Catalyst speciation and deactivation in the ruthenium mediated transformation of ethynyl-β-Ionol to α,β-Unsaturated esters for vitamin A synthesis

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Supplementary Information

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Table of Contents

1.0 General information

1.1 Materials

Unless otherwise stated, all manipulations were carried out under an inert atmosphere of argon using classical Schlenk and glovebox (MBraun UNIlab plus) techniques. Prior to use, all glassware were oven dried to 120 °C. All reactions were performed with anhydrous and degassed non-deuterated solvents; DCM (dried over CaH₂), Methanol (dried over magnesium), n-hexane (dried over potassium), THF (dried over potassium), and toluene (dried over sodium) were freshly distilled before use. Acetone, acetonitrile, chloroform, and ethyl acetate were purchased from commercial suppliers (Acro seal) and were used without further purification. All solvents were passed through 0.2 μm syringe filters (VWR 514-0070) prior to use.

All chemicals used were obtained either from Sigma Aldrich, Fisher chemicals or DSM, at reagent grade or higher, and used as received unless otherwise specified.

1.2 Analytical

NMR spectra were recorded on a Bruker Ultrashield 500 MHz Avance IIIHD equipped with a nitrogen cooled BBO Prodigy CryoProbe, Bruker AVIII 500 MHz and Bruker AVIII 400 MHz (BBO probe) at 25 °C, unless otherwise specified. Multiplicity of signals in the NMR data are represented by their peaks: s is singlet, d is doublet, t is triplet, q is quartet, qu is quintet, m is multiplet, dd is doublet of doublets, br is broad, etc. Coupling constants (*J*) were reported to the nearest 0.1 Hz. All NMR spectra were processed with the Bruker software TopSpin 4.0.6 or DynamicCentre 2.5.6. Further NMR processing and data plotting was carried on Microsoft Excel and/or Origin 2017.

GC-FID/MS data was collected using a Shimadzu GCMS-QP2020 chromatograph equipped with a nitroterephthalic acid modified polyethylene glycol column of high polarity (DB-FFAP; 30 m x 0.25 mm x 0.25 μm) and a flame ionization detector (FID). Helium was used as a carrier gas with electron ionisation for mass spectrometry. Method details used are included when GC-FID/MS data is shown.

Column chromatographic separations were carried out on a BÜCHI Reveleris X2 Flash Chromatography System using a BÜCHI FlashPure Silica 24 g flash cartridge as a stationary phase silica. The mobile phase contained a gradient mixture of ethyl acetate and *n*-hexane.

IR spectrometric data was collected using a Bruker Alpha II FT-IR between $400 - 4,000$ cm⁻¹ with 24 scans at 2 cm⁻¹ resolution. Multiplicity of signals in the IR data are represented by their peaks: s is sharp, m is medium, br is broad, w is wide etc.

2.0 Experimental Details

2.1 Preparation of $[Ru''(\eta^3-CH_2C(Me)CH_2)_2(dppe)]$ ([(dppe)Ru(MA)₂])¹

Synthesis: A 250 mL round bottom flask was charged with 1,2-bis(diphenyl-phosphino)ethane (5.33 g, 13.38 mmol) and $[{\text{(methylally)}}_2{\text{Ru(cod)}}]$ (4.11 g, 12.78 mmol). The flask was equipped with a reflux condenser and n-hexane (56 mL) was added. The reaction mixture was heated to reflux for 6 h with stirring, cooled to room temperature and then stored at 4 °C for 19 h. The resulting yellow suspension was filtered and the solid washed with n-hexane (2 x 40 mL). The solid was dried for 2 h at 30 °C (10 mbar) and 2 h under high vacuum at room temperature to obtain 6.49 g yellow crystals (89% yield).

Purification: A Schlenk flask was charged with [Ru^{ll}(n³-CH₂C(Me)CH₂)₂(dppe)] (350 mg, 0.51 mmol) and dissolved in anhydrous acetone (35 mL). The yellow-grey solution was syringe-filtered through a 0.22 μm PTFE membrane (removing some fine, black solids) and then concentrated to half volume. *n*hexane was added (18 mL) and the mixture stored at 4 °C overnight. The mixture was filtered and the solid dried under high vacuum leaving bright yellow, free-flowing micro crystals (280 mg, yield = 80%).

¹H NMR (500 MHz, (CD₃)₂CO), δ (ppm) = 7.89 (t, ³J_{HH} = 8.1 Hz, 4H, Ph_{ortho}), 7.45 (m, 6H, Ph_{meta and para),} 7.14 (m, 6H, Ph_{meta and para), 6.80 (t, ³J_{HH} = 7.3 Hz, 4H, Ph_{ortho}), 3.57 (dd, ³J_{HH} = 8.6, 35.7 Hz, 2H, CH₂C**H**₂),} 2.66 (br, 2H, C**H2**CH2), 2.05 (s, 6H, allylic (C**H3**)**2**), 1.71 (s, 2H, allylic C**H2**), 1.37 (s, 2H, allylic C**H2**), 0.83 (dd, ³/_{HH} = 4.9, 14.78 Hz, 2H, allylic CH₂), 0.00 (d, ³/_{HH} = 14.6 Hz, 2H, allylic CH₂).

 $31P{1H}$ NMR (202 MHz, (CD₃)₂CO), δ (ppm) = 83.9 (s).

Figure S1. ¹H NMR of $[Ru''/\eta^3$ -CH₂C(Me)CH₂)₂(dppe)] in (CD₃)₂CO.

Figure S3. ¹H-³¹P HMBC NMR of [RuII(η³ -CH2C(Me)CH2)2(dppe)] in (CD3)2CO.

2.2 Preparation of $[Ru''(\eta^3-CH_2C(Me)CH_2)_2(dppm)]^1$

A flame dried Schlenk flask was charged with bis(diphenylphosphino)methane (821 mg, 0.7 mmol), [(methylallyl)₂Ru(COD)] (399 mg, 1.25 mmol) and *n*-hexane (15.6 mL). The mixture was heated to reflux for 6 h with stirring, then cooled to room temperature and stored at 4 °C overnight. The resulting suspension was cannula filtered under argon, and the yellow solid was washed with cold *n*-hexane (2 x 5 mL). The residue was dried under high vacuum overnight giving free-flowing dark yellow micro crystals (yield = 368 mg, 92%).

¹H NMR (500 MHz, (CD₃)₂CO), *δ* (ppm) = 8.01 (t, ³J_{HH} = 8.1 Hz, 4H, Ph_{ortho}), 7.53 (m, ³J_{HH} = 7.2, 6H, Ph_{meta} and para), 7.16 (m, 6H, Ph_{meta and para}), 6.88 (t, ³J_{HH} = 7.3 Hz, 4H, Ph_{ortho}), 4.94 (t, ³J_{HH} = 9.5 Hz, 2H, PC**H**₂P), 2.65 (dd, 2H, allylic CH₂), 1.89 (s, 6H, allylic CH₃), 1.25 (s, 2H, allylic CH₂), 0.74 (d, ³J_{HH} = 10.5 Hz, 2H, allylic CH₂), 0.64 (br, 2H, allylic CH₂).

³¹P{¹H} NMR (202 MHz, (CD₃)₂CO), δ (ppm) = 12.7 (s).

2.2 Purification of carboxylic acids

2.2.1 Pivalic acid

A Schlenk flask was charged with pivalic acid (30 g, 0.3 mol) and attached in a bulb-to-bulb distillation apparatus. The flask with crude pivalic acid was heated to 180 °C with an oil bath and the receiving flask was cooled to 0 °C. The head temperature of the distillation was 175 °C. The first 15% of distillate was discarded. Clear viscous liquid was distilled over 2 h into a Schlenk flask (yield = 24 g, 80%).

2.2.2 Adamantane carboxylic acid and benzoic acid

A Schlenk flask was charged with adamantane carboxylic acid (30 g, 0.167 mol) and recrystallised from anhydrous benzene (100 mL). The crystals were filtered and dried overnight under dynamic vacuum, with free-flowing white micro crystals isolated (yield = 24.2 g, 82%).

A Schlenk flask was charged with benzoic acid (30 g, 0.25 mol) and recrystalised from anhydrous ethanol. The crystals were filtered and dried overnight under dynamic vacuum, with free-flowing white micro crystals isolated (yield = 21.6 g, 72%).

2.3 Preparation of ethynyl-β-ionol $(2a)^2$

Ethynol-β-ionol was produced by DSM using a proprietary method. Crude ethynyl-β-ionol (30 g, 0.14 mol) and butylated hydroxytoluene (stabiliser; 150 mg, 0.68 mmol) were transferred into separate round bottom flasks in a Vigreux distillation apparatus. The round bottom flask with crude ethynyl-βionol was heated (120 °C / 0.5 mbar) with an oil bath. The receiving round bottom flask containing butylated hydroxytoluene (BHT) was cooled to 0 °C. The head temperature of the distillation was 122 °C at 0.4 mbar. Clear viscous liquid was distilled over the duration of a couple of hours. (Yield = 24 g, 80%).

