

## Supporting Information Available

### **Novel half Salphen cobalt(III) complexes: synthesis, DNA binding and anticancer studies**

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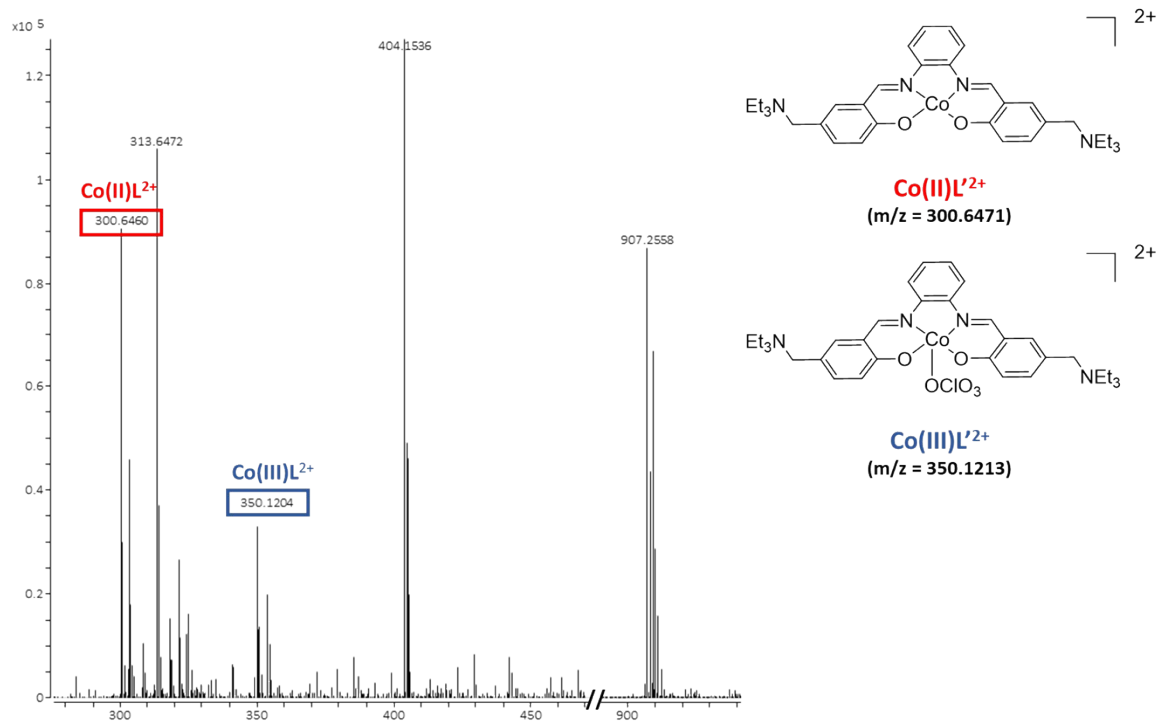


Figure S1. ESI-MS spectrum in acetonitrile of the pH = 6 reaction.

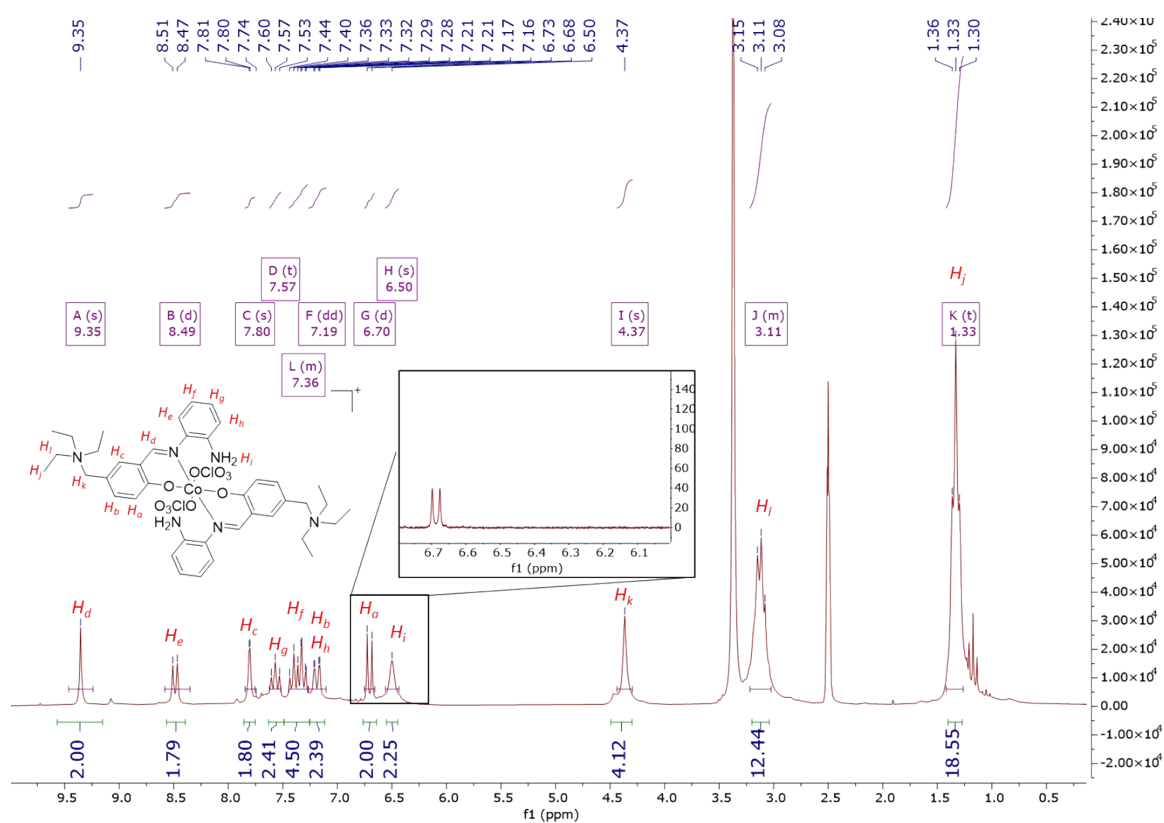


Figure S2. <sup>1</sup>H NMR of **1** in d<sub>6</sub>-DMSO. In the inset, H<sub>k</sub> signal disappearance upon D<sub>2</sub>O addition.

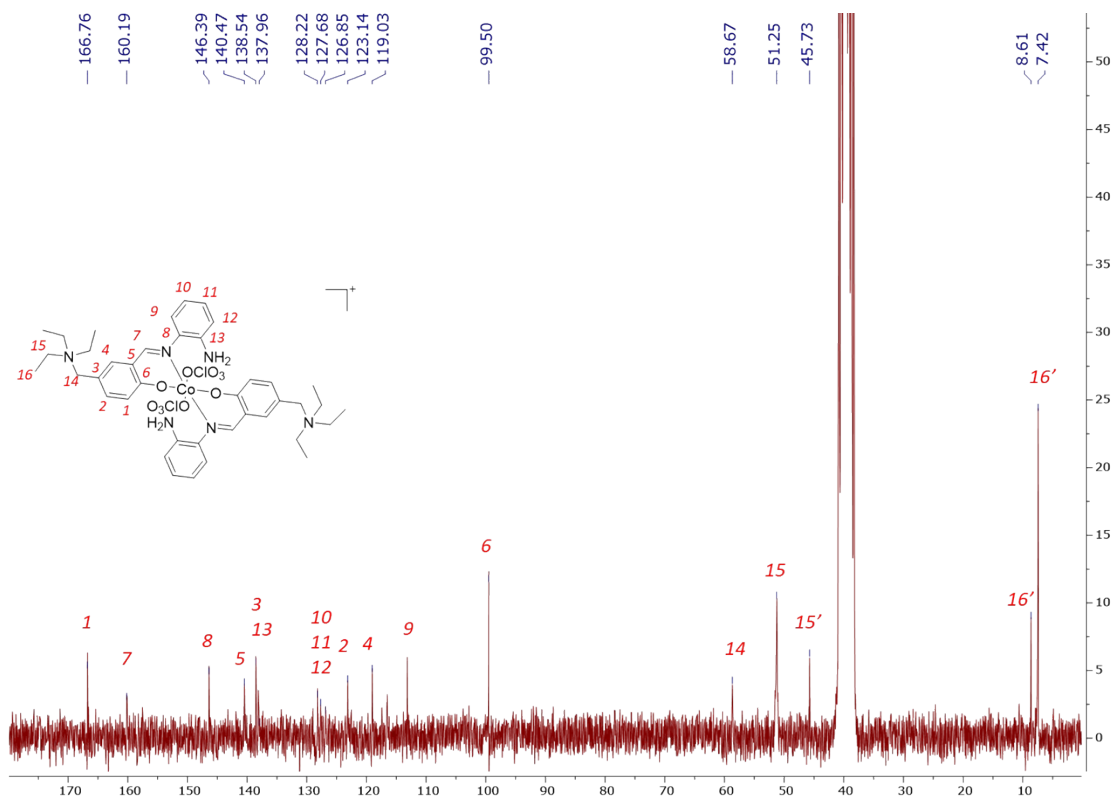


Figure S3.  $^{13}\text{C}$  NMR of **1** in  $d_6$ -DMSO.

