Electronic Supplementary Information for

Ni-catalyzed undirected and regioselective acceptorless dehydrogenative silylation of primary benzylic C(sp³)–H bonds

Qing Yu,^a Takafumi Yatabe,^{*ab} Takehiro Matsuyama,^a Tomohiro Yabe^a and Kazuya Yamaguchi^{*a}

^aDepartment of Applied Chemistry, School of Engineering, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8656, Japan.

^bPrecursory Research for Embryonic Science and Technology (PRESTO), Japan Science and Technology Agency (JST), 4-1-8 Honcho, Kawaguchi, Saitama 332-0012, Japan.

*e-mail: kyama@appchem.t.u-tokyo.ac.jp, yatabe@appchem.t.u-tokyo.ac.jp

Table of Contents

Supplementary Experimental Methods	S2–S5
Spectral Data of Products	S6–S13
Supplementary Figures	S14–S38
Supplementary Tables	S39–S50
Supplementary References	S51
NMR Spectra	S52–S67

Supplementary Experimental Methods

Instrumental and Reagents: GC analyses were performed on Shimadzu GC-2014 equipped with a flame ionization detector (FID) and an InertCap-5 using Shimadzu C-R8A Chromatopac Data Processor for area calculations. GC-MS spectra were recorded on Shimadzu GCMS-QP2020 equipped with an InertCap-5 capillary column at an ionization voltage of 70 eV. Liquid-state NMR spectra were recorded on JEOL JNM-ECA-500. ¹H, ²H, and ¹³C NMR spectra were measured at 500.16, 76.78, and 125.76 MHz, respectively. ¹H NMR chemical shifts were referenced to tetramethylsilane (TMS) signal (0 ppm). ¹³C NMR chemical shifts were referenced to residual solvent peaks. Column chromatography on silica gel was performed by using Biotage Isolera. ICP-AES analyses were performed on Shimadzu ICPS-8100. STEM observations were performed on a JEOL JEM-ARM200F. XRD patterns were recorded on a Rigaku SmartLab diffractometer (Cu Ka, $\lambda = 1.5405$ Å, 45 kV, 20 mA). The XPS spectra were measured on an Ulvac-Phi PHI5000 VersaProbe at the Advanced Characterization Nanotechnology Platform of the University of Tokyo. The samples were embedded in In foil and brought in the introduction chamber without exposing them to air by using the transfer vessel. The binding energies were calibrated by using the C 1s signal at 284.8 eV. Ni K-edge X-ray absorption spectroscopy (XAS) was carried out at the BL14B2 beamline of SPring-8 (Proposal No. 2022A1803, 2021A1620, and 2019B1820). XAFS measurements were conducted in fluorescence mode using a Si (111) crystal monochromator. Each sample was formed into a pellet, doubly wrapped in an Al laminate pack, and heat-sealed in an Ar-filled glovebox. XANES and EXAFS data were analyzed using Athena and Artemis software (Demeter, ver. 0.9.025; Bruce Ravel). The k^3 -weighed EXAFS spectra were Fourier-transformed into R-space at 3-12 Å⁻¹. Ni foil and Ni(OH)₂ were used as standard samples. IFFEFIT (Athena and Artemis) programs were used for data analysis, treatment, and modeling. FEFF6 was used to generate Feff.inp files for analysis by Artemis using the atomic coordinates of the relevant theoretical references calculated using X-ray crystallographic methods available in the literature: Ni and Ni(OH)₂. IR-ATR spectra were measured on Jasco FT/IR-4100. DRIFTS was performed using Jasco FT/IR-6700. BET surface areas were measured on micromeritics ASAP 2010 and calculated from the N_2 adsorption isotherm with the BET equation. CeO₂ (BET: 50 m² g^{-1} after calcination at 550 °C for 3 h, Cat. No. 544841-25G, Aldrich), Al₂O₃ (BET: 160 m² g⁻¹ after calcination at 550 °C for 3 h, Cat. No. KHS-24, Sumitomo Chemical), TiO₂ (BET: 316 m² g⁻ ¹, Cat. No. ST-01, Ishihara Sangyo Kaisya), ZrO₂ (Cat. No. 37022-35, Nacalai Tesque), Co₃O₄ (Cat. No. 039-08792, FUJIFILM Wako Pure Chemical), and hydroxyapatite (HAP, BET surface area: 11 m² g⁻¹, Cat. No. 011-14882, FUJIFILM Wako Pure Chemical) were acquired from commercial sources. Solvents and substrates were obtained from Tokyo Chemical Industry, Aldrich, Kanto Chemical, or FUJIFILM Wako Pure Chemical (reagent grade), and purified prior to being used, if necessary.

Preparation of Ni(OH)_x/CeO₂: The aqueous solution of NiCl₂·6H₂O (118.8 mg, 8.3 mM) was mixed with CeO₂ (2.0 g). The mixture was vigorously stirred at room temperature for 15 min, then the pH of the mixture was adjusted to 10.0 by dropping the aqueous solution of NaOH (1.0 M). The resulting slurry was continuously stirred for 24 h. The precipitation was filtered off and washed by water (3 L) then dried in vacuo to obtain Ni(OH)_x/CeO₂ as pale yellow powder. The Ni(OH)_x supported on other supports such as Ni(OH)_x/TiO₂, Ni(OH)_x/ZrO₂, Ni(OH)_x/Al₂O₃, Ni(OH)_x/Co₃O₄ and Ni(OH)_x/HAP was prepared in the same method using TiO₂, ZrO₂, Al₂O₃,

Co₃O₄, and HAP instead of CeO₂. In the same way, various metal hydroxide supported on CeO₂ such as $Mn(OH)_x/CeO_2$, $Fe(OH)_x/CeO_2$, $Co(OH)_x/CeO_2$, $Cu(OH)_x/CeO_2$ and $Zn(OH)_x/CeO_2$ were prepared using $MnCl_2$. $4H_2O$, $FeCl_3$. $6H_2O$, $CoCl_2$. $6H_2O$, $CuCl_2$. $2H_2O$ and $ZnCl_2$ as the metal source respectively.

Preparation of Ni/CeO₂-HBpin: By modifying the method reported in our previous report,^{S1} Ni/CeO₂-HBpin was prepared as follows. Ni(OH)_x/CeO₂ was dried in vacuo for 15 min at 150 °C and moved into an Ar-filled glovebox. The Ni(OH)_x/CeO₂ (1.0 g) was mixed with pinacolborane (HBpin) (8.0 mmol) and cyclopentyl methyl ether (CPME) (10 mL) in a Pyrex glass reactor (volume: ~100 mL), and a Teflon-coated magnetic stir bar was placed in the reactor. The mixture was vigorously stirred at 100 °C for 30 min under an Ar atmosphere, then moved into the glovebox after cooled down to room temperature. The solid was filtered off and washed by diethyl ether (30 mL), then dried in vacuo for 30 min. Ni/CeO₂-HBpin (Ni content: 1.4 wt%) was obtained as dark gray or black powder. The Ni(OH)_x supported on other supports and other metal hydroxide supported on CeO₂ reduced by HBpin was treated in the same method.

Preparation of Ni/CeO₂-NaNaph: According to our previous report, ^{S2} Ni/CeO₂-NaNaph was prepared as follows. In an Ar-filled glovebox, Ni(OH)_x/CeO₂ (500 mg) was mixed with the THF solution (3.0 mL, 0.17 M) of sodium naphthalenide and vigorously stirred at room temperature for 12 h. Methanol (2 mL) was dropped into the reaction system to quench the reduction, and then the solid was filtered off and washed by methanol (30 mL) and diethyl ether (30 mL), then dried in vacuo for 30 min. Ni/CeO₂-NaNaph was obtained as black powder.

