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Supplementary Material

Regulating the Work Function and Surface Hydrophobicity of Indium Tin Oxide Electrodes for Enhanced Electrochemiluminescence Analysis

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S1. Reagents and materials

All chemicals of analytical grade or higher were used as received without further purification. Ultrapure water (18.2 M Ω cm) was employed to prepare aqueous solutions. Sodium dihydrogen phosphate dihydrate (NaH₂PO₄·2H₂O, 99.0%), disodium hydrogen phosphate dodecahydrate (Na₂HPO₄·12H₂O, 99.0%) and potassium chloride (KCl,99.0%) were bought from Sinopharm Chemical Reagent. Tri-*n*-propylamine (TPrA,99%) was purchased from Aladdin. Tris(2,2'-bipyridyl)ruthenium dichloride hexahydrate (Ru(bpy)₃Cl₂·6H₂O, 99.95%) was obtained from Sigma-Aldrich. Indium tin oxide (ITO) coated glass (surface resistivity <17 Ω /cm², thickness 100 ± 20 nm) were purchased from Suzhou Refor Biomedical Co.,Ltd. Amino-functionalized polystyrene bead (3.8 µm, 2.5% w/v, PSB) was purchased from Tianjin Baseline Chrom Tech Research Centre.

S2. Instruments and measurements

All electrochemical measurements were conducted on an electrochemical workstation (CHI920, Shanghai) in a classic three-electrode configuration. Synchronous acquisition of ECL intensity and electrochemical current was performed on a commercial MPI-E ECL analytical system or a custom system consisting of a photon-counting detector and counting unit (Hamamatsu Photonics Co., Ltd., China). ECL and photoluminescence (PL) imaging was performed on a Nikon ECLIPSE microscope (LV100ND) equipped with a matching objective lens (Nikon CFI Apo 40×, N.A. 0.8) and an EMCCD camera (Andor iXon Ultra 897). In a typical experiment, 100 μ L of Ru-PSB suspension was firstly diluted with 900 μ L of ultrapure water and then 20 μ L of the suspension was deposited onto the electrode. The exposure time for PL imaging of single Ru-PSBs was 0.2 seconds. For ECL imaging, the exposure time was 5 seconds, during which a constant potential of +1.5 V (vs. Ag/AgCI) was applied.

The X-ray photoelectron spectra (XPS) were measured on Thermo Scientific ESCALAB 250Xi, utilizing an Al K α light source at 1486.6 eV. The relative surface potential was measured using Kelvin probe force microscopy (KPFM) from Bruker.

S3. Preparation of Ru-PSBs

100 μ L of PSB suspension was washed with 900 μ L of 0.05 M Tris buffer (pH 7.4, Tris), followed by centrifugation at 12000 rpm for 5 minutes. The washed PSBs were then resuspended in an additional 900 μ L of the same Tris buffer. Subsequently, a mixture containing 19.17 mg of EDC and 10 mg of Ru(bpy)₂(cpbpy)(PF₆)₂ dissolved in 100 μ L of DMSO was added to the PSB suspension. This mixture was incubated at room temperature for six hours under magnetic stirring at a speed of 400 rpm. After incubation, the PSBs were washed sequentially with 1 mL of PB (0.2 M, pH 7.4) solution and ultrapure water for ten and three times, respectively. The beads were separated from the solution via centrifugation at a speed of 12000 rpm for five minutes. Finally, Ru(bpy)₃²⁺-labeled PSB (Ru-PSB) was suspended in one milliliter of ultrapure water and stored at 4 °C.

S4. ECL routes



Fig. S1. Different pathways for ECL generation of the $Ru(bpy)_{3}^{2+}/TPrA$ system: (a) the oxidativereduction route, (b) the low oxidation potential (LOP) route and (c) the catalytic route. Ru^{2+} , Ru^{3+} and Ru^{2+*} represent $Ru(bpy)_{3}^{2+}$, $Ru(bpy)_{3}^{3+}$ and $Ru(bpy)_{3}^{2+*}$, respectively.

S5. Preparation and characterization of modified ITO Electrodes

Preparation of CI-ITO electrode: The ITO electrode was immersed in a mixture of methanol (12 mL) and trichloromethane (28 mL) containing 1 mM ethephon for 16 h. The ITO electrode was then rinsed with a mixture of methanol and trichloromethane (7:3 v/v) and dried with N_2 stream.