¹H NMR (500 MHz, (CD₃)₂CO), δ (ppm) = 6.39 (d, 1H, ³J_{HH} = 16.2 Hz, H-7), 5.56 (d, 1H, ³J_{HH} = 15.9 Hz, H-6), 4.57 (s, 1H, H-5), 2.92 (s, 1H, H-1), 2.00 (t, 2H, ³J_{HH} = 6.3 Hz, H-12), 1.68 (s, 3H, H-14), 1.63 (m, 2H, H-11), 1.54 (s, 3H, H-4), 1.47 (m, 2H, H-10), 1.01 (s, 6H, H-15-16)

¹³C{¹H} NMR (125 MHz, (CD₃)₂CO), δ(ppm) = 205.5 (s, C-3), 138.3 (s, C-6), 136.5 (s, C-8), 128.1 (s, C-13), 125.7 (s, C-7), 87.7 (s, C-9), 72.3 (s, C-1), 66.9 (s, C-2), 39.3 (s, C-10), 32.4 (s, C -12), 30.7 (s, C-4), 28.2 (d, *J* = 4.08 Hz, C-15-16), 20.7 (s, C-14), 19.12 (s, C-11).

Figure S6. ¹H NMR of ethynyl-β-ionol in (CD3)2CO.

Figure S7. ¹³C{¹H} NMR of ethynyl-β-ionol in (CD3)2CO.

2.4 Preparation of α,β-unsaturated carboxylates

2.4.1 3-hydroxy-3-methyl-5-(2,6,6-trimethylcyclohex-1-en-1-yl) penta-1,4-dien-1-yl pivalate (3ab)

Procedure adapted from literature for the isolation of the ester-adducts. ² A Schlenk flask was charged with pivalic acid (1.32 g, 11 mmol), pivalic anhydride (0.4 ml, 2 mmol), ethynyl-β-ionol (2.18 g, 10 mmol), and ethyl acetate (10 mL). Subsequently, $[Ru''(\eta^3-CH_2C(Me)CH_2)_2(dppe)]$ (61 mg, 0.1 mmol) was dissolved in ethyl acetate (5 mL) and syringed into the Schlenk flask and stirred for 40 hours for maximum conversion. The residue was purified by flash chromatography on a silica packed column with ethyl acetate as the eluent to give the product over several fractions. Fractions were poured into one Schlenk flask and concentrated under pressure to a brown oil (2.36 g, 72% yield).

¹H NMR (500 MHz, (CD₃)₂CO), δ (ppm) = 6.95 (d, ³J_{HH} = 7.2 Hz, 1H, H-7), 6.12 (d, ³J_{HH} = 16.2 Hz, 1H, H-11), 5.72 (d, 3 *J*_{HH} = 16.2 Hz, 1H, H-10), 5.19 (d, 3 *J*_{H-H} = 8 Hz, 1H, H-8), 1.96 (m, 2H, H-15), 1.65, (s, 3H, H-3), 1.61 (m, 2H, H-16), 1.58 (s, 3H, H-4), 1.44 (m, 2H, H-17), 1.25 (s, 9H, H-1), 0.94 (d, ³J_{HH} = 6.3 Hz, 6H, H-2)

Figure S8. ¹H NMR of 3ab in (CD₃)₂CO.

2.4.2 3-hydroxy-3-methyl-5-(2,6,6-trimethylcyclohex-1-en-1-yl)penta-1,4-dien-1-yl adamantane-1-carboxylate (3ae)

A Schlenk flask was charged with adamantane carboxylic acid (1.98 g, 11 mmol), pivalic anhydride (0.4 ml, 2 mmol), ethynyl-β-ionol (2.18 g, 10 mmol), and ethyl acetate (10 mL). Subsequently, [Ru^{||}(η³- $CH₂C(Me)CH₂$ ₂(dppe)] (61 mg, 0.1 mmol) was dissolved in ethyl acetate (5 mL) and syringed into the Schlenk flask and stirred for 40 hours for maximum conversion. The residue was purified by flash chromatography on a silica packed column with ethyl acetate as the eluent to give the product over several fractions. Fractions were poured into one Schlenk flask and concentrated under pressure to a brown oil (68% yield).

¹H NMR (500 MHz, CDCl₃), δ (ppm) = 7.09 (d, ³J_{HH} = 7.2 Hz, 1H, H-1), 6.11 (d, ³J_{HH} = 16.2 Hz, 1H, H-4), 5.65 (d, ³J_{HH} = 16.2 Hz, 1H, H-3), 5.08 (d, ³J_{HH} = 7.2 Hz, 1H, H-2), 2.04 (s, 3H, H-16), 2.00, (s, 1H, H-18), 1.97 (m, 2H, H-15), 1.92 (m, 10H, H-17), 1.65 (s, 3H, H-6), 1.6 (m, 2H, H-14), 1.53 (s, 3H, H-8), 1.45 (m, 2H, H-13), 0.98 (d, 6H, H-7)

Figure S9. ¹H NMR of 3ae in CDCl₃ containing methyl tert-butyl ether (MTBE) and EtOAc.

2.5 Synthetic procedure for generating compounds suitable for crystal structure determination

2.5.1 A_p' [(dppe)Ru^{ll}(η²-O₂CC^tBu)(η¹-O₂CC^tBu)(^tBuCCO₂H)]

A Schlenk flask was charged with ~100 eq. of pivalic acid (1 g, 9.8 mmol) and [Ru^{||}(n³-CH₂C(Me)CH₂)₂(dppe)] (60 mg, 0.1 mmol), which were dissolved in anhydrous ethyl acetate (15 mL) and stirred for 1 hour. The solvent was concentrated at 40 °C for 2 hours. The remaining solid was dissolved into dry n-hexane at first signs of boiling and allowed to cool gradually from +60 °C to -20 °C for the formation of yellow microcrystalline crystals (yield = 32 mg, 48.4%).

¹H NMR (500 MHz, (CD₃)₂CO), *δ* (ppm) = 7.57 (br, 8H, Ph), 7.36 (m, 12H, Ph), 2.72 (s, 4H, CH₂-CH₂), 0.58 $(s, 18H, (CH₃)₃)₂)$

 $31P{^1H}$ NMR (202 MHz, (CD₃)₂CO), *δ* (ppm) = 89.5 (s)

¹³C{¹H} NMR (125 MHz, (CD₃)₂CO), *δ* (ppm) = 193.4 (s, COO), 136.7 (s, Ph), 133.5 (t, *J*_{CH} = 4.7 Hz, Ph), 130.3 (s, Ph), 128.7 (t, *J*_{CH} = 4.7 Hz, Ph), 39.8 (s, CH₂-CH₂), 38.9 (s, CH₂-CH₂), 27.6 (s), 27.14(s, (CH₃)₃)₂)

Figure S10. ¹H NMR of [(dppe)Ru^{||}(η^2 -O2CC^tBu)(η^1 -O2CC^tBu)(^tBuCCO₂H)] (A_P) crystals dissolved in (CD₃)2CO

Figure S11. ³¹P{¹H} NMR of [(dppe)Ru^{||}(η ²-O2CC^tBu)(η ¹-O2CC^tBu)(^tBuCCO2H)] (A_P) crystals dissolved in (CD3)2CO

Figure S12. ¹³C(¹H) NMR of [(dppe)Ru^{||}(η ²-O₂CC^tBu)(η ¹-O₂CC^tBu)(^tBuCCO₂H)] crystals dissolved in (CD₃)₂CO.

2.5.2 A_{ad}' [(dppe)Ru^{||}(η²-O₂CC(CH₂)₆(CH)₃C)(η¹-O₂CC(CH₂)₆(CH)₃C)((CH)₃(CH₂)₆CCCO₂H)]

A Schlenk flask was charged with adamantane carboxylic acid (0.25 g, 1.4 mmol) and $\left[\text{Ru}^{\text{II}}(\eta^3-\eta)\right]$ CH₂C(Me)CH₂)₂(dppe)] (60 mg, 0.1 mmol) which were dissolved in anhydrous acetone (10 mL). After 0.5 h the solvent was evaporated for 2 hours (40 °C). The remaining solid was dissolved in a supersaturated solution with anhydrous acetone and set up forslow diffusion with *n*-hexane to obtain sharp yellow crystals in the glovebox (yield = 28 mg, 45%).

¹H NMR (500 MHz, (CDCl₃), δ (ppm) = 7.50 (br, 8H, Ph), 7.31 (m, 12H, Ph), 2.63 (br, 4H, CH₂-CH₂), 1.71 (br, 6H, (CH)₆), 1.50 (d, J_{HH} = 11.93 Hz, 6H, (CH₂)₃), 1.41 (d, J_{HH} = 11.93 Hz, 6H, (CH₂)₃), 1.23 (s, 12H, $(CH_2)_6$).

 $31P{1H}$ NMR (202 MHz, CDCl₃), *δ* (ppm) = 90.7 (s)

Figure S13. ¹H NMR of [(dppe)Ru^{||}(η^2 -O2CC(CH2)6(CH)3C)(η^1 -O2CC(CH2)6(CH)3C)((CH)3(CH2)6CCCO2H)] (A'ad) crystals *dissolved in CDCl3.*

Figure S14. ³¹P{¹H} NMR of [(dppe)Ru^{||}(η ²-O₂CC(CH₂)₆(CH)₃C)(η ¹-O₂CC(CH₂)₆(CH)₃C)((CH)₃(CH₂)₆CCCO₂H)] (**A'**_{ad}) *crystals dissolved in CDCl3*

2.5.3 **B_p'** [{(dppe)Ru^{||}(η¹-O₂CC^tBu)}₂(μ-O₂CC^tBu)₂(μ-H₂O)]

A Schlenk flask was charged with pivalic acid (0.5 g, 4.9 mmol) and [Ru^{II}(n³-CH₂C(Me)CH₂)₂(dppe)] (60 mg, 0.1 mmol) which were dissolved in anhydrous ethyl acetate (10 mL). After 0.5 h the solvent was evaporated for 2 hours (40 °C). The remaining solid was dissolved in $(CD₃)₂CO$ and set up for slow diffusion with *n*-hexane to obtain sharp yellow crystal formation in the glovebox (yield = 0.33 mg, 26.6%).

