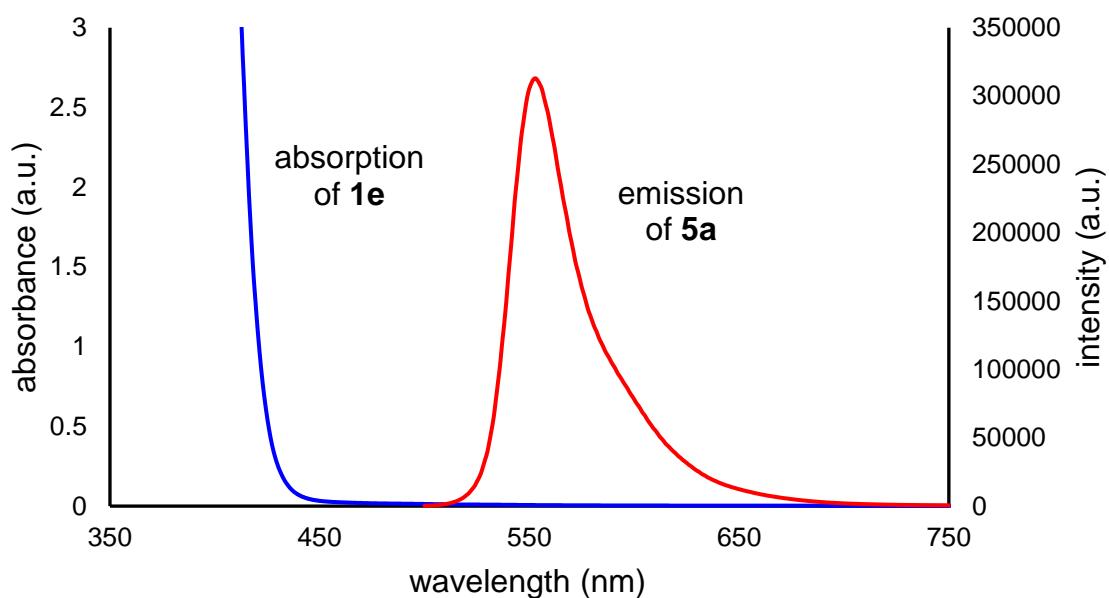
Scheme S8. Photochemical Tyrosine Bromination with Blue Light



Scheme S9. Fluorescence-Quenching Experiments

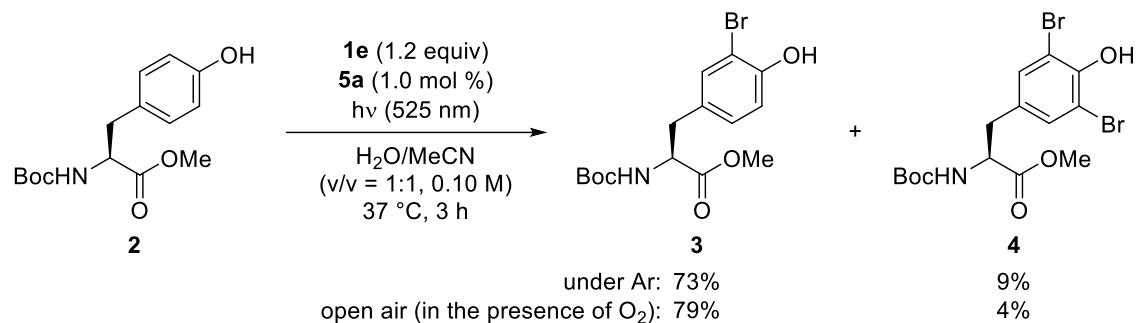


Scheme S10. Absorption of **1e** and Emission of **5a**

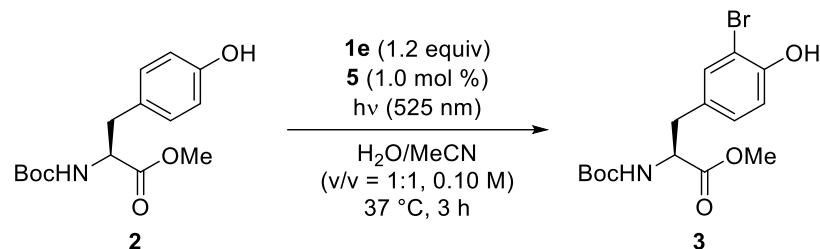


No overlap observed between the absorbance of **1e** and the emission of **5a**. The mechanism of Dexter energy transfer does not require their overlap, which is crucial for Förster energy transfer. Therefore, we believe that the present reaction proceeds through the mechanism of Dexter energy transfer.

Scheme S11. Reaction in the presence of O₂

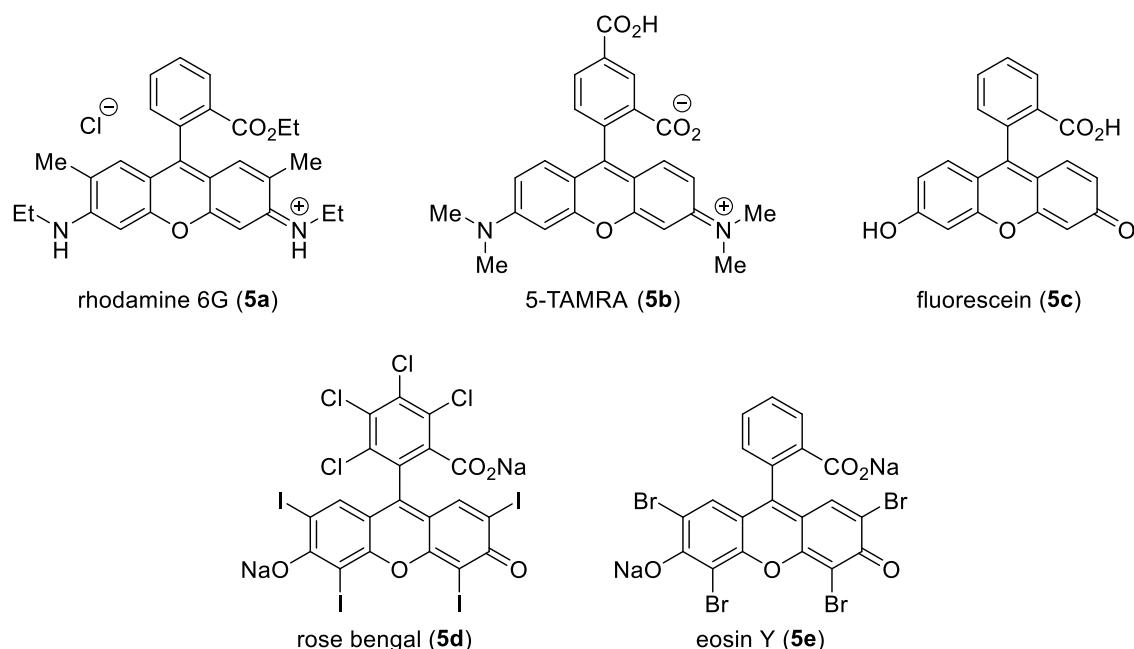


Scheme S12. Investigations of Photocatalysts^a



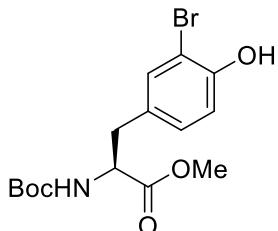
entry	5	yields without light (%)	yields with light (%)
1	5a	<1	76
2	5b	<1	47
3	5c	<1	54
4	5d	<1	64
5	5e	<1	57

^a Reactions were run using **2** (0.10 mmol), **1e** (0.12 mmol), and **5** (0.0010 mmol) in H₂O/MeCN (v/v = 1:1, 1.0 mL). ASAHI SPECTRA CL-1503/CL-H1-450-9-1-B (525 nm) was used as a light source. In all cases, yields of **4** were <5%.



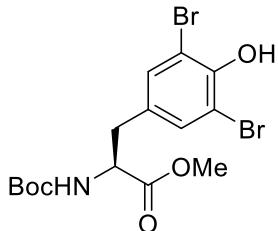
Characterization Data of Products

Methyl (S)-3-(3-bromo-4-hydroxyphenyl)-2-((*tert*-butoxycarbonyl)amino)propanoate (3): CAS RN [139517-74-1].



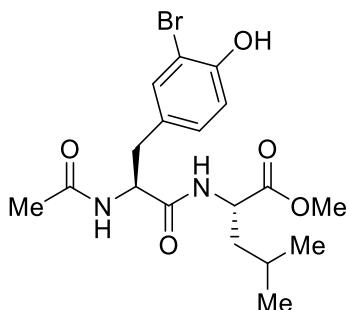
Yield: 76% (HPLC), white solid. ^1H NMR (CDCl_3) δ 7.22 (s, 1H), 6.97 (d, $J = 8.4$ Hz, 1H), 6.92 (d, $J = 8.4$ Hz, 1H), 5.57 (s, 1H), 5.00 (m, 1H), 4.52 (dd, $J = 13.6, 6.0$ Hz, 1H), 3.72 (s, 3H), 3.05 (m, 1H), 2.95 (m, 1H), 1.43 (s, 9H). ^{13}C NMR (CDCl_3) δ 172.1, 155.0, 151.4, 132.6, 130.0, 129.6, 116.1, 110.1, 80.1, 54.4, 52.3, 37.2, 28.3. TLC: R_f 0.32 (hexane/EtOAc = 3:1).

Methyl (S)-2-((*tert*-butoxycarbonyl)amino)-3-(3,5-dibromo-4-hydroxyphenyl)propanoate (4): CAS RN [355857-30-6].



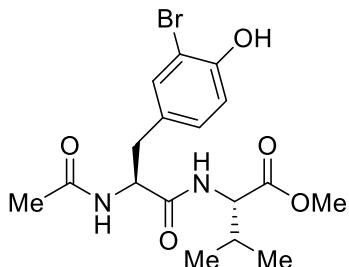
Yield: 27% (HPLC), white solid. ^1H NMR (CDCl_3) δ 7.22 (s, 2H), 5.82 (s, 1H), 5.03 (d, $J = 7.2$ Hz, 1H), 4.51 (m, 1H), 3.74 (s, 3H), 3.06 (dd, $J = 14.0, 6.0$ Hz, 1H), 2.92 (dd, $J = 14.0, 6.0$ Hz, 1H), 1.44 (s, 9H). ^{13}C NMR (CDCl_3) δ 171.8, 154.9, 148.4, 132.7, 130.8, 109.7, 80.2, 54.3, 52.4, 36.9, 28.3. TLC: R_f 0.38 ($\text{CHCl}_3/\text{EtOAc} = 20:1$).

Methyl ((S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl)-L-leucinate (7a).



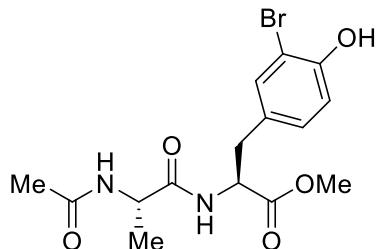
Yield: 75% (HPLC), white solid. ¹H NMR (CDCl₃) δ 7.28 (d, *J* = 2.4 Hz, 1H), 7.00 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.83–6.800 (m, 2H), 6.73 (s, 1H), 6.67 (d, *J* = 8.4 Hz, 1H), 4.72 (dd, *J* = 15.0, 6.6 Hz, 1H), 4.51 (m, 1H), 3.70 (s, 3H), 2.97–2.88 (m, 2H), 1.97 (s, 3H), 1.61–1.47 (m, 3H), 0.87 (d, *J* = 6.0 Hz, 6H). ¹³C NMR (CDCl₃) δ 172.9, 171.1, 170.5, 151.8, 133.2, 130.0, 129.7, 116.2, 110.1, 54.4, 52.5, 51.4, 41.3, 37.5, 24.8, 23.2, 22.8, 21.9. Mp. 72.0–72.2 °C. TLC: R_f 0.60 (CH₂Cl₂/MeOH = 10:1). IR (neat): 1743, 1645, 1539, 1508, 1499, 1209, 1153, 671, 594, 439 cm⁻¹. HRMS (ESI) Calcd for C₁₈H₂₅BrN₂O₅Na: [M+Na]⁺, 451.08445, 453.08241. Found: *m/z* 451.08342, 453.08111.

Methyl ((S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl)-L-valinate (7b).



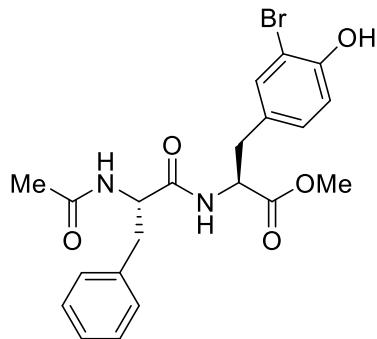
Yield: 73% (HPLC), white solid. ¹H NMR (CDCl₃) δ 7.35 (d, *J* = 1.8 Hz, 1H), 7.02 (dd, *J* = 8.4, 1.8 Hz, 1H), 6.78 (d, *J* = 8.4 Hz, 1H), 4.59 (dd, *J* = 8.4, 6.6 Hz, 1H), 4.29 (d, *J* = 6.0 Hz, 1H), 3.67 (s, 3H), 2.95 (dd, *J* = 13.8, 6.6 Hz, 1H), 2.76 (dd, *J* = 13.8, 8.4 Hz, 1H), 2.09 (m, 1H), 1.91 (s, 3H), 0.92 (d, *J* = 7.2 Hz, 3H), 0.91 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (CDCl₃) δ 171.8, 171.2, 170.5, 151.8, 133.1, 129.9, 129.7, 116.3, 110.1, 57.5, 54.6, 52.4, 37.6, 31.2, 23.1, 19.0, 17.8. Mp. 118.5–120.4 °C. TLC: R_f 0.26 (CH₂Cl₂/MeOH = 10:1). IR (neat): 1641, 1539, 1533, 1508, 1487, 1278, 1209, 1182, 1150, 752 cm⁻¹. HRMS (ESI) Calcd for C₁₇H₂₃BrN₂O₅Na: [M+Na]⁺, 437.06880, 439.06679. Found: *m/z* 437.06820, 4439.06609.

Methyl (S)-2-((S)-2-acetamidopropanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7c).



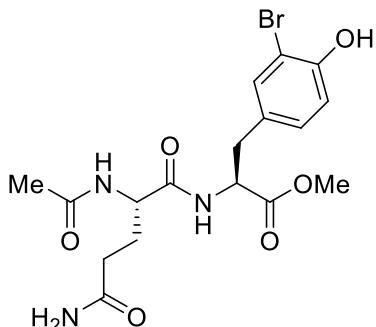
Yield: 51% (HPLC), white solid. ^1H NMR (CDCl_3) δ 7.26 (d, $J = 1.8$ Hz, 1H), 6.98 (dd, $J = 7.8, 1.8$ Hz, 1H), 6.78 (d, $J = 7.8$ Hz, 1H), 4.56 (dd, $J = 8.4, 6.0$ Hz, 1H), 4.31 (q, $J = 7.2$ Hz, 1H), 3.66 (s, 3H), 3.02 (dd, $J = 14.4, 6.0$ Hz, 1H), 2.88 (dd, $J = 14.4, 8.4$ Hz, 1H), 1.93 (s, 3H), 1.26 (d, $J = 7.2$ Hz, 3H). ^{13}C NMR (CDCl_3) δ 173.6, 171.8, 171.7, 153.0, 133.3, 129.3, 129.1, 115.8, 109.2, 53.8, 51.3, 48.9, 35.7, 21.1, 16.5. Mp. 170.3–170.5 °C. TLC: R_f 0.30 (EtOAc). IR (neat): 1737, 1625, 1604, 1465, 1444, 1296, 1225, 1188, 835, 513 cm^{-1} . HRMS (ESI) Calcd for $\text{C}_{15}\text{H}_{19}\text{BrN}_2\text{O}_5\text{Na}$: $[\text{M}+\text{Na}]^+$, 409.0369, 411.03491. Found: m/z 409.03682, 411.03444.

Methyl (S)-2-((S)-2-acetamido-3-phenylpropanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7d).



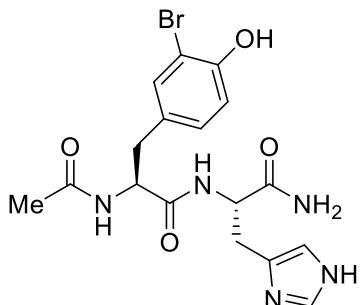
Yield: 69% (HPLC), pink solid. ^1H NMR (CD_3OD) δ 7.27–7.17 (m, 6H), 6.96 (dd, $J = 8.4, 2.0$ Hz, 1H), 6.77 (d, $J = 8.4$ Hz, 1H), 4.61–4.55 (m, 2H), 3.64 (s, 3H), 3.07–2.99 (m, 2H), 2.87–2.76 (m, 2H), 1.85 (s, 3H). ^{13}C NMR (CD_3OD) δ 172.1, 171.7, 171.6, 153.0, 137.0, 133.4, 129.2, 129.1, 128.9, 128.0, 126.4, 115.8, 109.2, 54.5, 53.8, 51.4, 37.5, 35.8, 21.1. Mp. 163.3–163.7 °C. TLC: R_f 0.48 (hexane/EtOAc = 1:3). IR (neat): 1653, 1647, 1533, 1420, 1179, 748, 698, 488, 422, 417 cm^{-1} . HRMS (ESI) Calcd for $\text{C}_{21}\text{H}_{23}\text{BrN}_2\text{O}_5\text{Na}$: $[\text{M}+\text{Na}]^+$, 485.06880, 487.06676. Found: m/z 485.06859, 487.06637.

Methyl (S)-2-((S)-2-acetamido-5-amino-5-oxopentanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7e).



Yield: 36% (HPLC, the mass balance was low probably because of the high hydrophilicity of **7e**, which decreased the efficiency of the extraction during the workup, while no byproduct was detected), white solid. ¹H NMR (CD₃OD) δ 7.29 (d, *J* = 2.4 Hz, 1H), 6.99 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.79 (d, *J* = 8.4 Hz, 1H), 4.59 (dd, *J* = 8.4, 5.4 Hz, 1H), 4.35 (dd, *J* = 8.4, 6.0 Hz, 1H), 3.69 (s, 3H), 3.04 (dd, *J* = 14.4, 6.0 Hz, 1H), 2.88 (dd, *J* = 14.4, 8.4 Hz, 1H), 2.26 (t, *J* = 7.8 Hz, 2H), 2.01 (ddt, *J* = 14.4, 8.4, 7.8 Hz, 1H), 1.96 (s, 3H), 1.86 (ddt, *J* = 14.4, 5.4, 7.8 Hz, 1H). ¹³C NMR (CD₃OD) δ 177.8, 173.7, 173.3, 173.2, 154.3, 134.8, 130.8, 130.4, 117.3, 110.6, 55.4, 53.9, 52.7, 37.2, 32.4, 28.9, 22.5. Mp. 184.5–185.5 °C. TLC: R_f 0.23 (CH₂Cl₂/MeOH = 10:1). IR (neat): 1734, 1653, 1541, 1437, 1290, 1221, 1084, 1043, 583 cm⁻¹. HRMS (ESI) Calcd for C₁₇H₂₂BrN₃O₆Na: [M+Na]⁺, 466.05842, 468.05660. Found: *m/z* 466.05789, 468.05550.

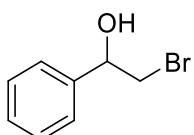
(S)-2-Acetamido-N-((S)-1-amino-3-(1*H*-imidazol-4-yl)-1-oxopropan-2-yl)-3-(3-bromo-4-hydroxyphenyl)propanamide (7f).



Yield: 14% (HPLC, the mass balance was low probably because of the high hydrophilicity of **7f**, which decreased the efficiency of the extraction during the workup, while no byproduct was detected), white solid. ¹H NMR (CD₃OD) δ 7.67 (s, 1H), 7.36 (s, 1H), 7.34 (d, *J* = 2.4 Hz, 1H), 7.01 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.89 (s, 1H), 6.79 (d, *J* = 8.4 Hz, 1H), 4.55 (dd, *J* = 7.8, 4.8 Hz, 1H), 4.46 (dd, *J* = 9.6, 6.0 Hz, 1H), 3.09 (dd, *J* =

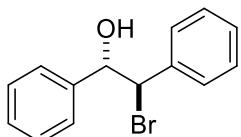
15.0, 4.8 Hz, 1H), 2.98 (dd, J = 14.4, 6.0 Hz, 1H), 2.97 (dd, J = 15.0, 7.8 Hz, 1H), 2.73 (dd, J = 14.4, 9.6 Hz, 2H), 1.91 (s, 3H). ^{13}C NMR (CD_3OD) δ 175.5, 173.6, 173.5, 154.2, 136.2, 134.7, 134.1, 130.9, 130.3, 117.2, 112.1, 110.6, 56.6, 54.4, 37.2, 30.1, 22.4. Mp. 147.5–148.5 °C. TLC: R_f 0.28 ($\text{CH}_2\text{Cl}_2/\text{MeOH} = 5:1$). IR (neat): 3254, 2924, 2853, 1636, 1646, 1539, 1429, 1373, 1288, 1203, 1134, 995, 588 cm^{-1} . HRMS (ESI) Calcd for $\text{C}_{17}\text{H}_{21}\text{BrN}_5\text{O}_4$: $[\text{M}+\text{H}]^+$, 438.07714, 440.07565. Found: m/z 438.07635, 440.07379.

