

Supporting Information for

**Markovnikov selective hydroaminocarbonylation of alkenes over  
a porous monophosphine polymer supported palladium catalyst**

Jiajun Li <sup>a,b</sup>, Kang Zhao<sup>a</sup>, Xinjiang Cui<sup>a</sup>, Lailai Wang<sup>a,\*</sup>, Feng Shi <sup>a,\*</sup>

<sup>a</sup>. State Key Laboratory of Low Carbon Catalysis and Carbon Dioxide Utilization,

State Key Laboratory for Oxo Synthesis and Selective Oxidation

Lanzhou Institute of Chemical Physics, Chinese Academy of Sciences

No.18, Tianshui Middle Road, Lanzhou, 730000, China

<sup>b</sup>. University of Chinese Academy of Sciences, No. 19A, Yuquanlu, Beijing, 100049, People's

Republic of China

E-mail: wll@licp.cas.cn, fshi@licp.cas.cn

## Contents

<b>1. General Information</b> .....	<b>3</b>
Chemicals and materials .....	3
Instrumental measurements and physical characterization .....	3
<b>2. Synthetic procedures of the ligands</b> .....	<b>4</b>
<b>3. Synthetic procedures of the porous organic polymer catalysts</b> .....	<b>5</b>
<b>4. General procedures for the hydroaminocarbonylation</b> .....	<b>6</b>
<b>5. Optimization of the reaction conditions</b> .....	<b>7</b>
<b>6. Controlled experiment</b> .....	<b>11</b>
<b>7. NMR data of the products</b> .....	<b>12</b>
<b>8. Catalyst characterization</b> .....	<b>20</b>
Experimental PXRD profiles .....	20
<b>In-stiu DRIFTS spectra</b> .....	21
Scanning electron micrographs .....	22
Transmission electron micrographs .....	23
N <sub>2</sub> adsorption-desorption analysis .....	25
TGA analysis .....	28
X-ray photoelectron spectroscopy analysis .....	29
<b>9. Reaction mechanism</b> .....	<b>31</b>
<b>10. Copies of quantitative GC and NMR spectra</b> .....	<b>32</b>
<b>11. References</b> .....	<b>64</b>

## 1. General Information

### Chemicals and materials

All solvents and chemicals, unless otherwise noted, were obtained commercially and were used as received without further purification. All glassware was dried before using. Analytical thin layer chromatography (TLC) was performed using pre-coated Jiangyou silica gel HSGF254 (0.2mm  $\pm$  0.03mm). Flash chromatography was performed using silica gel 60, 0.063-0.2 mm, 200-300 mesh (Jiangyou, Yantai) with the indicated solvent system.

### Instrumental measurements and physical characterization

Gas chromatography analysis was performed on Agilent 7890A GC equipped with a HP-5 capillary column and FID detector. GC-MS analysis was in general recorded on an Agilent 5977A MSD GC-MS. The contents of Pd in the catalysts were measured by inductively coupled plasma-atomic emission spectrometry (ICP-AES), using Iris advantage Thermo Jarrel Ash device.

Fourier transform infrared (FT-IR) spectrum were recorded with a Bruker VERTEX 70FTIR spectrometer.

The liquid nuclear magnetic resonance spectra (NMR) were recorded on a Bruker Avance<sup>TM</sup> III 400 MHz in deuterated chloroform unless otherwise noted. Data are reported in parts per million (ppm) as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, m = multiplet, dd = doublet of doublet and br = broad signal), coupling constant in Hz and integration.

Powder X-ray diffraction (PXRD) measurements were conducted by a STADIP automated transmission diffractometer (STOE) equipped with an incident beam curved germanium monochromator selecting CuK $\alpha$ 1 radiation and a 6 $^\circ$  position sensitive detector (PSD) (step size: 0.014 $^\circ$ , step time: 25.05 s). The XRD patterns were scanned in the 2 $\theta$  range of 0-80 $^\circ$ .

Nitrogen adsorption-desorption isotherms were measured at 77 K using an American Quantachrome iQ<sub>2</sub> automated gas sorption analyzer. The samples were outgassed at 120  $^\circ$ C for 12 h before the measurements. Surface areas were calculated from the adsorption data using Langmuir and Brunauer-Emmett-Teller (BET) methods. The pore-size-distribution curves were obtained from the adsorption branches using non-local density functional theory (NLDFT) method.

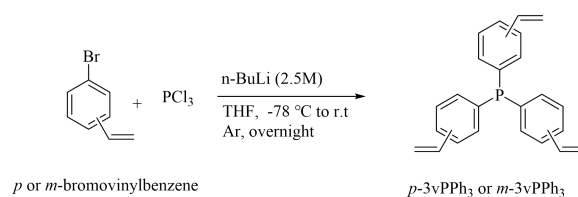
X-ray photoelectron spectroscopy (XPS) measurements were carried out by a VG ESCALAB 210 instrument equipped with a dual Mg/Al anode X-ray source, a hemispherical capacitor analyzer, and a 5 keV Ar<sup>+</sup> ion gun. All spectra were recorded by using AlK $\alpha$  (1361 eV) radiation. The electron binding energy was referenced to the C1s peak at 284.8 eV.

The thermal properties of Pd/POL-*m*-3v-PPh<sub>3</sub> catalysts were evaluated using a METTLER TOLEDO simultaneous thermal analyzer over the temperature range from 30 to 800  $^\circ$ C under nitrogen atmosphere (20 mL/min) with a heating rate of 5  $^\circ$ C/min.

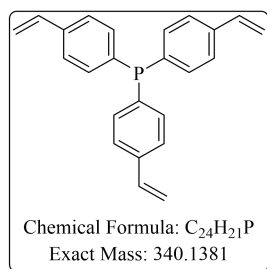
Field emission scanning electron microscopy (SEM) observations were performed on a Hitachi S-4800 microscope operated at an accelerating voltage of 5.0 kV.

High-resolution transmission electron microscope (HR-TEM) analysis was carried out on a Talos F200S operating at 200 kV.

## 2. Synthetic procedures of the ligands



Organic phosphine monomer was synthesized according to literature reports<sup>1,2</sup>. First of all, a solution of *n*-butyllithium in hexane (2.5 M, 12.6 ml, 31.5 mmol) was added dropwise over a period of 20 min to a solution of *p* or *m*-bromovinylbenzene (6.04 g, 33 mmol) in anhydrous tetrahydrofuran (40 ml) at -78 °C under argon atmosphere. The solution was stirred for 1 h and then phosphorus trichloride (1.37 g, 10 mmol) dissolved in anhydrous tetrahydrofuran (5 ml) was added dropwise over a period of 5 min. The mixture was continued to stir at -78 °C for 1 h and the system was recovered to room temperature, and allowed to react overnight. The reaction was quenched with 2M HCl solution. The mixture was extracted with ethyl acetate and water for 3 times, the combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel using EtOAc–petroleum ether mixture as an eluent to afford the desired compound *p*-3vPPh<sub>3</sub> or *m*-3vPPh<sub>3</sub> as a white solid.

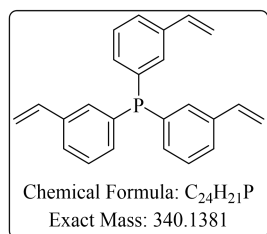


**Tris(4-vinylphenyl)phosphane (*p*-3vPPh<sub>3</sub>):** white solid, 2.4 g, 70% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.36 (d, *J* = 7.0 Hz, 6H), 7.27 (t, *J* = 7.7 Hz, 6H), 6.69 (dd, *J* = 17.6, 10.9 Hz, 3H), 5.76 (d, *J* = 17.6 Hz, 3H), 5.26 (d, *J* = 10.9 Hz, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 138.09, 136.81, 136.70, 136.47, 134.09, 133.90, 126.47, 126.40, 114.82.

<sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ -6.85.



**Tris(3-vinylphenyl)phosphane (*m*-3vPPh<sub>3</sub>):** white solid, 2.4 g, 70% yield.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.39 (d, *J* = 8.1 Hz, 6H), 7.34 – 7.24 (m, 3H), 7.17 (t, *J* = 7.2 Hz, 3H), 6.64 (dd, *J* = 17.6, 10.9 Hz, 3H), 5.67 (d, *J* = 17.6 Hz, 3H), 5.21 (d, *J* = 11.0 Hz, 3H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 137.80 (d, *J* = 7.7 Hz), 137.33 (d, *J* = 11.1 Hz), 136.59, 133.13 (d, *J* = 16.5 Hz), 132.01 (d, *J* = 23.0 Hz), 128.85 (d, *J* = 6.3 Hz), 126.63, 114.55.

<sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ -5.15.

### 3. Synthetic procedures of the porous organic polymer catalysts

#### Preparation of POL-*p*-3vPPh<sub>3</sub> or POL-*m*-3vPPh<sub>3</sub>

Under an argon atmosphere, 0.6 g of *p*-3vPPh<sub>3</sub> (*m*-3vPPh<sub>3</sub>) was dissolved in 6 ml of THF. Then, 15 mg of AIBN was added to the solution. The resulting mixture was transferred into an autoclave and stirred at room temperature for 30 minutes. Afterward, it was heated to 100 °C for 24 h without stirring. When the polymerization was complete, stop heating and allow to cool to room temperature. The resulting white solid was filtered, washed with THF (20 ml × 3), and dried under vacuum at 60 °C for 12 h. Finally, POL-*p*-3vPPh<sub>3</sub> was obtained.

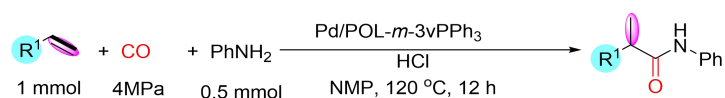
#### Preparation of Pd/POL-*p*-3vPPh<sub>3</sub> Or Pd/POL-*m*-3vPPh<sub>3</sub>

Under argon atmosphere, 13 mg of Pd (CH<sub>3</sub>CN)<sub>2</sub>Cl<sub>2</sub> was dissolved in 10 mL of THF, followed by the addition of 200 mg of POL-*p*-3vPPh<sub>3</sub> or POL-*m*-3vPPh<sub>3</sub>. After stirring for 24 h under argon atmosphere at room temperature, the resulted yellowish solid was filtered, washed with THF (20 ml × 3), and dried under vacuum at 60 °C for 12 h, quantitative yield of the Pd/POL-*m*-3vPPh<sub>3</sub> was obtained 3.066 wt.% of Pd contents was determined by inductively coupled plasma-atomic emission spectrometry (ICP-AES).

#### Preparation of Pd/POL-*m*-3vPPh<sub>3</sub>-NaBH<sub>4</sub>

The preparation of the Pd/POL-*m*-3vPPh<sub>3</sub>-NaBH<sub>4</sub> has undergone some modifications according to previous literature<sup>3</sup>. Under argon atmosphere, 76 mg sodium borohydride was added to the mixture of methanol and ethanol (v: v=10:10), and then the mixture was added drop by drop to 100 mg Pd/POL-*m*-3vPPh<sub>3</sub>, stirring at room temperature for 24 h. The solvent is removed by filtration, the black solid was filtered, washed with THF (10 ml × 3) to obtain Pd/POL-*m*-3vPPh<sub>3</sub>-NaBH<sub>4</sub> as black solid with yield of 83%.

#### 4. General procedures for the hydroaminocarbonylation



As a typical hydroaminocarbonylation recipe, the as-prepared Pd/POL-*m*-3vPPh<sub>3</sub> catalyst (20 mg), styrene (104 mg, 1.0 mmol), aniline (47 mg, 0.5 mmol), HCl (0.06 mmol) and NMP (3 ml) were added into an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. GC yield was obtained by GC analysis using n-dodecane as the internal standard, isolated yield was obtained by flash column chromatography on silica gel using EtOAc–petroleum ether mixture as an eluent.

For recycling, the Pd/POL-*m*-3vPPh<sub>3</sub> catalyst was separated by centrifugation, washed with MeOH (8.0 ml×3), and dried under vacuum at 50 °C for 3 hours. The Pd contents of the used catalyst and filtrate after each run were determined by inductively coupled plasma-atomic emission spectrometry (ICP-AES).

## 5. Optimization of the reaction conditions

### Comparison of activities of different palladium catalysts

First, five catalysts including PdCl<sub>2</sub>, PdCl<sub>2</sub>+PPh<sub>3</sub>(P/Pd=2:1), Pd/ POL-*p*-3vpph<sub>3</sub>, Pd/ POL-*m*-3vPPh<sub>3</sub>, Pd/ POL-*m*-3VPPh<sub>3</sub>-NaBH<sub>4</sub> were tested, and the dosage of Pd was 0.5 mg. The catalyst, styrene (104 mg, 1.0 mmol), aniline (47 mg, 0.5 mmol), HCl (0.6 mmol) and NMP (3 ml) were added into an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. GC yield was obtained by GC analysis using n-dodecane as the internal standard.

### Screen of the reaction solvents

Five common solvents (THF, 1,4-Dioxane, CH<sub>3</sub>CN, Toluene, NMP) were screened. Pd/POL-*m*-3vPPh<sub>3</sub> catalyst (20 mg), styrene (104 mg, 1.0 mmol), aniline (47 mg, 0.5 mmol), HCl (0.6 mmol) and solvent (3 ml) were added into an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. GC yield was obtained by GC analysis using n-dodecane as the internal standard.

### Screen of acids

Different acids were screened. Pd/POL-*m*-3vPPh<sub>3</sub> catalyst (20 mg), styrene (104 mg, 1.0 mmol), aniline (47 mg, 0.5 mmol), acid (0.6 mmol) and NMP (3 ml) were added into an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. GC yield was obtained by GC analysis using n-dodecane as the internal standard.

### Screen of the HCl dosage

The dosage of hydrochloric acid (X=0, 0.024, 0.06, 0.12, 0.6) was screened. Pd/POL-*m*-3vPPh<sub>3</sub> catalyst (20 mg), styrene (104 mg, 1.0 mmol), aniline (47 mg, 0.5 mmol), HCl (X mmol) and NMP (3 ml) were added into an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with

5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. GC yield was obtained by GC analysis using n-dodecane as the internal standard.

#### **Screen of CO pressure**

The CO pressure ( $X=0.5, 1, 2, 4$ ) was screened. Pd/POL-*m*-3vPPh<sub>3</sub> catalyst (20 mg), styrene (104 mg, 1.0 mmol), aniline (47 mg, 0.5 mmol), HCl (0.06 mmol) and NMP (3 ml) were added into an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to X MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. GC yield was obtained by GC analysis using n-dodecane as the internal standard.

#### **Screen of the reaction time**

The reaction time ( $X=0.5, 1, 2, 3, 6, 9, 12$ ) was screened. Pd/POL-*m*-3vPPh<sub>3</sub> catalyst (20 mg), styrene (104 mg, 1.0 mmol), aniline (47 mg, 0.5 mmol), HCl (0.06 mmol) and NMP (3 ml) were added into an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to X MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for X h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. GC yield was obtained by GC analysis using n-dodecane as the internal standard.



**Table S1.** Screen of the reaction temperature <sup>a</sup>

<b>Entry</b>	<b>T (°C)</b>	<b>Yield (%)<sup>b</sup></b>	<b>b/l<sup>c</sup></b>
1	60	trace	-
2	80	10	99:1
3	100	35	99:1
4	120	97	98:2

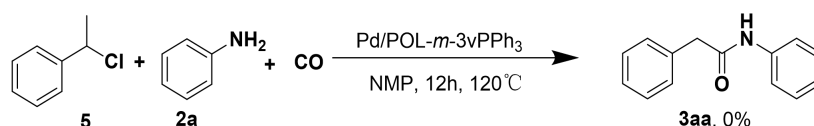
<sup>a</sup> Reaction conditions: Styrene (1.0 mmol), aniline (0.5 mmol), CO (4 MPa), Pd/POL-*m*-3vPPh<sub>3</sub> (20 mg), HCl (aq., 37wt.%, 0.12 equiv.), NMP (3 ml), 120 °C, 12 h. <sup>b</sup> Determined by GC analysis using n-dodecane as the internal standard. <sup>c</sup> Determined by GC-MS of the crude products.

**Table S2.** Pd contents in the catalysts and the filtration after each cycle, and the corresponding reaction performance <sup>a</sup>

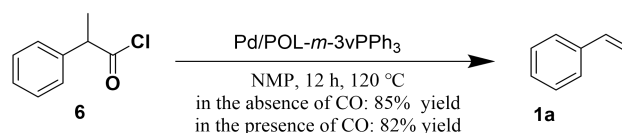
<b>Entry</b>	<b>Recycling</b>	<b>Pd contents in the catalysts (wt.%)</b>	<b>Yield (%)<sup>b</sup></b>	<b>b/I<sup>c</sup></b>
1	fresh	3.097	97	98:2
2	The 1 <sup>st</sup> run	3.066	97	98:2
3	The 2 <sup>nd</sup> run	2.917	97	98:2
4	The 3 <sup>rd</sup> run	3.035	42	97:3

<sup>a</sup> Pd content was determined by ICP-AES. Reaction conditions: Styrene (1 mmol), aniline (0.5 mmol), CO (4 MPa), Pd/POL-*m*-3vPPh<sub>3</sub> (20 mg), HCl (aq., 37wt.%, 0.12 equiv.), NMP (3 ml), 120 °C, 12 h. <sup>b</sup> Determined by GC analysis using n-dodecane as the internal standard. <sup>c</sup> Determined by GC-MS of the crude products.

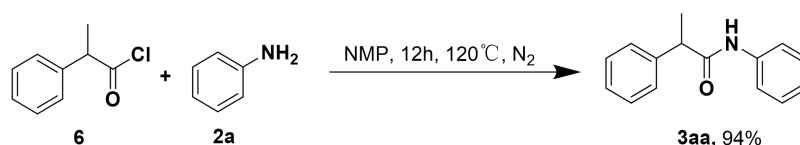
## 6. Controlled experiment



(1-chloroethyl)benzene **5** (1.0 mmol), aniline (0.5 mmol), Pd/POL-*m*-3vPPh<sub>3</sub> (20 mg) and anhydrous NMP (3 ml) were added to an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with CO for three times, the pressure of CO was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to give the ratio of b/l of the products. No amide **3aa** was observed.

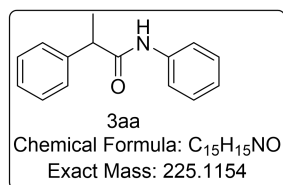


2-phenylpropanoyl chloride **6** (1.0 mmol), Pd/POL-*m*-3vPPh<sub>3</sub> (20 mg) and anhydrous NMP (3 ml) were added to an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with N<sub>2</sub> (CO) for three times, the pressure of N<sub>2</sub> (CO) was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to detected the products. The yield of styrene **1a** was 85% (82%).



2-phenylpropanoyl chloride **6** (1.0 mmol), aniline (0.5 mmol), Pd/POL-*m*-3vPPh<sub>3</sub> (20 mg) and anhydrous NMP (3 ml) were added to an 80 ml stainless-steel autoclave with a magnetic stir bar. After the autoclave was sealed and purged with N<sub>2</sub> for three times, the pressure of N<sub>2</sub> was adjusted to 4 MPa and the autoclave was put into a preheated reactor, then stirring at 120 °C for 12 h. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. Subsequently, the reaction mixture was diluted with ethyl acetate (6 ml), and the catalyst was removed from the system by centrifugation and analyzed by gas chromatography (Agilent 7890A GC equipped with a HP-5 capillary column with 5 wt.% phenyl groups and the FID detector) to detected the products. The yield of amide **3aa** was 94%.

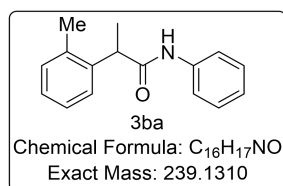
## 7. NMR data of the products



**N,2-diphenylpropanamide**<sup>3</sup>: white solid, 111 mg, 97% yield, 98% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.46 – 7.22 (m, 9H), 7.07 (dd, *J* = 18.3, 10.9 Hz, 2H), 3.71 (q, *J* = 7.1 Hz, 1H), 1.59 (d, *J* = 7.2 Hz, 3H).

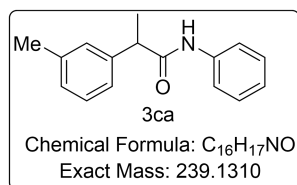
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.41, 141.05, 137.97, 129.29, 129.04, 127.84, 127.71, 124.37, 119.81, 48.25, 18.69.



**N-phenyl-2-(o-tolyl)propanamide**: white solid, 99 mg, 84% yield, 98% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.37 (t, *J* = 7.2 Hz, 3H), 7.28 – 7.20 (m, 5H), δ 7.05 (t, *J* = 7.4 Hz, 2H), 3.93 (q, *J* = 7.2 Hz, 1H), 2.36 (s, 3H), 1.59 (d, *J* = 7.2 Hz, 3H).

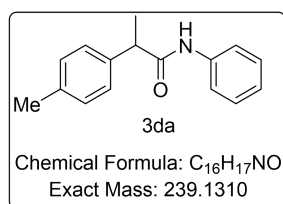
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.83, 138.92, 137.89, 136.51, 131.14, 128.98, 127.69, 127.09, 127.06, 124.36, 119.91, 44.71, 19.72, 17.77.



**N-phenyl-2-(m-tolyl)propanamide**: white solid, 102 mg, 88% yield, 99% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.61 (s, 1H), 7.44 (d, *J* = 7.8 Hz, 2H), 7.27 – 7.18 (m, 3H), 7.17 – 7.00 (m, 4H), 3.68 (q, *J* = 7.1 Hz, 1H), 2.31 (s, 3H), 1.54 (d, *J* = 7.1 Hz, 3H).

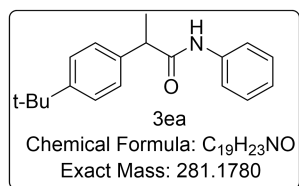
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.80, 140.98, 138.76, 138.07, 128.95, 128.89, 128.39, 128.28, 124.73, 124.23, 119.94, 47.87, 21.49, 18.62.



**N-phenyl-2-(p-tolyl)propanamide**: white solid, 106 mg, 88% yield, 96% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.50 – 7.38 (m, 3H), 7.23 (dt, *J* = 8.2, 3.8 Hz, 4H), 7.15 (d, *J* = 7.9 Hz, 2H), 7.08 – 7.00 (m, 1H), 3.67 (q, *J* = 7.1 Hz, 1H), 2.33 (s, 3H), 1.55 (d, *J* = 7.1 Hz, 3H).

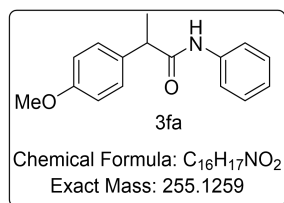
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.87, 138.04, 137.99, 137.23, 129.82, 128.91, 127.63, 124.24, 119.90, 47.61, 21.13, 18.64.



**2-(4-(tert-butyl)phenyl)-N-phenylpropanamide:** white solid, 128 mg, 89% yield, 98% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.43 (d, *J* = 7.7 Hz, 2H), 7.40 – 7.35 (m, 2H), 7.31 – 7.22 (m, 5H), 7.04 (t, *J* = 7.4 Hz, 1H), 3.70 (q, *J* = 7.1 Hz, 1H), 1.57 (d, *J* = 7.1 Hz, 3H), 1.31 (s, 9H).

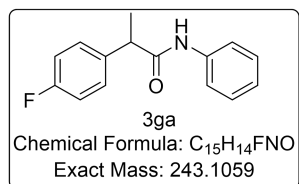
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.80, 150.49, 138.07, 137.91, 128.96, 127.44, 126.09, 124.26, 119.89, 47.65, 34.60, 31.42, 18.64.



**2-(4-methoxyphenyl)-N-phenylpropanamide:** white solid, 119 mg, 92% yield, 98% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.66 (s, 1H), 7.44 (d, *J* = 7.9 Hz, 2H), 7.30 – 7.12 (m, 4H), 7.12 – 6.92 (m, 1H), 6.85 (d, *J* = 8.6 Hz, 2H), 3.76 (s, 3H), 3.66 (q, *J* = 7.1 Hz, 1H), 1.53 (d, *J* = 7.1 Hz, 3H).

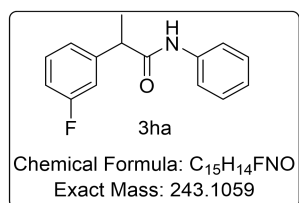
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 173.11, 158.89, 138.06, 133.02, 128.88, 128.74, 124.20, 119.92, 114.40, 55.30, 47.03, 18.70.



**2-(4-fluorophenyl)-N-phenylpropanamide:** white solid, 108 mg, 94% yield, 99% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.02 (s, 1H), 7.44 (d, *J* = 7.8 Hz, 2H), 7.32 – 7.18 (m, 4H), 7.05 (t, *J* = 7.4 Hz, 1H), 6.95 (t, *J* = 8.7 Hz, 2H), 3.67 (q, *J* = 7.1 Hz, 1H), 1.50 (d, *J* = 7.1 Hz, 3H).

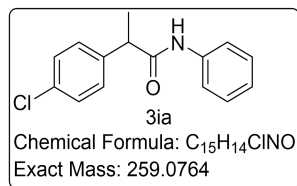
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.85, δ 162.06 (d, *J*<sub>C-F</sub> = 245.8 Hz), 137.91, 136.80 (d, *J*<sub>C-F</sub> = 3.2 Hz), 129.17 (d, *J*<sub>C-F</sub> = 8.0 Hz) 128.92, 124.50, 120.28, 115.71 (d, *J*<sub>C-F</sub> = 21.3 Hz), 46.91, 18.80.



**2-(3-fluorophenyl)-N-phenylpropanamide<sup>4</sup>:** white solid, 106 mg, 85% yield, 96% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.52 (s, 1H), 7.43 (d, *J* = 7.7 Hz, 2H), 7.33 – 7.22 (m, 3H), 7.16 – 7.04 (m, 3H), 6.97 (td, *J* = 8.4, 1.9 Hz, 1H), 3.69 (q, *J* = 7.1 Hz, 1H), 1.55 (d, *J* = 7.1 Hz, 3H).

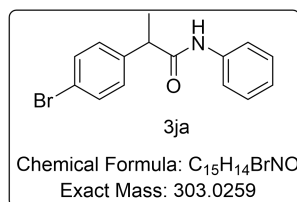
$^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  171.98, 163.16 (d,  $J_{\text{C-F}} = 246.8$  Hz), 143.51 (d,  $J = 7.1$  Hz), 137.81, 130.60 (d,  $J_{\text{C-F}} = 8.3$  Hz), 129.03, 124.57, 123.39 (d,  $J_{\text{C-F}} = 2.9$  Hz), 120.10, 115.01 (d,  $J = 36.6$  Hz), 114.72 (d,  $J_{\text{C-F}} = 21.6$  Hz), 47.73, 18.67.



**2-(4-chlorophenyl)-N-phenylpropanamide:** white solid, 122 mg, 91% yield, 98% branched selectivity.

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.43 (d,  $J = 7.8$  Hz, 2H), 7.35 – 7.21 (m, 7H), 7.08 (t,  $J = 7.4$  Hz, 1H), 3.67 (q,  $J = 7.1$  Hz, 1H), 1.56 (d,  $J = 7.1$  Hz, 3H).

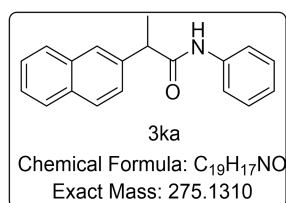
$^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  171.93, 139.53, 137.80, 133.52, 129.34, 129.12, 129.09, 124.59, 119.94, 47.60, 18.87.



**2-(4-bromophenyl)-N-phenylpropanamide:** white solid, 130 mg, 80% yield, 92% branched selectivity.

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.48 (d,  $J = 8.3$  Hz, 2H), 7.43 (d,  $J = 8.0$  Hz, 2H), 7.34 – 7.22 (m, 4H), 7.16 (s, 1H), 7.08 (t,  $J = 7.3$  Hz, 1H), 3.65 (d,  $J = 7.1$  Hz, 1H), 1.56 (d,  $J = 7.1$  Hz, 3H).

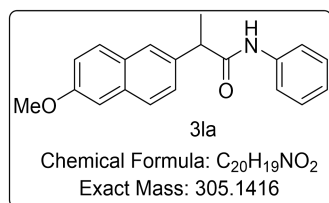
$^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  171.81, 140.05, 137.78, 132.30, 129.48, 129.10, 124.60, 121.61, 119.93, 47.69, 18.84.



**2-(naphthalen-2-yl)-N-phenylpropanamide<sup>5</sup>:** white solid, 115 mg, 83% yield, 99% branched selectivity.

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.91 – 7.77 (m, 4H), 7.53 – 7.38 (m, 5H), 7.28 – 7.19 (m, 3H), 7.08 – 7.00 (m, 1H), 3.87 (q,  $J = 7.1$  Hz, 1H), 1.67 (d,  $J = 7.1$  Hz, 3H).

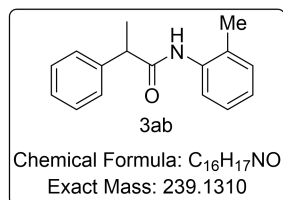
$^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  172.39, 138.48, 137.93, 133.68, 132.83, 129.14, 129.01, 127.90, 127.85, 126.60, 126.25, 125.75, 124.39, 119.86, 48.32, 18.68.



**2-(6-methoxynaphthalen-2-yl)-N-phenylpropanamide<sup>5</sup>**: white solid, 140 mg, 87% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.70 (dd, *J* = 11.5, 9.1 Hz, 3H), 7.41 (d, *J* = 8.5 Hz, 3H), 7.32 (s, 1H), 7.23 (dd, *J* = 9.5, 6.3 Hz, 2H), 7.18 – 7.09 (m, 2H), 7.03 (t, *J* = 7.4 Hz, 1H), 3.90 (s, 3H), 3.83 (q, *J* = 7.1 Hz, 1H), 1.64 (d, *J* = 7.1 Hz, 3H).

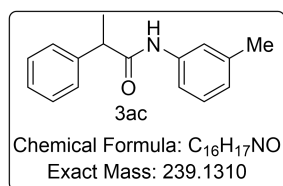
**<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)** δ 172.69, 157.94, 137.98, 136.09, 133.97, 129.36, 129.11, 128.97, 127.91, 126.39, 126.25, 124.32, 119.85, 119.38, 105.78, 55.43, 48.07, 18.67.