¹H NMR (500 MHz, (CD₃)₂CO), *δ* (ppm) = 7.58 (br, 8H, Ph), 7.37 (m, 12H, Ph), 2.86 (br, 4H, CH₂-CH₂), 0.7 (s, 18H, (C**H3**)**3**)**2**)

 $31P{1H}$ NMR (202 MHz, (CD₃)₂CO), δ (ppm) = 89.5 (s), 29.3 (s)

¹³C{¹H} NMR (125 MHz, (CD₃)₂CO), δ (ppm) = 205.22 (s), 132.7 (t, *J*_{CH} = 5.1 Hz), 129.49 (s), 127.9 (t, *J*_{CH} = 5.1 Hz), 26.62 (s), 26.1 (s)

Figure S17. ¹³C{¹H} NMR of B_p' [{(dppe)Ru^{||}(η ¹-O₂CC^tBu)}₂(μ -O₂CC^tBu)₂(μ -H₂O)] dissolved in (CD₃)₂CO.

2.5.4 **B_{ad}'** [{(dppe)Ru^{||}(η¹-O₂CC(CH₂)₆(CH)₃)}₂(μ-O₂CC(CH₂)₆(CH)₃)₂(μ-H₂O)]

A Schlenk flask was charged with adamantane carboxylic acid (0.25 g, 1.4 mmol) and $\left[\text{Ru}^{\text{II}}(\eta^3-\eta)\right]$ CH2C(Me)CH2)2(dppe)] (60 mg, 0.1 mmol) which were dissolved in anhydrous acetone (10 mL). After 0.5 h the solvent was evaporated for 2 hours (40 °C). The remaining solid was dissolved in a supersaturated solution with anhydrous toluene and slow diffusion of *n*-hexane were set up for yellow sharp crystal formation in the glovebox. The crystals were collected and washed with cold hexane before being concentrated. The residue was in CDCl₃ for analysis.

¹H NMR (500 MHz, (CD₃)₂CO, 298 K) *δ* (ppm) = 7.50 (br, 8H, Ph), 7.31 (m, 12H, Ph), 2.63 (br, 4H, CH₂-CH2), 1.73 (br, 6H, (C**H**)**6**), 1.51(d, *JH-H* = 11.93 Hz, 6H, (C**H2**)**3**), 1.41 (d, *JH-H* = 11.93 Hz, 6H, (C**H2**)**3**), 1.23 (s, 12H, (C**H2**)**6**) ³¹P{¹H} NMR (202 MHz, (CD₃)₂CO, 298 K) δ (ppm) = 90.4 (s)

Figure S19. ³¹P{¹H} NMR of B_{ad}' [{(dppe)Ru^{||}(n¹-O2CC(CH2)6(CH)3)}2(µ-O2CC(CH2)6(CH)3)2(µ-H2O)] dissolved in *CDCl3.*

2.5.5 B_{tf}' [{(dppe)Ru^{||}(η ¹-O₂CCF₃}₂(μ -O₂CCF₃)₂(μ -OH₂)]

A Schlenk flask was charged with trifluoracetic acid carboxylic acid (0.55 mL, 0.5 mmol) and [Ru^{ll}(n³-CH2C(Me)CH2)2(dppe)] (30 mg, 0.05 mmol) which were dissolved in anhydrous acetone (10 mL). After 0.5 h the solvent was evaporated for 2 hours (40 °C). The remaining solid was dissolved in a supersaturated solution with anhydrous acetone and slow diffusion of *n*-hexane were set up for yellow sharp crystal formation in the glovebox.

2.5.6 **B_b'** [{(dppe)Ru^{||}(η¹-O₂CPh}₂(μ-O₂CPh)₂(μ-H₂O)]

A Schlenk flask was charged with benzoic acid (0.25 g, 2.1 mmol) and [Ru^{ll}(η^3 -CH₂C(Me)CH₂)₂(dppe)] (60 mg, 0.1 mmol) which were dissolved in anhydrous acetone (10 mL). After 0.5 h the solvent was evaporated for 2 hours (40 °C). The remaining solid was dissolved to make a supersaturated solution with anhydrous acetone and set up for slow diffusion with *n*-hexane in the glovebox. This resulted in the formation of yellow crystals.

¹H NMR (500 MHz, (CD₃)₂CO), δ (ppm) = 8.07 (d, *J*_{HH} = 8.0 Hz 1H, Ph), 7.61 (d, *J*_{HH} = 7.4 Hz, 3H, Ph), 7.52 (t, J_{HH} = 7.4 Hz and 15.2 Hz, 1H, Ph), 7.4 (t, J_{HH} = 7.4 Hz and 14.9 Hz, 2H, Ph), 7.26 (m, 3H, Ph), 7.64 (br, 8H, Ph), 7.27 (m, 12H, Ph), 2.74 (m, 4H, CH₂CH₂) $31P{1H}$ NMR (202 MHz, (CD₃)₂CO), δ (ppm) = 89.9 (s), 78.2 (s)

Figure S20. ¹H NMR of B_b' [{(dppe)Ru^{||}(n¹-O2CPh}2(µ-O2CPh)2(µ-H2O)] dissolved in (CD3)2CO showing a mixture of *A^b and B^b shifts.*

Figure S21. ³¹P{¹H} NMR of **B_b'** [{(dppe)Ru^{||}(η ¹-O₂CPh}₂(µ-O₂CPh)₂(µ-H₂O)] dissolved in (CD₃)₂CO.

Figure S22. ¹H-³¹P HMBC NMR of a mixture of $[Ru''(\eta^3 - CH_2C(Me)CH_2)_2(dppe)]$ A_b and B_b in EtOAc

2.5.7 A_b [(dppe)Ru^{||}(η¹-O₂CPh)₂]

The formation of B_b was found to form due to the presence of moisture in the system. Here an experiment was performed with all reagents prepared as anhydrously as possible to yield only A_b . A Schlenk flask was charged with anhydrous benzoic acid (20 mg, 0.16 mmol) and [Ru^{ll}(n³- $CH_2C(Me)CH_2)_2(dppe)$] (30 mg, 0.05 mmol) which were dissolved in anhydrous (CD₃)₂CO (1 mL).

¹H NMR (500 MHz, (CD₃)₂CO), *δ* (ppm) = 7.64 (br, 8H, Ph), 7.61 (d, J_{HH} = 7.4 Hz, 4H, Ph), 7.38 (t, J_{HH} = 7.4 Hz, 2H, Ph), 7.27 (m, 12H, Ph), 7.24 (m, 4H, Ph), 2.76 (m, 4H, J_{HH} = 14.3 Hz, CH₂CH₂) $31P{1H}$ NMR (202 MHz, (CD₃)₂CO), *δ* (ppm) = 89.9 (s)

Figure S23. ¹H NMR of A_b [(dppe)Ru^{||}(η ¹-O₂CPh}₂] in (CD₃)₂CO without the presence of moisture and the *formation of isobutene in (CD3)2CO.*

Figure S24. ³¹P{¹H} NMR of A_b [(dppe)Ru^{||}(n^1 -O₂CPh}₂] in (CD₃)₂CO.

2.5.8 **X²_b•H₂O**</sub> [(dppe)Ru^{||}(η¹-*O*₂*CPh*)₂(CO)(H₂O)]

A Schlenk flask was charged with benzoic acid (1.81 g, 15 mmol), ethynyl-β-ionol (2.18 g, 10 mmol), tri-o-tolyl phosphate (461 mg, 1.25 mmol) (internal standard) and anhydrous acetone (10 mL). In a second flask, [Ru^{ll}(η³-CH₂C(Me)CH₂)₂(dppe)] (61 mg, 0.1 mmol, 1 mol%) was dissolved in anhydrous acetone (5 mL) and syringed into the first Schlenk flask to trigger the reaction. After reaction completion a ${}^{31}P{^{1}H}$ NMR was recorded (Figure S25) and the sample was then concentrated and layered with *n*-hexane (20 mL). Yellow crystals were observed at the bottom of the vial.

 $31P{^1H}$ NMR (202 MHz, (CH₃)₂CO), *δ* (ppm) = 53.3

Figure S25. ${}^{31}P_1{}^{1}H$ } NMR of X^2b , obtained after the reaction of ethynyl-6-ionol (0.66 M) with benzoic acid (1 M) *catalysed by [RuII(η³ -CH2C(Me)CH2)2(dppe)] (0.5mol%) in acetone.*
2.5.9 X^2 _b [(dppe)Ru^{||}(η¹-O₂CPh)₂(CO)₂]

A Schlenk flask was charged with benzoic acid (1.81 g, 15 mmol), [Ru^{||}(n³-CH₂C(Me)CH₂)₂(dppe)] (61 mg, 0.1 mmol, and anhydrous acetone (10 mL) under inert conditions and stirred for 3 hours producing a 50:50 mixture of [(dppe)Ru^{||}(η²-OOCPh)₂] and [(dppe)Ru^{||}(η²-OOCPh)(η¹-OOCPh)(H₂O)] in solution. CO was bubbled in the reaction flask *via* PEEK tubing for 10 seconds. The flask was left overnight, and the solution slowly changed from yellow to white with some precipitate visible. The solution was concentrated under vacuum and washed with cold hexane (2 x \approx 5 mL). The white crystals were redissolved in acetone and layered with *n*-hexane (20 mL). White crystals were observed at the bottom of the vial. The crystals were separated and redissolved in CDCl₃ for characterisation.