2-Bromo-1-phenylethan-1-ol (9a): CAS RN [2425-28-7].



Yield: 46%, yellow liquid. ^1H NMR (CDCl_3) δ 7.39–7.32 (m, 5H), 4.94 (ddd, J = 8.8, 3.2, 3.2 Hz, 1H), 3.65 (dd, J = 10.8, 3.2 Hz, 1H), 3.55 (dd, J = 10.8, 8.8 Hz, 1H), 2.63 (d, J = 3.2 Hz, 1H). ^{13}C NMR (CDCl_3) δ 140.2, 128.7, 128.5, 126.0, 73.8, 40.3. TLC: R_f 0.23 (hexane/EtOAc = 9:1).

(1*S,2*R**)-2-Bromo-1,2-diphenylethan-1-ol (9b):** CAS RN [10368-43-1].

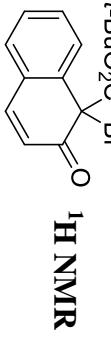


Yield: 9%, yellow liquid. ^1H NMR (CDCl_3) δ 7.40–7.29 (m, 10H), 5.21 (dd, J = 6.6, 3.0 Hz, 1H), 5.09 (d, J = 6.6 Hz, 1H), 2.40 (d, J = 3.0 Hz, 1H). ^{13}C NMR (CDCl_3) δ 139.7, 137.6, 128.9, 128.8, 128.5, 128.4, 128.2, 127.0, 78.1, 58.9. TLC: R_f 0.38 (hexane/EtOAc = 10:1).

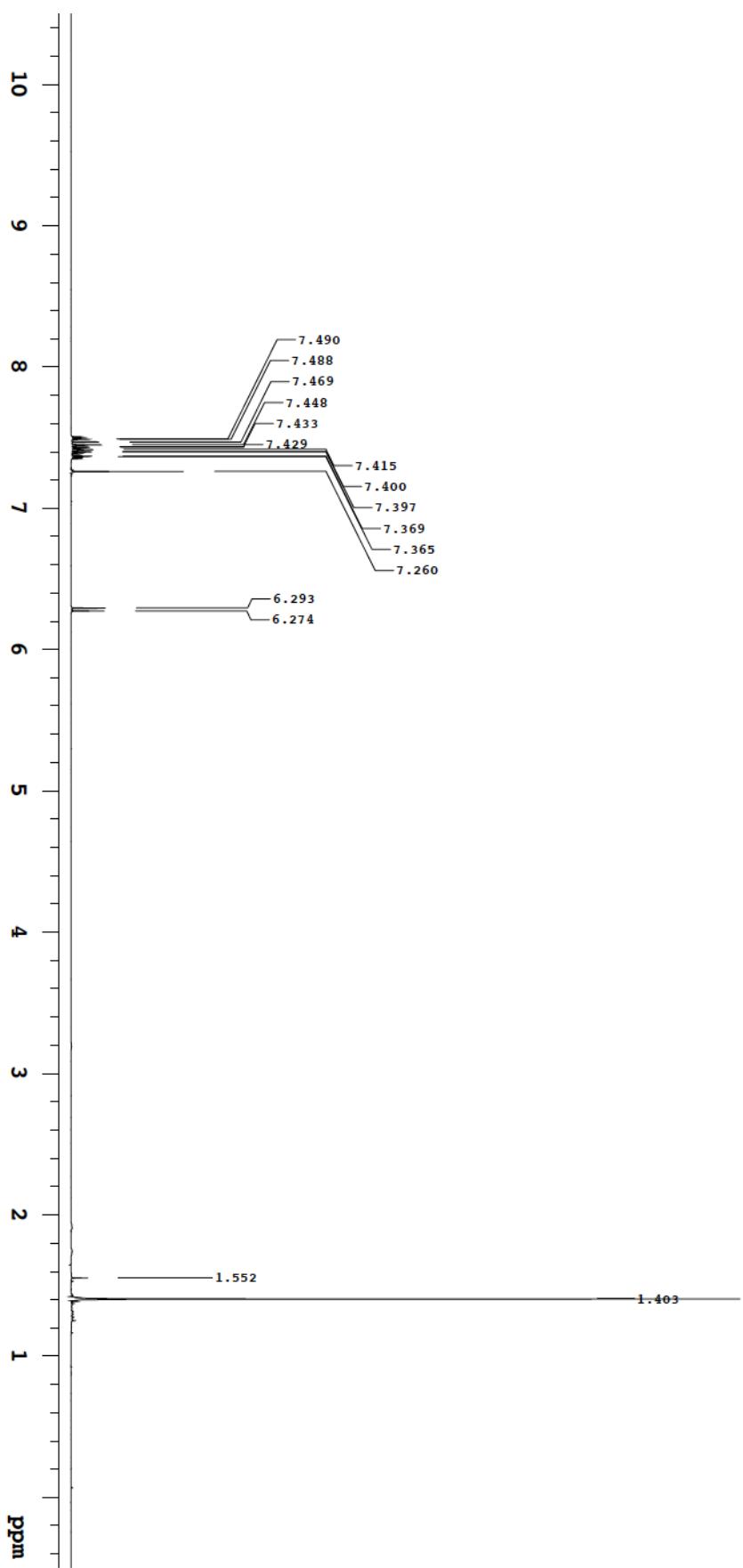
References

1. Takeshima, A.; Shimogaki, M.; Kano, T.; Maruoka, K. *ACS Catal.* **2020**, *10*, 5959.
2. Dogo.-Isonagie, C. ; Bekele, T.; France, S.; Wolfer, J.; Weatherwax, A.; Taggi, A. E.; Lectka, T. *J. Org. Chem.* **2006**, *71*, 8946.
3. Zhou, Y.-M.; Ping, Y.-J.; Xu, Z.-J.; Che, C.-M. *Asian J. Org. Chem.* **2021**, *10*, 674.
4. Zhang, Z.; Sun, Q.; Xu, D.; Xia, C.; Sun, W. *Green Chem.* **2016**, *18*, 5485.
5. Ashida, Y.; Sato, Y.; Suzuki, T.; Ueno, K.; Kai, K.; Nakatsuji, H.; Tanabe, Y. *Chem. Eur. J.* **2015**, *21*, 5934.
6. Fang, L.-Z.; Shen, J.-M.; Lv, Q.-H.; Yan, F.-L. *Asian J. Chem.* **2011**, *23*, 3425.

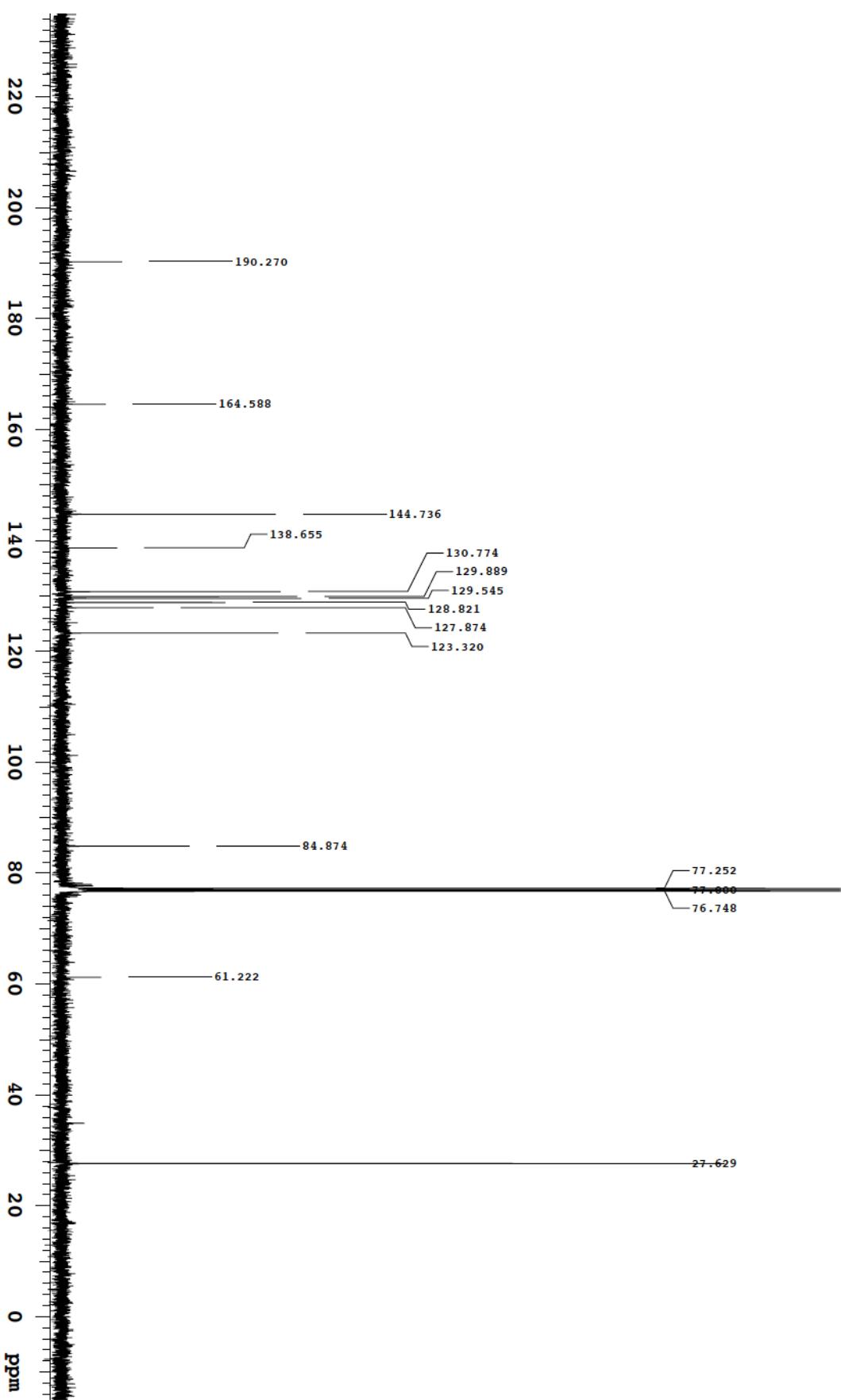
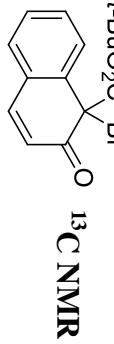
tert-Butyl 1-bromo-2-oxo-1,2-dihydronaphthalene-1-carboxylate (**1e**)



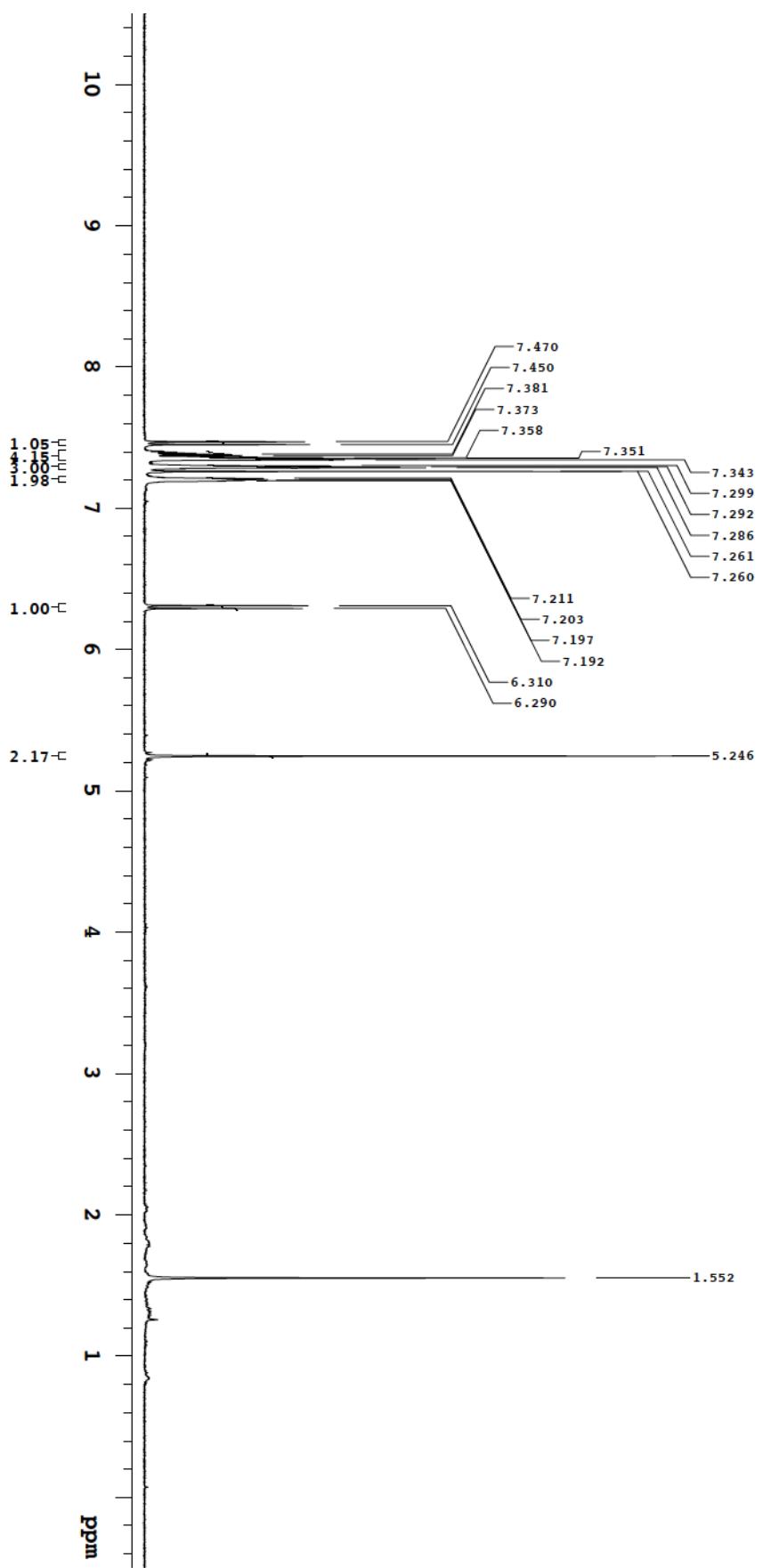
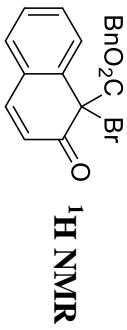
NMR Spectra (¹H, ¹³C) of BODNs



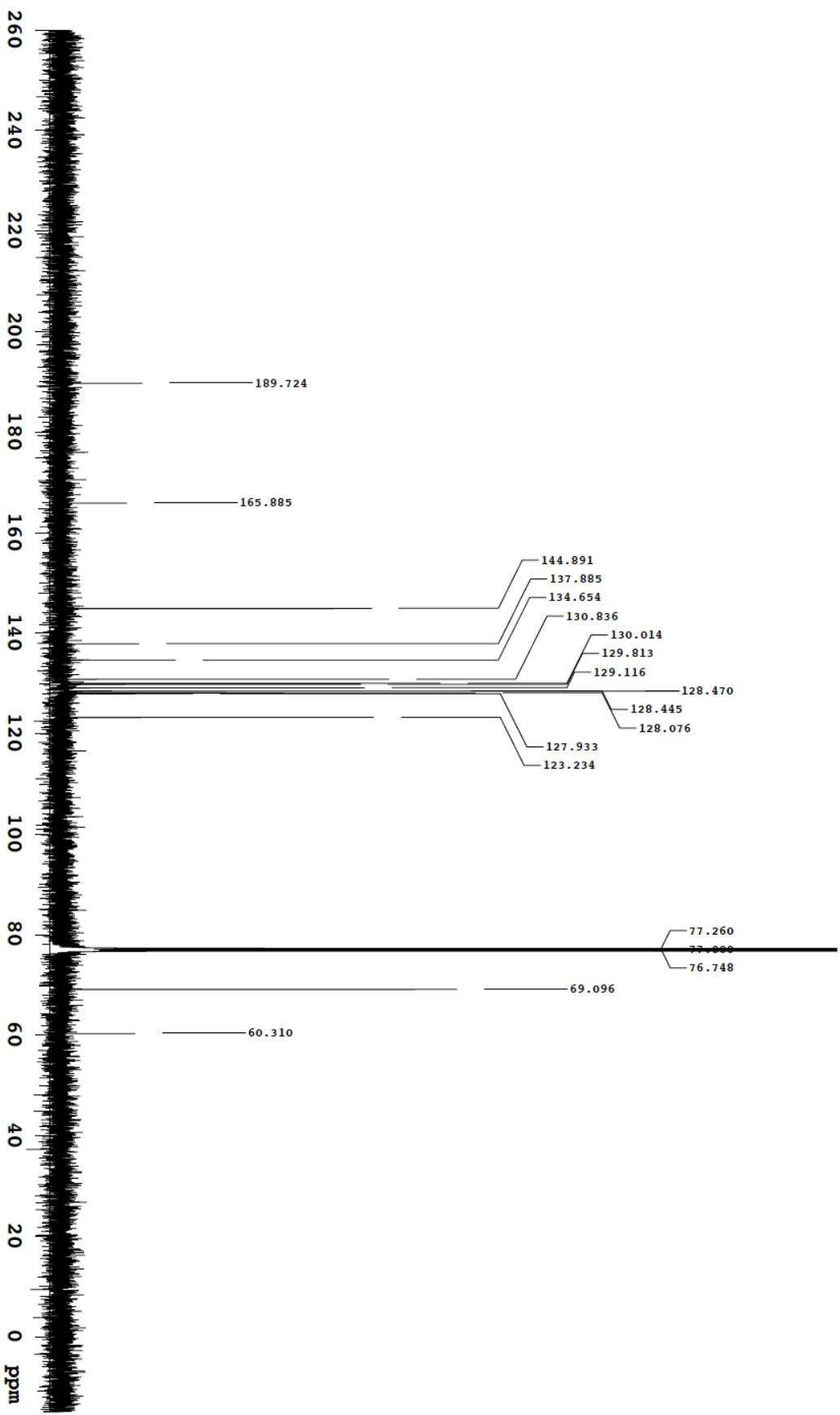
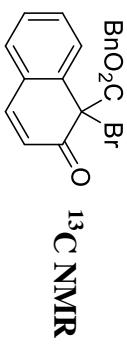
tert-Butyl 1-bromo-2-oxo-1,2-dihydronaphthalene-1-carboxylate (**1e**)



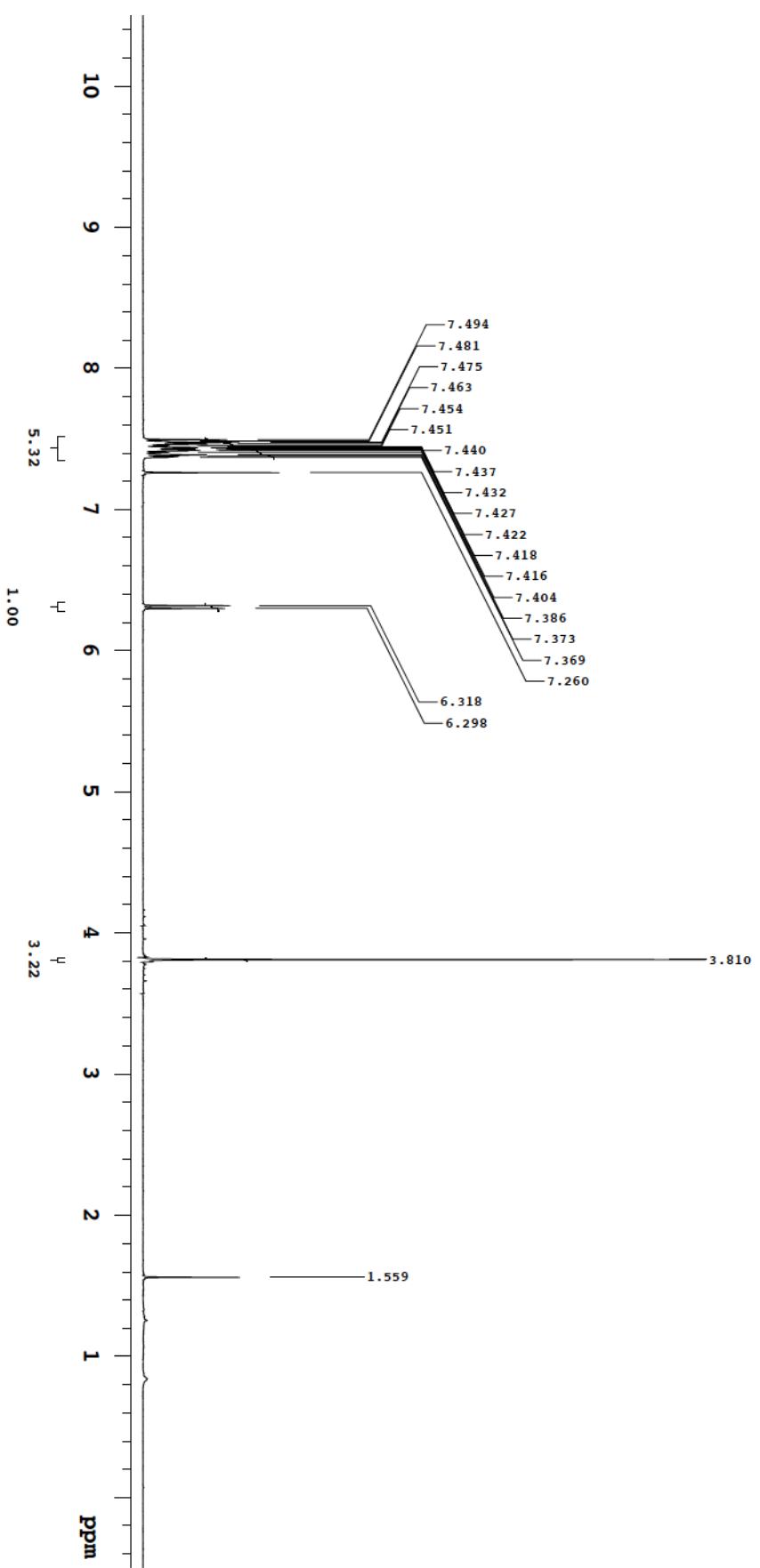
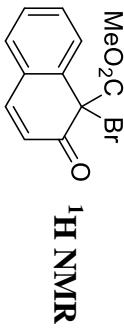
Benzyl 1-bromo-2-oxo-1,2-dihydroronaphthalene-1-carboxylate (1f)