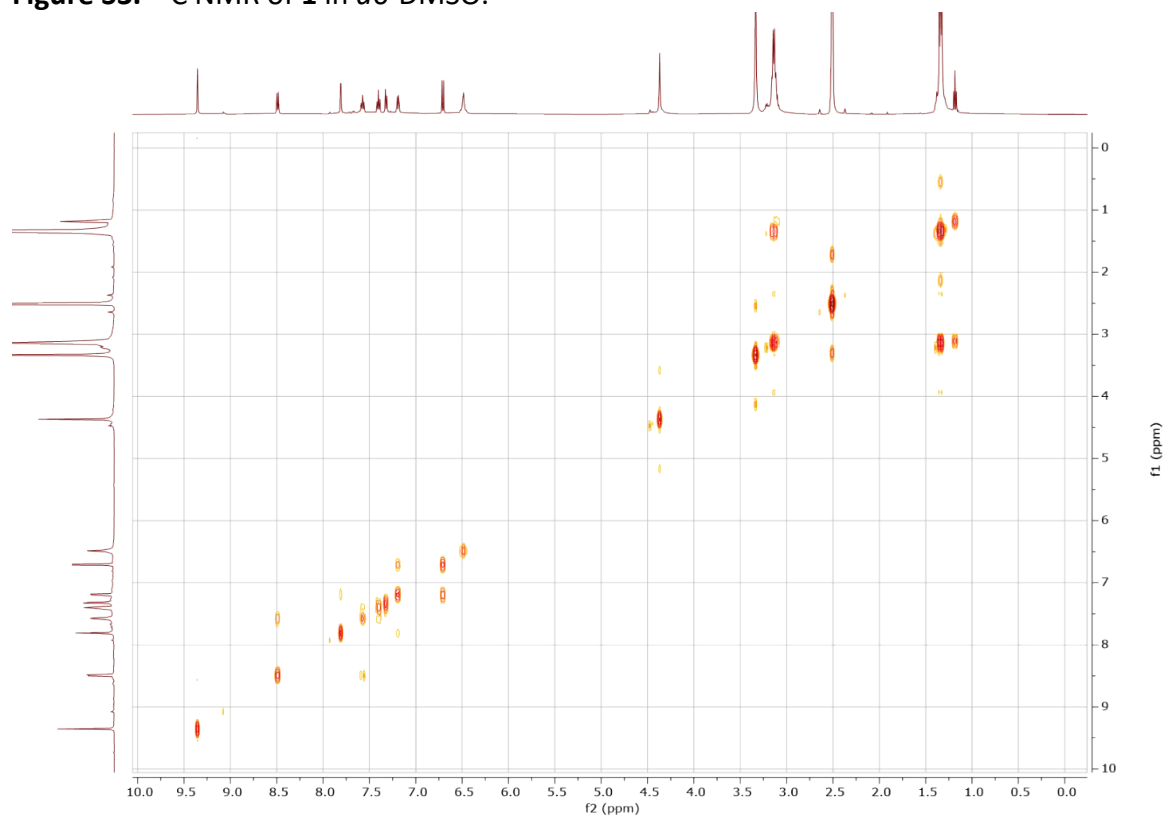


Figure S4.  $^1\text{H}$ - $^1\text{H}$  COSY NMR of **1** in  $d_6$ -DMSO.

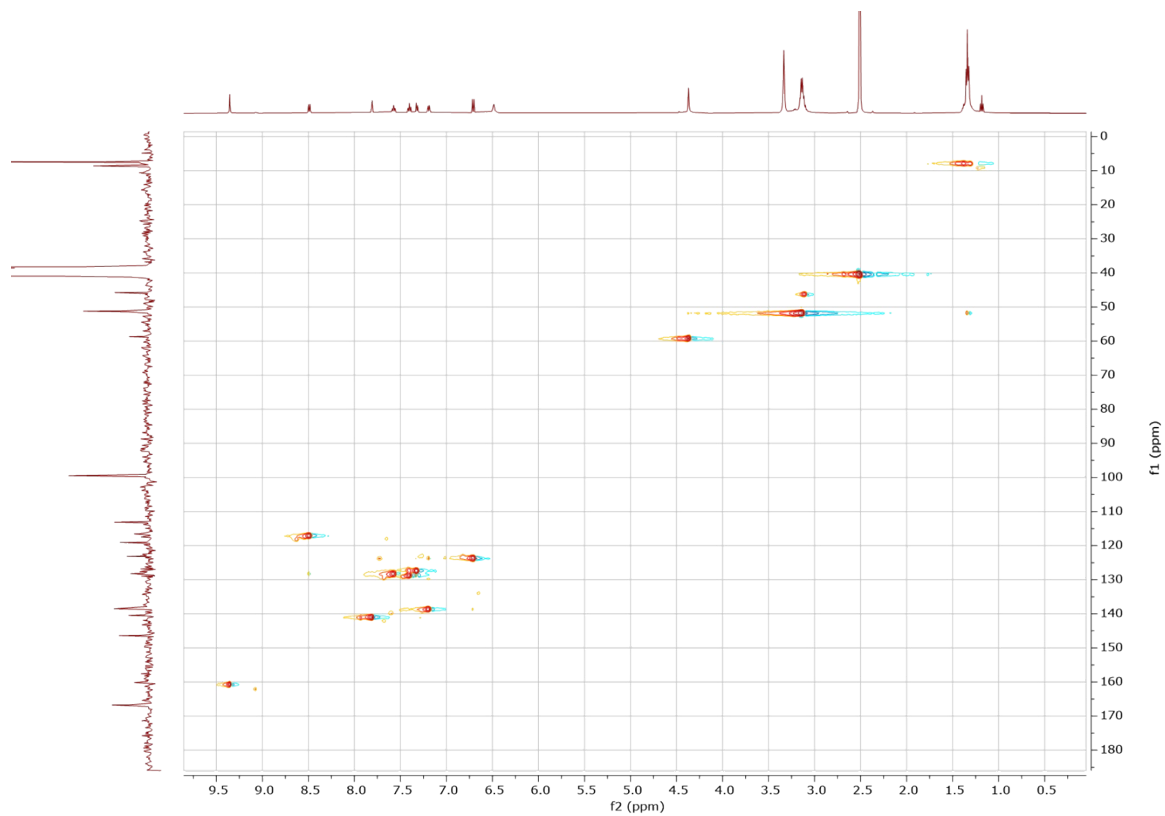
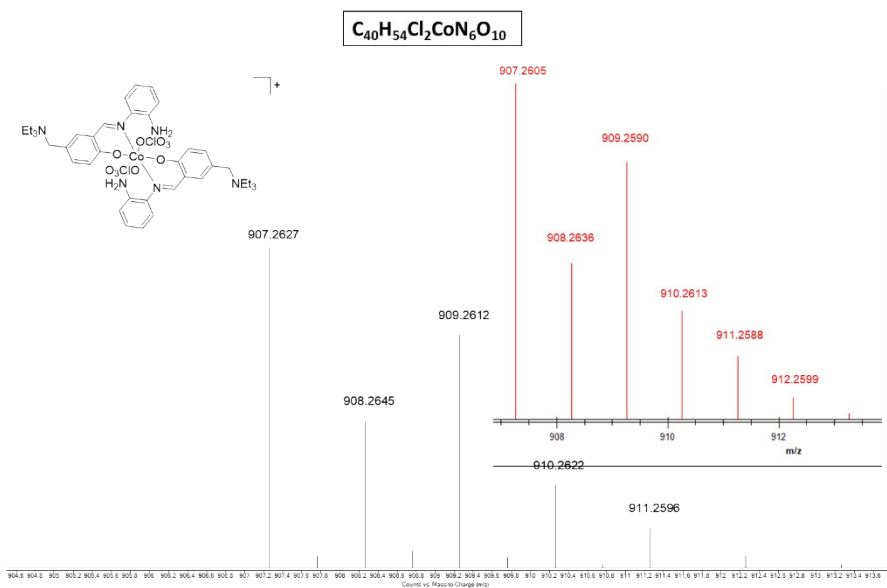
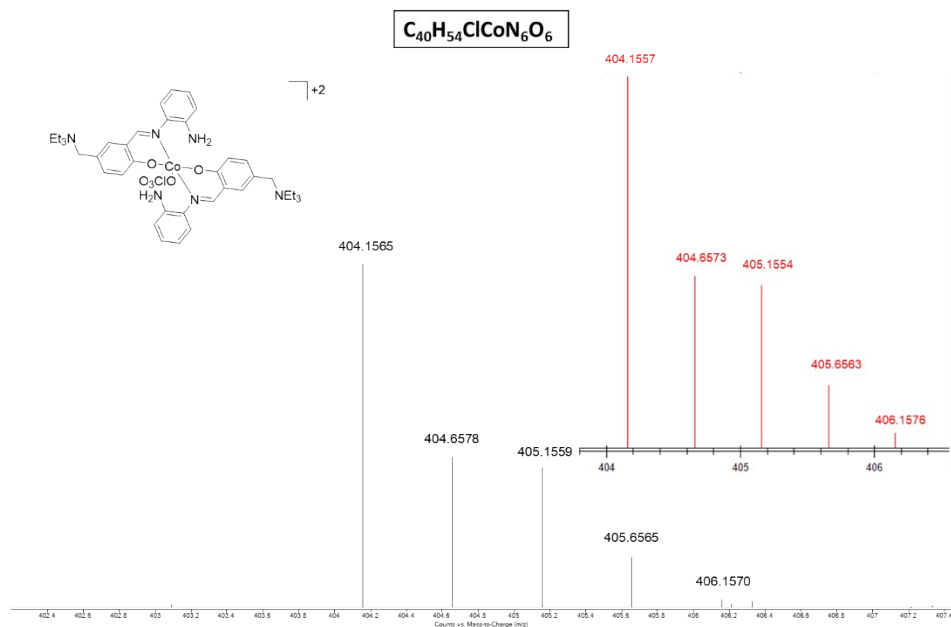


