

## **Endogenous Oxygen Self-Supplied Nanoplatform with GSH-Depleted and NIR-II Triggered Electron-hole Separation for Enhance Photocatalytic Anti-tumor Therapy**

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## **Materials and Methods**

Copper chloride ( $\text{CuCl}_2$ ), sodium sulphide ( $\text{Na}_2\text{S}$ ), Polyvinylpyrrolidone (PVP-K30), Sodium hydroxide ( $\text{NaOH}$ ), hydrazine anhydrous solution ( $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$ ), glucose, acetic acid, hydrogen tetrachloroaurate (III) ( $\text{HAuCl}_4\cdot 3\text{H}_2\text{O}$ ), Sodium borohydride ( $\text{NaBH}_4$ ) was purchased from from Shanghai Macklin Biochemical Co., Ltd. (Shanghai, China). 5, 5'-dithiobis (2-nitrobenzoic acid) (DTNB), glutathione (GSH), 2',7'-dichlorodihydrofluorescein (DCFH-DA) and 1,3-diphenylisobenzofuran (DPBF) were purchased from Sinopharm Chemical Reagent Co., Ltd. All compounds were utilized just as they were supplied, with no additional purification.

## **Characterization**

The crystallographic phase structure was examined via powder X-ray diffraction (XRD, Rigaku D/max 2500 system operating at 40 kV and 250 mA). Transmission electron microscopy was performed by JEM-2100F (JEM-2010, JEOL, Japan) with an accelerating voltage of 200 kV. The UV-Vis adsorption spectral values were measured on a U-3310 spectrophotometer (Hitachi). The surface chemical composition was evaluated by X-ray photoelectron spectrometer (XPS, ESCALab 250Xi, Thermo Fisher Scientific, USA).

## **Synthesis of Cu<sub>7</sub>S<sub>4</sub> nanospheres**

The Cu<sub>7</sub>S<sub>4</sub> nanospheres were synthesized according to previously reported method with minor modification<sup>1</sup>. Briefly, 100  $\mu\text{L}$   $\text{CuCl}_2$  solution (0.5 M) was added

to 25 mL deionized water containing 0.24 g PVP-K30 under magnetic stirring at room temperature. Then, 25 mL NaOH solution (pH=9.0) was added, followed by addition of 6.4  $\mu\text{L}$   $\text{N}_2\text{H}_4\cdot\text{H}_2\text{O}$  (50 %) to form a bright-yellow suspension of  $\text{Cu}_2\text{O}$  spheres. After 5 min, 200  $\mu\text{L}$   $\text{Na}_2\text{S}$  aqueous solution ( $320 \text{ mg mL}^{-1}$ ) was added to the suspension. The solution was heated for 2 h at  $60^\circ\text{C}$ . Finally,  $\text{Cu}_7\text{S}_4$  nanospheres were centrifuged at 10000 rpm for 6 min, and washed three times with deionized water and ethanol. Then, the  $\text{Cu}_7\text{S}_4$  nanospheres were dispersed in 20 mL ethanol.

### **Synthesis of hollow $\text{Cu}_7\text{S}_4@\text{Au}$ nanospheres**

The  $\text{Cu}_7\text{S}_4@\text{Au}$  nanospheres were one-step synthesized according to previously reported literature with some modifications. Briefly, 1.0 mL  $\text{Cu}_7\text{S}_4$  suspension above prepared was dispersed in 1.2 mL ethanol, followed by the addition of 0.01 g PVP-K30. After stirring for 30 min,  $\text{HAuCl}_4\cdot 3\text{H}_2\text{O}$  aqueous solution with different volume (1.6, 3.2, 4.8, 6.4 and 9.6 mL) (0.3 mM) was added and the mixture was stirred for 10 min. Afterwards, 0.5 mL  $\text{NaBH}_4$  (3 mM) was added and the mixture was stirred for another 30 min. The products were collected by centrifugation at 10000 rpm for 5 min and washed three times with ethanol. Then, the  $\text{Cu}_7\text{S}_4@\text{Au}$  nanospheres were dispersed in water.

### **Synthesis of hollow $\text{Cu}_7\text{S}_4@\text{Au}@ \text{MnO}_2$**

1 mL  $\text{Cu}_7\text{S}_4@\text{Au}$  nano-material was mixed with 20 mg  $\text{KMnO}_4$  in 20 mL water and stirred for 10 min. After the solution was changed to brown, the obtained  $\text{Cu}_7\text{S}_4@\text{Au}@ \text{MnO}_2$  nanocatalysts were collected by centrifugation and washed with

water several times. The contents of Cu and Au in  $\text{Cu}_7\text{S}_4@\text{Au}@MnO_2$  are determined by ICP.