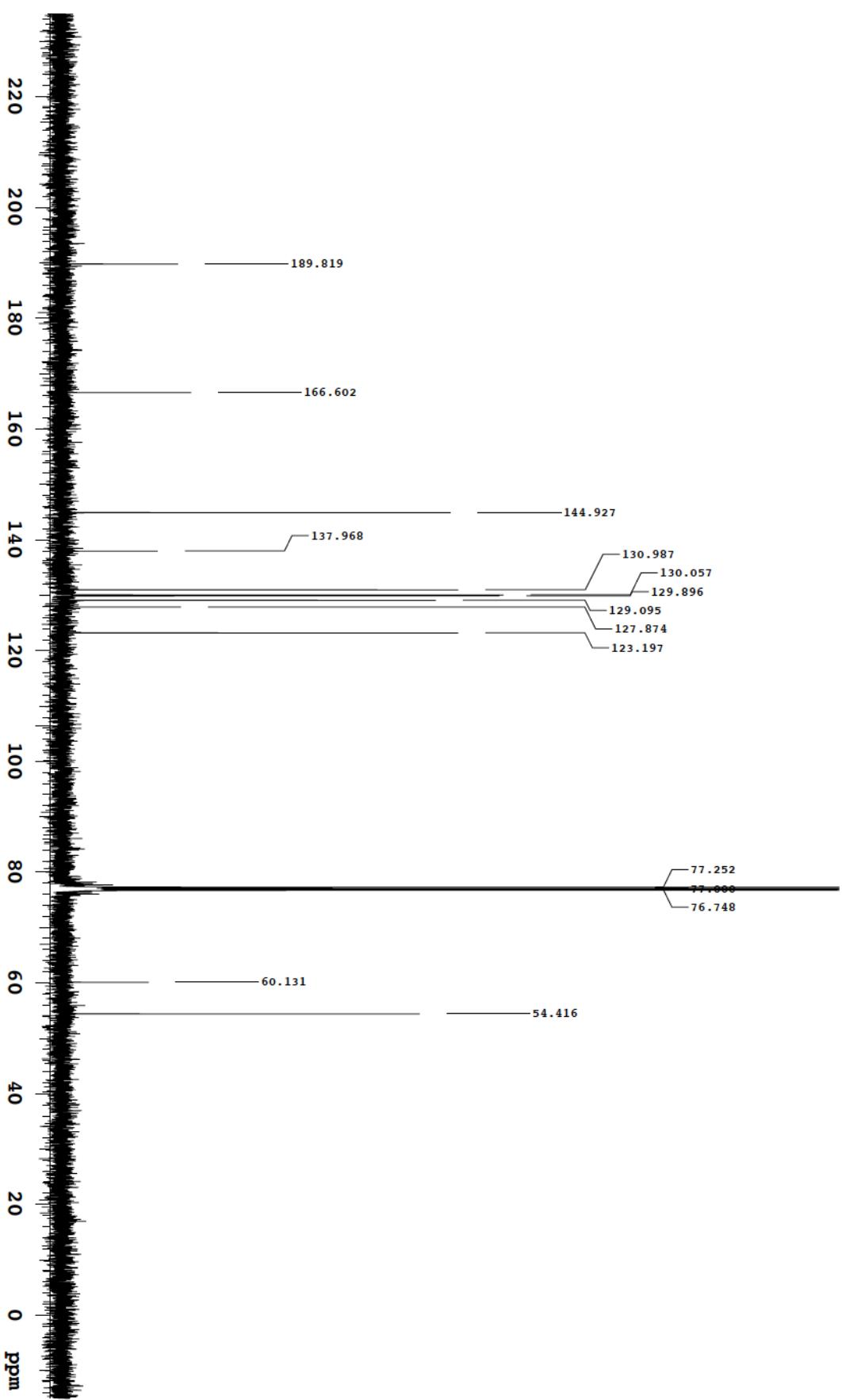
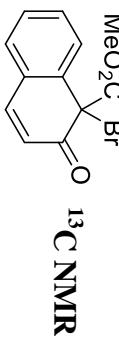
Benzyl 1-bromo-2-oxo-1,2-dihydronaphthalene-1-carboxylate (1f)



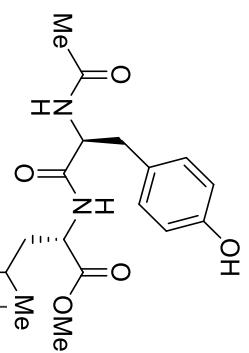
Methyl 1-bromo-2-oxo-1,2-dihydroronaphthalene-1-carboxylate (1g)



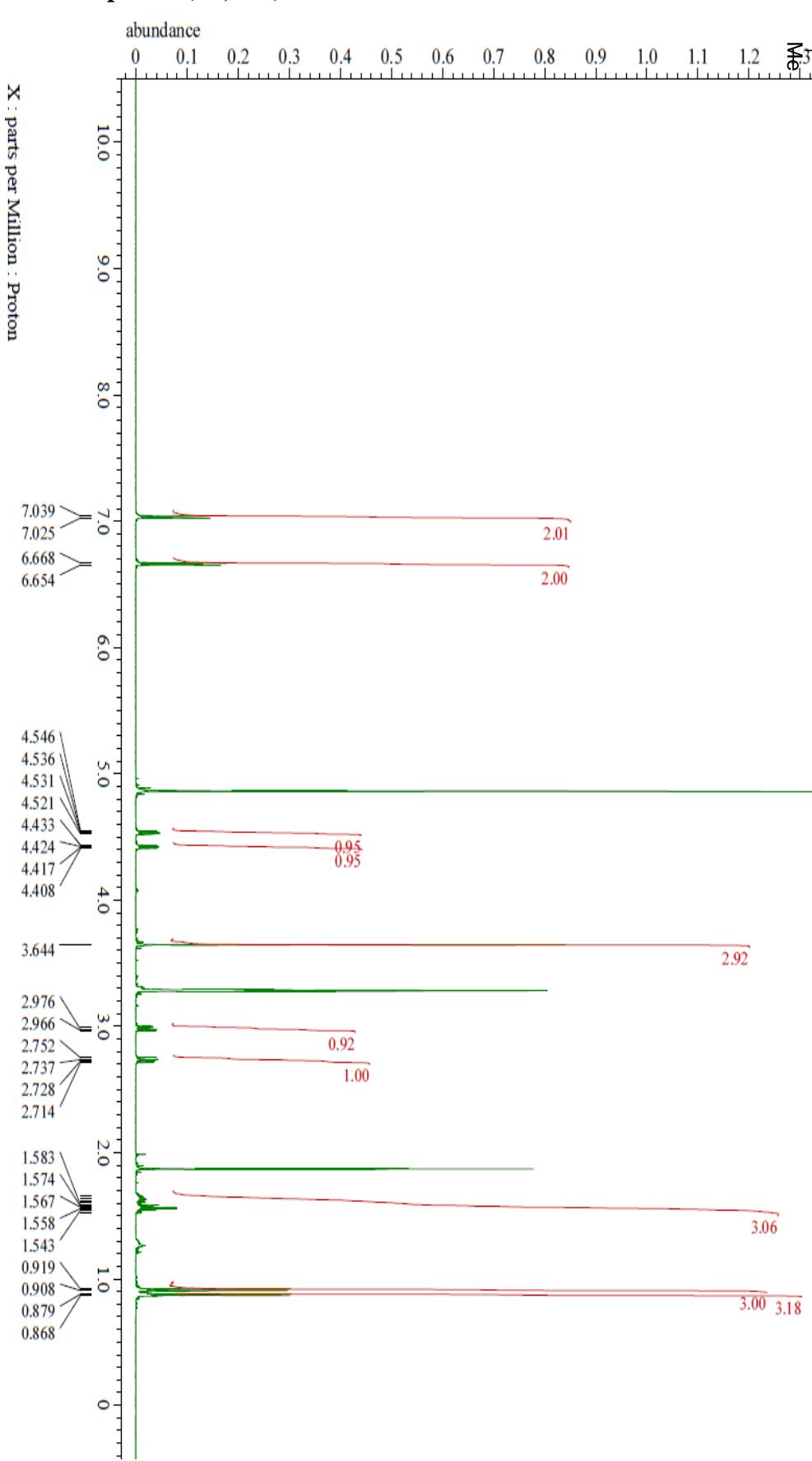
Methyl 1-bromo-2-oxo-1,2-dihydronaphthalene-1-carboxylate (1g)



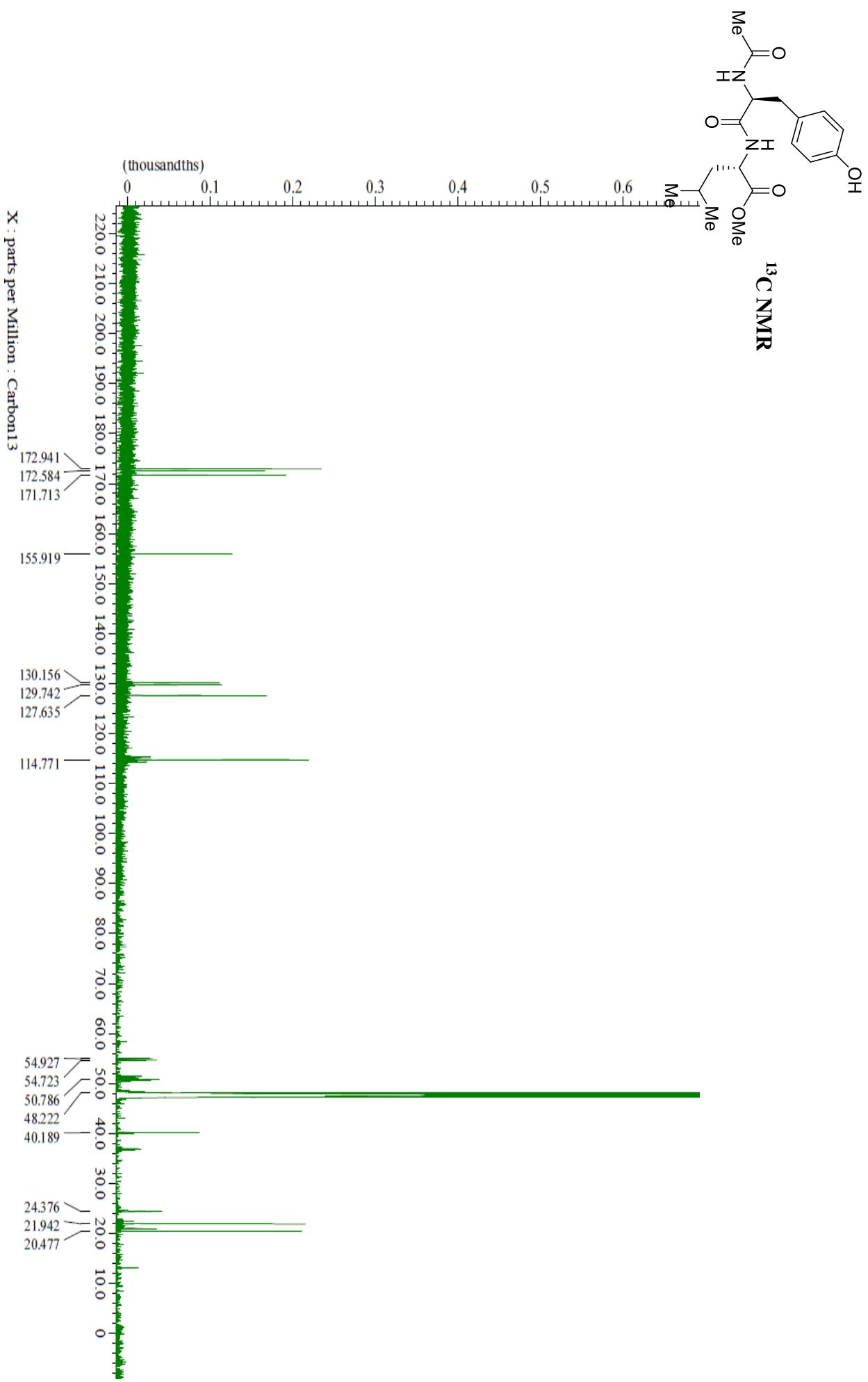
N-Acetyl-L-tyrosyl-L-leucin methyl ester (**6a**)



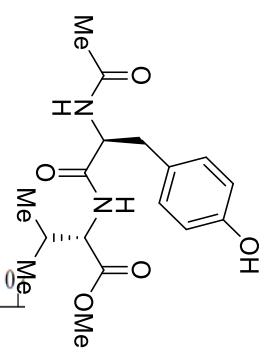
¹H NMR



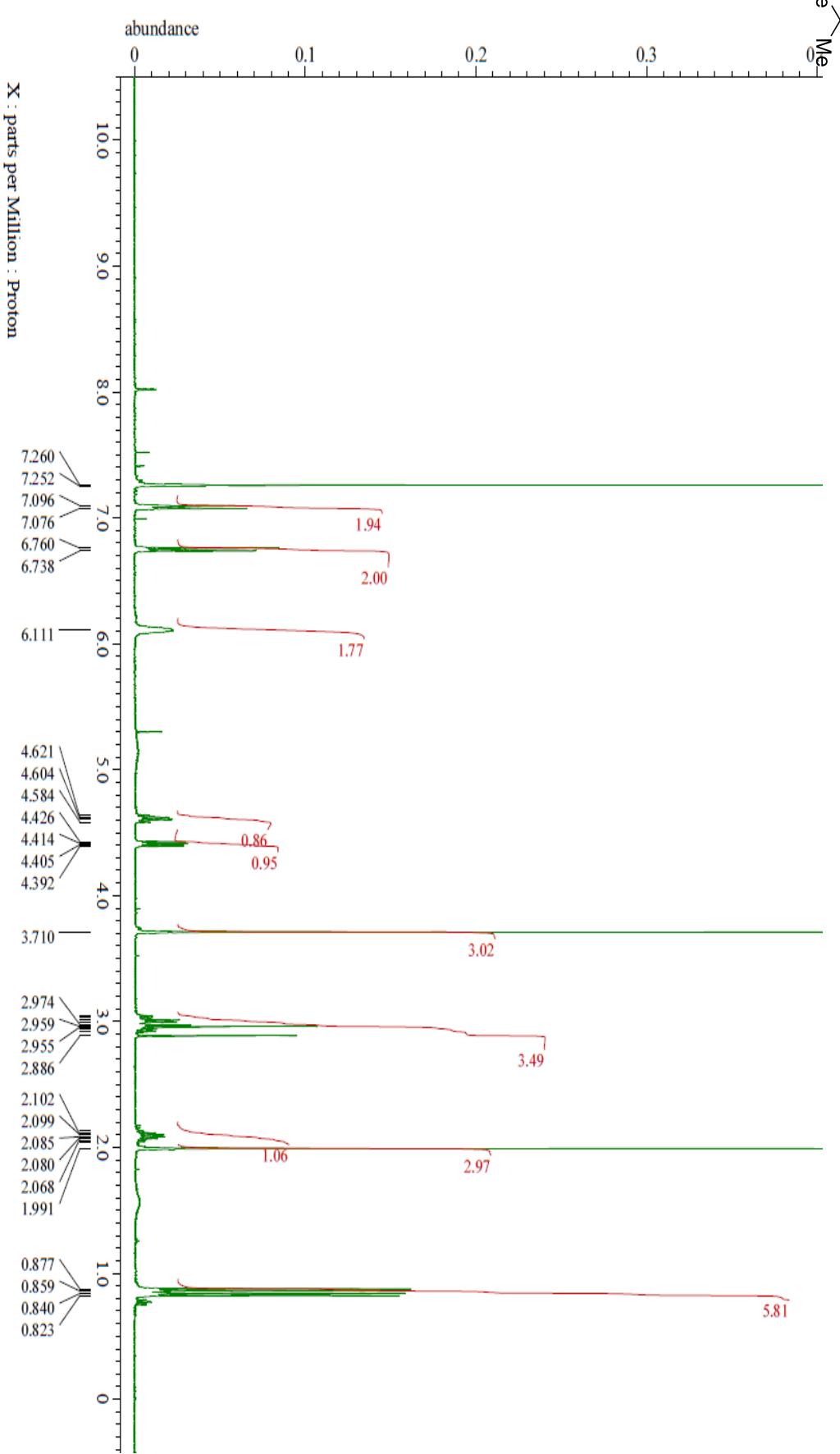
N-Acetyl-L-tyrosyl-L-leucin methyl ester (**6a**)



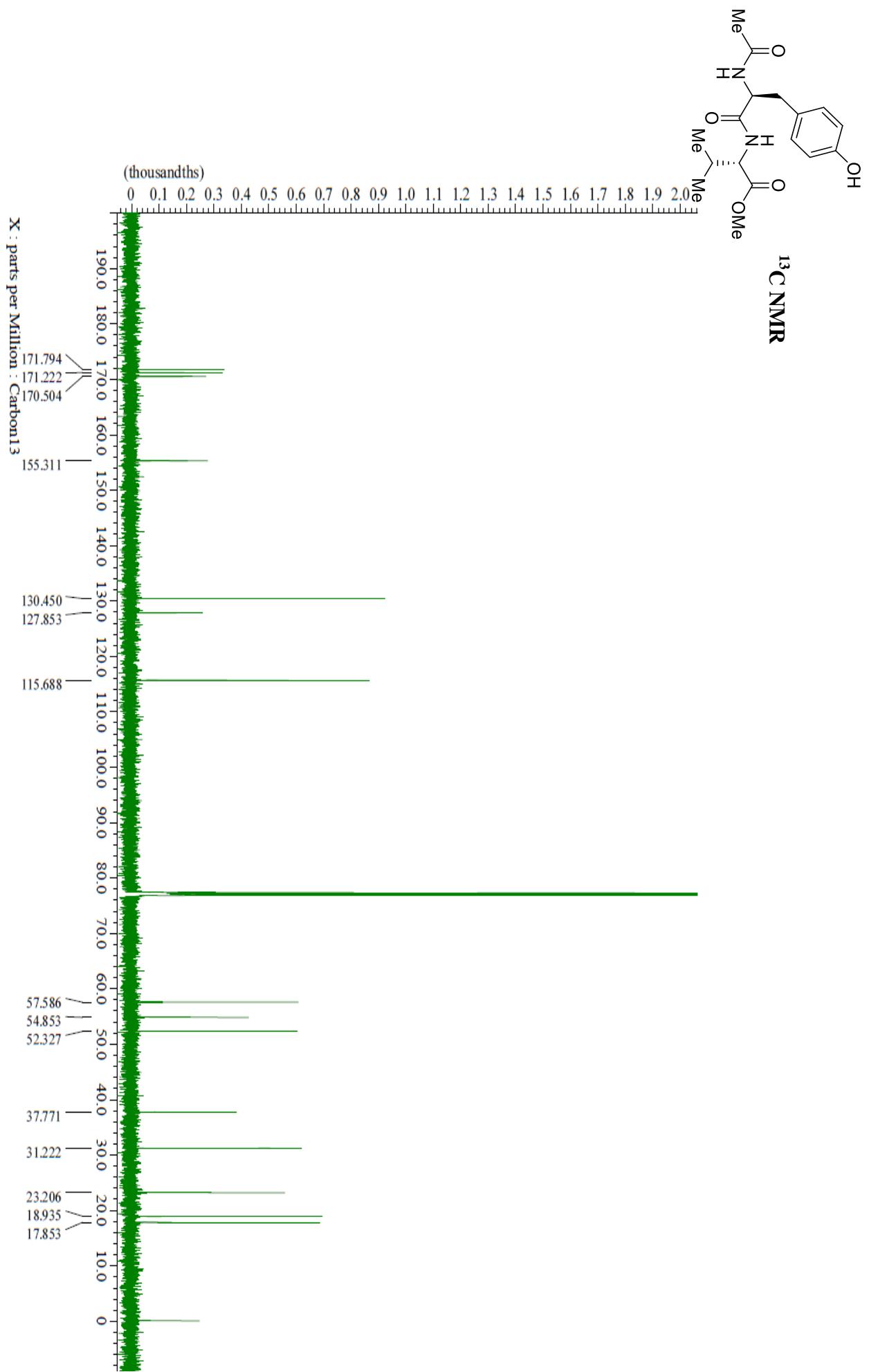
N-Acetyl-L-tyrosyl-L-valine methyl ester (**6b**)



