On the mechanism of sp2 C-H borylation using ortho-Nsubstituted pyridinium cation

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Supplementary Information

Table of content

General considerations

Unless otherwise noted, all reactions and manipulations were performed under an argon atmosphere in a glovebox (Mbraun Unilab) or using standard Schlenk techniques with dried and degassed solvents. All Schlenk flasks and materials were dried prior to their use. All solvents were purified by common distillation techniques: Dichloromethane (CH_2Cl_2) , dimethylformamide (DMF), ethyl acetate (EtOAc) and dichloroethane (DCE) were distilled over CaH₂. n-Hexane (C₆H₁₄), toluene (C₇H₈) and diethyl ether (Et₂O) were distilled under argon over sodium/benzophenone. d6-Benzene (C_6D_6) , d8-Toluene (C_7D_8) , CDCl₃ and CD_2Cl_2 were dried over molecular sieves (4Å). Deuterated solvents were purchased from Eurisotop. [Li(Et2O)2,5][B(C6F5)4] (lithium tetrakis(pentafluorophenyl)borate diethylether complex (1:2.5), 97%) was purchased from ABCR and used as received. Catecholborane (CatBH), pentafluoropiridine, morpholine, 2-bromopyridine, N,N-dimethylpyridin-2-amine, benzo[b]thiophene n-butyllithium (n-BuLi, 2M, cyclohexane solution), thiophene, 2 methylthiophene, 3-methylthiophene, lithium dimethylamide (LiNMe₂), 2-fluoropiridine, piperidine, aniline, 1-phenylpyrrolidine, 1-phenylpiperidine, hexamethylbenzene (HMB), 3,4 ethylenedioxythiophene, HCl (1M, diethylether solution), bromobenzene, N-phenylpyrrole, B(C₆F₅)₃, 4-phenylmorpholine and 2,2,6,6-tetramethylpiperidine (TMP) were purchased from Sigma-Aldrich, TCI, Alfa Aesar, Fluorochem and were used as received

All new compounds have been characterized by ${}^{1}H$, ${}^{13}C$, ${}^{19}F$, ${}^{11}B$ NMR spectroscopy and ESI-HRMS. The structures of compound **3a** have been authenticated by Xray diffraction studies and their corresponding CIF files deposited in the Cambridge Crystallographic Data Centre with nos. CCDC-2288120.

Borylations were performed in J. Young valve NMR tubes purchased from Fisher Scientific. Schlenk vessels were equipped with gas-tight Teflon valves and Glindemann PTFE sealing rings.

NMR spectra were recorded at Bruker Avance NEO 400 MHz or 500 MHz spectrometers. If not otherwise stated: NMR spectra were recorded at 25 $^{\circ}$ C; ¹³C spectra were ¹H decoupled; ¹¹B spectra were not ¹H decoupled. Chemical shifts for the ¹H and ¹³C spectra were referenced to the residual ¹H/¹³C resonances of the deuterated solvent: C₆D₆: δ = 7.16; δ = 128.06; CD₂Cl₂: δ = 5.32; δ = 53.84; CDCl₃: δ = 7.26; δ = 77.16; and are reported as ppm relative to Me₄Si. Chemical shifts for the ¹¹B and ¹⁹F spectra were referenced to external standard (BF3·Et2O, CFCl3, respectively).

Gas chromatographic analyses were carried out an Agilent 6890N GC equipped with 5973 MSD using a HP-5MS UI (30m, 0.25 mm, 0.25 μm) column.

High-resolution mass spectra (ESI-HRMS) were recorded on Bruker microTOF mass spectrometer by electrospray ionization time of flight reflectron experiments. The mass detector was calibrated with HCOONa solution in 1:1 H_2O : iPrOH. All HRMS experiments were recorded with positive and negative ion detector and the recorded masses were under 5 ppm error unless otherwise noted.

Preparation

4-butyl-2,3,5,6-tetrafluoropyridine (1)

Compound **1** was synthesized by modifying the published procedure.¹ To a solution of pentafluoropyridine (3 g, 17.75 mmol) dissolved in diethyl ether F 40 mL at -93 °C (frozen acetone bath) in a 100 mL Schlenk tube, n-BuLi (9.76 mL, 19.52 mmol, 2M solution in cyclohexane) was added dropwise F N F over 30 min via syringe. The reaction was then allowed to warm up to room

temperature while stirring overnight immersed in the cooling bath. After 16 hours, the redorange solution was extracted with CH_2Cl_2 (3x50 mL). Combined organic layers were washed with Brine and dried over $MqSO₄$. After drying, $MqSO₄$ filtered on a short silica pad and the volatiles were removed in the rotary evaporator. The residue yellowish oil was purified by flash column chromatography on silica gel (0→5% CH₂Cl₂ in Hexane, v/v) to afford colorless liquid **1** (3.12 g, 85%). R_f 0.5 (CH₂Cl₂/Hexane – 1:19, v/v). Spectroscopic data are in agreement with literature².

¹**H** NMR (400 MHz, CDCl₃, 25 °C) δ 2.81 (tt, *J* = 7.7, 1.5 Hz, 2H, CH₂CH₂CH₂CH₃), 1.63 (p, *J* = 7.5 Hz, 2H, CH₂CH₂CH₂CH₃), 1.40 (dq, *J* = 14.6, 7.3 Hz, CH₂CH₂CH₂CH₃), 0.96 (t, *J* = 7.3 Hz, 3H, CH2CH2CH2C**H3**).

¹⁹F { ¹H} NMR (376 MHz, CDCl3, 25 °C) δ -92.09 (m), -145.74 (m).