¹H NMR (500 MHz, (CDCl3), *δ* (ppm) = 7.5 (m, 8H, Ph), 7.2 (m, 4H, Ph), 7.2 (m, 6H, Phacid), 7.14 (m, 8H, Ph), 7.04 (m, 4H, Ph_{acid}), 3.03 (d, J_{PH} = 17.8 Hz, 4H)

 $31P{1H}$ NMR (202 MHz, CDCl₃), *δ* (ppm) = 52.0 (s)

 $31P{^1H}$ NMR (202 MHz, (CD₃)₂CO), *δ* (ppm) = 53.7 (s)

¹³C{¹H} NMR (125 MHz, (CD₃)₂CO, 298 K) δ (ppm) = 195.7 (dd, J_{CH} = 106.7 Hz, 18.9, Hz, C=O), 172.2 (s, COO), 134.1 (s, Ph_{acid}), 132.4 (t, *J*_{CH} = 5.4 Hz, Ph), 130.8 (s, Ph_{acid}), 129. 9 (s, Ph), 129.3 (s, Ph), 129.0 (t, *J*_{CH} = 5.4 Hz, Ph), 128.5 (s, Ph_{acid}), 127.0 (s, Ph_{acid}), 25.8 (dd, *J*_{CH} = 22.3 Hz, 20.0 Hz, (CH₂)₂)

T-IR (cm-1): 3061 (w) ν CH-benzoic, 2043 (s) ν C=O, 1996 (s) ν C=O, 1611 (s) ν CO-benzoic, 1433 (w) ν Ph(P–Ph), 1333 (s) ν C-O-benzoic (coordinated), 1102 (w) δ (C–CH in the plane), 742 (m) δ (C–C out of the plane), 688 (s) δ (C–C in the plane), 521 (s) ν Ru–P

Figure S26. ¹H NMR spectrum of the isolated [(dppe)Ru^{||}(CO)₂(η ¹-O₂CPh)₂] (**X²b**) in CDCl₃.

S39

Figure S29. 1H-¹³C HMBC NMR of [(dppe)RuII(CO)2(η¹ -O2CPh)2] in CDCl3.

Figure S30. FTIR ATR dry spectrum of the isolated [(dppe)RuII(CO)2(η¹ -O2CPh)2, recorded under Ar atmosphere.

The IR spectrum of *[(dppe)RuII(CO)2(η¹ -O2CPh)2] shows two distinct shifts at* 2043 cm-1 and 1996 cm-1 corresponding to the terminal carbonyl stretching modes of the Z-carbonyl structure.^{3,4}

2.6 Standard procedure for the ruthenium-mediated transformation of ethynyl-βionol into the α,β-unsaturated ester adduct

A Schlenk flask was charged with pivalic acid (1.32 g, 11 mmol), pivalic anhydride (0.4 mL, 2 mmol), ethynyl-β-ionol (2.18 g, 10 mmol), 1,3,5-trimethoxybenzene (420 mg, 2.5 mmol) and anhydrous acetone (10 mL). In a second flask, [Ru^{ll}(n³-CH₂C(Me)CH₂)₂(dppe)] (61 mg, 0.1 mmol, 1 mol%) was dissolved in anhydrous acetone (5 mL) and syringed into the first Schlenk flask to start the reaction. The reaction was periodically monitored with quantitative 1 H NMR and ${}^{31}P{}^{1}H$ } NMR spectroscopy by integration relative to the internal standard 1,3,5-trimethoxybenzene.

Figure S31. Exemplary ¹H NMR spectrum recorded during a Ru-mediated transformation of ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) catalysed by [(dppe)Ru(MA)2] in anhydrous acetone (15 mL) at 20 °C. Product yields obtained from quantitative ex-situ ¹H NMR spectroscopy against trimethoxybenzene (167 mM) as internal standard. Spectrum taken after 12 hours of reaction initiation, intentionally showing both substrate and product peaks.

Figure S32. ¹H NMR spectra showing the Ru-mediated *transformation of ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) into 3ab catalysed by [(dppe)Ru(MA)2] in anhydrous acetone (15 mL) at 20 °C from quantitative ¹H FlowNMR monitoring, at 4 mL/min, at two different time points, red: start of the reaction (5 minutes), blue: after 12 hours. The purpose is to clearly provide an example to visually illustrate the best peaks to use for quantification of all FlowNMR data.*

Compound	Peak name	$F2$ (ppm)	$T_1(s)$
Ethynyl-β-ionol	$H - 7$	6.39	3.7(3)
Ethynyl-β-ionol	$H-1$	2.84	8.9(9)
3ab	$H - 7$	6.95	4.6(3)
3ab	H-8	5.19	4.2(4)
Internal standard	TMB (aliphatic)	3.75	2.4(1)
Internal Standard	TMB (aromatic)	6.03	6.5(10)

Table S1. T¹ times for all peaks of interest in the catalytic transformation of ethynyl*-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) into 3ab catalysed by [(dppe)Ru(MA)2] in anhydrous acetone, determined via inversion recovery NMR experiment.*

2.7 FlowNMR Apparatus

The FlowNMR apparatus (Figure S33) consisted of 3 main components: the reaction vessel (highlighted in red by heat exchanger 3), the flow path (highlighted in yellow by heat exchanger 1) and the flow tube (highlighted in blue by heat exchanger 2). The flow path was composed of PEEK tubing (PEEK, OD 1/16", ID 0.76 mm, Upchurch Scientific) which connected the reaction vessel to the UV-Visible flow cell (PEEK SMA-Z, Ocean Optics) and InsightMR flow tube (Bruker). The InsightMR flow tube consisted of PEEK (OD 1/32", ID 0.51 mm, Upchurch Scientific) transfer lines housed within concentrically aligned Teflon tubing that connects to the flow tip (Figure S33). A Vapourtec SF-10 peristaltic pump was connected to the FlowNMR apparatus *via* PEEK tubing (O.D 1/16", I.D 0.76 mm, Upchurch Scientific). Connections to reaction vessels were secured *via* rubber septa, HPLC-type (1/16") or Swagelok (1/16") fittings. Thermal regulation of the flow path and flow tube was maintained by 2 heat exchangers (Julabo CORIO CD-300F, thermofluid 50:50 ethylene glycol/water). Thermal regulation of the reaction vessel was controlled by either a heat exchanger (Julabo CORIO CD-300F, thermofluid 50:50 ethylene glycol/water) connected to a reactor jacket (DrySyn Snowstorm ONE) or a hotplate (IKA RCT B S002) with a reactor jacket (paraffin oil bath).

Figure S33. Piping and instrumentation schematic of the closed-loop recirculating FlowNMR setup, with exemplary length scales, tubing ID (indicated by the line thickness), and showing sequential temperature control by use of independent heat exchangers (colour coded for clarity).

2.7.1 Data acquisition

For all FlowNMR reactions, non-deuterated solvents were used, and the frequency lock was switched off. Automated shimming and tuning were performed whilst the flow was turned off. Accurate quantification of spectra acquired during flow is complicated by flow effects which effect the observed relaxation times and the equilibrium magnetisation of the sample. To account for these flow effects, a correction factor was needed for each peak of interest in the spectra that were quantified, as has been described previously.⁵ Briefly, spectra of starting materials, products, internal standards and (where possible) intermediates were recorded under static conditions (0 mL min⁻¹) with a long relaxation delay (60 seconds) to ensure they were fully quantitative. A spectrum was also recorded with flowrate of the reaction (4 mL min⁻¹) to calculate the comparison of the integral area of peaks. For each peak of interest, a correction factor (CF) could be calculated (Equation 1; *I* = peak integral), which could then be applied to subsequent spectra acquired under flowing conditions (Equation 2).

Equation 1:
$$
CF = \frac{I_{Static}}{I_{Flow}}
$$

Equation 2: $I_{\text{Corrected}} = \text{CF} \times I$

Data acquisition was started using InsightMR software (Bruker) queuing up interleaved ¹H NMR and $31P{1}$ H} NMR experiments every 4 minutes with the following acquisition parameters:

¹H NMR (Number of scans (NS) = 8, Receiver gain (RG) = 4, Dummy scans (DS) = 0 s, Acquisition time $(AQ) = 1.6$ s, Relaxation delay $(D_1) = 1$ s)

 $^{31}P{^1H}$ NMR (NS = 48, RG = 203, DS= 0 s, AQ = 0.4, D₁ = 0.5 s)

2.7.2 FlowNMR reaction setup

The FlowNMR apparatus was purged for 15 minutes with argon to remove traces of air and moisture. Anhydrous acetone (25 mL) was pumped through the flow apparatus, which was then further purged with argon for 15 minutes. In a typical reaction, a Schlenk flask charged with pivalic acid (1.13 g, 11 mmol), pivalic anhydride (0.4 mL, 2 mmol), ethynyl-β-ionol (2.18 g, 10 mmol), 1,3,5 trimethoxybenzene (420 mg, 2.5 mmol) and anhydrous acetone (10 mL) was connected to the system and the homogeneous mixture circulated around the flow apparatus. After the data acquisition had been started, [Ru^{ll}(η³-CH₂C(Me)CH₂)₂(dppe)] (61 mg, 0.1 mmol, 1mol%) was dissolved in anhydrous acetone (5 mL) and syringed into the Schlenk flask containing all other reagents to initiate the reaction.