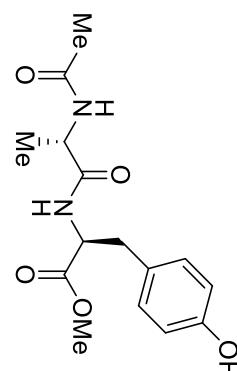
^1H NMR



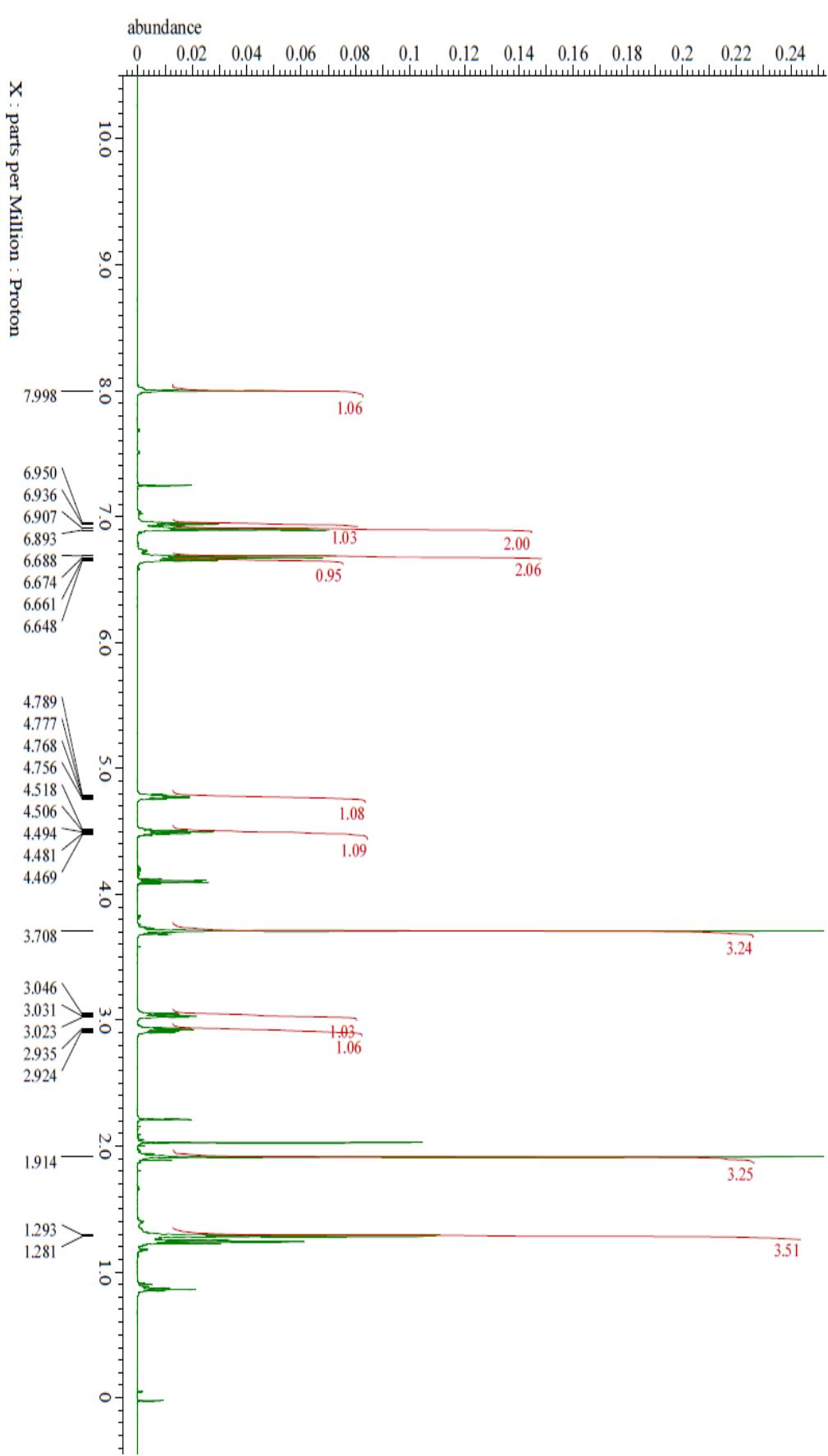
N-Acetyl-L-tyrosyl-L-valine methyl ester (**6b**)



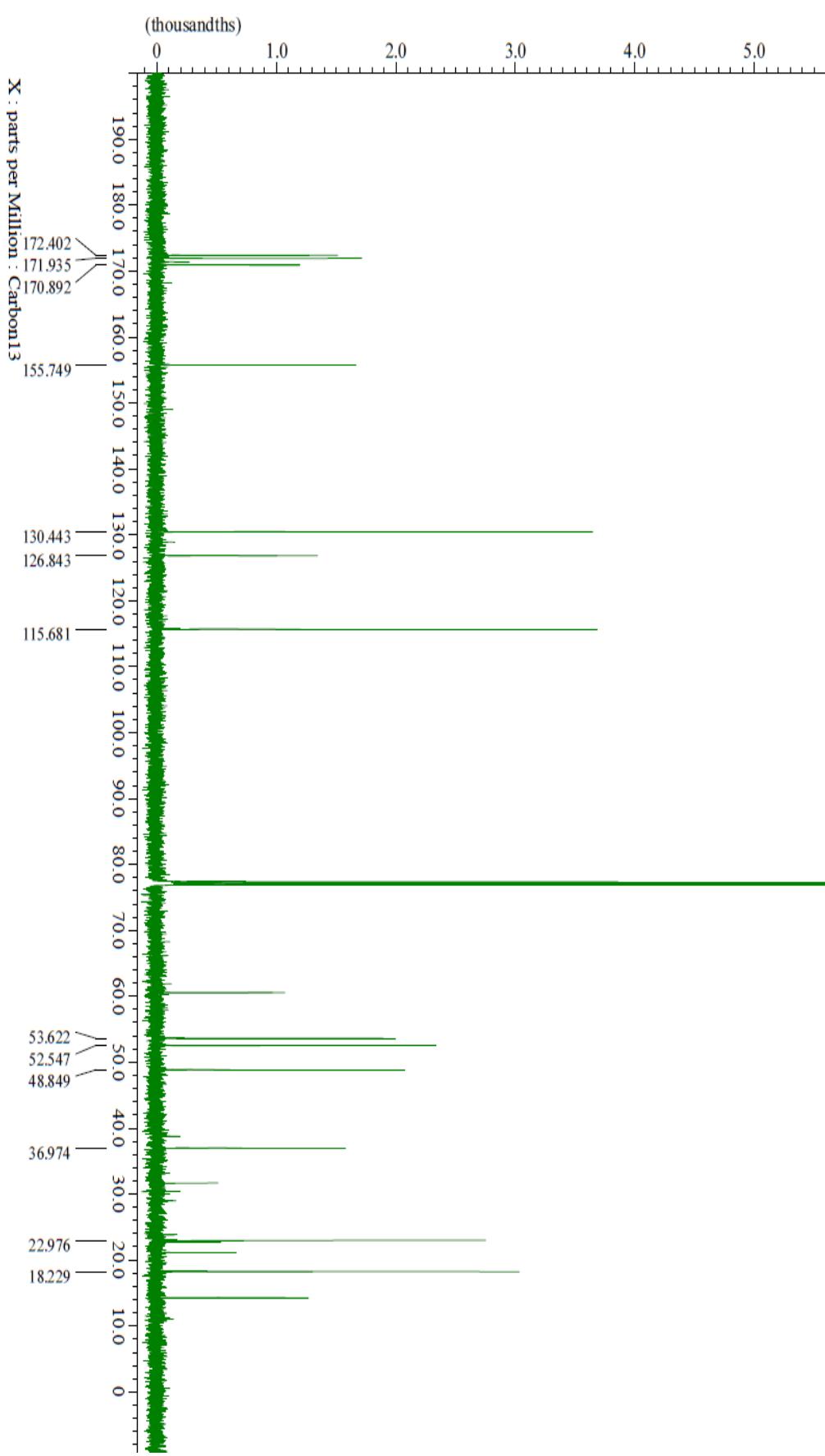
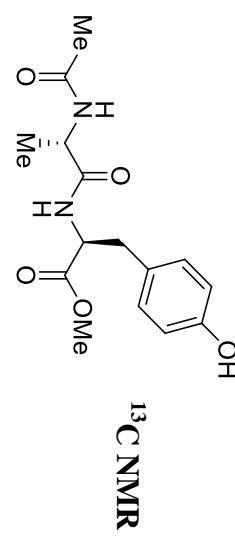
N-Acetyl-L-alanyl-L-tyrosine methyl ester (6c)



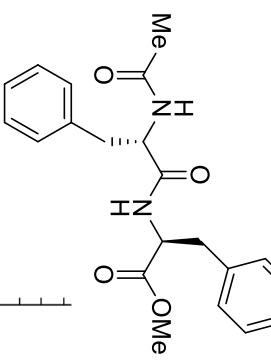
¹H NMR



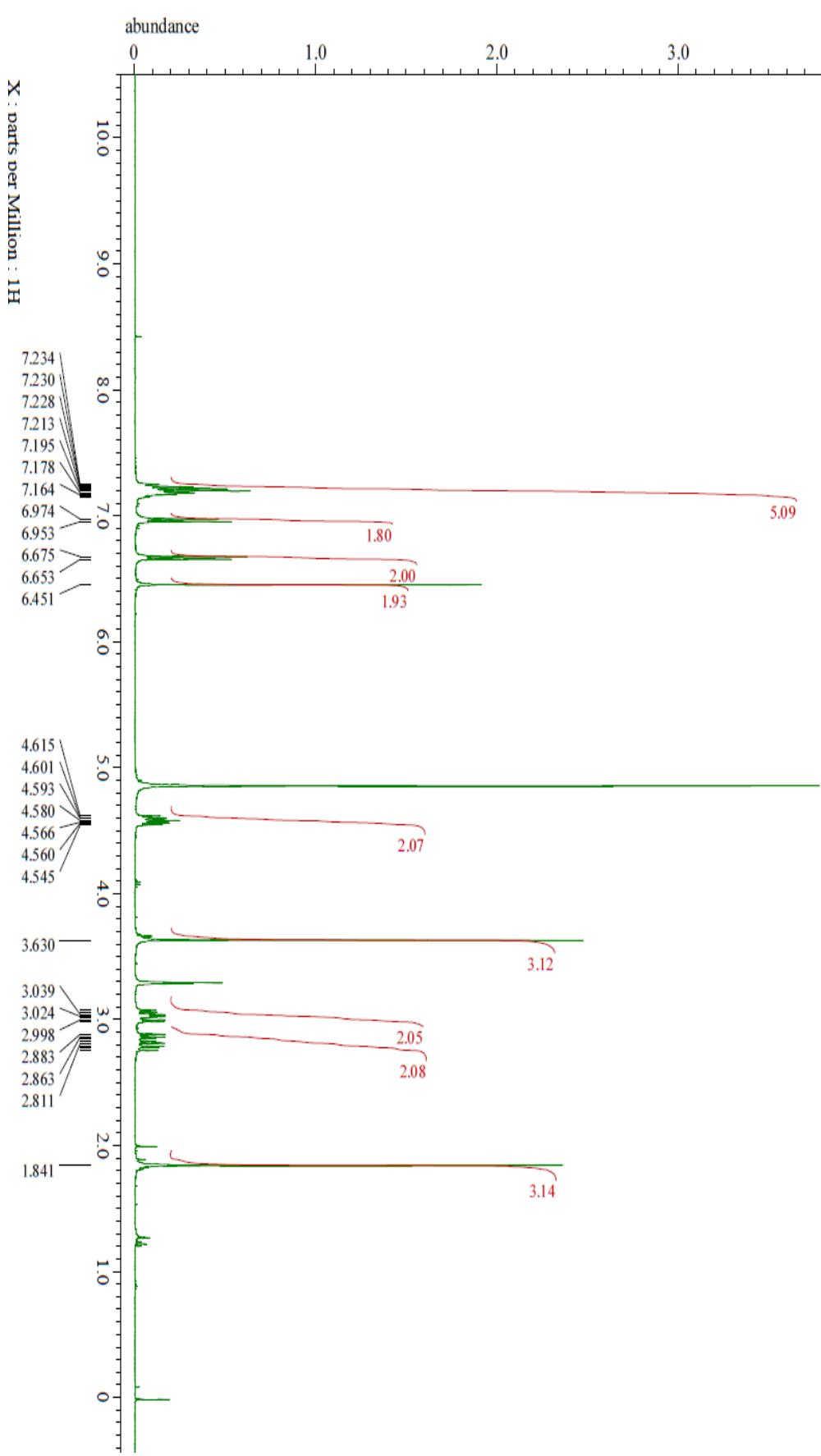
N-Acetyl-L-alanyl-L-tyrosine methyl ester (6c)



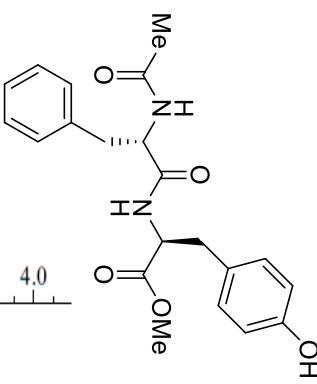
N-Acetyl-L-phenylalanyl-L-tyrosine methyl ester (**6d**)



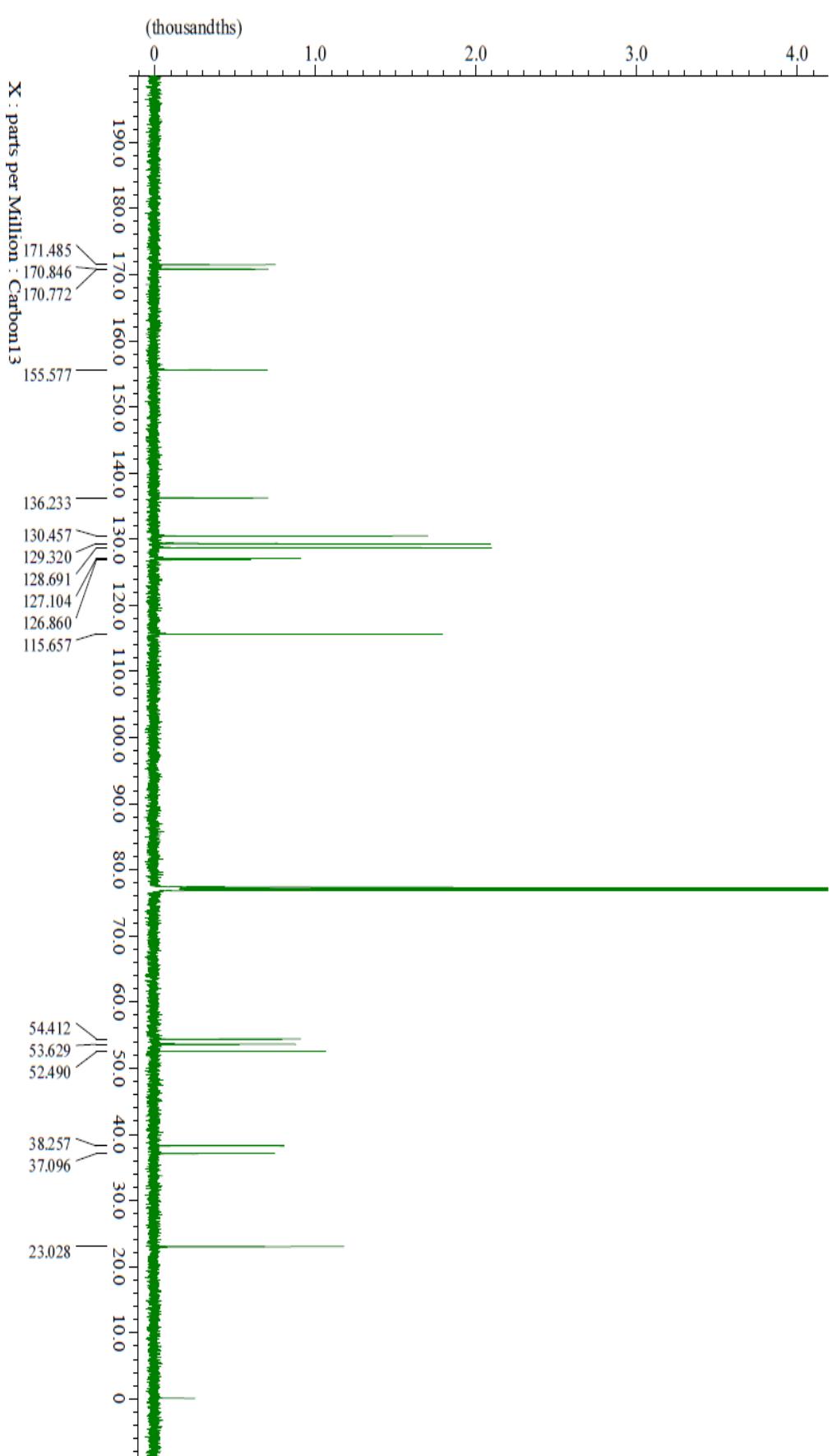
¹H NMR



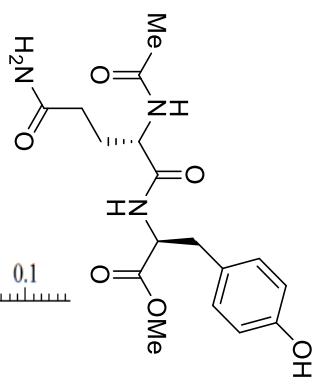
N-Acetyl-L-phenylalanyl-L-tyrosine methyl ester (**6d**)