**2-phenyl-N-(o-tolyl) propanamide<sup>3</sup>**: white solid, 115 mg, 94% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.76 (d, *J* = 8.0 Hz, 1H), 7.42 – 7.24 (m, 5H), 7.19 – 6.94 (m, 4H), 3.74 (q, *J* = 7.2 Hz, 1H), 1.86 (s, 3H), 1.59 (d, *J* = 7.2 Hz, 3H).

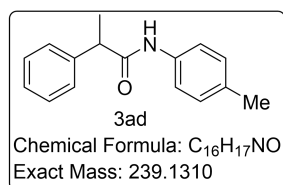
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.49, 141.08, 135.70, 130.30, 129.12, 128.85, 127.80, 127.59, 126.60, 124.94, 122.63, 47.79, 18.08, 17.18.



**2-phenyl-N-(m-tolyl) propanamide<sup>3</sup>**: white solid, 112 mg, 81% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.78 (s, 1H), 7.38 – 7.18 (m, 7H), 7.09 (t, *J* = 7.8 Hz, 1H), 6.84 (d, *J* = 7.5 Hz, 1H), 3.70 (q, *J* = 7.1 Hz, 1H), 2.22 (s, 3H), 1.54 (d, *J* = 7.1 Hz, 3H).

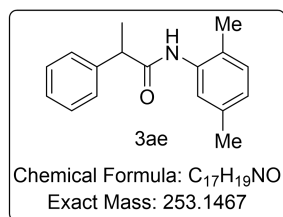
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.80, 141.09, 138.74, 137.94, 128.96, 128.67, 127.63, 127.38, 125.04, 120.68, 117.12, 47.78, 21.40, 18.62.



**2-phenyl-N-(p-tolyl) propanamide<sup>3</sup>**: white solid, 102 mg, 87% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.75 (s, 1H), 7.35 – 7.18 (m, 7H), 7.01 (d, *J* = 8.2 Hz, 2H), 3.68 (q, *J* = 7.2 Hz, 1H), 2.24 (s, 3H), 1.52 (d, *J* = 7.1 Hz, 3H).

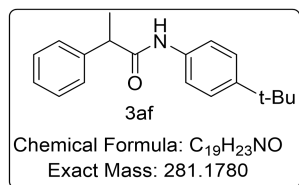
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.74, 141.15, 135.44, 133.83, 129.33, 128.94, 127.66, 127.35, 120.19, 47.70, 20.85, 18.62.



**N-(2,5-dimethylphenyl)-2-phenylpropanamide:** white solid, 105 mg, 80% yield, 98% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.69 (s, 1H), 7.45 – 7.24 (m, 5H), 6.94 (d, *J* = 7.7 Hz, 2H), 6.81 (d, *J* = 7.6 Hz, 1H), 3.76 (q, *J* = 7.2 Hz, 1H), 2.27 (s, 3H), 1.83 (s, 3H), 1.62 (d, *J* = 7.2 Hz, 3H).

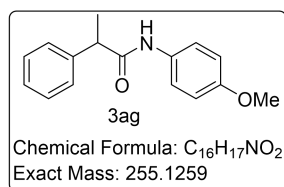
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.42, 141.11, 136.51, 135.54, 130.14, 129.25, 127.90, 127.73, 125.61, 125.22, 122.86, 48.06, 21.17, 18.12, 16.76.



**N-(4-(tert-butyl) phenyl)-2-phenylpropanamide:** white solid, 126 mg, 86% yield, 97% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.46 (s, 1H), 7.39 – 7.30 (m, 6H), 7.29 – 7.21 (m, 3H), 3.71 (q, *J* = 7.1 Hz, 1H), 1.56 (d, *J* = 7.1 Hz, 3H), 1.26 (s, 9H).

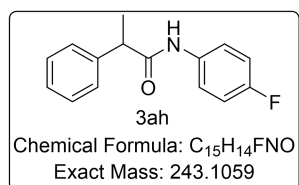
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.51, 147.23, 141.18, 135.43, 129.08, 127.73, 127.48, 125.74, 119.69, 47.93, 34.38, 31.41, 18.67.



**N-(4-methoxyphenyl)-2-phenylpropanamide:** white solid, 117 mg, 91% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 – 7.14 (m, 8H), 6.78 (d, *J* = 8.8 Hz, 2H), 3.74 (s, 3H), 3.69 (q, *J* = 7.1 Hz, 1H), 1.57 (d, *J* = 7.1 Hz, 3H).

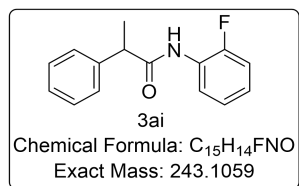
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.40, 156.45, 141.22, 131.11, 129.14, 127.78, 127.54, 121.81, 114.10, 55.54, 47.90, 18.72.



**N-(4-fluorophenyl)-2-phenylpropanamide<sup>4</sup>:** white solid, 104 mg, 87% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.89 (s, 1H), 7.43 – 7.19 (m, 7H), 6.88 (t, *J* = 8.5 Hz, 2H), 3.69 (q, *J* = 6.8 Hz, 1H), 1.54 (d, *J* = 6.9 Hz, 3H).

**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.95, 159.37 (d, *J*<sub>C-F</sub> = 243.5 Hz), 140.94, 133.97 (d, *J*<sub>C-F</sub> = 2.7 Hz), 129.03, 127.62, 127.52, 122.06 (d, *J*<sub>C-F</sub> = 7.8 Hz), 115.45 (d, *J*<sub>C-F</sub> = 22.4 Hz), 47.65, 18.59.

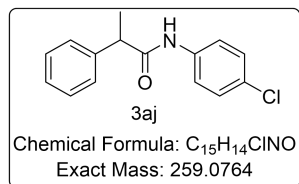




**N-(2-fluorophenyl)-2-phenylpropanamide**<sup>6</sup>: white solid, 128mg, 95% yield, 99% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.28 (t, *J* = 8.0 Hz, 1H), 7.49 (d, *J* = 31.2 Hz, 1H), 7.40 – 7.23 (m, 5H), 7.10 – 6.92 (m, 3H), 3.76 (q, *J* = 7.1 Hz, 1H), 1.59 (d, *J* = 7.2 Hz, 3H).

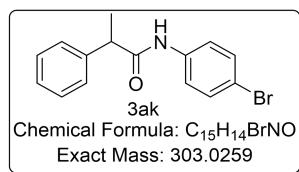
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.48, 152.47 (d, *J*<sub>C-F</sub> = 243.2 Hz), 140.64, 129.16, 127.67, 126.39 (d, *J*<sub>C-F</sub> = 10.0 Hz), 124.49 (d, *J*<sub>C-F</sub> = 3.7 Hz), 124.32 (d, *J*<sub>C-F</sub> = 7.7 Hz), 121.73, 114.71 (d, *J*<sub>C-F</sub> = 19.2 Hz), 48.13, 18.47.



**N-(4-chlorophenyl)-2-phenylpropanamide**: white solid, 127 mg, 95% yield, 99% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.79 (s, 1H), 7.36 – 7.27 (m, 7H), 7.17 – 7.13 (m, 2H), 3.68 (q, *J* = 7.1 Hz, 1H), 1.53 (d, *J* = 7.1 Hz, 3H).

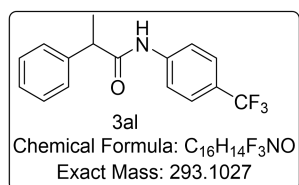
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 173.00, 140.76, 136.49, 129.32, 129.10, 128.87, 127.62, 121.43, 47.79, 18.58.



**N-(4-bromophenyl)-2-phenylpropanamide**: white solid, 92 mg, 86% yield, 99%, branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.55 (s, 1H), 7.40 – 7.25 (m, 9H), 3.68 (q, *J* = 7.1 Hz, 1H), 1.55 (d, *J* = 7.1 Hz, 3H).

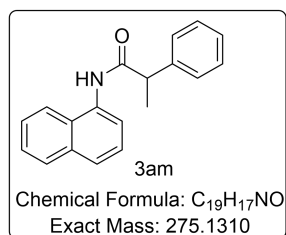
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.77, 140.75, 137.01, 131.87, 129.20, 127.68, 121.62, 116.94, 47.97, 18.63.



**2-phenyl-N-(4-(trifluoromethyl)phenyl) propanamide**: white solid, 138 mg, 95% yield, 99% branched selectivity.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.85 (s, 1H), 7.49 (dd, *J* = 30.5, 8.3 Hz, 4H), 7.40 – 7.22 (m, 5H), 3.73 (q, *J* = 6.8 Hz, 1H), 1.57 (d, *J* = 6.7 Hz, 3H).

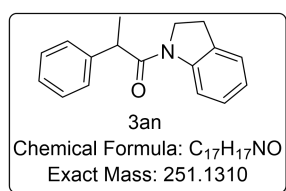
<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 173.21, 141.01, 140.58, 129.24, 127.79, 127.65, 126.12 (q, *J*<sub>C-F</sub> = 3.7 Hz), 125.65 (d, *J*<sub>C-F</sub> = 43.6 Hz) 122.78, 119.67, 48.05, 18.59.



**N-(naphthalen-1-yl)-2-phenylpropanamid** : white solid, 107 mg, 95% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.98 (s, 1H), 7.66 (dd, *J* = 17.3, 7.7 Hz, 2H), 7.50 (d, *J* = 8.2 Hz, 1H), 7.37 – 7.18 (m, 9H), 3.75 (q, *J* = 7.0 Hz, 1H), 1.50 (d, *J* = 7.1 Hz, 3H).

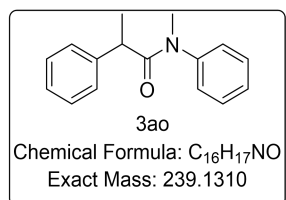
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 173.33, 141.24, 133.92, 132.31, 129.00, 128.42, 127.77, 127.43, 125.99, 125.74, 125.67, 125.43, 121.04, 120.78, 47.44, 18.30.



**1-(indolin-1-yl)-2-phenylpropan-1-one<sup>7</sup>**: white solid, 107 mg, 80% yield, 97% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.34 (d, *J* = 8.1 Hz, 1H), 7.35 – 7.27 (m, 4H), 7.25 – 7.14 (m, 2H), 7.09 (d, *J* = 7.3 Hz, 1H), 6.97 (td, *J* = 7.4, 0.8 Hz, 1H), 4.05 (td, *J* = 10.4, 6.5 Hz, 1H), 3.84 (q, *J* = 6.8 Hz, 1H), 3.73 (td, *J* = 10.4, 6.6 Hz, 1H), 3.18 – 3.00 (m, 1H), 3.00 – 2.88 (m, 1H), 1.52 (d, *J* = 6.9 Hz, 3H).

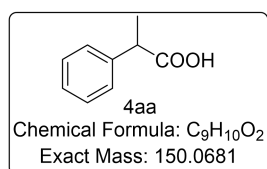
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 171.99, 143.36, 141.13, 131.16, 129.00, 127.55, 127.49, 127.03, 124.50, 123.68, 117.22, 47.68, 46.26, 28.01, 20.56.



**N-methyl-N,2-diphenylpropanamide<sup>4</sup>**: colorless oily liquid, 98 mg, 77% yield, 91% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.41 – 7.30 (m, 3H), 7.23 – 7.14 (m, 3H), 7.07 – 6.95 (m, 4H), 3.64 (q, *J* = 6.9 Hz, 1H), 3.23 (s, 3H), 1.39 (d, *J* = 6.9 Hz, 3H).

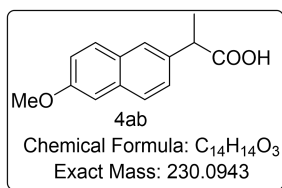
**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 174.05, 143.82, 141.97, 129.57, 128.38, 127.85, 127.56, 126.63, 43.12, 37.74, 20.35.



**2-phenylpropanoic acid**: Colorless liquid, 64 mg, 81% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 11.72 (s, 1H), 7.37 – 7.24 (m, 5H), 3.73 (q, *J* = 7.2 Hz, 1H), 1.51 (d, *J* = 7.2 Hz, 3H).

**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 181.09, 139.89, 128.81, 127.73, 127.52, 45.52, 18.21.



**2-(6-methoxynaphthalen-2-yl)propanoic acid:** white solid, 97 mg, 84% yield, 99% branched selectivity.

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.74 (d, J = 9.2 Hz, 3H), 7.45 (dd, J = 8.5, 1.7 Hz, 1H), 7.21 – 7.12 (m, 2H), 4.23 – 3.64 (m, 4H), 1.63 (d, J = 7.2 Hz, 3H).

**<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 180.99, 157.84, 135.00, 133.96, 129.44, 129.03, 127.37, 126.33, 126.28, 119.17, 105.73, 55.43, 45.41, 18.24.

## 8. Catalyst characterization

Experimental PXRD profiles

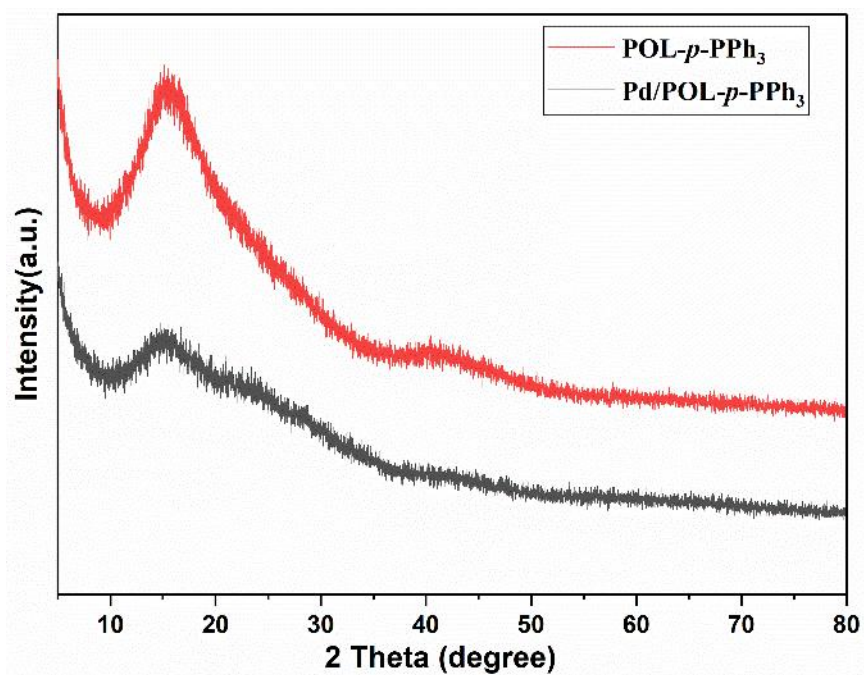
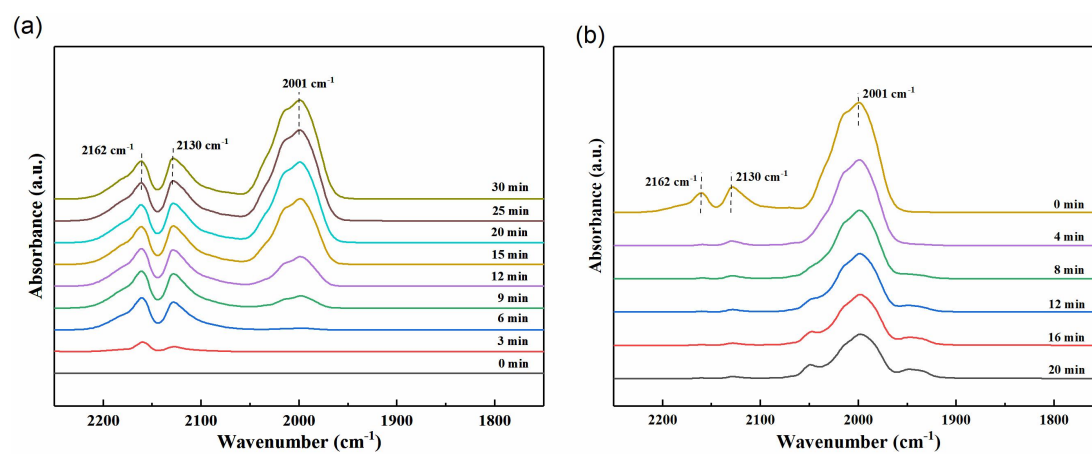


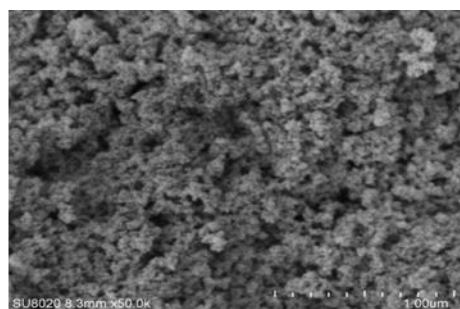
Fig. S1 PXRD profiles of POL-*p*-3vPPh<sub>3</sub>, Pd/POL-*p*-3vPPh<sub>3</sub>

## In-stiu DRIFTS spectra

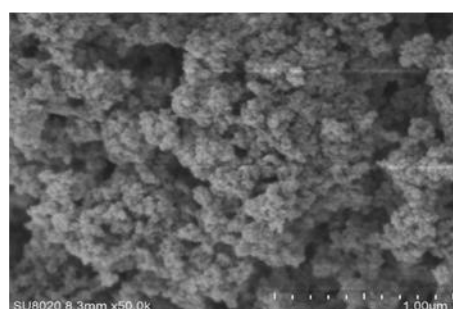


**Fig. S2** In stiu DRIFTS spectra using CO probe. (a) after exposure to CO and (b) Ar purging at 303 K.

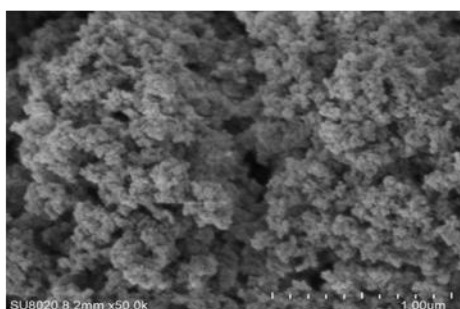
### Scanning electron micrographs



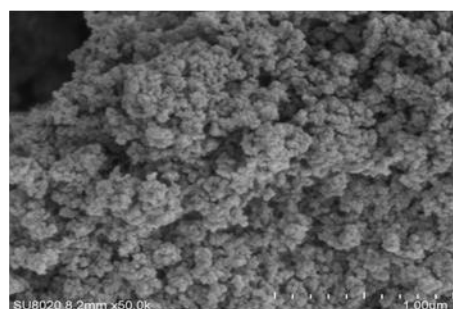
POL-*p*-3vPPh<sub>3</sub>



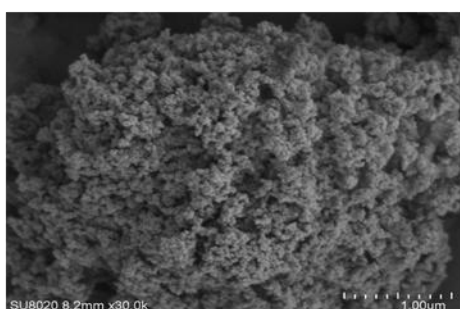
Pd/POL-*p*-3vPPh<sub>3</sub>



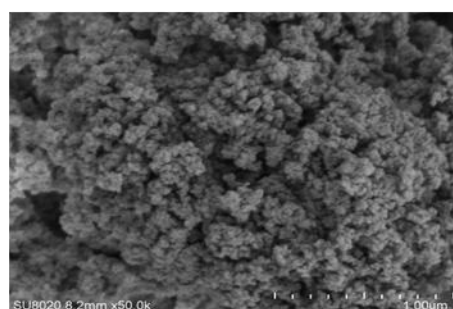
POL-*m*-3vPPh<sub>3</sub>



Pd/POL-*m*-3vPPh<sub>3</sub>-reused-1<sup>st</sup>



Pd/POL-*m*-3vPPh<sub>3</sub>-reused-2<sup>nd</sup>



Pd/POL-*m*-3vPPh<sub>3</sub>-reused-3<sup>rd</sup>

**Fig. S3** SEM images of the POL-*p*-3vPPh<sub>3</sub>, Pd/POL-*p*-3vPPh<sub>3</sub>, POL-*m*-3vPPh<sub>3</sub>, Pd/POL-*m*-3vPPh<sub>3</sub>-reused-1<sup>st</sup>, Pd/POL-*m*-3vPPh<sub>3</sub>-reused-2<sup>nd</sup> and Pd/POL-*m*-3vPPh<sub>3</sub>-reused-3<sup>rd</sup>.

Transmission electron micrographs

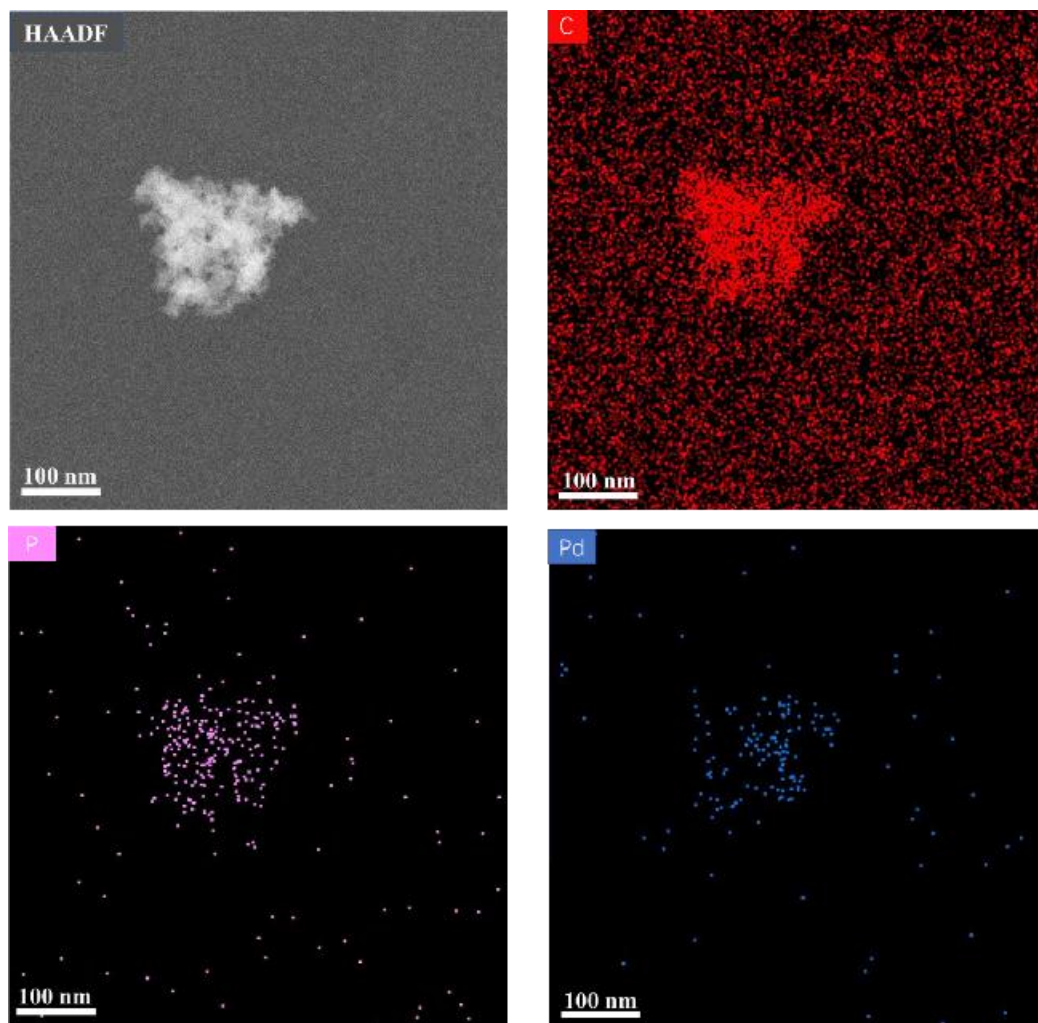
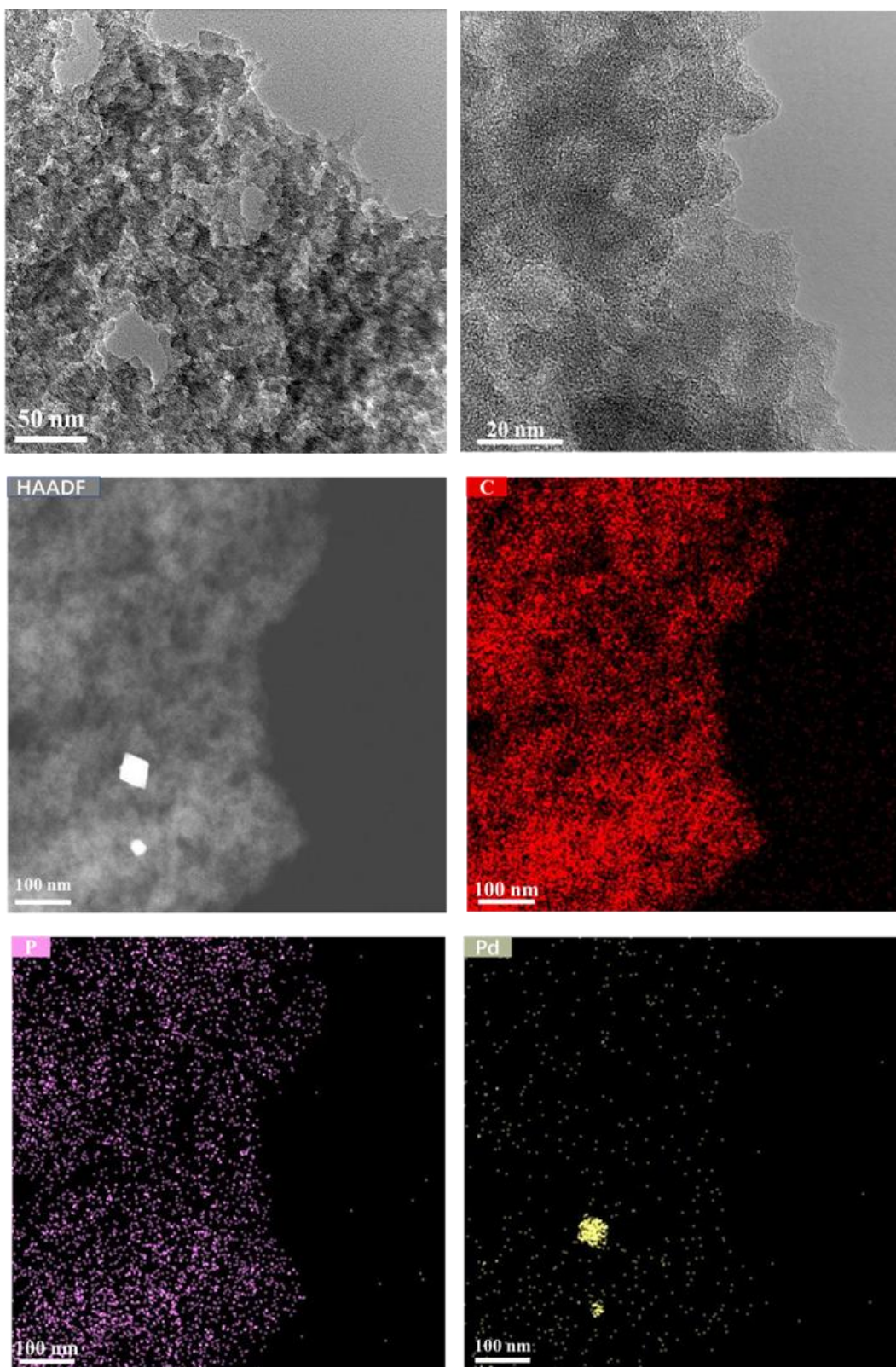


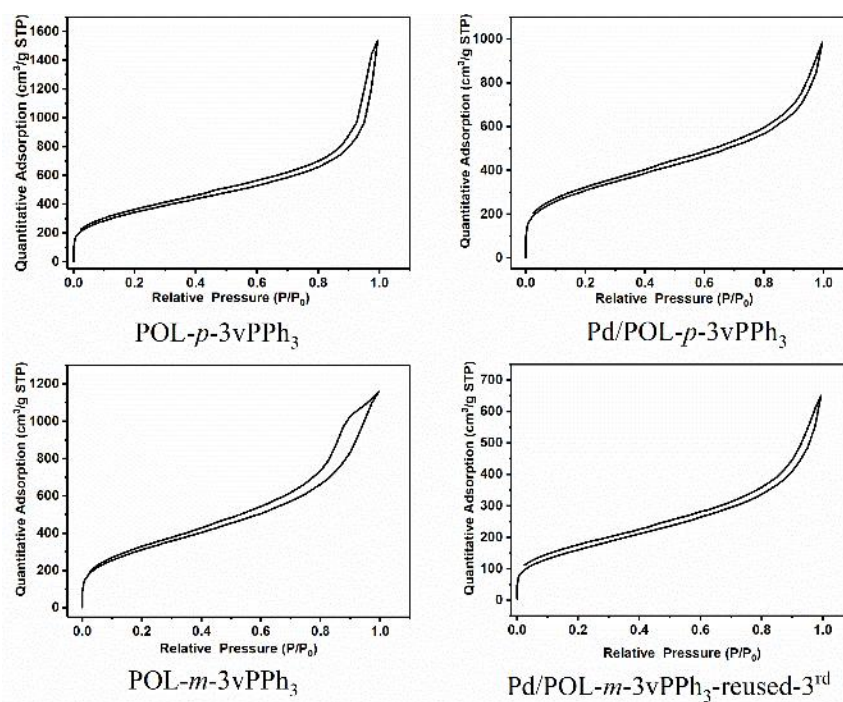
Fig. S4 TEM images of the Pd/POL-*m*-3v-PPh<sub>3</sub>



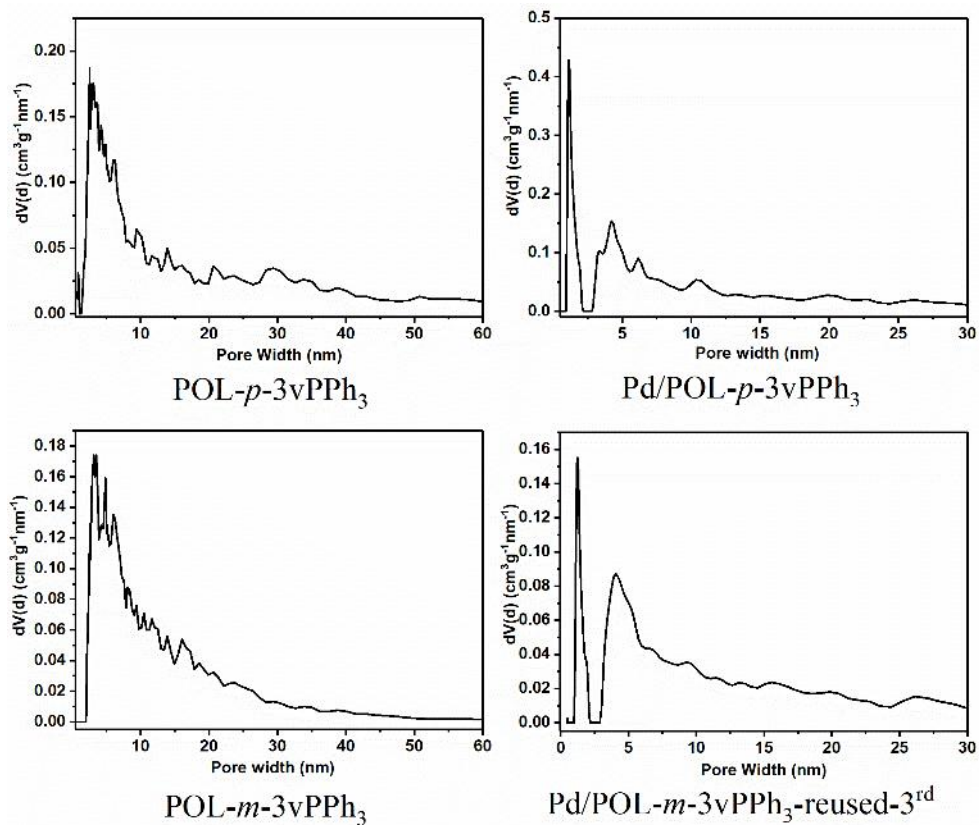
**Fig. S5** TEM images of the Pd/POL-*m*-3v-PPh<sub>3</sub>-reused-3<sup>rd</sup>



## N<sub>2</sub> adsorption-desorption analysis



**Fig. S6** N<sub>2</sub> adsorption-desorption isotherm of POL-*p*-3vPPH<sub>3</sub>, Pd/POL-*p*-3vPPH<sub>3</sub>, POL-*m*-3vPPH<sub>3</sub>, and Pd/POL-*m*-3vPPH<sub>3</sub>-3<sup>rd</sup>.