Variations of catalytic runs consisting of different starting reagents were added in same amounts unless stated otherwise.

3.0 X-Ray Crystallography

Crystals were selected using the oil drop technique, in perfluoropolyether oil and mounted at 150(2) K with an Oxford Cryostream N₂ cooling device. Intensity data were collected on a Rigaku SuperNova Dual EosS2 single crystal diffractometer using monochromated Cu-Kα radiation ($λ = 1.54184$ Å) or on a Rigaku Xcalibur, EosS2 single crystal diffractometer using graphite monochromated Mo-Kα radiation (λ = 0.71073 Å). Unit cell determination, data collection, data reduction and absorption correction were performed using the CrysAlisPro software.⁶ The structures were solved with SHELXT⁷ and refined by a full-matrix least-squares procedure based on F^2 (SHELXL-2018-19).⁷ All non-hydrogen atoms were refined anisotropically. Hydrogen atoms were placed onto calculated positions and refined using a riding model.

Data analysis and graph display were carried out using the SHELXle⁸ and Mercury⁹ software packages

3.1 Crystal data for *[RuII(η³ -CH2C(Me)CH2)2(dppe)]*

3.2 Crystal data for Ap' *[(dppe)RuII(η² -O2CC^tBu)(η¹ -O2CC^tBu)(^tBuCCO2H)]*

3.3 Crystal data for Bp' *[{(dppe)RuII(η¹ -O2CC^tBu)}2(µ-O2CC^tBu)2(µ-H2O)]*

3.4 Crystal data for Bb' *[{(dppe)RuII(η¹ -O2CPh}2(µ-O2CPh)2(µ-H2O)]*

3.5 Crystal data for A_{ad} '

 $[(dppe)Ru''/\eta^2-O_2CC(CH_2)_{6}(CH)_3C)/(\eta^1$ -2CC(CH2)₆(CH)₃C)((CH)₃(CH₂)₆CCCO₂ H)]

3.6 Crystal data for B_{ad}' [{(dppe)Ru^{||}(η^1 -O₂CC(CH₂)₆(CH)₃)}₂(μ -O₂CC(CH₂)₆(CH)₃)₂(μ -H₂O)]

3.7 Crystal data for Btf' *[{(dppe)RuII(η¹ -O2CCF3}2(µ-O2CCF3)2(µ-OH2)]*

3.8 Crystal data for X 2 ^b•H2O *[(dppe)RuII(η¹ -O2CPh)2(CO)(H2O)]*

3.9 Crystal data for X 2 ^b*[(dppe)RuII(η¹ -O2CPh)2(CO)2]*

4.0 Supplementary data

Figure S34. ¹H NMR spectra of the crude product (3aa) mixture of the Ru-mediated transformation (sum of Z and E adduct) of ethynyl-β-ionol (0.667 M), benzoic acid (1 M) and [(dppe)Ru(MA)2] (1 mol%) at room temperature after 38 hours under inert atmosphere in a) acetone – 93% product yield and b) ethyl acetate – 96% product yield. Illustrating clean ¹H spectrum with a small amount of substrate still observed in both spectra. The E-isomer also highlighted.

4.2 Influence of the carboxylic acid

Table S2. Screening of different carboxylic acids used in the Ru-mediated transformation of ethynyl-β-ionol into their respective ester-adducts under inert atmosphere with anhydrous ethyl acetate at different temperatures, reaction times and volumes.

*Sample was not homogeneous

Figure S35. Example ¹H NMR spectrum showing aldehyde adduct (4a) formation from the Ru-mediated transformation of ethynyl-β-ionol (0.5 M) with benzoic acid (1.5 equiv.) to form 3aa under inert atmosphere in anhydrous ethyl acetate (10 mL) at 20 °C after 24 hours.

4.3 Ligand effects

Table S3. A selection of ruthenium precursors screened with 27 different P, P/P. N/P ligands for the regio- and stereo-selective anti-Markovnikov addition of pivalic acid to ethynyl-β-ionol. Reaction conditions: Substrate (0.1 mmol); catalyst (0.003 mmol; 3 mol%); monodentate ligands (0.0033 mmol) or bidentate ligands (0.0066 mmol), pivalic acid (0.15 mmol), EtOAc (1.2 mL), monitored after 19 hours.

No Ru precursors except $[Ru(cod)(me-ally)]$ showed catalytic activity. We have identified a few ligands giving a small amount of the desired product, albeit lower than dppe (**1-4, 7, 10-23, 25-27).** The ligands **13**, **16** and **20** selectively form only one isomer of the product.

Figure S36. Steric and electronic parameters as well as reaction yields for selected chelating diphosphines used for the Ru-catalysed [(MA2Ru(COD)] transformation of ethynyl-β-ionol (0.1 mmol) with pivalic acid (0.15 mmol) using a catalyst loading of 3 mol% to form 3ab in anhydrous ethyl acetate (1.2 mL) at 20 °C . Yields were determined after 19 h. L1=dppe, L2=dmpe, L3=(R,R)-Dipamp, L4=(S,S)-Norphos, L5=dppm, L6=dppb, L7=dcypp, L8=dppp, L9= (R,R)-DIOP, L10= DPE-phos, L11= D ⁱPrF.

4.4 TON limitation

Table S4. TONs for the ruthenium catalysed transformation of ethynyl-β-ionol (*sum of Z and E adduct) to 3ab at various catalyst loadings (0.1 – 2 mol%). In all cases the reaction was run to maximum yield. Ethynyl-β-ionol (10 mmol); pivalic acid 1.5 equiv, under inert atmosphere with anhydrous acetone (15 mL) with TMB (0.25 equiv, internal standard) was used.*

4.5 Substrate effects

Figure S37. GC-chromatogram of distilled ethynyl-β-ionol.

Sample prepared in methanol: split ratio 10:0, column pressure = 314.2 kPa, and column flow = 3.17 mL/min. The oven temperature increased by 20 °C min⁻¹ from 40 °C to 220 °C (held at 5.5 min). The temperature program was set to 220 °C, 1 min; ramp of 5 °C min⁻¹ to 250 °C, and finally held at 280 °C for 6 min (total run time 29 min). Analyte's retention times were: methanol (2.5 min), (7.83 min), (8.47 min), (8.55 min), ethynyl-β-ionol (8.85 min), (8.88 min), (9.05 min). Impurities identified: α/β-ionone, α-ethynyl-β-ionol, ethynyl-β-ionol -oxide, ethynyl-β-ionol -dimer and dehydrated ethynyl-β-ionol.

Figure S38. Mass spectrum fragmentation of ethynyl-β-ionol.

The mass spectrometry was operated in an EI positive mode, scanning mass ions in the range 10 to 550 (4–29 min) containing a helium carrier gas. The mass fragmentation spectrum of ethynyl-β-ionol $(m/z = 218 [M]^+, 100\%)$ is shown in Figure S37.

Table S5. Product yield of the Ru-mediated *transformation in a range of alkynols (0.66 M) and phenylacetylene (0.66 M) under inert atmosphere in anhydrous acetone (15 mL) with pivalic acid (1 M) TMB (0.167 M), (internal standard) at room temperature after 18 hours and 40 hours*

The peaks used to monitor reaction progress of **3eb** are highlighted: Substrate (**2e**), *Z*-adduct (**3eb**), *E*-adduct (**3eb**), Internal standard

Figure S39. ¹H stacked NMR spectra of the crude product mixture of the Ru-mediated transformation (sum of Z and E adduct) of dimethyloctinol (0.667 M), pivalic acid (1 M) and [(dppe)Ru(MA)2] (1 mol%) at room temperature under inert atmosphere in acetone after a) 1 hour b) 18 hours c) 40 hours. Illustrating reaction progression and near complete product formation of dimethyloctinol -pivalic adduct (97% yield after 40 hours).

4.6 Poisoning tests

To test for possible catalyst poisons, either present in the substrate or formed by the reaction in small amounts, the effect of different functional group additives on the Ru-mediated transformation of phenylacetylene was examined. Considering the context of ethynyl-β-ionol in vitamin A synthesis (Scheme 1), the most relevant functional groups to test included unsaturated aldehydes, unsaturated alcohols and an olefin. **1** and **2f** are minor impurities present in ethynyl-β-ionol (Figures S36-S37), and so were added to the reaction in higher amounts see if they have a negative effect on the catalysis. Other possible poisons/inhibitors tested included water and cyclooctadiene. Following the reaction progress with 0.75 mol% [(dppe)Ru(MA)2] under optimised conditions showed that none of the tested additives caused significant levels of deactivation or inhibition of the catalysis at 10-20% loading, with all product yields falling between 79-82% after 44 hours (Figure 2).

Figure S40. Product yield of the Ru-mediated transformation of phenylacetylene (0.66 M) with pivalic acid (1 M) catalysed by [(dppe)Ru(MA)2] (0.75 mol% loading) in anhydrous acetone (15 mL) at 20 °C in the presence of various additives as detailed in the legend.

4.7 *Operando* ¹H FlowNMR analysis

Table S6. Selected FlowNMR experiments performed to further optimise reaction conditions for kinetic analysis of the Ru-mediated transformation of ethynyl-β-ionol (0.66 M) catalysed by [(dppe)Ru(MA)2] at 1 mol% loading in anhydrous acetone or ethyl acetate (15 mL) at 20 °C from quantitative ¹H FlowNMR spectroscopy at 4 mL/min unless stated otherwise.