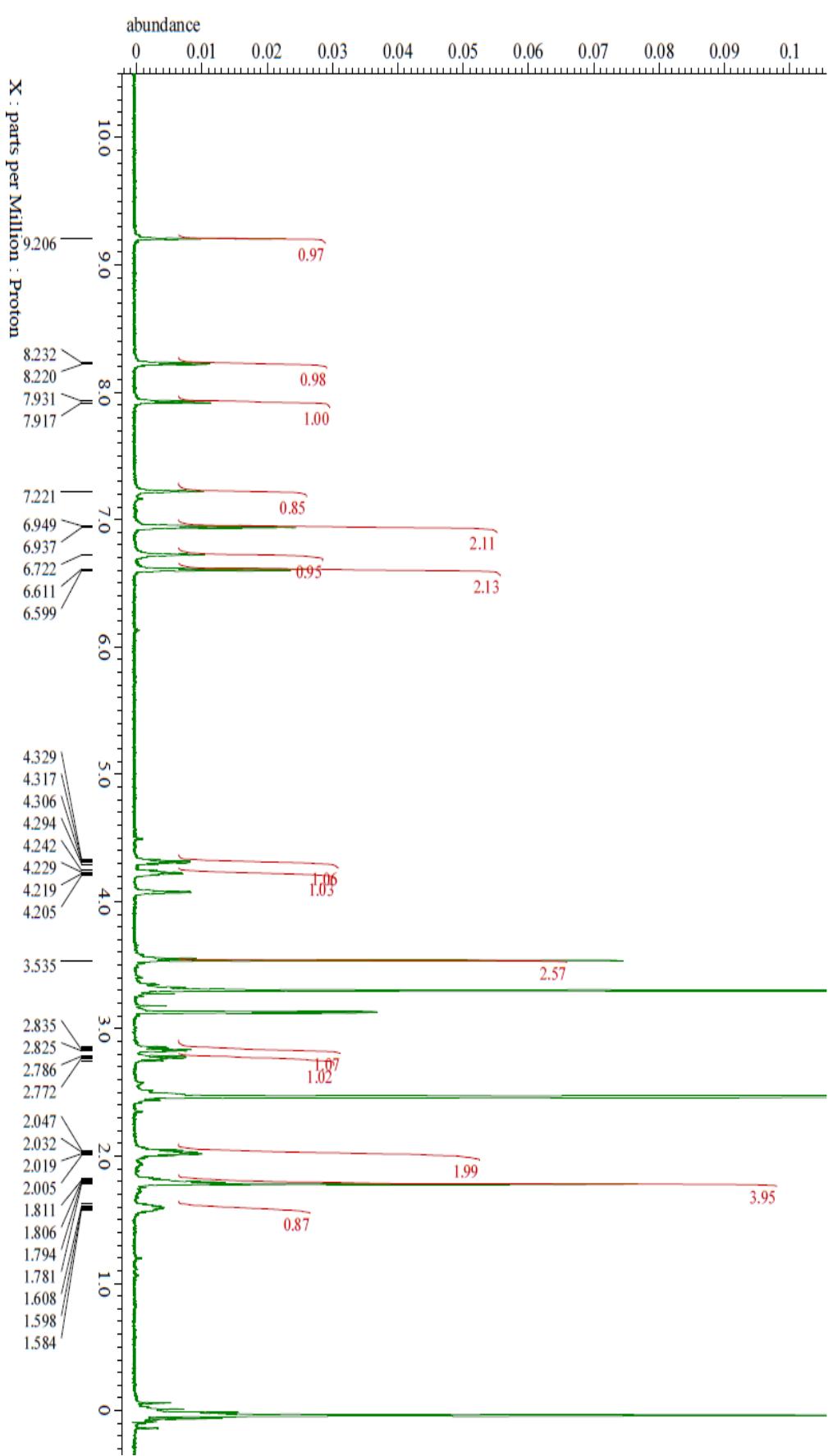
¹³C NMR



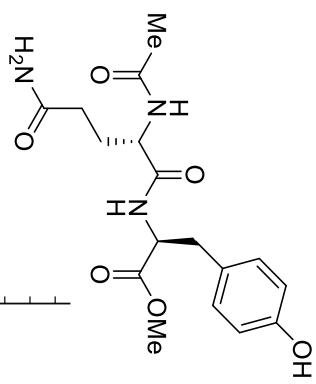
N-Acetyl-L-glutaminyl-L-tyrosine methyl ester (**6e**)



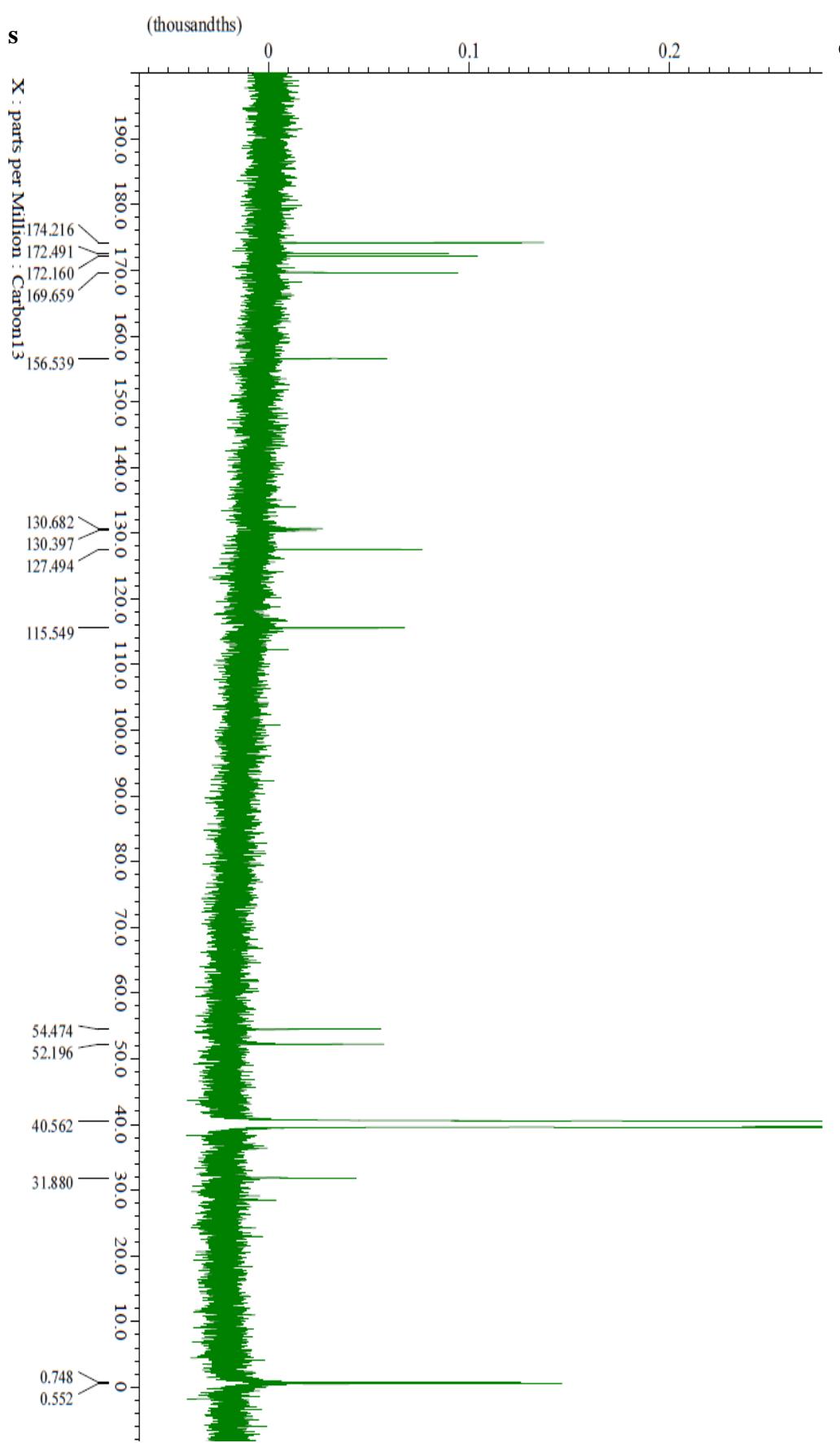
¹H NMR



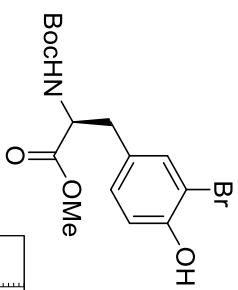
N-Acetyl-L-glutaminyl-L-tyrosine methyl ester (**6e**)



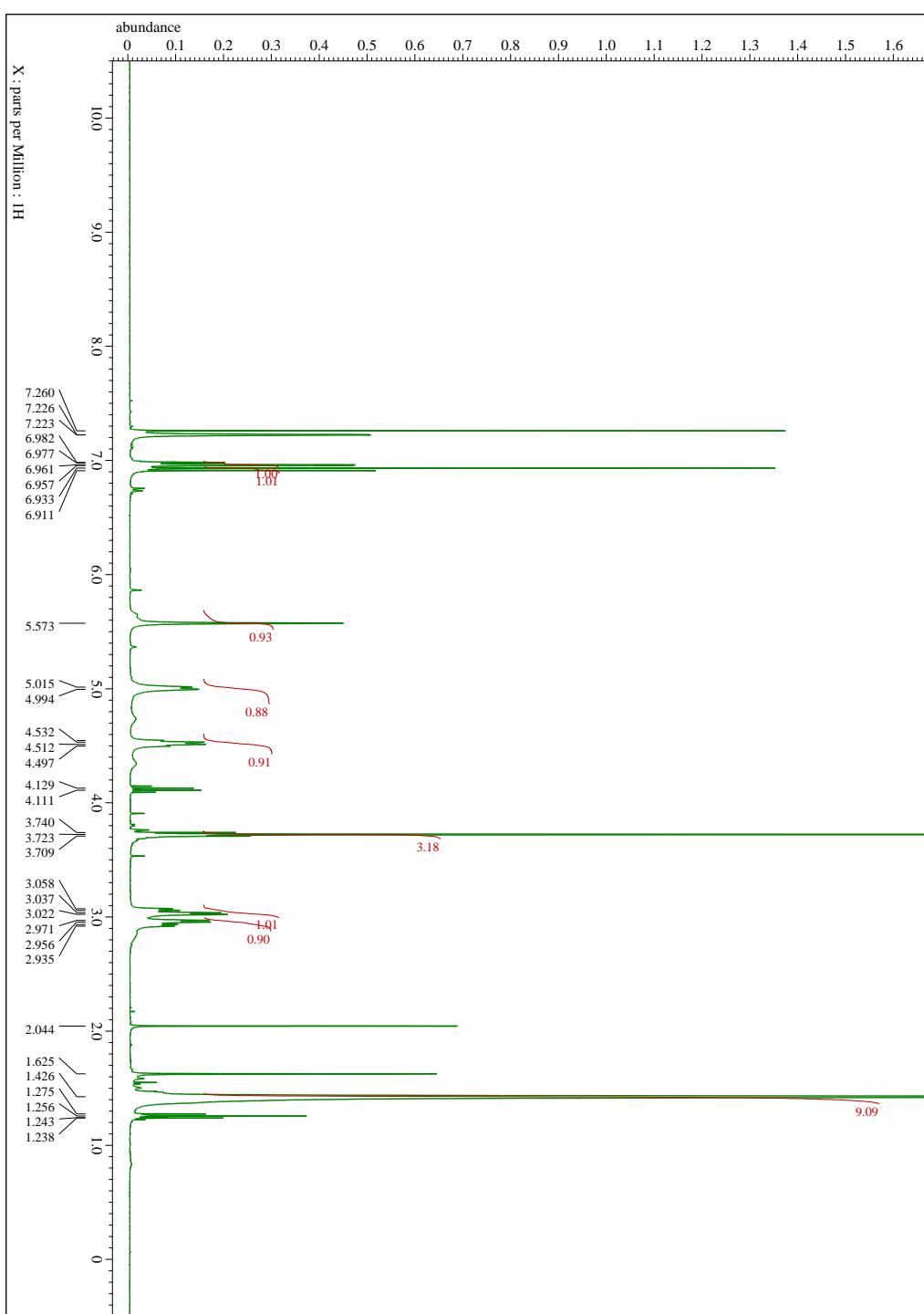
¹³C NMR



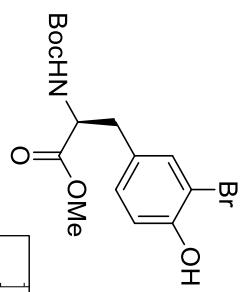
Methyl (S)-3-(3-bromo-4-hydroxyphenyl)-2-((*tert*-butoxycarbonyl)amino)propanoate (3)



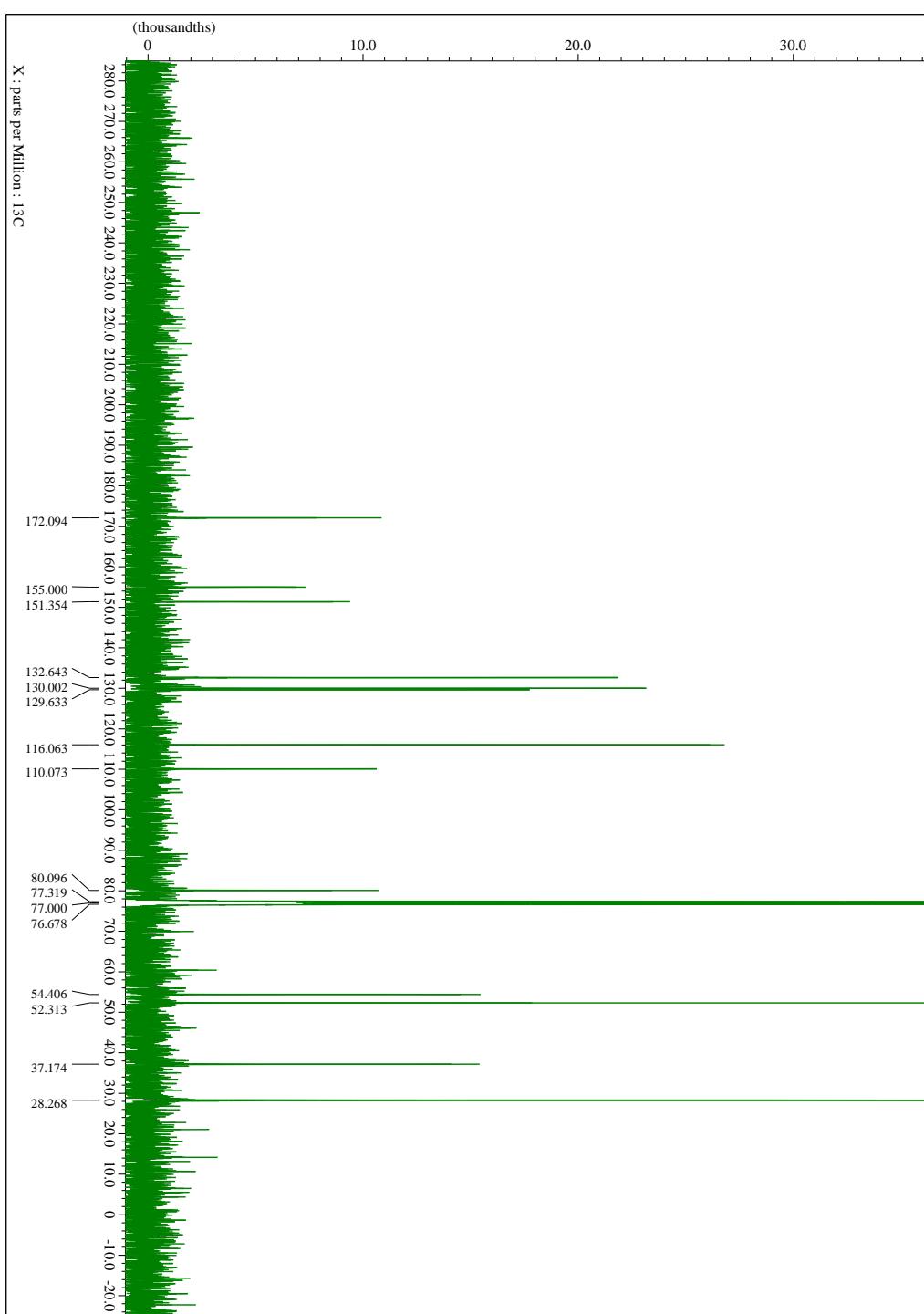
^1H NMR



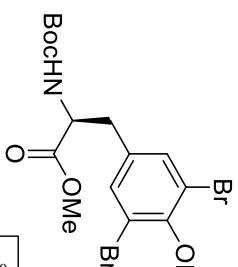
Methyl (S)-3-(3-bromo-4-hydroxyphenyl)-2-((*tert*-butoxycarbonyl)amino)propanoate (3)



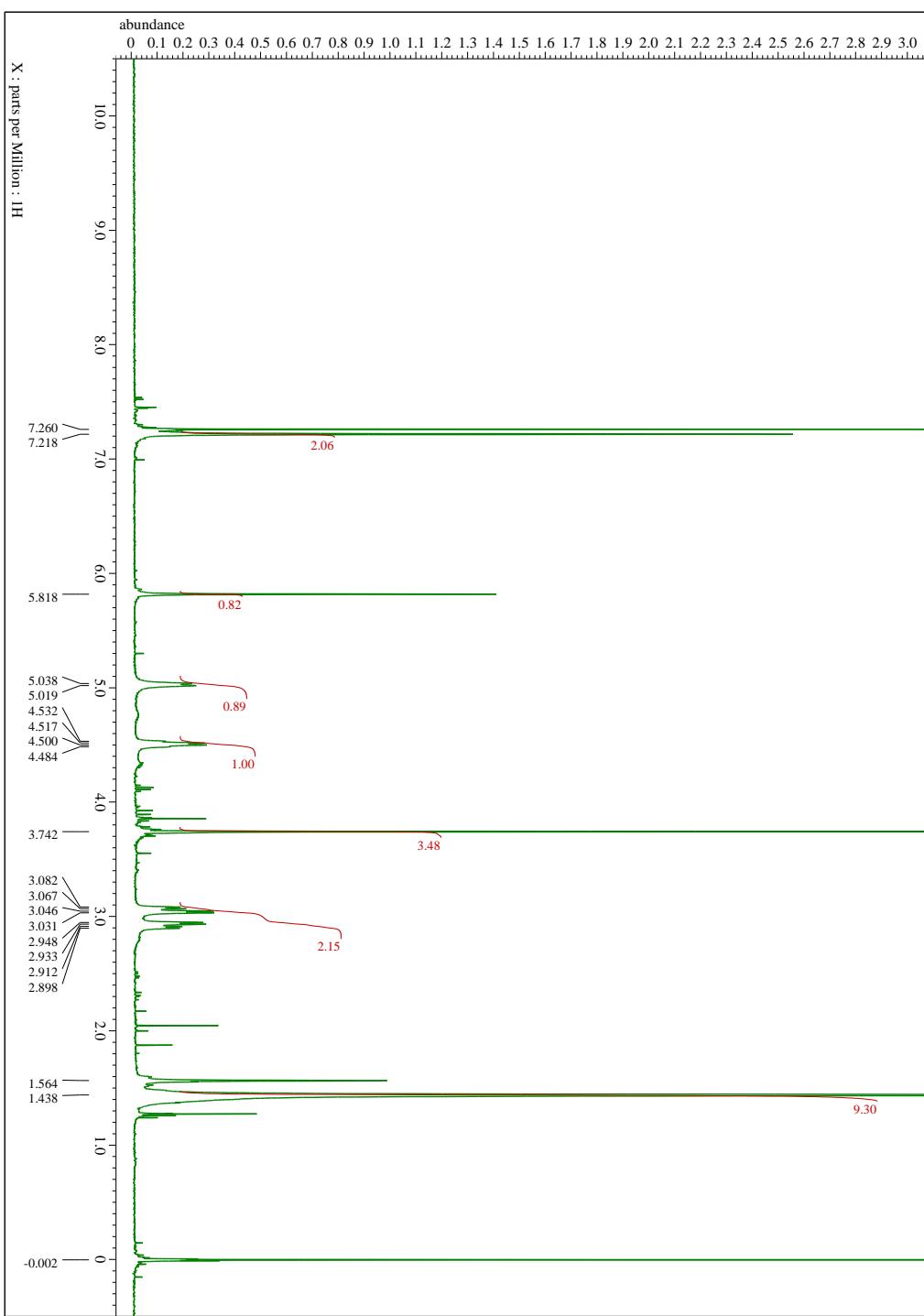
^{13}C NMR



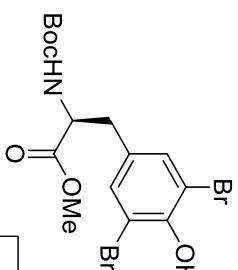
Methyl (S)-2-((*tert*-butoxycarbonyl)amino)-3-(3,5-dibromo-4-hydroxyphenyl)propanoate (4)