**Fig. S7** Pore size distribution of POL-*p*-3vPPh<sub>3</sub>, Pd/POL-*p*-3vPPh<sub>3</sub>, POL-*m*-3vPPh<sub>3</sub> and Pd/POL-*m*-3vPPh<sub>3</sub>-reused-3<sup>rd</sup>.

**Table S3.** The physical properties of the POPs catalysts.

Entry	Catalyst	SA (m <sup>2</sup> /g) <sup>a</sup>	APW (nm) <sup>a</sup>	PV (cm <sup>3</sup> /g) <sup>a</sup>
1	POL- <i>p</i> -3vPPh <sub>3</sub>	1235	2.58	2.38
2	Pd/POL- <i>p</i> -3vPPh <sub>3</sub>	1100	1.14	1.52
3	POL- <i>m</i> -3vPPh <sub>3</sub>	1119	3.18	1.80
4	Pd/POL- <i>m</i> -3vPPh <sub>3</sub>	1022	1.24	1.62
5	Pd/POL- <i>m</i> -3vPPh <sub>3</sub> -reused-3 <sup>rd</sup>	577	1.25	1.01

<sup>a</sup> Determined by an IQ<sub>2</sub> automated gas sorption analyzer. SA: BET surface area; APW: average pore width; PV: pore volume.

**Table S4.** Comparison of the reaction performance of Pd/POL-*m*-3vPPh<sub>3</sub> reported in this work with other heterogeneous catalysts.

Alkene/Amine	Cat.	CO(MPa)	T (°C)	Yield (%)	Amide (b/l)	Ref.
Styrene/Aniline (1:0.5)	Pd/POL- <i>m</i> -3vPPh <sub>3</sub>	4	120	97	98:2	This Work
Styrene/Aniline (4:1)	Pd-610	4	130	89	71:29	6
Phenylacetylene/ <i>p</i> -toluidine (1.5:1)	Pd/DVB-0.2-PAM-Naph	4	120	69	-	8
Styrene/Aniline (0.5:0.6)	Pd@POC	4	110	97	77:1	3

We summarized the precedent heterogeneous catalysts for the hydroaminocarbonylation of aromatic alkenes with branched selectivity in Table S4, suggesting that Pd/POL-*m*-3vPPh<sub>3</sub> catalyst exhibited superior activity and branched selectivity.

TGA analysis

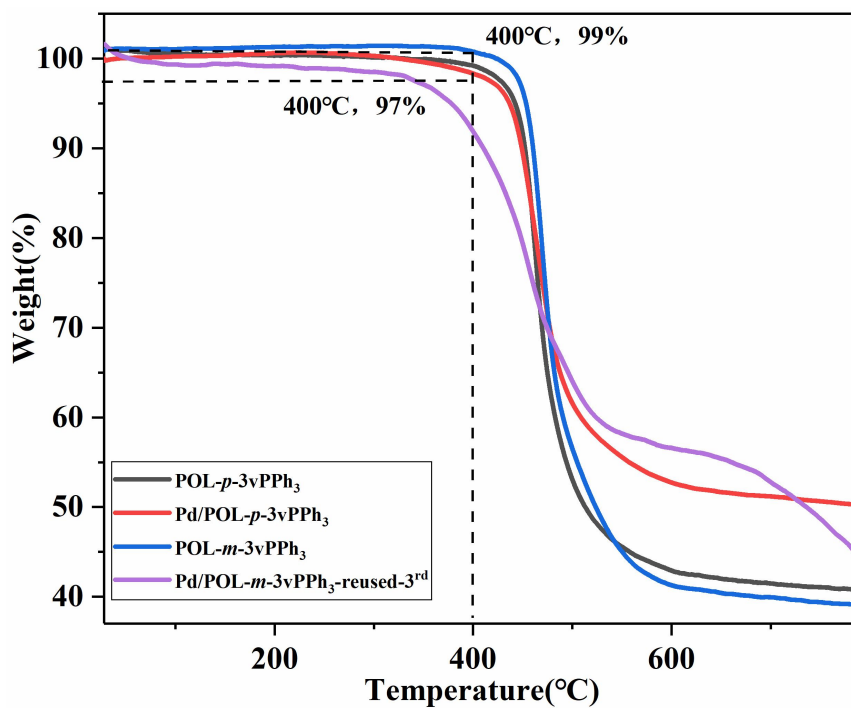
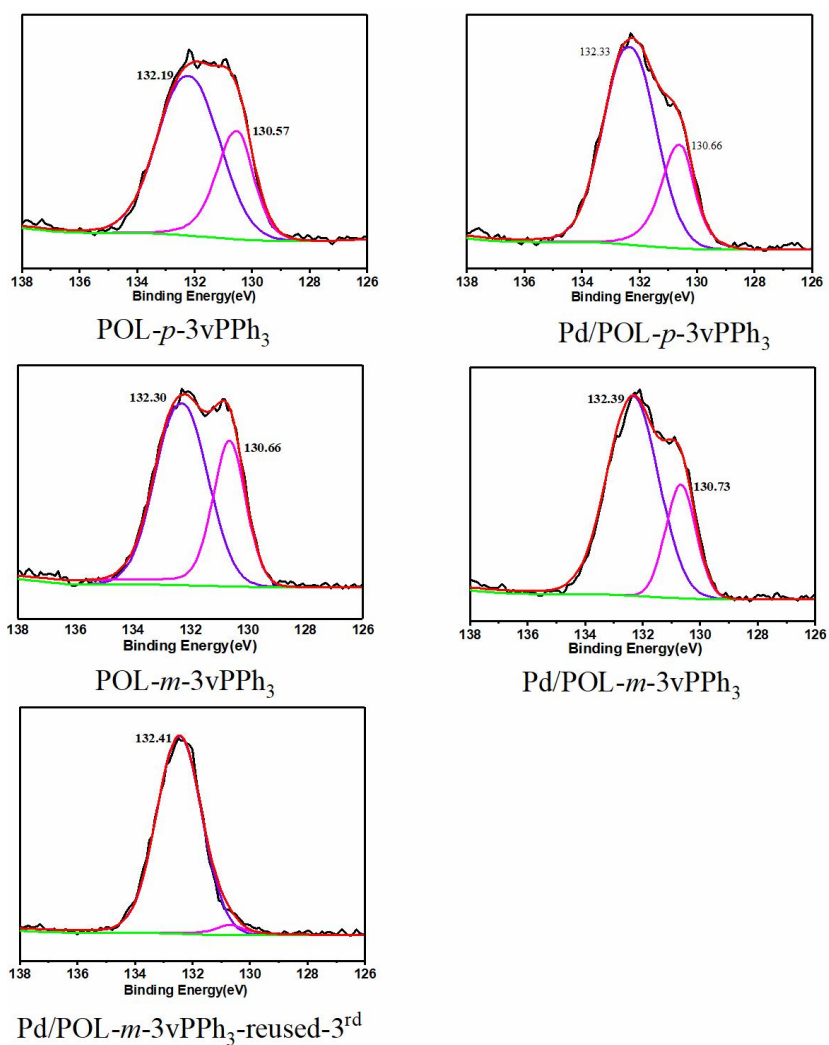
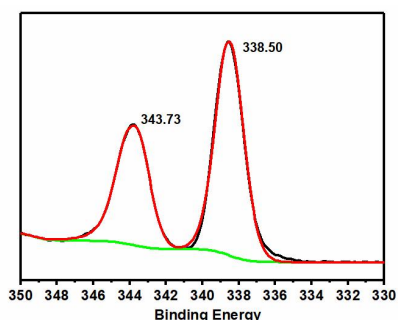


Fig. S8 TG curve of the POL-*p*-3vPPh<sub>3</sub>, Pd/POL-*p*-3vPPh<sub>3</sub>, POL-*m*-3vPPh<sub>3</sub> and Pd/POL-*m*-3vPPh<sub>3</sub>-reused-3<sup>rd</sup>.

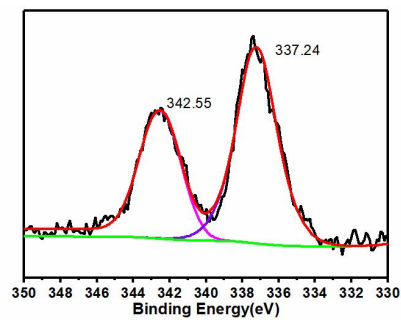
## X-ray photoelectron spectroscopy analysis



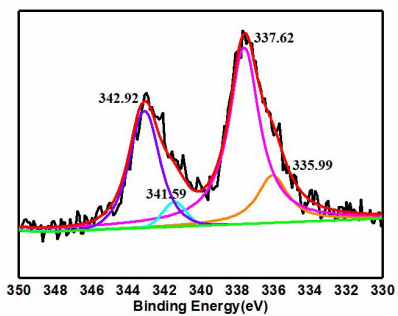
**Fig. S9** P 2p XPS analysis of POL-*p*-3vPPh<sub>3</sub>, Pd/POL-*p*-3vPPh<sub>3</sub>, POL-*m*-3vPPh<sub>3</sub>, Pd/POL-*m*-3vPPh<sub>3</sub>, Pd/POL-*m*-3vPPh<sub>3</sub>-reused-3<sup>rd</sup>.



$\text{Pd}(\text{CH}_3\text{CN})_2\text{Cl}_2$



$\text{Pd}/\text{POL-}p\text{-3vPPh}_3$



$\text{Pd}/\text{POL-}m\text{-3vPPh}_3\text{-reused-3}^{\text{rd}}$

**Fig. S10** Pd 3d XPS analysis of  $\text{Pd}(\text{CH}_3\text{CN})_2\text{Cl}_2$ ,  $\text{Pd}/\text{POL-}p\text{-3vPPh}_3$ ,  $\text{Pd}/\text{POL-}m\text{-3vPPh}_3\text{-reused-3}^{\text{rd}}$ .

## 9. Reaction mechanism

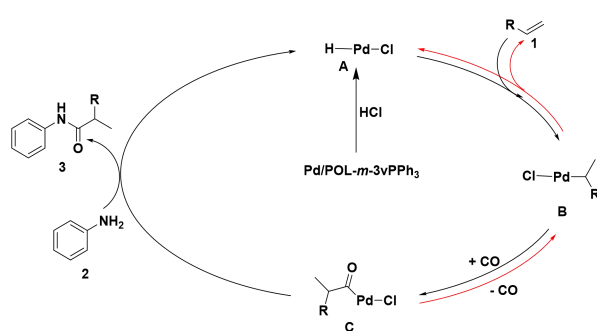
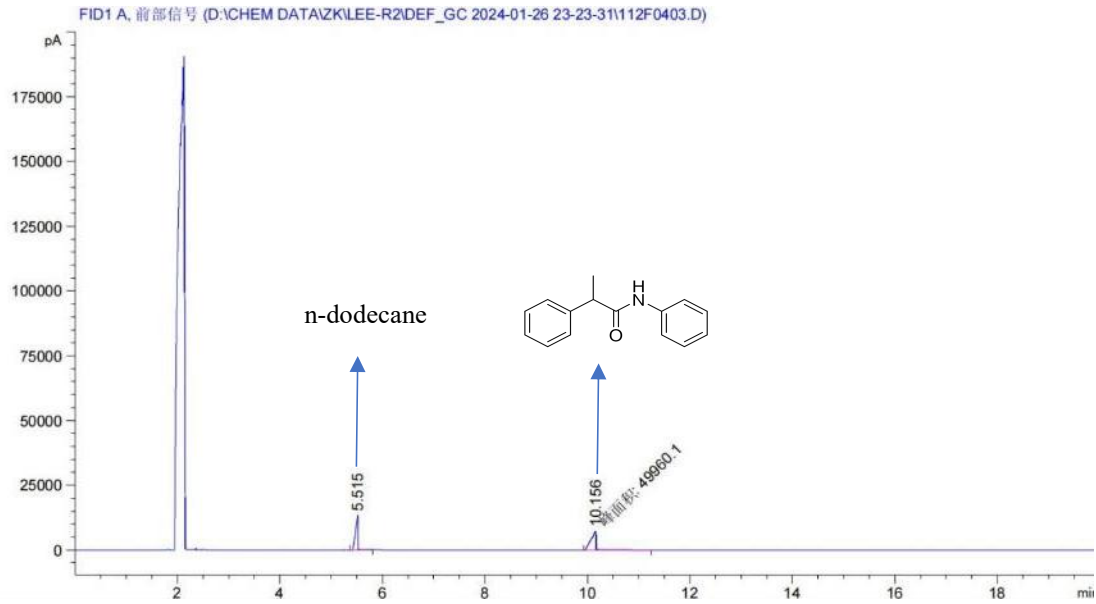


Fig. S11 Proposed reaction mechanism

## 10. Copies of quantitative GC and NMR spectra

### Quantitative GC spectra of styrene hydroaminocarbonylation

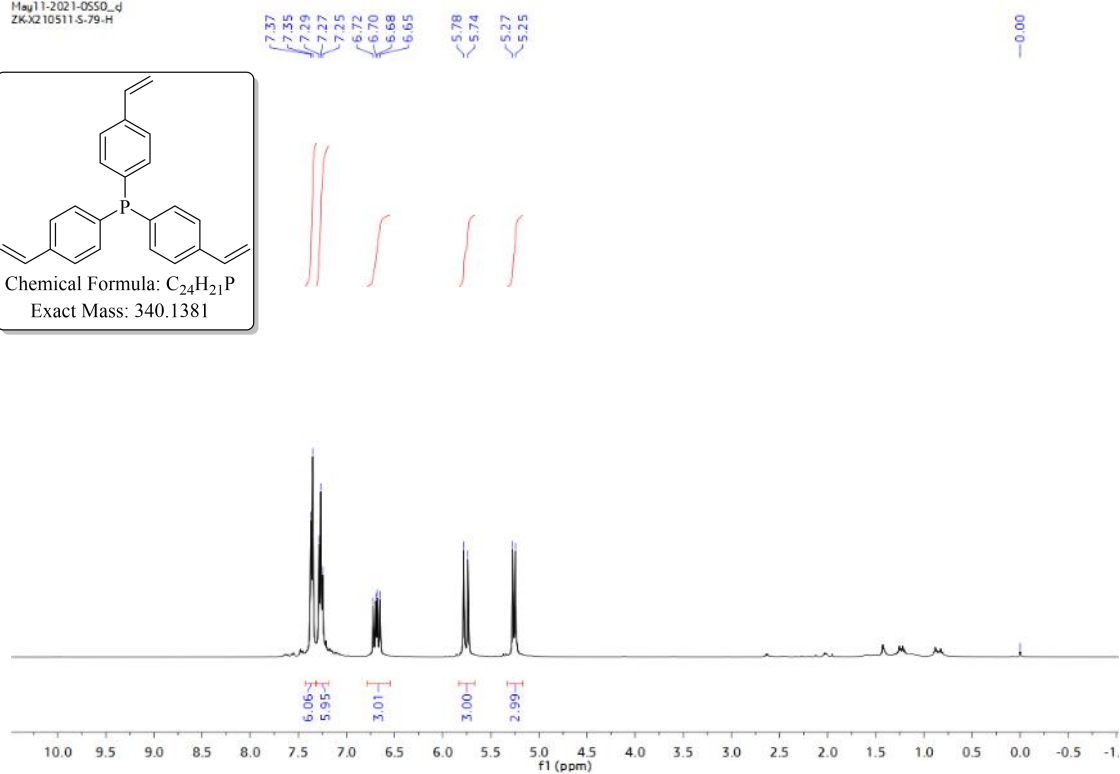
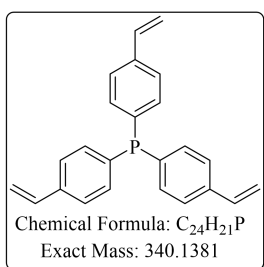


Peak No.	Ret. time	Area	Area%
1	5.515	45436	47.6289
2	10.156	49960	52.3711
Total		95396	100

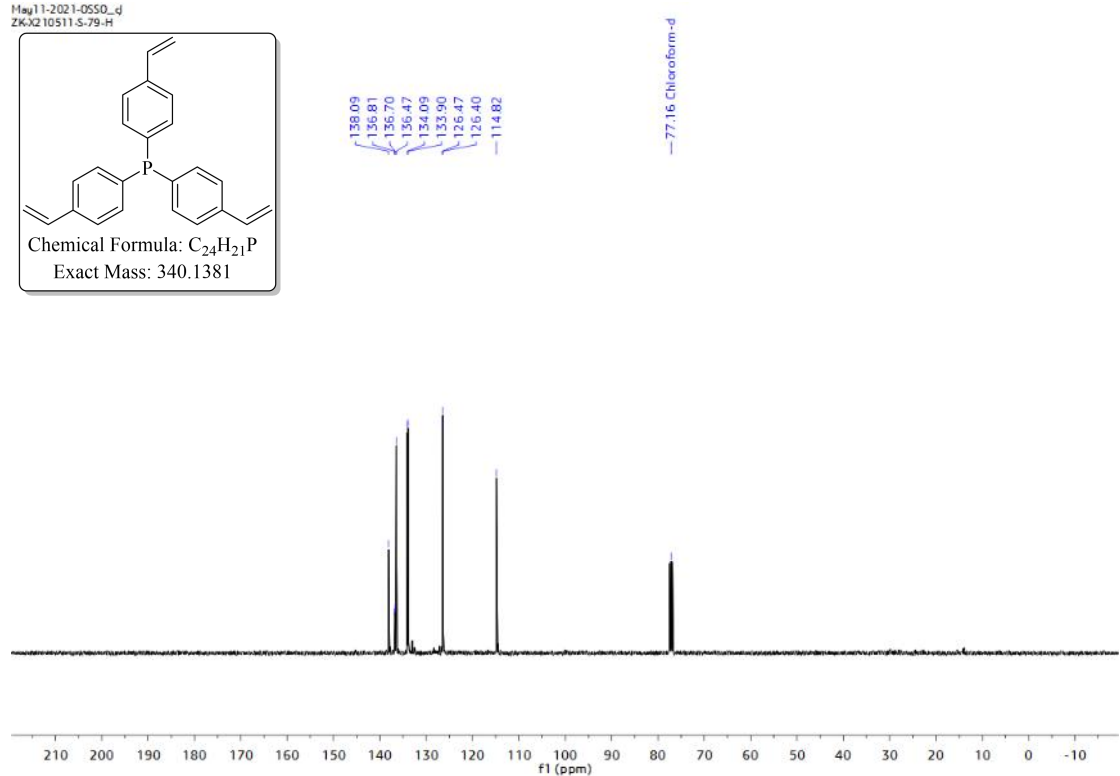
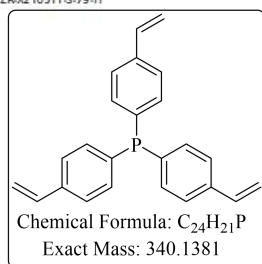


## NMR spectra of the ligands

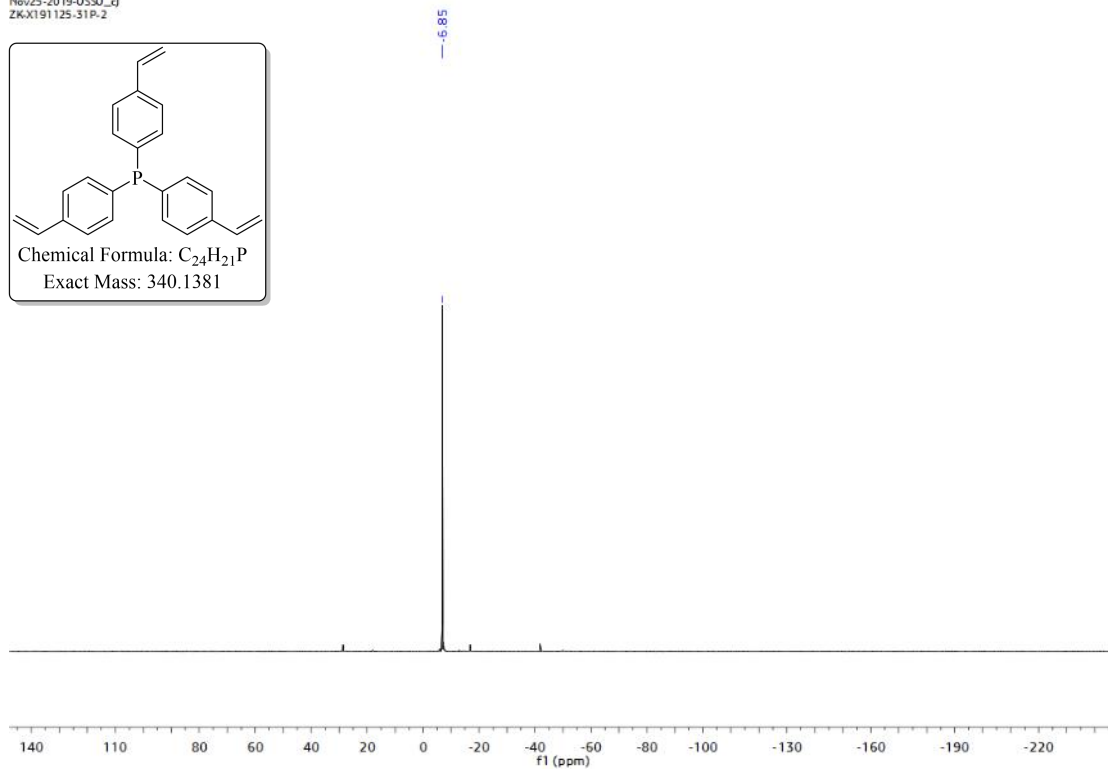
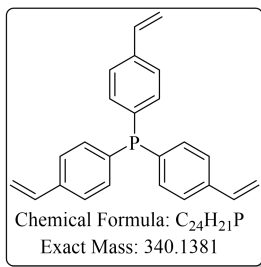
May11-2021-0550\_d  
ZKX210511 S-79-H



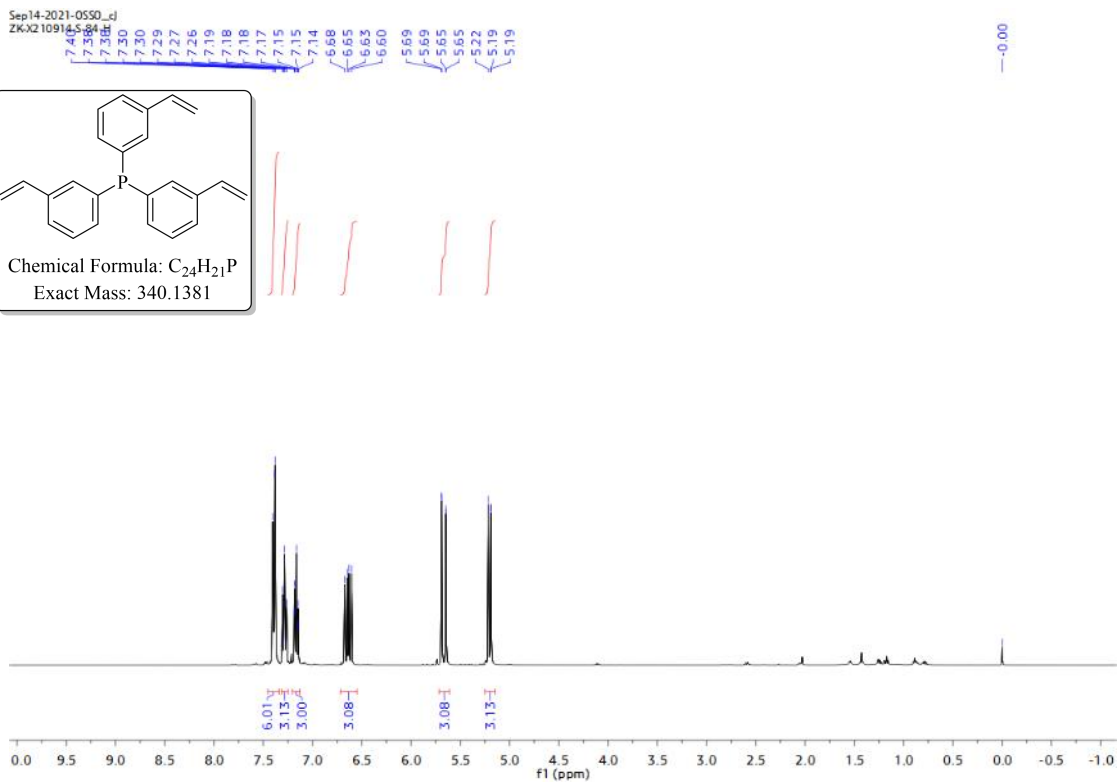
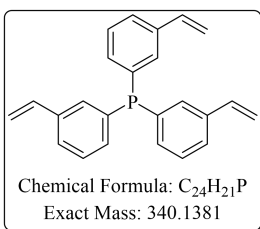
May11-2021-0550\_d  
ZKX210511 S-79-H



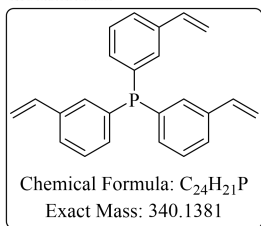
Nov25-2019-0550\_d  
ZKX191125-31P-2



Sep14-2021-0550\_d  
ZKX210914-5-84

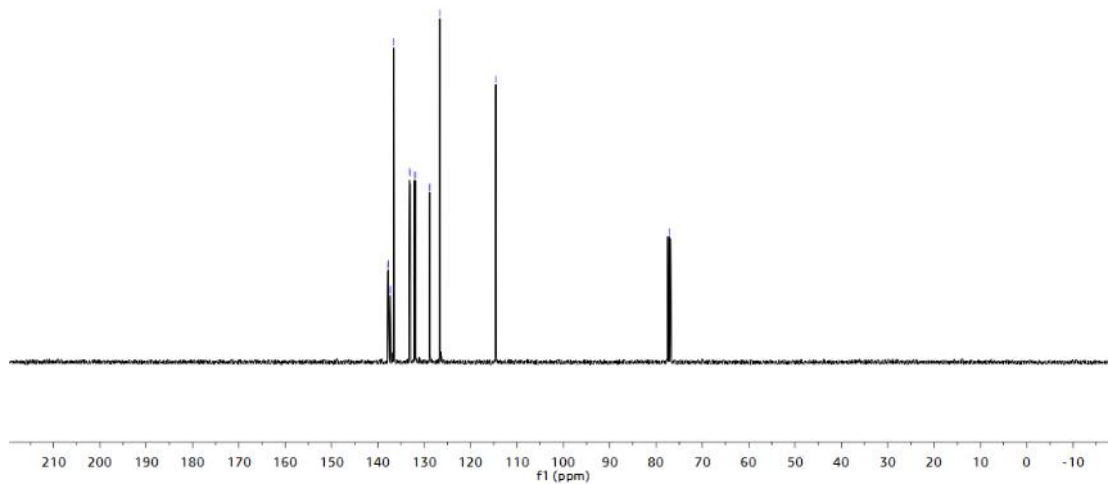


Sep14-2021-0550\_dj  
ZKX210914 S-84-C

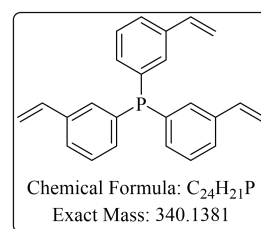


137.85  
137.77  
137.70  
137.28  
136.59  
133.21  
133.04  
132.13  
131.90  
128.88  
126.63  
114.55