Figure S5.  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR of **1** in  $d_6$ -DMSO.

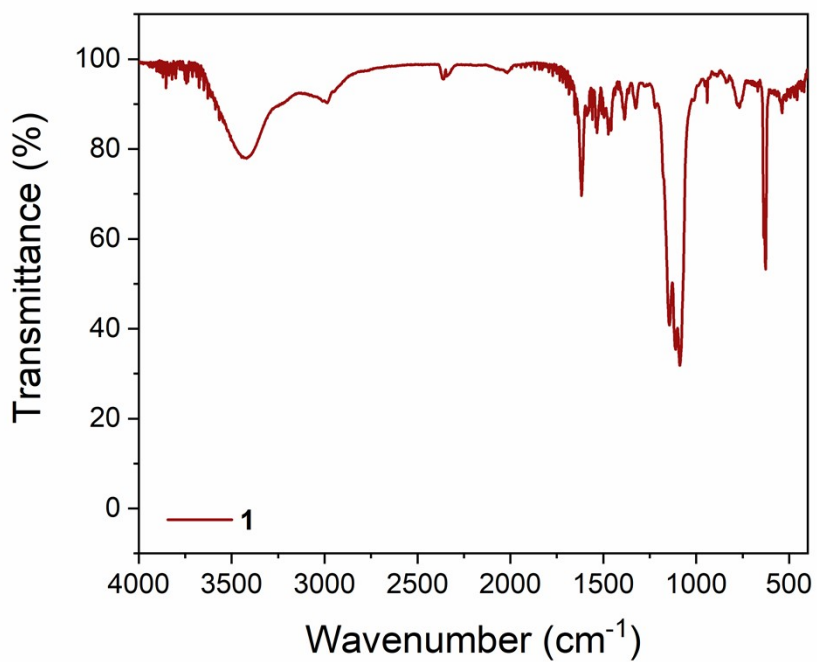
a)



b)



**Figure S6.** ESI-MS spectra of the two species found for **1** in acetonitrile: **[1]<sup>+</sup>** (a) and **[1-ClO<sub>4</sub>]<sup>+2</sup>** (b). Simulated spectra are depicted in red as insets.



**Figure S7.** IR spectrum of **1** in KBr.

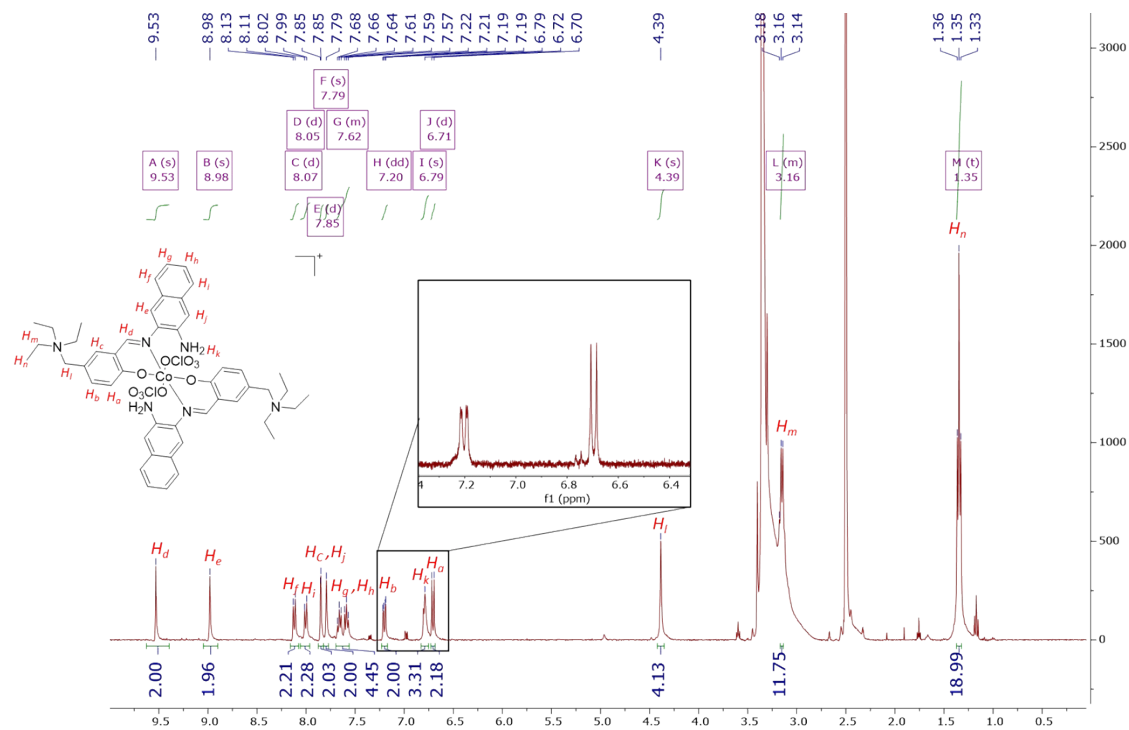


Figure S8.  $^1\text{H}$  NMR of **2** in  $d_6$ -DMSO. In the inset,  $H_k$  signal disappearance upon  $\text{D}_2\text{O}$  addition.