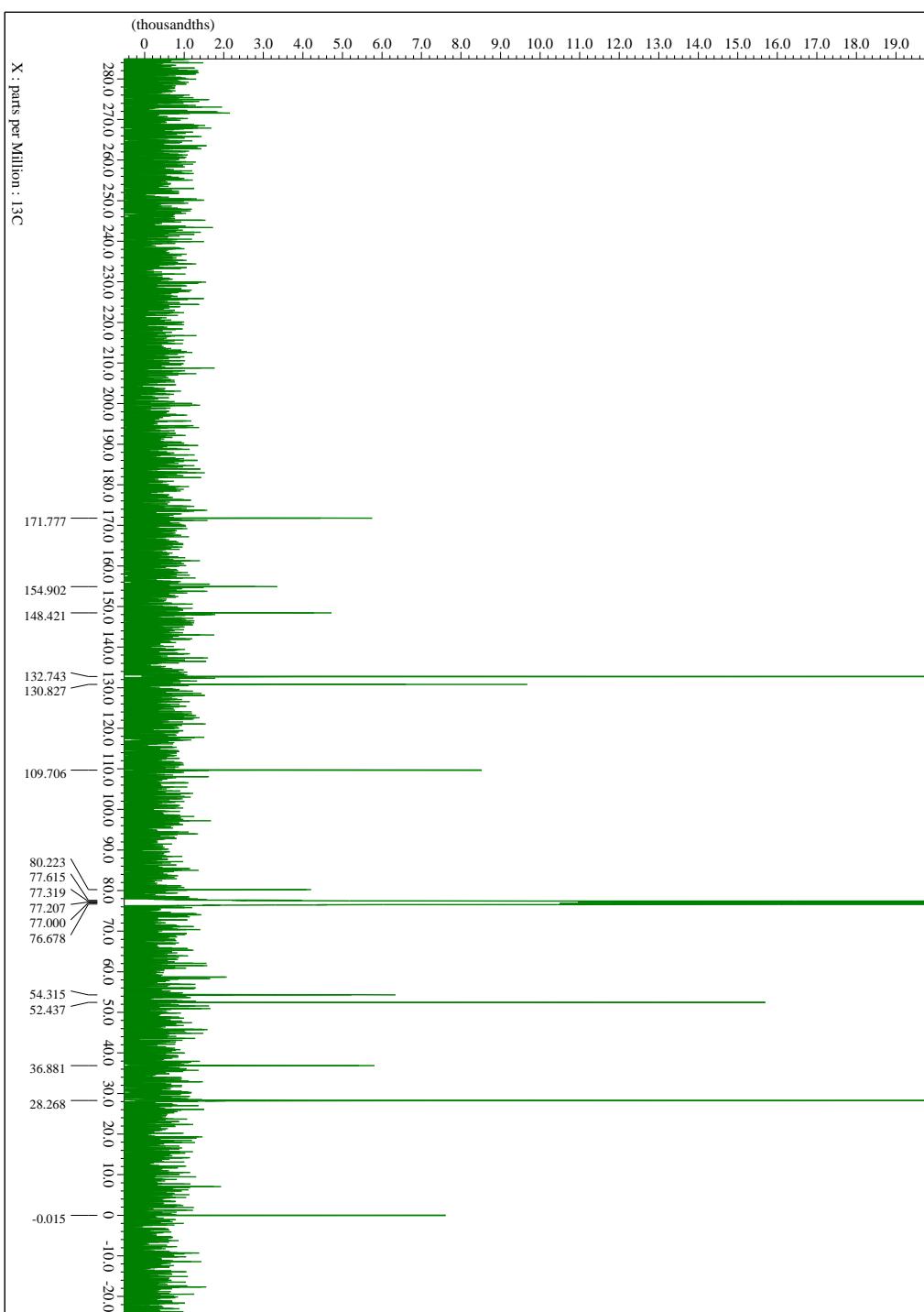
¹H NMR



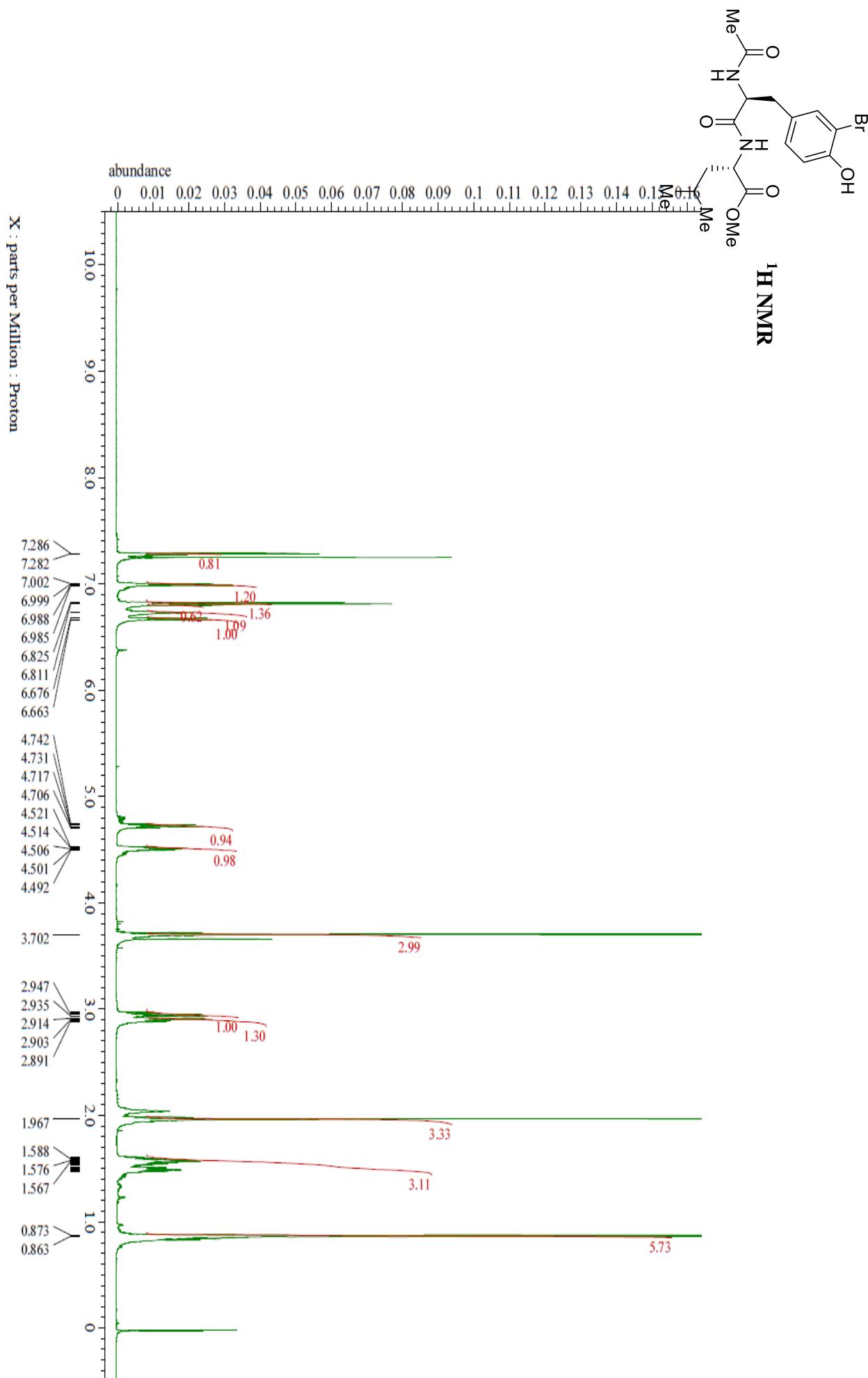
Methyl (S)-2-((*tert*-butoxycarbonyl)amino)-3-(3,5-dibromo-4-hydroxyphenyl)propanoate (4)



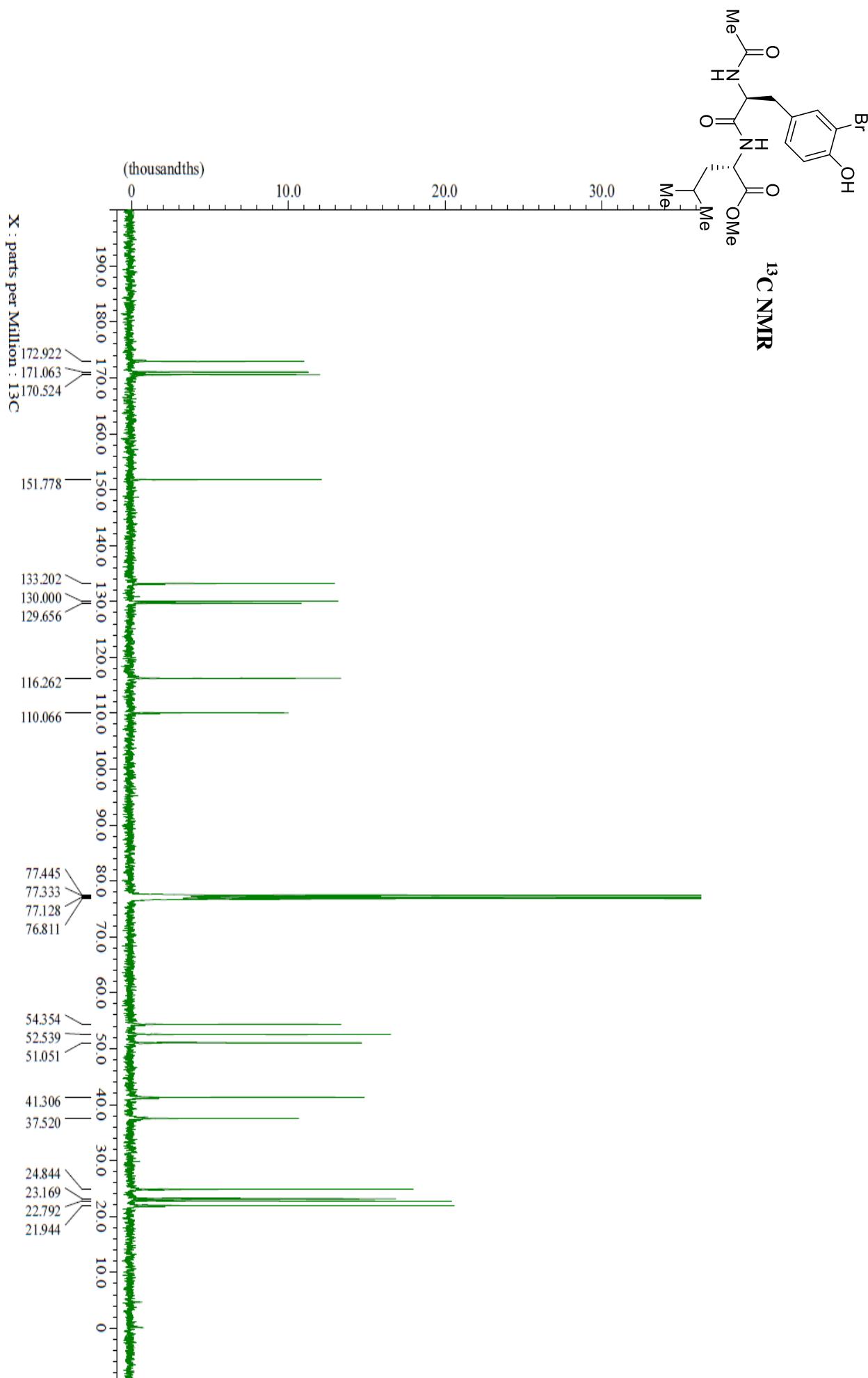
¹³C NMR



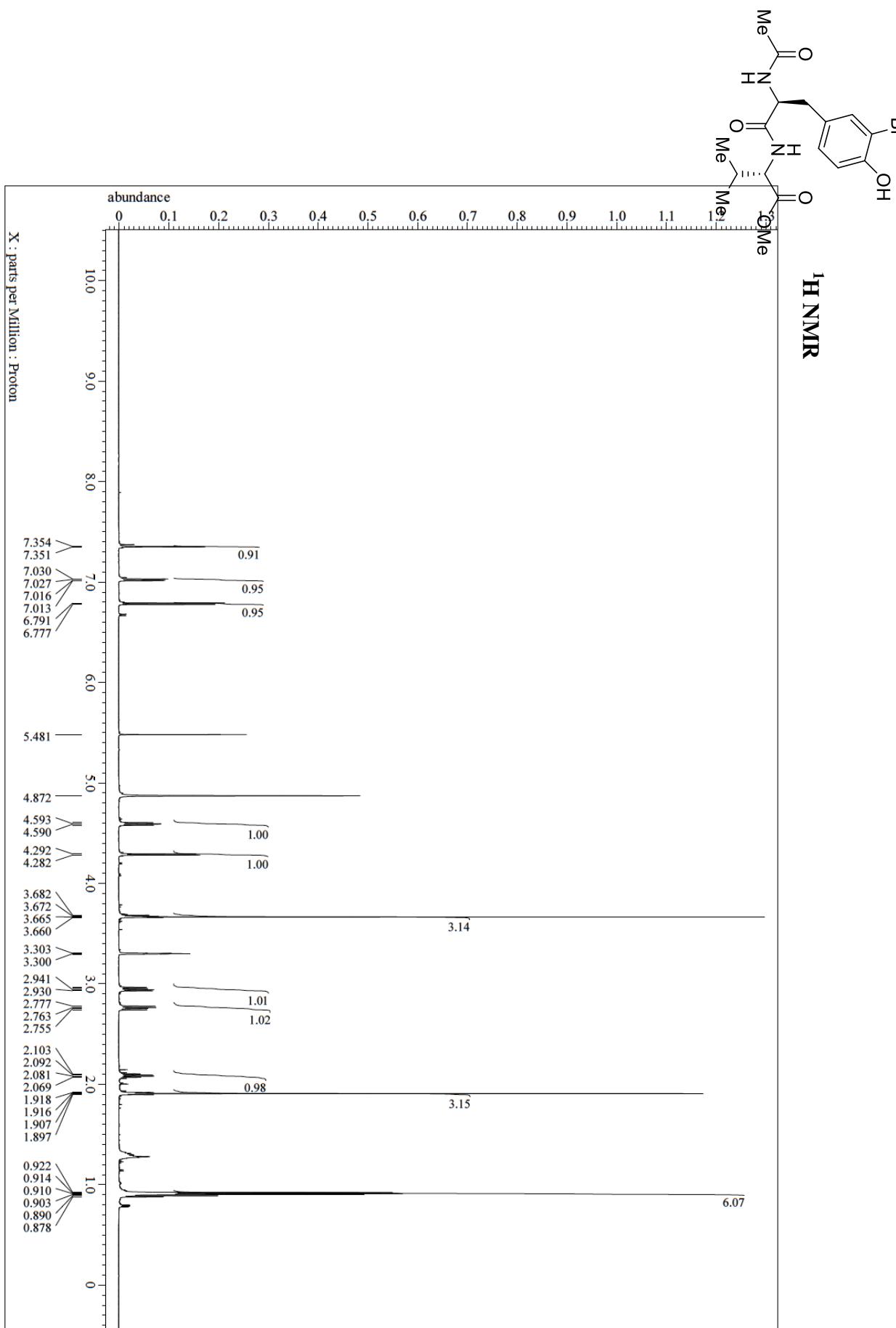
Methyl ((S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl)-L-leucinate (7a)



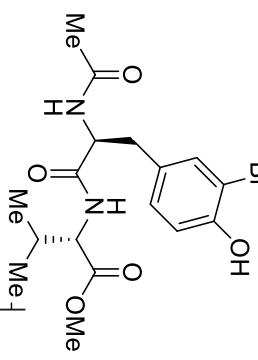
Methyl ((S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl)-L-leucinate (7a)



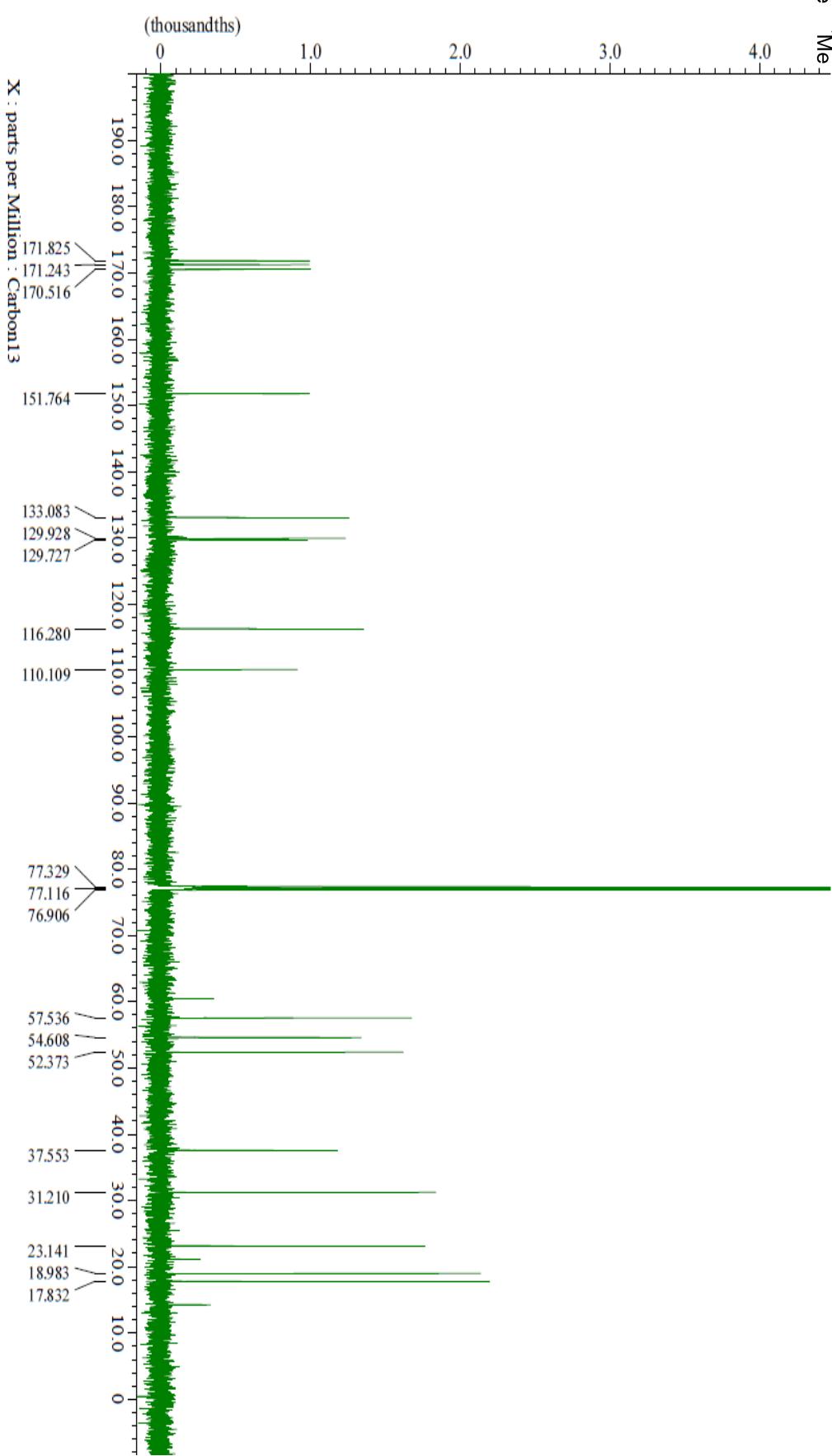
Methyl ((S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl)-L-valinate (7b)



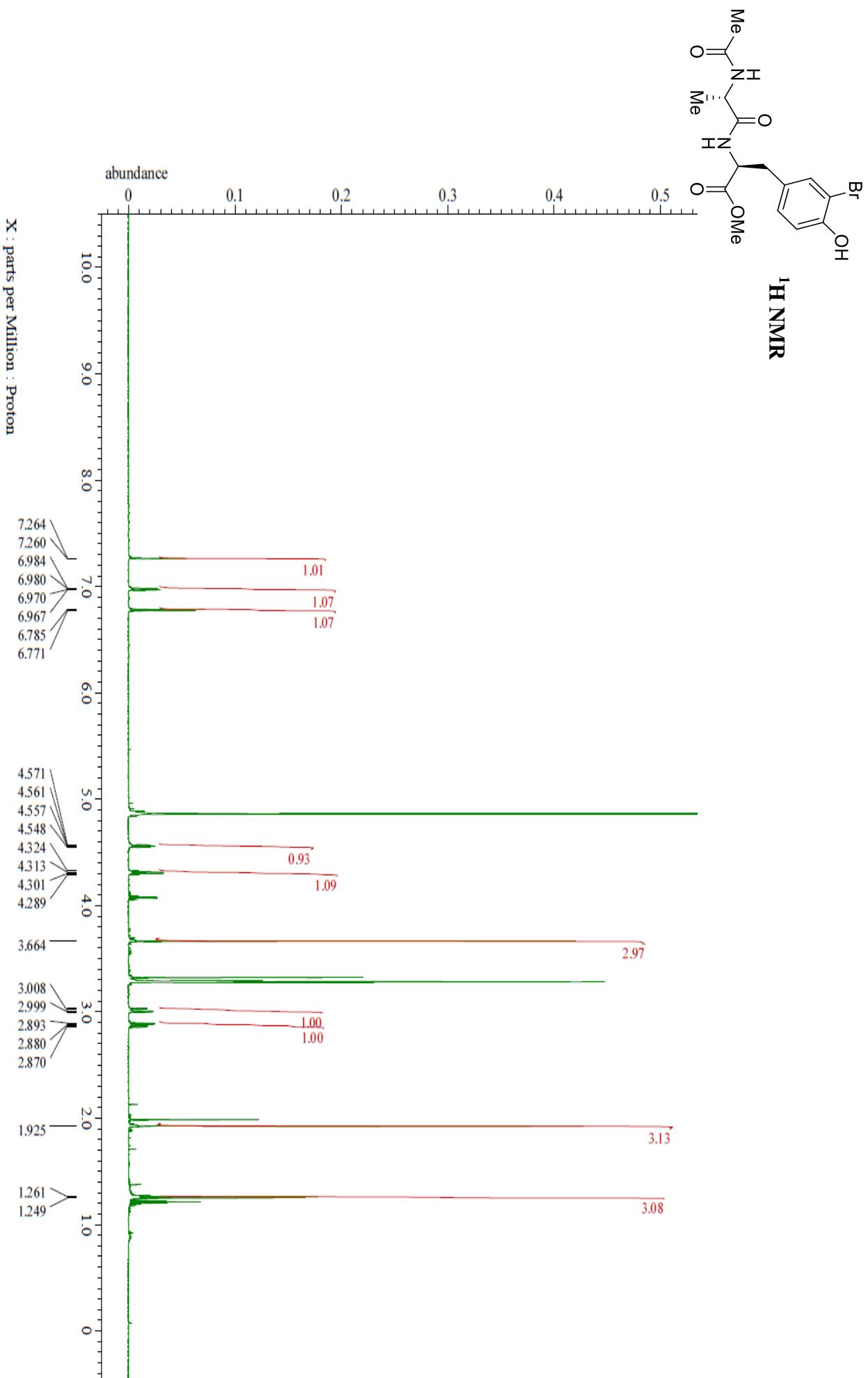
Methyl ((S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl)-L-valinate (7b)