¹³C { ¹H} NMR (101 MHz, CDCl3, 25 °C) 14.25, 22.81, 23.62, 31.74, 135.62, 139.55, 139.20, 142.03, 144.62.

4-Butyl-3,5,6-trifluoro-*N***,***N***-dimethylpyridin-2-amine (2a)**

Compound **2a** was synthesized by modifying the published procedure.¹ To a suspension of lithium dimethylamide (285 mg, 5.31 mmol) in diethyl ether 20 mL at -93 °C (frozen acetone bath) in a 100 mL Schlenk tube, **1** (0.8 mL, 4.83 mmol) was added dropwise via syringe. The reaction was allowed to warm up to room temperature while stirring overnight N⁷ immersed in the cooling bath. After 48 hours, formed red-orange solution was extracted with CH_2Cl_2 (3x50 mL). Combined organic layers were

washed with Brine and dried over MgSO₄. After drying, MgSO₄ filtered on a short silica pad and the volatiles were removed in the rotary evaporator. The residue oil was purified by flash column chromatography on silica gel (5 \rightarrow 25% CH₂Cl₂ in Hexane, v/v) to afford colorless liquid **2a** (868 mg, 77%). R_f 0.33 (CH₂Cl₂/Hexane – 1:9, v/v). Spectroscopic data are in agreement with literature $^{\rm 1}.$

¹H NMR (400 MHz, CDCl3, 25 °C) δ 3.00 (s, 3H, NC**H3**), 3.00 (s, 3H, NC**H3**), 2.68 (tt, *J* = 7.7, 1.7 Hz, 2H, C**H2**CH2CH2CH3), 1.58 (p, *J* = 7.4 Hz, 2H, C**H2**CH2CH3), 1.38 (h, *J* = 7.3 Hz, 2H, C**H2**CH3), 0.94 (t, *J* = 7.3 Hz, 3H, C**H3**).

¹⁹F { ¹H} NMR (376 MHz, CDCl3, 25 °C) δ -94.18 (dd, *J* = 31.2, 25.7 Hz), -137.57 (d, *J* = 31.2 Hz), -158.77 (d, *J* = 25.7 Hz).

¹³C {¹H} NMR (101 MHz, CDCl₃, 25 °C) δ 145.58 (d, J = 17.1 Hz), 143.64 – 142.68 (m), 144.60 – 141.43 (m), 136.93 – 133.11 (m), 131.75 (ddd, J = 20.0, 16.5, 3.3 Hz), 40.15, 40.09, 31.05, 23.24, 22.52, 13.85.

4-butyl-2,3,5-trifluoro-6-(piperidin-1-yl)pyridine (2b)

To a solution of piperidine (324 mg, 3.8 mmol) in diethyl ether 10 mL at 0 °C in a 50 mL Schlenk tube, n-BuLi (2 mL, 4 mmol, 2M in cyclohexane solution) was added dropwise via syringe. After 10 minutes stirring, mixture was frozen to -93°C (frozen acetone bath). Upon -93°C, **1** (750 mg, 3.62 mmol) was added via syringe. After 48 N hours, formed red-orange solution was extracted CH₂Cl₂ (3x50 mL). Combined organic layers were washed with Brine and dried over MgSO4. After drying, MgSO⁴ filtered on a short silica pad and the volatiles were removed in the rotary evaporator. The residue oil was

purified by flash column chromatography on silica gel (5 \rightarrow 25% CH₂Cl₂ in Hexane, v/v) to afford colorless liquid **2b** (596 mg, 60%). R_f 0.15 (CH₂Cl₂/Hexane – 1:19, v/v).

¹H NMR (400 MHz, C6D6, 25 °C) δ 3.24 (m, 4H, C**H2**NC**H2**), 2.43 (tt, J = 7.7, 1.7 Hz, 2H, C**H2**CH2CH2CH3), 1.41 (m, 6H, C**H2**C**H2**C**H2**), 1.29 (m, 2H, C**H2**CH2CH3), 1.14 (h, J = 7.3 Hz, 2H, C**H2**CH3), 0.76 (t, J = 7.3 Hz, 3H, C**H3**).

¹⁹F { ¹H} NMR (376 MHz, C6D6, 25 °C) δ -93.20 (dd, J = 31.6, 26.0 Hz), -135.70 (d, J = 31.9 Hz), -156.69 (d, $J = 26.4$ Hz).

¹³C { ¹H} NMR (101 MHz, C6D6, 25 °C) δ 146.37, 144.11 (d, J = 251.4 Hz), 143.74 (d, J = 65.0 Hz), 136.35 (dd, J = 250.1, 31.1 Hz), 131.77 (m), 49.11 (d, J = 5.4 Hz), 31.11, 25.94, 24.78, 23.22, 22.57, 13.81.

ESI-HRMS m/z: exp. for 273.1566 [M+H]⁺ (calcd. for C₁₄N₂F₃H₂₀ 273.157

4-(4-butyl-3,5,6-trifluoropyridin-2-yl)morpholine (2c)

To morpholine (205 mg, 5.8 mmol) in diethyl ether 10 mL at 0 °C in a 50 ml Schlenk tube, n-BuLi (2.9 mL, 5.8 mmol, 2M solution in cyclohexane) was added dropwise via syringe. After 10 minutes stirring, mixture was frozen to -93°C (frozen acetone bath). Upon -93°C, **1** (1 g, 4.83 mmol) was added via syringe. After 18 hours, formed red-orange solution was extracted with CH_2Cl_2 (3x50 mL). Combined organic layers were washed with Brine and dried over $MgSO₄$. After drying, $MgSO₄$ filtered on a short silica pad and the

volatiles were removed in the rotary evaporator. The residue oil was purified by flash column chromatography on silica gel (0→2% EtOAc in CH2Cl2, v/v) to afford colorless liquid **2c** (650 mg, 49%). R_f 0.4 (CH₂Cl₂).