Preparation of Ni/CeO₂-NaBH₄: In an Ar-filled glovebox, Ni(OH)_x/CeO₂ (500 mg) was mixed with the isopropanol solution (10.0 mL, 0.15 M) of sodium borohydride and vigorously stirred at 80 °C for 90 min. The solid was filtered off and washed by ethanol (30 mL), then dried in vacuo for 30 min. Ni/CeO₂-NaNaph was obtained as black powder.

Preparation of Ni/CeO₂-HSi(OEt)₃: In an Ar-filled glovebox, Ni(OH)_x/CeO₂ (1.0 g) was mixed with cyclopentyl methyl ether (CPME) solution of triethoxysilane (HSi(OEt)₃) (5.0 mL, 0.4 M) and vigorously stirred at 100 °C for 30 min. The solid was filtered off and washed by diethyl ether (30 mL), then dried in vacuo for 30 min. Ni/CeO₂-HSi(OEt)₃ was obtained as gray powder.

Preparation of Ni/CeO₂-H₂: According to our previous report,^{S1} Ni/CeO₂-H₂ was prepared as follows. Ni(OH)_x/CeO₂ (575 mg) was dried in vacuo at 400 °C for 30 min in a Schlenk flask (volume: ~30 mL). The dried Ni(OH)_x/CeO₂ was heated at 400 °C for 1 h with an H₂ balloon, and it was dried in vacuo for 30 min while the reactor was cooled down to 150 °C. Ni/CeO₂-H₂ was obtained as black powder and stored in an Ar-filled glovebox.

A Typical Procedure for the Dehydrogenative Benzylic Silylation of Alkylarenes: In an Arfilled glovebox, Ni/CeO₂-HBpin (Ni: 1.8 mol%, 32 mg), alkylarene (2 mmol), *n*-hexadecane (0.1 mmol, internal standard), triethoxysilane (0.4 mmol), methylcyclohexane (1 mL) were mixed in a in a Pyrex glass reactor (volume: ~20 mL), and a Teflon-coated magnetic stir bar was placed in the reactor. The mixture was vigorously stirred at 120 °C for 24 h. Product yields were determined by GC analysis with *n*-hexadecane as the internal standard based on the initial amount of triethoxysilane. *n*-Hexadecane was not added for the product isolation. The catalyst was removed by filtration after the reaction. The filtrate was moved into an eggplant flask (volume: \sim 10 mL) and washed by dichloromethane. The filtrate was concentrated by rotary evaporation, then purified by silica-gel column chromatography. The solvent of the pure products was finally removed by rotary evaporation at 50 °C, 6 mbar for 30 min. The isolated products were identified by GC-MS and NMR (¹H and ¹³C).

Synthesis of Heptylbenzene-1,1- d_2 : By referring to previous reports,^{S1,S3} heptylbenzene-1,1- d_2 was synthesized as follows. Pd/C (N.E. CHEMCAT K-type, 5 wt%, 350 mg) was placed in a Schlenk flask (volume: ~ 50 mL) with a Teflon-coated magnetic stir bar and dried in vacuo. Then, the atmosphere was filled in an H₂ gas with a balloon. Heptylbenzene (10 mmol) and D₂O (10 mL) were then added by syringe. The mixture was stirred at room temperature for 4.5 days. After the reaction, the catalyst was removed by filtration. The filtrate was extracted with diethyl ether, washed with brine, dried over MgSO₄, then concentrated with the rotary evaporator at 45 °C, 6 mbar for 30 min to give the product (deuteration ratio: 92%, determined by ¹H NMR).



heptylbenzene-1,1-*d*₂ (CAS number: 2596115-67-0): GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 8.3 min. Rotary evaporator (45 °C, 6 mbar, 30 min) yielded 1.19 g (67% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 179 (5), 178 (36) [*M*⁺], 177 (6), 135 (8), 107 (8), 106 (5), 95 (8), 94 (100), 93 (99), 92 (29), 91 (9), 78 (8), 67 (5), 66 (7). ¹H NMR (500 MHz, CD₃OD): δ 7.19–7.22 (m, 2H), 7.09–7.13 (m, 3H), 2.52–2.56 (m, 0.16H), 1.55–1.58 (m, 2H), 1.27–1.31 (m, 8H), 0.86–0.89 (m, 3H). The NMR spectral data accord with the previous report.^{S1}

A Procedure for the Dehydrogenative Benzylic Silylation of 1a in a Gram-Scale: In an Arfilled glovebox, Ni/CeO₂-HBpin (Ni: 1.8 mol%, 463.8 mg), toluene (1a) (29 mmol), *n*-hexadecane (1.45 mmol, internal standard), triethoxysilane (5.8 mmol), methylcyclohexane (7 mL) were added into a 40 mL pressure tube possessing a valve, and a Teflon-coated magnetic stir bar was placed in the reactor. The mixture was vigorously stirred at 120 °C for 24 h with the cooling to room temperature and degassing by immediate open/close a valve per 1 h till 3 h after the reaction started. The product yields were determined by GC analysis with *n*-hexadecane as the internal standard based on the initial amount of triethoxysilane.

Leaching Test: A magnetic stir bar, *n*-hexadecane (0.1 mmol), $HSi(OEt)_3$ (0.4 mmol), toluene (2 mmol), methylcyclohexane (1 mL) and Ni/CeO₂-HBpin (32 mg, 1.8 mol%) were added to a Pyrex glass reactor (volume: ~ 20 mL), and the reaction was carried out at 120 °C in an Ar-filled glovebox. The catalyst was removed by hot filtration after the reaction started for 7.5 min, when

the yield of **2a** reached 55%. The solution was moved to another reactor which had been heated to 120 °C with a magnetic stir bar, and then the reaction solution was stirred at 120 °C continuously. As a result, the reaction immediately stopped after the hot filtration (Fig. S11). To determine the leaching amount of Ni species into the reaction solution, the solution after 1 h reaction was filtered by a disposable syringe and concentrated by a rotary evaporator. Next, aqua regia (1 mL) was added into the concentrated mixture, and the amount of Ni species leached into the reaction solution was measured by ICP-AES. Ni species in the solution after the reaction were hardly observed by ICP-AES (0.7% of the Ni species used for the reaction). Therefore, this Ni/CeO₂-HBpin-catalyzed reaction was confirmed to be truly heterogeneous.

Reuse Test: After the dehydrogenative benzylic silvlation of **1a** for 1 h under the optimized conditions, the Ni/CeO₂-HBpin catalyst was filtered off in an Ar-filled glovebox, washed by methanol (30 mL), and dried in vacuo for 30 min. Then, the retrieved catalyst was applied to the dehydrogenative benzylic silvlation of **2a** again for 1 h.