4.8 Variable Time Normalisation Analysis

4.8.1 Change in catalyst loading

Figure S41. Maximum catalyst TONs of the Ru-mediated transformation of ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) giving 3ab, catalysed by various amounts of [(dppe)Ru(MA)2] in anhydrous acetone (15 mL) at 20 °C. Determined from quantitative ¹H FlowNMR spectroscopy at 4 mL/min.

4.8.2 Change in pivalic acid loading

Figure S42. Product concentration profiles (sum of Z and E adduct) of the Ru-mediated transformation of ethynylβ-ionol (0.66 M) to form 3ab. This was catalysed with [(dppe)Ru(MA)2] (1mol%) with various amounts of pivalic acid (10-20mmol) in anhydrous acetone (15 mL) at 20 °C. Determined from quantitative ¹H FlowNMR spectroscopy, acquired at 4 mL/min.

Figure S43. Time adjusted reaction progress profiles of data from Figure S42 for a reaction order in [acid] = 0.

4.8.2 Change in ethynyl-β-ionol loading

*Figure S44. Product concentration profiles (sum of Z and E adduct) of the Ru-mediated transformation of ethynylβ-ionol (at different loadings 5 - 15 mmol) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M)) catalysed with [(dppe)Ru(MA)*²] (1 *mol%)* in anhydrous acetone (15 mL) at 20 °C from quantitative ¹H FlowNMR *spectroscopy, acquired at 4 mL/min.*

Figure S45. Time adjusted reaction progress profiles (data from Figure S44) for a reaction order in [Substrate] = 1.

4.8.3 Phenylacetylene concentration profile

Figure S46. Product concentration profiles (sum of Z and E adduct of 3ab) of the Ru-mediated transformation of phenylacetylene (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) catalysed by various amounts of [(dppe)Ru(MA)2] in anhydrous acetone (15 mL) at 20 °C. Determined from quantitative ¹H FlowNMR spectroscopy, acquired at 4 mL/min.

Figure S47. ¹H NMR spectra showing the crude product mixture of the Ru-mediated transformation (sum of Z and E adduct) of phenylacetylene (0.667 M), pivalic acid (1 M) and [(dppe)Ru(MA)2] (1 mol%) at room temperature under inert atmosphere in acetone after a) 1 hour b) 44 hours. This figure illustrates the reaction progression and near complete product formation of the phenylacetylene-pivalic adduct (97% yield after 44 hours). 7 and 8 represent the protons associated to the ester adduct, Z represents Z-ester adduct and E represents E-ester adduct (a ration of ~96/4).

4.9 Rate law derivation

4.9.1 Irreversible literature mechanism

Proposed mechanism for the regioselective ruthenium catalysed anti-Markovnikov addition of carboxylic acids to terminal alkynes giving α , β -unsaturated carboxylates.¹¹⁻¹³

Assumptions:

- $k_1 \gg k_2$, k_3 , k_4 therefore can be neglected and $[1] \approx 0$
- Steady-state Approximation

From the mechanism:

- 1. *Ru*: [Ru(MA)₂(dppe)]
- 2. *A*: [Ru(carboxylate)₂(dppe)]
- 3. *sub*: Substrate (alkynol) (**1**)
- 4. *CA*: Carboxylic acid (**2**)
- 5. *P*: Product (**3**)

Step 1: Activation of the Ru precursor: $Ru \overset{k_1}{\rightarrow} A$ **Step 2:** Coordination of the substrate to **A** to form the vinylidene intermediate (**III**). $A + sub \stackrel{k_2}{\rightarrow} III$ **Step 3:** Transformation to Intermediate (**IV**) $III \overset{k_3}{\rightarrow} IV$ **Step 4:** Addition of carboxylic acid and product ejection / catalyst regeneration. $IV + CA \overset{k_4}{\rightarrow} A + P$ **Equation 1** – Rate Law for Step 1: *Rate* $1 = k_1[Ru]$

Equation 2 – Rate Law for Step 2: *Rate* 2 = $k_2[A][sub]$ **Equation 3** – Rate Law for Step 3: *Rate* $3 = k_3$ [*III*] **Equation 4** – Rate Law for Step 4: *Rate* $4 = k_4[V][CA]$

Using the steady-state approximation for **III** and **IV**:

For III:
$$
\frac{d (III)}{dt} = \text{Rate 2} - \text{Rate 3} = 0
$$

Rearranging Equation (2) and Equation (3): $k_2[A][sub] = k_3[III]$

Equation 5 – [*III*] = $\frac{k_2 [A][sub]}{k}$ k_3 For **IV**: $\frac{d (IV)}{dt}$ $\frac{d\mathbf{r}}{dt}$ = Rate 3 – Rate 4 = 0

Since Rate 3 = Rate 4 - Rearranging Equation (3) and Equation (4): k_3 [III] = k_4 [IV][CA]

Substituting [III] into the equation for IV: $[IV] = k_3(\frac{k_2 [A][Sub]}{k_3}$ $\frac{k_3[3uv]}{k_3}$ = $k_4[IV][CA]$

Equation 6 – [IV] = $(k_2 \frac{[A][Sub]}{k_2 [CA]}$ $\frac{A_1[3uv]}{k_4[CA]}$

Now, the overall rate at which the product is formed is Rate 4: Rate_{overall} = k₄[IV][CA]

Substituting [IV] from Equation (6) into Equation (4): Rate_{overall} = $k_4(k_2 \frac{[A][Sub]}{k_1 [CA]}$ $\frac{A_1[3ab_1]}{k_4[CA]}$ [CA]

Without assuming the rate determining step, the unbiased rate law derived from the mechanism and the steady state approximation is:

Equation 7 – Rate_{overall} = k_2 [A][Sub]

Applying the steady-state approximation to IV (a different intermediate prior to product formation):

The rate of formation of **IV** (from **Equation 3**):

Equation $3 - Rate_{formation} = Rate 3 = k_3$ [III]

The rate of consumption of **IV** - Equation (**4**) Repeated:

Rate_{consumption} = Rate $4 = k_4$ [**IV**][CA]

According to the steady-state approximation: **Equation 8** – $\frac{d (IV)}{dt}$ $\frac{d\mathbf{r}}{dt}$ = Rate formation 3 – Rate consumption $4 = 0$

 k_3 [**III**] − k_4 [**IV**][CA] = 0 → k_3 [**III**] = k_4 [**IV**][CA]

Now, from the steady-state approximation for III, we derived: $\text{[III]} = (k_2 \frac{[A][sub]}{k_2})$ $\frac{1}{k_3}$) (Equation 5)

Substitute this expression into the equation for **IV** from **Equation 8**:

Equation 9: $k_3(k_2 \frac{[A][sub]}{k_2})$ $\frac{1}{k_3}$ = k_4 **[IV**] [CA]

This simplifies to: $k_2[A][Sub] = k_4[IV][CA] \rightarrow [IV] = k_2 \frac{[A][sub]}{k_2[A]}$ $k_4[A]$

The overall rate at which the product is formed is determined by Rate 4 (**Equation 4**):

 $Rate_{overall} = k_4$ [**IV**][CA]

Substitute our derived expression for [**IV**] from **Equation 9**:

Equation 10 – *Rate*_{overall} = k_4 (k_2 $\frac{[A][sub]}{k_4$ [*CA*] $\frac{A_1[3au]}{k_4[CA]}$ [CA]

This simplifies to: *Rateoverall* = *k*2[*A*][*Sub*]

This result is the same as the one we derived from the steady-state approximation for **III**. This demonstrates the internal consistency of the mechanism and shows that the rate law is robust to the choice of intermediate for which the steady-state approximation is applied.

4.9.2 Reversible literature mechanism

Revised Mechanism with reversibility for steps 2 and 3:

Step 1 (assumed irreversible): $Ru \overset{k_1}{\rightarrow} A$ Step 2 (reversible): $A + Sub \begin{matrix} k_2 \\ \rightleftarrows \\ k_{-2} \end{matrix}$ III Step 3: III $\frac{k_3}{2}$ IV Step 4: *IV* + CA ^{*k*₄} *A* + *P*

Rate Laws:

- 1. Rate $1 = k_1[Ru] -$ **Equation 1** (repeated)
- 2. Rate $2_{formation} = k_2[A][Sub] Equation 2 (repeated)$
- 3. Rate 2dissociation = k−2[**III**] **Equation 11**
- 4. Rate $3 = k_3$ [III] **Equation 3** (repeated)
- 5. Rate $4 = k_4[V][CA] -$ **Equation 4** (repeated)

Steady-State Approximation for [III]: Given that the formation and consumption of **III** is in a steady state: **Equation 12** $\frac{d (III)}{dt}$ = *Rate 2*_{formation} – *Rate 2*_{dissociation} – *Rate* 3 = 0

Substituting in the rate laws: $k_2[A][Sub] - k_{-2}[III] - k_3[III] = 0$

From this equation, we can rearrange and isolate [III]: [III] = $k_2 \frac{[A][sub]}{k_2 + k_3}$ $k_{-2}+k_3$

Solving for [**IV**]**: Equation 13** - $\frac{d (IV)}{dt}$ $\frac{dV}{dt}$ = Rate 3 – Rate 4 = 0

Substitute the rate laws: k_3 [III] = k_4 [IV][CA]

Substitute for [III] from **Equation 12**: $k_3 (k_2 \frac{[A][sub]}{k_3+k_4})$ $\frac{[A_1|3uv]}{k_{-2}+k_3}$ = k_4 [**IV**][CA]

Equation 14 - Solve for [IV]: $k_3 \frac{k_3 k_2 [A][sub]}{(k_1 + k_2)k_3}$ $(k_{-2}+k_3)k_4[CA]$

Overall Rate Law:

- 1. The overall rate is governed by Rate 4: Rate_{overall} = k_4 [**IV**][CA]
- **2.** Substitute for [**IV**] from **Equation 14**:

Equation 15: Rate_{overall} = $k_4 \left(\frac{k_3 k_2 [A][sub]}{(k_1 + k_2)k_1 [S]} \right)$ $\frac{k_3k_2[A][3ab]}{(k_{-2}+k_3)k_4[CA]}$ [CA]

3. Simplify: Rate_{overall} = $k_3 k_2 \frac{[A][sub]}{(k_3+k_3)}$ $\frac{[A_1|3ab_1]}{(k_{-2}+k_3)}$

This rate law indicates that the overall rate is dependent on the concentrations of **A** and Sub, as well as the forward and reverse rate constants for substrate coordination (k_2 and k_2). The rate law also shows that the system's behaviour will be influenced by the relative magnitudes of k_2 and k_3 , with the latter not being influenced by the concentration of CA as it only appears in the last step.