¹H NMR (400 MHz, CDCl3, 25 °C) δ 3.86 – 3.76 (m, 4H, morpholine), 3.42 – 3.32 (m, 4H, morpholine), 2.70 (t, J = 7.6 Hz, 2H, CH₂CH₂CH₂CH₃), 1.58 (p, J = 7.5 Hz, 2H, C**H2**CH2CH3), 1.38 (h, J = 7.3 Hz, 2H, C**H2**CH3), 0.94 (t, J = 7.3 Hz, 3H, C**H3**). **¹⁹F { ¹H} NMR** (376 MHz, CDCl3, 25 °C) δ -93.41 (dd, J = 31.2, 25.0 Hz), -135.81 (d, J = 31.2 Hz), -154.98 (d, J = 25.7 Hz). **¹³C { ¹H} NMR** (101 MHz, CDCl3, 25 °C) δ 145.83, 144.28 (d, J = 252.1 Hz), 142.98 (d, J = 132.2 Hz), 138.98 – 134.44 (m), 132.86 – 131.09 (m), 66.84, 48.27, 30.99, 23.24, 22.50,

13.83.

ESI-HRMS m/z: exp. for 297.1190 [M+Na]⁺ (calcd. for C₁₃H₁₇F₃N₂ONa 297.1185)

4-butyl-2,3,5-trifluoro-6-(piperidin-1-yl)pyridine (2d)

Compound **2d** was synthesized by modifying the published procedure.³ A 50 mL round-bottom flask was charged with 2-bromopyridine (1.5 g, 9.49 mmol), CuI (180.8 mg, 0.95 mmol), 2-picolinic acid (233.7 mg, 1.90 mmol), anhydrous K_2CO_3 (1.97 g, 14.24 mmol), piperidine (808.4 mg, 9.49 mmol) and DMF (8 mL). Then, the flask was equipped with bulb condenser and the reaction mixture was degassed and purged with argon gas 3 times. The reaction was heated in the oil bath at 130°C and

stirred for 48 h. Then, the reaction mixture was cooled to room temperature, diluted with water (20 mL) and extracted with ethyl acetate (3x20 mL). The combined organic layer was washed with water (3x20 mL), brine (50 mL) and dried over anhydrous $MqSO₄$. After drying, MgSO⁴ filtered on a short silica pad and the volatiles were removed in the rotary evaporator. The residue oil was purified by flash column chromatography on silica gel ($0\rightarrow 10\%$ EtOAc in Hexane, v/v) to afford colorless liquid **2d** (1,003 g, 65%). R_f 0.5 (EtOAc/Hexane – 1:9, v/v). Spectral data are in agreement with literature³.

¹H NMR (400 MHz, CDCl3, 25 °C) δ 8.17 (ddd, J = 4.9, 2.1, 1.0 Hz, 1H), 7.43 (ddd, J = 8.8, 7.1, 2.0 Hz, 1H), $6.66 - 6.61$ (m, 1H), 6.54 (ddd, $J = 7.2$, 4.9, 1.0 Hz, 1H), $3.56 - 3.48$ (m, 4H, C**H2**NC**H2**), 1.69 – 1.60 (m, 6H, C**H2**C**H2**C**H2**). **¹³C { ¹H} NMR** (101 MHz, CDCl3, 25 °C) δ 159.89, 148.07, 137.41, 112.51, 107.22, 46.44, 25.64, 24.87.

2-(2,2,6,6-tetramethylpiperidin-1-yl)pyridine (2e)

To 2,2,6,6-tetramethylpiperidine (8.08 mg, 61.8 mmol) in diethyl ether 20 mL at 0 °C in a 100 mL Schlenk tube, n-BuLi (8.24 mL, 20.6 mmol, 2.5M solution in hexane) was added dropwise via syringe. After 10 minutes stirring, 2-fluoropyridine (1 g, 10.3 mmol) was added via syringe. After 2 hours, reaction was extracted with EtOAc (3x50 mL). Combined organic layers were washed with Brine and dried over MgSO4. After drying, MgSO⁴ filtered on a short silica pad and the volatiles were removed in

the rotary evaporator. The residue oil was purified by flash column chromatography on silica gel (0→10% EtOAc in Hexane, v/v) to afford colorless liquid **2e** (802 mg, 36%). R^f 0.5 (EtOAc/Hexane – 1:9, v/v).

¹H NMR (400 MHz, CDCl3, 25 °C) δ 8.49 (ddd, J = 4.8, 2.2, 0.8 Hz, 1H), 7.59 – 7.52 (m, 1H), 7.07 (ddd, J = 7.4, 4.8, 1.2 Hz, 2H), 7.03 (dt, J = 7.9, 1.1 Hz, 1H), 1.79 – 1.72 (m, 1H), 1.58 – 1.53 (m, 3H), 1.09 (s, 12H).

¹³C { ¹H} NMR (101 MHz, CDCl3, 25 °C) δ 18.32, 29.35, 41.75, 54.40, 120.68, 128.81, 136.23, 147.96, 161.04.