Preparation of PDMS-ITO electrode: As reported previously,^{S1, S2} The ITO electrode was positioned on top of a PDMS monolith with a distance of ca. 2 mm using glass spacers. The assembly was then heated at 100 °C for different durations to deposit PDMS oligomers on the surface of ITO electrode.

Preparation of CIP-ITO electrode:



Fig. S2. XPS (a) and FT-IR (b) characterization of CI-ITO electrode.

Table S1. Relative surface potential ($\Delta V = V_{\text{sample}} - V_{\text{bare ITO}}$) of Cl-ITO and PDMS-ITO electrode

	CI-ITO	PDMS-ITO	CIP-ITO
$\Delta V / mV$	160.5	- 15.4	96.5



Fig. S3. Variation of the contact angle of ITO electrode with the duration of PDMS modification.



Fig. S4. XPS (a) and MALDI-TOF (b) characterization of PDMS-ITO electrode.

S6. Calculation of the standard electrochemical oxidation rate constants

The electrochemical oxidation of TPrA on the ITO electrode is too slow to show a current peak. In this case, k^0_{TPrA} can be estimated using the following equations according to the Tafel plot method, ^{S3, S4}

$$i_{a} = i^{0} \exp\left[(1 - \alpha)\frac{nF}{RT}(E - E^{0})\right]$$
(S1)
$$\ln(i_{a}) = \ln(i^{0}) + (1 - \alpha)\frac{nF}{RT}(E - E^{0})$$
(S2)

$$i^0 = nFk^0[\text{TPrA}] \tag{S3}$$

where i_a and i^0 represent the anodic current density and exchange current density. *F*, *R*, *T*, and α are the Faraday constant, gas constant, temperature, and charge transfer coefficient, respectively. k^0 and E^0 are the standard rate constant of electrochemical oxidation and the standard redox potential. Considering the acid-base equilibrium of TPrA, the molecular concentration in the actual electrochemical reaction (deprotonated TPrA) was calculated in terms of the relationship between pK_a (10.4) and experimental pH (7.4). In this case, the amount of electroactive TPrA was roughly 0.1% of the analytical concentration. In the case of Ru(bpy)₃²⁺, the standard rate constant of electrochemical oxidation (k^0_{Ru}) can be derived according to the Nicholson equation,

$$k^{0} = \psi \left(\frac{\pi D n F v}{RT}\right)^{\frac{1}{2}}$$
(S4)

where D (5.9 × 10⁻⁶ cm²/s) is the diffusion coefficient of Ru(bpy)₃²⁺ and v is the potential scan rate. The value of ψ refers to the table given by Nicholson.^{S5}

Electrodes	Bare ITO	CI-ITO	PDMS-ITO
$k^{0}_{\rm TPrA} imes 10^{6} / {\rm cm} \cdot {\rm s}^{-1}$	4.31±0.34	9.99 ± 1.54	73.23 ± 7.53

Table S2. k^{0}_{TPrA} at different electrodes



Fig. S5. (a) CVs of different electrodes in 0.2 M PB (pH 7.4) solution containing 0.5 mM Ru(bpy)₃²⁺ (b) A comparison of k_{Ru}^0 for three different electrodes. Standard deviation, n = 3

S7. Calibration of the bias voltages of PMT



Fig. S6. The calibration of bias voltages of PMT (450 V and 600 V) was conducted using ITO electrode in 0.2 M PB (pH 7.4) solution containing 5 μ M Ru(bpy)₃²⁺ and 5 mM TPrA.

S8. Finite element simulations

If applying a sufficiently positive potential, we can assume that the ECL process is dominated by the "oxidative reduction route" and "catalytic route", which include the following elementary steps, where k_i represents the rate constant of each step and P is $(CH_3CH_2CH_2)_2^{N+} = CHCH_2CH_3^{-S6-S8}$

$$\mathsf{TPrA} + \mathsf{H}^+ \underbrace{\overset{k_1}{\longleftarrow}}_{k_2} \mathsf{TPrAH}^+$$
(S5)

$$TPrA \xrightarrow{k_2} TPrA^{+\bullet} + e^{-1}$$
(S6)

$$\mathsf{TPrA}^{+\bullet} \xrightarrow{k_3} \mathsf{TPrA}^{\bullet} + \mathsf{H}^{+} \tag{S7}$$