Table 1. The weight ratio between Cu, Au and Mn in  $\text{Cu}_7\text{S}_4@\text{Au}@MnO_2$

Cu	Au	Mn
1	0.27	0.09

### **Tumor animal models**

The  $8 \times 10^6$  4T1 cells were subcutaneously injected into the BALB/c mice to establish the 4T1 tumor-bearing mice models. The 4T1 tumor-bearing mice were used for NIR-II light triggered on-demand enhanced photocatalytic therapy.

### ***In vitro* cytotoxicity and NIR-II triggered on-demand enhanced photocatalytic therapy**

*In vitro* cell viability was evaluated by using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) proliferation assay method. The 4T1 cells were first cultured in a 96-well microplate and kept at 37 °C under 5% CO<sub>2</sub> incubation 3 h. For cell toxicity tests, the cells were treated with different concentrations of different nano-drugs for 12 h at 37 °C. Finally, the cell viabilities were tested by using MTT method. For *in vitro* therapy of cancer cells, the cells were divided into four groups:(I) Control, (II)  $\text{Cu}_7\text{S}_4@\text{Au}+\text{NIR-II}$ , (III)  $\text{Cu}_7\text{S}_4@\text{Au}@MnO_2$ , (IV)  $\text{Cu}_7\text{S}_4@\text{Au}@MnO_2+\text{NIR-II}$ . The cells were then cultured at 37 °C for 6 h. Finally, the cell viabilities were tested referring to the MTT method.

### ***In vitro* oxidative stress detection**

The 4T1 cells were first treated with (I) Control, (II) Cu<sub>7</sub>S<sub>4</sub>@Au+NIR-II, (III) Cu<sub>7</sub>S<sub>4</sub>@Au@MnO<sub>2</sub>, (IV) Cu<sub>7</sub>S<sub>4</sub>@Au@MnO<sub>2</sub>+NIR-II incubated for 12 h. Next, the 4T1 cells were stained with DCFH-DA for 30 min. The fluorescence images were then acquired by using a fluorescence microscope.

### **Test of GSH depletion**

The consumption of GSH was monitored by UV-vis spectroscopy. The Cu<sub>7</sub>S<sub>4</sub>@Au@MnO<sub>2</sub> (2 mL, 5 μM) was mixed with (40 μL) GSH (1 mM). At different time points, 40 μL DTNB (10 mg/mL) was added. The absorbance spectrum of the supernatant was measured by UV-vis spectroscopy.

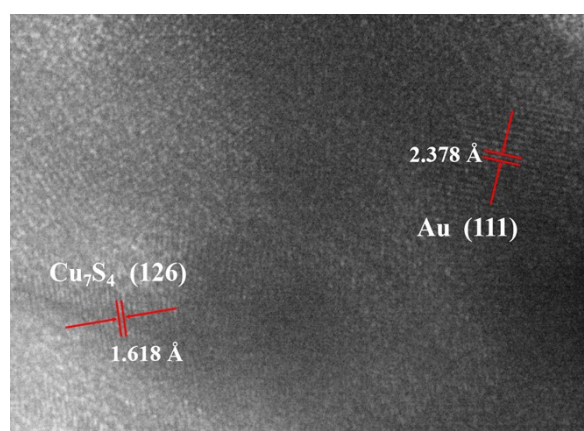
### **Photocurrent test**

The photocurrent properties of the samples were tested using a three-electrode method. The platinum electrode was used as the counter electrode, the Ag/AgCl electrode was used as the reference electrode, and the working electrode was made of a titanium plate encapsulated with a silica gel. The electrolyte solution was Na<sub>2</sub>SO<sub>4</sub> (0.5 M) solution. The samples were tested using a CHI-660 electrochemical workstation, and the results were measured under 1064 nm laser irradiation. The linear sweep voltammetry scan rate is 10 mV s<sup>-1</sup>.

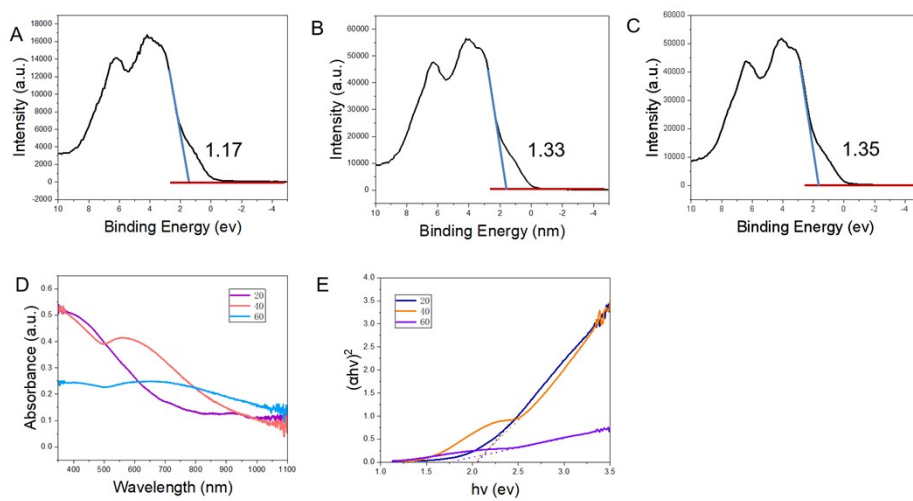
### **Tumor biopsies**

For hematoxylin and eosin (H&E) and Terminal deoxynucleotidyl transferase

dUTP nick end labeling (TUNEL) staining, the tumors of 4T1 tumor-bearing mice and bilateral 4T1 tumor-bearing mice after different treatments were dissected. And then the tumors were stained with H&E and TUNEL for histological test.