ESI-HRMS m/z: exp. for 219.1861 [M+H]⁺ (calcd. for C₁₄H₂₃N₂ 219.1816)

Compounds **3a-f** were synthesized according to the published procedure.¹ $[Li(Et2O)2,5][B(C6F5)4]$ was used instead of $[Li(Et2O)4][B(C6F5)4]$.

To a solution of corresponding aminopyridine (1 eq) in CH_2Cl_2 5 mL in a 50 mL flask, HCl (1 eq, ether solution) was added. After 5 minutes, a solution of [Li(Et2O)_{2,5}][B(C₆F₅)₄] (1 eq) in CH_2Cl_2 10 ml was added. The solution was stirred for 30 min, the precipitate formed was filtered and washed with CH_2Cl_2 5 mL. Mother liquor was removed in the rotary evaporator and dried in vacuum (1 mbar) at room temperature for 1h. The solid was redissolved in $CH₂Cl₂$ 10 mL, and the filtration and evaporation procedure was repeated. Dried in vacuum (1 mbar) at room temperature for 1 h gave 77-96% of powder **3a-f**.

4-Butyl-2-(dimethylamino)-3,5,6-trifluoropyridinium tetrakis(pentafluorophenyl)borate(1-) (3a)

Compound **3a** was synthesized according to the general procedure using aminopyridine **2a** (342 mg, 1.47 mmol).¹ The yield is 96% (1,29 g).

¹H NMR (400 MHz, CD₂Cl₂, 25 °C) δ 4.38 (br. s, 1H, NH), 3.38 (s, 6H, NH(CH₃)₂), 2.87 (tt, *J* = 7.7, 1.5 Hz, 2H, C**H2**CH2CH2CH3), 1.63 (p, *J* = 7.5 Hz, 2H, C**H2**CH2CH3), 1.41 (h, *J* = 7.3 Hz, 2H, C**H2**CH3), 0.95 (t, *J* = 7.3 Hz, 3H, C**H3**).

¹⁹F { ¹H} NMR (376 MHz, CD2Cl2, 25 °C) δ -85.28 (dd, J = 28.4, 21.5 Hz, 1F, cation), -132.30 (d, $J = 18.0$ Hz, 1F, cation), -133.18 - -133.44 (m, 8F, anion), -134.22 (dd, $J = 28.4$, 8.3 Hz, 1F, cation), -163.41 (t, J = 20.5 Hz, 4F, anion), -167.45 (t, J = 20.1 Hz, 8F, anion). **¹¹B NMR** (128 MHz, CD₂Cl₂, 25 °C) δ -16.68.

¹³C { ¹H} NMR (101 MHz, CD2Cl2, 25 °C) δ 13.59, 22.79, 24.03, 30.79, 46.10, 135.66, 138.12, 147.38, 149.67.

ESI-HRMS m/z: exp. for 233.1249 [M+H]⁺ (calcd. for C₁₁H₁₆N₂F₃ 233.1260)

ESI-HRMS m/z: exp. for 678.9779 [M][.] (calcd. for C₂₄F₂₀B 678.9772)

1-(4-butyl-3,5,6-trifluoropyridin-2-yl)piperidin-1-ium tetrakis(pentafluorophenyl)borate(1-) (3b)

Compound **3b** was synthesized according to the general procedure using aminopyridine **2b** (228 mg, 0.837 mmol).¹ The yield is 96% (764 mg).

¹**H** NMR (400 MHz, CD₂Cl₂, 25 °C) δ 3.69 (d, J = 5.6 Hz, 4H, CH₂NHCH₂), 3.51 (br. s., 1H, N**H**), 2.84 (tt, J = 7.7, 1.6 Hz, 2H, C**H2**CH2CH2CH3), 2.04 (p, J = 6.0 Hz, 4H, C**H2**CH2NHCH2C**H2**), 1.82 (p, J = 6.0 Hz, 2H, NHCH2CH2C**H2**), 1.61 (p, J = 7.5 Hz, 2H, C**H2**CH2CH3), 1.39 (h, J = 7.3 Hz, 2H, C**H2**CH3), 0.94 (t, J = 7.4 Hz, 3H, C**H3**). **19F {1H} NMR** (376 MHz, CD₂Cl₂, 25 °C) δ -86.43 (br.s., 1F, cation), -133.24 (m, 9F, anion + cation) -134.27 (dd, J = 28.4, 7.6 Hz, 1F, cation), -163.46 (t, J = 20.5 Hz, 4F, anion), - 167.46 (t, J = 19.8 Hz, 8F, anion). **¹¹B NMR** (128 MHz, CD₂Cl₂, 25 °C) δ -16.67. **¹³C { ¹H} NMR** (101 MHz, CD2Cl2, 25 °C) δ 13.60, 21.57, 22.76, 23.93, 24.33, 30.84, 56.52, 135.85, 137.94, 147.38, 149.75.

ESI-HRMS m/z: exp. for 273.1561 [M+H]⁺ (calcd. for C₁₄H₂₀N₂F₃ 273.1573)

ESI-HRMS m/z: exp. for 678.9761 [M]⁻ (calcd. for C₂₄F₂₀B 678.9772)

4-(4-butyl-3,5,6-trifluoropyridin-2-yl)morpholin-4-ium tetrakis(pentafluorophenyl)borate(1-) (3c)

Compound **3c** was synthesized according to the general procedure using aminopyridine **2c** (300 mg, 1.09 mmol).¹ The yield is 94% (980 mg).