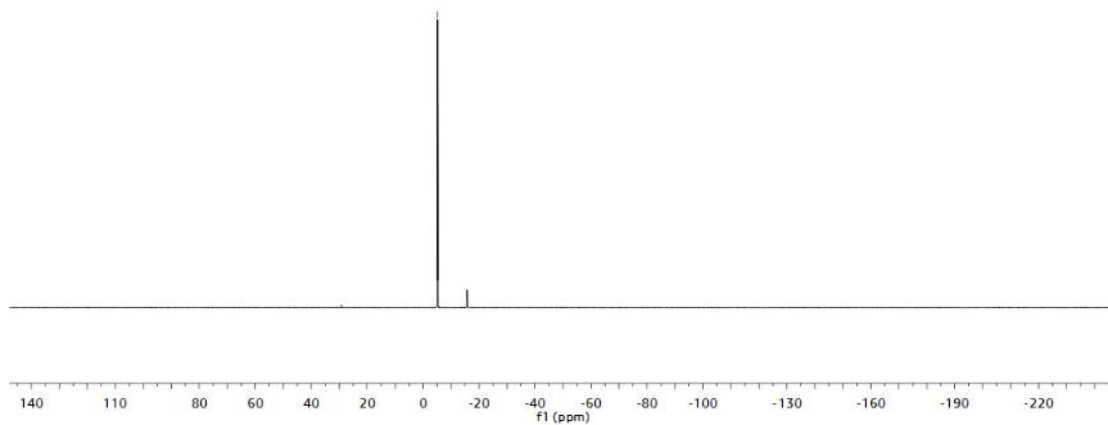
77.16 Chloroform-d



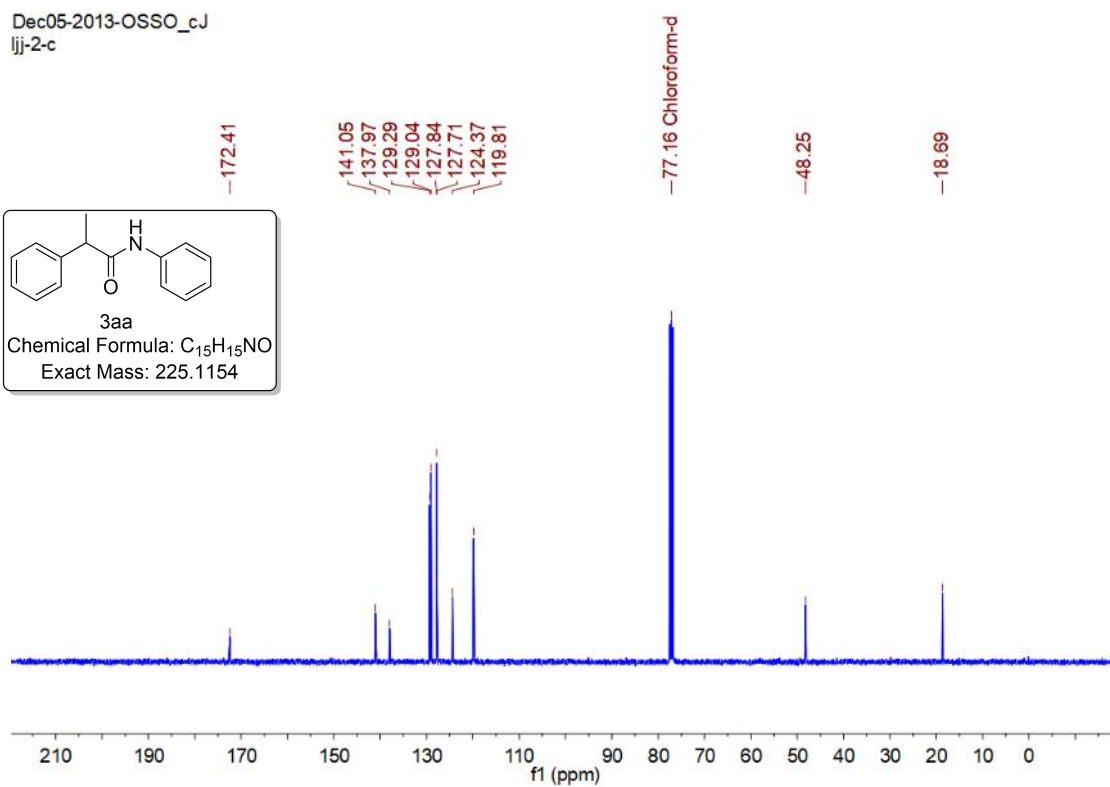
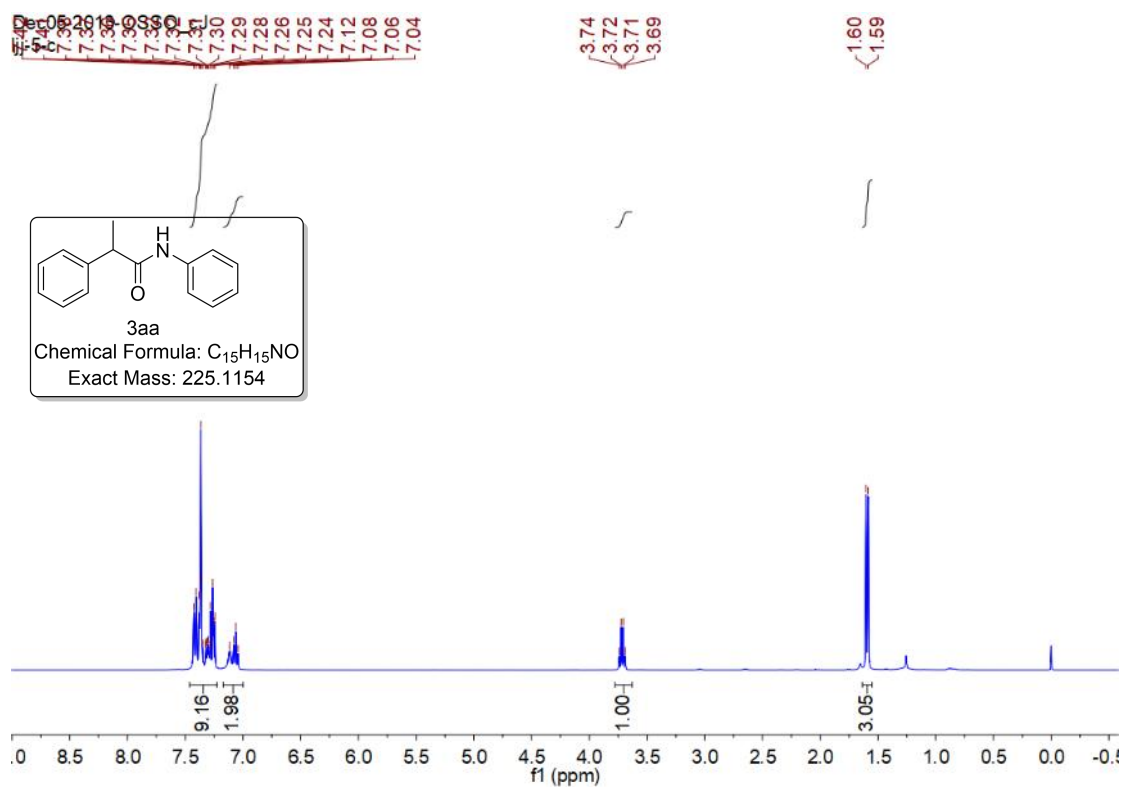
Sep14-2021-0550\_dj  
ZKX210914 S-84-P

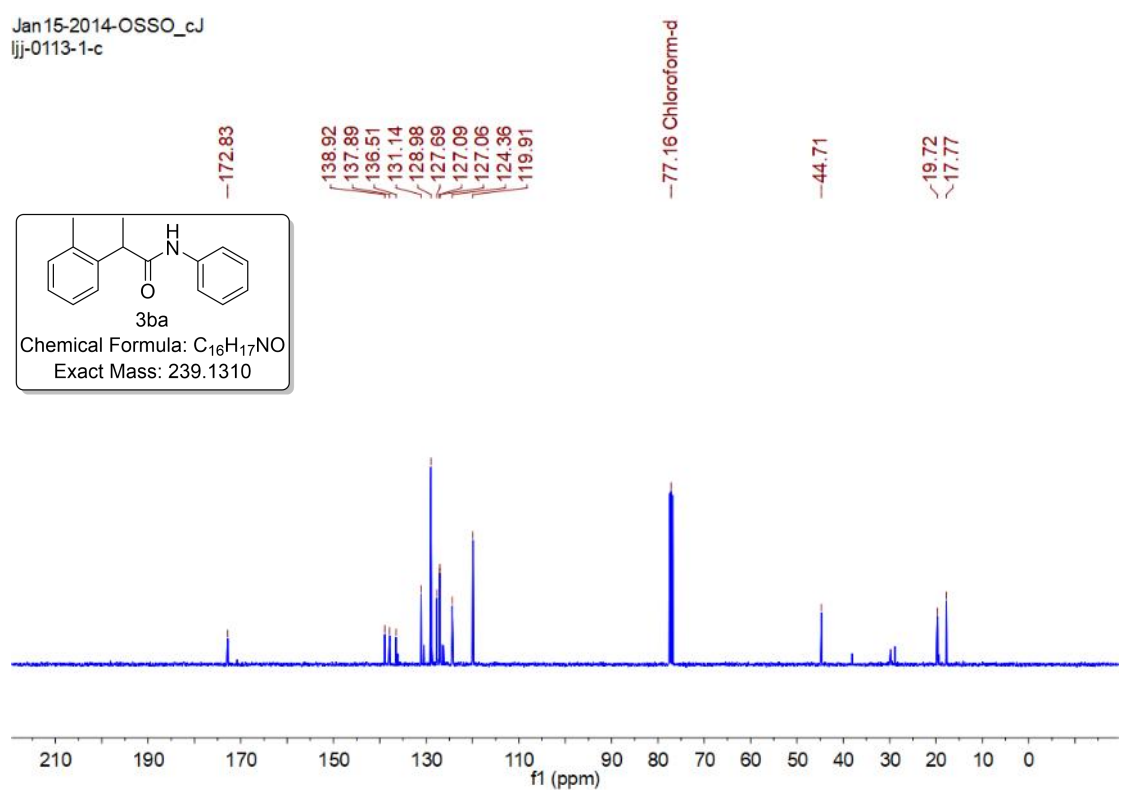
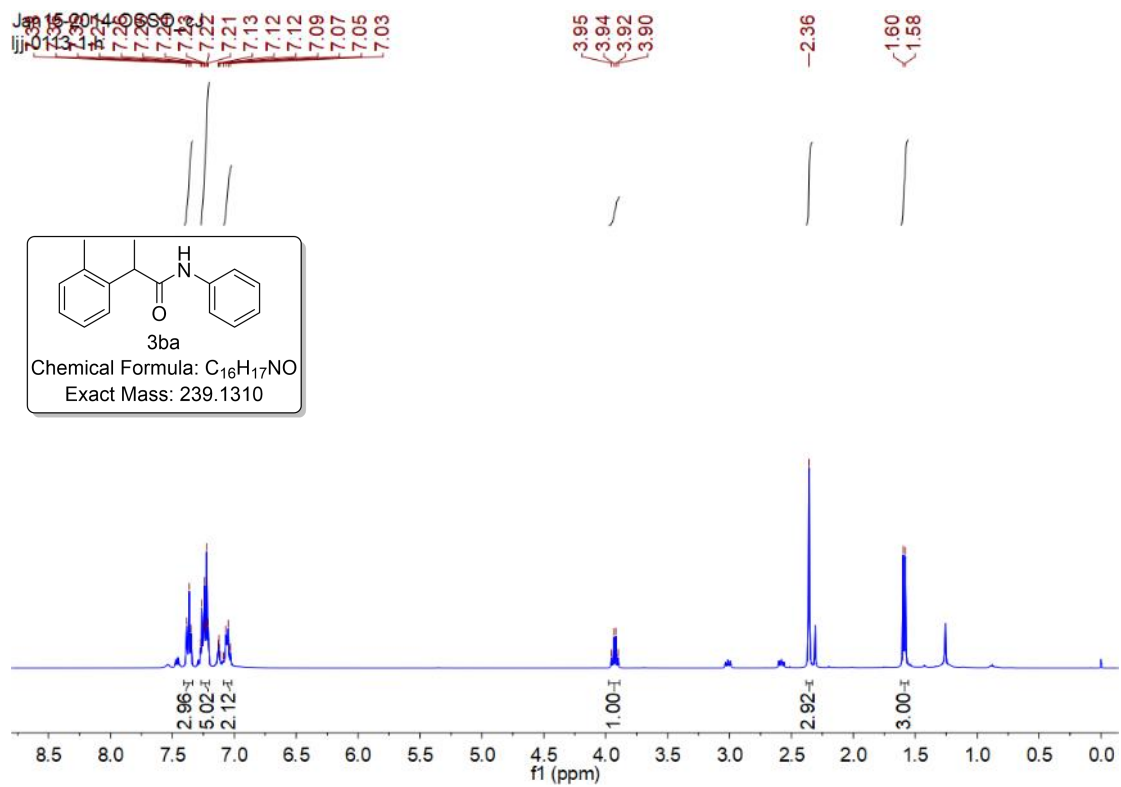


-5.15



### NMR spectra of the products





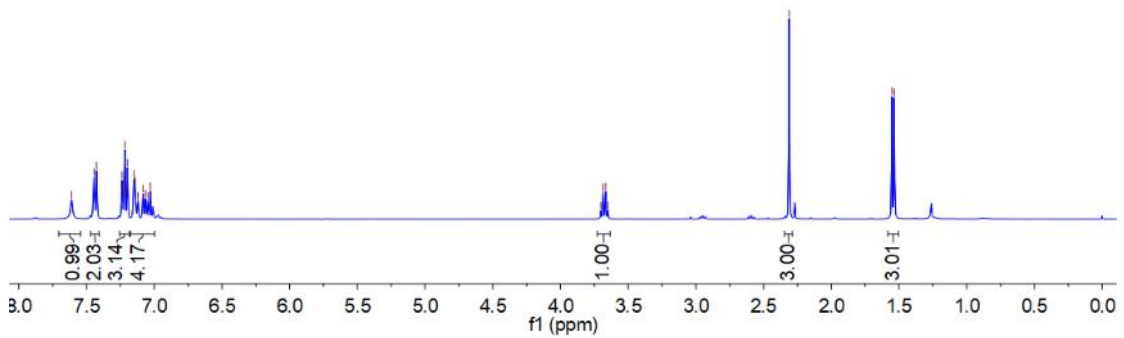
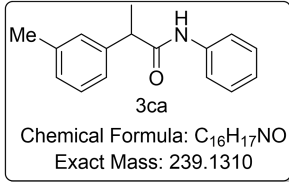
Apr29-2014-OSSO\_cj  
ljj-2-20240428-c



3.70  
3.68  
3.67  
3.65

-2.31

1.55  
1.53



Apr29-2014-OSSO\_cj  
ljj-2-20240428-c

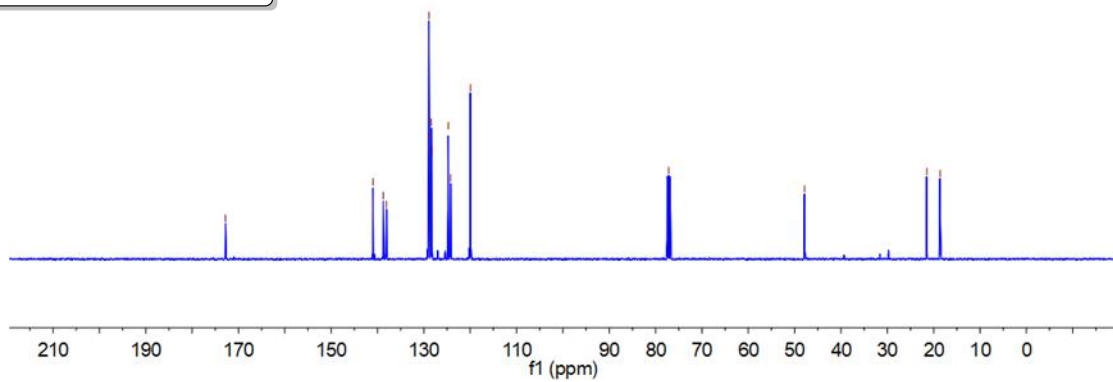
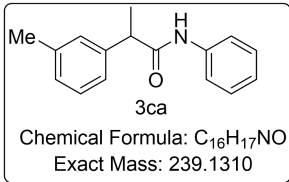
-172.80

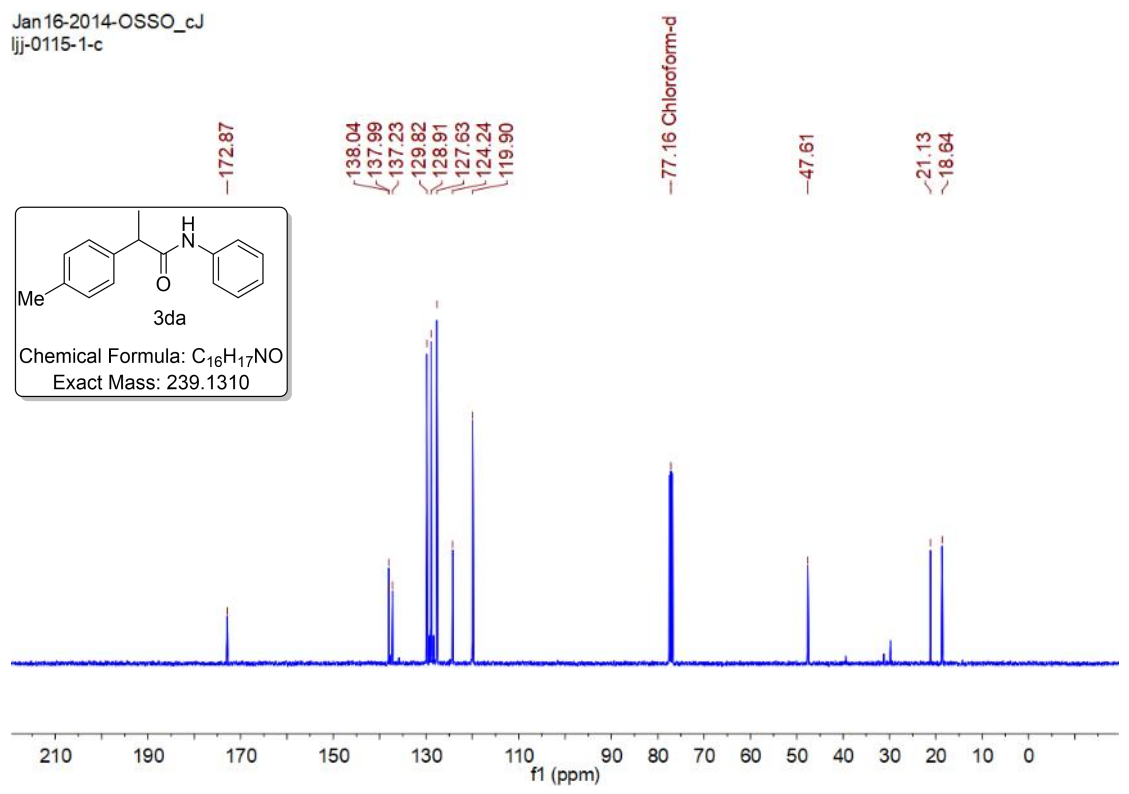
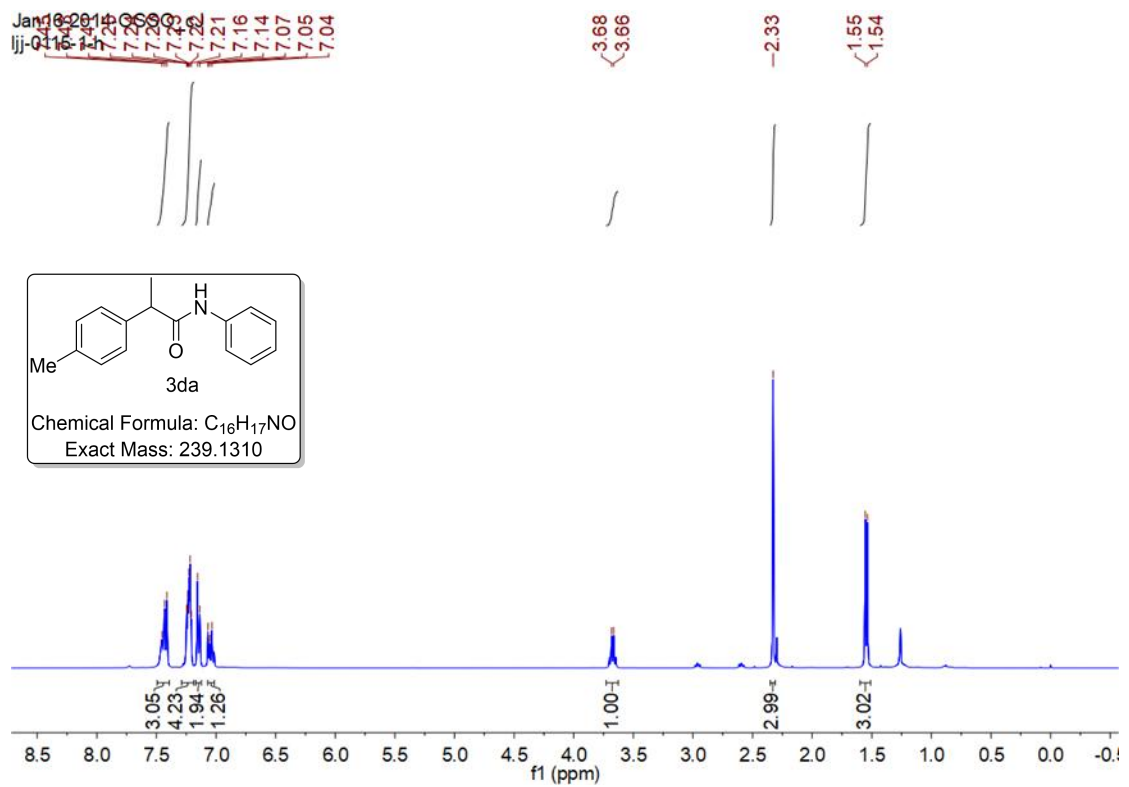
140.98  
138.76  
138.07  
128.95  
128.89  
128.39  
128.28  
124.73  
124.23  
119.94

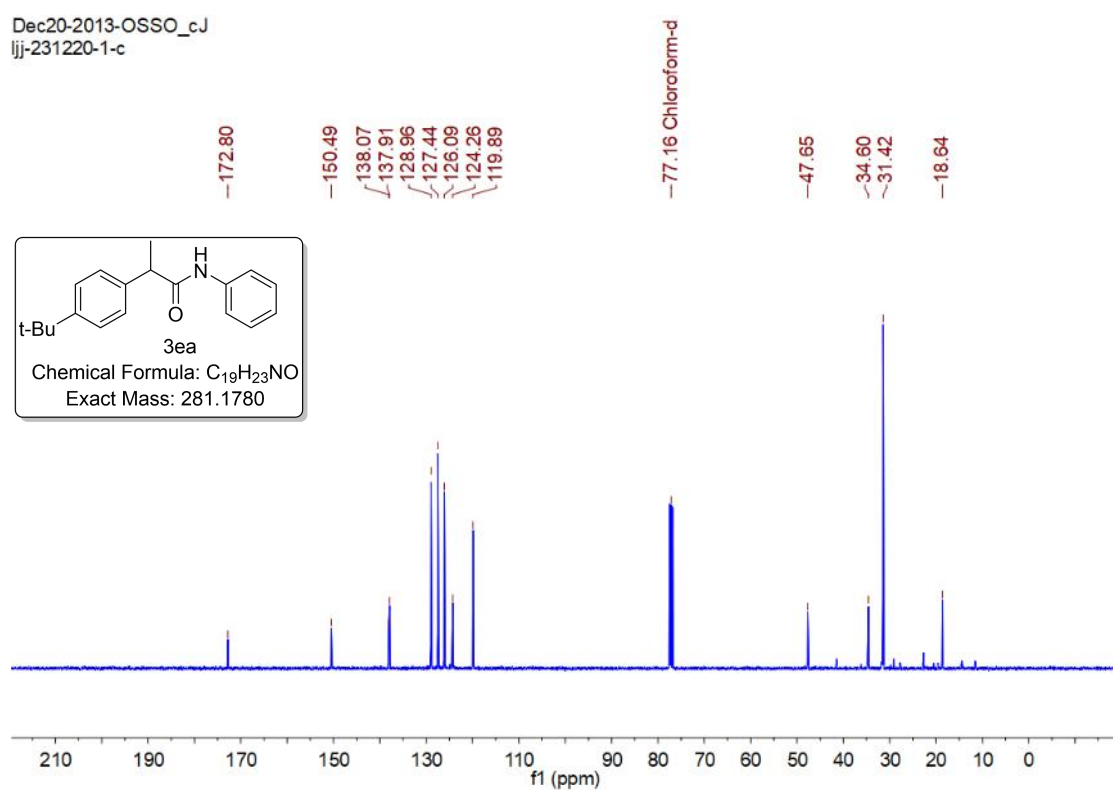
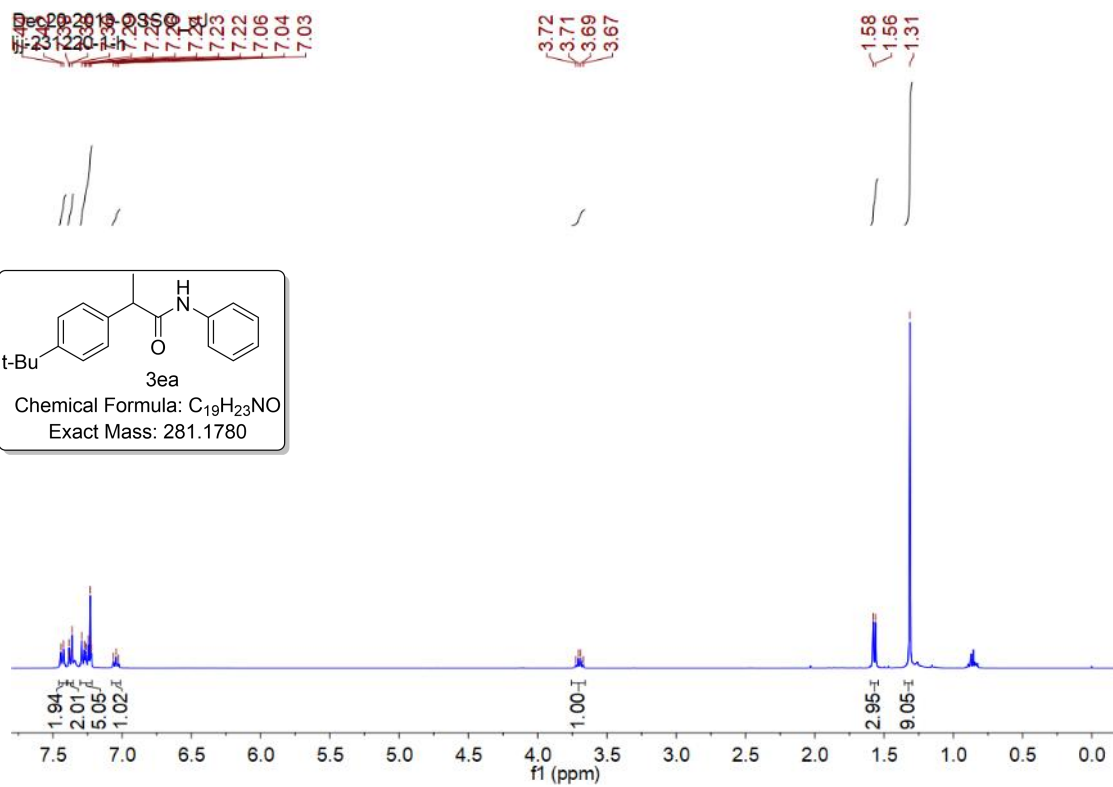
-77.16 Chloroform-d