Spectral Data of Products



Benzyltriethoxysilane (2a) (CAS number: 2549-99-7): 93% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 9.2 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.747 (calculated by calibration curve using an isolated substance). Rotary evaporator (60 °C, 6 mBar, 30 min) yielded 79.5 mg (79% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 254 (16) [*M*⁺], 165 (9), 164 (12), 163 (100), 135 (14), 121 (6), 120 (6), 119 (55), 107 (15), 91 (33), 79 (20), 65 (5), 63 (7), 45 (7). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.16–7.23 (m, 4H), 7.07–7.10 (m, 1H), 3.75 (q, *J* = 7.0 Hz, 6H), 2.20 (s, 2H), 1.16 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 137.68, 128.99, 128.31, 124.69, 58.81, 20.50, 18.30. The NMR spectral data accord with the previous report.^{S4}



Triethoxy(2-methylbenzyl)silane (**2b**) (CAS number: 1428345-75-8): 74% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 10.2 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.810 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.33) yielded 49.6 mg (45% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 163 (100), 268 (24) [*M*⁺], 164 (12), 135 (13), 119 (47), 107 (11), 105 (11), 91 (7), 79 (16). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.00–7.15 (m, 4H), 3.73 (q, *J* = 7.5 Hz, 6H), 2.32 (s, 3H), 2.18 (s, 2H), 1.16 (t, *J* = 7.5 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 136.28, 135.81, 130.05, 129.42, 125.77, 124,85, 58.75, 20.32, 18.28, 18.05. The NMR spectral data accord with the previous report.⁸⁴



Triethoxy(3-methylbenzyl)silane (**2c**) (CAS number: 1443303-79-4): 85% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL·min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C·min⁻¹ (1 min), 10 °C·min⁻¹ (20 min), 0 °C·min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 10.0 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.810 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 9/1, Rf = 0.44) yielded 54.9 mg (50% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 268 (24) [*M*⁺], 164 (12), 163 (100), 135 (15), 120 (5), 119 (50), 107 (12), 105 (14), 91 (9), 79 (17). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.10 (t, *J* = 7.5 Hz, 1H), 6.96–7.00 (m, 2H), 6.90 (d, *J* = 7.5 Hz, 1H), 3.75 (q, *J* = 7.0 Hz, 6H), 2.29 (s, 3H), 2.17 (s, 2H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 137.70, 137.46, 129.84, 128.17, 125.99, 125.41, 58.77, 21.46, 20.29, 18.28.



Triethoxy(4-methylbenzyl)silane (2d) (CAS number: 1443341-91-0): 77% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL·min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C·min⁻¹ (1 min), 10 °C·min⁻¹ (20 min), 0 °C·min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 10.2 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.810 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.25) yielded 65.5 mg (61% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 268 (21) [*M*⁺], 164 (12), 163 (100), 135 (12), 119 (47), 107 (11), 105 (12), 91 (8), 79 (20), 77 (5), 63 (6). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.00–7.08 (m, 4H), 3.75 (q, *J* = 7.0 Hz, 6H), 2.28 (s, 3H), 2.16 (s, 2H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 134.29, 133.94, 128.99, 128.83, 58.74, 21.02, 19.82, 18.29.



Triethoxy(2-methoxybenzyl)silane (2e) (CAS number: 1443323-43-0): 63% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 11.3 min; relative sensitivity for quantification (vs n-hexadecane, internal standard), 0.747 (calculated by the relative sensitivity of 2a and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.20) yielded 22.9 mg (56% yield) of a colorless oil. MS (EI, 70 eV): m/z (%): 286 (6), 285 (21), 284 (100) [*M*⁺], 270 (9), 269 (48), 241 (5), 240 (6), 239 (22), 225 (21), 223 (15), 197 (8), 196 (6), 195 (28), 183 (13), 182 (6), 181 (22), 179 (11), 169 (6), 168 (8), 167 (22), 165 (7), 163 (10), 153 (7), 151 (16), 150 (28), 149 (75), 148 (21), 147 (95), 137 (6), 136 (7), 135 (50), 134 (7), 133 (18), 131 (8), 123 (5), 122 (27), 121 (15), 120 (8), 119 (58), 118 (64), 117 (17), 107 (25), 106 (7), 105 (50), 104 (6), 103 (27), 99 (11), 93 (14), 92 (13), 91 (53), 90 (9), 89 (9), 84 (12), 79 (29), 78 (13), 77 (14), 73 (6), 63 (13), 62 (6). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.16–7.18 (m, 1H), 7.07–7.10 (m, 1H), 6.79–6.85 (m, 2H), 3.81 (s, 3H), 3.75 (q, J = 7.0 Hz, 6H), 2.23 (s, 2H), 1.15 (t, J = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 156.90, 130.36, 126.36, 125.87, 120.44, 110.15, 58.61, 55.32, 18.28, 13.26.



Triethoxy(3-methoxybenzyl)silane (**2f**) (CAS number: 1443302-78-0): 62% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 11.7 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.747 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.30) yielded 11.2 mg (37% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 285 (9), 284 (43) [*M*⁺], 165 (5), 164 (12), 163 (100), 135 (14), 121 (11), 120 (6), 119 (57), 107 (14), 91 (19), 79 (18), 63 (6). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.13 (t, *J* = 7.8 Hz, 1H), 6.75–6.77 (m, 2H), 6.64–6.67 (m, 1H), 3.78 (s, 3H), 3.76 (q, *J* = 7.0 Hz, 6H), 2.19 (s, 2H), 1.18 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 159.65, 139.28, 129.20, 121.56, 114.56, 110.39, 58.83, 55.21, 20.66, 18.34.



Triethoxy(4-methoxybenzyl)silane (2g) (CAS number: 1443324-33-1): 77% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL·min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C·min⁻¹ (1 min), 10 °C·min⁻¹ (20 min), 0 °C·min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 12.0 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.747 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.25) yielded 58.4 mg (52% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 285 (6), 284 (30) [*M*⁺], 271 (6), 270 (19), 269 (100), 195 (6), 181 (5), 164 (8), 163 (66), 135 (15), 121 (45), 120 (6), 119 (64), 107 (18), 91 (22), 79 (26), 77 (9), 63 (8), 45 (6), 78 (6). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.08–7.11 (m, 2H), 6.76–6.79 (m, 2H), 3.75 (q, *J* = 7.0 Hz, 6H), 3.76 (s, 3H), 2.14 (s, 2H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 157.08, 129.79, 129.39, 113.80, 58.74, 55.31, 19.10, 18.30. The NMR spectral data accord with the previous report.⁸⁵



N,*N*-dimethyl-4-((triethoxysilyl)methyl)aniline (2h): 49% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 13.6 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.856 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 13/1, Rf = 0.15) yielded 41.9 mg (35% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 298 (14), 297 (66) [*M*⁺], 282 (22), 251 (6), 163 (22), 135 (15), 134 (100), 133 (9), 119 (26), 118 (13), 107 (6), 98 (7), 91 (11), 79 (12). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.04–7.06 (m, 2H), 6.66–6.68 (m, 2H), 3.76 (q, *J* = 7.0 Hz, 6H), 2.88 (s, 6H), 2.11 (s, 2H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 148.45, 129.55, 125.54, 113.46, 58.71, 41.19, 18.68, 18.34.



Triethoxy(3-(trifluoromethyl)benzyl)silane (2i): 46% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 8.9 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.747 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.30) yielded 41.9 mg (35% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 322 (1) [*M*⁺], 164 (6), 163 (48), 141 (12), 140 (100), 135 (9), 120 (5), 119 (42), 107 (11), 91 (9), 79 (16), 63 (6). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.45 (s, 1H), 7.31–7.37 (m, 3H), 3.76 (q, *J* = 7.0 Hz, 6H), 2.25 (s, 2H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 139.02, 132.29, 130.56 (q, *J* = 32.4 Hz), 128.61, 125.69 (q, *J* = 3.6 Hz), 124.48 (q, *J* = 272.3 Hz), 121.53 (q, *J* = 3.6 Hz), 58.93, 20.76, 18.26.



