# Small Lectin Ligands as a Basis for Applications in Glycoscience and Glycomedicine

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Fig. S1 Haworth chair perspectives of selected monosaccharides and oligosaccharides and corresponding symbol nomenclature for glycans (SNFG).



**Fig. S2** A. Equilibrium concentrations of glucopyranoses. B. Hyperconjugation explanation for increased stabilisation for axial anomer (endo-anomeric effect). C. Equilibrium concentrations of cyclohexanol conformers. D. Crystal structure for Man $\alpha$ 1-2Man $\alpha$ 1-OMe (Cambridge Crystallographic Database Centre identifier = FABYOW11, www.ccdc.cam.ac.uk) shows  $\phi$ = -g (-55, -60) at both glycosidic linkages consistent with influence of the exo-anomeric effect and minimisation of steric repulsive interactions. The angle for the 1-2 linkage in the X-ray structure is +17 which approaches  $\psi$ = syn.

### Supplementary notes on interglycosidic torsional angles. For a fuller account on glycan structural preferences please consult sources referred to in the main text.

<u>Φ for α-glycopyranosides</u>: Fig. S3 depicts staggered conformers that result from rotation about the C1-O1 bond of α-Man*p*; the exo-anomeric effect stabilises exo-anti-Φ and exo-syn-Φ conformers, but exo-syn-Φ is preferred as it has only one steric repulsive gauche interaction, compared to two for exo-anti, showing the combined effect of stereoelectronic and steric repulsive interaction. The glycoside torsion Φ, defined herein by atoms H1-C1-O1-R', approximates to -60° for the exo-syn-Φ. 1 These values are estimates typical for conformational analysis of hydrocarbons. Deviation is expected when there are oxygen substituents due to differences in C-O bond lengths and COC bond angles. The exo-syn conformer could be defined as within the range -60±30°.  $\phi$  is ~+60° if defined for O5-C1-O1-R' as is the definition in many publications.



Fig. S3 The exo-anomeric effect is the stabilisation of exo-syn and exo-anti over non-exo, by hyperconjugation. Steric repulsive interactions imply that exo-syn is preferred.

<u> $\Phi$  for  $\beta$ -D-glycopyranosides</u>: Then exo-syn- $\Phi$  conformer is again preferred (see Fig. S4C), with  $\Phi$ , defined by atoms H1-C1-O1-R', approximating in this case to +60°.

<u>Φ for NeuAc-Gal linkages.</u> The presence of the carboxylic acid substituent at the glycosidic carbon, reduces the energy difference between NeuAc's exo-syn-Φ and exo-anti-Φ conformers.<sup>1</sup> Thus, Φ is potentially more flexible in NeuAc( $\alpha$ 2-3)Gal and NeuAc( $\alpha$ 2-6)Gal, than Φ for β-galactopyranosides and α-mannopyranosides. The crystal structure analysis by Wormwald et al<sup>13</sup> showed exo-syn-Φ frequently found in bound conformers of NeuAc( $\alpha$ 2-3)Gal.

<u> $\Psi$  when the glycoside is derived from a secondary alcohol</u>. While there is no exo-anomeric effect for  $\Psi$ , (defined by CY-OY-CX'-HX', where Y = anomeric C/O), there is a preference for syn- $\Psi$  (Fig. 4D), observed in disaccharides such as lactose (Gal $\beta$ 1-4Glc), NeuAc( $\alpha$ 2-3)Gal, Man $\alpha$ 1-2Man, where the CY-OY and CX'-HX' bonds have a near eclipsed conformation (syn- $\Psi$ ). Organic Chemistry students learn that eclipsed conformations of ethane are disfavoured compared to staggered conformers. We understand the near eclipsing syn (non-staggered) preference for  $\Psi$  is influenced by (i) repulsive syn-1,3-diaxial (syn pentane/Hassel Ottar) interactions that occur in staggered disaccharide  $\Psi$  conformers and (ii) a lower barrier to rotation about a C-O bond linkage (~1 kcal/mol for the C-O bond in methanol, compared to ~3 kcal/mol for the ethane C-C bond).<sup>2</sup> If there is a higher barrier to rotation or if repulsive syn-1,-3-diaxial interactions are less significant then  $\Psi$  (anti) may have significant population in such disaccharides.

<u> $\psi$  when the glycoside is derived from a primary alcohol.</u> The staggered anti- $\Psi$  conformer, which places large substituents anti-periplanar, has increased preference compared to syn- $\Psi$  in NeuAc( $\alpha$ 2-6)Gal and Man $\alpha$ 1-6Man<sup>3</sup> and related linkages, as repulsive syn-1,3-diaxial type interactions are absent (Fig. 4E).