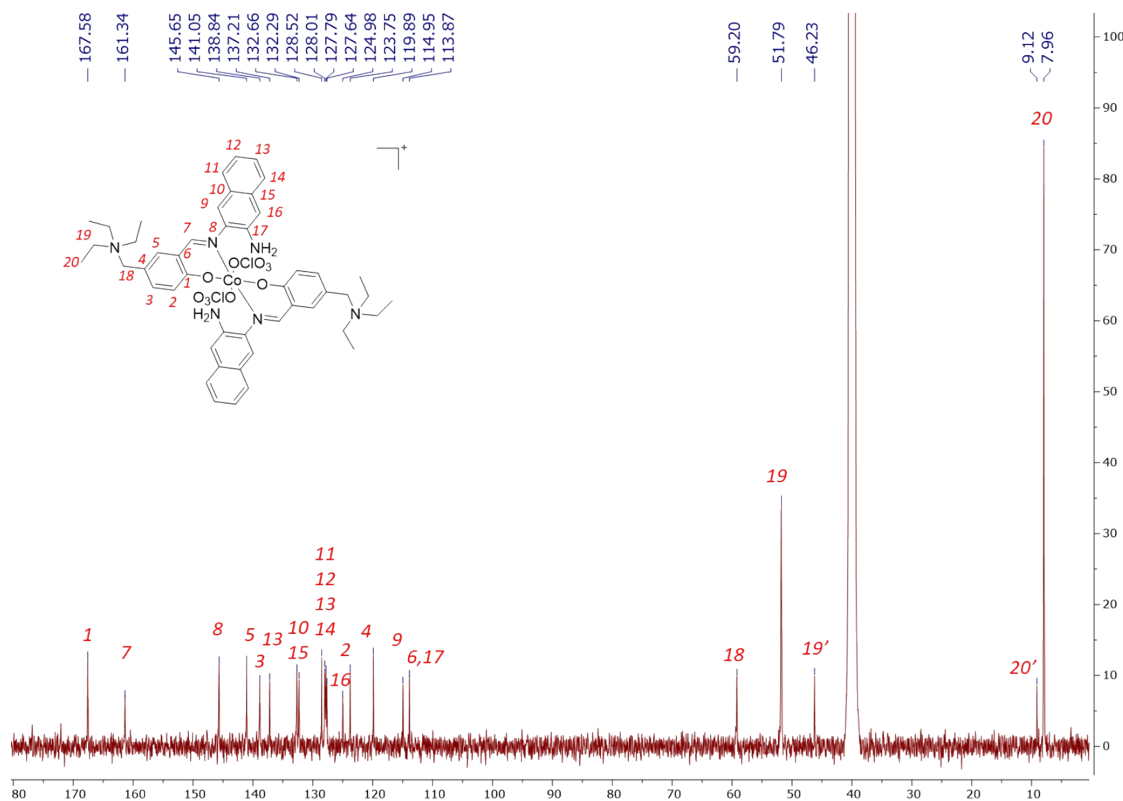
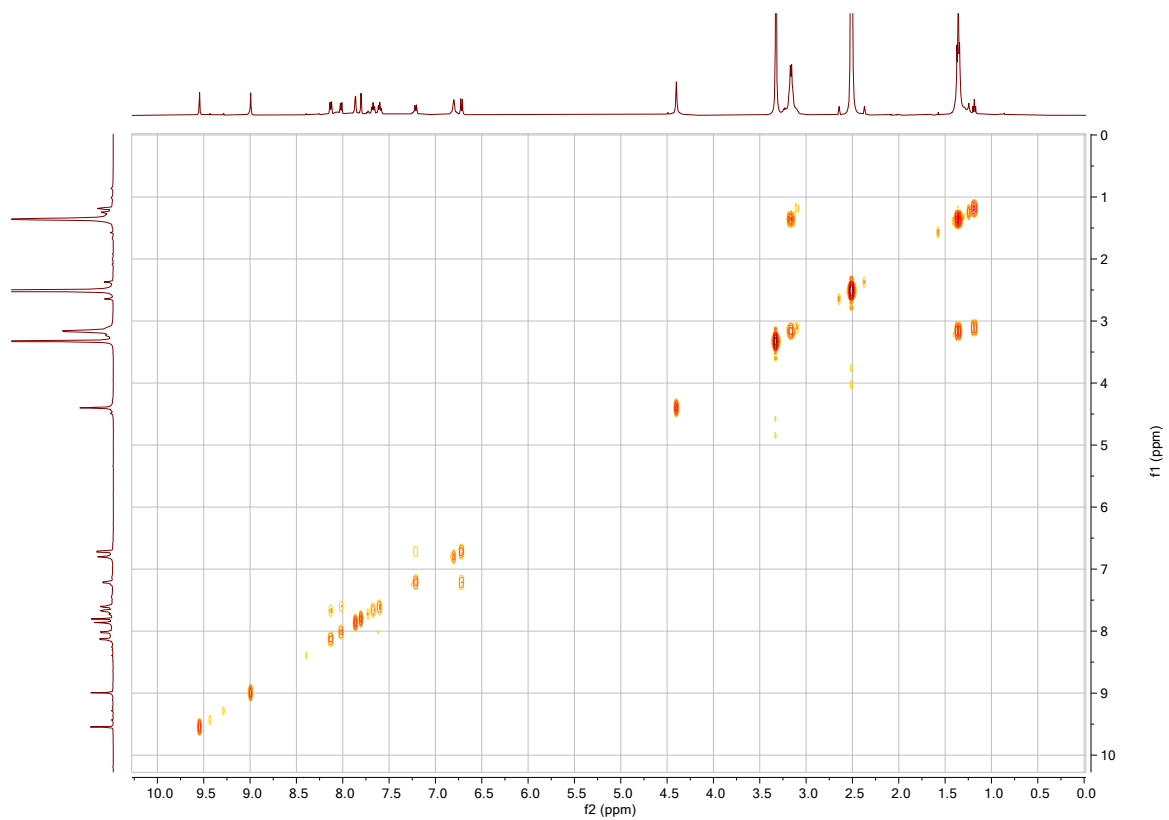
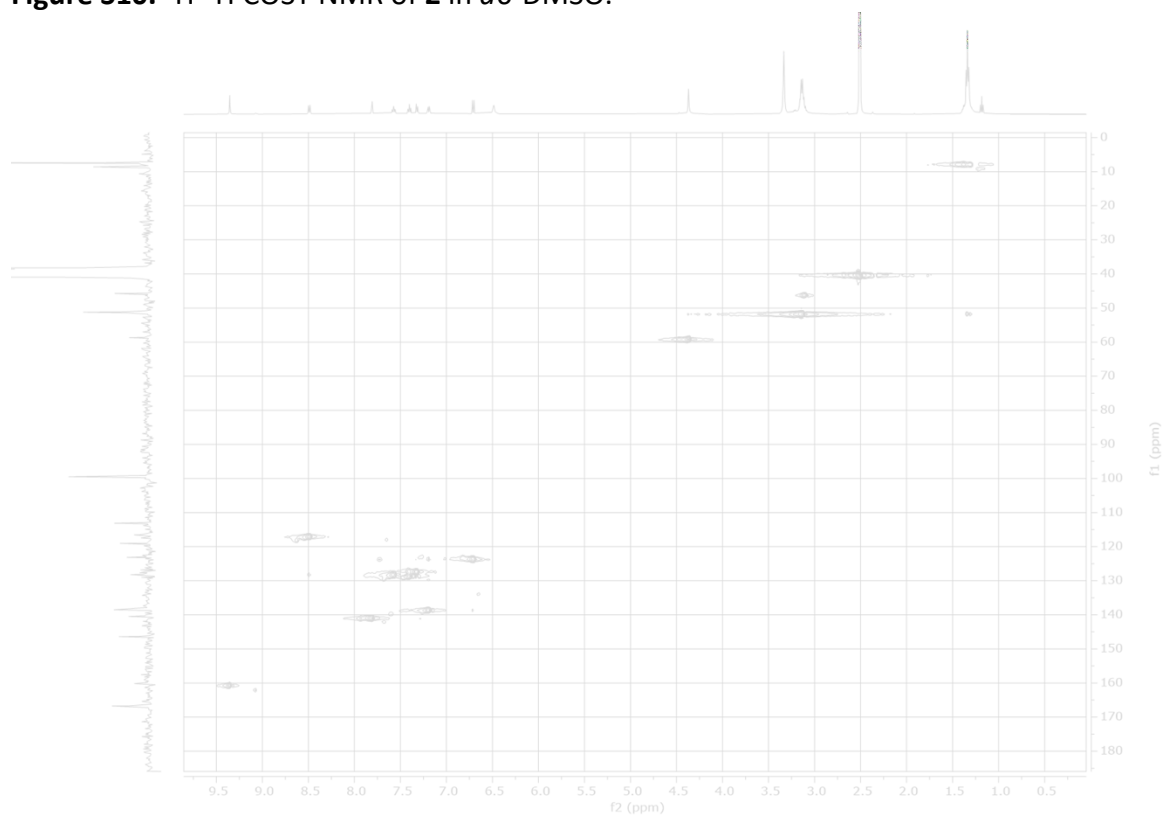


Figure S9.  $^{13}\text{C}$  NMR of **2** in  $d_6$ -DMSO.