Triethoxy(4-(trifluoromethyl)benzyl)silane (2j): 41% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 9.1 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.747 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.3) yielded 7.4 mg (11% yield) of a colorless oil. MS (EI, 70 eV): m/z (%): 322 (1), 277 (5), 233 (6), 164 (7), 163 (56), 159 (9), 141 (12), 140 (100), 135 (9), 120 (5), 119 (45), 109 (6), 107 (12), 91 (9), 79 (18), 63 (6). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.46–7.48 (m, 2H), 7.26–7.28 (m, 2H), 3.76 (q, *J* = 7.0 Hz, 6H), 2.25 (s, 2H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 142.49, 129.13, 127.06 (q, *J* = 32.4 Hz), 125.19 (q, *J* = 3.6 Hz), 124.68 (q, *J* = 272.3 Hz), 58.94, 21.05, 18.31.



methyl 4-((triethoxysilyl)methyl)benzoate (**2k**): 60% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 14.1 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.747 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.14) yielded 53.1 mg (43% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 312 (13) [*M*⁺], 163 (28), 135 (8), 119 (42), 118 (100), 107 (9), 91 (10), 90 (19), 79 (15), 63 (5). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.89–7.92 (m, 2H), 7.22–7.25 (m, 2H), 3.89 (s, 3H), 3.75 (q, *J* = 7.0 Hz, 6H), 2.26 (s, 2H), 1.16 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 167.42, 144.05, 129.66, 128.89, 126.66, 58.89, 51.99, 21.39, 18.27.



(3,5-dimethylbenzyl)triethoxysilane (2I) (CAS number: 1443323-10-1): 80% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL·min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C·min⁻¹ (1 min), 10 °C·min⁻¹ (20 min), 0 °C·min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 10.8 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.872 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.28) yielded 17.0 mg (45% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 282 (19) [*M*⁺], 165 (5), 164 (12), 163 (100), 135 (11), 120 (7), 119 (53), 107 (10), 91 (13), 79 (17), 63 (6), 45 (5). ¹H NMR (500 MHz, CDCl₃, TMS): δ 6.79 (s, 2H), 6.73 (s, 1H), 3.76 (q, *J* = 7.0 Hz, 6H), 2.25 (s, 6H), 2.13 (s, 2H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 137.61, 137.29, 126.89, 126.35, 58.78, 21.38, 20.09, 18.28.



Triethoxy(4-ethylbenzyl)silane (**2m**) (CAS number: 1443323-41-8): 55% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 11.2 min; relative sensitivity for quantification (vs biphenyl, internal standard), 0.872 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.36) yielded 21.3 mg (19% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 283 (10), 282 (43) [*M*⁺], 253 (9), 238 (17), 237 (10), 236 (25), 223 (6), 210 (10), 209 (53), 207 (9), 192 (7), 191 (21), 190 (11), 165 (7), 164 (14), 163 (100), 147 (9), 146 (5), 136 (6), 137 (6), 135 (58), 120 (6), 119 (61), 118 (32), 117 (18), 115 (6), 107 (21), 105 (14), 103 (7), 91 (31), 90 (5), 79 (31), 77 (8), 63 (16), 45 (9). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.08–7.10 (m, 2H), 7.04–7.06 (m, 2H), 3.76 (q, *J* = 6.8 Hz, 6H), 2.59 (q, *J* = 7.5 Hz, 2H), 2.17 (s, 2H), 1.20 (t, *J* = 7.8 Hz, 3H), 1.17 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 140.52, 134.55, 128.92, 127.82, 58.78, 28.51, 19.83, 18.32, 15.85.



Triethoxy(4-propylbenzyl)silane (**2n**) : 70% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 12.2 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.935 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 19/1, Rf = 0.35) yielded 66.3 mg (sum of 4 runs, 56% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 296 (15) [*M*⁺], 164 (12), 163 (100), 135 (9), 133 (6), 119 (39), 107 (9), 104 (6), 91 (13), 79 (15). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.07–7.09 (m, 2H), 7.02–7.03 (m, 2H), 3.75 (q, *J* = 7.0 Hz, 6H), 2.52 (t, *J* = 7.8 Hz, 2H), 2.17 (s, 2H), 1.60 (sext, *J* = 7.4 Hz, 2H), 1.16 (t, *J* = 7.3 Hz, 9H), 0.91 (t, *J* = 7.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 138.89, 134.56, 128.81, 128.43, 58.75, 37.70, 24.75, 19.84, 18.29, 13.88.



Ethyl 2-(4-((triethoxysilyl)methyl)phenyl)acetate (**2o**): 30% GC yield. GC conditions and analysis: InertCap-5 capillary column, 0.25 mm × 30 m, GL Science Inc.; carrier gas (N₂) flow rate, 1.6 mL· min⁻¹; initial column temp., 80 °C; final column temp., 280 °C; progress rate, 0 °C· min⁻¹ (1 min), 10 °C· min⁻¹ (20 min), 0 °C· min⁻¹ (9 min); injection temp., 250 °C; detection temp., 280 °C; retention time, 15.0 min; relative sensitivity for quantification (vs *n*-hexadecane, internal standard), 0.872 (calculated by the relative sensitivity of **2a** and the effective carbon number concept). Silica gel column chromatography (eluent, hexane/ethyl acetate = 24/1, Rf = 0.08) yielded 30.8 mg (sum of 2 runs, 23% yield) of a colorless oil. MS (EI, 70 eV): *m/z* (%): 341 (7), 340 (31) [*M*⁺], 294 (6), 268 (7), 267 (27), 266 (67), 265 (25),209 (12), 165 (6), 164 (12), 163 (100), 135 (21), 133 (6), 132 (25), 120 (6), 119 (66), 107 (20), 104 (72), 103 (9), 105 (15), 91 (19), 79 (35), 78 (7), 77 (6), 63 (10), 45 (6). ¹H NMR (500 MHz, CDCl₃, TMS): δ 7.11–7.15 (m, 4H), 4.13 (q, *J* = 7.3 Hz, 2H), 3.75 (q, *J* = 6.8 Hz, 6H), 3.55 (s, 2H), 2.18 (s, 2H), 1.23 (t, *J* = 7.0 Hz, 3H), 1.16 (t, *J* = 7.0 Hz, 9H). ¹³C NMR (126 MHz, CDCl₃, TMS): δ 171.99, 136.41, 130.43, 129.16, 129.13, 60.82, 58.79, 41.16, 20.08, 18.29, 14.27.

Supplementary Figures

Chauvier et al. (2023): homogeneous catalysis



Fig. S1 Comparison of the present Ni-catalyzed undirected dehydrogenative benzylic silylation system in terms of green chemistry indicators such as atom economy (AE), *E*-factor, and turnover frequency (TOF) with the previously reported efficient systems showing comparatively broad substrate scopes^{S5,S6} by utilizing the cases of dehydrogenative benzylic silylation of toluene. The catalytic system using a *tert*-butyl-substituted silyldiazene^{S5} shows worse AE and *E*-factor than those of the present catalytic system due to the silylating reagent. The UiO-Co-catalyzed system using hydrosilane as the same silylating reagent^{S6} showed much lower TOF than that of the present catalytic system even when comparing them at almost the same temperature. Also, much longer reaction time and solvent amount of alkylarenes were required in the UiO-Co-catalyzed system. Overall, all of the indicators were tremendously improved in this study.