¹**H** NMR (400 MHz, CDCl₃, 25 °C) δ 4.03 (t, J = 4.8 Hz, 4H, morpholine), 3.66 (t, J = 4.9 Hz, 4H, morpholine), 2.78 (t, J = 7.7 Hz, 2H, CH₂CH₂CH₂CH₃), 2.23 (s, 1H, NH), 1.60 – 1.50 (m, 2H, C**H2**CH2CH3), 1.34 (h, J = 7.3 Hz, 2H, C**H2**CH3), 0.88 (t, J = 7.3 Hz, 3H, C**H3**). **¹⁹F { ¹H} NMR** (376 MHz, CDCl3, 25 °C) δ -84.42 (br.s., 4F, cation), -131.21 (d, J = 22.2 Hz, 9F, cation + anion), -132.12 (br.s., 1F, cation), -161.16 (dq, J = 40.2, 20.5 Hz, 4F anion), -165.24 (br.s., 8F, anion).

11**B NMR** (128 MHz, CDCl₃, 25 °C) δ -14.76.

¹³C { ¹H} NMR (101 MHz, CDCl3, 25 °C) δ 13.16, 22.27, 23.42, 30.37, 31.52, 64.07, 134.97, 137.42, 146.82, 149.14.

ESI-HRMS m/z: exp. for 275.1360 [M+H]⁺ (calcd. for C₁₃H₁₈F₃N₂O 275.1366) **ESI-HRMS** m/z: exp. for 678.9785 [M]⁻ (calcd. for C₂₄F₂₀B 678.9772)

1-(pyridin-2-yl)piperidin-1-ium tetrakis(pentafluorophenyl)borate(1-) (3d)

Compound **3d** was synthesized according to the general procedure using aminopyridine **2d** (100 mg, 0.616 mmol).¹ The yield is 77% (400 mg).

1H NMR (400 MHz, CD₂Cl₂, 25 °C) δ 9.98 (br.s, 1H, NH), 7.98 – 7.91 (m, 1H), 7.69 (td, J = 6.5, 2.4 Hz, 1H), 7.14 (dd, J = 9.5, 2.6 Hz, 1H), 6.90 (t, J = 6.8 Hz, 1H), 3.64 – 3.55 (m, 4H C**H2**NHC**H2**), 1.83 – 1.76 (m, 6H, C**H2**C**H2**C**H2**).

¹⁹F { ¹H} NMR (376 MHz, CD2Cl2, 25 °C) δ -133.01 – -133.34 (m, 8F), -163.53 (t, J = 20.5 Hz, 4F), -167.47 (t, J = 20.1 Hz, 8F).

11**B NMR** (128 MHz, CD₂Cl₂, 25 °C) δ -16.67.

¹³C {¹H} NMR (101 MHz, CD₂Cl₂, 25 °C) δ 23.26, 25.22, 47.87, 113.35, 113.50, 135.07, 135.50, 145.32, 147.30, 149.75, 151.57.

ESI-HRMS m/z: exp. for 163.1232 [M+H]⁺ (calcd. for C₁₀H₁₅N₂ 163.1230)

ESI-HRMS m/z: exp. for 678.9737 [M]⁻ (calcd. for C₂₄F₂₀B 678.9772)

1-(pyridin-2-yl)-2,2,6,6-tetramethylpiperidin-1-ium tetrakis(pentafluorophenyl)borate(1-) (3e)

Compound **3e** was synthesized according to the general procedure using aminopyridine **2e** (100 mg, 0.458 mmol).¹ The yield is 95% (392 mg) of the crude product. Differents attempt to purify it was not successful.

1H NMR (400 MHz, CD₂Cl₂, 25 °C) δ 8.56 (d, J = 3.7 Hz, 1H), 8.18 – 8.07 (m, 1H), 7.70 (dd, J = 7.6, 4.8 Hz, 1H), 7.57 (d, J = 8.3 Hz, 1H), 6.75 (s, 1H, N**H**), 2.00 (m, 6H, C**H2**C**H2**C**H2**), 1.41 (br.s, 12H, C(C**H3**)2)2). **¹⁹F {¹H} NMR** (376 MHz, CD₂Cl₂, 25 °C) δ -133.02 – -133.14 (m), -163.61 (t, J = 20.5 Hz), $-167.37 - -167.60$ (m). **¹¹B NMR** (128 MHz, CD₂Cl₂, 25 °C) δ -16.66. **¹³C { ¹H} NMR** (101 MHz, CD2Cl2, 25 °C) δ 150.30, 148.59 (d, J = 244.1 Hz), 140.30, 136.74 (d, J = 245.2 Hz), 127.19 (d, J = 32.0 Hz), 40.05, 31.45, 22.29, 16.41. **ESI-HRMS** m/z: exp. for 219.1851 [M+H]⁺ (calcd. for C₁₄H₂₃N₂ 219.1856) **ESI-HRMS** m/z: exp. for 678.9735 [M]⁻ (calcd. for C₂₄F₂₀B 678.9772)

N,N-dimethylpyridin-2-aminium tetrakis(pentafluorophenyl)borate(1-) (3f)

Compound **3f** was synthesized according to the general procedure using *N*,*N*-dimethylpyridin-2-amine (100 mg, 0.819 mmol).¹ The yield is 87% (573 mg).

¹**H NMR** (400 MHz, CD₂Cl₂, 25 °C) δ 9.75 (s, 1H, NH), 7.97 (ddd, J = 8.9, 7.1, 1.7 Hz, 1H), 7.74 (ddd, J = 6.6, 1.8, 0.9 Hz, 1H), 7.06 (dt, J = 9.4, 0.9 Hz, 1H), 6.96 (td, J = 6.8, 1.1 Hz, 1H), 3.28 (s, 6H, C**H3**NHC**H3**).