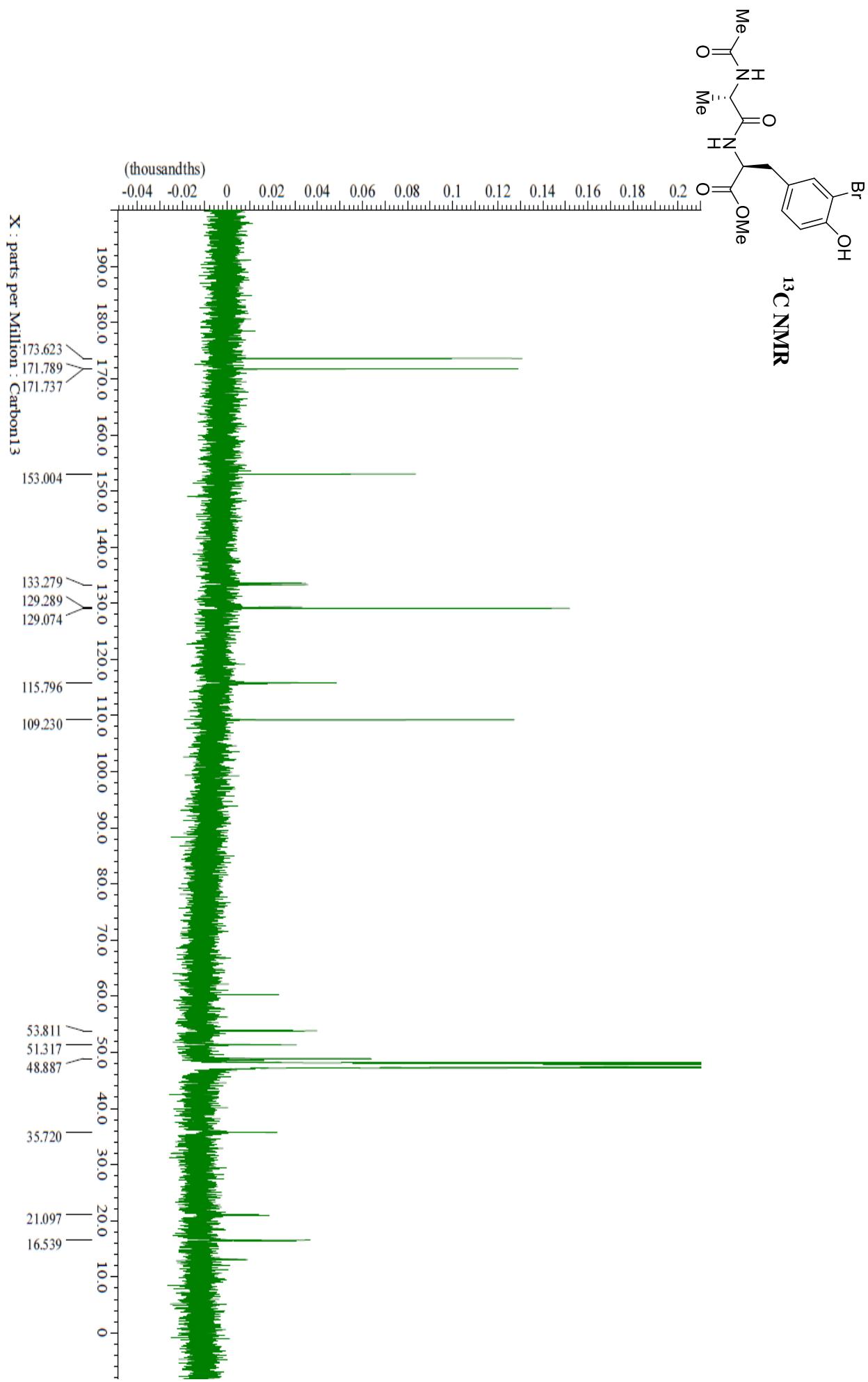
¹³C NMR



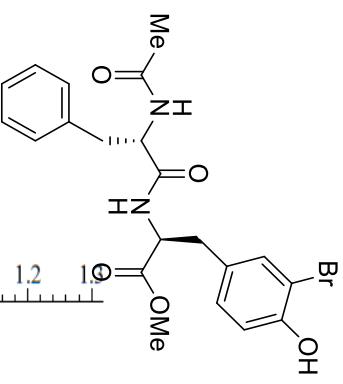
Methyl (S)-2-((S)-2-acetamidopropanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7c)



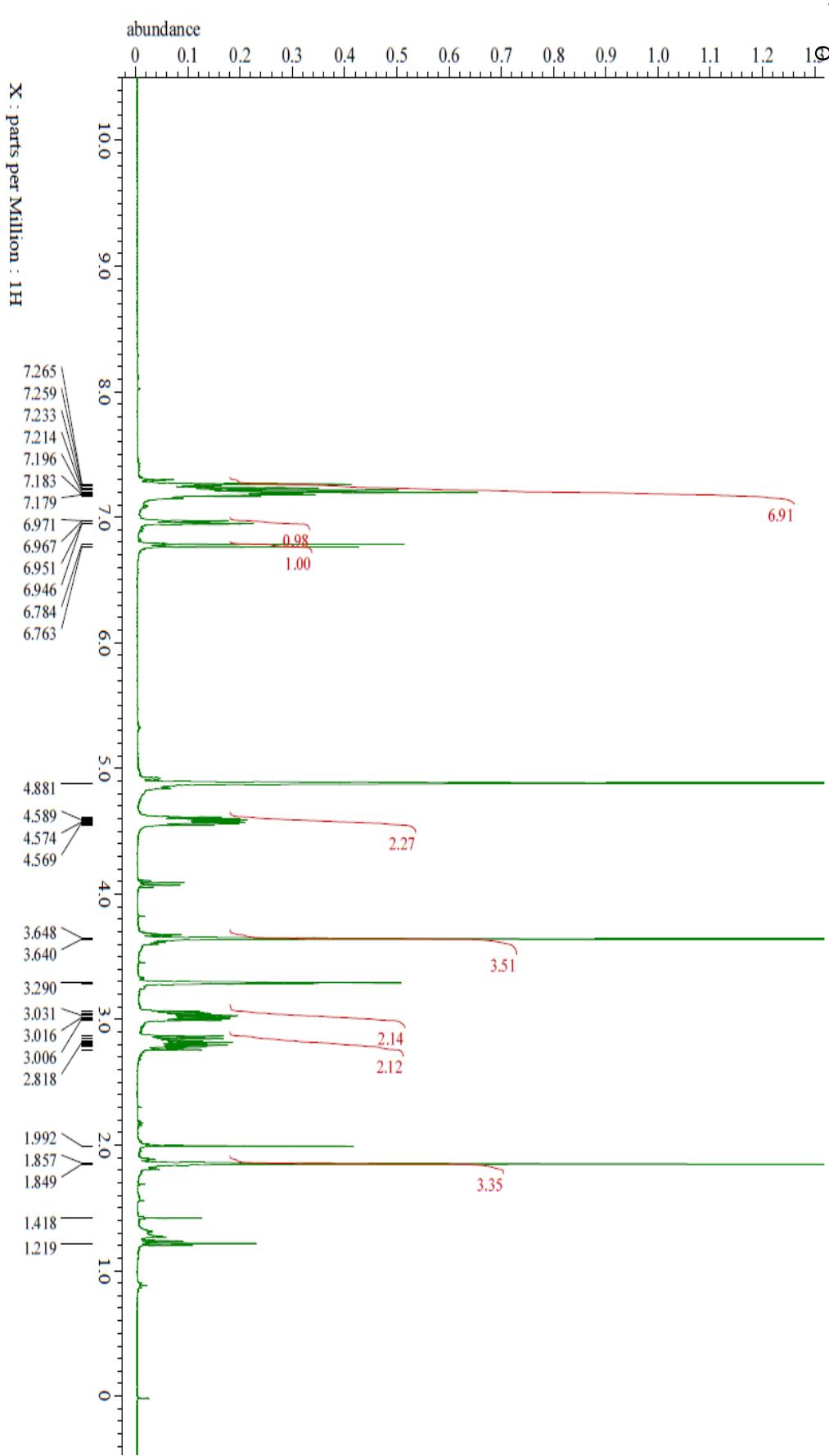
Methyl (S)-2-((S)-2-acetamidopropanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7c)



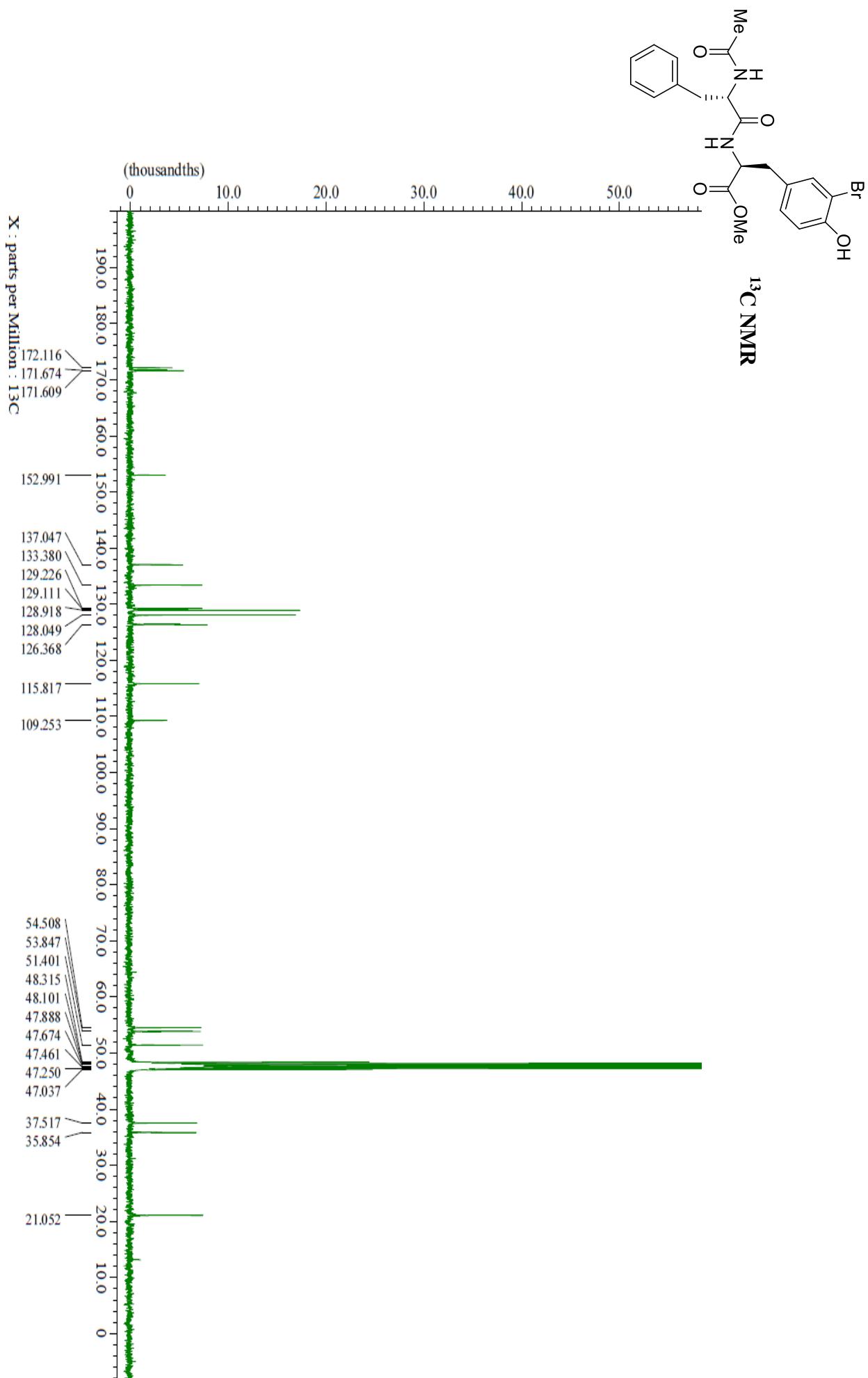
Methyl (S)-2-((S)-2-acetamido-3-phenylpropanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7d)



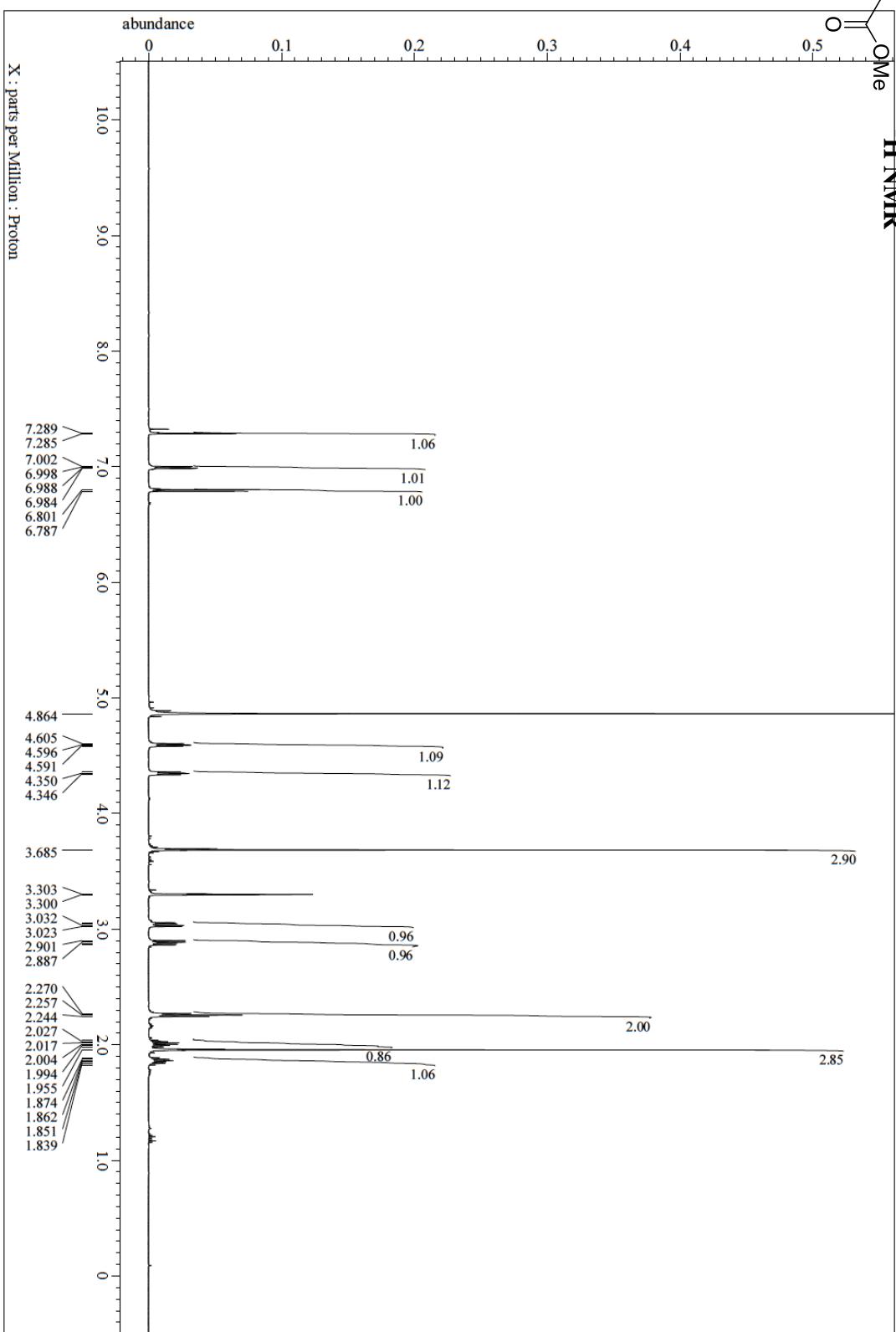
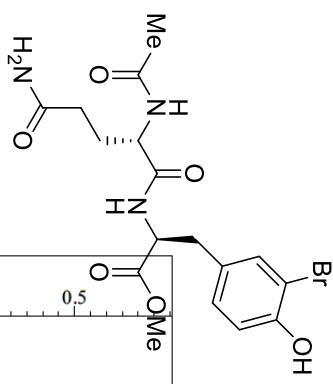
¹H NMR



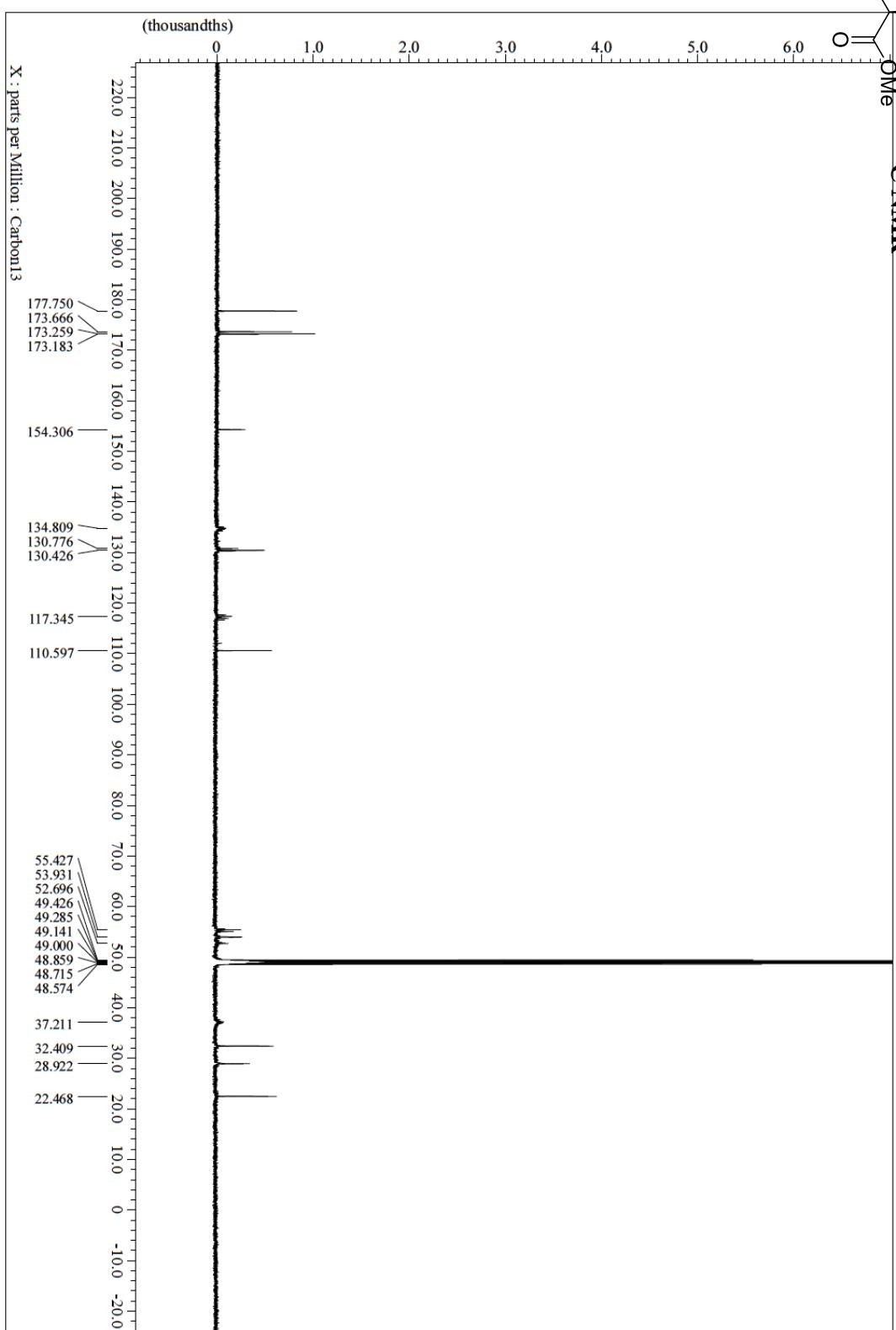
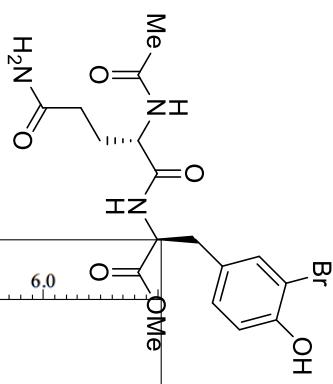
Methyl (S)-2-((S)-2-acetamido-3-phenylpropanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7d)