AE: molecular weight (MW) of desired product divided by total MW of reactants. *E*-factor: weight of waste divided by weight of desired product (in this case, MW of byproducts were divided by MW of desired product). TOF was calculated using demonstrated yields, amounts of metal catalysts used for the reactions, and reaction times. THF = tetrahydrofuran. MCH = methylcyclohexane.



Fig. S2 Linear combination fittings of Ni K-edge XANES spectra of (a) Ni/CeO₂-HBpin, (b) Ni/CeO₂-HBpin-used, (c) Ni/CeO₂-4HBpin.



Fig. S2 (continued) Linear combination fittings of Ni K-edge XANES spectra of (d) Ni/CeO₂-HSi(OEt)₃ and (e) Ni/CeO₂-NaBH₄.



(i)

Fig. S3 (i) k^3 -weighted Fourier-transformed Ni K-edge EXAFS spectra of Ni-foil, (a) Ni(OH)_x/CeO₂, (b) Ni/CeO₂-HBpin, (c) Ni/CeO₂-HSi(OEt)₃, (d) Ni/CeO₂-NaBH₄, (e) Ni/CeO₂-NaNaph, (f) Ni/CeO₂-HBpin-used, (g) Ni/CeO₂-0.5HBpin, and (h) Ni/CeO₂-4HBpin. Experimental data and fitted data are shown. FFT parameters, *k*-range: 3–12, window: Hanning. All *R*-space spectra are shown without phase correction.



(ii)

Fig. S3 (continued) (ii) Raw *k*³-weighted Ni K-edge EXAFS spectra of Ni-foil, (a) Ni(OH)*x*/CeO₂, (b) Ni/CeO₂-HBpin, (c) Ni/CeO₂-HSi(OEt)₃, (d) Ni/CeO₂-NaBH₄, (e) Ni/CeO₂-NaNaph, (f) Ni/CeO₂-HBpin-used, (g) Ni/CeO₂-0.5HBpin, and (h) Ni/CeO₂-4HBpin. Experimental data and fitted data are shown. FFT parameters, *k*-range: 3–12, window: Hanning. All *R*-space spectra are shown without phase correction.



Fig. S4 (a) HAADF-STEM image of Ni/CeO₂-HBpin; (b, c) STEM-EDS mapping of Ni/CeO₂-HBpin, showing the distribution of O in blue (panel (b)), and the distribution of B in yellow (panel (c)); (d) Overlap of the STEM-EDS mapping of Ni/CeO₂-HBpin, showing the distributions of Ni in magenta and Ce in cyan; (e) Overlap of the STEM-EDS mapping of Ni/CeO₂-HBpin, showing the distributions of Ce in cyan and B in yellow; (f) Overlap of the STEM-EDS mapping of Ni/CeO₂-HBpin, showing the distributions of B in yellow; (f) Overlap of the STEM-EDS mapping of Ni/CeO₂-HBpin, showing the distributions of B in yellow; (f) Overlap of the STEM-EDS mapping of Ni/CeO₂-HBpin, showing the distributions of B in yellow and Ni in magenta



Fig. S5 X-ray photoelectron spectra of (a) Ni/CeO₂-HBpin and (b) Ni/CeO₂-NaBH₄ around the B 1s region. The binding energies were calibrated by using the C 1s signal at 284.8 eV.



Fig. S6 X-ray photoelectron spectra of (a) Ni/CeO₂-HBpin, (b) Ni/CeO₂-NaNaph around the Ni 2p region. Black lines indicate the data plots. The gray dotted lines indicate the deconvoluted signals. The red dotted lines indicate the sum of the lines of the deconvoluted signals. The binding energies were calibrated by using the C 1s signal at 284.8 eV.



Fig. S7 The images of Ni(OH)_x/CeO₂ before/after the reaction of borylation or silylation of **1a** using HBpin or HSi(OEt)₃, respectively. Reaction conditions: **1a** (2 mmol), HBpin or HSi(OEt)₃ (0.4 mmol), catalyst (Ni: 1.8 mol%), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 15 min.



Fig. S8 X-ray photoelectron spectra of (a) Ni/CeO₂-NaBH₄ and (b) Ni/CeO₂-NaNaph around the Na 1s region. The binding energies were calibrated by using the C 1s signal at 284.8 eV.



Fig. S9 X-ray photoelectron spectra of (a) Ni/CeO₂-NaNaph and (b) Ni/CeO₂-HBpin around the C 1s region. Black lines indicate the data plots. The gray dotted lines indicate the deconvoluted signals. The red dotted lines indicate the sum of the lines of the deconvoluted signals. The π - π * satellite in (a) is highlighted in violet line.^{S7} The binding energies were calibrated by using the C 1s signal at 284.8 eV.



Fig. S10 X-ray photoelectron spectra of (a) Ni/CeO₂-HBpin, (b) Ni/CeO₂-NaBH₄ around the Ce 3d region. Black lines indicate the data plots. The green lines indicate the deconvoluted signals of Ce⁴⁺ species, including the satellite peaks u''', u'', v''', v'', and the main peaks u, v. The red lines indicate the deconvoluted signals of Ce³⁺ species, including the satellite peaks u', v', and the main peaks u', v', and the main peaks u, v. The red lines indicate the sum of the lines of the deconvoluted signals. The ratios of Ce³⁺/Ce⁴⁺ was calculated based on the area of Ce 3d_{5/2} region (~875–895 eV). The equation to

calculate the percentages of Ce³⁺ species was as follows: $\frac{Area(Ce^{3+}) \times 100}{Area(Ce^{3+}) + Area(Ce^{4+})}$.⁸⁸ The

binding energies were calibrated by using the C 1s signal at 284.8 eV.



Ce³⁺ : Ce⁴⁺ = 1 : 6.3

25000

20000



$$Area(Ce^{3+}) \times 100$$

V

Vo

calculate the percentages of Ce^{3+} species was as follows: $\overline{Area(Ce^{3+}) + Area(Ce^{4+})}$. se The binding energies were calibrated by using the C 1s signal at 284.8 eV.

(c)



Fig. S11 Ni K-edge XANES spectra of Ni-foil, (a) Ni(OH)_x/CeO₂, (b) Ni/CeO₂-HBpin, (c) Ni/CeO₂-0.5HBpin, (d) Ni/CeO₂-HBpin-used, (e) Ni/CeO₂-4HBpin.



Fig. S12 Effect of removal of Ni/CeO₂-HBpin catalyst on the dehydrogenative benzylic silylation of toluene (**1a**). Reaction conditions: **1a** (2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), HSi(OEt)₃ (0.4 mmol), methylcyclohexane (1 mL), 120 °C, Ar (1 atm). Yields were determined by GC analysis using *n*-hexadecane as an internal standard.



Fig. S13 IR-ATR spectra of fresh (marked in black) and used Ni/CeO₂-HBpin (marked in blue). The used catalyst was obtained under the conditions in Table 1, entry 11.



Fig. S14 Ni/CeO₂-HBpin-catalyzed benzylic C(sp³)–H dehydrogenative of **1a** in a gram-scale. Reaction conditions: **1a** (29 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), HSi(OEt)₃ (5.8 mmol), methylcyclohexane (7 mL), 120 °C, 24 h, Ar (1 atm). Yields were determined by GC analysis using *n*-hexadecane as an internal standard.



Fig. S15 Limitation of substrates. Reaction conditions are indicated in the figure.