¹⁹F { ¹H} NMR (376 MHz, CD2Cl2, 25 °C) δ -133.06 – -133.28 (m, 8F), -163.43 (t, J = 20.5 Hz, 4F), -167.21 – -167.67 (m, 8F).

11**B NMR** (128 MHz, CD₂Cl₂, 25 °C) δ -16.67.

13C {1H} NMR (101 MHz, CD₂Cl₂, 25 °C) δ 152.26, 148.54 (d, J = 236.5 Hz), 145.06, 138.63 (d, J = 244.1 Hz), 136.70 (d, J = 240.5 Hz), 135.01, 113.39, 112.81, 39.48. **ESI-HRMS** m/z: exp. for 145.0731 [M+Na]⁺ (calcd. for C₇H₁₀N₂Na 145.0736) **ESI-HRMS** m/z: exp. for 678.9768 [M]⁻ (calcd. for C₂₄F₂₀B 678.9772)

Borylations

Comparison of different ortho-N-substituted pyridinium catalysts in sp² C-H borylation of 3-methylthiophene

In a glovebox, J. Young-NMR tube was charged with the appropriate catalyst (**3a-f**) (5% mol.), 50 mg of 2-methylthiophene (0,5 mmol), 500 µl DCE and 100 µl C_6D_6 . HMB was measured individually for each reaction (3-5 mg) and added. Then, ¹H NMR was recorded from samples as a reference point. J. Young-NMR tubes were heated in the oil bath at 110°C for 24 h. After heating, ¹H and ¹¹B NMR were recorded from samples. Yields were calculated based on hexamethylbenzene signal and the most characteristic signal of the product on the ¹H NMR.

Optimization of sp² C-H borylation of 3-methylthiophene with catalyst 3a

All reactions were carried out in the J. Young-NMR tubes and were charged in the glovebox according to the presented table based on 50 mg of 3-methylthiophene. Before the reaction, ¹H NMR was acquired from samples as a reference point. After reaction, ¹H and ¹¹B NMR were recorded from samples. Convertions were calculated based on reference spectra and the spectra of reaction on the ¹H NMR.

Conversion higher than 75% is coloured green. DCE – 1,2-dichloroethane. a) Entries 26 and 27 are entries and 24 respectively with prolonged heating.

Substrate scope of sp² C-H borylation with catalyst 3a

In a glovebox, J. Young-NMR tube was charged with catalyst **3a** (5% mol.), corresponding substrate (exact mass is in the description of each spectrum for the substrate scope), 500 μ l DCE and 100 μ l C₆D₆. Hexamethylbenzene was measured individually for each reaction (3-6 mg) and added. Then, ¹H NMR was recorded from samples as a reference point (d_1 = 10 sec). J. Young-NMR tubes were heated in the oil bath at 110°C for 24 h. After heating, ¹H and ¹¹B NMR were recorded from samples (d_1 = 10 sec). After that, 10 µl aliquots were drawn and diluted in 1 mL EtOAc. Prepared samples were screened on GC-MS. Yields were calculated based on hexamethylbenzene (HMB) signal and the most characteristic signal of the product on the ¹H NMR.

signals are aligned to C_6D_6 . All signals are shifted from original place in C_6D_6 because of high amount DCE. Conversion is determined by integration of the signals of starting material from δ 2.43 and the products from δ 2.83 (1st product) and δ 2.50 (2nd product).

signals are aligned to C_6D_6 . All signals are shifted from original place in C_6D_6 because of high amount DCE. Conversion is determined by integration of the signals of starting material from δ 2.64 and the product from δ 2.71.

GC-MS data after reaction. Mass spectrum depicts peak of the product at 10.40 seconds.

are aligned to C_6D_6 . All signals are shifted from original place in C_6D_6 because of high amount DCE. Conversion is determined by integration of the signals of starting material from δ 7.49 and the product from δ 7.90 and δ

GC-MS data after reaction. Mass spectrum depicts peak of the product at 9.7 seconds.

signals are aligned to C_6D_6 . All signals are shifted from original place in C_6D_6 because of high amount DCE.

Conversion is determined by integration of the signals of starting material from δ 7.67-7.60 and the product from δ 7.26.

All signals are aligned to DCE at 3.79. All signals are shifted from original place in C_6D_6 because of high

amount DCE. Conversion is determined by integration of the signals of starting material from δ 2.99 and the product from δ 3.10.

GC-MS data after reaction. Mass spectrum depicts peak of the product at 12.66 seconds.

signals are aligned to C_6D_6 . All signals are shifted from original place in C_6D_6 because of high amount DCE. Conversion is determined by integration of the signals of starting material from δ 3.41 and the product from δ

S19

GC-MS data after reaction. Mass spectrum depicts peak of the product at 14.31 seconds.

¹H NMR spectra before reaction and after. 54.4 mg of 1-phenylmorpholine and 6.2 mg of HMB were added. All signals are aligned to DCE at 3.80. All signals are shifted from original place in C_6D_6 because of high amount DCE. Conversion is determined by integration of the signals of starting material from δ 3.26 and the

S21

GC-MS data after reaction. Mass spectrum depicts peak of the product at 14.34 seconds.

signals are aligned to C_6D_6 . All signals are shifted from original place in C_6D_6 because of high amount DCE.

Conversion is determined by integration of the signals of starting material from δ 6.52 and the product from δ 3.41.

GC-MS data after reaction. Mass spectrum depicts peak of the product at 14.28 seconds.