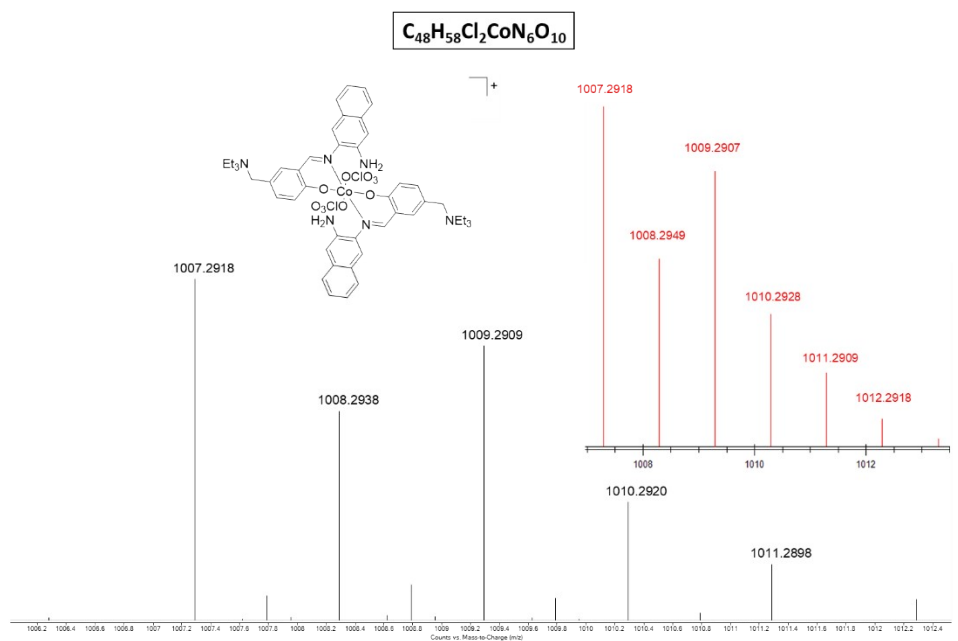


**Figure S10.**  $^1\text{H}$ - $^1\text{H}$  COSY NMR of **2** in  $d_6$ -DMSO.

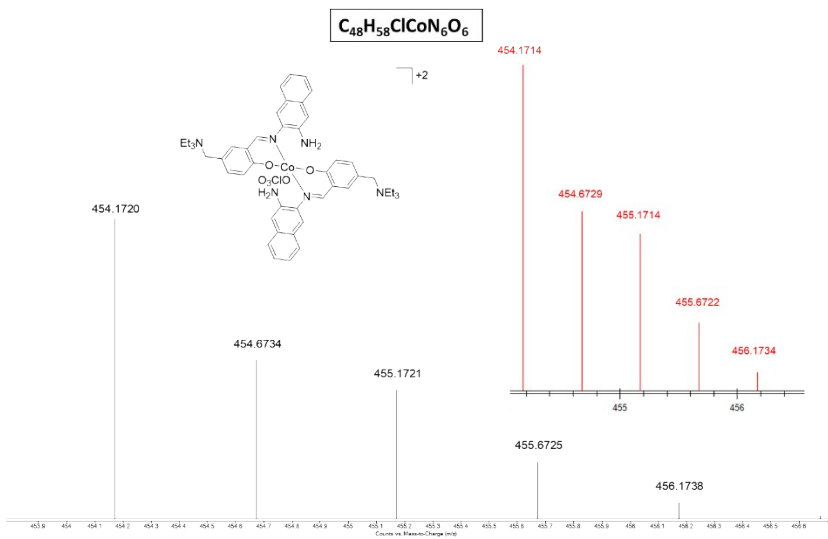


**Figure S11.**  $^1\text{H}$ - $^{13}\text{C}$  HSQC NMR of **1** in  $d_6$ -DMSO.

a)

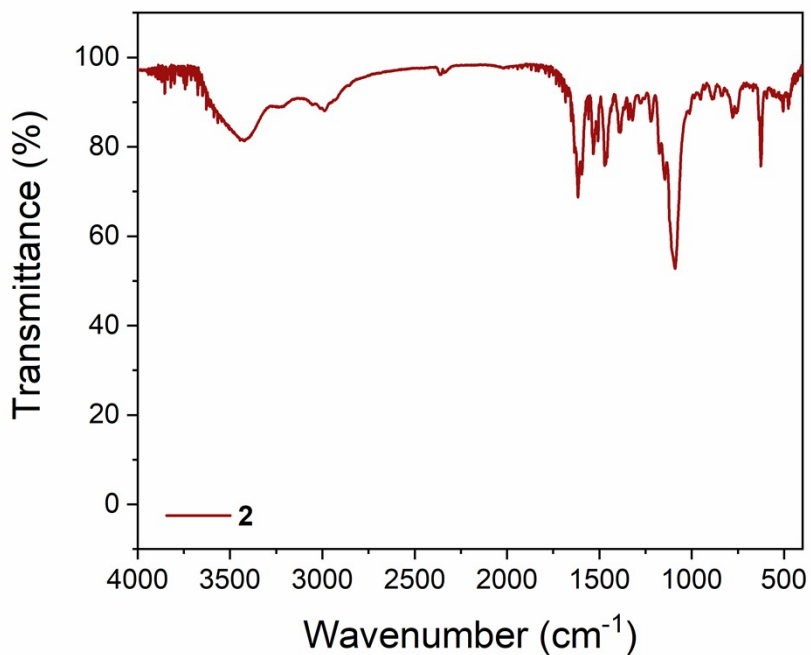


b)

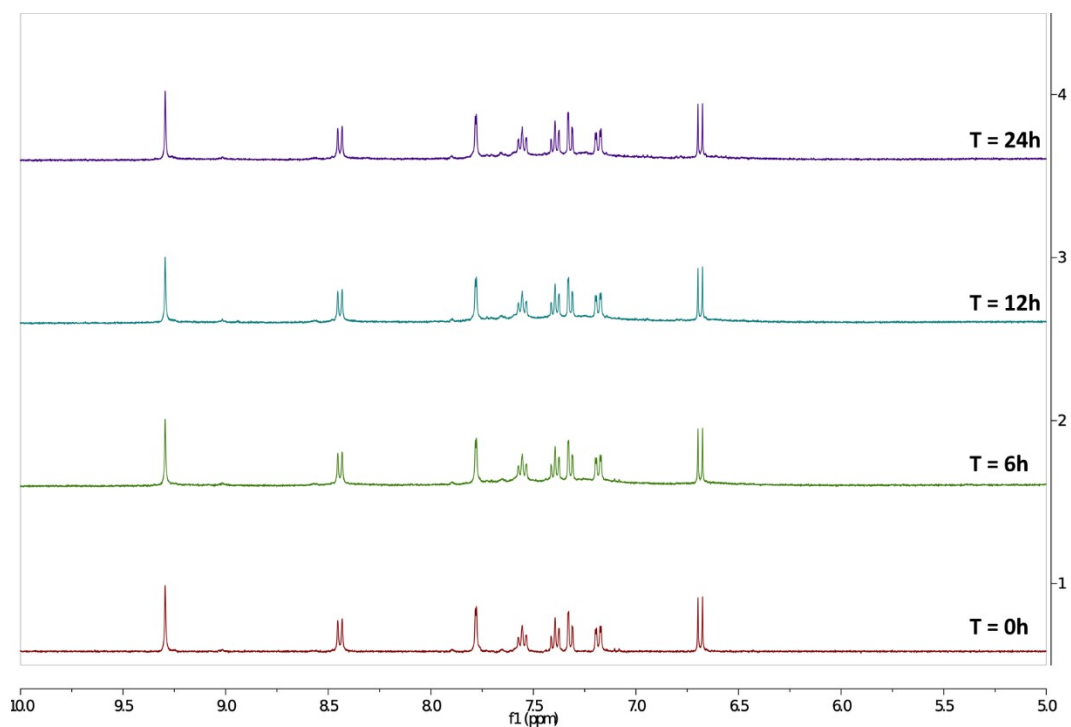


**Figure S12.** ESI-MS spectra of the two species found for **2** in acetonitrile:  $[2]^+$  (a),  $[2-\text{ClO}_4]^{+2}$  (b). Simulated spectra are depicted in red as insets.