Fig. S16 The reaction profile of the dehydrogenative benzylic silylation of toluene (**1a**) with or without 3,5-di-*tert*-butyl-4-hydroxytoluene (BHT). Reaction conditions: **1a** (2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), HSi(OEt)₃ (0.4 mmol), BHT (0 or 0.4 mmol), methylcyclohexane (1 mL), 120 °C, Ar (1 atm). Yields were determined by GC analysis using *n*-hexadecane as an internal standard.



Fig. S17 (a) m/z distribution of heptylbenzenes between m/z = 176.2 and m/z = 182.2 and (b) ¹H NMR spectra (500 MHz, CD₃OD) of heptylbenzenes before/after the reaction. Reaction conditions: hepylbenzene- d_2 (0.2 mmol), HSi(OEt)₃ (0 or 0.4 mmol), Ni/CeO₂-HBpin (Ni: 1.8 mol%), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 4 h.



Fig. S17 (continued) (c) ²H NMR spectra (77 MHz, CH₃OH) of the heptylbenzene- d_2 during the reaction catalyzed by Ni/CeO₂-HBpin with or without HSi(OEt)₃. Reaction conditions: hepylbenzene-1,1- d_2 (0.2 mmol), HSi(OEt)₃ (0.4 mmol), Ni/CeO₂-HBpin (Ni: 1.8 mol%), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 4 h.



Fig. S18 GC-MS spectra of (a) $HSi(OEt)_3$ and (b) $HSiMe_2Ph$ before/after the reactions with toluene- d_8 in the presence of Ni/CeO₂-HBpin. Reaction conditions: $HSi(OEt)_3$ or $HSiMe_2Ph$ (0.4 mmol), toluene- d_8 (0.2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 3.6 mol%), methylcyclohexane (1 mL), 120 °C, 4 h, Ar (1 atm).



Fig. S19 Dependence of the amount of HSi(OEt)₃ on the production rate. Reaction conditions: **1a** (2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), HSi(OEt)₃ (0.1, 0.2, 0.4, 1, or 2 mmol), methylcyclohexane (1 mL), 120 °C, Ar (1 atm).



Fig. S20 Dependence of the amount of toluene (1a) on the production rate. Reaction conditions: 1a (0.4, 1, 2, 4, or 10 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), HSi(OEt)₃ (0.4 mmol), methylcyclohexane (1 mL), 120 °C, Ar (1 atm).



Fig. S21 Initial production rates of the dehydrogenative benzylic silylation of toluene (**1a**, marked in black) and toluene- d_8 (**1a**- d_8 , marked in orange). Reaction conditions: **1a** or **1a**- d_8 (2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), HSi(OEt)₃ (0.4 mmol), methylcyclohexane (1 mL), 120 °C, Ar (1 atm). The product concentrations were calculated by GC analysis.



Fig. S22 (a) Initial production rates and (b) reaction profiles of the dehydrogenative benzylic silylation of toluene (**1a**) in an Ar atmosphere (marked in black) or in an H₂ atmosphere (marked in green). Reaction conditions: **1a** (2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), HSi(OEt)₃ (0.4 mmol), methylcyclohexane (1 mL), 120 °C, Ar or H₂ (1 atm). The product concentrations and yields were calculated by GC analysis.



Fig. S23 Ni/CeO₂-HBpin-catalyzed benzylic C(sp³)–H dehydrogenative (a) borylation or (b) silylation of heptylbenzene. Reaction conditions: heptylbenzene (0.2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 3.6 mol%), pinacolborane or HSi(OEt)₃ (0.4 mmol), methylcyclohexane (1 mL), 120 °C, 4 h, Ar (1 atm).

Supplementary Tables

Reference	Silylating reagent	Scope of Substrates (Number of examples)	Reaction time	Regioselectivity
This work	HSi(OEt) ₃	alkylarenes (15)	1–24 h	primary benzylic
S9	HSiMe ₃	$HSiMe_3(1)$	425 h	_
S10	HSiEt ₃ , Me ₃ SiSiMe ₃	toluene (1) (trace)	16.5 h	aryl C(sp ²)–H
S11	1,2-benzenedisilane	mesitylene (1)	30 h	benzylic
S12	HSiEt ₃	$HSiEt_3(1)$	12 h	—
S13	H ₃ SiMes, H ₂ SiPh ₂	CH ₄ , cyclopropane (2)	168–480 h	_
S5	HSiEt ₃ , HSi(OEt) ₃	alkylarenes (9)	48–144 h	benzylic
S14	H_2SiEt_2	toluene, ethylbenzene (2)	20 h	benzylic
S15	HSiEt ₃	alkylarenes (4)	96–120 h	benzylic
S6	tBu–N=N–SiR ₃	alkylarenes (>30)	1.5–18 h	benzylic

Table S1. Comparison of this work with previous reports on undirected $C(sp^3)$ -H silylation.

Sample	Shell	C.N.	<i>R</i> (Å)	ΔE_{j0} (eV)	$\sigma^{2 \times 10^{2}}$ (Å ²)	R-factor (%)
	Ni–O	2.6±0.7	2.02	-4.72	1.15	
Ni/CeO ₂ -HBpin	Ni–Ni (in Ni)	2.0±0.6	2.45	-10.0	1.03	1.40
_	Ni–Ni (in Ni(OH) ₂)	1.1±1.1	3.12	-10.0	1.88	
	Ni–O	2.9±0.9	2.03	-2.34	1.47	
Ni/CeO ₂ -4HBpin	Ni–Ni (in Ni)	2.5±0.8	2.45	-9.33	1.05	2.48
	Ni–Ni (in Ni(OH) ₂)	0.5±0.3	3.13	10.0	1.50	
Ni/CeO ₂ -HSi(OEt) ₃	Ni–O	4.5±0.4	2.02	-6.44	0.76	0.88
	Ni–O	3.0±0.4	2.03	-3.95	0.66	
Ni/CeO ₂ -NaBH ₄	Ni–Ni (in Ni)	1.2±0.5	2.46	-10.0	10.0	0.74
	Ni–Ni (in Ni(OH) ₂)	0.7±0.4	3.16	10.0	0.86	
	Ni–O	3.0±0.2	2.03	-4.03	0.088	
Ni/CeO ₂ -HBpin (used)	Ni–Ni (in Ni)	1.3±0.6	2.46	-10.0	1.27	0.79
	Ni–Ni (in Ni(OH) ₂)	1.0±0.4	3.11	-10.0	1.41	
	Ni–O	3.2±0.6	2.04	-3.45	0.69	
Ni/CeO ₂ -0.5HBpin	Ni–Ni (in Ni)	1.8±0.9	2.54	-0.22	1.27	1.94
_	Ni–Ni (in Ni(OH) ₂)	0.7±0.4	3.18	10	1.22	
	Ni–O	1.4	2.04	0.27	0.63	
	Ni–Ni (in Ni)	3.1	2.48	-7.88	0.81	
Ni/CeO ₂ -NaNaph ^{S2}	Ni–Ni (in Ni(OH) ₂)	0.4	3.17	10.66	0.59	1.01
	Ni–Ni (in Ni)	3.3	4.39	-4.89	1.07	
	Ni–Ni (in Ni)	2.5	5.04	-9.16	0.93	

 Table S2. Fitted parameters from Ni K-edge EXAFS.

Table S3. Effect of catalysts on dehydrogenative benzylic silylation of toluene (1a) for 24 h.