-47.87

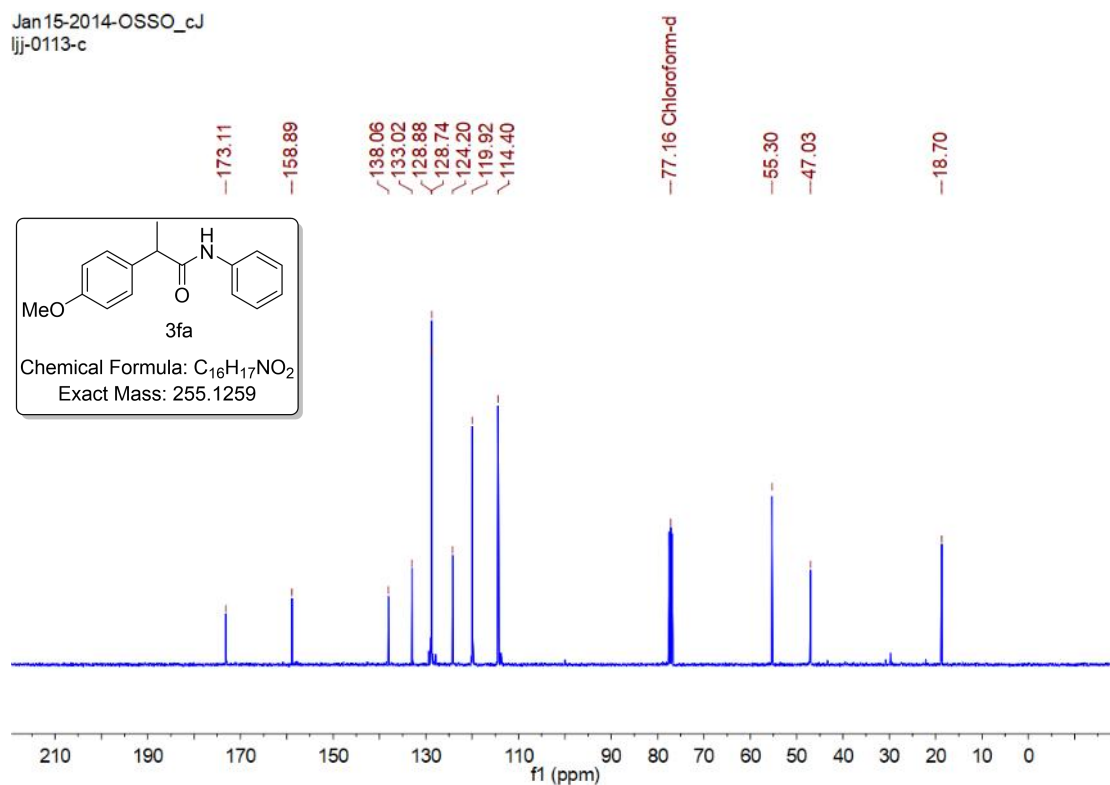
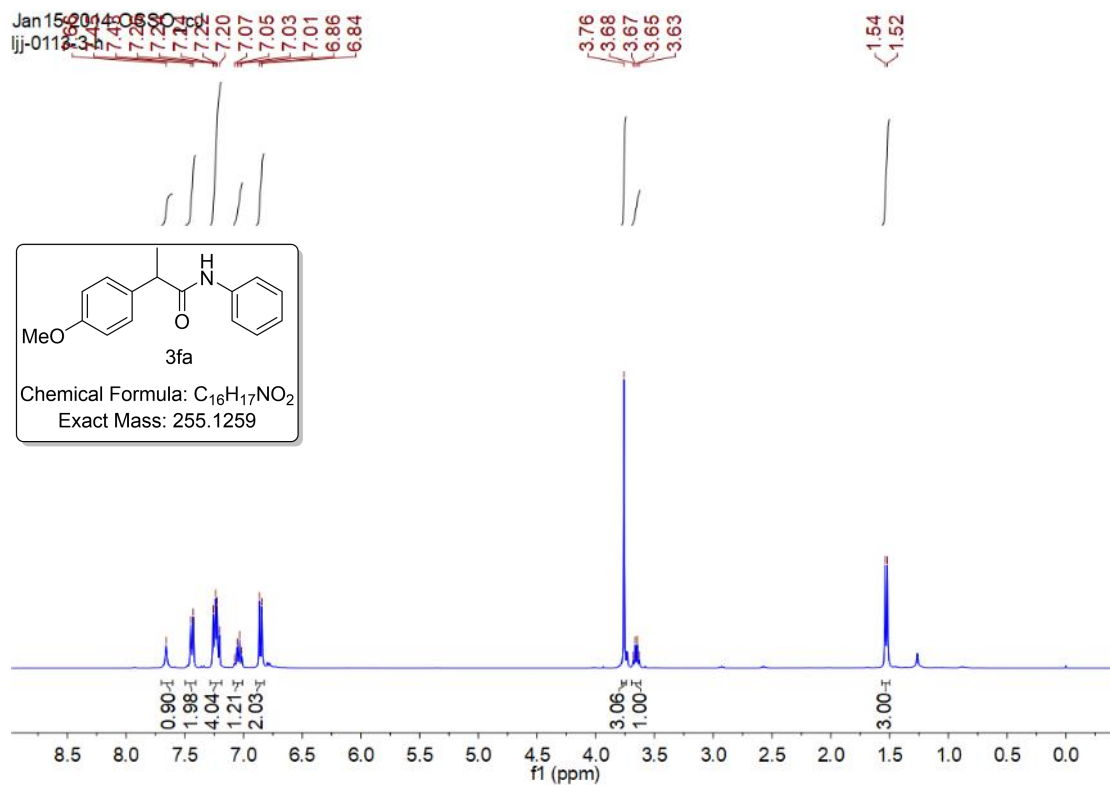
21.49  
18.62



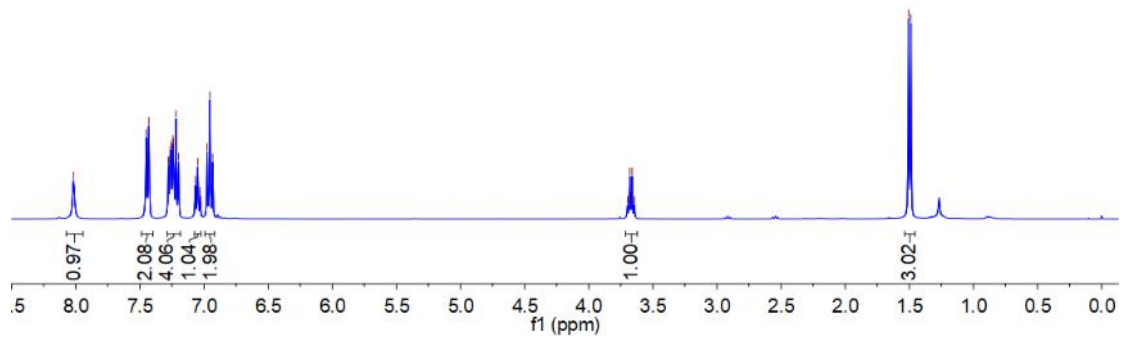
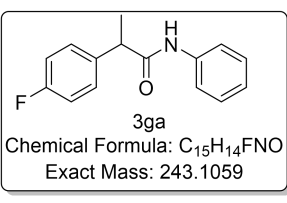








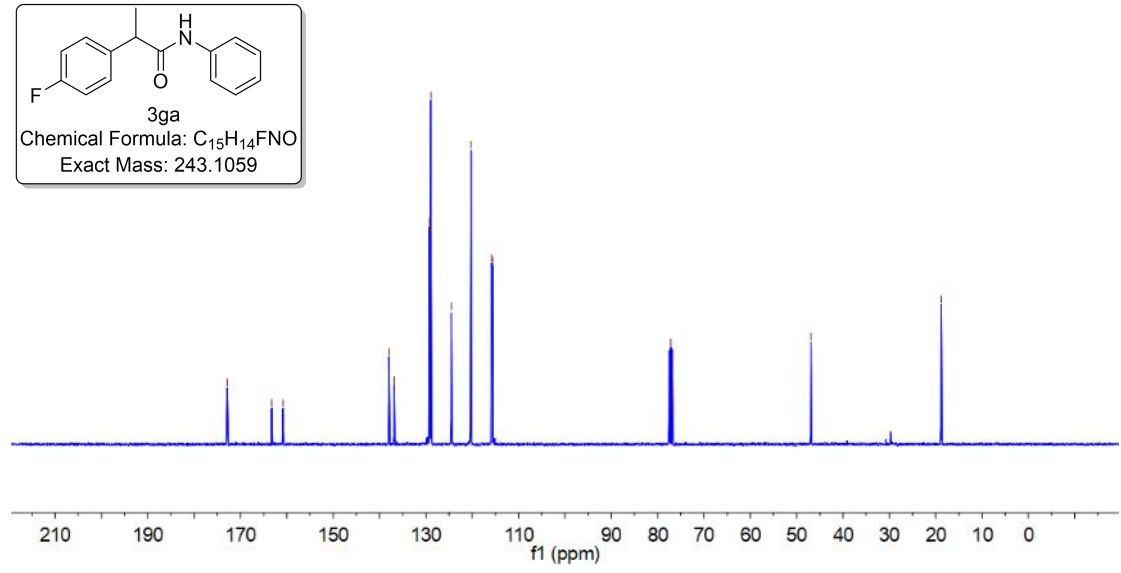
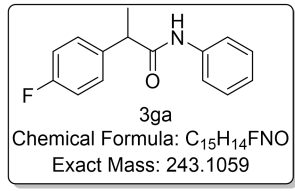
Apr25-2014-OSSO\_c  
ljj-120240424-b



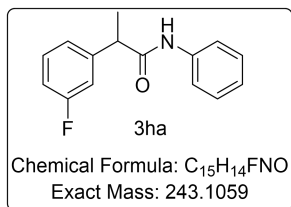
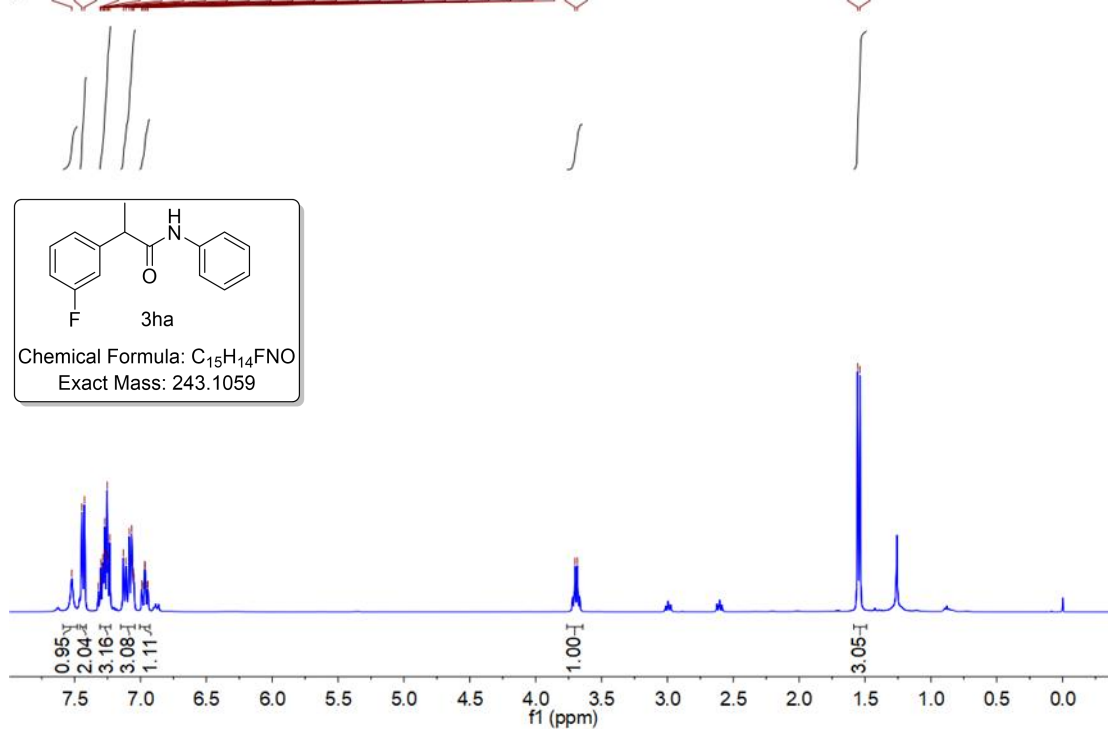
Apr25-2014-OSSO\_c\_j  
ljj-120240424-c

172.85  
163.27  
160.82  
137.91  
136.81  
136.77  
129.19  
129.11  
128.92  
124.50  
120.28  
115.80  
115.59

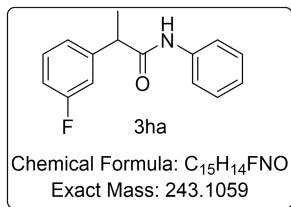
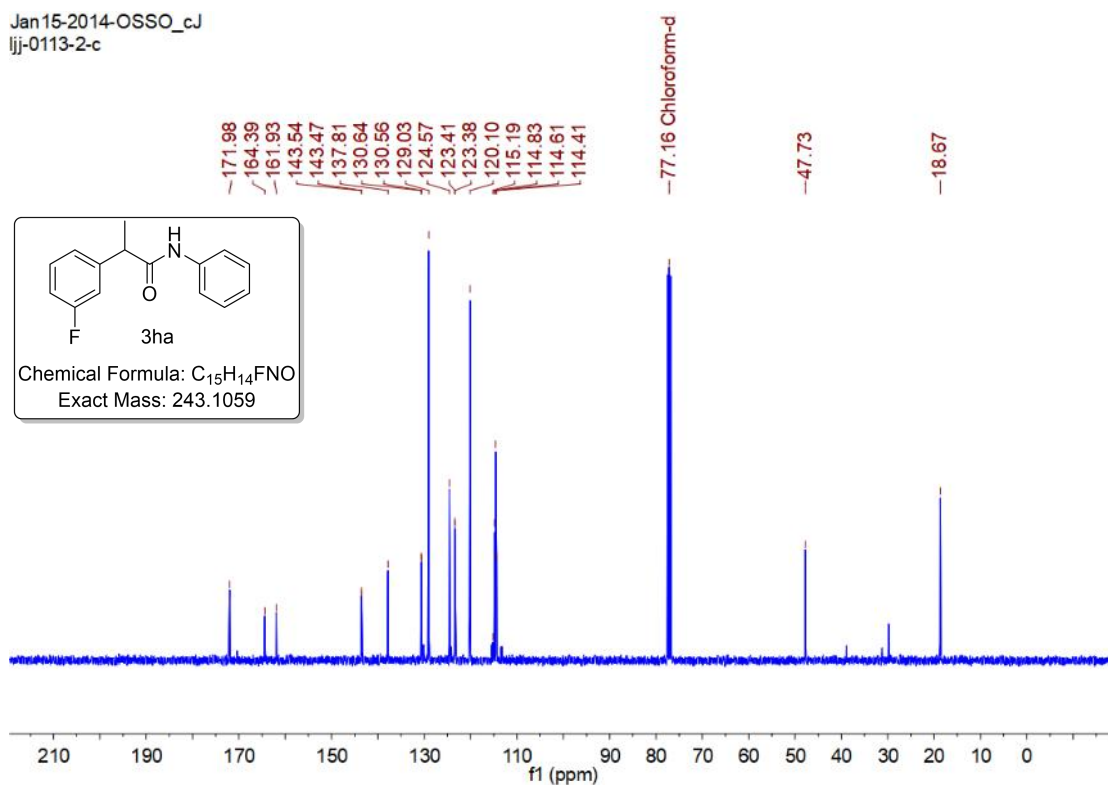
77.16 Chloroform-d  
46.91  
18.80



Jan 15-2014-OSSO\_c  
ljj-0113-2-c



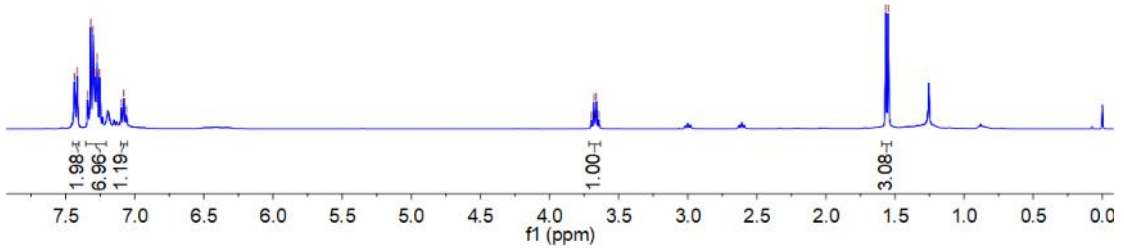
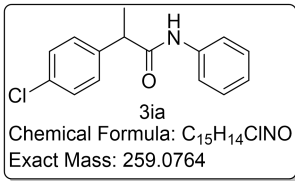
Jan 15-2014-OSSO\_c\_j  
ljj-0113-2-c



Apr24-2014-OSSO\_cj  
ljj-1-23240423-c

3.70  
3.68  
3.66  
3.64

1.57  
1.55



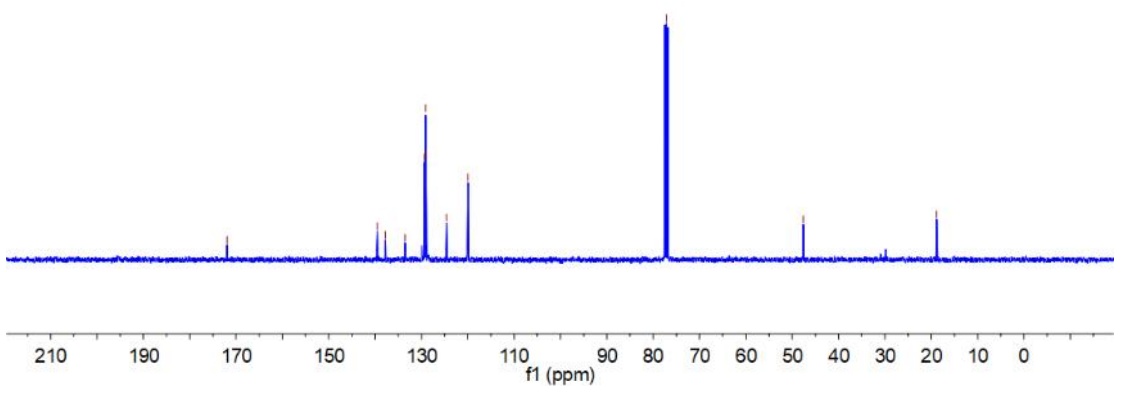
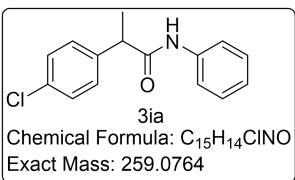
Apr24-2014-OSSO\_cj  
ljj-1-23240423-c

-77.16 Chloroform-d

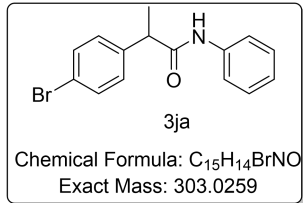
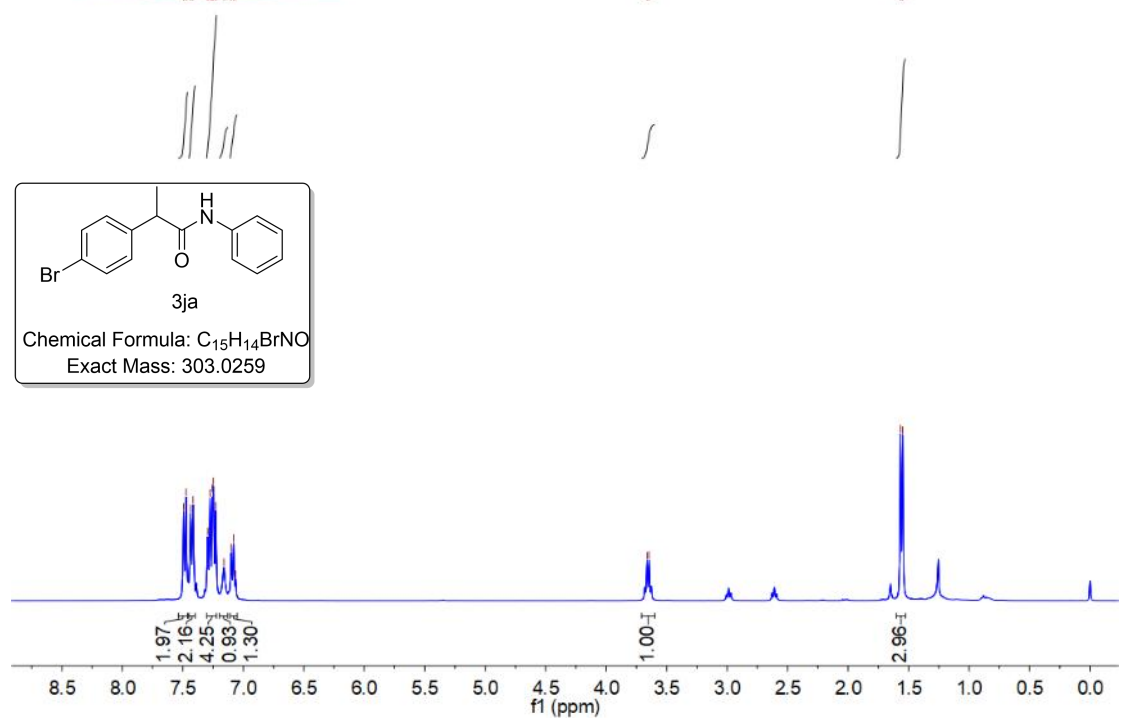
171.93  
139.53  
137.80  
133.52  
129.34  
129.12  
129.09  
124.59  
119.94

-47.60

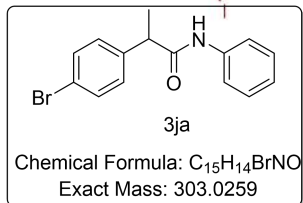
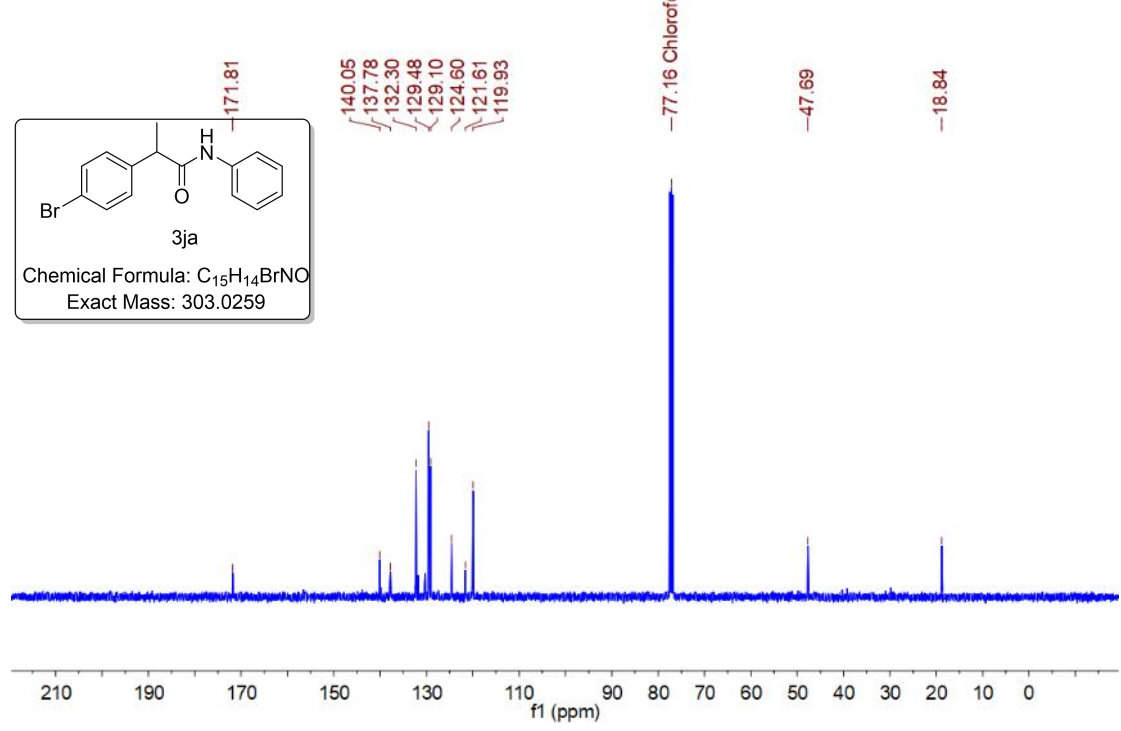
-18.87

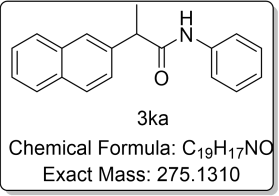
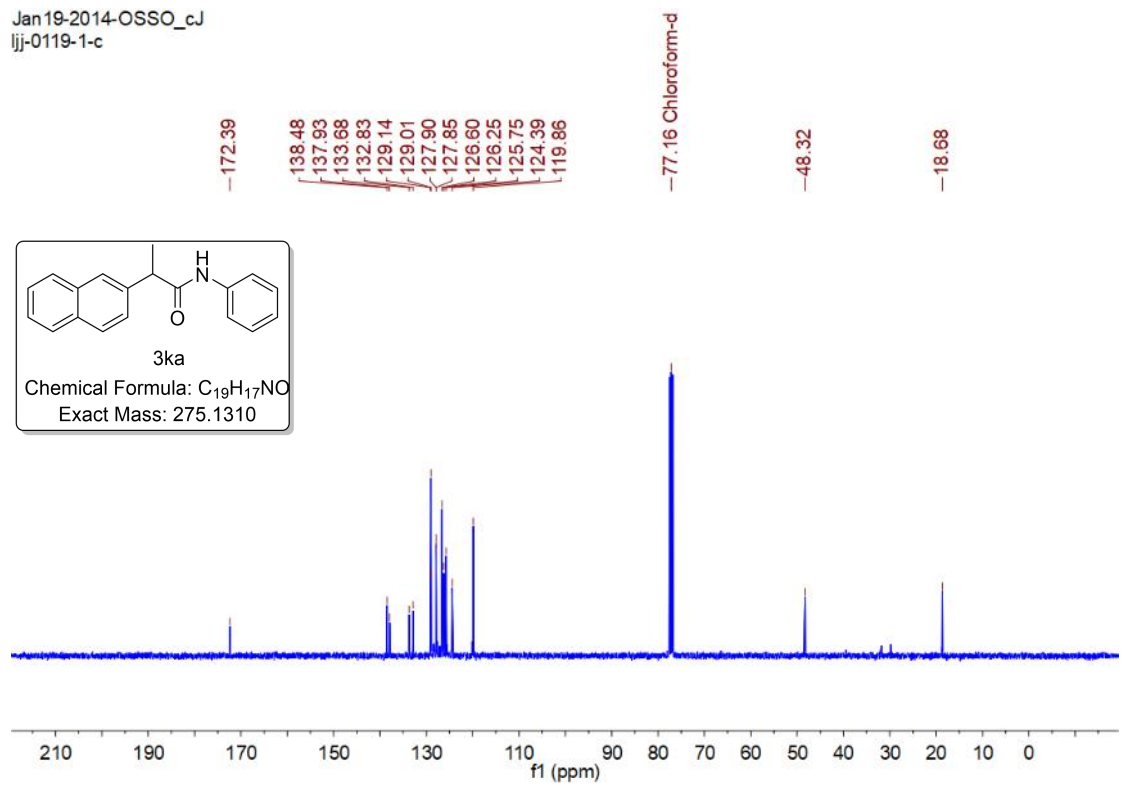
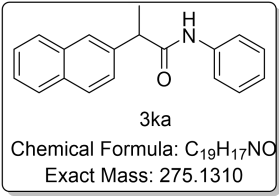
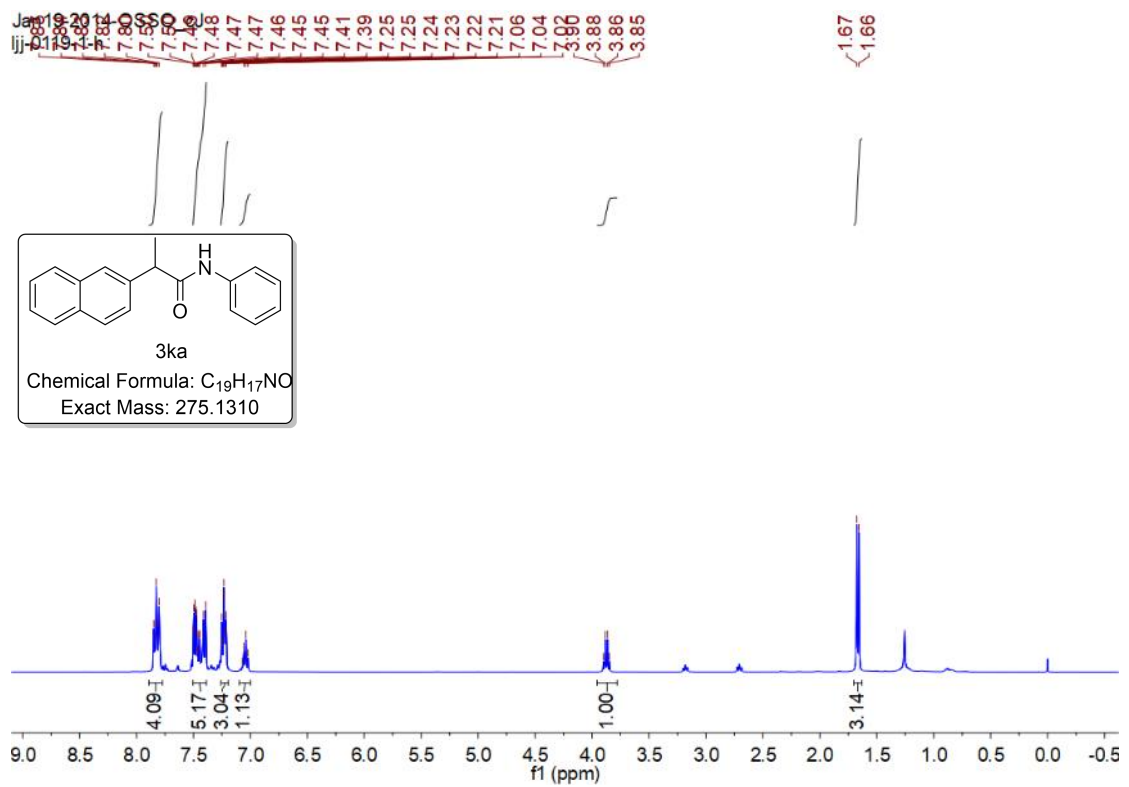