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<u>The  $\omega$  torsion</u>. Free rotation about the C5-C6 single bond ( $\omega$ -torsion) of pyranosides is influential for 1,6-linked disaccharides and NeuAc $\alpha$ -2,6Gal derivatives. The  $\omega$ -torsion adopts the gauche-gauche (*gg*), trans-gauche (*tg*) or gauche-trans (*gt*) conformers (Fig. 5). There are high preferences for *gg* and *gt* conformers, and a low population of *tg*, for glucopyranose.<sup>4,5</sup> In contrast, for galactopyranose in water, that there is increased preference (>70%) for the *gt* conformer with a low *gg* population.<sup>6</sup> Molecular dynamics (MD) calculations using explicit water accurately predicts the  $\omega$  torsion population, showing that water disrupts intramolecular hydrogen bonding allowing the *gg*, *gt* and *tg* populations, to be determined, not by intramolecular H-bonding, but by internal electronic and steric repulsions.<sup>7</sup> For Man and GlcNAc, the conformer preferences are like those of glucopyranoses.<sup>8</sup>



**Fig. S4** A. Hassel Ottar (syn 1,3-diaxial) interactions,<sup>9</sup> destabilise  ${}^{1}C_{4}$  relative to  ${}^{4}C_{1}$  B. X-ray crystal structure of lactose has exo-syn- $\Phi$ ; syn- $\Psi$ . C. Exo-anomeric effect and steric repulsive interactions favour exo-syn  $\Phi$  for  $\beta$ -Galp. D. Low energy conformers from O1-C4' bond rotation in disaccharides like lactose, favours syn- $\Psi$  in 1-4 linkage; 1,3-syn-diaxial interactions disfavours staggered anti- $\Psi$ . E. 1-6 linkage favours anti- $\Psi$ .



Fig. S5 Newman projections showing gg, tg and gt conformers viewing along C-5 to C-6 bonds of: A. Glc/Man/GlcNAc (gg and gt preferred); B. Gal/GalNAc (tg and gt preferred).

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Ligand	Galectin	Φ	Ψ	PDB code	
Lactose	1	61	10	1W6O	
Lactose	3	51	12	1KJL	
Lactose	4C	53	10	4YM3	
3'-Sulfo-lactose	4C	61	10	4YM2	
3'-Sulfo-lactose	4N	55	19	5DUW	
Lactose	7	61	4	4GAL	
Lactose	8C	61	-7	3VKM	
Lactose	8N	50	18	3AP4	
3'-Sialyl Lactose	8N	54	14	3AP7	
3'-Sulfo-lactose	8N	53	11	3AP6	
3'-Sialyl Lactose	9C	54	19	3NV4	
Lactose	9N	60	20	<b>3EAK</b>	
Lactose	10	54	-20	6A1T	

## Table S1: $\Phi$ and $\Psi$ (°) for interglycosidic torsions between galactopyranose and glucopyranose residues in galectin ligands from co-crystal structures

Data is taken from co-crystal structures with coordinates downloaded from the protein databank (<u>www.rcsb.org</u>).  $\Phi$  is defined by H1'-C1'-O'-C4 and  $\Psi$  by C'1-O1'-C4-H4. While there are no H atoms in the crystal structures, the H atoms were added to the coordinates downloaded from <u>www.rcsb.org</u> using Maestro (freely available for academic use from schrodinger.com) and this was used to measure the torsion angles.



Fig. S6. A selection of potential lectin-ligand binding interactions depicted with Chemdraw. Shown are A. Fuc-LecB; B. Gal-LecA; C. β-Galactoside (sulfatide)-galectin-8N; D. Neu5Ac-mouse siglec-1); E. Neu5Ac-hemagglutinin; F. Man-FimHLD. The ligand-lectin interactions have affinities from mM to mM range. PDB identifier codes are provided.



Fig. S7. 3'SL binding mode with galectin-8N. Charged H-bonding interactions with arginine from the carboxylate give higher affinity interactions with galectin-8. The wider H-bonding network arises and CH-p interactions common for galectin recognition of Gal. For sulfatide, the CH-Pi interactions involves the  $\alpha$ -Gal face defined by H atoms ~3A distance from indole atoms are shown. The ligand bound structure has glycosidic torsion angles that correspond to a low energy ligand conformation.



Table S2: Ligand Binding Affinities for Galectins	( <i>K</i> d µM) References	specific to data cite	ed are
given below Table.			