1a (2 mmol)	catalyst (1.8 mol%) HSi(OEt) ₃ 0.4 mmol methylcyclohexane (MCH) 120 °C, 24 h	Si(OEt) ₃ + 2a	2a'
onter	aatalvat	3	vield [%]
entry	cataryst	2a	2a'
1	Ni/CeO ₂ -HBpin	80	<1
2	Ni/CeO ₂ -HSi(OEt) ₃	16	<1
3	Ni/CeO ₂ -NaBH ₄	<1	<1
4	Ni/CeO ₂ -NaNaph	<1	<1

Reaction conditions: **1a** (2 mmol), $HSi(OEt)_3$ (0.4 mmol), catalyst (Ni: 1.8 mol%), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 24 h. Yields were determined by GC analysis using *n*-hexadecane as an internal standard.

Table S4. Effect of the additional catalysts on Ni/CeO₂-HBpin-catalyzed 1a silylation with HSi(OEt)₃.



entry	additional catalyst —	yield	1 [%]
		2a	2a'
1	-	79	<1
2	CeO_2 (32 mg)	77	3
3	CeO ₂ -HBpin (32 mg)	65	<1
4	Ni/CeO ₂ -NaBH ₄	7	<1
5	Ni/CeO ₂ -NaNaph	18	<1

Reaction conditions: **1a** (2 mmol), $HSi(OEt)_3$ (0.4 mmol), Ni/CeO_2 -HBpin (Ni: 1.8 mol%), additive (Ni: 1.8 mol% or 32 mg), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 15 min. Conversions and yields were determined by GC analysis using *n*-hexadecane as an internal standard.

Table S5.	Effect of the pretreated CeO ₂ supp	oorts on the conversio	n of HSi(OEt) ₃	without 1a	ı or Ni
catalysts.					

	HSi(OEt) ₃ ——— 0.4 mmol	Catalyst (32 mg) <u>MCH 1 mL</u> → Si(OEt) ₄ 120 °C, 1 h 3 1 atm Ar 3	
entry	catalyst	conv. [%]	yield of 3 [%]
1	CeO ₂ -HBpin	18	2
2	CeO ₂ -NaBH ₄	56	30
3	CeO ₂ -3NaNaph	>99	34

Reaction conditions: $HSi(OEt)_3$ (0.4 mmol), catalyst (32 mg), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 1 h. Conversions and yields were determined by GC analysis using *n*-hexadecane as an internal standard.

1a (2 mmol)	catalyst (1.8 mol%) HSi(OEt) ₃ 0.4 mmol methylcyclohexane (MCH) 120 °C, 15 min	2a	^Si(OEt) ₃ +	2a'	3
ontry	ootolyst			yield [%]	
entry	cataryst		2a	2	2a'
1	Ni/CeO ₂ -4HBpin		44	~	<1
2	Ni/CeO ₂ -HBpin		79	~	<1
3	Ni/CeO ₂ -0.5HBpin		46	<	<1

Table S6. Optimization of the amount of reducing agent used in the catalyst pretreatment for silylation of toluene (1a).

Reaction conditions: **1a** (2 mmol), $HSi(OEt)_3$ (0.4 mmol), catalyst (Ni: 1.8 mol%), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 15 min. Yields were determined by GC analysis using *n*-hexadecane as an internal standard.

1a (2 mmol)	catalyst (1.8 mol%) HSi(OEt) ₃ 0.4 mmol methylcyclohexane (MCH) 120 °C, 15 min	Si(OEt) ₃ + 2a	2a'
out my			yield [%]
entry	catalyst	2a	2a'
1	Ni/CeO ₂ -0.5HBpin	46	<1
2	Mn/CeO ₂ -0.5HBpin	<1	<1
3	Fe/CeO ₂ -0.5HBpin	<1	<1
4	Co/CeO ₂ -0.5HBpin	<1	<1
5	Cu/CeO ₂ -0.5HBpin	<1	<1
6	Zn/CeO ₂ -0.5HBpin	<1	<1

Table S7. Effect of metals on dehydrogenative benzylic silylation of toluene (1a).

Reaction conditions: **1a** (2 mmol), $HSi(OEt)_3$ (0.4 mmol), catalyst (metal: 1.8 mol%), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 15 min. Yields were determined by GC analysis using *n*-hexadecane as an internal standard.

Table S8. Effect of solvents on dehydrogenative benzylic silylation of toluene (1a).

	Ni/CeO ₂ -HBpin (1.8 mol%) HSi(OEt) ₃ 0.4 mmol solvent 1 mL	Si(OEt) ₃ +	DEt) ₃
1a (2 mmol)	120 °C, 15 min, Ar 2a	2a'	
		yield [9	%]
entry	sorvent	2a	2a'
1	heptane	81	<1
2	methylcyclohexane (MCH)	79	<1
3	cyclopentyl methyl ether (CPME)	73	<1
4	1,4-dioxane	77	1
5	N,N-dimethylacetamide (DMA)	53	<1

Reaction conditions: **1a** (2 mmol), $HSi(OEt)_3$ (0.4 mmol), Ni/CeO_2 -HBpin (Ni: 1.8 mol%), solvent (1 mL), Ar (1 atm), 120 °C, 15 min. Yields were determined by GC analysis using *n*-hexadecane as an internal standard.

1a (2 mmol)	Ni/CeO ₂ -HBpin (1.8 HSi(OEt) ₃ 0.4 m methylcyclohexane temp., time, A	mol%) mol (MCH) Si((r 2a	DEt) _{3 +}	Si(OEt) ₃
ontra	town [°C]	timo [min]	yield	1 [%]
entry	temp. [C]		2a	2a'
1	80		15	<1
2	100	15	62	<1
3	120	15	79	<1
4	140		81	<1
5	80		41	<1
6	100	60	86	<1
7	120	00	93	<1
8	140		91	<1

 Table S9. Effect of temperature on dehydrogenative benzylic silylation of toluene (1a).

Reaction conditions: **1a** (2 mmol), $HSi(OEt)_3$ (0.4 mmol), Ni/CeO_2 -HBpin (Ni: 1.8 mol%, 1.5 wt%), methylcyclohexane (1 mL), Ar (1 atm). Yields were determined by GC analysis using *n*-hexadecane as an internal standard.

1a (X mmol)	Ni/CeO ₂ -HBpin (1.8 mol%) HSi(OEt) ₃ 0.4 mmol methylcyclohexane (MCH) 120 °C, 1 h, Ar	Si(OEt) ₃ + (2 2a	[⊥] Si(OEt) ₃ 2a'
antm	amount of 1 [mmol]	yie	ld [%]
entry		2a	2a'
1^a	0.2	58	<1
2	0.4	57	<1
3	1	87	<1
4	2	93	<1

 Table S10. Effect of the amount of 1a on dehydrogenative benzylic silylation of 1a.

Reaction conditions: **1a** (0.2, 0.4, 1, or 2 mmol), $HSi(OEt)_3$ (0.4 mmol), Ni/CeO_2 -HBpin (Ni: 1.8 mol%, based on the silvlating reagents), methylcyclohexane (1 mL), Ar (1 atm), 120 °C. Yields were determined by GC analysis using *n*-hexadecane as an internal standard (based on HSi(OEt)₃). ^{*a*}24 h. The yields were calculated based on **1a**.