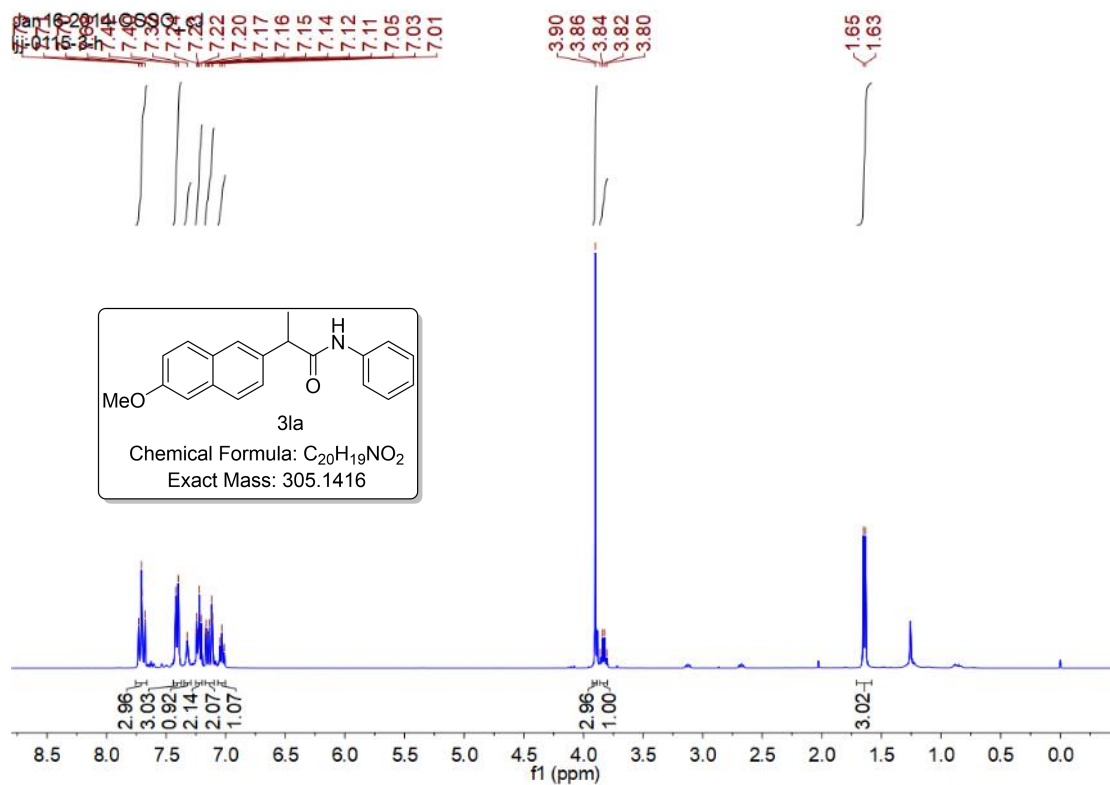
Jan17-2014-OSSO  
ljj-0117-1-c



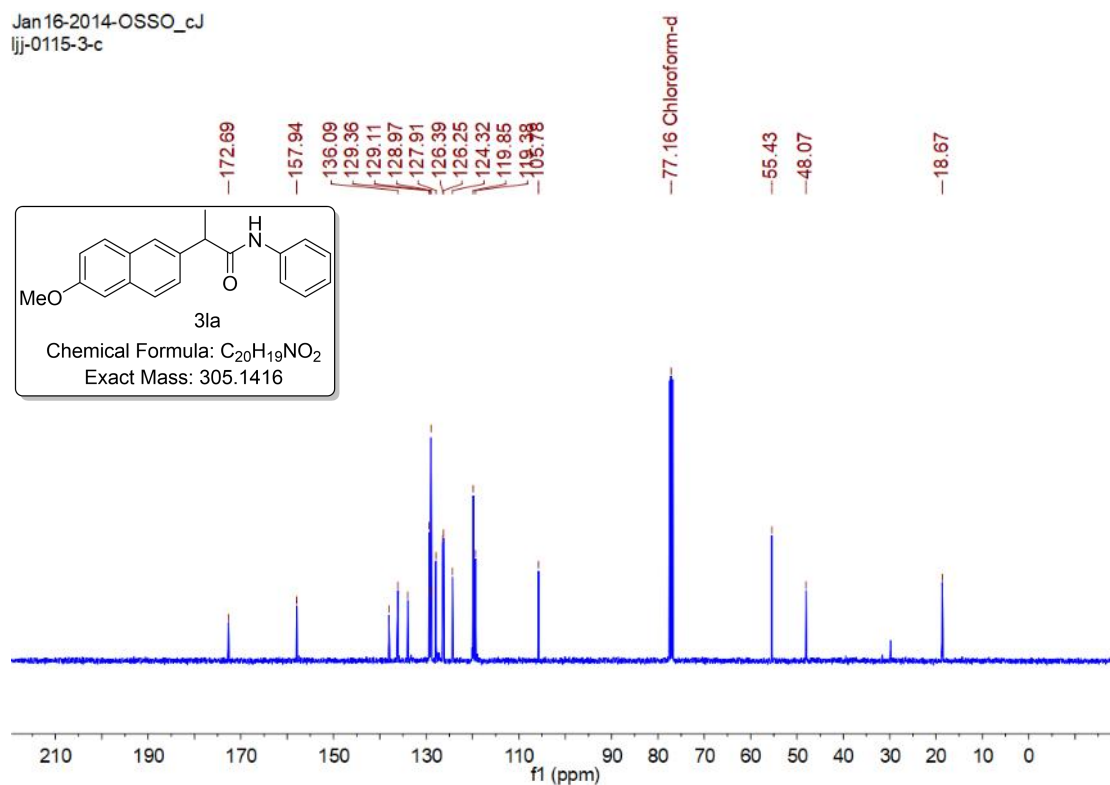
Jan17-2014-OSSO\_cj  
ljj-0117-1-c



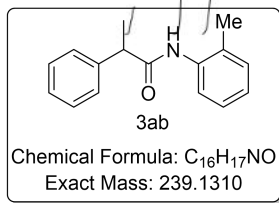
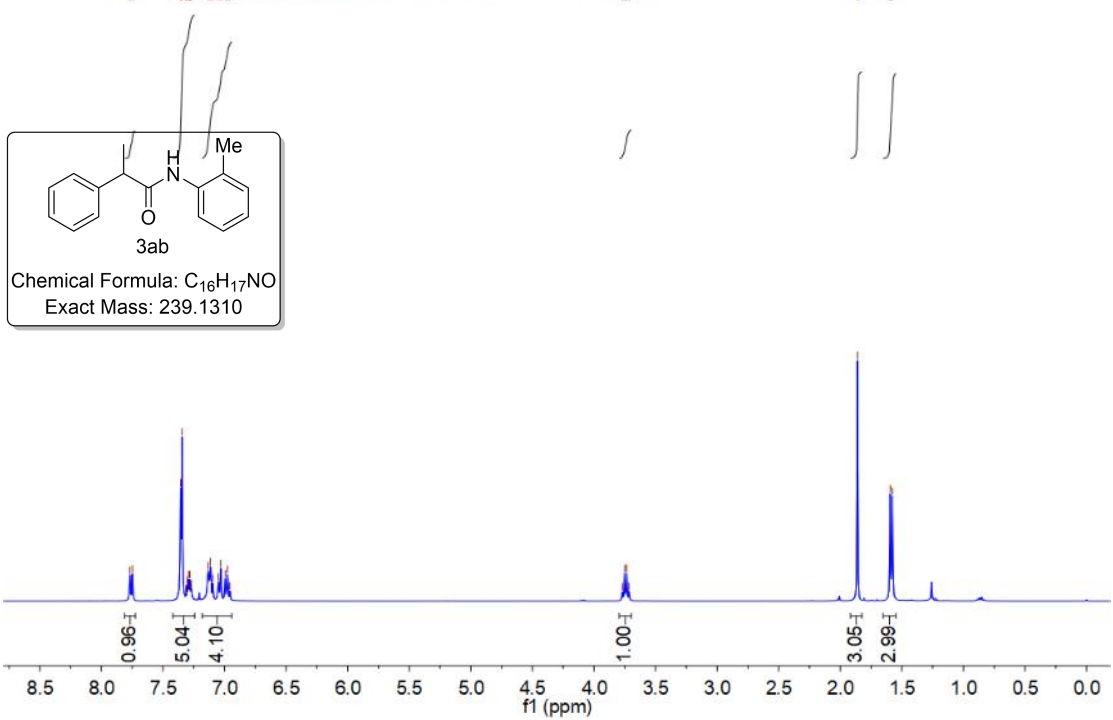




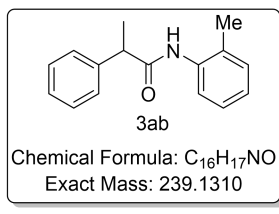
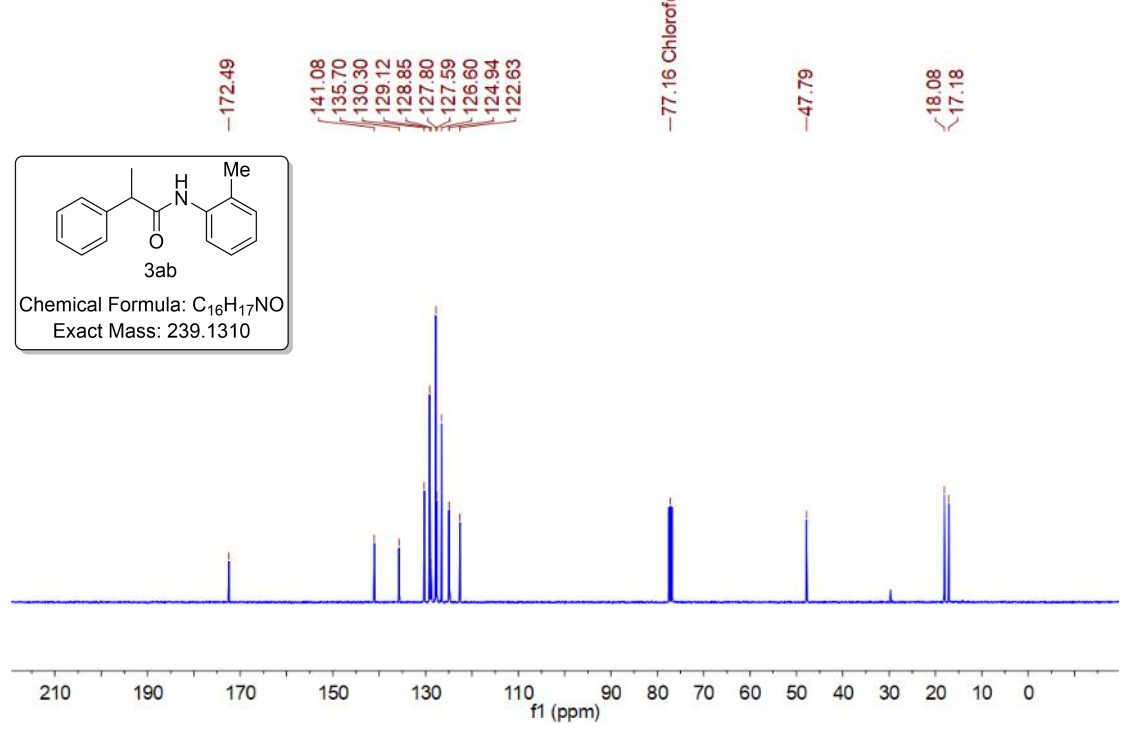
Jan16-2014-OSSO\_cj  
ljj-0115-3-c



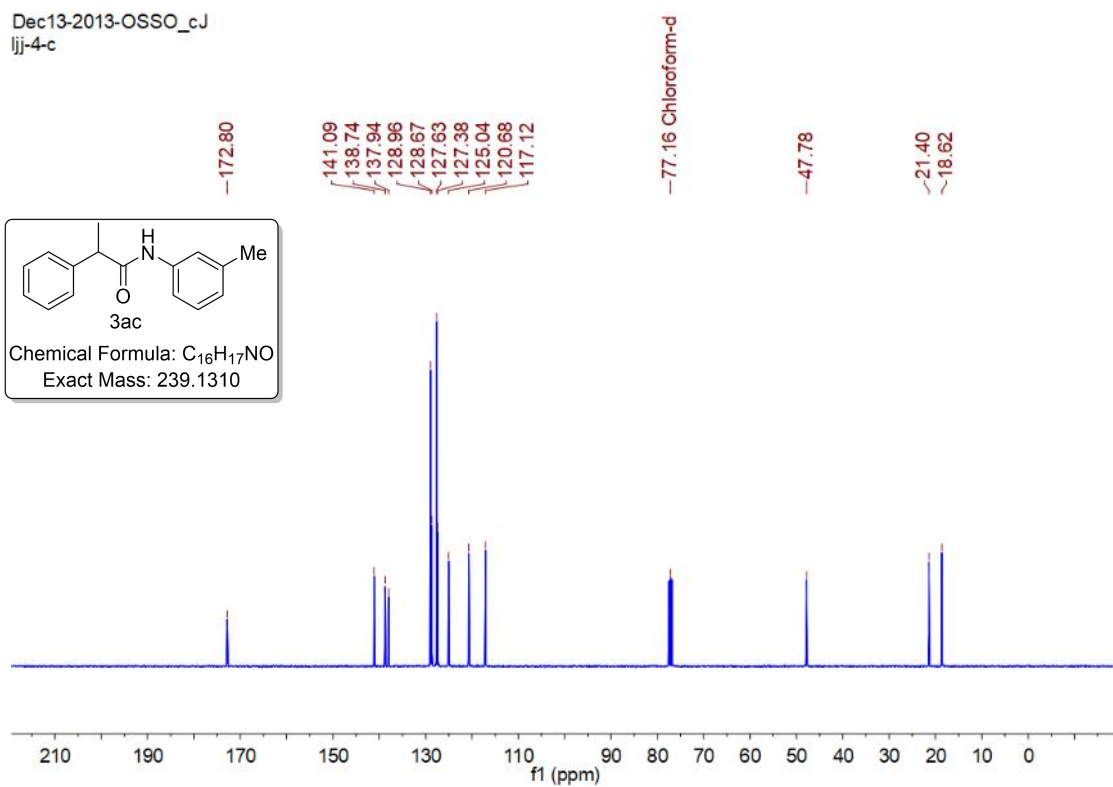
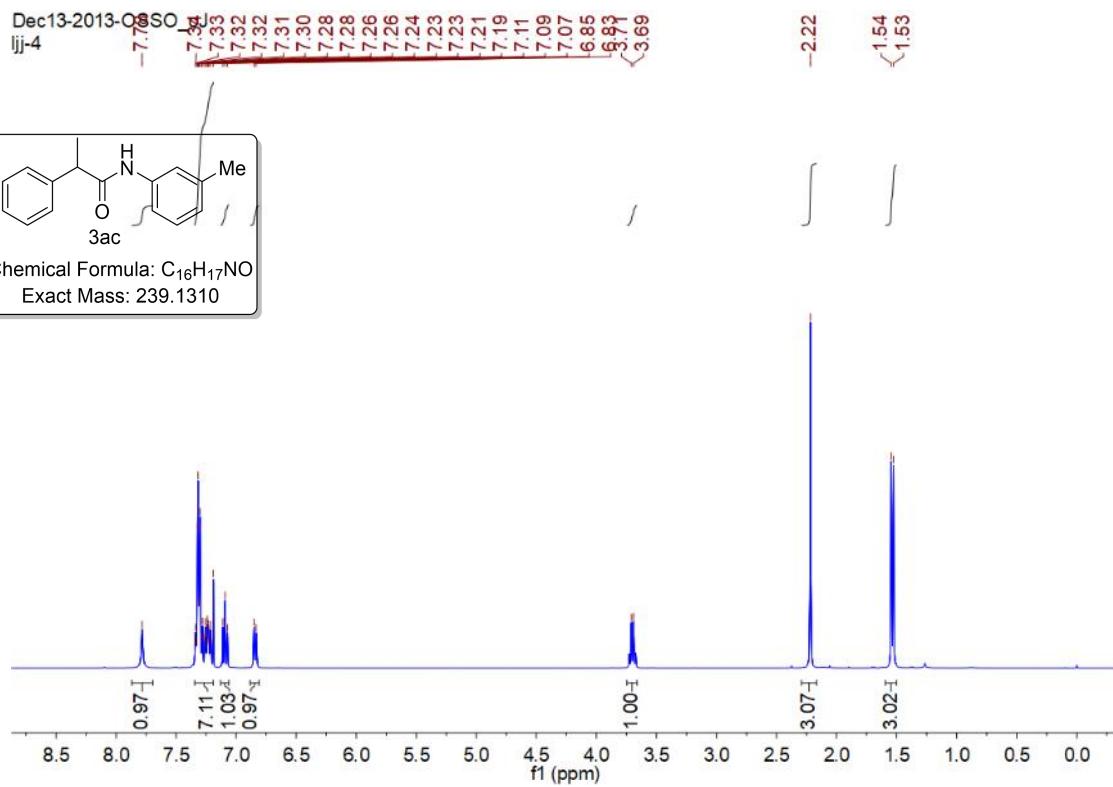
Dec13-2013-OSSO\_c  
 ljj-1213-3-h



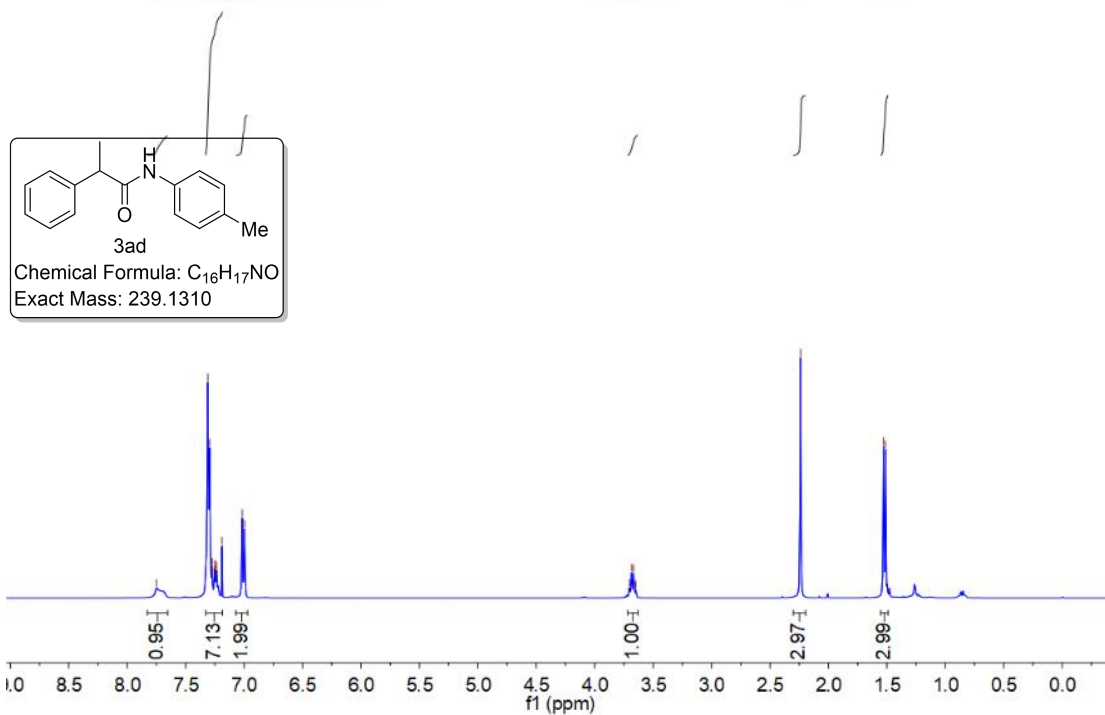
Dec13-2013-OSSO\_c\_j  
 ljj-1213-3-c



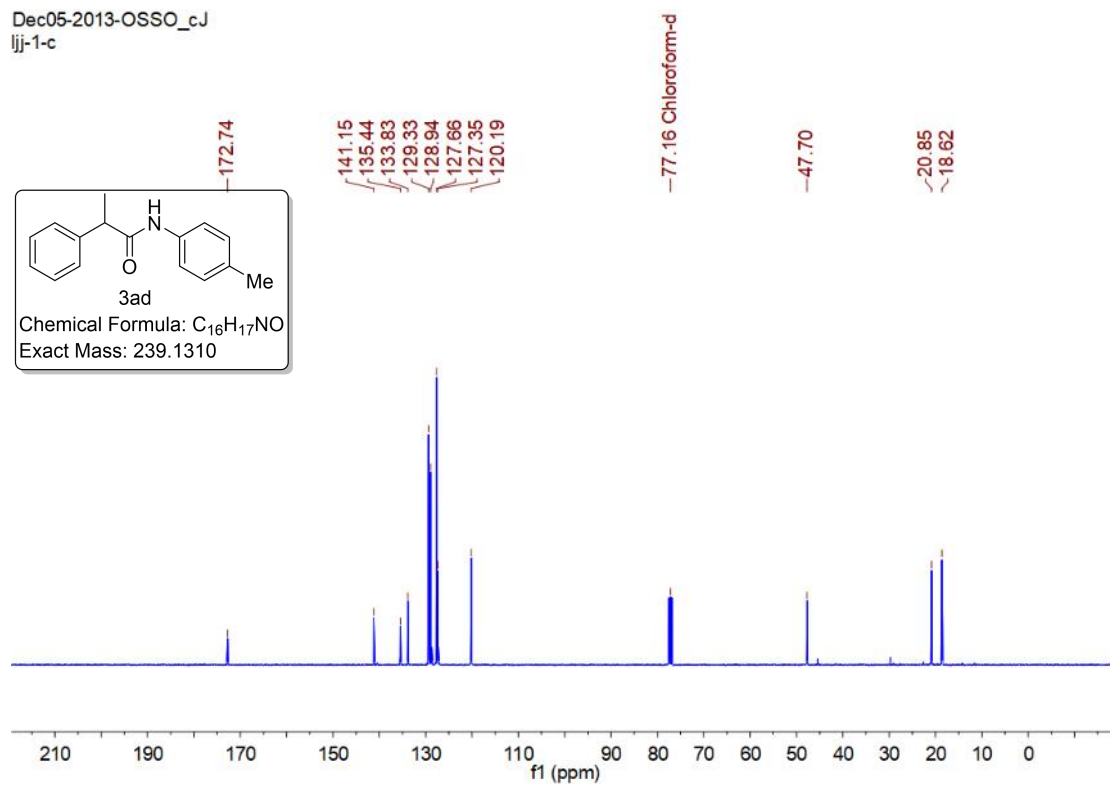


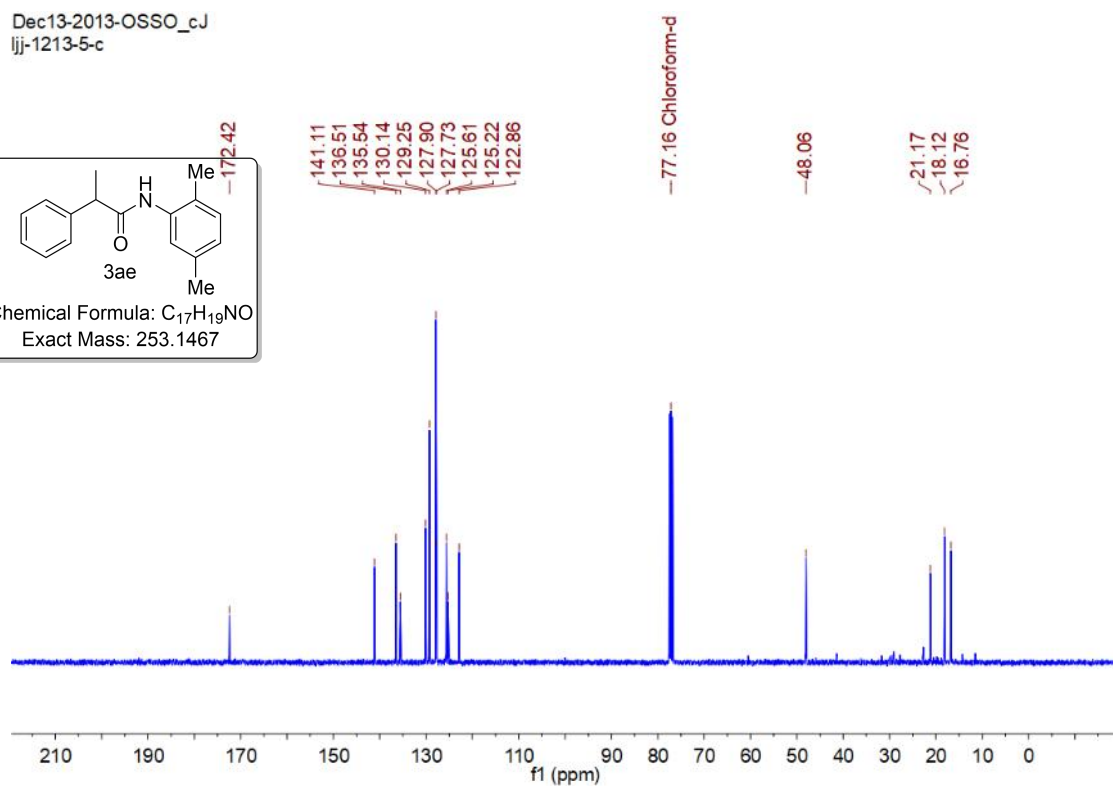
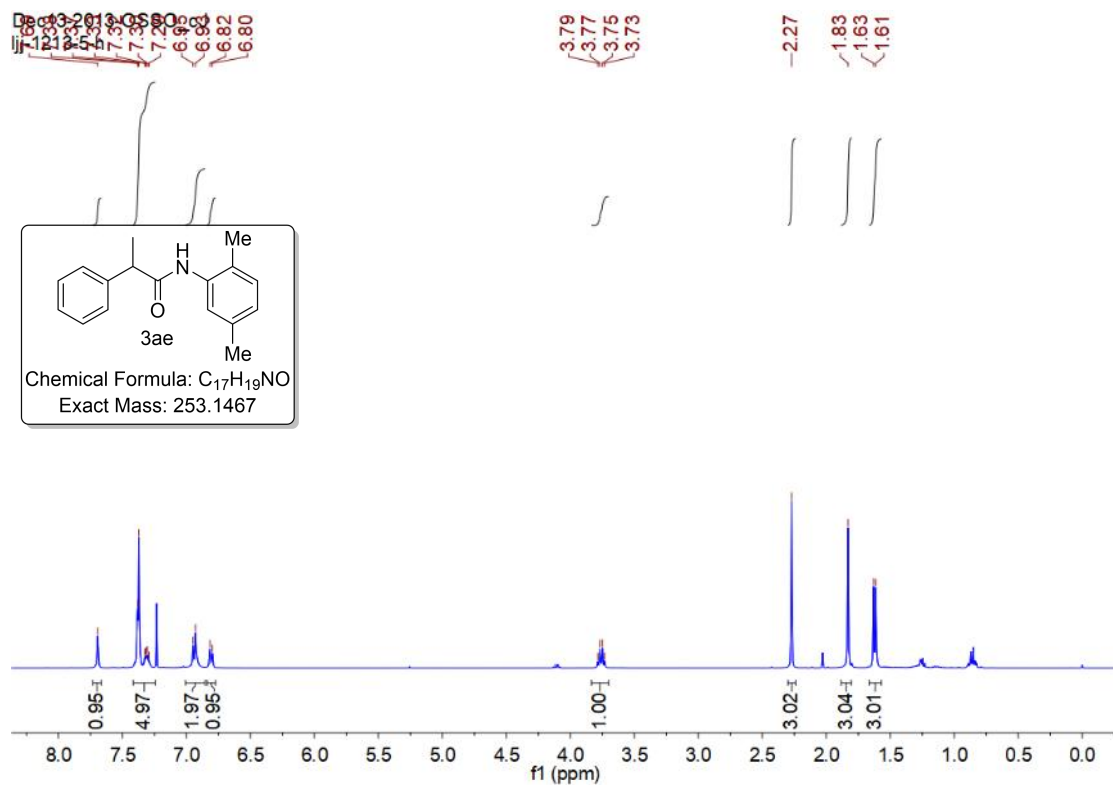