Compound	Gal-1	Gal-2	Gal-3	Gal-4C	Gal-4N	Gal-7	Gal-8C	Gal-8N	Gal-9C	Gal-9N
Methyl β-D-Gal	10000 <sup>1</sup>	13000 <sup>10</sup>	4100 <sup>1</sup>	10000 <sup>1</sup>	6600 <sup>1</sup>	4800 <sup>9</sup>	>300001	6300 <sup>1</sup> 4370 <sup>4</sup>	8600 <sup>1</sup>	3300 <sup>1</sup>
Methyl α-D-Gal	>10000 <sup>2</sup>	>20000 <sup>2</sup>	2700 <sup>2</sup>	>20000 <sup>2</sup>	>20000 <sup>2</sup>	11000 <sup>2</sup>	>20000 <sup>2</sup>	6300 <sup>2</sup>	6200 ±220 <sup>2</sup>	2800 <sup>2</sup>
Methyl β-D-Glc	10000 <sup>1</sup>	10000 <sup>1</sup>	NB	11000 <sup>1</sup>	ND	ND	NB	NB	NB	NB
Man	>10000 <sup>14</sup>	ND	>10000 <sup>14</sup>	ND	ND	>10000 <sup>14</sup>	ND	>10000 <sup>14</sup>	ND	>10000 14
Lactose (Galβ1-4Glc)	104 <sup>3</sup>	ND	231 <sup>3</sup>	1900 <sup>7</sup>	130011	ND	331.1 <sup>13</sup>	91 <sup>3</sup> 47.4 ±1.1 <sup>13</sup>	ND	ND
Galβ1-4GlcNAc	ND	ND	1.815	21800 <sup>7</sup>	ND	ND	ND	420 <sup>15</sup> (SPR) 9.7 <sup>15</sup> (FA)	ND	ND
Galβ1-4Glcβ1-OMe	187 <sup>8</sup> 190 <sup>12</sup>	ND	160 <sup>8</sup> 220 <sup>12</sup>	1200 <sup>12</sup>	540 <sup>12</sup>	110 <sup>8</sup> 91 <sup>12</sup>	ND	109 <sup>4</sup> 62 <sup>8</sup> 52 <sup>12</sup>	ND	23 <sup>8</sup>
Galβ1-4GlcNAcβ1- OMe	65 <sup>8</sup>	ND	59 <sup>8</sup>	ND	ND	550 <sup>8</sup>	ND	1000 <sup>8</sup>	ND	490 <sup>8</sup>
TDG (Thiodigalactoside)	24±11 <sup>5,8</sup>	340±19 <sup>6</sup>	49±11 <sup>5,8</sup>	1450±260 <sup>5</sup>	410±21 <sup>5</sup>	160±18 <sup>6,8</sup>	ND	61±17 <sup>5</sup>	42±1.1 <sup>6</sup>	38±8 <sup>6,8</sup>

ND = not determined or not reported

#### **References for Table S2:**

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#### Energy surface plots for various disaccharides: Figures S6-S9:

**Fig. S9**: This figure was reproduced from S. Kumar, M. Frank, R. Schwartz-Albiez, PLoS ONE, 2013, 8(3):e59761 (doi.org/10.1371/journal.pone.0059761). The conformational space of glycosidic linkages in blood group antigens was analyzed over 10 ns of MD simulations in the gas phase. Panel A shows the space for blood group antigen A (BGA), and Panel B for blood group antigen B (BGB). The  $\phi$  and  $\psi$  values were determined using NMR definitions where  $\phi$  is defined by H1-C1-O1-C<sub>x</sub> and  $\psi$  as C1-O1-C<sub>x</sub>-H<sub>x</sub>.



**Fig. S10**: This figure was reproduced from V. Palivec, C. Johannessen, J. Kaminsky, H. Martinez-Seara, PLoS Computational Biol, 2022, 18(1):e1009678 (doi.org/10.1371/journal.pcbi.1009678). Left: Calculated free energy surface of Manα(1-3)ManαOMe. Middle: Calculated FES, together with 250 sampled structures from unbiased 500 ns MD simulations. Right: Calculated FES, together with 250 sampled structures for various biased 200 ns MD simulations. White regions represent area with the free energy >40 kJ/mol.  $\phi_1$  is defined by atoms H1-C1-O1-C3';  $\phi_2$  is defined by C1-O1-C3'-H3'. 1Rad = 180/ $\pi$  = 57.296°.



**Fig. S11:** This figure was reprinted from P. Vidal, J. Jiménez-Barbero, J. F. Espinosa, Carbohydrate Research 2016, 433, 36-40 (doi.org/10.1016/j.carres.2016.06.009) with permission from Elsevier. The left side shows an adiabatic map and the right side displays the population distribution of Gal $\beta$ (1-3)Glc $\beta$ OMe, calculated using the MM3\* force field ( $\epsilon$  = 80). Isoenergy contours are marked at 0.5 kcal/mol intervals. Probability contours indicate populations at 10%, 1%, and 0.1%. The three possible conformational families are A (syn), B (anti- $\psi$ ), and C (anti- $\phi$ ). The  $\phi$  and  $\psi$  torsions correspond to H1 -C1 -O1 -C3' and C1 -O1 -C3'-H3' dihedral angles respectively.



**Fig. S12:** This figure was reprinted from S. Sabesan, K. Bock, J. C. Paulson, Carbohydrate Research., 1991, 218, 27-54 (doi: 10.1016/0008-6215(91)84084-r) with permission from Elsevier. It shows the energy-contour map for sialoside linkage of D-NeuAc(2-3) $\beta$ -D-Gal-OMe. Contour lines indicate energy differences of 1 kcal/mol, with the outermost line around each energy well representing a level 10 kcal/mol above the minimum-energy conformer.  $\phi$  is defined by C1-C2-O3'-C3';  $\psi$  is defined by C2-O3'-C3'-H3'.