Mechanistic studies

Time-depemdent ¹H and ¹¹B NMR of C-H borylation of 3-methylthiophene with catalyst 3a

In a J. Young-NMR tube, 3-methylthiophene (50mg, 0.509 mmol), **3a** (18,59 mg, 0.020 mmol) and CatBH (61.08mg, 0.509 mmol) were mixed in 650 μ L C₆D₆ and shaked for 1 minute. Before heating, ¹H and ¹¹B NMR of sample were recorded as a reference. Further, J. Young-NMR tube were placed in oil bath at 80°C. After 2, 4, 8, 16, 24, 36 and 48 hours, ¹H and ¹¹B NMR of sample were recorded. Conversion was calculated based on integration of ¹H spectrum between 1.5 and 3.0 ppm. Hydrogen $(H₂)$ peak increased over time.

Stacked aliphatic part of ¹H NMR.

Stacked ¹H NMR of hydrogen molecule $(H₂)$ area.

Stochiometric reaction between CatBH and 3a at RT

In a J. Young-NMR tube, **3a** (70.5 mg, 0.077 mmol) and CatBH (9.3 mg, 0.077 mmol) were mixed in 700 μ L CD₂Cl₂ and shaked for 1 minute upon RT. After 45 minutes, NMR spectra were recorded. New broad singlet at 7.96 was observed. Hydrogen $(H₂)$ at 4.60 ppm was observed. Formation of Cat₃B₂ was observed at 22.38 ppm in ¹¹B NMR.

¹H NMR (400 MHz, CD_2Cl_2) δ 7.96 (s, 1H), 7.47 – 6.87 (m, 5H), 3.47 (s, 6H), 2.89 (s, 2H), 1.62 (s, 2H), 1.41 (s, 2H), 0.94 (s, 3H).

 19 F NMR (376 MHz, CD₂Cl₂) δ -84.44 (dd, J = 28.1, 21.2 Hz), -130.05 (dd, J = 21.5, 9.0 Hz), $-133.32, -134.26$ (dd, $J = 27.7, 9.0$ Hz), -163.26 (t, $J = 20.5$ Hz), -167.40 (t, $J = 19.1$ Hz).

Stochiometric reaction between CatBH and 3a at -23°C

In a J. Young-NMR tube, 3a (70.5 mg, 0.077 mmol) was dissolved in 700 μL CD₂Cl₂. The sample was frozen at -80°С. The frozen sample was transferred in glovebox and CatBH (9.3 mg, 0.077mmol) was added. The J. Young-NMR tube was delivered in precooled NMR spectrometer at -23°C. After 30 minutes, NMR spectra were recorded.

1H NMR (500 MHz, CD₂Cl₂) δ 7.91 (s, 1H), 7.28 – 7.20 (m, 1H), 7.15 – 7.08 (m, 1H), 7.08 – 6.93 (m, 1H), 3.45 (d, J = 5.2 Hz, 6H), 2.85 (t, J = 7.8 Hz, 2H), 1.57 (p, J = 7.7 Hz, 2H), 1.36 (h, J = 7.3 Hz, 2H), 0.90 (t, J = 7.4 Hz, 3H).

 $1H-1H$ COSY NMR (500 MHz, CD₂Cl₂). Cross-peak at {3.45, 7.90} shows correlation between NH and dimethylamino group. This cross-peak demonstrates artifact, where it does not have visible symmetric counterpart at {7.90, 3.45}. Such artifacts can occur when there are big differences in T_2 -values of the coupling partners and the acquisition times in indirectly detected dimension f_1 , t_{1max} , and in directly detected f_2 -dimension , t_2 , are very different (as is commonly the case). The severity then is dependent on the utilized window functions⁵. Here, the observed COSY cross peak between NH and dimethyl group indicate existing $2J_{HH}$ and thus acidic proton is still bonded to dimethylamino group.

NOESY NMR (500 MHz, CD_2Cl_2). NOESY cross-peaks at {7.90, 3.47} and {3.46, 7.87} show correlation between NH and dimethylamino group, indicating proximity of dimethylamino group and acidic proton.

Stochiometric reaction between CatBH, 2a and BCF

In a J. Young-NMR tube, **2a** (15 mg, 0.065 mmol), CatBH (7.7 mg, 0.065 mmol) and BCF (33.1 mg, 0.065 mmol) were dissolved in 700 μ L C $_6$ D $_6$. 11 B NMR spectra were recorded after 1h and 24h. Then, the sample was heated 24h additionally at 80 $^{\circ}$ C and ¹¹B NMR was recorded again. [CatBH-**2a**] was observed at -12 ppm (doublet). After heating the sample all CatBH, BCF and CatB-oligomers are decomposed forming C_6F_5BC at signal at 29.5 ppm.³

Crystal Structure Determination of 3a

The single-crystal X-ray diffraction study was carried out on a STOE IPDS 2T diffractometer with STOE image plates detector at 120(2) K using Mo-K α radiation (λ = 0.71073 Å). Dual space methods (SHELXT)⁵ were used for structure solution and refinement was carried out using SHELXL-2014 (full-matrix least-squares on *F²*).⁶ Hydrogen atoms were localized by difference electron density determination and refined using a riding model (H(N) free). An absorption correction by Gaussian integration (see cif-file for details) was applied. The absolute structure could not be determined reliably.⁷

3a colourless crystals, $C_{11}H_{16}F_3N_2 \cdot C_{24}BF_{20}$, $M_r = 912.31$, crystal size 0.25 × 0.13 × 0.08 mm, orthorhombic, space group *Pca*2₁ (No. 29), $a = 20.8363(9)$ Å, $b = 11.7036(4)$ Å, $c =$ 14.1501) Å, *V* = 3450.6(2) Å³, *Z* = 4, *ρ* = 1.756 Mg/m⁻³, *μ*(Mo-K_α) = 0.19 mm⁻¹, *F*(000) = 1808, $2\theta_{\text{max}}$ = 55.8°, T = 120 K, 24592 reflections, of which 8188 were independent (R_{int}) = 0.061), 556 parameters, 2 restraints, R_1 = 0.039 (for 6724 I > 2 $\sigma(1)$), w R_2 = 0.089 (all data), *S* = 1.06, largest diff. peak / hole = 0.21 / -0.21 e Å-3 . x = 0.4(4).