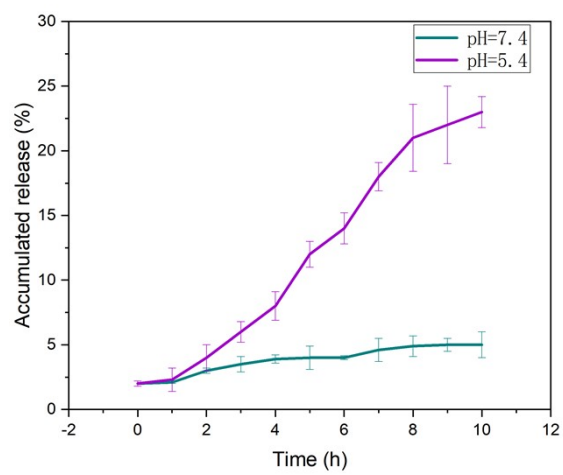


**Figure S1.** The high-resolution images of  $\text{Cu}_7\text{S}_4@Au@MnO_2$ .

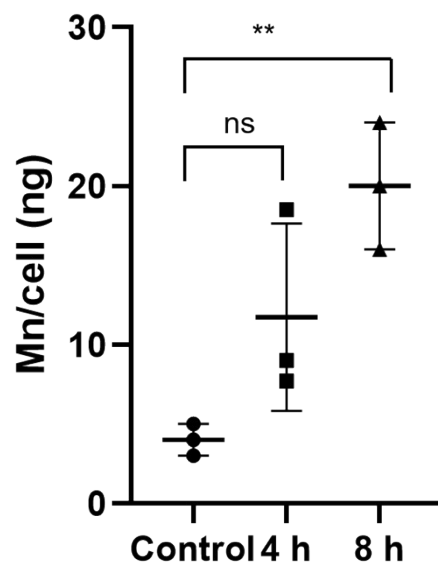


**Figure S2.** VB-XPS spectra, UV-vis spectra and Plot of  $(\alpha hv)^2$  versus photon energy  $(hv)$  of Cu<sub>7</sub>S<sub>4</sub>@Au (HAuCl<sub>4</sub>·3H<sub>2</sub>O aqueous solution with 20, 40 and 60 μL) nanocatalyst.

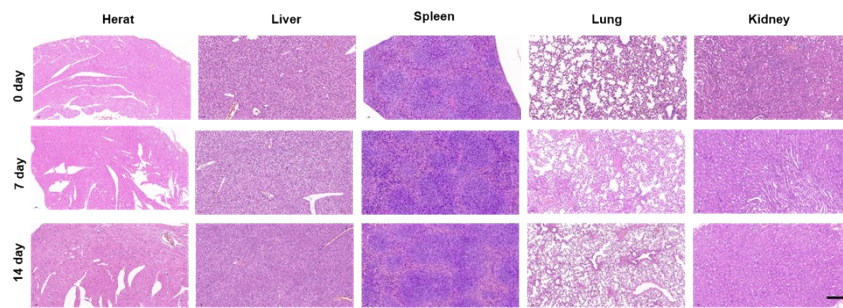




**Figure S3.** The oxygen release ability of  $\text{Cu}_7\text{S}_4@\text{Au}@\text{MnO}_2$  nanomaterials.



**Figure S4.** Content of Mn in 4T1 cells after coincubating with  $\text{Cu}_7\text{S}_4@\text{Au}@\text{MnO}_2$  nanomaterials for 4 h and 8 h, respectively.



**Figure S5.** Images of several organs stained with H&E. Scale bar : 200  $\mu\text{m}$ .

## Reference

1. Deng, X. R.; Li, K.; Cai, X. C.; Liu, B.; Wei, Y.; Deng, K. R.; Xie, Z. X.; Wu, Z. J.; Ma, P. A.; Hou, Z. Y.; Cheng, Z. Y.; Lin, J. A Hollow-Structured Cu<sub>7</sub>S<sub>4</sub>@Cu<sub>2</sub>S@Au Nanohybrid: Synergistically Enhanced Photothermal Efficiency and Photoswitchable Targeting Effect for Cancer Theranostics, *Advanced Materials*, 2017, 29, 1701266.