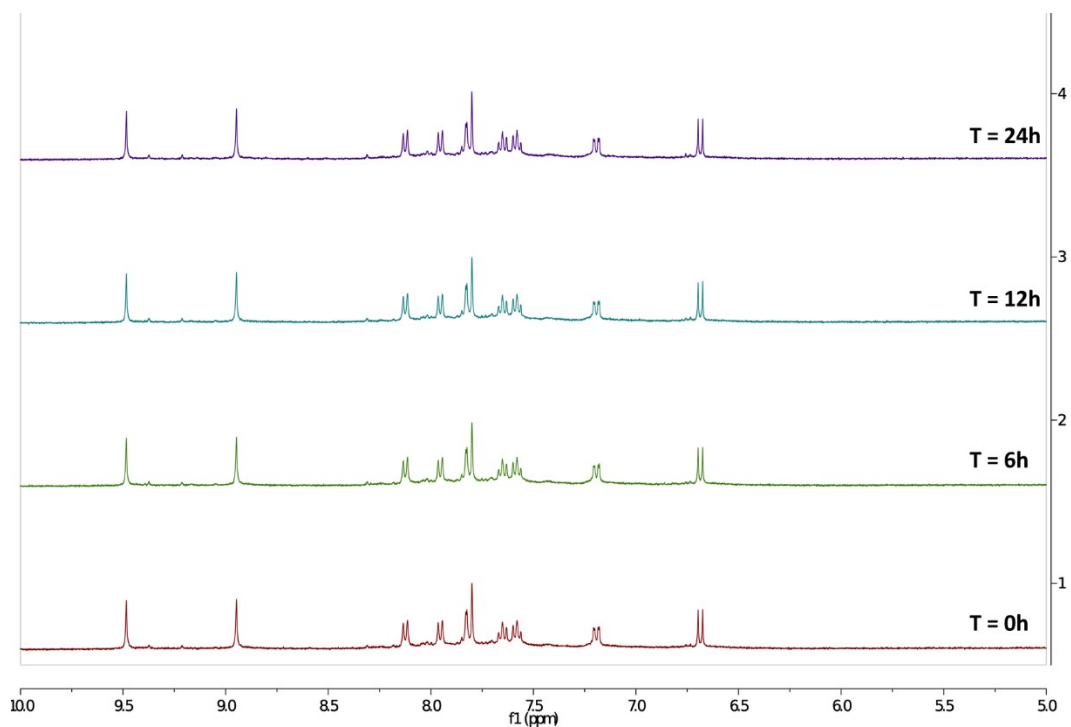




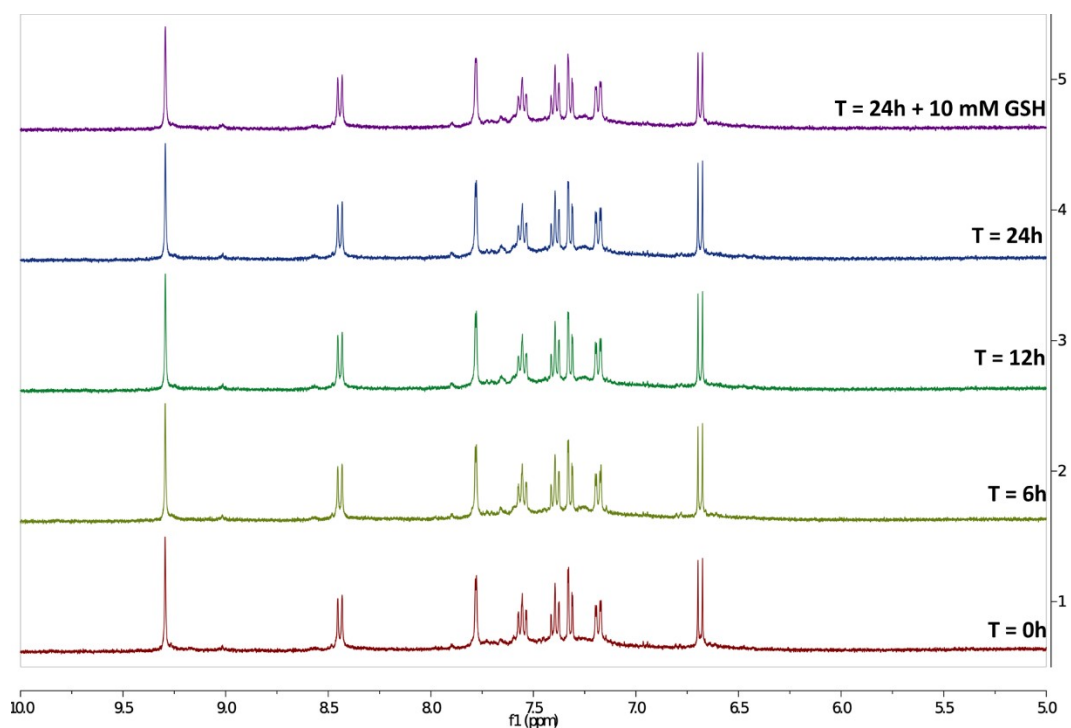
**Figure S13.** IR spectrum of **2** in KBr.



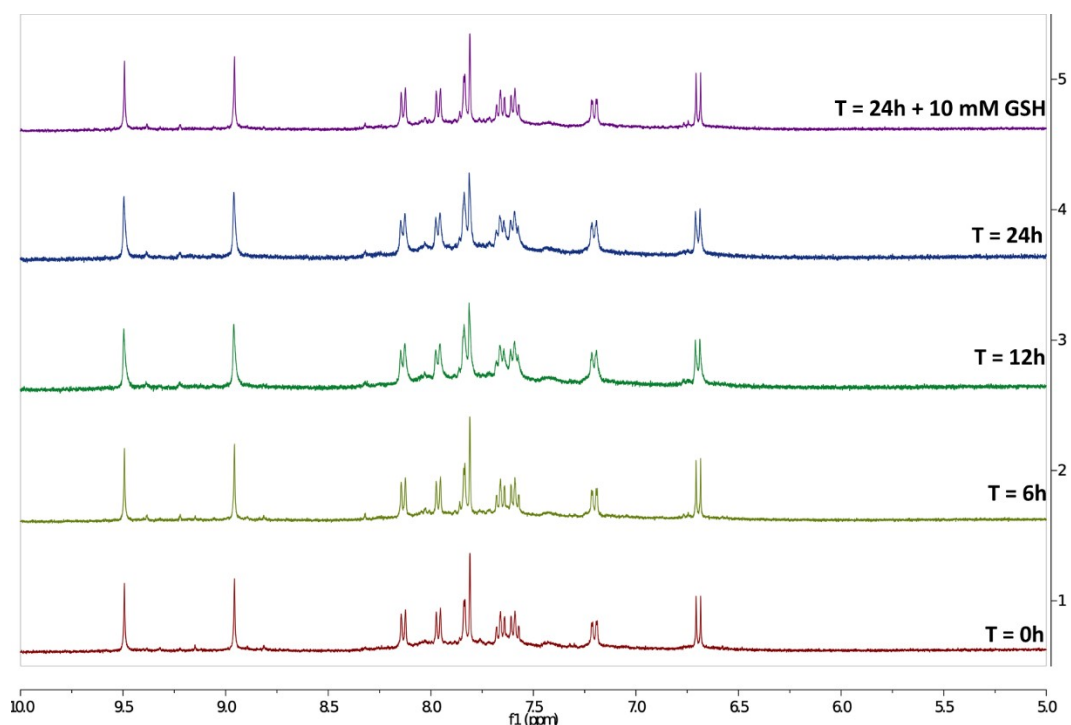
**Figure S14.**  $^1\text{H}$  NMR stability studies of **1** in  $d_6$ -DMSO: $\text{D}_2\text{O}$  (80:20) over 24 h. The selected timepoints are reported on the right.



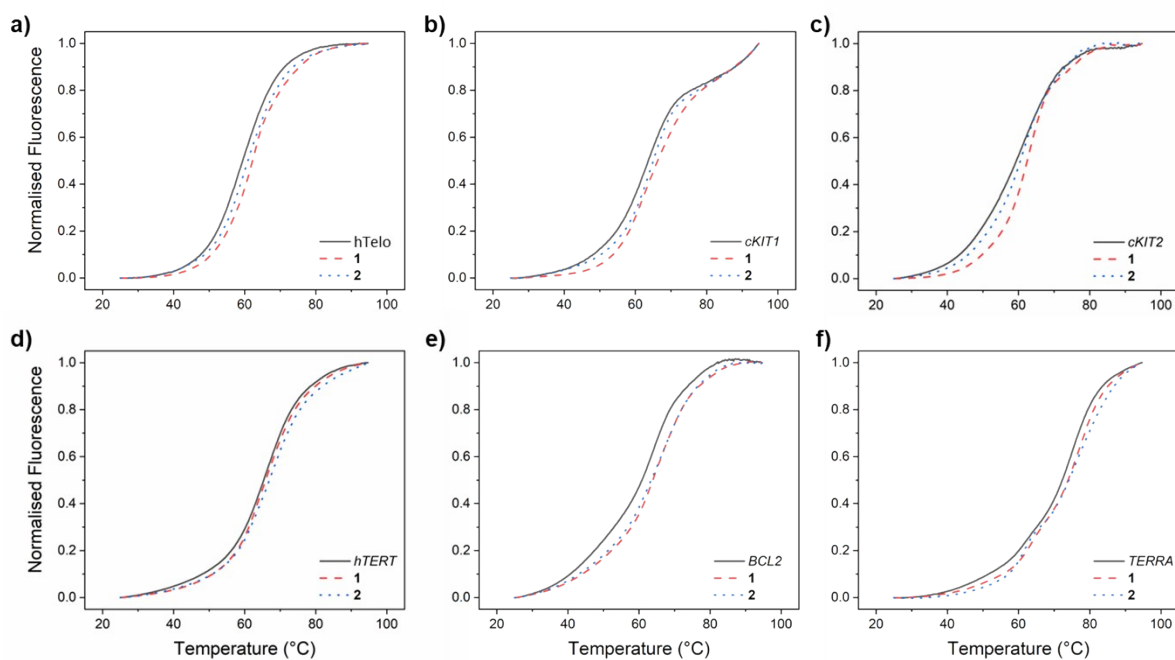
**Figure S15.**  $^1\text{H}$  NMR stability studies of **2** in  $d_6$ -DMSO: $\text{D}_2\text{O}$  (80:20) over 24 h. The selected timepoints are reported on the right.