R ^{II} + 1 (2 mmol)	H <mark>[Si]</mark> (0.4 mmol)	(1.5 wt%, Ni; 1.8 m [Si] = Si(OEt) ₃ , MCH 120 °C, 24 h, Ar 1 a	ol%) 1 mL <mark>➤</mark> R atm	2	`[Si] ₊		$ \begin{array}{c} \begin{array}{c} & & Si(OEt)_3 \\ + & Si(OEt)_4 & + O \\ \hline \\ \hline \\ \hline \\ \hline \\ \hline \\ \hline \\ \\ \\ \hline \\ \\ \\ \hline \\ \\ \\ \\ \hline \\$
Entry	Substrate	Conv. of HSi(OEt) ₃ [%]	Yield [%]				
			2	3	4	5	disilylated product
1^b	1a	>99	93	1	1	<1	<1
2^c	1b	99	74	1	10	<1	5
3^d	1c	99	85	<1	4	<1	2
4	1d	99	77	<1	4	6	5
5^e	1e	96	63	<1	8	<1	<1
6 ^f	1f	96	62	<1	5	1	<1
7^d	1g	98	77	5	8	<1	1
8^d	1h	>99	49	<1	13	1	<1
9	1i	>99	46	2	8	<1	2
10 ^g	1j	91	41	2	20	1	3
11	1k	99	60	<1	4	6	1
12 ^f	11	95	80	<1	<1	<1	4
13 ^h	1m	77	55 (1)	<1	<1	1	<1
14 ^e	1n	95	70 (1)	<1	<1	<1	<1
15 ^g	10	90	30 (< 1)	<1	<1	<1	<1

Table S11. The details about of the investigation of the substrate scope of the present regioselective dehydrogenative silylation.^a

Ni/CeO₂-HBpin

aReaction conditions: 1 (2 mmol), HSi(OEt)₃ (0.4 mmol), Ni/CeO₂-HBpin (32 mg, Ni: 1.8 mol%), methylcyclohexane (1 mL), Ar (1 atm), 120°C, 24 h. Yields were determined via GC using nhexadecane as an internal standard. The regioisomer yields (secondary C(sp³)-H silylation) determined via GC are shown in parentheses. ^b1 h. ^c6 h. ^d8 h. ^eHSi(OEt)₃ (0.1 mmol), 140°C. ^fHSi(OEt)₃ (0.1 mmol). ^gHSi(OEt)₃ (0.2 mmol). ^hUsing biphenyl as an internal standard.

1a (2 mmol)	Ni/CeO ₂ -HBpin (1.8 mol%) H[Si] 0.4 mmol methylcyclohexane (MCH) 120 °C, 15 min, Ar	2a Si(OEt) ₃	+ Si(OEt) ₃ 2a'		
ontra	cilulating reagant	yield [%]			
entry	sirylating reagent	2a	2a'		
1	HSi(OEt) ₃	79	<1		
2	HSi(OMe) ₃	8	<1		
3	HSiEt ₃	<1	<1		
4	HSiMe ₂ Ph	<1	<1		
5	HSiPh ₃	<1	<1		

Table S12. Effect of different kinds of silvlating reagents on dehydrogenative benzylic silvlation of toluene (1a).

Reaction conditions: **1a** (2 mmol), Ni/CeO₂-HBpin (Ni: 1.5 wt%, 1.8 mol%), silylating reagent (0.4 mmol), methylcyclohexane (1 mL), Ar (1 atm), 120 °C, 15 min. Yields were determined by GC analysis using *n*-hexadecane as an internal standard.

Supplementary References

S1 D. Yoshii, T. Yatabe, T. Yabe and K. Yamaguchi, ACS Catal., 2021, 11, 2150–2155.

S2 T. Matsuyama, T. Yatabe, T. Yabe and K. Yamaguchi, ACS Catal., 2021, 11, 13745–13751.

S3 T. Kurita, K. Hattori, S. Seki, T. Mizumoto, F. Aoki, Y. Yamada, K. Ikawa, T. Maegawa, Y. Monguchi and H. Sajiki, *Chem. Eur. J.*, 2008, **14**, 664–673.

S4 Z. Wang and S. Chang, Org. Lett., 2013, 15, 1990–1993.

S5 K. Manna, P. Ji, Z. Lin, F. X. Greene, A. Urban, N. C. Thacker and W. Lin, *Nat Commun.*, 2016, **7**, 12610.

S6 B. Neil, L. Saadi, L. Fensterbank and C. Chauvier, *Angew. Chem. Int. Ed.* 2023, **62**, e202306115.

S7 M. Campos and P. A. P. Nascente, Synth. Met., 2010, 160, 1513–1519.

S8 (a) E. Paparazzo, *Mater. Res. Bull.*, 2011, 46, 323–326; (b) D. R. Mullins, *Surf. Sci. Rep.*, 2015, 70, 42–85.

S9 L. J. Procopio and D. H. Berry, J. Am. Chem. Soc., 1991, 113, 4039-4040.

S10 T. Sakakura, Y. Tokunaga, T. Sodeyama and M. Tanaka, Chem. Lett., 1987, 2375–2378.

S11 M. Ishikawa, S. Okazaki, A. Naka and H. Sakamoto, Organometallics, 1992, 11, 4135–4139.

S12 P. I. Djurovich, A. R. Dolich and D. H. Berry, J. Chem. Soc., Chem. Commun., 1994, 1897–1898.

S13 A. D. Sadow and T. D. Tilley, Angew. Chem. Int. Ed., 2003, 42, 803–805.

S14 T. Baba, A. Kato, H. Yuasa, F. Toriyama, H. Handa and Y. Ono, *Catal. Today*, 1998, 44, 271–276.

S15 A. A. Toutov, W.-B. Liu, K. N. Betz, A. Fedorov, B. M. Stoltz and R. H. Grubbs, *Nature*, 2015, **518**, 80–84.

NMR Spectra



¹H NMR spectrum (500 MHz, CDCl₃) of 2a



¹³C NMR spectrum (126 MHz, CDCl₃) of **2b**



S53

¹H NMR spectrum (500 MHz, CDCl₃) of **2c**



¹³C NMR spectrum (126 MHz, CDCl₃) of **2c**



¹H NMR spectrum (500 MHz, CDCl₃) of **2d**



¹H NMR spectrum (500 MHz, CDCl₃) of **2e**



¹³C NMR spectrum (126 MHz, CDCl₃) of 2e



¹H NMR spectrum (500 MHz, CDCl₃) of **2f**



¹³C NMR spectrum (126 MHz, CDCl₃) of **2f**



¹H NMR spectrum (500 MHz, CDCl₃) of **2g**



¹³C NMR spectrum (126 MHz, CDCl₃) of **2g**



¹H NMR spectrum (500 MHz, CDCl₃) of **2h**



¹³C NMR spectrum (126 MHz, CDCl₃) of **2h**



¹H NMR spectrum (500 MHz, CDCl₃) of 2i



¹³C NMR spectrum (126 MHz, CDCl₃) of 2i



¹H NMR spectrum (500 MHz, CDCl₃) of **2j**



¹H NMR spectrum (500 MHz, CDCl₃) of **2k**



¹³C NMR spectrum (126 MHz, CDCl₃) of **2k**



S62

¹H NMR spectrum (500 MHz, CDCl₃) of **2l**



¹H NMR spectrum (500 MHz, CDCl₃) of **2m**



¹³C NMR spectrum (126 MHz, CDCl₃) of **2m**







X : parts per Million : Proton

¹³C NMR spectrum (126 MHz, CDCl₃) of **2n**



¹H NMR spectrum (500 MHz, CDCl₃) of **20**



¹³C NMR spectrum (126 MHz, CDCl₃) of **20**







²H NMR spectrum (77 MHz, CH₃OH) of heptylbenzene-1,1- d_2