If *k***−² is much smaller than** *k***3**: This condition implies that once **III** is formed, it transforms rapidly to **IV** without significant reverse reaction back to A + Sub. Under this condition, the reverse rate *k*−2[**III**] becomes negligible compared to k_3 [III].

Therefore, with this approximation, $\text{[III]} \approx (k_2 \frac{[A][sub]}{(k_2)}$ (k_3)

Considering Rate 4 governs the overall rate: $Rate_{overall} = k_4[IV][CA]$.

Since **IV** is formed from **III**, and assuming CA is in excess (its concentration doesn't significantly change), the rate of formation of **IV** can be approximated to be proportional to the rate at which **III** is formed, i.e., Rate 2.

Thus, under the assumption that k₋₂≪k₃ and CA is in excess, Rate_{overall} simplifies to the rate of formation of **III**, which is $k_2[A][Sub]$.

The overall rate law, Rate_{overall} $\approx k_2[A][Sub]$, can be derived under the condition that the reverse reaction in step 2 is much slower than the forward reaction (i.e., k-2 is much smaller than k₃). Simplifying this allows us to focus on the forward reaction rates, particularly the rate of substrate coordination with **A**, as the main causes of the overall reaction rate.

4.10 Identification of reaction intermediates by ³¹P{¹H} NMR

4.10.1 Precursor activation

a) With benzoic acid

Figure S49. Quantitative ³¹P{¹H} NMR (inverse-gated) showing a mixture of [(dppe)Ru^{||}(η^2 -O₂CPh)₂] (A_b) and *[(dppe)RuII(η² -O2CPh) (η¹ -O2CPh)(H2O)] (Bb) in solution.*

Figure S50. 1H-³¹P HMBC after the addition of benzoic acid to [(dppe)RuII(MA)2] showing correlation assigned peaks of intermediates A^b and B^b in solution from the ¹H and ³¹P{¹H} NMR.

Figure S51. ³¹P{ ¹H} DOSY, acquired 45 minutes after the addition of 2 eq. benzoic acid to [(dppe)RuII(MA)2], in ethyl acetate.

For the DOSY experiment, 64 scans were performed with a delay of 1 second and 0 dummy scans. The diffusion delay, Δ, was set to a value of 0.06 s, and the gradient pulse length, δ/2, was set to 2000 μs. After acquisition, Fourier transform, phase and baseline correction in TopSpin, the diffusion data was processed in dynamics centre to obtain diffusion coefficients for $[(\text{dppe})Ru^{\text{II}}(MA)_2]$, A_b and B_b .

T*able S7. The effect of different amounts of moisture on the ratio of intermediate A^b and B^b in toluene once 2 equivalents of benzoic acid has been completely reacted with [(dppe)[Ru^{|I}(MA)₂] where the starting ratios of* A_b *and* B_b *were 98 and 2.Loading of moisture is in respect to [(dppe)[Ru^{II}(MA)₂].*

Figure S52. ³¹P{¹H} NMR showing the addition of H2O to a ~1:1 mixture of A^b and B^b generated from [(dppe)Ru(MA)2] and benzoic acid in acetone at room temperature resulting in the shift of distribution in favour of B^b

Figure S53. $^{31}P_1^4H$ } NMR spectrum showing relative amounts of Ab [(dppe)Ru"(η^2 -O2CPh)2] and Bb [(dppe)Ru"(η^2 -*O2CPh)(η¹ -* O2CPh)(H2O)] resulting *from the addition of 15 equiv of benzoic acid to [(dppe)[RuII(MA)2].*

Figure S54. ³¹P{¹H} NMR spectrum showing relative amounts of **A**_b [(dppe)Ru^{ll}(η ²-O2CPh)2] and **B**_b [(dppe)Ru^{ll}(η ²-*O2CPh)(η¹ - O2CPh)(H2O)] from the addition of 11 equiv of benzoic acid and 2 equiv of benzoic anhydride to [(dppe)[RuII(MA)2].*

b) With pivalic acid and adamantane carboxylic acid

 ${}^{31}P{^{1}H}T_1$ of [(dppe)Ru^{||}(MA)₂]

Figure S55. ³¹P inversion recovery NMR experiment performed to obtain the T₁ value of [(dppe)[Ru^{II}(MA)₂] in *acetone. Value of 1.1 s obtained.*

 ${}^{31}P{^1H}T_1$ of [(dppe)Ru^{||}(η^2 -O₂C^tBu)₂]

Figure S56. ³¹P inversion recovery NMR experiment performed to obtain the T¹ value of A^p in acetone. Value of 0.74 s obtained.

Figure S57. Stacked ³¹P{¹H} NMR showing the formation of A using different carboxylic acids a) pivalic, b) acetic, c) adamantane carboxylic acids.

Figure S58. Exemplary ¹H NMR spectrum of the formation of A^p in (CD3)2CO.

Figure S59. Exemplary ³¹P{¹H} NMR spectrum observed after for the addition of H₂O to [(dppe)Ru^{II}(n²-O2C^tBu)₂] *(Ap).*

Table S8. A list of selected bond lengths from the crystallographic data to show how the central ruthenium atom is coordinated to the carboxylates in the various analogues of A, and to the methyl allyls in [(dppe)Ru(MA)2]. Also showing selected C-O bond lengths. Refer to pages S45-S62 for corresponding labels. Similar structures for comparison angles and lengths can be found here.¹⁴

4.10.2 Monitoring catalyst speciation under catalytic conditions using ³¹P{¹H} NMR

 $65 60 55 50$ 45 40 35 30 25 20 $-5 - 10$ ppm 105 100 95 90 85 80 75 70 15 10 $\frac{1}{5}$ $\dot{\mathbf{o}}$ *Figure S60. Exemplary ³¹P{¹H} FlowNMR spectra showing the reaction progress of the Ru-mediated transformation of ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.14 M) forming 3ab catalysed by [(dppe)Ru(MA)2] (1 mol%) in anhydrous acetone (15 mL) at 20 °C. Clearly depicting the consumption of A^p over time and the formation of other reaction species. Determined from ³¹P{¹H} FlowNMR spectroscopy, acquired at 4 mL/min.*

Figure S62. Stacked ³¹P{¹H} NMR showing the reaction progress of the Ru-mediated transformation of ethynylβ-ionol (0.66 M) using pivalic acid under different conditions, recorded at 30 h under inert atmosphere at rt unless stated: a) in acetone with pivalic acid (1 M) at 1 mol% catalyst, b) with phenylacetylene in acetone with pivalic acid (0.66 M) and pivalic anhydride (0.17 M) at 1 mol% catalyst, c) in acetone with pivalic acid (1 M) at 0.5 mol% catalyst, d) in ethyl acetate with pivalic acid (1 M) at 1 mol% catalyst, e) in acetone with pivalic acid (0.66 M) and pivalic anhydride (0.17 M)) at 0.5 mol% catalyst. Determined from ³¹P{¹H} FlowNMR, acquired at 4 mL/min.

Figure S63. Stacked ³¹P{¹H} NMR showing the reaction progress of the Ru-mediated transformation of ethynylβ-ionol (0.66 M) with various carboxylic acids catalysed by [(dppe)Ru(MA)2] in anhydrous acetone (15 mL) at 20 °C after 40 hours. Determined from *³¹P{¹H}* FlowNMR spectroscopy, acquired at 4 mL/min.

Figure S64. ³¹P{¹H} NMR stack plot showiing a) A^p under inert atmosphere b) A^p opened to air after 48 hours.

Figure S65. ³¹P{¹H} NMR concentration profiles showing the comparison of the consumption of A^p with the addition of 2 mM of DPPE oxide at the start of the reaction (1) and without (2) to a reaction solution of ethynylβ-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) catalysed by [(dppe)Ru(MA)2] (1mol%) in anhydrous acetone (15 mL) at 20 °C. Determined by quantitative ³¹P{ ¹H} FlowNMR spectroscopy at 4 mL/min.