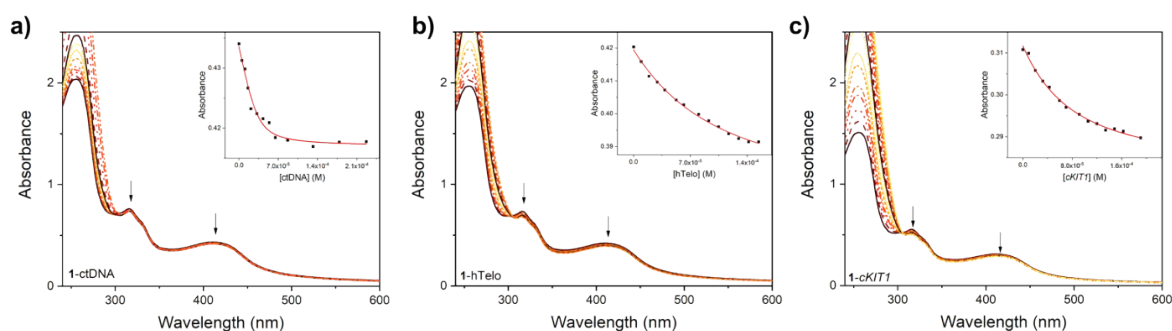
**Figure S16.**  $^1\text{H}$  NMR stability studies of **1** recorded over 24 h in a 2 mM GSH  $d_6$ -DMSO: $\text{D}_2\text{O}$  (80:20) mixture. In the top spectrum,  $^1\text{H}$  NMR stability studies of **1** in the presence of an excess of GSH (10 mM), collected in the same solvent as above. The selected timepoints are reported on the right.



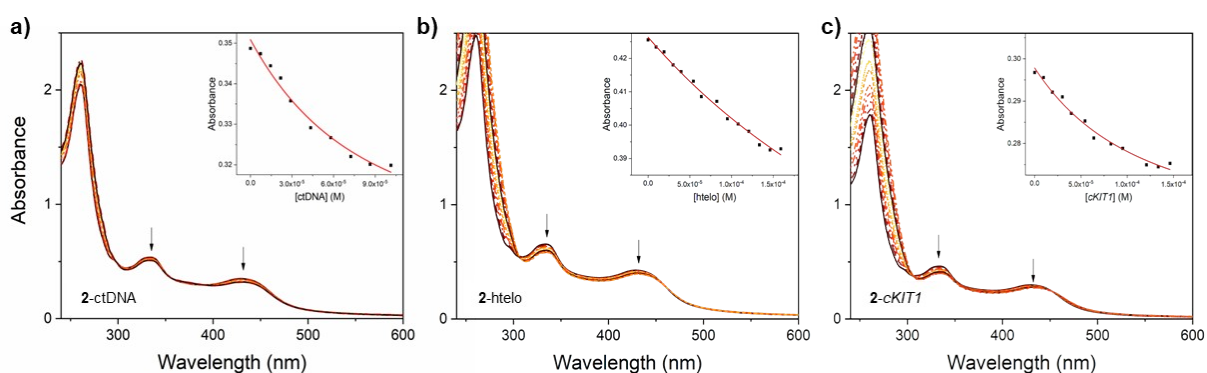
**Figure S17.**  $^1\text{H}$  NMR stability studies of **2** recorded over 24 h in a 2 mM GSH  $d_6$ -DMSO: $\text{D}_2\text{O}$  (80:20) mixture. In the top spectrum,  $^1\text{H}$  NMR stability studies of **1** in the presence of an excess of GSH (10 mM), collected in the same solvent as above. The selected timepoints are reported on the right.



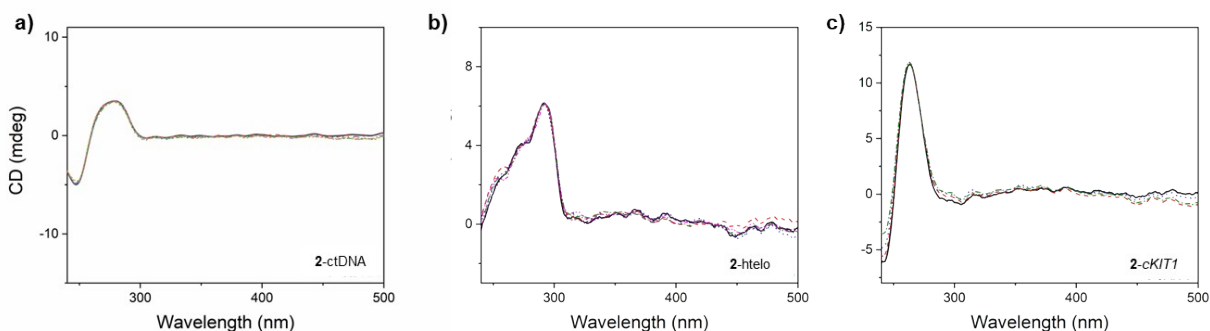
**Figure S18.** Representative FRET melting profiles of 200 nM hTelo (a), *cKIT1* (b), *cKIT2* (c), *hTERT* (d) *BCL2* (e) or *TERRA* (f) G4-DNA alone (black lines) and in the presence of 5 equivalents of the **1** or **2**, in 60 mM KAcodylate buffer (pH = 7.4).



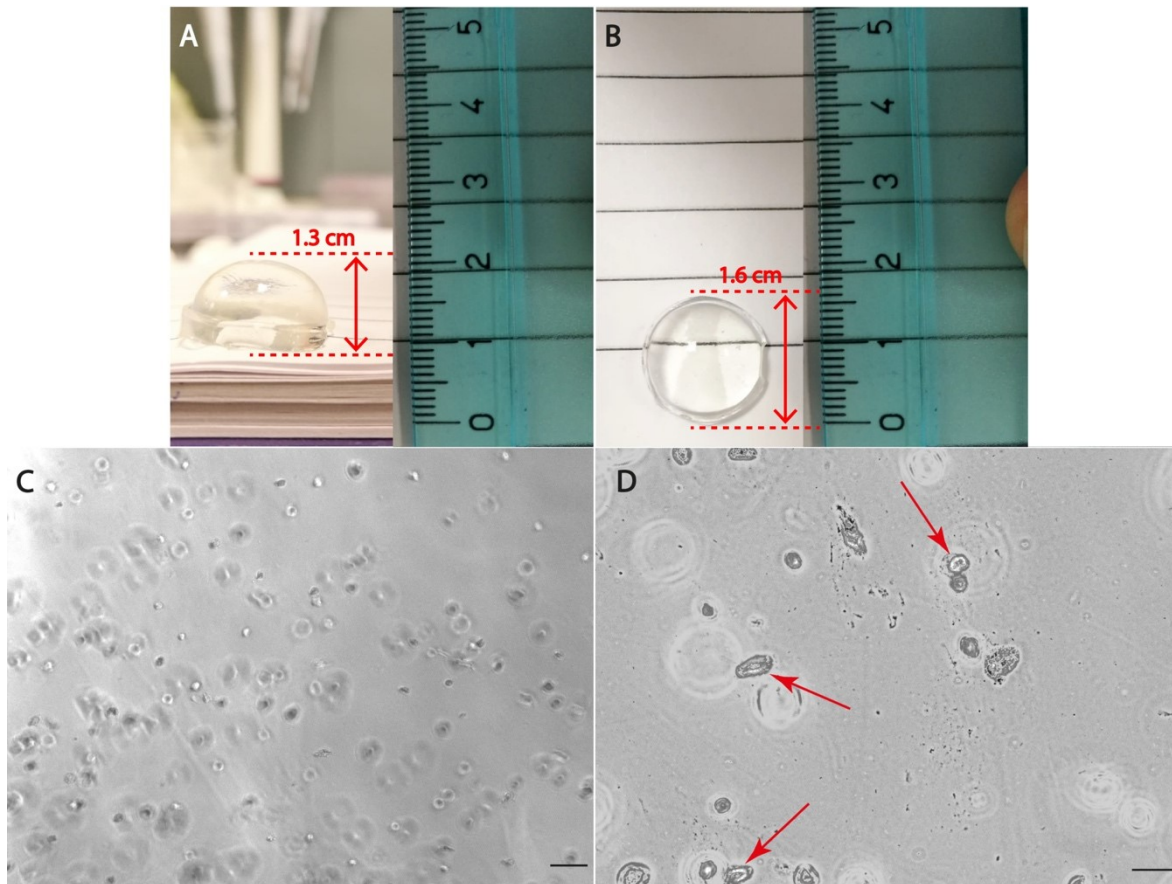
**Figure S19.** UV-Vis spectra of **1** in the presence of increasing amount of DNA collected in 100 mM KCl and 50 mM Tris-HCl buffer. (a)  $[\mathbf{1}] = 33.0 \mu\text{M}$ ,  $[\text{ctDNA}] = 0.0 - 227.0 \mu\text{M}$ ; (b)  $[\mathbf{1}] = 31.0 \mu\text{M}$ ,  $[\text{hTelo}] = 0.0 - 7.0 \mu\text{M}$ ; (c)  $[\mathbf{1}] = 23.7 \mu\text{M}$ ,  $[\text{cKIT1}] = 0.0 - 8.7 \mu\text{M}$ . In the inset representative data fits.