$$TPrA^{\bullet} \xrightarrow{k_{4}} P_{0} + e^{-}$$
(S8)
$$Ru(bpy)_{2}^{2+} + TPrA^{\bullet} \xrightarrow{k_{5}} Ru(bpy)_{2}^{+} + P_{1}$$
(S9)

$$\operatorname{Ru}(\operatorname{bpy})_{3}^{*} + \operatorname{TPrA}^{*\bullet} \xrightarrow{k_{6}} \left[\operatorname{Ru}(\operatorname{bpy})_{3}^{2^{+}}\right]^{*}$$
(S10)

$$\left[\operatorname{Ru}(\operatorname{bpy})_{3}^{2^{+}}\right]^{*} \xrightarrow{k_{7}} \operatorname{Ru}(\operatorname{bpy})_{3}^{2^{+}} + h\nu \tag{S11}$$

$$\operatorname{Ru}(\operatorname{bpy})_{3}^{2+} \xrightarrow{k_{g}} \operatorname{Ru}(\operatorname{bpy})_{3}^{3+} + e^{-}$$
(S12)

$$Ru(bpy)_{3}^{3+} + TPrA \xrightarrow{k_{g}} Ru(bpy)_{3}^{2+} + TPrA^{+\bullet}$$
(S13)

$$\operatorname{Ru}(\operatorname{bpy})_{3}^{3+} + \operatorname{TPrA}^{\bullet} \xrightarrow{k_{10}} \left[\operatorname{Ru}(\operatorname{bpy})_{3}^{2+} \right]^{*} + \operatorname{P}_{2}$$
(S14)

$$Ru(bpy)_{3}^{3+} + Ru(bpy)_{3}^{+} \xrightarrow{k_{32}} \left[Ru(bpy)_{3}^{2+} \right]^{*} + Ru(bpy)_{3}^{2+}$$
(S15)

Name	Value	Description
r _e	250 μm	Length of the radial electrode
L	200 μm	Length of the axial solution boundary
<i>c</i> ₀ _Ru ²⁺	500 μΜ, 50 μΜ, 1 μΜ	Initial concentration of Ru(bpy) ₃ ²⁺
c _{o_} TPrA	(1 or 200) × 0.001 mM	Initial concentration of TPrA
c ₀ _TPrAH⁺	(1 or 200) × 0.999 mM	Initial concentration of TPrAH ⁺
<i>с</i> ₀_Н	10 ^{-7.4} M	Initial concentration of H ⁺
D _{Ru}	$5 imes 10^{-6}$ (cm²/s)	Diffusion coefficients of Ru(bpy) ₃ ²⁺ , Ru(bpy) ₃ ³⁺ and Ru(bpy) ₃ ^{2+*}
D _{TPrA}	5 × 10 ^{−6} (cm²/s)	Diffusion coefficient of TPrA, TPrAH⁺, TPrA⁺• and TPrA•
D _H	$5 imes 10^{-5}$ (cm ² /s)	Diffusion coefficient of H ⁺
<i>k</i> ₁	$k_{0r} \times e^{38.92 \times (1-\alpha_1) \times (E-1.05)}$	Forward rate constant of eq (S5)
<i>k</i> _1	$k_{0r} \times e^{38.92 \times (-\alpha_1) \times (E-1.05)}$	Backward rate constant of eq (S5)
k ₂	0.01, 0.2 (cm/s)	Rate constant of eq (S6)
<i>k</i> ₃	3500 (s ⁻¹)	Rate constant of eq (S7)
<i>k</i> 4	k ₂	Rate constant of eq (S8)
<i>k</i> 5	300 m³/(s·mol)	Rate constant of eq (S9)
<i>k</i> ₆	300 m³/(s·mol)	Rate constant of eq (S10)
k ₇	$3.3 imes 10^{6} \ (s^{-1})$	Rate constant of eq (S11)
k ₈	0.1, 0.3 m/s	Rate constant of eq (S12)
<i>k</i> 9	1000 m³/(s·mol)	Rate constant of eq (S13)
k ₁₀	1× 10 ⁶ m³/(s⋅mol)	Rate constant of eq (S14)
k ₁₁	10000 m³/(s·mol)	Rate constant of eq (S15)

Table S3. Definitions of simulation parameters

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