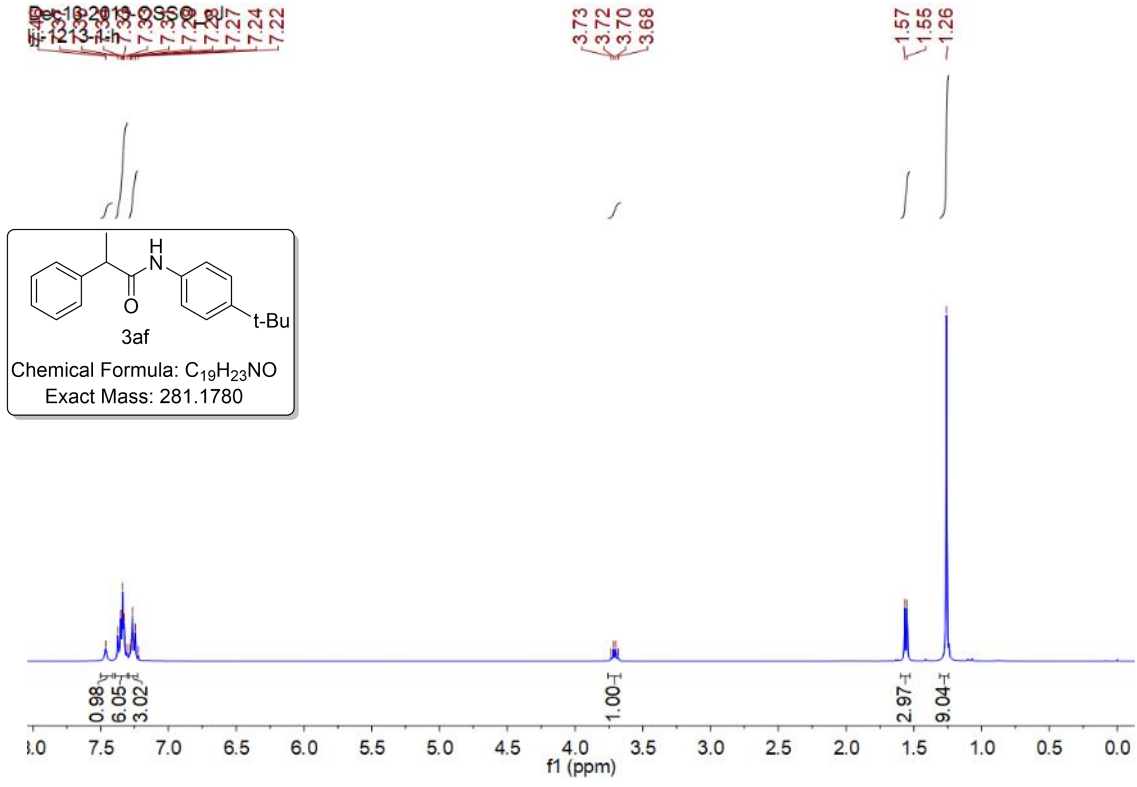
Dec05-2013-OSSO\_cj  
ljj-1



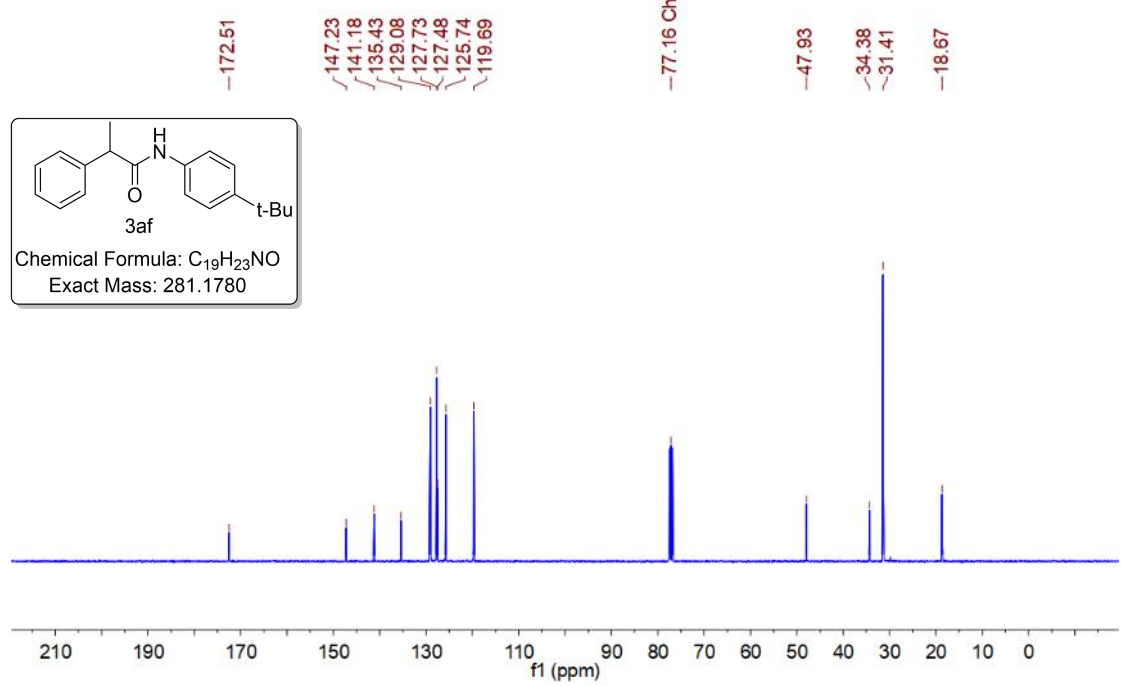
Dec05-2013-OSSO\_cj  
ljj-1-c

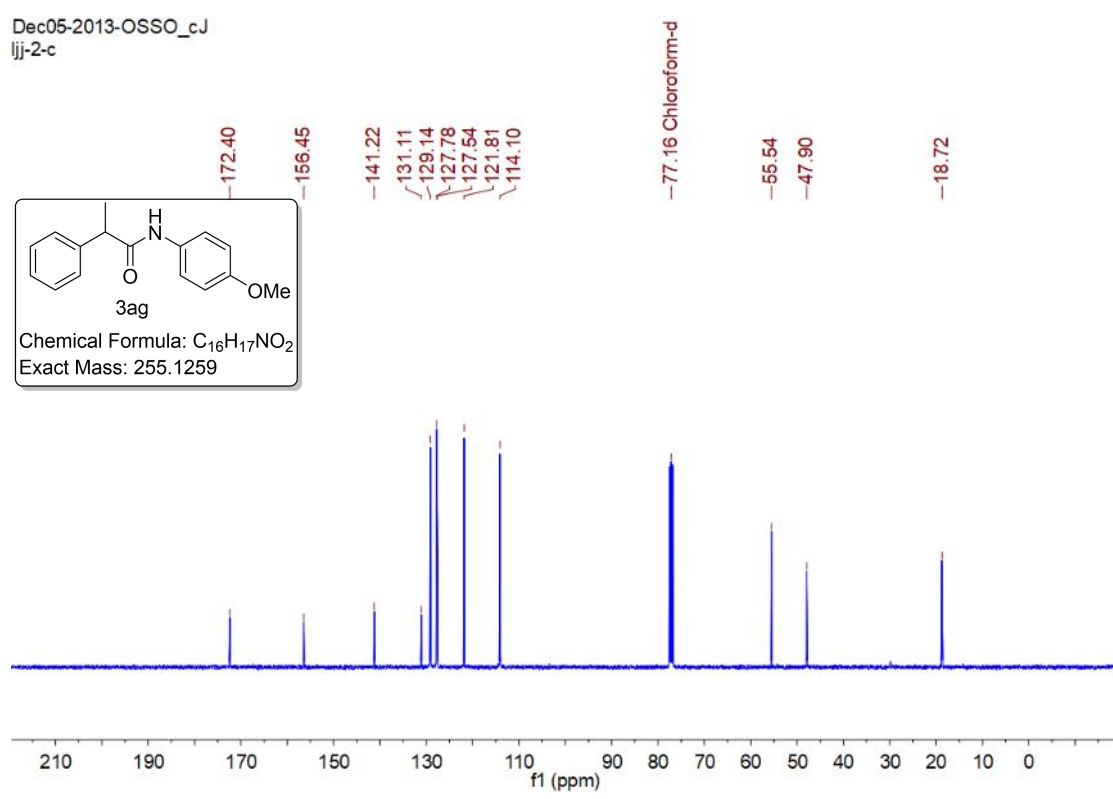
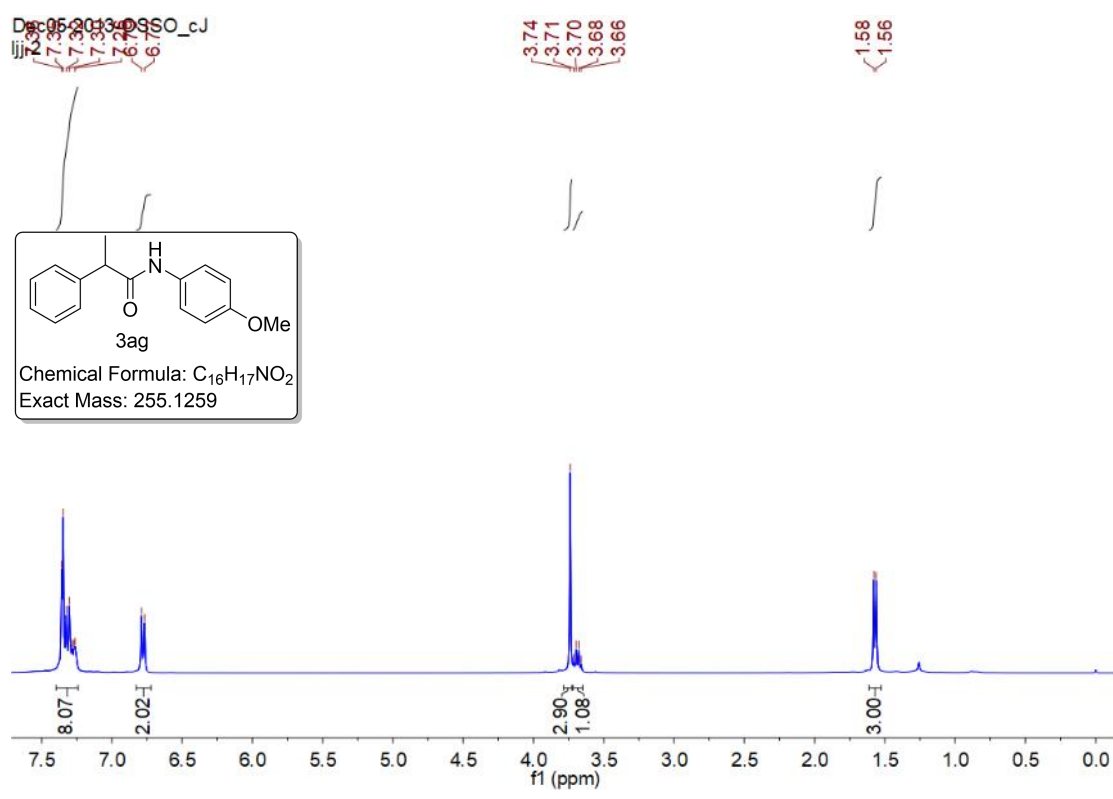




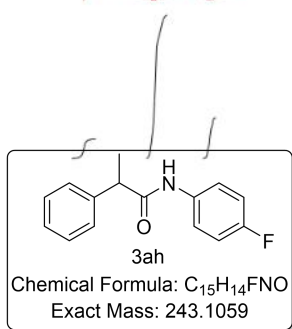


Dec13-2013-OSSO\_c.J  
ljj-1213-1-c



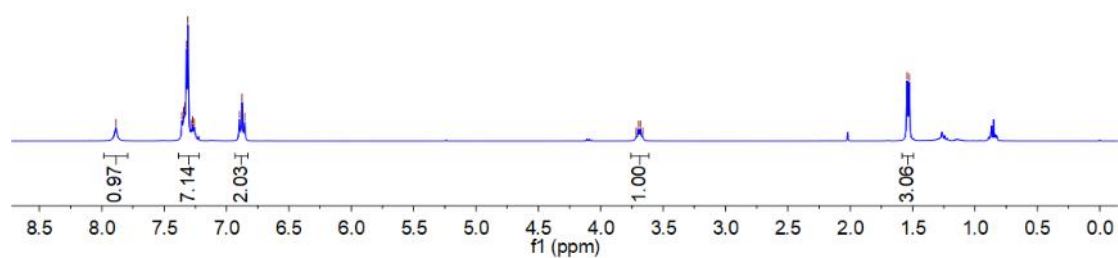


Dec07-2013-OSSO\_cJ  
ljj-1207-1-h

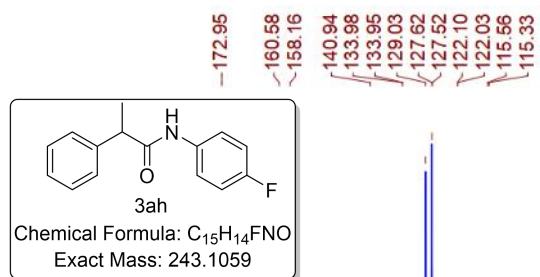


3.72  
3.70  
3.68  
3.66

1.54  
1.53



Dec07-2013-OSSO\_cJ  
ljj-1207-1-c

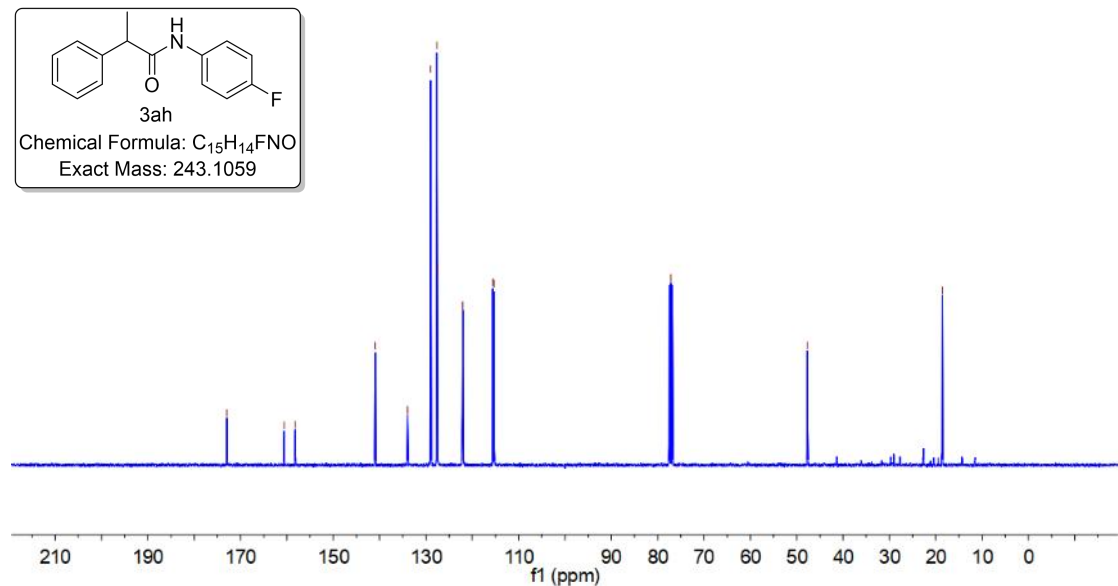


172.95  
160.58  
158.16  
140.94  
133.98  
133.95  
129.03  
127.62  
127.52  
122.10  
122.03  
115.56  
115.33

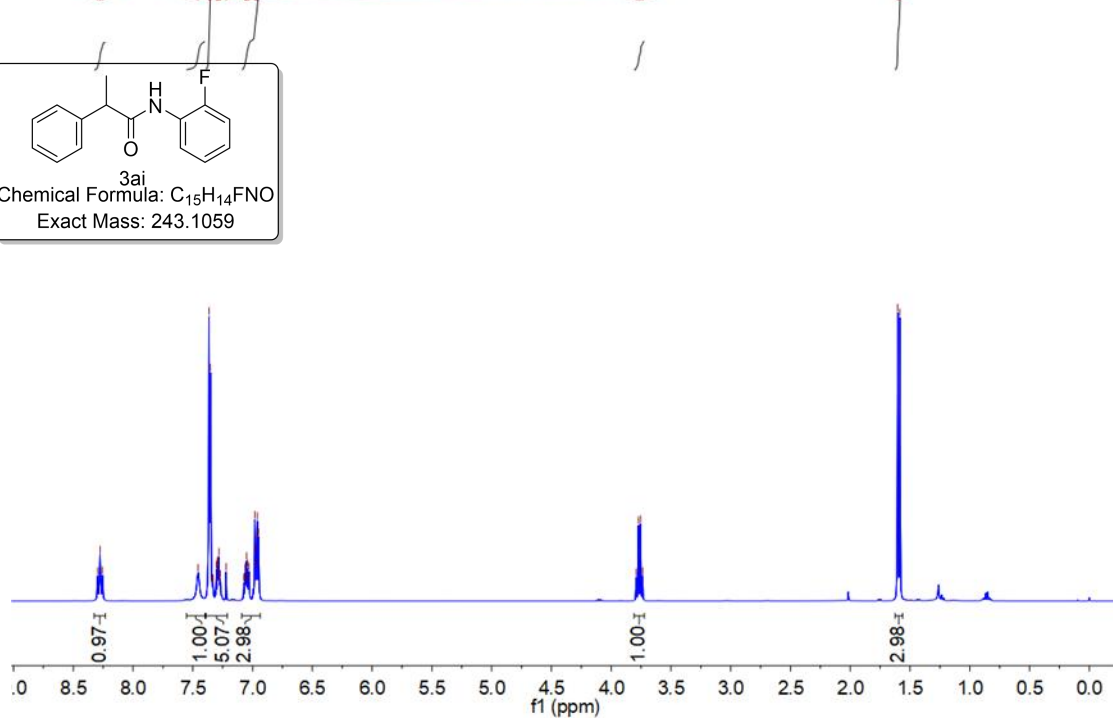
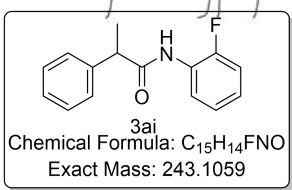
77.16 Chloroform-d

47.65

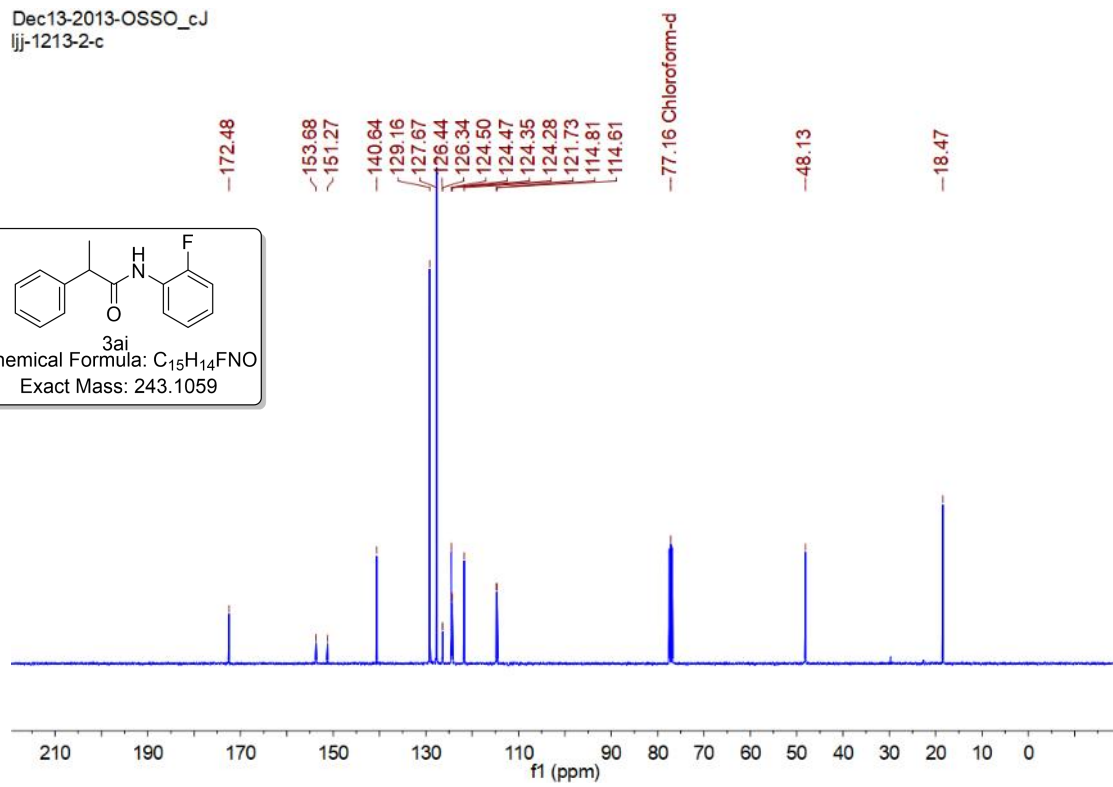
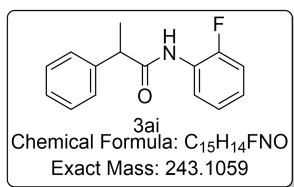
18.59



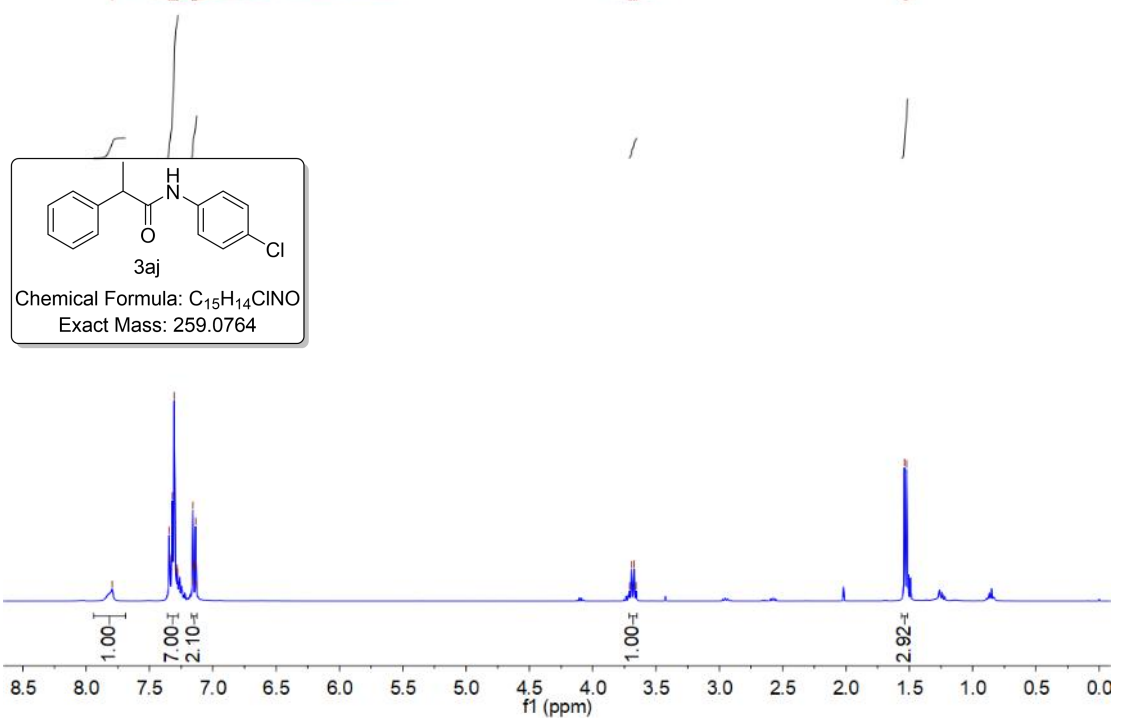
Dec13-2013-OSSO\_cj  
 ljj-1213-2-c



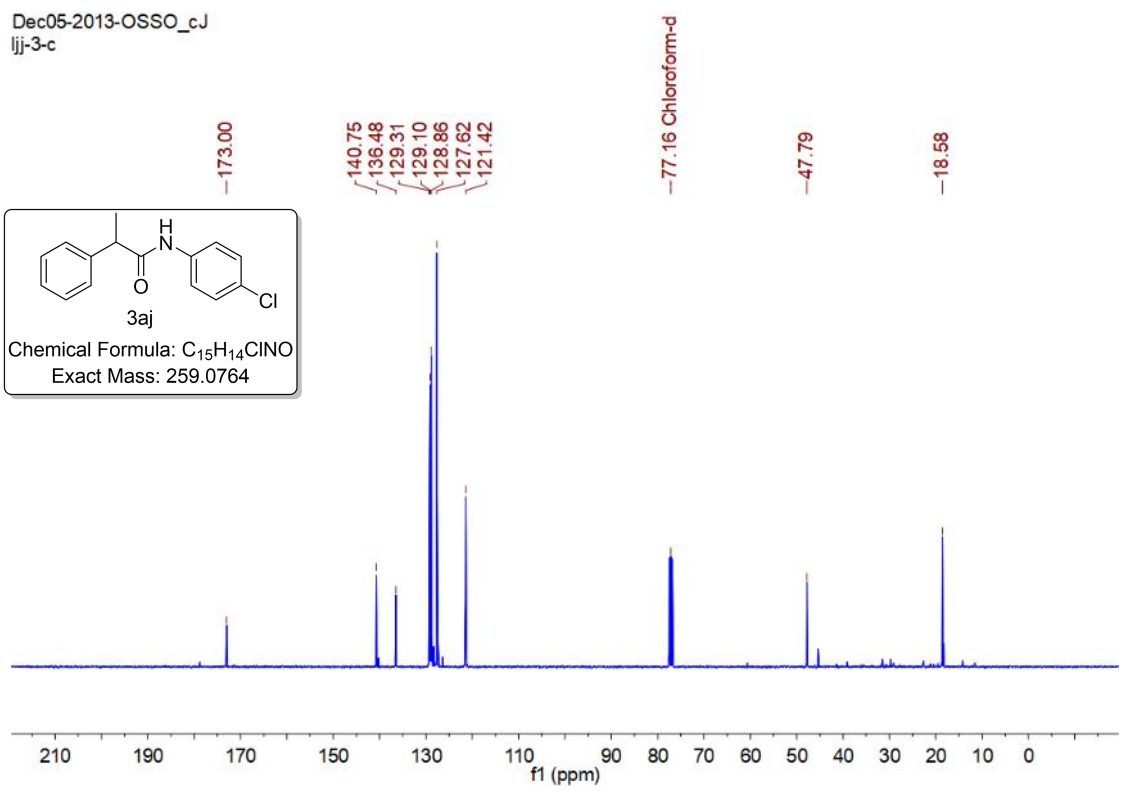
Dec13-2013-OSSO\_cj  
 ljj-1213-2-c



Dec05-2013-OSSO\_cj  
lji-3



Dec05-2013-OSSO\_cj  
lji-3-c

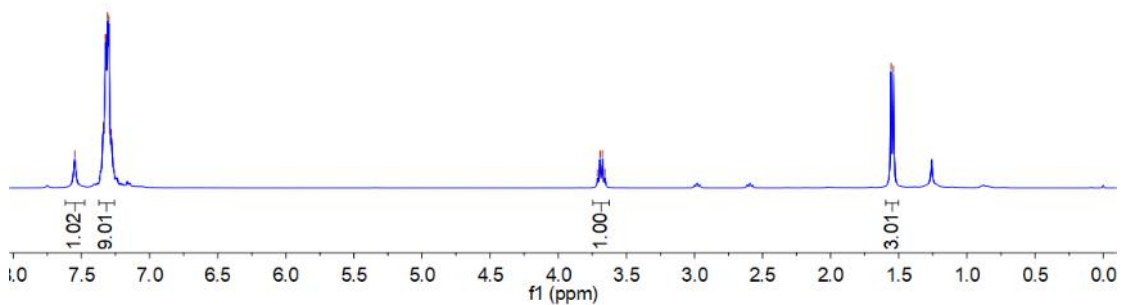
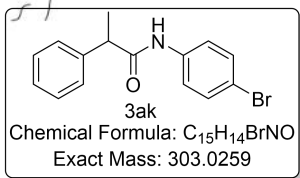




Apr25-2014-OSSO\_cj  
ljj-2-20240424-h

3.71  
3.69  
3.68  
3.66

1.56  
1.54



Apr25-2014-OSSO\_cj  
ljj-2-20240424-c

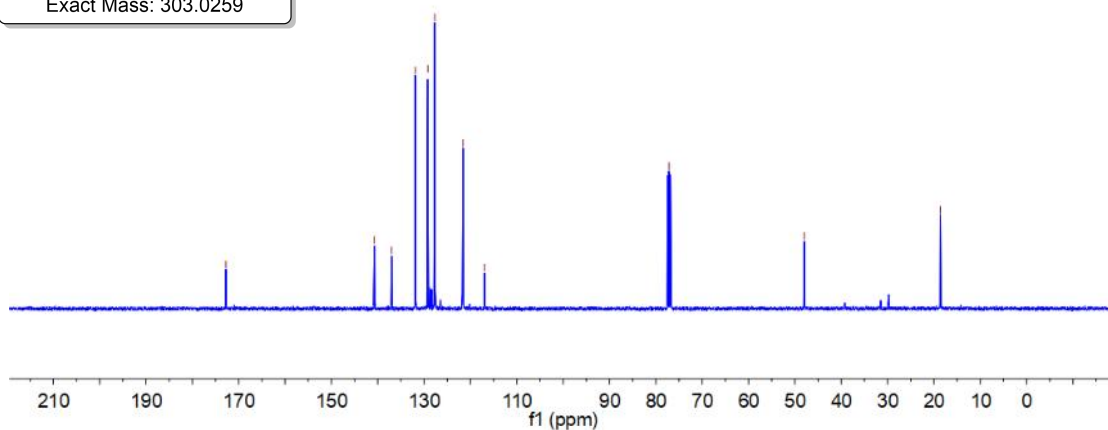
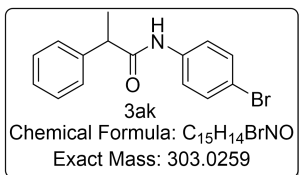
-77.16 Chloroform-d

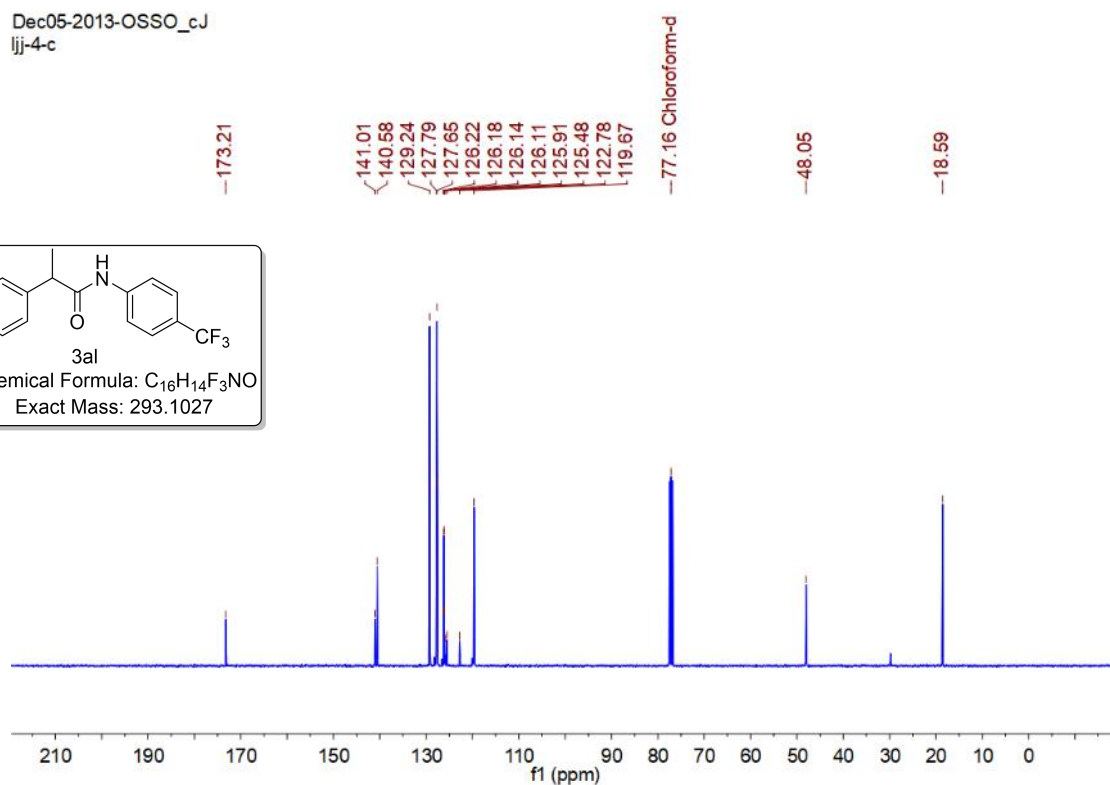
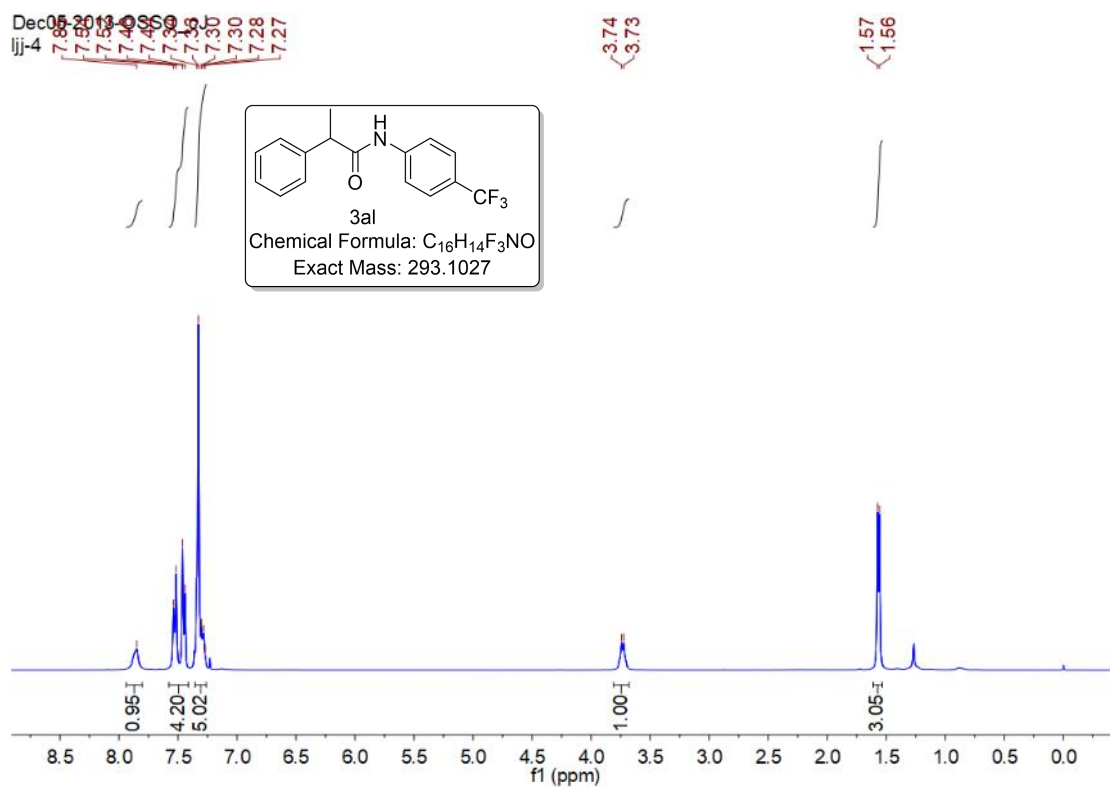
-172.77