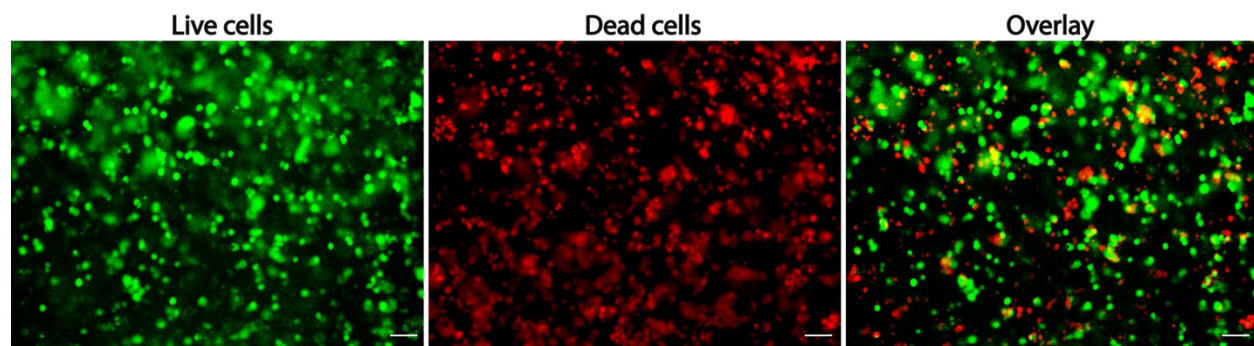
**Figure S20.** UV-Vis spectra of **2** in the presence of increasing amount of DNA collected in 100 mM KCl and 50 mM Tris-HCl buffer. (a)  $[\mathbf{2}] = 27.5 \mu\text{M}$ ,  $[\text{ctDNA}] = 0.0 - 101.0 \mu\text{M}$ ; (b)  $[\mathbf{2}] = 33.5 \mu\text{M}$ ,  $[\text{hTelo}] = 0.0 - 7.2 \mu\text{M}$ ; (c)  $[\mathbf{2}] = 23.5 \mu\text{M}$ ,  $[\text{cKIT1}] = 0.0 - 6.7 \mu\text{M}$ . In the inset representative data fits.



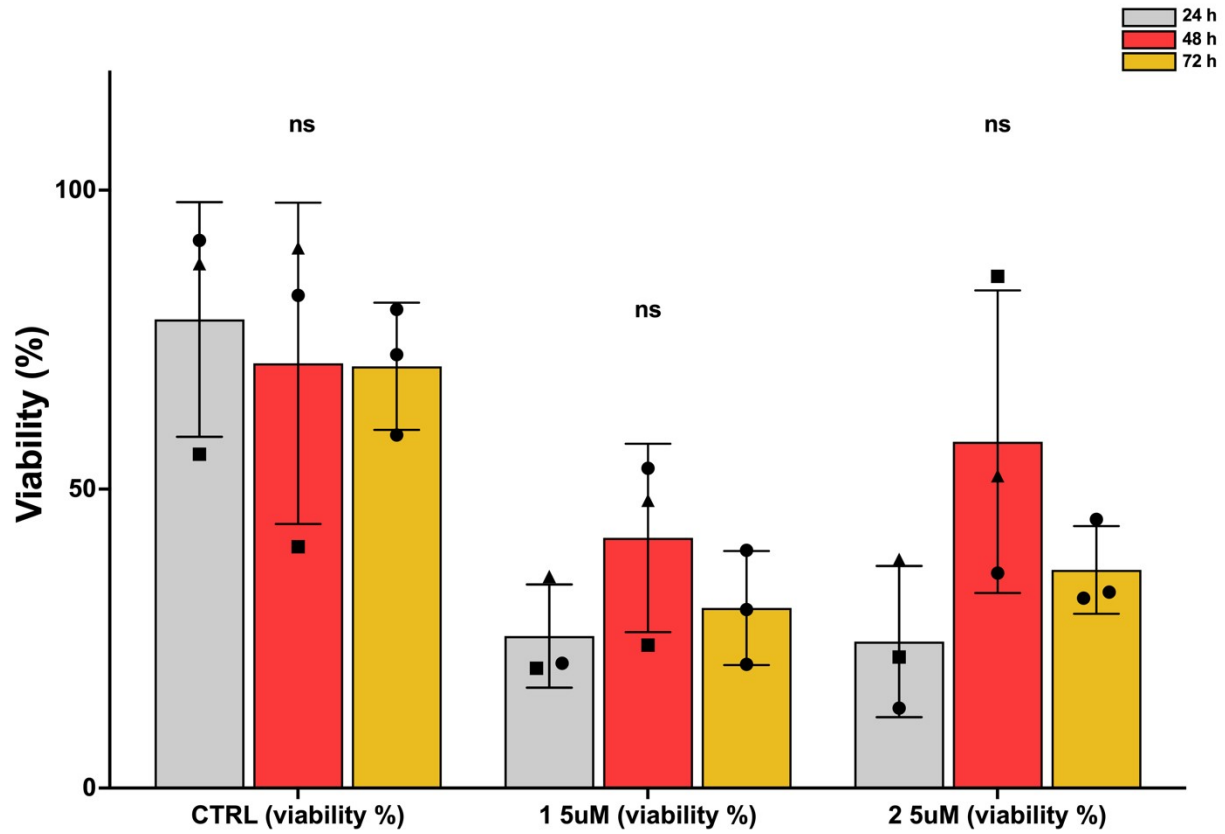
**Figure S21.** CD spectra of duplex- and G4-DNA solutions recorded in 100 mM KCl and 50 mM Tris-HCl buffer, in the presence of increasing amounts of **2**. (a)  $[\text{ctDNA}] = 50.0 \mu\text{M}$ ,  $[\mathbf{2}] = 0.0 - 39.7 \mu\text{M}$ ; (b)  $[\text{hTelo}] = 1.5 \mu\text{M}$ ,  $[\mathbf{2}] = 0.0 - 8.0 \mu\text{M}$ ; (c)  $[\text{cKIT1}] = 1.3 \mu\text{M}$ ,  $[\mathbf{2}] = 0.0 - 31.0 \mu\text{M}$ .



**Figure S22.** Macroscopic image of a GelMA scaffold in frontal (A), and top (B) view. Rounded shaped SW-1353 cells cultured within the scaffold at day 0 (C) and day 7 (D). Pericellular matrix deposition is clearly visible at day 7 (D, red arrows). Scale bar = 50  $\mu\text{m}$  (C) and 25  $\mu\text{m}$  (D).



**Figure S23.** Representative images from LIVE/DEAD assay performed on GelMA scaffolds cellularized with SW-1353 cells captured by fluorescence microscopy. The assay relies on a mixture of two fluorescent probes, calcein AM and ethidium homodimer (EthD-1), able to selectively stain live and dead cells respectively. Scale bar = 100  $\mu\text{m}$ .



**Figure S24.** Graphical representation of viability data grouped by time-point. Two-way ANOVA followed by Tukey's post-hoc test. Ns = not significant