CCDC 2288120 (**3a**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Molecular structure of **3a** (displacement parameters are drawn at 50 % probability level). Selected bond lengths (Å): C1—N7 = 1.470(5); C1—N6 = 1.322(5); N7—C9 = 1.499(5); N7—C8 = 1.513(5); Selected bond angles (°): N6—C1—N7 = 115.1(3); C9—N7—C8 = 111.6(3).

Molecular structure of the cation of **3a** (displacement parameters are drawn at 50 % probability level).

Computing details

Data collection: *X-AREA* WinXpose 2.0.22.0⁸; cell refinement: *X-AREA* Recipe 1.37.0.0⁹; data reduction: *X-AREA* Integrate 2.5.1.0⁹, *X-AREA X-RED* 2.3.0.0⁹; program(s) used to solve structure: SHELXT⁷ ; program(s) used to refine structure: *SHELXL2014*/7⁷ .

3-butyl-2,4,5-trifluoro-N,N-dimethylpyridinium tetrakis(perfluorophenyl)borate (3a)

Crystal data for 3a

Data collection for 3a

Refinement for 3a

Computational details

All computational work reported here was carried out at the density functional theory (DFT) level, using Gaussian16 (Revision C.02).¹¹ The exchange correlation functional PBE1PBE¹²**-**¹⁴ was employed in conjunction with the Def2-TZVP15,16 basis set, Grimme's empirical dispersion correction (GD3BJ)¹⁷ and an ultrafine integration grid for the optimization and frequency calculations. Singlepoint calculations were performed for the Def2-TZVP optimized geometries at the PBE1PBE-GD3BJ/Def2-TZVP (PCM, solvent = dichloroethane)^{14,16,18} level. The nature of the stationary points and transition states was confirmed by analytical frequency calculations at the Def2-TZVP level and are characterized by zero (minima) or one (transition state) imaginary frequency. Intrinsic reaction coordinate (IRC) calculations were done to the optimized TS structures to ensure the transition states are connected to the respective minima on the potential energy surface.

Optimised xyz-coordinates

3a + HBcat

46

SCF done: -1243.838719

 12 SCF Done: -592.0493486

TS1

Int1

TS2

56

TS

C -0.588922 0.034714 1.505134 N 0.253397 -2.123106 -1.216361 C 0.242979 -3.562645 -0.962716 F -2.517707 -2.339695 -1.064920 C -4.162446 -0.987154 0.788850
C -4.837512 0.028303 -0.139986 C -4.837512 0.028303 -0.139986
C -6.352178 -0.084357 -0.100436 C -6.352178 -0.084357 -0.100436

C -7.030568 0.918720 -1.015925
F -2.576909 0.581147 2.587515 F -2.576909 0.581147 2.587515 F 0.050765 0.744850 2.403434 C 0.024085 -1.797988 -2.623574 H 2.083602 -1.716681 -0.621982 H 6.304385 -0.732401 -0.276462 H 5.349007 -1.566021 2.005858 H 3.142837 -2.342892 3.039092

 -0.126105

-0.060646

Product

NMR data

 $\frac{10}{210} \quad \frac{10}{200} \quad \frac{10}{190} \quad \frac{180}{180} \quad \frac{170}{170} \quad \frac{160}{150} \quad \frac{150}{150} \quad \frac{140}{140} \quad \frac{130}{120} \quad \frac{120}{120} \quad \frac{110}{110} \quad \frac{100}{190} \quad \frac{90}{190}$ $\overline{80}$ $\frac{1}{70}$ $\frac{1}{20}$ $\overline{10}$ $\overrightarrow{0}$ -10 60 50 40 $\frac{1}{30}$

 $\begin{array}{c} \begin{array}{c} \text{30.84} \\ \text{43.33} \\ \text{43.33} \\ \text{43.33} \\ \text{43.33} \\ \text{43.33} \\ \text{44.33} \\ \text{45.33} \\ \text{46.33} \\ \text{47.33} \\ \text{48.33} \\ \text{49.33} \\ \text{40.33} \\ \text{41.33} \\ \text{42.33} \\ \text{43.33} \\ \text{44.33} \\ \text{45.33} \\ \text{46.33} \\ \text{47.33} \\ \text{48.$

S46

 $\frac{1}{20}$ 10 0 10 20 30 40 50 60 -70 80 90 100 110 120 130 140 150 170 180 190 200 210 22
11 (ppm)

 $\begin{array}{ccccccccccccccccccccc} 210 & 200 & 190 & 180 & 170 & 160 & 150 & 140 & 130 & 120 & 110 & 100 & 90 & 80 & 70 & 60 & 50 & 40 & 30 & 20 \end{array}$ 10 $\overrightarrow{0}$

 -10

 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -22
 $f1(ppm)$ $\frac{1}{20}$ 10 $\frac{1}{\sqrt{2}}$

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