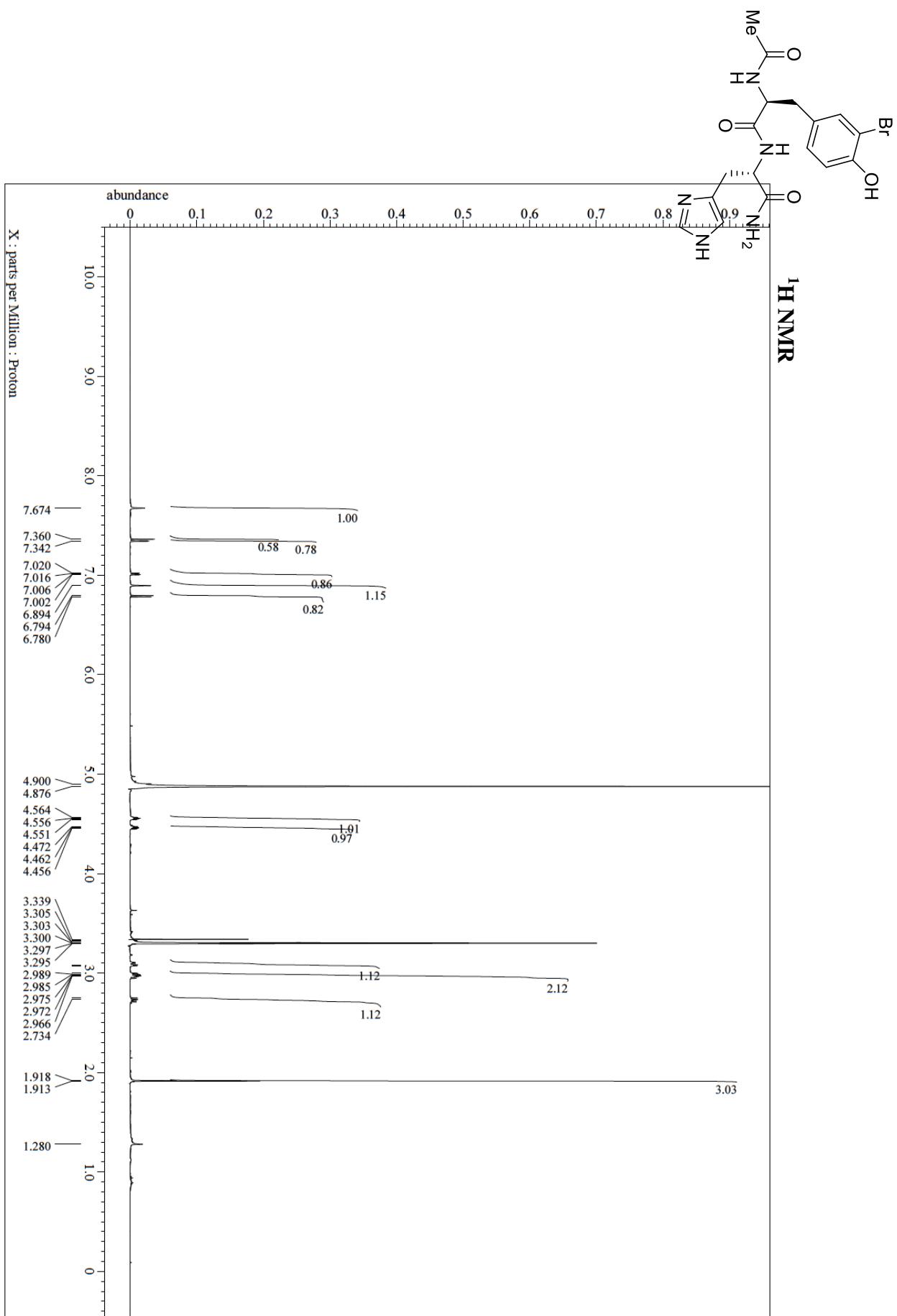
Methyl (S)-2-((S)-2-acetamido-5-amino-5-oxopentanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7e)



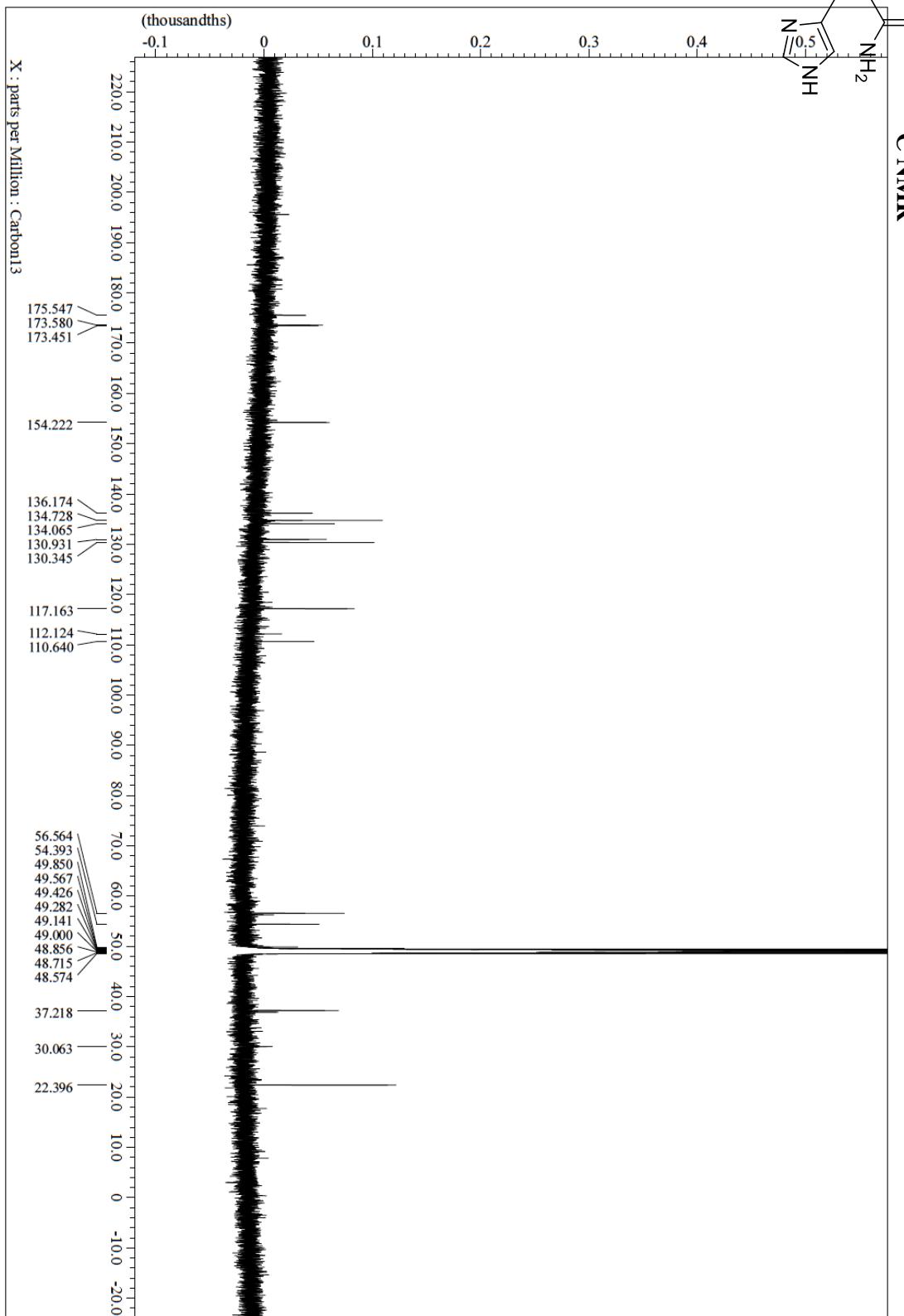
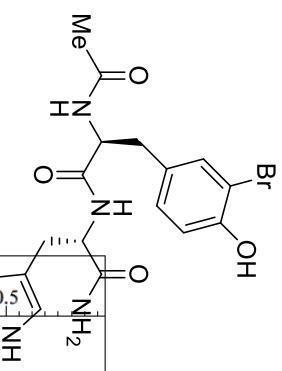
Methyl (S)-2-((S)-2-acetamido-5-amino-5-oxopentanamido)-3-(3-bromo-4-hydroxyphenyl)propanoate (7e)



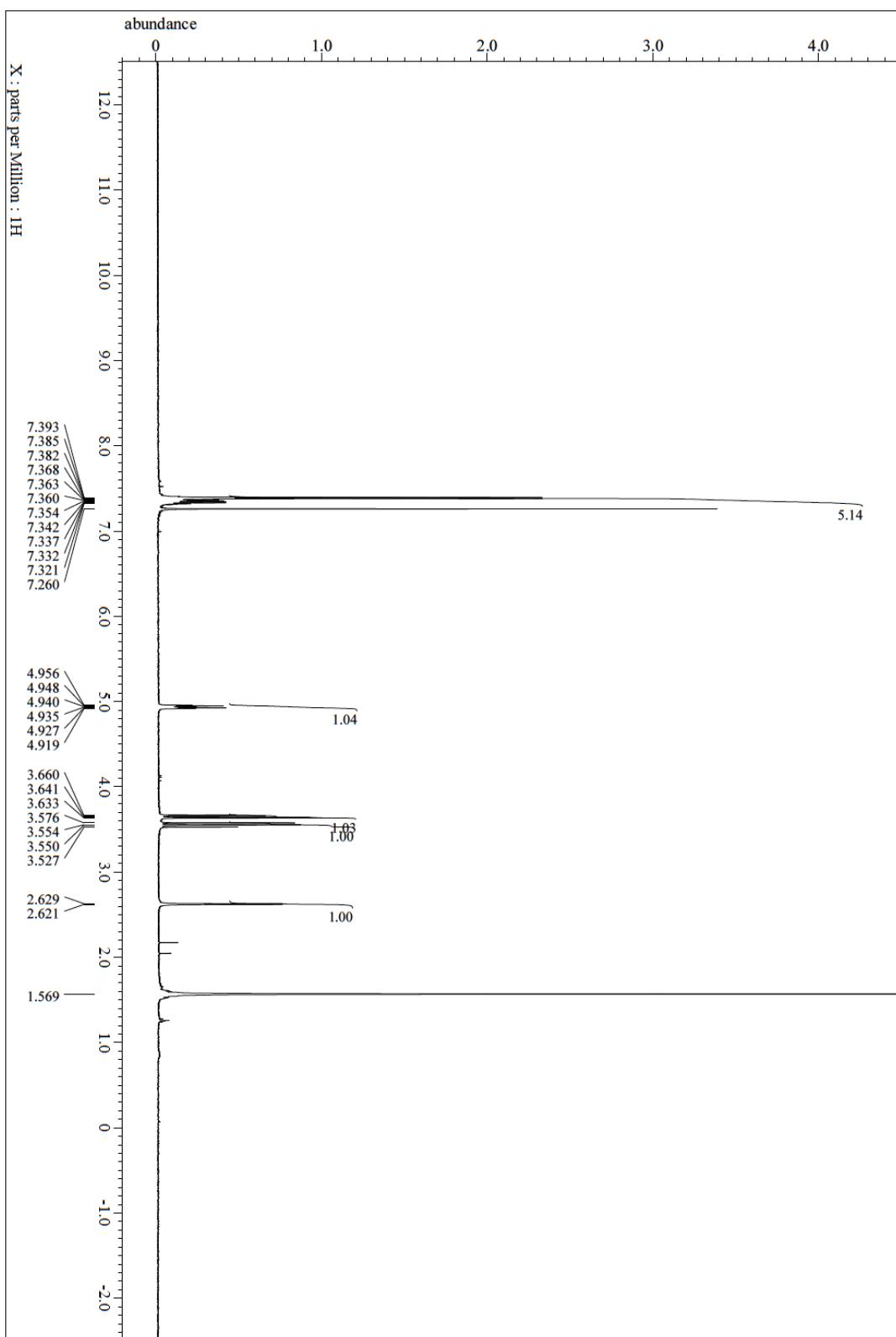
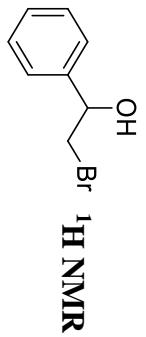
((S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl)-L-histidinamide (7f)



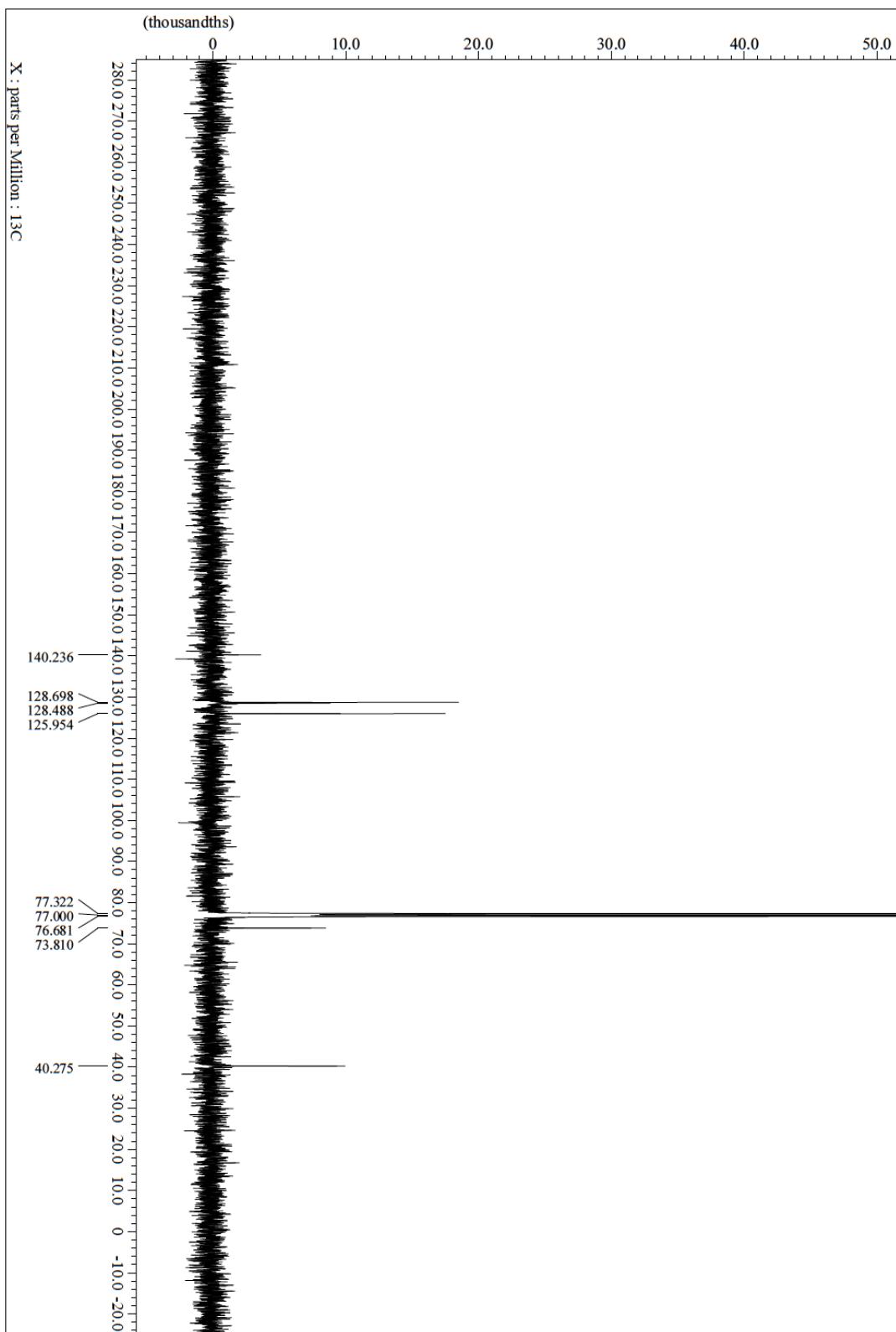
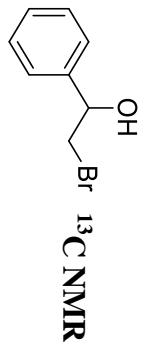
(S)-2-acetamido-3-(3-bromo-4-hydroxyphenyl)propanoyl-L-histidinamide (7f)



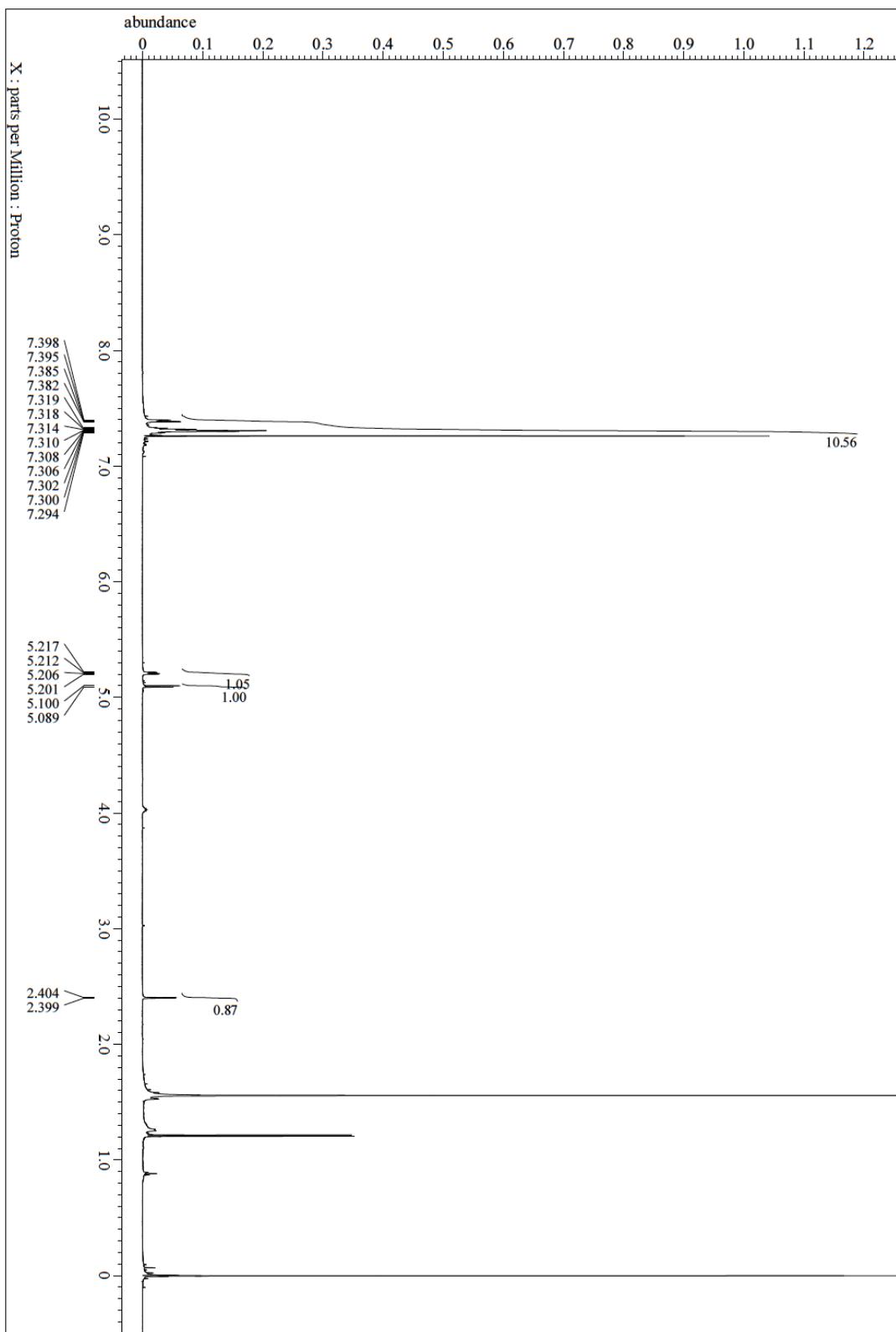
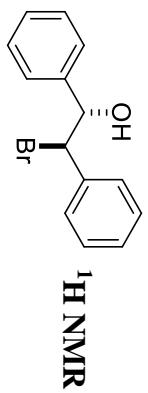
2-Bromo-1-phenylethan-1-ol (9a)



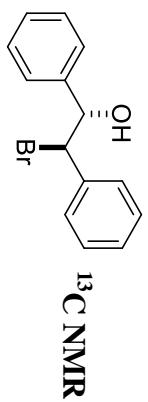
2-Bromo-1-phenylethan-1-ol (9a)



(1*S*^{*},2*R*^{*})-2-Bromo-1,2-diphenylethan-1-ol (9b)



(1*S*^{*},2*R*^{*})-2-Bromo-1,2-diphenylethan-1-ol (9b)



¹³C NMR