Figure S66. ³¹P{¹H} NMR spectrum showing the formation of two mutually coupling triplets at 54.0 ppm and 57.2 ppm in the Ru-mediated transformation of ethynyl-β-ionol (0.66 M) with pivalic acid (0.1 M) catalysed by [(dppe)Ru(MA)2] (0.5 mol%) in anhydrous acetone (15 mL) at 20 °C with at 0.5 equiv. of DPPE added at the start of the reaction, resulting in no product yield observed. Peak at -13 ppm shows free dppe present and tri-o-tolyl phosphate at -16 ppm.

Table S9. Comparison of the crystallographic data of selected bond lengths of [(dppe)Ru^{||}(η²-O₂CPh)₂(CO)(H₂O)] and [(dppe)RuII(η 2 -*O2CPh)2(CO)2]. Refer to pages S45-S62 for corresponding labels.¹¹*

Figure S67. ³¹P{¹H} NMR spectrum showing the isolated [(dppe)Ru^{||}(¹³CO)₂(η ²-O₂CPh)₂] in CDCl₃.

Figure S68. ¹³C{¹H} NMR spectrum showing the isolated [(dppe)Ru^{||}(¹³CO)₂(η ²-O₂CPh)₂] in CDCl₃.

 ${}^{13}C{^1H}$

 ${}^{31}P{^1H}$

Figure S69. ¹³C{¹H} (top) and ³¹P{¹H} (bottom) simulations of [(dppe)Ru^{ll}(¹³CO)₂(η ²-O₂CPh)₂]. In each case the real *spectrum is on top, simulated is on the bottom. Impurity peaks are marked with an asterisk. Only the relevant carbonyl region is shown in the ¹³C{¹H} spectrum, and impurity peaks are marked with an asterisk. Simulations were carried out using gNMR (IvorySoft) version 5.0.6.0. Simulations were achieved with J_{cc} = 4.36; J_{PP} = -25.84; JPC = 113.24 (E) and -13.45 (Z).*

M) with benzoic acid (1 M) and [(dppe)Ru^{||}(CO)(OH₂)(η^2 -O₂CPh)₂] (X^2b , 1 mol%) in anhydrous acetone (15 mL) at *20 °C at a) t = 30 minutes b) t = 48 hours. No product formation was observed in this time.*

Figure S72. ³¹P{¹H} NMR showing no change in the chemical shift of X ²b in the Ru-mediated transformation of ethynyl-6-ionol (0.66 M) with benzoic acid (1 M) and [(dppe)Ru"(CO)(OH₂)(n²-O₂CPh)₂] (X²b, 1 mol%) in anhydrous *acetone (15 mL) at 20 °C at a) t = 30 minutes b) t = 48 hours.*

Figure S73. ¹H NMR spectra showing the post reaction mixture of the Ru-mediated transformation of ethynyl-βionol (0.66 M) with a) benzoic acid to form 3aa b) adamantane carboxylic acid to form 3ae, catalysed by [(dppe)Ru(MA)2] (1 mol%) in anhydrous acetone (15 mL) at 20 °C, identifying the characteristic shift for the formation of the aldehyde adduct.

4.11 Mechanistic relevance of [(dppe)Ru^{||}(η²-O₂CR)₂]

Figure S74. Product concentration profiles (sum of Z and E adduct) overlayed with deactivation of A showing the Ru-mediated transformation of ethynyl-β-ionol (0.66 M) or phenylacetylene (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) catalysed by [(dppe)Ru(MA)2] at various loadings in anhydrous acetone (15 mL) at 20 °C from quantitative ¹H and ³¹P{¹H} FlowNMR spectroscopy at 4 mL/min.

Figure S75. Initial rates of [Ap] decay against the total initial concentration of A^p for the Ru-mediated transformation of *ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) catalysed by various amounts of [(dppe)Ru(MA)2] (0.25 - 2 mol%) in anhydrous acetone (15 mL) at room temperature.*

Figure S76. ln([Ap]) vs ln(dAp/dt) showing catalyst deactivation to be half order in [A] through the gradient.

Figure S77. Rate of decay of A against substrate consumption in the Ru-mediated *transformation of ethynyl-βionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) to form 3ab, catalysed by [(dppe)Ru(MA)₂] (1 mol%) in anhydrous acetone (15 mL) at room temperature.*

Figure S78. Concentration profiles showing the consumption of A^p for the Ru-mediated transformation of ethynyl-β-ionol at three different loadings (0.33 M, 0.66 M, 1 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) to form 3ab, catalysed by [(dppe)Ru(MA)2] (6.66mM) in anhydrous acetone (15 mL) at 20 °C. Determined from quantitative 31P{1H} FlowNMR spectroscopy at 4 mL/min.

Figure S79. Time adjusted reaction progress profiles (from Figure S78) for the deactivation rate as a function of [Acid] = 0

Figure S80. Overlay of the decline of A^p over time under different catalytic conditions at room temperature: a) phenylacetylene (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) with [(dppe)Ru(MA)2] (1 mol%) in anhydrous acetone in anhydrous acetone, b) ethynyl-β-ionol (0.66 M) with pivalic acid (1 M) with [(dppe)Ru(MA)2] (1 mol%), c) ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) with [(dppe)Ru(MA)2] (0.5 mol%) in anhydrous acetone, d) ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) with [(dppe)Ru(MA)2] (1 mol%) in anhydrous ethyl acetate, e) ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) with [(dppe)Ru(MA)2] (1 mol%) in anhydrous acetone. Determined from quantitative 31P{1H} FlowNMR spectroscopy at 4 mL/min.

4.12 Maximising catalyst productivity

Figure S81. Conversion profiles from ¹H FlowNMR data of the Ru-mediated transformation of ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) catalysed by 0.5 mol% [(dppe)Ru(MA)2] in anhydrous acetone (15 mL) at different temperatures.

Figure S82. Decline of A from ³¹P{¹H} FlowNMR data of the Ru-mediated transformation of ethynyl-β-ionol (0.66 M) with pivalic acid (0.73 M) and pivalic anhydride (0.13 M) catalysed by 0.5 mol% [(dppe)Ru(MA)2] in anhydrous acetone (15 mL) at different temperatures.

The peaks used to monitor reaction progress of **3ae** are highlighted: Substrate (**2a**), *Z*-adduct (**3ae**), *E*-adduct (**3ae**), Internal standard

Figure S83. The Ru-mediated transformation of ethynyl-β-ionol with adamantane carboxylic acid and [(dppe)Ru(MA)2] in anhydrous acetone under inert atmosphere (final product yields after 45 hours obtained from quantitative ex-situ ¹H NMR spectroscopy against trimethoxybenzene as internal standard) at: a) 2 M substrate loading and 6.67 mM Ru loading b) 5.23 M substrate loading with 3.34 mM Ru loading c) 5.23 M substrate loading with 6.67 mM Ru loading at 40 °C. With example substrate and product peak highlight for quantification.

Table S10. The Ru-mediated transformation of phenylacetylene with adamantane carboxylic acid with [(dppe)Ru(MA)2] in anhydrous acetone under inert atmosphere (final product yields after 60 hours obtained from quantitative ex-situ ¹H NMR spectroscopy against trimethoxybenzene as an internal standard).

Entry	Substrate	Catalyst	Catalyst	Acid (M)	Temperature	Conversion	TON
			loading		(°C)	$(\%)$	
	1.333M	6.67 mM	0.5%	1.8	20	100	200
2	5.23M	6.67 mM	0.125%	5.5	20	100	800
3	10.46 M	6.67 mM	0.0625%	11	20	98	1568
4	20.98 M	6.67 mM	0.03125%	22	20	66.8	2138

5.0 References

- 1. H. Doucet, B. Martin-Vaca, C. Bruneau and P. H. Dixneuf, *J. Org. Chem.*, 1995, **60**, 7247-7255.
- 2 F. Aquino, W. Bonrath, F. Pace, P. Ruckstuhl and K. Witzgall, WO2020025512A1, 2020.
- 3. J. G. Małecki and A. Maroń, *Transit. Met. Chem.*, 2012, **37**, 727-734.
- 4. M. S. Quinby and R. D. Feltham, *Inorg. Chem.*, 1972, **11**, 2468-2476.
- 5. A. Hall, PhD Thesis, University of Bath, 2019.
- 6. CrysAlisPro 1.171.42.49 (Rigaku Oxford Diffraction, **2022**).
- 7. G. M. Sheldrick, *Acta Cryst.*, 2015, C71, 3-8.
- 8. C. B. Hübschle, G. M. Sheldrick and B. Dittrich, J. Appl. Cryst., 2001, **44**, 1281-1284.
- 9. C. F. Macrae, P. R. Edgington, P. McCabe, E. Pidcock, G. P. Shields, R. Taylor, M. Towler and M. van der Streek, *J. Appl. Crystallogr.*, 2006, **39**, 453-457.
- 10. D. A. Engel and G. B. Dudley, *Org. Biomol. Chem.*, 2009, **7**, 4149-4158
- 11. M. Picquet, C. Bruneau and P. H. Dixneuf, *Chem. Commun.,* 1997, **1997**, 1201-1202.
- 12. H. Doucet, B. Martin-Vaca, C. Bruneau and P. H. Dixneuf, *J. Org. Chem.,* 1995, **60**, 7247-7255.
- 13. M. Picquet, A. Fernández, C. Bruneau and P. H. Dixneuf, *Eur. J. Org. Chem.,* 2000, **2000**, 2361- 2366.
- 14. X. L. Lu, S. Y. Ng, J. J. Vittal, G. K. Tan, L. Y. Goh and T. S. A. Hor, *J. Organomet. Chem.*, 2003, **688**, 100-111.