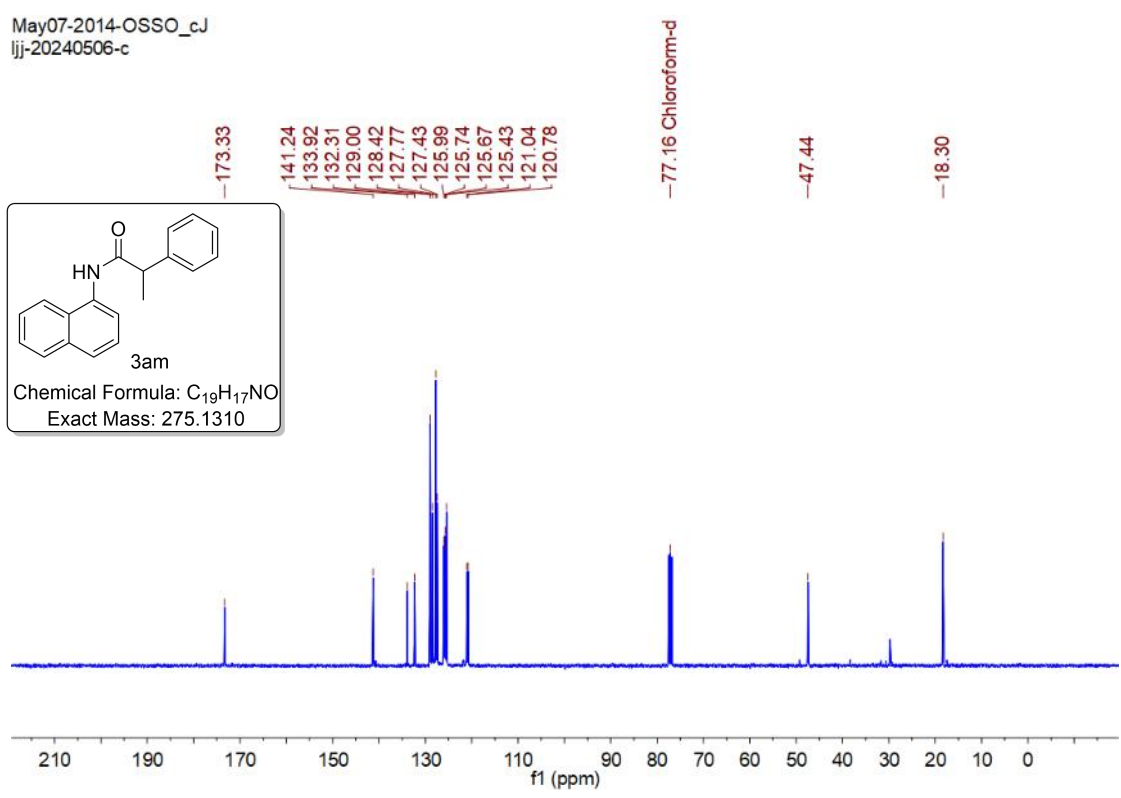
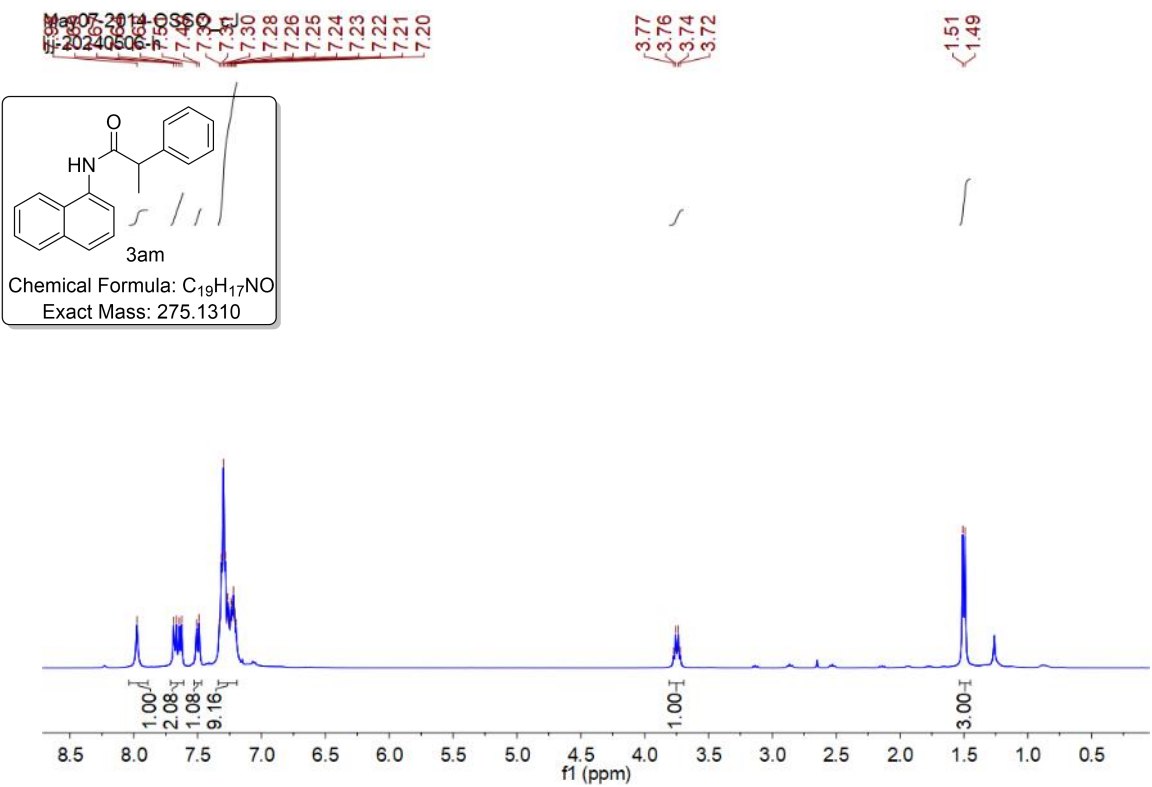
140.75  
137.01  
131.87  
129.20  
127.68  
121.62  
116.94

-47.97

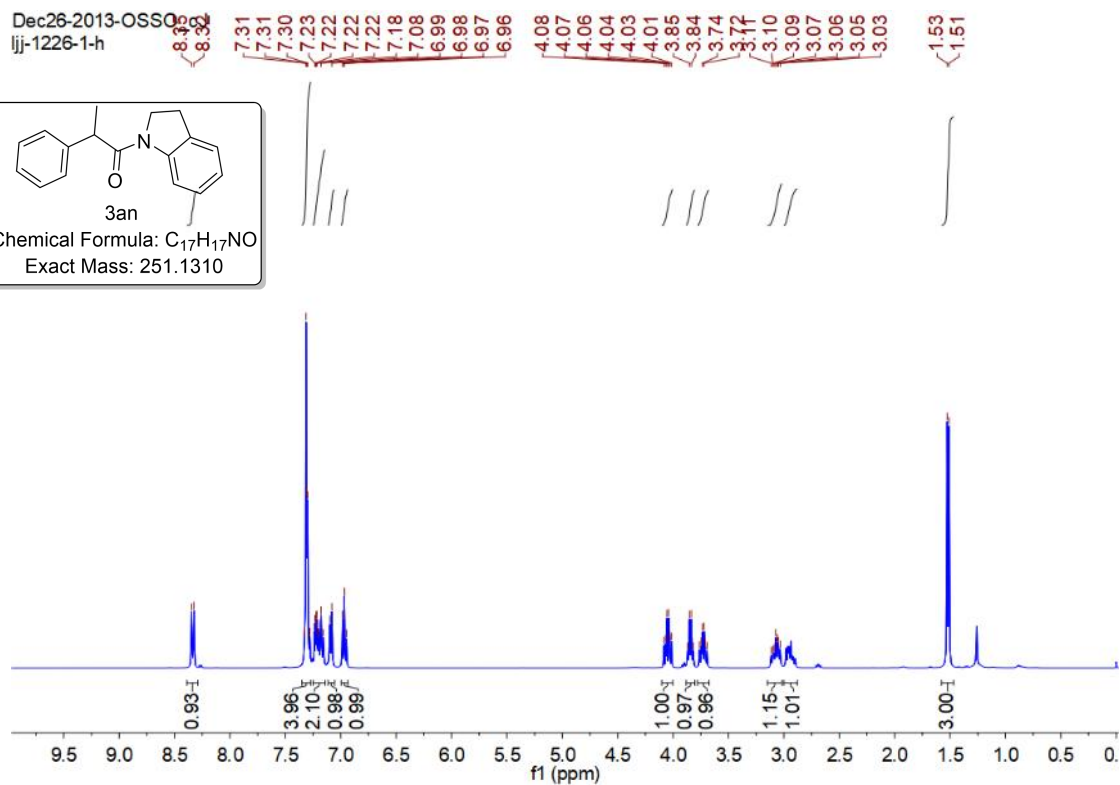
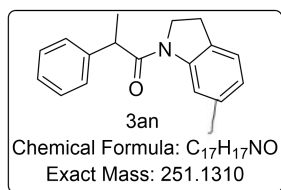
-18.63



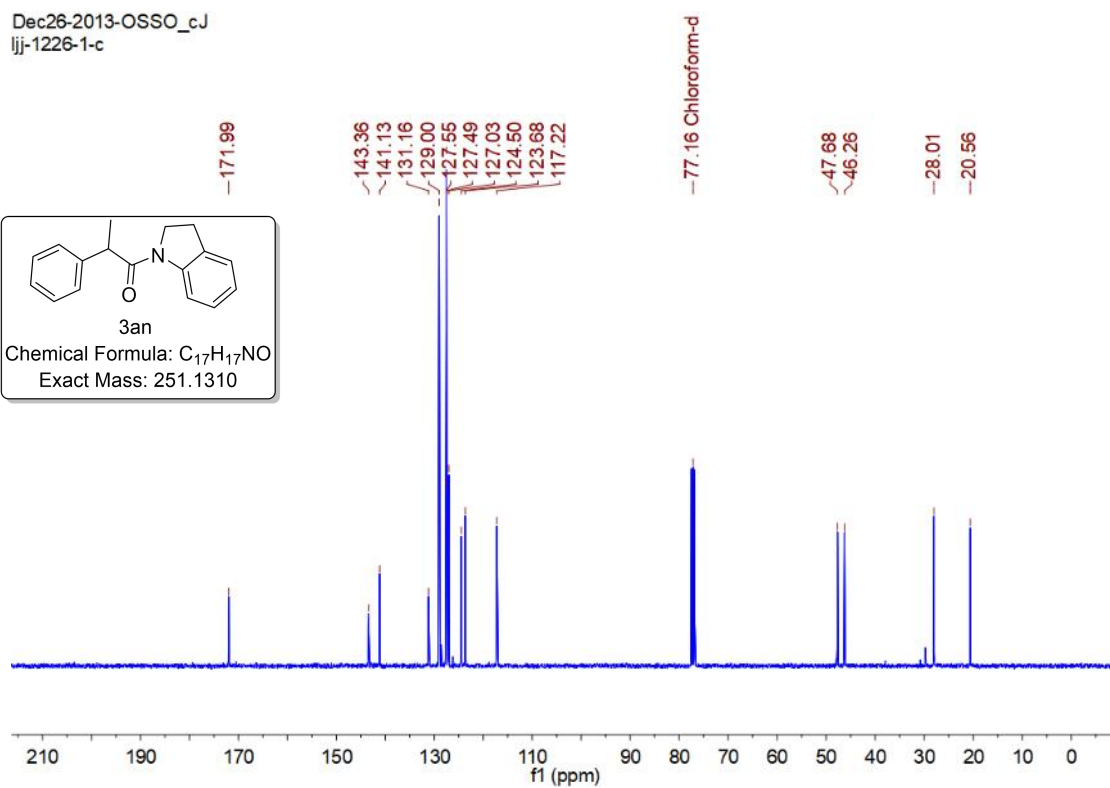
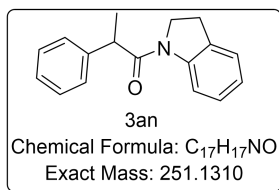


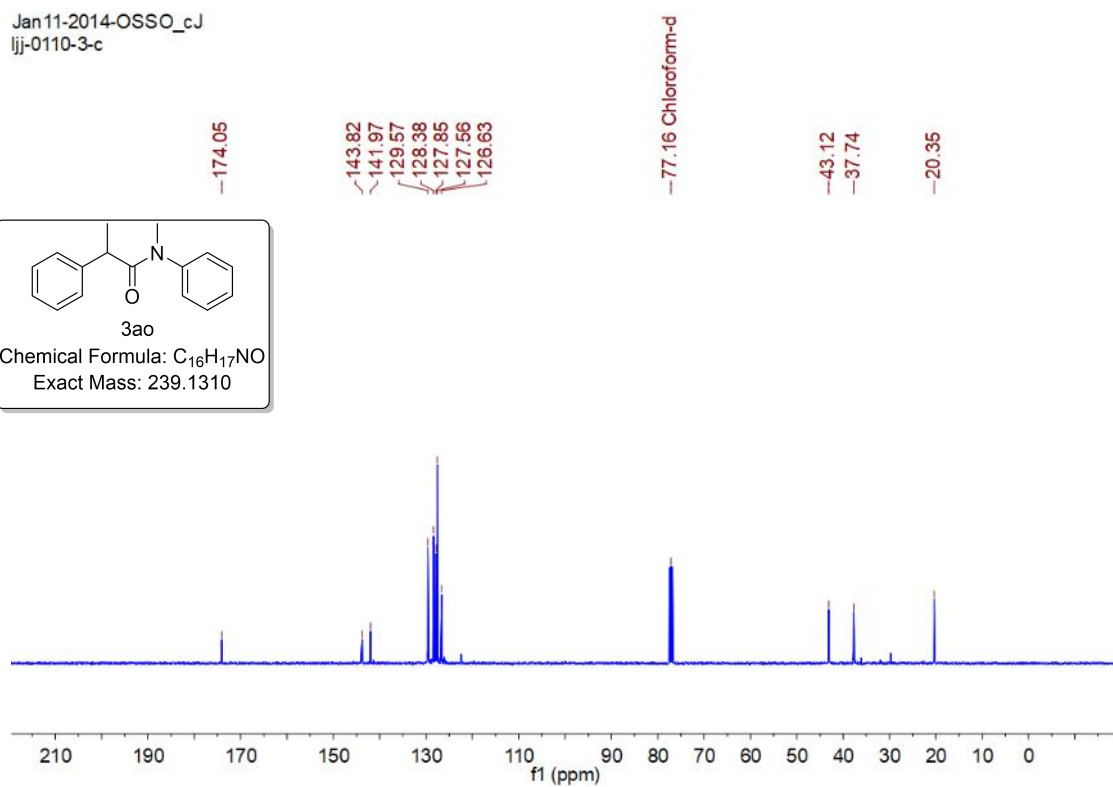
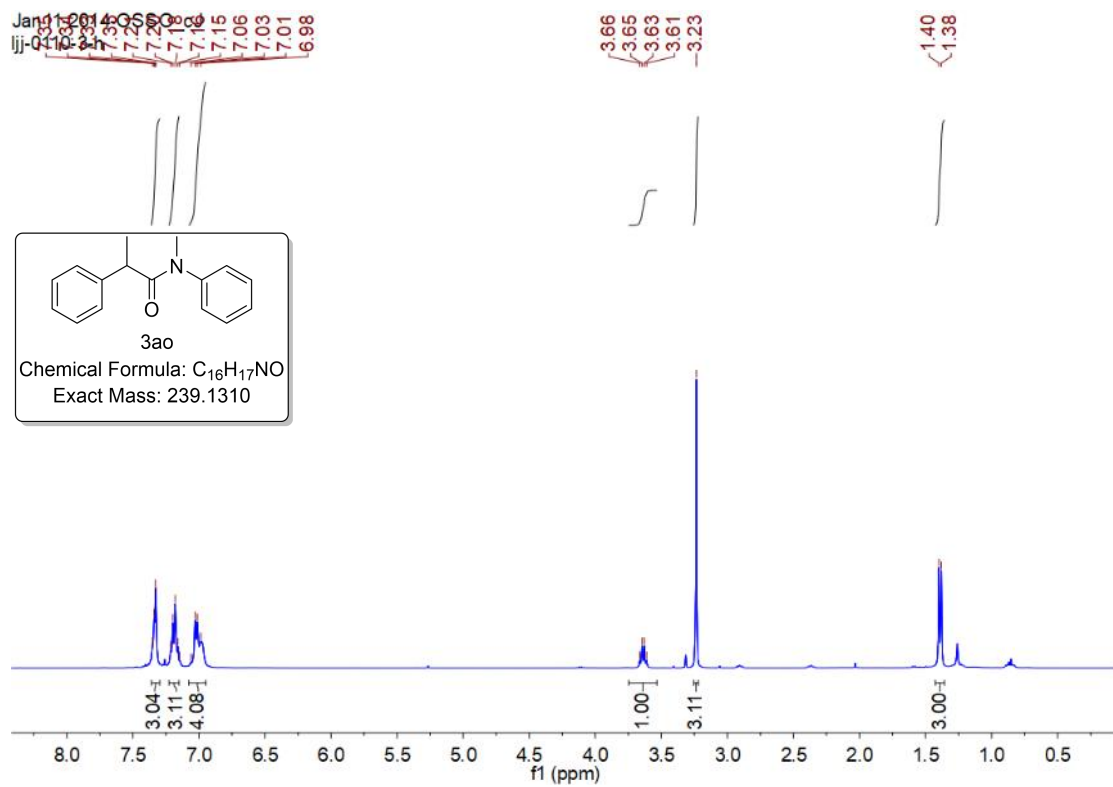


Dec26-2013-OSSO\_cj  
ljj-1226-1-h

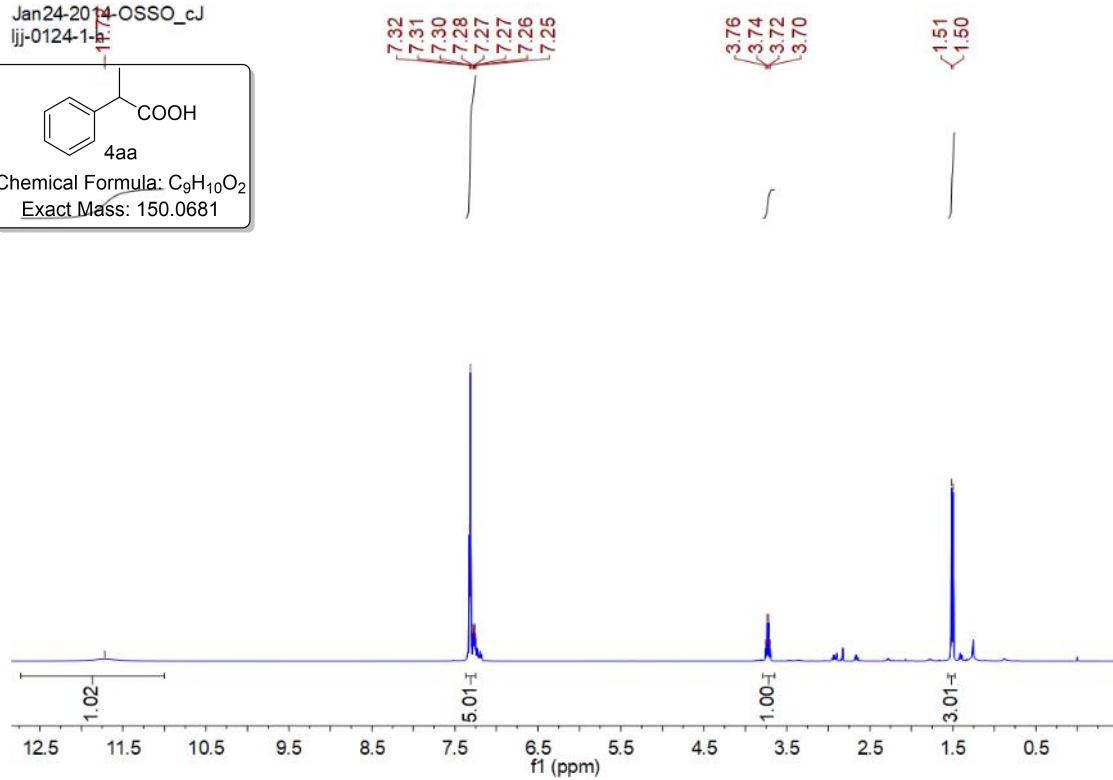
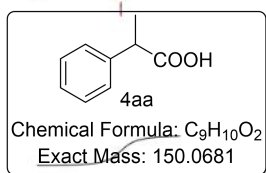


Dec26-2013-OSSO\_cj  
ljj-1226-1-c

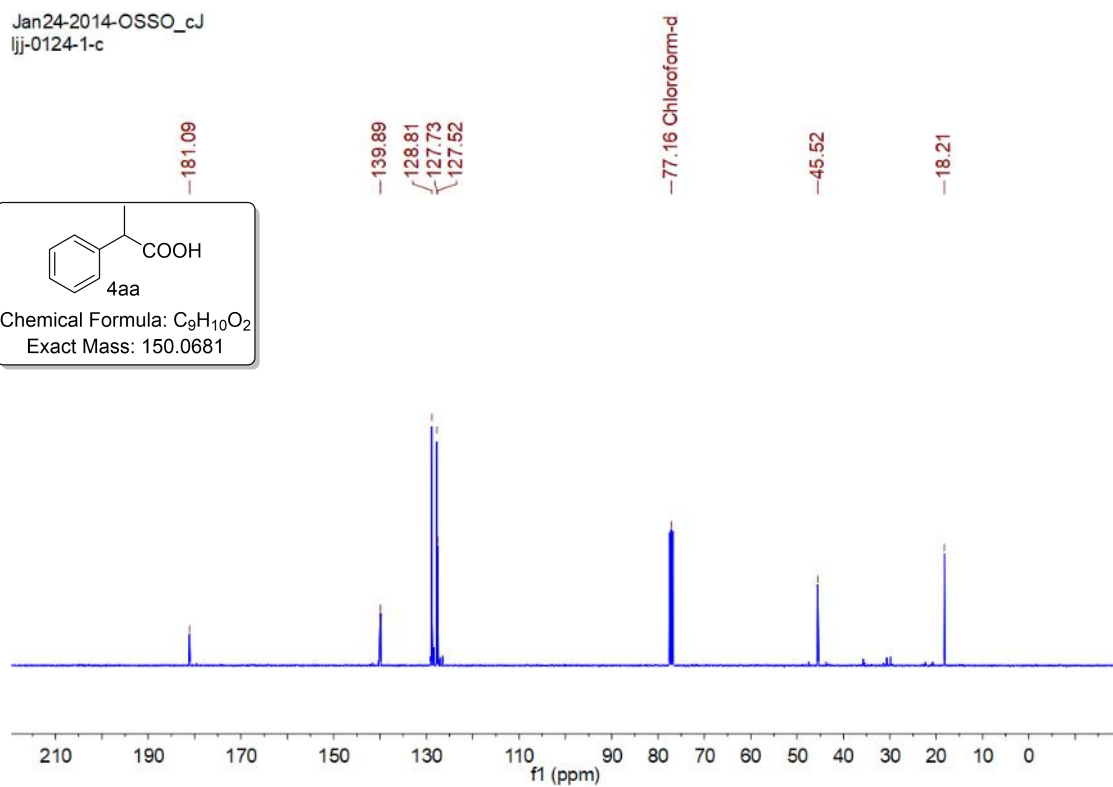
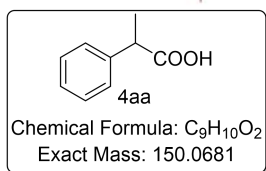




Jan24-2014 OSSO\_cj  
ljj-0124-1-b



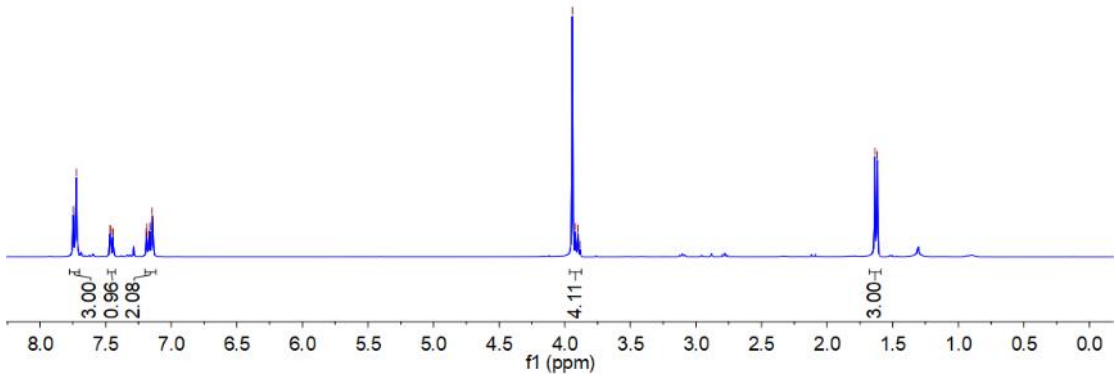
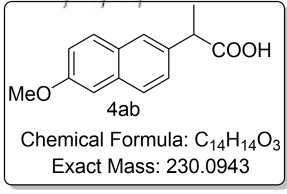
Jan24-2014-OSSO\_cj  
ljj-0124-1-c



May17-2014-OSSO\_cj  
ljj-20240515-h

3.94  
3.92  
3.90  
3.88

1.64  
1.62



May17-2014-OSSO\_cj  
ljj-20240515-c

-180.99

-157.84

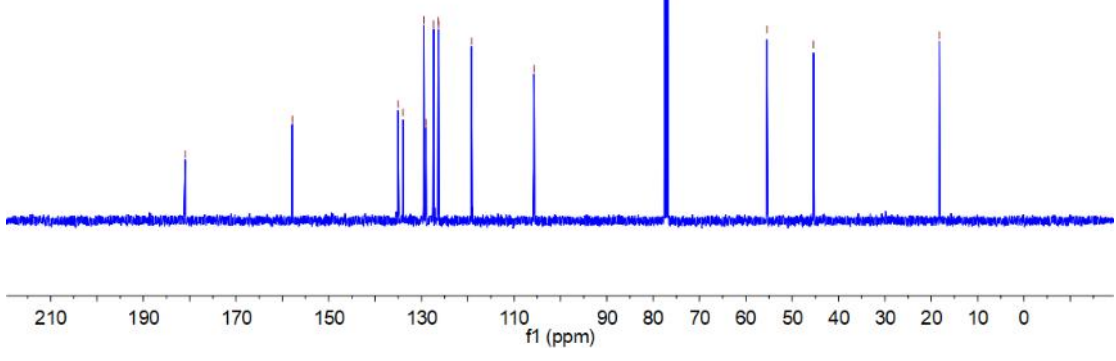
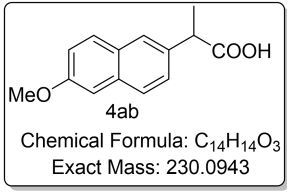
135.00  
133.96  
129.44  
129.03  
127.37  
126.33  
126.28  
119.17  
106.73

-77.16 Chloroform-d

-55.43

-45.41

-18.24



## 11. References

1. G. Ji, C. Li, D. Xiao, G. Wang, Z. Sun, M. Jiang, G. Hou, L. Yan and Y. Ding, *J. Mater. Chem. A*, 2021, **9**, 9165-9174.
2. K. Zhao, H. Wang, X. Wang, X. Cui and F. Shi, *Chem. Commun.*, 2022, **58**, 8093-8096.
3. X. Zhou, Z. Wang, B. Yu, S. Kuang, W. Sun and Y. Yang, *Green Chem.*, 2022, **24**, 4463-4469.
4. J. B. Pan, Z. C. Yang, X. G. Zhang, M. L. Li and Q. L. Zhou, *Angew. Chem., Int. Ed.*, 2023, **62**, e202308122.
5. S. Cuesta-Galisteo, J. Schörgenhumer, C. Hervieu and C. Nevado, *Angew. Chem., Int. Ed.*, 2024, **63**, e202313717.
6. S. Liu, H. Wang, X. Dai and F. Shi, *Green Chem.*, 2018, **20**, 3457-3462.
7. Y.-H. Yao, H.-Y. Yang, M. Chen, F. Wu, X.-X. Xu and Z.-H. Guan, *J. Am. Chem. Soc.*, 2020, **143**, 85-91.
8. H. Wang, H. Yuan, X. Wang, J. Zhao, D. Wei and F. Shi, *Adv. Synth. Catal.*, 2020, **362**, 2348-2353.